



Review Rapeseed Meal and Its Application in Pig Diet: A Review

Hao Cheng ¹^(b), Xiang Liu ¹, Qingrui Xiao ¹, Fan Zhang ¹, Nian Liu ¹, Lizi Tang ¹, Jing Wang ¹, Xiaokang Ma ^{1,*}, Bie Tan ¹^(b), Jiashun Chen ¹ and Xianren Jiang ²

- ¹ College of Animal Science and Technology, Hunan Agricultural University, Changsha 410128, China; chenghao19970316@163.com (H.C.); liuxiang1138120110@163.com (X.L.); 15773772597@163.com (Q.X.); fzhang4056@163.com (F.Z.); ln18874045069@163.com (N.L.); t1523102483@163.com (L.T.); jingwang023@hunau.edu.cn (J.W.); bietan@hunau.edu.cn (B.T.); llh0920@163.com (J.C.)
- ² Key Laboratory of Feed Biotechnology of Ministry of Agriculture and Rural Affairs, Institute of Feed Research, Chinese Academy of Agricultural Sciences, Beijing 100081, China; jiangxianren@caas.cn
- Correspondence: maxiaokang@hunau.edu.cn

Abstract: Rapeseed is the second largest plant protein resource in the world with an ideal profile of essential amino acids. Rapeseed meal (RSM) is one of the by-products of rapeseed oil extraction. Due to the anti-nutritional components (glucosinolates and fiber) and poor palatability, RSM is limited in livestock diets. Recently, how to decrease the anti-nutritional factors and improve the nutritional value of RSM has become a hot topic. Therefore, the major components of RSM have been reviewed with emphasis on the methods to improve the nutritional value of RSM as well as the application of RSM in pig diets.

Keywords: rapeseed meal; pig; plant protein; livestock



Citation: Cheng, H.; Liu, X.; Xiao, Q.; Zhang, F.; Liu, N.; Tang, L.; Wang, J.; Ma, X.; Tan, B.; Chen, J.; et al. Rapeseed Meal and Its Application in Pig Diet: A Review. *Agriculture* **2022**, *12*, 849. https://doi.org/10.3390/ agriculture12060849

Academic Editor: Javier Álvarez-Rodríguez

Received: 21 April 2022 Accepted: 10 June 2022 Published: 12 June 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

1. Introduction

The imbalance between supply and demand of protein feed resources in developing countries has been increasing acutely. The feed production industry is facing a demand challenge. Therefore, the effective utilization of feed resources has attracted more attention. The United States Department of Agriculture states that rapeseed, one of the excellent plant protein resources, is the second-largest protein resource in the world with a global output of 73.09 million tons in 2017, which is only less than soybean [1,2]. Data from the National Bureau of Statistics shows that the yield of rapeseed in China has reached 14.05 million tons in 2020, which has become one of the major protein crops (Figure 1). The rapeseed oil is mainly utilized for human diet and the rapeseed meal (RSM) is a co-product which is commonly used as a protein source in animal diets after the oil has been extracted [3].

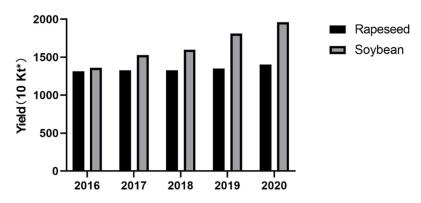


Figure 1. Soybean and rapeseed yield from 2016–2020 in China; Kt* = kiloton.

RSM is abundant in crude protein and has well-balanced essential amino acids, which is very close to the published values of soybean protein from the United Nations Food and Agriculture Organization [4]. However, the application of RSM in nutrition is limited because of the existence of anti-nutritional factors and toxic substances.

In recent years, the use of RSM in feeds for swine has increased with the development of low glucosinolate rapeseed by Canadian and European plant breeders [5]. Meanwhile, many technologies have been developed to improve the nutrient digestibility of RSM in growing pigs efficiently, including fermentation and hull-removing [6,7]. However, there are few systematic reviews of the application of RSM in pig diets. To maximize the benefits of RSM, it is vital to understand its nutritional characteristics and develop technologies to enhance its utilization cost-effectively [8]. Therefore, this review summarized the limitation of the utilization of RSM in animal diets from the nutritional value standpoint, and some improvement methods to enhance the quality of RSM.

