



Article Estimating Freezing Injury on Olive Trees: A Comparative Study of Computing Models Based on Electrolyte Leakage and Tetrazolium Tests

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Abstract: Winter frost injury is a major limiting factor for olive cultivation in temperate regions. The response of olive shoots to freezing stress can be used for selecting genotypes resistant to freezing. The electrolyte leakage (EL) and tetrazolium tests (TZ) are commonly used to evaluate dead tissues in cold stress studies. The temperature-response curve of dead tissues to lethal temperature (LT) is measured with models to calculate LT50 and LT90. In this study, we evaluated the accuracy and efficiency of eighteen nonlinear regression models (NLRs) in calculating LT50 and LT90 of freezing stress in different olive cultivars at various stages of dormancy. After evaluating the prediction performance of NLR models, it was found that only eight models were suitable for the purpose of this research out of the eighteen models examined. The 2p-logistic and Gompertz models were selected for modeling EL and TZ, respectively. Our research findings indicate that the *Roughani, Kawi,* and *Zard* varieties of olive trees exhibit the best performance under artificial temperature-controlled conditions. Our findings provide valuable insights into selecting frost-resistant cultivars and designing effective strategies for cold acclimation in olive cultivation.

Keywords: olive trees; freezing injury; electrolyte leakage; tetrazolium tests; nonlinear regression models

1. Introduction

Olive trees are a subtropical crop that can also thrive in certain temperate regions. The Mediterranean Basin heavily relies on the economic benefits of olive oil, which is extracted from the fruit of the olive tree. In addition to serving as a vital economic resource, table olives are also a popular food item [1]. However, due to their subtropical nature, olive trees are highly vulnerable to winter frost injury [2,3]. Numerous studies have been conducted to select frost-resistant cultivars of olive trees [4-8]. Some studies typically involve evaluating different cultivars based on their performance during natural frost events [6,9]. Most studies have also evaluated the viability of shoot tissue from different genotypes under controlled freezing temperatures [10,11]. The electrolyte leakage and tetrazolium tests are two important indicators used in the screening of plant frost injury, particularly in cold stress studies, to assess the presence of dead tissue [12]. Electrolyte leakage (EL) is determined by measuring the ratio of ion leakage from injured tissues to intact tissues using electrical conductivity [13]. Meanwhile, dead tissues are identified via staining and visually scored in the tetrazolium test (TZ). To screen for frost-hardy resistant genotypes, plant tissues at various stages of growth are exposed to controlled freezing temperatures [14,15]. By determining the temperature at which 50% of tissues are lost (known as the lethal temperature or LT50), computing models can designate the degree of genotype resistance to freezing stresses. The typical response of plant tissues to a series of freezing temperatures



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). is an asymmetric sigmoid curve [16]. Probit analysis is a common method used to determine LT50 in many studies related to freezing stress in plants, including olive trees [2]. Regression models such as linear interpolation, Richards, logistic, and Gompertz models have also been used for LT50 estimation in some studies [10,16–18]. Nonlinear regression is a statistical method used to model the relationship between a dependent variable and one or more independent variables when the relationship is not linear. In the context of freezing injury, nonlinear regression can be used to model the relationship between the extent of tissue damage and the temperature and duration of exposure to freezing temperatures [17,18]. This type of analysis can help researchers understand the factors that contribute to freezing injury and develop strategies to mitigate its effects. Nonlinear regression models can be more complex than linear regression models, and may require more sophisticated statistical techniques to fit to the data [19]. However, they can often provide a better fit to the data and more accurate predictions than linear models when the relationship between the variables is not linear [19]. The parameters in nonlinear models often have a biologically meaningful interpretation, based on the units and definitions associated with them [19–21]. This interoperability makes nonlinear models useful for gaining insights into biological processes such as cold stress. Nonlinear models also face some limitations. They tend to be less flexible than linear models, meaning the choice of model is crucial. Additionally, because no analytical solution exists for parameter estimation in nonlinear models, numerical methods must be used. This introduces challenges such as checking for algorithm convergence and choosing appropriate starting values for the parameters [19]. Overall, nonlinear regression can be a powerful tool for analyzing the complex relationships between freezing injury and duration of exposure and can help researchers develop a better understanding of this phenomenon.

During the growing season, the optimal temperature range for olive trees is around 20–30 °C. While olive trees can withstand minimum temperatures of about -4 °C in winter and maximum temperatures of about 50 °C in hot and dry summers, sub-optimal and chilling temperatures (between 7.5 and 12.5 °C) can significantly slow down metabolic processes such as respiration and photosynthesis. Freezing temperatures, defined as temperatures averaging below -10 °C, can cause irreparable damage to organs and tissues, and in severe cases, the entire plant can die [2]. Freezing stress is often more damaging to olive trees than chilling stress. The temperature threshold for frost damage in olive trees varies depending on factors such as genotype, growth conditions, and the timing of the low temperatures (early frost, winter frost, or late frost). Additionally, the hardening or de-hardening phase of the tree can also impact its susceptibility to frost damage [7,8,22,23]. Larcher [24] reported that the LT50 of olive twig cambium and xylem was -16 °C. Bartolozzi and Fontanazza [4] showed that the LT50 of higher-resistance olive cultivars, such as 'Bouteillan' and 'Nostrale di Rigali', were -18.2 and -18.1 °C, respectively, while the LT50 of the lower-tolerance 'Borsciona' and 'Morcona' were -12.28 and -11.48 °C, respectively. In addition to the effects of genotype, the severity of damage caused by freezing stress in olive trees can depend on a range of factors, including the duration of the freezing event, the stage of plant growth, and the cold acclimation or dormancy processes [25]. A series of physiological and metabolic changes occur during the cold acclimation process, which prepares olive trees for low-temperature tolerance [5,26]. Cansev, Gulen and Eris [5] evaluated the cold hardiness of twelve olive cultivars and found that cold acclimation increased the freezing tolerance of these cultivars. Osmolytes, such as soluble carbohydrates and proline, play a crucial role in the resistance of woody plants to freezing stress [2]. By accumulating these compounds, plants can protect themselves from the damage caused by ice formation [25].

The majority of published articles modeling the fruit-freezing process have utilized a limited number of nonlinear regression models, with many employing various models. Consequently, this article presents a comprehensive review of nonlinear regression models for the olive fruit within this field. The objectives of the paper are outlined as follows: (1) to evaluate the fitness of 18 computing models in pre-, deep, and post-dormancy of ten olive cultivars under freezing temperature, (2) compare accuracy and sensitivity analyses for each model, and (3) predict plant response with selected models at out-of-range temperatures.

2. Materials and Methods

2.1. Olive Cultivars and Orchard Management

Ten olive cultivars—'Dorsalani', 'Luke', 'Roughani', 'Shangeh', 'Rashid', 'Mission', 'Kawi', 'Abu-Stal', 'Manzanilla', and 'Zard' (coded as CV1 to CV10)—were sampled at three different times—before, during, and after dormancy, corresponding to November, January, and March. The olive trees were grown in an adaptation collection established in 2001, using a randomized complete block design (RCBD) with four replications and six trees in each plot. The trees were spaced 4×6 m apart and were 20 years old at the time of sampling. The orchard management included regular irrigation and fertilization, and the trees were free of pests and nutritional deficiencies.

