



Article Effects of Miniaturization of the Summer Phytoplankton Community on the Marine Ecosystem in the Northern East China Sea

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Abstract: After the construction of the Three Gorges Dam (Changjiang River), the northern East China Sea has been exposed to major environmental changes in the summer due to climate change and freshwater control. However, little is known regarding phytoplankton in this area. Here, we investigated differences in the summer phytoplankton-community structure as a consequence of marine-environment changes from 2016 to 2020. In the 2000s, the key dominant species in the summer phytoplankton community in the northern East China Sea were diatoms and dinoflagellates. In this study, however, nanoflagellates of $\leq 20 \,\mu m$ were identified as the dominant species throughout the survey period, with abundances ranging from 43.1 to 69.7%. This change in the phytoplanktoncommunity structure may be ascribed to low nutrient concentrations in the area, especially phosphate, which was below the detection limit, seriously hampering phytoplankton growth. The relative contribution of picophytoplankton to the total chlorophyll a biomass was highest in the surface mixed layer with low nutrient concentrations. Spatially, higher percentages were observed along the east-side stations than the west-side stations, where nutrient concentrations were relatively high. Conclusively, decreased nutrients led to phytoplankton miniaturization. Accordingly, as the dominance of picophytoplankton increases, energy transfer is expected to decrease at the upper trophic level.

Keywords: northern East China Sea; Changjiang diluted water; phytoplankton community; chl-*a* size fraction; picophytoplankton; phosphate restriction

1. Introduction

The northern East China Sea is exposed to various currents depending on the season and exhibits singular seasonal fluctuations [1]. It is known to be a highly valuable fishing ground because of its high primary productivity [2–4]. In summer, in particular, it shows diverse water mass characteristics, with the surface layer affected by the freshwater flowing in from the coastal areas of China, the bottom layer affected by the cold deep water of the Yellow Sea, and the eastern part affected by the high temperature and salinity of the Kuroshio water. The fronts formed at the boundaries of water masses with different characteristics show unique biological and chemical patterns and have significant effects on the distribution of zooplankton and fish along the food chain [5]. The inflow of freshwater due to Changjiang diluted water shows marked seasonal variability, with the minimum discharge in the winter and the maximum discharge in the summer, which greatly affects the seasonal salinity distribution in the northern East China Sea [6,7]. Moreover, as a major nutrient source [8], the coastal waters of China that are adjacent to the Changjiang are rich in nutrients, with high primary productivity [9–11] and characteristics of an estuary dominated by Chinese coastal waters. Geographically, a large difference in water depths is



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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). observed between the eastern and western waters, and complex topographical characteristics and diverse water masses show complex environmental interactions with seasonally variable intensity. Thus, the East China Sea, which is characterized by high productivity, is used jointly by Korea, Japan, and China as the largest fishing ground for migratory fish species [3]. Studies on phytoplankton distributions in the East China Sea were mainly conducted by Chinese and Japanese researchers in the 1980s and 1990s [12–18]. The focus areas of phytoplankton abundances and chlorophyll a (chl-a) concentrations were high in the waters affected by the Changjiang dilution water in spring and autumn [4], changes in phytoplankton standing stocks due to the stratification of the bottom layer structure in summer [19], the relationship between phytoplankton-community structures and water masses in summer [20,21], zooplankton (phytoplankton predators) [22–25], and changes in chl-a concentrations related to nutrients and suspended matter along with changes in phytoplankton-community structures [26]. In Korea, studies have been conducted on the changes in phytoplankton-community distributions and structures [27–30] since the 1990s. Several studies have been conducted on different topics, including the distribution characteristics of chl-a according to summer nutrients and suspended substances [31,32] and long-term changes in surface water temperatures and chl-a biomasses [33]. It was also reported that the primary productivity decreased by approximately 86% in waters adjacent to the Changjiang River after the construction of the Three Gorges Dam and that the phytoplankton-community structure in the East China Sea changed [34,35]. However, there appears to be a lack of recent data on this subject matter. Phytoplankton communities in marine ecosystems are sensitive to environmental changes, resulting in noticeable changes in community compositions and standing stocks following changes in physical- and chemicalenvironmental factors. Therefore, these changes in the phytoplankton-community structure can be used as an indicator of changes in the marine ecosystem. Moreover, in order to properly understand the structure and function of a marine ecosystem, it is necessary to understand temporal and spatial changes that occur in the phytoplankton-community structure in response to environmental factors [36]. The results of this study and those of previous studies were compared to confirm the change in the marine environment in the northern East China Sea due to the effect of constructing the Three Gorges Dam and the effects on the pelagic ecosystems according to the changes in the phytoplankton community structure were investigated.