2. Composition and Characteristics of Rapeseed Meal

2.1. Crude Protein and Amino Acids

As a raw material of protein feed, the content of crude protein (CP) in RSM is usually between 35% and 40% (Table 1). The protein content of RSM is affected by rapeseed type, growing environment, fiber content and so on. It has been found that the RSM produced from yellow-seeded Brassica juncea contained, on a dry matter basis, higher crude protein than that produced from traditional black-seeded Brassica napus canola and yellow-seeded Brassica juncea [9]. Earlier studies have demonstrated a sharp negative relationship between protein and dietary fiber content in meals [10] and crude protein content of RSM is significantly increased after the shell was removed [11].

	Rapeseed Meal	Double-Low Rapeseed Meal	Soybean Meal
Crude protein (%)	37.6~38.2	39.4~43.6	43.8~49.9
Arginine (g/kg)	20.6~22.1	20.8~24.1	34.9~37.8
Histidine (g/kg)	10.0~10.1	10.4~12.0	12.1~13.2
Isoleucine (g/kg)	14.6~15.3	13.8~15.6	21.5~27.8
Leucine (g/kg)	26.6~27.0	25.4~27.6	36.6~39.2
Lysine (g/kg)	17.2~19.5	19.4~24.1	29.9~32.2
Methionine (g/kg)	7.4~7.6	7.6~9.7	6.0~6.9
Phenylalanine (g/kg)	15.1~15.3	12.2~13.6	23.5~30.0
Threonine (g/kg)	17.5~17.6	17.6~19.1	18.9~20.3
Tryptophan (g/kg)	5.0~5.1	4.6~5.4	6.6~7.5
Valine (g/kg)	18.6~19.7	21.5~23.8	22.4~26.7
	Mosenthin et al. [12]	Li et al. [13]	Banaszkiexicz [14]

Table 1. Composition of crude protein and essential amino acids of rapeseed meal and soybean meal(as fed basis).

The protein from RSM is less digestible than that from soybean meal (SBM), while the amino acids balance of RSM is similar to that of SBM [15]. Compared with SBM, the concentration of lysine in RSM is lower, while the concentration of sulfur-containing amino acid in RSM is higher [16]. Therefore, it is better to mix RSM with SBM with a reasonable ratio when used for livestock diet due to the complementarity of amino acids content. Salazar-Villanea et al. found that toasting time has no influence on the CP content, but decreased the content of lysine, alanine, and glutamic acid and there was a significant correlation between the rate of protein hydrolysis and lysine [17]. It is worth of noting that heat treatment and other steps during processing may change the physical and chemical structure of protein while having no effects on the nitrogen content. Therefore, the content of available protein and amino acids in RSM can't be determined by the crude protein content while the content of lysine might be a better parameter to indicate the protein quality in RSM.

2.2. Crude Fat

RSM is the residue of rapeseed oil extracted by different processing techniques. The concentration of crude fat varied in dry matter of RSM from 1.42% to 14.55% with a mean of 7.78% [18]. The type of rapeseed, impurity content, processing technology, planting conditions in different producing areas, such as climate, and other factors can all affect the crude fat content in RSM [19]. Theodoridou and Yu found that the concentration of crude fat was higher from the brown-seeded Brassica napus canola press cake than from the yellow Brassica juncea and the browm-seeded Brassica napus canola meal [20]. During the drying, softening, rolling, steaming, and reserving processes, the value of the crude fat content of RSM as fed basis changes with the change of moisture content. Bojanowska reported that in comparison with fresh samples, significant differences in the crude fatcontent were observed in RSM after storage with increased saturated fatty acids and decreased unsaturated fatty acids [21]. However, as the utilization of RSM in swine diets is limited, relatively few studies are available on the crude fat content of RSM, and further research is needed.