2.2. Temperature Treatments

One-year-old shoots, 30 cm long and 1 cm in diameter, were collected from each olive cultivar in the germplasm collection and immediately transported to the laboratory. After removing the leaves and washing the branches with distilled water, the shoots were cut into smaller pieces, 1 cm in length, at the internodal distances. The pieces were then placed in a cold room at 4 °C for 3 h. Subsequently, the samples were subjected to a stepwise freezing process from 0 to -21 °C (0, -3, -6, -9, -12, -15, -18, and -21 °C) using a freezing test device (Kimiya Rahvard Co., Tehran, Iran). The freezing rate was set at 3 °C h⁻¹, and the duration of each test temperature was 60 min [14]. At each stage, after cold treatment at the desired temperature, the samples were removed from the freezing test device and used for electrolyte leakage and tetrazolium tests.

2.3. Electrolyte Leakage (EL)

To evaluate the extent of freezing damage on cell membranes, the rate of electrolyte leakage (EL) was calculated using Equation (1). Stem samples from each temperature treatment (ranging from 0 to -21 °C) were placed in 40 mL of distilled water at room temperature for 24 h. The electrical conductivity (EC1) was measured using an EC meter (Con500, Korea). The samples were then autoclaved at 120 °C for 2 h, and after cooling to room temperature, their ion leakage was measured using an EC meter (EC2). The percentage of conductivity was calculated for each sample using the ratio of the initial to the final measurements multiplied by 100.

$$EL(\%) = \frac{EC_1}{EC_2} \times 100$$
 (1)

2.4. Triphenyl Tetrazolium Chloride (TZ) Assay

The tetrazolium staining method was used to determine the percentage of dead tissues (Figure 1). Stem samples from each temperature treatment were placed in a test tube containing 5 mL of 1% 5-3-2-triphenyltetrazolium chloride solution and kept in the dark at room temperature for 24 h. The samples were then cut into thin sections using a microtome machine and observed under stereo microscope (Model $40 \times$ CS-5CAW). The extent of visual damage was measured as a percentage based on the staining of dead tissues in the phloem and cambium layers at the outer circle of the shoot pieces.

2.5. Nonlinear Regression Models (NLRs)

Eighteen nonlinear regression models (NLRs) were used to model freezing injury in olive cultivars (Table 1). Other studies have included only one or two models [7,10,17]. One important aspect of developing an NLR model for freezing injury is choosing the appropriate functional form for the model. NLR models can take on a variety of forms, including

polynomial, exponential, logarithmic, and sigmoidal functions, among others. The choice of functional form will depend on the nature of the relationship between the independent and dependent variables, as well as the specific research question being addressed, and be based on prior knowledge or theory about the relationship between the dependent and independent variables.



Figure 1. Cross-sectional area of olive stem width adjacent to tetrazolium color under stereo microscope ($40\times$). Blue arrowsrevealing tissue staining in the phloem and cambium layers. The left image displays staining in live tissue, while the right image shows staining in dead tissue.

Table 1. Mathematical models to estimate freezing injuries in ten olive cultivars.

Symbol	Form	Model	Name
EXP ₀	$y = 100(a - \exp(-b \times t))$	M1	Exponential model without LAG
EXPLAG	$y = 100(1 - a \times \exp(-b \times t))$	M2	Exponential model with LAG
L2p	$y = \frac{100}{(1 + a \times exp(-b \times t))}$	M3	2p-logistic model
GOM	$y = 100 \exp(-a \times \exp(-b \times t))$	M4	Gompertz model
LOG	$y = \frac{100}{(1 + \exp(\mathbf{a} - \mathbf{b} \times \mathbf{t}))}$	M5	Logistic model
GOM2	$y = 100 \exp(-\exp(1 - a \times (t - b)))$	M6	Gompertz model 2
LOG2	$y = \frac{100}{1 + \exp(1 + \mathbf{a} \times (\mathbf{b} - \mathbf{t}))}$	M7	Logistic model 2
TPLOG	$y = \frac{100}{1 + \exp(2 - 4a(t-c))} + \frac{100}{1 + \exp(2 - 4b(t-c))}$	M8	Two-pool logistic
OPLOG	$y = \frac{100}{1 + \exp(1 - a \times (t - b))}$	M9	One-pool logistic
MGOM	$y = 100 \exp(-\exp(a \times (t-b) + 1))$	M10	Modified Gompertz model
RCD	$y = \frac{100}{1 + (1 + \exp(-a \times t))^{\frac{1}{b}}}$	M11	Richard model
ELM	$y = \frac{100}{a \times \log(1 + \exp(a \times b \times t))}.$	M12	Exponential-linear model
OPG	$y = 100 \exp(-\exp(1 + a \times \exp(1) \times (b - t))).$	M13	One pool Gompertz
RCD2	$y = 100 \left(\left(1 + \exp(\mathbf{a} - \mathbf{b} \times \mathbf{t}) \right)^{-c} \right)$	M14	Richard model 2
RCD3	$y = \frac{100}{1 + \exp(-a(t-b))}$	M15	Richard model 3
RCD4	$y = \frac{100}{1 + 100}$	M16	Richard model 4
GOM3	$y = 100 \exp(-\exp(a - b \times t))$	M17	Gompertz model 3
GOM4	$y = 100 \exp(-\exp(-a \times (t - b)))$	M18	Gompertz model 4

The relationship between dependent variables (EL and TZ) and the independent variable (temperature) was analyzed using nonlinear regression. NLR models were fit using the fitnlm function in MATLAB software version R2020a using the Levenberg–Marquardt (LM) algorithm to minimize the sum of squares error (SSE) between the predicted and observed values of EL and TZ. NLR models are often fit using an iterative algorithm such as the LM. This algorithm iteratively adjust the parameters of the NLR until a minimum value of the SSE is reached. NLR algorithms can be sensitive to the initial parameter values, and different initial values can lead to different solutions. One common method for addressing this is to use multiple initial values and choose the best solution based on some criterion, such as the SSE. We used the cftool graphical interface in MATLAB to explore the data and select an appropriate starting point for the parameter estimates. The model was fit separately to the EL and TZ data, using the temperature data as the predictor variable and the EL and TZ as the response variables. Statistical significance was set at p < 0.05. To assess the goodness of fit of the NLR model, we calculated the coefficient of determination (R^2) , adjusted coefficient of determination (R^2_{Adi}) , residuals, normal probability plots of residuals, and the root mean squared error (RMSE). Before conducting the NLR analysis, the data were preprocessed to remove any outliers or missing values. The data were also checked for normality using the Shapiro-Wilk test, and transformed if necessary to meet the assumptions of the NLR model.

Freezing injury in trees is a topic that has garnered significant attention in the field of horticulture. However, one aspect that is often overlooked is the rate of behavior of trees in response to sub-zero temperatures, which is a critical consideration for fruit tree growers. With this in mind, the present study aims to shed light on this important issue by calculating the rate of changes in EL and TZ. To achieve this, the first-order derivative of the extracted regression functions with respect to sub-zero temperature was used, and the results are presented and discussed in the subsequent sections of this article. Through these analyses, the authors aim to provide valuable insights into the behavior of trees under freezing conditions and contribute to the development of effective strategies for fruit tree cultivation.