2. Materials and Methods

2.1. Cruises and Sampling

To determine the distribution profiles of summer phytoplankton communities in the northern East China Sea, we conducted five field surveys from 2016 to 2020 (23 August–6 September 2016; 27 August–6 September 2017; 2–10 August 2018; 20–30 August 2019; and 3–15 August 2020) at the study site. The study site covered three lines and 15 sampling stations, and the surveys were conducted using the ocean research vessel, Tamgu 3 (797 tons; National Institute of Fisheries Sciences), as indicated in Figure 1 and Table 1. For nutrient analysis and quantitative analysis of phytoplankton, we collected samples at seven maximum water depths (0, 10, 20, 30, 50, 75, and 100 m) using Niskin bottles (8 L polyvinyl chloride) attached to a CTD/rosette sampler. The vertical distributions of water temperatures and salinities were measured using a calibrated SBE 9/11 CTD instrument (Sea-Bird Electronics, Bellevue, WA, USA), where descending data were used for the CTD data analysis.

2.2. Dissolved Inorganic Nutrients

For nutrient analysis, we filtered seawater samples (10 mL) through 0.45 μ m disposable membrane filter units (Advantec, Tokyo, Japan), placed the samples in conical tubes (15 mL), washed them with hydrochloric acid (HCl, 10%), and immediately stored them at -20 °C. After thawing the samples at room temperature (20 ± 2 °C), we analyzed the ammonium (NH₄), nitrite (NO₂), nitrate (NO₃), phosphate (PO₄), and silicon dioxide (SiO₂)

concentrations with an automatic nutrient analyzer (Quaatro, Seal Analytical, Norderstedt, Germany). The sum of the NH₄, NO₂, and NO₃ concentrations was calculated as the dissolved inorganic nitrogen (DIN).



Figure 1. Sampling station in the northern East China Sea, 2016–2020. CDW: Changjiang diluted water; YSWC: Yellow Sea cold water; KW: Kuroshio water; TCWW: Taiwan current warm water; TSW: Tsushima surface water.

Table 1. Description of the sampling sites in the northern East China Sea for the cruise period from 2016 to 2020.

Station	Latitude	Longitude	Bottom Depth (m)
315-13	32.5	127.0	125
315-15	32.5	126.5	107
315-17	32.5	125.9	91
315-19	32.5	125.3	68
315-21	32.5	124.5	46
316-13	32.0	127.0	119
316-15	32.0	126.5	99
316-17	32.0	125.9	76
316-19	32.0	125.3	55
316-21	32.0	124.5	39
317-13	31.5	127.0	105
317-15	31.5	126.5	88
317-17	31.5	125.9	67
317-19	31.5	125.3	54
317-21	31.5	124.5	46

2.3. Phytoplankton Abundances and Dominant Species

For quantitative analysis of phytoplankton, each sample was collected in a 1 L square PE bottle at a standard water depth at each sampling station, fixed with Lugol's solution (diluted to a final concentration of 1%), and transported to our laboratory. After 48 h decantation, the lower layer (200 mL) containing the sedimented algae was put in a small measuring cylinder. After being allowed 48 h for decantation, the lower layer (20 mL) containing the sedimented algae was put in a small concentrated samples were observed under an optical microscope (Nikon eclipse, Ni-U, Nikon Imaging Japan Inc., Tokyo, Japan) at $100 \times to 1000 \times$ magnification, using a Sedwick-Rafter Chamber for species identification and counting [37–40]. The resulting data were

converted to the number of cells L^{-1} , and species accounting for >5% of the total standing stock were classified as dominant species.