2.3. Carbohydrate

The carbohydrates account for about one-third of the RSM and are mainly composed of monosaccharide, sucrose, oligosaccharide, starch, and NSP (non-starch polysaccharide). The content and structure of NSP in RSM vary with the type of rapeseed. Slominski and Campbell reported that the NSP content of Brassica campestris meal is between 16–22% while Pustjens et al. found the NSP content of Brassica napus meal is 24% [22,23]. In rapeseed, low-molecular-weight carbohydrates are mainly found in cotyledons while high-molecular-weight carbohydrates are mainly found in hulls [24]. Hulls contained 2.9% of the oil, 11.2% of protein, 73% of NDF (neutral detergent fiber), 80% of ADF (acidic detergent fiber) and 95% of ADL (lignin) and 6.0% of the glucosinolates of the whole rapeseed and RSM contained 48.3% protein (dry basis), 10.8% of NDF, 6.6% of ADF, and 0.5% of ADL after completely dehulled [25] The RSM has a limited utilization in swine diets due to the highly indigestible carbohydrates. Some studies have shown that rapeseed types and processing technologies affect the content of crude fiber in RSM, such as the dehulled RSM containing less crude fiber [19,25]. Along with the rapid development of alternative protein resources, it is necessary to develop some methods to improve the nutritional value of RSM.

2.4. Minerals and Vitamins

RSM is a relatively rich source of minerals including calcium, phosphorus, potassium, iron, zinc, and selenium which are important minerals for pigs [26]. Compared with SBM, RSM contains higher calcium and phosphorus contents [27]. In terms of vitamin composition, RSM contains a great amount of vitamin B such as biotin, folic acid, niacin, riboflavin and thiamin and vitamin E [28]. Moreover, RSM is rich in phenolic compounds and other bioactive compounds such as tocopherols and choline [29]. These studies above have shown that RSM has the potential to be a good protein feed resource.

2.5. Anti-Nutritional Factors

The nutritional value of RSM is close to that of SBM, but RSM is limited in practical appliances because it contains harmful substances and anti-nutritional factors, such as glucosinolates, phytic acid and fiber.

2.5.1. Glucosinolates

Glucosinolates are a large group of sulphur-containing secondary plant metabolites which are widely found in cruciferous plants and vegetables. The major forms of glucosinalates in RSM are gluconapin (3-butenyl), glucobrassicanapin (4-pentenyl), progoitrin (2-hydroxy-3-butenyl), gluconapoleiferin (2-hydroxy-4-pentenyl), and glucobrassicin (3-indolymethyl) [30–32]. Generally, the natural and non-hydrolyzed forms of glucosinolates are not harmful to animals or humans. But, glucosinolates will produce glucosinolate derivatives including nitriles, thiocyanates, isothiocyanates and 5-vinyloxazolidine-2-thione after being hydrolyzed which have been reported to interfere with iodine uptake by thyroid gland and induce goitrogenic effects in humans [33]. Moreover, gluconapin will affect the palatability of RSM due to its' pungent taste and the degradation product of progoitrin can be harmful to the health and function of thyroid [34,35]. Glucosinolates are the major anti-nutritional factor in RSM that reduces animal performance by affecting palatability of RSM and feed intake, impairing thyroid, liver, and kidney function and reducing feed utilization efficiency [36,37]. Velayudhan et al. demonstrated that replacing 33-66% of SBM with expeller extracted RSM (containing 1.90-2.78 µmol/g glucosinolates) in growing pig diets increased thyroid weight, changes in thyroid hormones, and linearly decreased the average daily feed intake [38]. Seneviratne et al. found that the tolerant level of glucosinolates in grower-finisher pigs was 1.70–3.40 μmol/g [39]. Whereas, Landero et al. reported that body weight gain, feed intake, and feed efficiency were reduced in weaned pigs even at the lowest dietary inclusion (6%) of juncea canola meal containing 0.65 μ mol/g glucosinolate in wheat-based diets, which indicates a high sensitivity of young pigs to glucosinolates [33]. Many studies have shown that as the early life production stressor, weanling not only changes structural and functional intestine, but also contributes to an intestinal inflammatory status which impairs gut barrier function [40–42]. We speculate that the fact young pigs are more sensitive to glucosinolates than older pigs may due to the immature gastrointestinal tract function [43].

2.5.2. Phytic Acid

Phytic acid (myoinositol hexaphosphoric acid) is the stored form of phosphorus in grains, legumes, nuts, and seeds [44]. Phytic acid is considered an anti-nutritional factor because it forms insoluble complexes by combining with proteins and several minerals (Zn, Ca, and Fe). This interaction may effect changes in protein structure and protein solubility which render them unavailable for intestinal absorption in humans and animals [45].

2.5.3. Tannin

Food tannins are polyphenolic compounds with molecular weights of 500–3000 Daltons, which can be divided into condensed or hydrolysable tannins and most of the tannins in rapeseed are condensed tannins [46]. Tannins can reduce the bioavailability of the nutrients due to the ability to form indigestible and bitter-tasting complexes with proteins [47].