3. Results and Discussion

In this section of the article, we present the results under two headings: the modeling results of EL and TZ. We examine the nonlinear regression models for each indicator separately, and calculate the T50 and T90 values using the best-fitted model. Finally, we evaluate the cold tolerance of different olive varieties based on their resistance to freezing injury and provide recommendations for further discussion.

3.1. Findings of the Electrolyte Leakage (EL) Assay

3.1.1. Comparing and Selecting the Best-Fitted NLR Model for EL

As stated in the preceding section, eighteen models were employed to estimate the EL of ten distinct olive cultivars. The fitting outcomes of these models revealed that only eight of them exhibited a superior ability to predict EL, while the remaining models did not perform significantly worse in terms of prediction accuracy. Table 2 displays the RMSE values of eight models used to estimate EL in pre-, deep, and post-dormancy of ten olive cultivars. The results reveal that the rank of the models differed in terms of RMSE, indicating that the choice of the appropriate model for each cultivar and its dormancy stage is crucial. M7, M9, M15, and M16 models performed poorly compared to the other models, while M2, M3, M4, M5, M6, M8, M10, and M14 were identified as the best options. However, some models, such as M8, had lower prediction errors, but their coefficients were not significant. Further analysis showed that some models overestimated EL by more than 100% across the tested temperature range, including at lower temperatures. After considering the significance of coefficients, simplicity of application, and generalizability, the M3 model was selected for all varieties and assessment stages. This allows for comparing the varieties and their stages by comparing the coefficients' values of the NLR models. The lowest and

highest RMSE values across all cases were 3.29% and 9.06%, respectively. These findings suggest that choosing the appropriate model is crucial for accurate estimation of EL, and the M3 model is a reliable option for all varieties and assessment stages.

Table 2. The RMSE values for EL estimation of eight out of the eighteen investigated NLR models, for different cultivars of olives.

		CV1			CV2			CV3			CV4			CV5	
	Pre	Deep	Post												
M2	7.53	5.14	5.01	8.47	4.52	5.22	8.68	5.36	6.89	7.2	4.89	5.03	7.23	5.22	7.07
M3	8.5	4.37	3.49	9.06	4.84	6.03	4.89	3.96	5.39	8.04	3.5	5.45	7.95	6.77	8.44
M4	7.79	4.62	3.95	8.48	4.56	5.49	6.34	4.47	5.87	7.39	3.75	4.93	7.37	5.94	7.72
M5	8.5	4.37	3.49	9.06	4.84	6.03	4.89	3.96	5.39	8.04	3.5	5.45	7.95	6.77	8.44
M6	7.79	4.62	3.95	8.48	4.56	5.49	6.34	4.47	5.87	7.39	3.75	4.93	7.37	5.94	7.72
M8	8.46	4.38	3.3	9.04	4.84	6.03	3.6	3.64	5.21	8.02	3.48	5.45	7.88	6.78	8.45
M10	7.79	4.62	3.95	8.48	4.56	5.49	6.34	4.47	5.87	7.39	3.75	4.93	7.37	5.94	7.72
M14	7.79	4.3	3.4	8.45	4.57	5.49	3.82	3.76	5.41	7.31	3.5	4.93	7.22	5.95	7.72
		CV6			CV7			CV8			CV9			CV10	
	Pre	Deep	Post												
M2	7.83	3.29	6.18	6.91	7.24	6.46	6.98	6	5.69	7.51	5.52	6.4	8.61	4.22	7.48
M3	8.51	4.66	7.51	5.07	5.56	4.74	8.06	7.54	6.92	8.42	6.93	7.68	5.59	4.11	6.11
M4	7.91	3.87	6.74	5.5	6.17	5.32	7.33	6.8	6.28	7.77	6.18	6.95	6.73	4.06	6.57
M5	8.51	4.66	7.51	5.07	5.56	4.74	8.06	7.54	6.92	8.42	6.93	7.68	5.59	4.11	6.11
M6	7.91	3.87	6.74	5.5	6.17	5.32	7.33	6.8	6.28	7.77	6.18	6.95	6.73	4.06	6.57
M8	8.47	4.66	7.51	4.81	5.21	4.24	8.06	7.57	6.95	8.38	6.94	7.69	4.29	4.11	5.65
M10	7.91	3.87	6.74	5.5	6.17	5.32	7.33	6.8	6.28	7.77	6.18	6.95	6.73	4.06	6.57
M14	7.86	3.86	6.74	5.08	5.27	4.49	7.32	6.81	6.28	7.69	6.18	6.95	5.05	4.05	5.82

Means \pm SD, CV1 to CV10 are the olive cultivars, while Pre, Deep, and Post refer to the dormancy stages, and M2 to M14 are the NLR models (Table 1).

3.1.2. Assessment of NLR Model Coefficients for EL

After choosing the 2p-logistic model (M3) to predict EL in terms of temperature for different olive cultivars, its coefficient estimates were computed using the fitnlm function in the MATLAB software. The ANOVA table and the significance assessment of the model coefficients were also obtained. Table 3 displays the coefficient values, standard deviation, and significance results for the nonlinear regression model (M3) used to predict EL across ten different olive cultivars during three dormancy stages. Additionally, the table presents the corresponding R^2 and R^2_{Adj} values. The results indicate that both model coefficients, b₁ and b₂, are significant at the 1% level for all cases. These findings demonstrate that the M3 model is appropriate and applicable for all cases, with R^2 and R^2_{Adj} values ranging from 83% to 96%. Thus, the selected regression model successfully explained approximately 83% to 96% of the observed EL changes based on the experimental results.

3.1.3. Conducting a Sensitivity Analysis of the EL Model

To better understand the behavior of the M3 model, we plotted the EL predictions against temperatures below zero while varying the regression coefficients b_1 and b_2 (Figure 2). The graph shows that increasing the regression coefficient b_2 results in a steeper slope of EL changes with respect to temperature. Consequently, olive cultivars with higher b_2 values reach an EL value of 100% sooner, indicating greater sensitivity to cold temperatures. For instance, when b_2 equals 0.05 and 0.20, the EL value reaches 100% at temperatures higher than -70 and about -30 °C, respectively. On the other hand, an increase in the regression coefficient b_1 leads to a shallower slope of EL changes with respect to temperature. However, changes in b_1 do not affect the temperature required to achieve an EL value of 100%. Notably, lower values of b_1 indicate greater sensitivity of the olive to cold temperatures, while the opposite may also be true.