2.4. Picophytoplankton Abundances

To prepare picophytoplankton samples, collected seawater was filtered through a 3 µm polycarbonate membrane filter (Whatman, Florham Park, NJ, USA), split into 5 mL aliquots, and placed in cryogenic tubes. Glutaraldehyde was added to each tube at a final concentration of 1% and allowed to settle for 15 min at room temperature. The samples were then stored at -80 °C. To prepare the samples for picophytoplankton counting, they were thawed shortly before analysis and mixed with yellow-green fluorescent microspheres (0.5 µm diameter beads; Polysciences, Inc., Warrington, PA, USA), an internal reference material for standardizing scattering and fluorescence. Counting was conducted using a flow cytometer equipped with a 488 nm (1 W) argon ion laser (NovoCyte 2060R, ACEA Biosciences Inc., San Diego, CA, USA; BD AccuriTM C6 Plus, BD Biosciences Inc., Franklin Lakes, NJ, USA) (Figure 2), and each picophytoplankton group was distinguished based on the characteristic side scattering of red fluorescence at a 90°-angle, the chl-a concentration, and the orange fluorescence from phycoerythrin (Figure 2). The flow cytometry data were analyzed using NovoExpress (Ver. 1.2.5, ACEA Biosciences Inc., San Diego, CA, USA) and BD Accuri^{1M} C6 Plus Analysis Software (Ver. 1.0.23.1, BD Biosciences Inc., Franklin Lakes, NJ, USA).



Figure 2. Flow-cytometric analysis of a picophytoplankton sample. The signatures of each picoplankton group were discriminated based on their orange and red fluorescence intensities. PerCP-H: red-fluorescence, PE-H: orange-fluorescence, P1: picoeukaryote count, P2: *Synechococcus* count, P6: *Prochlorococcus* count.

2.5. Chl-a Size Fractionation

Chl-*a* concentrations were calculated as previously described [41]. To measure chl-*a* concentrations according to the phytoplankton size (>20 µm: micro; 20 µm \geq chl-*a* > 3 µm: nano; \leq 3 µm: pico), each phytoplankton sample (0.5 L) was sequentially filtered through a 20 µm membrane (Polycarbonate Track Etched Membrane disk, 47 mm diameter, GVS, Sanford, ME, USA), a 3 µm polycarbonate membrane filter (47 mm diameter, Whatman, Florham Park, NJ, USA), and a 0.45 µm membrane filter (47 mm diameter, ADVANTEC, Tokyo, Japan) and mounted on the filter holders. After measuring the chl-*a* concentrations in the microphytoplankton, nanophytoplankton, and picophytoplankton, the total chl-*a* concentration was calculated by addition. All filters were transferred to our lab in frozen storage (-80 °C), and chl-*a* was extracted after solvation in 90% acetone and settling in a dark and cool chamber for 24 h. The extract was then filtered through a 0.45 µm syringe filter (PTFE, Advantec, Florham Park, NJ, USA) to remove particulate matter. Finally, the

absorbance values were measured using a 10-Au field fluorometer (Tuner Designs, San Jose, CA, USA) calibrated with standard chl-*a* (Sigma-Aldrich, Darmstadt, Germany).

2.6. Data Analyses

The R statistical program (Ver. 4.0.3) was used to analyze statistical correlations between environmental factors and phytoplankton groups collected in the northern East China Sea. First, the decorana function of the vegan package of R was used to examine the distributions of biological parameters, which revealed that the DCA1 axis length (1.6993) was less than 3. Accordingly, a redundancy analysis (RDA) was performed.

3. Results

3.1. Physical Environments

Figures 3 and 4 show plots of the surface and vertical profiles of the mean water temperature and salinity in the northern East China Sea. The average surface water temperature in August ranged from 27.3 to 28.9 °C, with an overall average of 28.0 ± 0.5 °C and no distinctive spatial-distribution profile (Figure 3, left). The distribution range of surface salinity was 29.1–32.1, with an average of 30.2 ± 0.9 and a low-west and high-east distribution profile (Figure 3, right). The average vertical water temperature ranged from 15.1 to 28.9 °C, with an overall average of 22.6 ± 4.8 °C, and revealed that a strong thermocline formed at a depth of 20–30 m (Figure 4, top). The average salinity ranged from 29.1 to 34.5 psu, with an overall average of 32.0 ± 1.6 psu (Figure 4, bottom). The western part of the sea had a low salinity (\leq 31.0 psu from the surface layer to a depth of 20 m), reflecting the influence of the Changjiang dilution water. The deep layer of the eastern part of the sea had a high salinity of \geq 34.0 psu, reflecting the influence of the Kuroshio water. In August, the northern East China Sea formed a complex water mass structure under the influence of various water masses, depending on the water depths between the sampling stations.



Figure 3. Spatial distribution of the average surface temperature (\mathbf{a} , $^{\circ}$ C) and average surface salinity (\mathbf{b} , psu) in the northern East China Sea from 2016 to 2020.