3. Improvements to the Nutritive Values of Rapeseed Meal

In order to improve the overall nutritive value of RSM, enhance the utilization rate of RSM in pig diets and reduce the waste of protein resources, it is necessary to detoxify RSM and improve the processing techniques. At present, the nutritional value of RSM can be enhanced by optimizing processing conditions, reducing fiber content, and plant breeding strategy.

3.1. Optimize Processing Conditions

The processing technology of RSM has a great influence on the quality of RSM. Woyengo et al. reported that the expeller-extracted canola meal had higher digestible amino acid, digestible energy, and metabolic energy contents than the solvent-extracted canola meal [48]. Moreover, heat treatment in the processing also affects the nutritional value of RSM. For cold-press canola cake, application of heat to the barrel of the press during oil extraction increased the apparent ileal digestibility and apparent total tract digestibility of energy of cold-pressed canola cake [49]. While for hot-press, medium seed conditioning temperature resulted in the highest ileal digestible energy and apparent metabolic energy compared with low and high temperature [50]. It can be seen from the above that proper temperature adjustment during the pressing process can improve the nutritional value of RSM. The optimization of the process requires the joint efforts of different feed and oil enterprises to detect the nutritional value of oil and RSM, and then find the best control point of processing technology.

3.2. Reduce Fiber Content

In recent decades, various approaches including breeding for low-fiber and dehulling of seed have been undertaken to reduce the fiber content and improve the nutrient value of RSM [10,51].

Selective breading of rapeseed is not only aimed at reducing glucosinolate content but also decreasing fiber content [51]. Earlier research has demonstrated that the yellow-seed canola contains more protein and sucrose, less fiber and similar amounts of oligosaccharides and minerals compared with brown-seeded [52]. Slominski et al. also demonstrated that RSM derived from yellow-seeded Brassica napus canola contained more protein (49.8 vs. 43.8% DM), more sucrose (10.2 vs. 8.8% DM), and less total dietary fiber (24.1 vs. 30.1% DM) in comparison with conventional black-seeded counterpart [10].

Removing the hull of rapeseed before oil extraction is a good method to reduce fiber content. Kracht et al. studied the effects of removing the hull of rapeseed on the composition of RSM and rapeseed cake before the rapeseed is pressed and found that removing the hull of rapeseed significantly decreased the crude fiber content in RSM and rapeseed cake by approximately 40%, the neutral detergent fiber content by 28% and 39%, and the acid detergent fiber content by 35% and 39% in RSM and rapeseed cake, respectively [53]. Additionally, the hull of RSM could be removed by sieving particle size or by air classification according to density after production. It has been observed that the acid detergent fiber and the neutral detergent fiber were reduced by 31.9% and 29.5% in the light-particle fraction and were enriched by 16.5% and 9.0% in the heavy-particle fraction compared with parent canola meal [54]. Meanwhile, the air classification of canola meal increased its energy and acid amino digestibility in the light-particle fraction because of the reduced dietary fiber content. These results are in agreement with those reported by Hansen et al., who reported that sieving and air classification could remove the fiber from RSM and the reduced fiber content and increased crude protein content resulted in a higher digestibility of crude protein and amino acids [55].

Taken together, breeding for low-fiber rapeseed and removing the hull of rapeseed could reduce fiber content and enhance nutritive value, which have been revealed as effective methods to improve the nutrient value of RSM.

3.3. Biological Method to Improve Utilization of Rapeseed Meals

3.3.1. Enzymic Method

A relatively abundant literature has described the application of dietary enzymes (phytase, protease and carbohydrases) on RSM for the purpose of facilitating phosphorus, protein, and energy utilization for animal feed (Table 2).

Generally, phytases are known to improve phosphate and mineral uptake in animals which can't metabolize phytate [68]. Maison et al. reported that the addition of microbial phytase in growing pig diet improved digestibility of dietary phosphorus in RSM [58]. Potocka et al. found similar results, indicating that addition of phytase additives in growing and finishing pig diets could improve phosphorus and calcium digestibility [57]. Moreover, Rodrigues et al. reported that compared with a simple extraction process under similar conditions, pretreatment of RSM with phytase combined with alkaline extraction could enhance protein extraction yield [69]. It can be seen from the above that in addition of phytase can improve the utilization of RSM. However, the effect of adding phytase on growing pig diets is also different due to the difference of the source, dose and method of phytase application.