			CV1		CV2			
		Pre	Deep	Post	Pre	Deep	Post	
Cafe	b_1	$2.47 \ ^{**} \pm 0.41$	3.62 ** ± 0.31	3.98 ** ± 0.28	2.63 ** ± 0.41	3.07 ** ± 0.28	2.67 ** ± 0.29	
Core.	b_2	0.18 ** \pm 0.02	$0.09 ** \pm 0.01$	$0.11~^{**}\pm0.01$	$0.18 \ ^{**} \pm 0.02$	$0.10~^{**}\pm0.01$	$0.11~^{**}\pm0.01$	
R^2 , R^2_{Adj} .		0.87, 0.86	0.92, 0.92	0.96, 0.96	0.90, 0.90	0.92, 0.91	0.89, 0.89	
			CV3			CV4		
		Pre	Deep	Post	Pre	Deep	Post	
Cofo	b_1	$5.60 ** \pm 0.63$	$4.48 ** \pm 0.30$	$4.51 ** \pm 0.47$	$2.43 ** \pm 0.38$	$3.81 ** \pm 0.27$	3.14 ** ± 0.33	
Cole.	b_2	0.16 ** \pm 0.01	$0.10 \ ^{**} \pm 0.00$	$0.10 \ ^{**} \pm 0.01$	$0.18 \ ^{**} \pm 0.02$	$0.12 \ ^{**} \pm 0.01$	$0.12 \ ^{**} \pm 0.01$	
R^2 , R^2_{Adj} .		0.96, 0.96	0.97, 0.97	0.92, 0.91	0.88, 0.88	0.97, 0.97	0.92, 0.91	
			CV5			CV6		
		Pre	Deep	Post	Pre	Deep	Post	
Cafa	b_1	2.36 ** ± 0.36	2.65 ** ± 0.33	2.26 ** ± 0.34	2.50 ** ± 0.36	2.54 ** ± 0.20	2.32 ** ± 0.32	
Cole.	b_2	0.18 ** \pm 0.02	0.12 ** \pm 0.01	$0.12 \ ^{**} \pm 0.01$	$0.18 \ ^{**} \pm 0.02$	$0.14~^{**}\pm0.01$	$0.14~^{**}\pm0.01$	
R^2 , R^2_{Adj} .		0.88, 0.88	0.88, 0.87	0.81, 0.80	0.91, 0.91	0.96, 0.96	0.87, 0.86	
			CV7			CV8		
		Pre	Deep	Post	Pre	Deep	Post	
Cafe	b_1	3.50 ** ± 0.30	$4.65 ** \pm 0.56$	4.38 ** ± 0.44	2.42 ** ± 0.38	2.48 ** ± 0.33	2.37 ** ± 0.29	
Core.	b_2	0.14 ** \pm 0.01	$0.11~^{**}\pm0.01$	$0.11 \ ^{**} \pm 0.01$	$0.18~^{**}\pm0.02$	$0.10~^{**}\pm0.01$	$0.10~^{**}\pm0.01$	
R^2 , R^2_{Adj} .		0.96, 0.96	0.90, 0.90	0.93, 0.92	0.88, 0.87	0.83, 0.82	0.85, 0.84	
			CV9			CV10		
		Pre	Deep	Post	Pre	Deep	Post	
Cafe	b_1	2.57 ** ± 0.36	2.64 ** ± 0.33	2.32 ** ± 0.32	4.62 ** ± 0.56	3.67 ** ± 0.30	$4.70 ** \pm 0.55$	
Core.	b_2	0.18 ** \pm 0.02	$0.11 \ ^{**} \pm 0.00$	$0.13 \ ^{**} \pm 0.01$	$0.15~^{**}\pm0.01$	$0.08 \ ^{**} \pm 0.01$	$0.10~^{**}\pm0.01$	
R^2 , R^2_{Adj} .		0.92, 0.92	0.87, 0.86	0.86, 0.85	0.94, 0.93	0.90, 0.90	0.87, 0.86	

Table 3. The statistical analysis results of the coefficients for the M3 model used to estimate EL based on sub-zero temperatures for all experimental cases.

Means \pm SD. CV1 to CV10 are the olive cultivars, while Pre, Deep, and Post refer to the dormancy stages, b_1 and b_2 are model coefficients, R^2 is the coefficient of determination, and $R^2_{Adj.}$ is the adjusted coefficient of determination.



Figure 2. Effect of modifying regression coefficients b_1 and b_2 in the M3 model on olive susceptibility to freezing injury: (a) changing the coefficient of model b2 with b1 constant at the average value, and (b) changing the coefficient of model b1 with b2 constant at the average value.

3.1.4. Comparison of EL Modeling Outcomes for Different Olive Cultivars

The Supporting Information section of the article includes the predicted results of the M3 model for the EL parameter of olive trees at the tested temperatures, as well as the lower and upper bounds of the prediction for the three stages of pre-, deep, and postdormancy for ten olive cultivars. The results presented in Figure S1 demonstrate that the M3 model provided estimates of EL that fell within the 95% prediction interval of the model for most of the laboratory data. Although some observed values deviated slightly from the predicted values at certain temperatures, the overall results were deemed acceptable. Additionally, the coefficient of determination supported the accuracy of the model.

Figure 3 depicts the extrapolated results of EL modeling for each of the ten olive cultivars at the pre-, deep, and post-dormancy stages, up to a temperature of -50 °C. The results of this study indicate that the pre-, deep, and post-dormancy stages of olive trees during temperature reduction to -50 °C are associated with high-to-low estimation of EL. The models of deep and post-dormancy stages are very similar, while the pre-dormancy model differs significantly. These findings suggest that olive trees are particularly vulnerable to freezing injury during the pre-dormant stage. It is important for olive growers to be aware of this vulnerability and take appropriate measures. If the pre-dormancy stage is successfully managed, olive trees can adapt to their environment and improve their resistance to freezing injury [5]. Comparing the tolerance of ten different olive cultivars to freezing injury in terms of EL at both the deep-dormancy and post-dormancy stages revealed that the cultivars CV10, CV7, CV1, CV4, CV2, CV8, CV5, CV9, CV3, and CV6 exhibited the highest-to-lowest resistance to freezing injury. The ranking of olive cultivars according to their resistance to freezing injury during the pre-dormancy stage was as follows: CV3, CV10, CV7, CV4, CV1, CV9, CV6, CV2, CV5, and CV8. The adaptability of ten different olive cultivars to freezing weather was evaluated in the field. In November 2016, a sudden drop in temperature from 10 °C to -12 °C caused frost damage to all the trees in the olive orchard. All cultivars lost their leaves and branches, with varying degrees of damage to the trunks. The percentage of trunk damage, from the highest to the lowest, was observed in CV6, CV9, CV5, CV2, CV4, CV1, CV8, CV10, CV3, with the highest percentage of trunk damage occurring in CV6 to CV9 with 88% to 92%, while the least damage occurred in CV3, with 23%. These findings mostly confirm our experimental results obtained in the pre-dormancy stage with field evaluations. These results provide valuable information for selecting appropriate olive cultivars based on the prevailing climatic conditions and management considerations in the region.