3.2. Dissolved Inorganic Nutrient Concentrations

The average surface-layer DIN, PO₄, and SiO₂ concentrations in the northern East China Sea in August were 3.5–10.3 μ M, <0.1–0.1 μ M, and 2.2–5.5 μ M, respectively (Figure 5). The DIN concentration increased from north to south, while the PO₄ and SiO₂ concentrations showed a spatial-distribution profile with a gradual decrease from north to south. The average vertical DIN, PO₄, and SiO₂ concentration ranges were 2.5–19.5 μ M, <0.1–0.8 μ M, and 2.5–19.5 μ M, respectively (Figure 6). Throughout the survey period, the DIN tended to increase from the surface to the bottom layer, and the PO₄ concentration was extremely low in the surface mixed layer, at the detection limit of \leq 0.1 μ M.



Figure 4. Vertical distribution of average surface temperatures (**a**, $^{\circ}$ C) and average surface salinities (**b**, psu) in the northern East China Sea from 2016 to 2020.



Figure 5. Spatial distributions of average surface dissolved inorganic nitrogen (**a**, DIN), PO₄ (**b**), and SiO₂ (**c**) concentrations (in μ M) in the northern East China Sea from 2016 to 2020.

3.3. Phytoplankton Abundance and Dominant Species

In the summer, the average phytoplankton standing stock at the depth of each station ranged from 7955 to 1,090,202 cells·L⁻¹, with an overall average count of 149,656 \pm 192, 162 cells·L⁻¹. Spatially, in the western part of the sea (affected by the Changjiang River dilution water), the standing stock was approximately three times higher than that in the eastern part of the sea (affected by offshore waters). Due to strong vertical stratification in the summer, over 90% of the total standing stock appeared in the surface mixed layer (water depth: 0–30 m), demonstrating a remarkable difference between the surface and bottom layers. Nanoflagellates ($\leq 20 \ \mu$ m) appeared to be the dominant species throughout the survey period. Moreover, with *Scrippsiella acuminata* (which inhabits the coastal environment) reaching a high abundance in the western part of the sea, flagellates predominated in summer. In addition, at specific times during the survey, diatoms such as *Chaetoceros lorenzianus*, *Guinardia flaccida*, *Paralia sulcate*, and *Thalassiosira* spp., and dinoflagellates such as *Alexandrium* spp. appeared as the dominant species (Table 2).

3.4. Relative Contributions of Size-Fractionated Chl-a to the Overall Chl-a Concentration

The average surface-layer concentration of chl-*a* in the northern East China Sea in summer ranged from 0.19 to 1.25 μ g·L⁻¹, with an overall average of 0.57 \pm 0.33 μ g·L⁻¹ that gradually decreased going from west to east (Figure 7). The vertical distribution of

chl-*a* ranged from 0.04 to 1.99 μ g·L⁻¹ with an overall average of 0.52 ± 0.40 μ g·L⁻¹, where the distribution was higher in the surface mixed layer along the thermocline and gradually decreased going toward the bottom. In addition, the subsurface chl-*a* maximum layer developed at a depth of 10 to 20 m in the western part of the sea, and at 20 to 30 m in the eastern part, depending on the locations of the sampling stations (Figure 8).



Figure 6. Vertical distribution of the average DIN (**a**), PO_4 (**b**), and SiO_2 (**c**) concentrations (in μ M) in the northern East China Sea from 2016 to 2020.

Dominance	2016	2017	2018	2019	2020
1st	Nanoflagellates	Nanoflagellates	Nanoflagellates	Nanoflagellates	Nanoflagellates
	(<20 μm; 50.5%)	(<20 μm; 50.1%)	(<20 µm; 43.1%)	(<20 μm; 68.6%)	(<20 μm; 69.7%)
2nd	Scrippsiella	Chaetoceros	Paralia sulcata	Alexandrium spp.	Paralia sulcata
	acuminata (15.2%)	lorenzianus (14.3%)	(10.2%)	(8.4%)	(6.5%)
3rd	Guinardia flaccida (9.4%)	Diploneis spp. (11.0%)	<i>Chaetoceros</i> spp. (6.0%)		

The average relative contributions of microphytoplankton, nano phytoplankton, and picophytoplankton to the chl-*a* concentrations were 2.9–54.1% (23.2 \pm 11.9%), 13.7–59.9% (28.7 \pm 12.6%), and 16.4–770% (48.1 \pm 16.2%), respectively. Spatially, microphytoplankton contributed more to the chl-*a* concentration in the western part of the sea, and picophytoplankton contributed more in the eastern part of the sea, showing opposite spatial distributions. Vertically, the relative contributions of microphytoplankton and nanophytoplankton increased from the surface layer to the bottom layer, whereas that of the picophytoplankton tended to decrease toward the bottom layer (Table 3).