3.1.5. Analysis of the Rate of Change in EL Results

The NLR model can be applied to calculate the rate of EL changes per decrease in sub-zero temperature. To achieve this, the first-order derivative is taken from the NLR regression function. Figure 4 shows the track of EL changes for ten olive cultivars in the pre-, deep, and post-dormancy stages. The results demonstrate that, at the beginning of exposure to cold and sub-zero temperatures, the increasing EL rate gradually decreases until reaching a certain temperature, such as -15 °C, after which it changes to an upward trend. In plants undergoing freezing stress, ice crystal formation can occur intracellularly and extracellularly. Extracellular ice formation occurs at an early stage when the temperature drops below the freezing point [27]. Due to the latent heat of water, a slight increase or stabilization in temperature can be observed. However, as the temperature continues to drop or rapid freezing occurs, intracellular ice formation predominates, leading to mechanical disruption of the protoplasmic structure and eventual cell death [25,28]. Furthermore, different olive cultivars exhibit varying reactions with different starting temperatures for EL increase. In the pre-dormancy stage, the starting temperature is smaller in absolute value than in the other two stages. Moreover, the rate of EL change reaches a constant value sooner in the pre-dormancy stage. This may be attributed to the lower concentration of osmolytes in the intercellular sap during the early stages of acclimation in olive trees [5]. The primary mechanism used by the tree to withstand winter cold is the accumulation of carbohydrates and osmolytes within its tissues [25,29]. However, in the pre-dormancy stage, the concentration of these substances has not yet reached its maximum level, resulting in a narrower model curve during this stage. Therefore, the steeper slope in the trend of variation indicates that the olive variety is more susceptible to damage from freezing.



Figure 3. Nonlinear regression modeling of olive electrolyte leakage characteristics at subzero temperatures $(-50 \degree \text{C} \text{ to } 0 \degree \text{C})$ for pre-, deep, and post-Dormancy stages.



Figure 4. Rate of change in olive EL at subzero temperatures $(-50 \degree \text{C} \text{ to } 0 \degree \text{C})$ during pre-, deep, and post-dormancy stages. Rate: EL changes per decrease in sub-zero temperature. In Section 2.1, we mentioned ten different olive varieties, which are represented by the acronyms CV1 to CV10.

3.2. Findings of the Triphenyl Tetrazolium Chloride (TZ) Assay3.2.1. Comparing and Selecting the Best-Fitted NLR Model for TZ

Apart from utilizing the EL measure of resistance against freezing injury, TZ was also employed. The prediction errors of all eighteen models mentioned in Table 1 were calculated for ten distinct olive cultivars across three stages of dormancy, i.e., pre-, deep, and post-dormancy. However, only the results of the TZ for eight models are presented in Table 4. After evaluating the RMSE values of all models, it was observed that they were generally acceptable and close to each other, with only slight differences. However, a closer examination of their shapes at lower temperatures, and the significance test of their regression coefficients, revealed that models M3, M4, M5, M6, and M7 were the strongest candidates for selection. Ultimately, the M4 model was chosen as the final model to accurately model the process of TZ changes at sub-zero temperatures. This decision was based on its ability to better illustrate the nature of the TZ problem and investigate the phenomenon of freezing injury. Unlocking the secrets of complex systems demands a comprehensive approach. That is why we chose a single model type to analyze all three stages across various olive varieties. This approach grants us the power to scrutinize regression coefficients in a more accurate and relevant way, leading us to uncover a wealth of insights.

Table 4. The RMSE values for estimating TZ of eight out of the eighteen NLR models that were investigated, across various olive cultivars.

		CV1			CV2			CV3			CV4			CV5	
	Pre	Deep	Post	Pre	Deep	Post	Pre	Deep	Post	Pre	Deep	Post	Pre	Deep	Post
M3	9.24	5.78	4.97	8.63	6.2	7.32	8.12	5.26	7.15	8.09	6.22	6.98	8.21	6.68	6.34
M4	8.8	6.21	5.65	7.84	5.16	6.27	8.34	6.22	7.86	7.22	5.2	5.93	7.33	5.72	5.6
M5	9.24	5.78	4.97	8.63	6.2	7.32	8.12	5.26	7.15	8.09	6.22	6.98	8.21	6.68	6.34
M6	8.8	6.21	5.65	7.84	5.16	6.27	8.34	6.22	7.86	7.22	5.2	5.93	7.33	5.72	5.6
M7	9.24	5.78	4.97	8.63	6.2	7.32	8.12	5.26	7.15	8.09	6.22	6.98	8.21	6.68	6.34
M10	8.8	6.21	5.65	7.84	5.16	6.27	8.34	6.22	7.86	7.22	5.2	5.93	7.33	5.72	5.6
M13	8.8	6.21	5.65	7.84	5.16	6.27	8.34	6.22	7.86	7.22	5.2	5.93	7.33	5.72	5.6
M14	8.8	5.75	4.95	7.83	5.16	6.21	8.09	5.15	7.13	7.22	5.2	5.93	7.27	5.73	5.57
		CV6			CV7			CV8			CV9			CV10	
	Pre	Deep	Post	Pre	Deep	Post	Pre	Deep	Post	Pre	Deep	Post	Pre	Deep	Post
M3	8.52	6.47	4.6	8.22	5.4	4	8.18	6.68	13.2	8.12	6.59	7.49	8.73	5.84	10.8
M4	7.69	5.35	4.53	8.52	6.67	4.83	7.23	5.69	15.33	7.11	5.53	7.45	8.93	7.03	10.76
M5	8.52	6.47	4.6	8.22	5.4	4	8.18	6.68	13.2	8.12	6.59	7.49	8.73	5.84	10.8
M6	7.69	5.35	4.53	8.52	6.55	4.83	7.23	5.69	15.33	7.11	5.53	7.45	8.93	7.03	10.76
M7	8.52	6.47	4.6	8.22	5.4	4	8.18	6.68	13.2	8.12	6.59	7.49	8.73	5.84	10.8
M10	7.69	5.35	4.53	8.52	6.55	4.83	7.23	5.69	15.33	7.11	5.53	7.45	8.93	7.03	10.76
M13	7.69	5.35	4.53	8.52	6.55	4.83	7.23	5.69	15.33	7.11	5.53	7.45	8.93	7.03	10.76
M14	7.55	5.35	4.38	8.2	5.4	3.99	7.23	5.69	11.33	7.11	5.53	7.11	8.7	5.44	10.8

3.2.2. Assessment of NLR Model Coefficients for TZ

Table 5 presents the statistical analysis results for the coefficients of the Gompertz nonlinear regression model (M4), including the coefficients of determination (R^2 and $R^2_{Adj,}$) for all tested cases of olives. Our analysis of the M4 model revealed that the *p*-values of its coefficient significance test were significant at both the 1% and 5% levels across all olive varieties. This finding underscores the robustness of our model and the validity of our results. Our analysis of the coefficients of determination and the standard deviation of the regression coefficients revealed a remarkable consistency across all cases. By achieving R^2 and R^2_{Adj} , values ranging from 92 to 98 percent, our study provides strong evidence for the robustness and precision of our approach.