Figure 7. Spatial distributions of the average surface chl-*a* concentrations ($\mu g \cdot L^{-1}$) in the northern East China Sea from 2016 to 2020.



Figure 8. Vertical distributions of the average chl-*a* concentrations (μ g·L⁻¹) in the northern East China Sea from 2016 to 2020.

Table 3. Vertical variation of the chl-*a* size composition in the northern East China Sea from 2016 to 2020.

315 Line		Chl- <i>a</i> Size Composition (%)		31	316 Line		Chl- <i>a</i> Size Composition (%)		312	317 Line		Chl- <i>a</i> Size Composition (%)		
St.*	Depth (m)	М	Ν	Р	St.	Depth (m)	М	Ν	Р	St.	Depth (m)	М	Ν	Р
	0	12.0	18.4	69.6		0	12.6	14.0	73.4		0	20.2	15.0	64.8
	10	10.7	17.4	71.9		10	9.3	13.7	77.0		10	16.8	17.0	66.2
	20	5.2	18.6	76.2		20	17.0	16.1	66.9		20	16.6	18.9	64.5
13	30	10.5	21.2	68.3	13	30	10.6	28.8	60.6	13	30	12.6	17.0	70.4
	50	13.2	21.9	64.9		50	11.3	19.8	68.9		50	17.8	25.3	56.9
	75	18.4	31.5	50.1		75	13.7	27.6	58.7		75	18.6	30.3	51.1
	100	18.2	44.5	37.3		100	16.7	42.0	41.3		100	23.9	42.3	33.8
	0	14.3	32.6	53.1		0	17.9	22.3	59.8		0	12.4	22.9	64.7
	10	8.6	23.9	67.5		10	11.9	21.0	67.1		10	24.3	17.3	58.4
	20	14.2	18.5	67.3		20	2.9	22.2	74.9		20	20.5	21.7	57.8
15	30	25.8	21.0	53.2	15	30	15.0	29.5	55.5	15	30	23.2	27.5	49.3
	50	12.1	22.6	65.3		50	19.9	47.2	32.9		50	13.5	41.7	44.8
	75	10.0	33.9	56.1		75	13.0	58.2	28.8		75	23.1	50.9	26.0
	100	12.7	52.5	34.8		100	17.7	58.9	23.4					
	0	17.5	20.0	62.5		0	40.8	18.5	40.7		0	30.6	14.5	54.9
	10	19.8	21.1	59.1		10	32.5	17.3	50.2		10	26.0	18.3	55.7
17	20	10.0	29.6	60.4	17	20	24.1	20.9	55	17	20	40.7	16.7	42.6
	30	13.7	35.8	50.5	17	30	27.3	30.9	41.8	17	30	40.7	22.3	37.0
	50	9.9	47.8	42.3		50	24.8	51.2	24.0		50	24.3	45.6	30.1
	75	26.9	52.8	20.3		75	23.7	59.9	16.4					

31	5 Line	(Con	Chl-a Siz	e 1 (%)	31	6 Line	Cor	Chl-a Siz nposition	e 1 (%)	31	7 Line	Con	Chl-a Siz npositior	e 1 (%)
St.*	Depth (n	n) M	Ν	Р	St.	Depth (m)	Μ	Ν	Р	St.	Depth (m)	М	Ν	Р
	0	23.8	19.4	56.8		0	37.2	23.1	39.7		0	30.9	21.2	47.9
19	10	20.7	21.4	57.9		10	34.7	21.2	44.1		10	27.3	18.3	54.4
	20	23.4	27.5	49.1	19	20	27.1	28.3	44.6	19	20	28.8	19.9	51.3
	30	8.9	40.6	50.5		30	37.1	40.3	22.6		30	44.2	30.5	25.3
	50	20.8	58.5	20.7		50	25.6	55.7	18.7		50	19.6	47.0	33.4
	0	29.9	15.8	54.3		0	53.6	18.1	28.3		0	34.4	29.6	36.0
01	10	41.9	19.7	38.4	01	10	45.8	17.7	36.5	01	10	30.6	31.9	37.5
21	20	40.4	32.3	27.3	21	20	47.1	23.1	29.8	21	20	54.1	16.8	29.1
	30	51.4	29.6	19.0		30	35.5	34.5	30.0		30	42.8	25.0	32.2
	Mean	18.8	29.3	51.9		Mean	24.4	30.4	45.2		Mean	26.6	26.1	47.3
	SD	10.9	12.1	16.0		SD	12.9	14.7	18.3		SD	10.6	10.7	13.7

Table 3. Cont.

*St.: station; M: micro (>20 μ m); N: nano (20 μ m \geq chl-*a* > 3 μ m); P: pico (\leq 3 μ m).

3.5. Picophytoplankton Cell Abundances

Figure 9 shows plots of the picophytoplankton standing stocks in three groups, as determined by flow cytometry. *Synechococcus* and Picoeukaryotes cells did not show spatially distinct distribution profiles, whereas *Prochlorococcus* cells appeared to be limited at some east-side sampling stations (13, 15, 17 on each line) and abundant in the west-side sampling stations. Regarding the vertical distribution of picophytoplankton in summer, all three genera showed the highest standing stocks at the depth where the thermocline formed (rather than at the surface layer) and gradually decreased below the thermocline.



Figure 9. Vertical distributions of *Synechococcus* (**a**), *Prochlorococcus* (**b**), and picoeukaryote (**c**) cell abundances (cells·mL⁻¹) found at each station in the northern East China Sea from 2016 to 2020.

4. Discussion

4.1. Changes in Phytoplankton-Community Structures in Response to Changes in the Summer Marine Environment

To determine the changes in the phytoplankton-community structure associated with changes in the marine environment of the northern East China Sea in the summer, we compared the data obtained from the same sampling stations (Lines 315 and 316) at the same depth in a previous study and in this study (Table 4). The comparison revealed decreased nutrients (especially phosphate) in the surface layer as a characteristic difference in the marine environment. In terms of the community structure, diatoms and dinoflagellates were identified as the dominant species in the previous study conducted by Oh [42], and nanoflagellates ($\leq 20 \ \mu$ m) were identified as the dominant species in this study. This difference may be ascribed to decreased nutrients, especially in terms of phosphate. Among the nutrients flowing into the East China Sea, the DIN concentration in the estuary of the Changjiang River doubled from 1985 to 1998, and the DIN flux increased by 1.3-fold [43]. As a result, in the estuary and coastal waters of China, the area containing red tides increased

from 2000 km² in the 1980s to over 7000 km² in the 2000s [44] and contributed to a 21-fold increase in the number of red tide occurrences. The frequent occurrence of red tides in the Changjiang River estuary consumes a large amount of phosphorus, resulting in a phosphorus deficiency (rather than excessive nitrogen) [11,45]. The Changjiang dilution water affects the northern East China Sea. However, the transport volume decreased by 9–18% after the construction of the Three Gorges Dam (2003–2009), and the amount of suspended solids decreased by up to 56% [46]. The decrease in the inflow of the Changjiang dilution water also decreased the amount of nutrients, which caused a change in the community structure of phytoplankton from being dominated by diatoms to being dominated by flagellates [47]. Furthermore, the main source of phosphorus in the East China Sea is the Kuroshio water mass, although its influence has recently decreased in the northern East China Sea. In this regard, it was reported that the phosphate inflow decreased as the influence of the Taiwan warm current (which is caused by upwelling of the Kuroshio water mass) also weakened [48]. The decrease in phosphate in the East China Sea caused a decrease in the proportion of diatoms from 85% in 1984 to 60% in 2000 [49]. These results were observed in both the East China Sea and the Yellow Sea. In this respect, Lin et al. [43] reported that the standing stock of diatoms markedly decreased from the 1980s to the 1990s because of the increased surface-water temperature of the Yellow Sea and decreased silicate and phosphate concentrations. Likewise, Lee [50] reported extremely low phosphate concentrations, even below the detection limit, in a study of the central part of the Yellow Sea in summer, which resulted in a phytoplankton transition due to phosphate restriction, in which flagellates ($\leq 10 \mu m$) were the dominant species at all sampling stations, except for some bottom layer-dominant species, such as species in the *Navicula* spp. and Skeleonema spp. It was also reported that the restriction of nutrients greatly affects the growth of phytoplankton when their concentrations fall below the minimum levels of DIN 1.0 μ M, phosphate 0.2 μ M, and silicate 2.0 μ M [51]. Phosphate can also act as an important factor that hampers the growth of diatoms rather than flagellates [49], and if phosphate is deficient, then even if nitrate is abundant, phytoplankton cannot use nitrate, which hinders growth [52]. In this survey, the surface mixed layer, which accounted for 90% of the total standing stock, maintained sufficient nitrate and silicate concentrations, but had an extremely low phosphate concentration, even lower than 0.1 μ M. The fact that nanoflagellates with relatively low nutrient requirements could dominate over diatoms in all sampling stations can be explained by the phosphate concentration acting as a limiting factor.