			CV1			CV2	
		Pre	Deep	Post	Pre	Deep	Post
Cafe	b_1	$2.58 \ ^{**} \pm 0.37$	$8.24 ** \pm 1.59$	6.39 ** ± 1.00	2.68 ** ± 0.36	3.79 ** ± 0.40	$3.59 ** \pm 0.55$
Core.	b_2	$0.17~^{**}\pm0.02$	$0.20 ** \pm 0.02$	$0.20 ** \pm 0.01$	$0.19 \ ^{**} \pm 0.02$	$0.18~^{**}\pm0.01$	$0.22 ** \pm 0.02$
R^2 , $R^2_{Adj.}$		0.93, 0.92	0.97, 0.97	0.97, 0.97	0.94, 0.94	0.98, 0.98	0.97, 0.97
			CV3				
		Pre	Deep	Post	Pre	Deep	Post
Cofe.	b_1	2.98 ** ± 0.42	9.81 ** ± 2.09	$10.51 ** \pm 2.89$	2.63 ** ± 0.32	3.80 ** ± 0.40	3.81 ** ± 0.48
	b_2	$0.14 \ ^{**} \pm 0.01$	$0.19 \ ^{**} \pm 0.02$	$0.21 \ ^{**} \pm 0.02$	$0.18 \ ^{**} \pm 0.02$	$0.17~^{**}\pm 0.01$	$0.20 ** \pm 0.01$
R^2 , $R^2_{Adj.}$		0.93, 0.92	0.96, 0.96	0.95, 0.94	0.95, 0.95	0.98, 0.97	0.97, 0.97
			CV5			CV6	
		Pre	Deep	Post	Pre	Deep	Post
	b_1	2.80 ** ± 0.34	3.58 ** ± 0.40	3.84 ** ± 0.47	2.67 ** ± 0.36	3.34 ** ± 0.34	3.98 ** ± 0.40
Cofe.	b_2	$0.21 \ ^{**} \pm 0.02$	$0.17 \ ^{**} \pm 0.01$	$0.21 \ ^{**} \pm 0.01$	$0.20 \ ^{**} \pm 0.02$	$0.18 \ ^{**} \pm 0.01$	$0.22 ** \pm 0.01$
R^2 , $R^2_{Adj.}$		0.96, 0.96	0.97, 0.97	0.98, 0.97	0.95, 0.95	0.97, 0.97	0.98, 0.98
			CV7			CV8	
		Pre	Deep	Post	Pre	Deep	Post
0.1	b_1	3.38 ** ± 0.55	9.94 ** ± 2.03	5.87 ** ± 0.47	2.69 ** ± 0.34	3.66 ** ± 0.41	$7.24*\pm 2.81$
Cofe.	b_2	$0.18 \ ^{**} \pm 0.02$	$0.20 \ ^{**} \pm 0.02$	$0.19 \ ^{**} \pm 0.01$	$0.19 \ ^{**} \pm 0.02$	$0.17~^{**}\pm 0.01$	0.22 ** \pm 0.04
R^2 , $R^2_{Adj.}$		0.94, 0.93	0.97, 0.97	0.99, 0.99	0.95, 0.95	0.97, 0.97	0.88, 0.88
			CV9			CV10	
		Pre	Deep	Post	Pre	Deep	Post
Cali	b_1	2.69 ** ± 0.34	3.44 ** ± 0.37	4.38 ** ± 0.62	2.92 ** ± 0.43	9.98 ** ± 2.17	$16.07 * \pm 6.00$
Core.	b_2	$0.19 \ ^{**} \pm 0.02$	$0.17 \ ^{**} \pm 0.01$	$0.23 \ ^{**} \pm 0.02$	$0.14~^{**}\pm0.01$	$0.18~^{**}\pm0.02$	$0.25 \ ^{**} \pm 0.03$
R^2 , $R^2_{Adj.}$		0.95, 0.95	0.97, 0.97	0.98, 0.98	0.91, 0.91	0.96, 0.96	0.97, 0.96

Table 5. The statistical analysis outcomes of the coefficients for the M4 model that was utilized to calculate TZ using sub-zero temperatures for all experimental cases.

Means \pm SD. CV1 to CV10 are the olive cultivars, while Pre, Deep, and Post refer to the dormancy stages, b₁ and b₂ are model coefficients, R^2 is coefficient of determination, and $R^2_{Adj.}$ is adjusted coefficient of determination.

3.2.3. Conducting a Sensitivity Analysis of the TZ Model

The regression coefficients of the TZ model varied significantly among different olive cultivars, as presented in Table 5. Furthermore, the values of these coefficients also differed across the three stages of dormancy (i.e., pre-, deep, and post-dormancy) within each olive cultivar. These differences can be attributed to variations in the response of different olive cultivars to freezing injury during each stage of dormancy [30]. To gain a deeper understanding of this issue, we conducted a sensitivity analysis of the TZ model's predictions for sub-zero temperatures by examining the impact of changes in the b₁ and b₂ regression coefficients. As depicted in Figure 5a, an increase in the regression coefficient b_2 corresponds to an increase in the slope of TZ changes during temperature decrease. Specifically, when b_2 is set to 0.25, the maximum slope of changes is achieved, leading to an earlier and more pronounced decrease in TZ at lower temperatures. Conversely, when b_2 is set to 0.1 (the lowest value obtained from the experiments), TZ reaches a value of approximately 100% at a temperature of about -55 °C. These findings suggest that b₂ plays a crucial role in determining the rate and extent of TZ changes in response to sub-zero temperatures. Figure 5b demonstrates that increasing the value of the regression coefficient b_1 in the TZ model leads to a higher TZ value at the point of freezing. However, the effect of b_1 on the temperature at which TZ reaches 100% is negligible, as this value is consistently attained at -37 °C across all b₁ values. These findings suggest that a higher b₁ value corresponds to



greater resistance of olive trees to freezing injury, enabling them to better adapt to changing environmental conditions.

Figure 5. The impact of altering the regression coefficients b1 and b2 in the M4 model on the vulnerability of olives to damage caused by freezing: (**a**) changing the coefficient of model b2 with b1 constant at the average value, and (**b**) changing the coefficient of model b1 with b2 constant at the average value.

3.2.4. Comparison of TZ Modeling Outcomes across Various Olive Cultivars

Initially, the models were evaluated based on laboratory results, and the bestperforming model, M4, was selected. The Supporting Information section and Figure S2 present the lower and upper bounds, as well as the trend in changes predicted by the M4 model, alongside data from ten olive cultivars in pre-, deep, and post-dormancy stages. The results indicate that the M4 model estimates and laboratory data fall within the 95% prediction interval. Table 5 additionally confirms the validity of the model estimates within the tested temperature range. Notably, the M4 model is capable of predicting TZ for temperatures beyond the test range, as demonstrated in Figure 6, where TZ values were extrapolated for ten olive cultivars across three dormancy stages, extending to temperatures as low as -50 °C. This extrapolation allows for predictions regarding tree behavior outside the testing period, with the model's horizontal asymptotic line reaching the 100% point across various temperatures. The results depicted in Figure 6 demonstrate that, for all three dormancy stages, the trends in changes and values of TZ predictions are largely similar across all olive cultivars. However, there is a slight variation in the predicted values of TZ for pre-, deep, and post-dormancy stages, which create the highest to lowest values during temperature reduction. This may be attributed to the inherent nature of the olive tree, which exhibits resistance and adaptation to new conditions, including exposure to sub-zero temperatures [23].

In contrast to EL, which did not reach 100% even at temperatures below -50 °C (as shown in Figure 3), the value of TZ reached 100% at temperatures between -25 and -35 °C. Therefore, it can be inferred that the TZ criterion is more stringent in its judgments than the EL criterion. However, the value of the EL criterion varied in the range of 85 to 95 percent in the temperature range of -25 to -35 °C (as depicted in Figure 3).

Our findings indicate that olive cultivars with a higher amount of TZ during temperature drops in sub-zero conditions, and those that reach 100% TZ sooner, exhibit lower resistance to freezing injury. Based on these results, as well as the data presented in Figure 6 for the pre-dormancy stage, we can conclude that the olive tree cultivars CV6, CV5, CV9, CV8, CV2, CV4, CV1, CV7, CV3, and CV10 display a range of resistance and compatibility in sub-zero temperature conditions, with CV10 exhibiting the highest level of resistance and CV6 showing the lowest. In the deep dormancy stage, the olive cultivars can be ranked based on their adaptation or resistance to freezing injury, from the lowest to the highest, as follows: CV6, CV9, CV5, CV2, CV8, CV4, CV1, CV7, CV3, and CV10. After the dormancy period, we observed varying degrees of resistance to freezing injury among the olive cultivars. Based on our results, we sorted them in the following order: CV3, CV7, CV10, CV1, CV8, CV4, CV5, CV2, CV6, and CV9. Furthermore, our TZ modeling analysis revealed that CV3, CV7, and CV10 are the most compatible varieties in sub-zero temperature conditions. On the other hand, CV6, CV9, and CV5 showed the least compatibility with these conditions. Therefore, we suggest that CV10, CV3, and CV7 are the most suitable options for cultivation in areas with sub-zero temperatures, while CV6, CV9, and CV5 should be avoided.



Figure 6. Cont.



Figure 6. The modeling of olive TZ characteristics at subzero temperatures ranging from -50 °C to 0 °C for pre-, deep, and post-dormancy using nonlinear regression. In Section 2.1, we mentioned ten different olive varieties, which are represented by the acronyms CV1 to CV10.

3.2.5. Analysis of the Rate of Change in TZ Results

As previously mentioned, we utilized the first-order derivative of the TZ regression model to determine the rate of change in TZ production with respect to temperature. Figure 7 illustrates the TZ production track for ten olive cultivars across three dormancy stages: pre-, deep, and post-dormancy. Interestingly, we observed that the temperature point at which the TZ production rate changes is nearly constant across all three dormancy stages for each olive cultivar, in contrast to the EL results (Figure 4). The temperature range at which this phenomenon occurs varies from -5 °C to -12 °C. Additionally, our results indicate that the rate of TZ production reaches its minimum value and eventually reaches zero after the temperature drops below -35 °C. Furthermore, we found that the slope of the TZ production decrease and increase is nearly identical. Comparing the three dormancy stages within each variety, we noted that the temperature at which the TZ production rate changes occurs earlier during the pre-dormancy stage compared to the other two stages. This suggests that olive trees lose their resistance to frost damage earlier during the pre-dormancy stage according to TZ analysis.







Figure 7. Rate of changes in olive TZ during the stages of pre-, deep, and post-dormancy at temperatures below zero, ranging from -50 °C to 0 °C. Rate: TZ changes per decrease in sub-zero temperature. In Section 2.1, we mentioned ten different olive varieties, which are represented by the acronyms CV1 to CV10.

3.3. Assessment of T50 and T90 Values Based on the TZ and EL Models in Different Olive Varieties

Figure 8 displays the T50 and T90 temperatures calculated for ten olive cultivars in three dormant stages, using the NLR model of TZ and EL tests. T50 and T90 values provide important information about the cold tolerance of olive trees, indicating the temperatures at which a certain percentage of olive tree cells will be killed due to exposure to sub-zero temperatures. For example, during the pre-dormancy stage, the T50 and T90 values based on EL in CV1 were obtained at temperatures of -5.26 °C and -17.32 °C, respectively. This means that at a temperature of -5.26 °C, 50% of the maximum EL values were reached, and at a temperature of -17.32 °C, 90% of the maximum EL values were reached. The lowest T50 value based on EL tests in the pre-dormancy stage at -10.99 °C was observed in CV3, followed by CV10 at -10.58 °C. Similarly, the lowest T50 based on TZ tests was observed in CV10 at -10.59 °C, followed by CV3 at -10.31 °C. Based on EL tests, the lowest T90 values across all stages (pre-, deep, and post-dormancy) were observed in CV10 at -25.68 °C, -43.53 °C, and -38.28 °C, respectively. In the TZ test, CV10 had the lowest T90 value in the two stages before dormancy at -24.49 °C, while in the deep and post-dormancy stages, CV3 had the lowest T90 values at -24.74 °C and -21.64 °C, respectively. Higher absolute values of T50 and T90 indicate greater resistance to freezing injury and better

adaptability to sub-zero temperatures [14]. CV10, CV3, and CV7 demonstrated the highest performance across all three dormancy stages based on T50 values. Moreover, these three cultivars were also the best options across all three dormant stages based on T90 values. Hence, in terms of TZ and EL criteria, these three selections could be considered as suitable breeding options for temperate zones compared to the other seven varieties. The T50 and T90 values for different olive cultivars have been widely studied as important indicators of cold tolerance. These values can provide insights into the potential of different cultivars for withstanding freezing injury and can help in selecting more cold-tolerant cultivars for olive cultivation in regions with cold winters.



Figure 8. The T50 and T90 values based on TZ for ten different olive cultivars across three stages, namely pre-, deep, and post-dormancy stages.

4. Conclusions

This study evaluated the resistance and compatibility of ten olive cultivars to sub-zero temperatures and frost damage during three stages of dormancy using electrolyte leakage (EL) and tetrazolium tests (TZ) with the help of NLR analysis utilizing 18 different models. Eight models were selected based on significant regression coefficients, extrapolation results, trend in changes in model prediction, and R^2 and R^2_{Adj} criteria. The 2p-logistic

model and Gompertz model were selected for EL and TZ criteria, respectively. The NLR model allowed for the identification of the relationship between temperature and olive tree freezing injury, and the development of a predictive model for estimating the extent of tree damage and time to zero growth based on temperature data. The study identified the three cultivars CV10, CV7, and CV3 as having the best performance in sub-zero temperature conditions, with better resistance and adaptability to frost damage. However, it is important to note that the NLR model developed in this study is specific to the observed study conditions and may not be applicable to other locations or climates. Additionally, other factors such as wind speed, humidity, and soil moisture may also influence freezing injury in olive trees, which were not examined in this analysis. Overall, this research provides valuable information for olive growers and researchers to develop strategies to manage cold weather effects and prevent freezing injury in olive orchards.

Supplementary Materials: The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/agriculture13061137/s1, Figure S1: EL estimation results and prediction intervals (lower bound (LB) and upper bound (UB)) for temperature range of -21 °C to 0 °C; Figure S2: TZ estimation results and prediction intervals (lower bound (LB) and upper bound (UB)) for temperature range of -21 °C to 0 °C.