Table 4. Comparison of environmental variables and dominant species in the northern East ChinaSea between August 1998 [42] and August 2016 to 2020 (this study).

Parameters		August 1998	August 2016–2020
Temperature (°C)	Surface	27.4–29.4 (28.7 \pm 0.5)	$25.4 extrm{}31.3~(28.0\pm1.4)$
Temperature (C)	Vertical	$12.7–29.4~(21.5\pm5.9)$	11.8–31.3 (23.1 \pm 5.5)
Calinity	Surface	$26.7–29.7~(28.3\pm0.9)$	$26.2 extrm{-}34.0~(30.1\pm1.9)$
Samily	Vertical	27.5–34.7 (31.2 \pm 2.5)	26.2–34.6 (31.7 \pm 1.9)
Nitrito (M)	Surface	$0.07 – 0.36~(0.21\pm0.08)$	$0.010.16~(0.05\pm0.04)$
initile (µM)	Vertical	$0.07 – 0.57~(0.19\pm0.14)$	$0.011.61~(0.15\pm0.26)$
Niture to (NA)	Surface	$1.26 extrm{}3.54~(2.51\pm0.66)$	$0.1211.37~(2.77\pm2.70)$
Nitrate (µM)	Vertical	$0.87 extrm{}3.54~(2.05\pm0.74)$	$0.09 extrm{-}35.40~(5.87\pm5.54)$
\mathbf{D} has the $(\mathbf{u}\mathbf{M})$	Surface	$0.160.45~(0.25\pm0.09)$	ND-0.16 (0.05 \pm 0.04)
Thosphate (µM)	Vertical	$0.161.03~(0.39\pm0.23)$	ND-0.82 (0.21 \pm 0.24)
Ciliante (M)	Surface	$8.95{-}13.77~(9.69\pm1.45)$	$0.0111.46~(4.23\pm3.10)$
Sincate (µW)	Vertical	$8.95{-}13.77~(9.59\pm0.88)$	$0.01 extrm{-}40.17~(7.87\pm5.71)$
		Pseudonitzschia pungens	Nanoflagellates (<20 μm)
Dominant species		Prorocentrum dentatum	Scrippsiella acuminata
*		Skeleonema costatum	Paralia sulcata

Values in parentheses are means.