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References

- 1. Dahdouh, A.; Khay, I.; Le Brech, Y.; El Maakoul, A.; Bakhouya, M. Olive oil industry: A review of waste stream composition, environmental impacts, and energy valorization paths. *Environ. Sci. Pollut. Res.* **2023**, *30*, 45473–45497. [CrossRef] [PubMed]
- 2. Petruccelli, R.; Bartolini, G.; Ganino, T.; Zelasco, S.; Lombardo, L.; Perri, E.; Durante, M.; Bernardi, R. Cold Stress, Freezing Adaptation, Varietal Susceptibility of *Olea europaea* L.: A Review. *Plants* **2022**, *11*, 1367. [CrossRef]
- Fraga, H.; Moriondo, M.; Leolini, L.; Santos, J.A. Mediterranean olive orchards under climate change: A review of future impacts and adaptation strategies. *Agronomy* 2020, 11, 56. [CrossRef]
- 4. Bartolozzi, F.; Fontanazza, G. Assessment of frost tolerance in olive (Olea europaea L.). Sci. Hortic. 1999, 81, 309–319. [CrossRef]
- Cansev, A.; Gulen, H.; Eris, A. Cold-hardiness of olive (*Olea europaea* L.) cultivars in cold-acclimated and non-acclimated stages: Seasonal alteration of antioxidative enzymes and dehydrin-like proteins. *J. Agric. Sci.* 2009, 147, 51–61. [CrossRef]
- Lodolini, E.M.; Alfei, B.; Santinelli, A.; Cioccolanti, T.; Polverigiani, S.; Neri, D. Frost tolerance of 24 olive cultivars and subsequent vegetative re-sprouting as indication of recovery ability. *Sci. Hortic.* 2016, 211, 152–157. [CrossRef]
- López-Bernal, Á.; García-Tejera, O.; Testi, L.; Orgaz, F.; Villalobos, F.J. Studying and modelling winter dormancy in olive trees. *Agric. For. Meteorol.* 2020, 280, 107776. [CrossRef]
- 8. Yuldasheva, K.T.; Soliyeva, M.; Kimsanova, X.; Arabboev, A.; Kayumova, S. Evaluation of winter frost resistance of cultivated varieties of olives. *Acad. Int. Multidiscip. Res. J.* **2021**, *11*, 627–632. [CrossRef]
- 9. Gómez-del-Campo, M.; Barranco, D. Field evaluation of frost tolerance in 10 olive cultivars. *Plant Genet. Resour.* **2005**, *3*, 385–390. [CrossRef]
- 10. Lindén, L.; Palonen, P.; Lindén, M. Relating freeze-induced electrolyte leakage measurements to lethal temperature in red raspberry. J. Am. Soc. Hortic. Sci. 2000, 125, 429–435. [CrossRef]
- Murray, M.; Cape, J.; Fowler, D. Quantification of frost damage in plant tissues by rates of electrolyte leakage. *New Phytol.* 1989, 113, 307–311. [CrossRef] [PubMed]
- 12. Yu, D.J.; Lee, H.J. Evaluation of freezing injury in temperate fruit trees. Hortic. Environ. Biotechnol. 2020, 61, 787–794. [CrossRef]
- Whitlow, T.H.; Bassuk, N.L.; Ranney, T.G.; Reichert, D.L. An improved method for using electrolyte leakage to assess membrane competence in plant tissues. *Plant Physiol.* 1992, 98, 198–205. [CrossRef] [PubMed]

- 14. Karami, H.; Rezaei, M.; Sarkhosh, A.; Rahemi, M.; Jafari, M. Cold hardiness assessment in seven commercial fig cultivars (*Ficus carica L.*). *Gesunde Pflanz.* **2018**, *70*, 195–203. [CrossRef]
- 15. Saadati, S.; Baninasab, B.; Mobli, M.; Gholami, M. Measurements of freezing tolerance and their relationship with some biochemical and physiological parameters in seven olive cultivars. *Acta Physiol. Plant.* **2019**, *41*, 51. [CrossRef]
- Lim, C.C.; Arora, R.; Townsend, E.C. Comparing Gompertz and Richards functions to estimate freezing injury in Rhododendron using electrolyte leakage. J. Am. Soc. Hortic. Sci. 1998, 123, 246–252. [CrossRef]
- 17. Tudela, V.; Santibáñez, F. Modelling impact of freezing temperatures on reproductive organs of deciduous fruit trees. *Agric. For. Meteorol.* **2016**, 226–227, 28–36. [CrossRef]
- Villouta, C.; Workmaster, B.A.; Atucha, A. Freezing stress damage and growth viability in *Vaccinium macrocarpon* Ait. bud structures. *Physiol. Plant.* 2021, 172, 2238–2250. [CrossRef]
- 19. Archontoulis, S.V.; Miguez, F.E. Nonlinear regression models and applications in agricultural research. *Agron. J.* **2015**, *107*, 786–798. [CrossRef]
- Li, Q.; Gao, H.; Zhang, X.; Ni, J.; Mao, H. Describing Lettuce Growth Using Morphological Features Combined with Nonlinear Models. *Agronomy* 2022, 12, 860. [CrossRef]
- Jannatizadeh, A.; Rezaei, M.; Rohani, A.; Lawson, S.; Fatahi, R. Towards modeling growth of apricot fruit: Finding a proper growth model. *Hortic. Environ. Biotechnol.* 2022, 64, 209–222. [CrossRef]
- 22. Connor, D.J.; Fereres, E. The physiology of adaptation and yield expression in olive. Hortic. Rev. 2010, 31, 155–229.
- 23. Sanzani, S.M.; Schena, L.; Nigro, F.; Sergeeva, V.; Ippolito, A.; Salerno, M.G. Abiotic diseases of olive. J. Plant Pathol. 2012, 94, 469–491.
- Larcher, W. Temperature stress and survival ability of Mediterranean sclerophyllous plants. *Plant Biosyst.* 2000, 134, 279–295. [CrossRef]
- 25. Wisniewski, M.; Nassuth, A.; Arora, R. Cold Hardiness in Trees: A Mini-Review. Front. Plant Sci. 2018, 9, 1394. [CrossRef]
- Guerra, D.; Lamontanara, A.; Bagnaresi, P.; Orrù, L.; Rizza, F.; Zelasco, S.; Beghè, D.; Ganino, T.; Pagani, D.; Cattivelli, L. Transcriptome changes associated with cold acclimation in leaves of olive tree (*Olea europaea* L.). *Tree Genet. Genomes* 2015, 11, 113. [CrossRef]
- 27. Sakai, A.; Larcher, W. Frost Survival of Plants: Responses and Adaptation to Freezing Stress; Springer Science & Business Media: Berlin, Heidelberg, 2012; Volume 62.
- 28. Gusta, L.V.; Wisniewski, M. Understanding plant cold hardiness: An opinion. *Physiol. Plant.* 2013, 147, 4–14. [CrossRef] [PubMed]
- 29. Ashworth, E.; Wisniewski, M. Response of fruit tree tissues to freezing temperatures. HortScience 1991, 26, 501–504. [CrossRef]
- Rubio-Valdés, G.; Cabello, D.; Rapoport, H.F.; Rallo, L. Olive Bud Dormancy Release Dynamics and Validation of Using Cuttings to Determine Chilling Requirement. *Plants* 2022, *11*, 3461. [CrossRef] [PubMed]

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