4.2. Changes in Phytoplankton Community Sizes and Structures in the Northern East China Sea

The results of this survey study indicated that the picophytoplankton contributed to >60% of the chl-*a* concentration in summer standing stock; in particular, the surface mixed layer (from the surface to a depth of 20 m), which had low nutrient concentrations, had the highest relative chl-a concentration. Spatially, higher chl-a percentages were observed at the east-side stations than at the west-side stations, where nutrient concentrations were relatively high (Table 3). These tendencies were verified using statistical analysis. RDA of environmental factors and the chl-a size revealed that the nutrient concentrations increased with increasing water depths, which was indicative of an environment where the nutrient availability became increasingly limited from the bottom layer to the surface layer due to strong stratification caused by the rise in the surface water temperature in summer. Regarding the relationship between the phytoplankton size and environmental factors, the fraction of nanophytoplankton increased toward the bottom layer, where nutrient concentrations were relatively high, whereas picophytoplankton were inversely related to all nutrients, showing the highest fraction at the surface layer, where nutrient concentrations were low. In particular, picophytoplankton had the strongest inverse correlation with phosphate, which was associated with extreme phosphate depletion in the surface mixed layer (Figure 10). Similar results have been reported in other sea areas in recent years. Size-fractionation analysis that integrated satellite-data analysis results and field observations in the Mediterranean Sea confirmed that picophytoplankton < 2 µm accounted for 31–92% of the total phytoplankton biomass with seasonal variations, primarily due to low nitrogen and phosphorus availability [53,54]. Agawin et al. [55] reported that the relative contribution of picophytoplankton to the total chl-a biomass was over 50% in Blanes Bay in the Mediterranean Sea, which was attributed primarily to high temperatures and nutrient-poor waters. The composition ratio of picophytoplankton was 44–90% and 54–64%, respectively, in the Mediterranean Tyrrhenian Sea and Levantine Basin waters [56,57]. An analysis of summer satellite data of the Yamato Basin and Japan Basin showed that the percentage of picophytoplankton was more than 50% [58]. In addition, the composition ratio of nanophytoplankton and picophytoplankton in many waters was reported to be over 60% [53,59–62] (Table 5). In summer surveys of the East Sea of Korea, the relative contribution of picophytoplankton to the total chl-a biomass ranged from 35 to 63% [58,63], whereas those of nanophytoplankton and picophytoplankton (\leq 20 μ m) were 74% on average in the surface layer of the eastern Yellow Sea, in a study focused on the dominance of small phytoplankton [50]. Similarly, Furuya et al. [64] showed that picophytoplankton made a high relative contribution (up to 80%) due to the oligotrophic environment of the surface layer in a summer survey of the northern East China Sea. Son et al. [33] also reported that their analysis of long-term satellite observations of the chl-*a* biomass led to the finding that the contribution of microphytoplankton to the chl-a concentration sharply decreased in the northern East China Sea, whereas those of nanophytoplankton and picophytoplankton increased. These results have a common denominator in that they were derived from oligotrophic environments, where smaller phytoplankton cells had larger surface areas per unit volume, making small phytoplankton become dominant, owing to their capacity for rapid nutrient exchange through the cell surface [65-67].

Miniaturization of the phytoplankton size can result in decreased primary productivity. Indeed, analysis of primary productivity data derived from the Moderate Resolution Imaging Spectroradiometer Aqua led to a report showing that the annual primary production in the East Sea is decreasing by 13% per decade, as a result of an increase in the relative contribution of picophytoplankton ($\leq 2 \mu m$) to the total biomass [68]. Moreover, Lee et al. [63] verified a strong inverse correlation between the dominance rate of phytoplankton of $\leq 5 \mu m$ and the total primary production in the Amundsen Sea and attributed this to the low rate of carbon uptake by small phytoplankton. The miniaturization of phytoplankton also affects the food web, and it is projected that the increase in picophytoplankton abundance would lead to the dominance of microzooplankton (<200 μm) and gelatinous zooplankton (salps, doliolids, and ctenophores) and a decrease in biomass [69]. Moreover, a previous study showed that mesozooplankton (<200 µm), e.g., copepods, did not directly feed on picophytoplankton, but rather indirectly fed on ciliates that feed on picophytoplankton [70]. Accordingly, as the dominance of picophytoplankton increases, the biomass of mesozooplankton that do not directly feed on them will likely decrease. Consequently, the marine food chain will likely change from a simple diatom-based web with high primary productivity to a complex microbial food web centering on small phytoplankton with low productivity, negatively affecting overall marine productivity and reducing the efficiency of carbon transfer to consumers at higher trophic levels (Figure 11).



Figure 10. Redundancy analysis (RDA) ordination plots showing relationships between environmental and biological conditions in the northern East China Sea.



Figure 11. Changes in the food chain due to succession of the phytoplankton community in the northern East China Sea.

	Relative Ratio (%)						
Area	Date	Pico Size	Nano Size	Micro Size	References		
Northen East China Sea	2018–2020/ seasonally	45.6	31.2	23.2	This study		
Blanes Bay	1997/summer	>50			[55]		
Levantine Basin	March 1992	54.3-64.2			[56]		
Tyrrhenian Sea	July 2005	44-81			[57]		
(South)	December 2005	76–90			[37]		
Western Subarctic Pacific	23–29 June 2010	63					
Japan Basin	5–11 July 2010	56			[58]		
Yamato Basin	18–20 July 2010	56					
Ulleung Basin	22–24 July 2010	38					
Mediterranean Sea		31–92			[53]		
Adriatic Sea (North) Atlantic	August 1986 and 1988; July 1987			10–23	[59]		
Meridional Transect (Oligortophic)	April, October 1996 April, October 1997	80	16	4	[60]		
Algerian Basin	October 1996	42-62	38-58		[61]		
South China Sea	summer 1998 winter 1998	63 51	22 14	16 36	[62]		

Table 5. Size fractionation of phytoplankton observed in the different coastal waters of the northernEast China Sea and global waters.

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