The use of non-starch polysaccharide enzymes, including xylanase, glucanase, cellulase, have been extensively studied in pig diet. It has been found that cellulase and alkaline feeding considerably changed the microbiome, as well as improved the overall degradation of RSM [70]. Li et al. also reported that multi-enzyme (cellulase, xylanase, glucanase, and protease) supplementation increased the crude protein digestion and all amino acids absorption and enhanced fiber degradation in double-low rapeseed co-products fed to pigs [60]. Similarly, Long and Venema also found that cellulose and pectinases could change microbial community composition, increase the abundance of microbial fiber-degrading enzymes and pathways, and increase acetic acid, propionic acid, butyric acid, and SCFA production [62]. In addition, non-starch polysaccharide enzymes could also reduce the viscosity of chyme, degrade plant cell walls and reduce the proliferation of pathogenic microorganisms in the intestine [71].

Method	Treatment	Animal	Main Results	Reference
Enzymic method	Aspergillus ficuum phytase	Weanling pig	Increase digestibilities of P and Ca and improve bone structure	Zhang et al. [56]
	Quantum Blue phytase	Growing and finishing pig	Improve P and Ca digestibility, and reduce P excretion	Małgorzata et al. [57] Maison et al. [58]
	Carbohydrases and phytase	Sow	Reduce the body weight loss and improve P digestibility post-farrowing	Velayudhan et al. [59]
	Cellulose, xylanase, glucanase and protease	Growing and finishing pig	Increased the standardized ileal digestibility of crude protein and all amino acids and enhanced fiber degradation Change microbial community and	Li et al. [60] Torres et al. [61]
	Cellulose and pectinase	Growing pig	increase the abundance of microbial fibre-degrading enzymes and pathways	Long and Venema [62]
	Cellulase, pectinase, amylase, protease and phytase	Gestating and lactating sow	Improve the standard ileal digestibility of amino acid	Velayudhan et al. [63]
Microbiological fermentation	Bacillus subtilis and Lactobacillus fermentum	Weaned piglet	Reduce the incidence of diarrhea and improve the gut microbiota	Czech et al. [64]
	Aspergillus niger 41258	Growing barrow	Increase P digestibility and digestible amino acid content and decrease P excretion	Shi et al. [65]
	Lactobacillus	Pregnant sow	Improve the structure and mechanical properties of compact bone in offspring	Tomaszewska et al. [66
	Lactobacillus, cellulose and pectinase	Growing pig	Increase reducing sugars and reduce the glucosinolate, total short-chain fatty acid and acetic acid content	Zhu et al. [67]

Table 2. Biological method application of rapeseed meal on pig.

3.3.2. Microbiological Fermentation

Microbial fermentation RSM refers to the use of complex microbial community and complex microbial enzyme system to ferment and decompose the toxic components and enzymatic hydrolysis products of RSM to achieve the purpose of detoxification. Microbial fermentation has an excellent detoxification effect on glucosinolates in RSM, which is the main anti-nutritional factor in RSM (Table 3). Zhang et al. found that under the fermentation of *Lactobacillus delbrueckii* and *Bacillus subtilis*, the content of glucosinolates in RSM decreased from 64.56 µmol/g to 3.47 µmol/g, and the degradation rate was high as 94.62% [72]. It has been reported that the soluble protein content, lactic acid content and total amino acid content in RSM increased significantly, whereas the glucosinolate content and neutral detergent fiber content decreased significantly after fermentation of *Bacillus licheniformis*, *Yeast* and *Lactobacillus* [73]. The lactic acid could contribute to the acidic taste and denature protein to reduce chewiness. Vig and Walia employed solid state fermentation of RSM and found that the contents of glucosinolates, thiooxazolidones, phytic acid and crude fibre declined by 43.1%, 34%, 42.4% and 25.5%, respectively, following inoculation with *Rhizopus oligosporus* [74].

Taken together, these studies have shown that microbial fermentation not only decreases the content of anti-nutritional material (glucosinolate, phytic acid, and crude fiber), but improves the palatability and the nutrient digestibility of RSM. Moreover, detoxified RSM by microbial fermentation has the advantages of obvious detoxification effect, mild condition, and the low expenture of process and add the basis for the increase in attention [79].

Table 3. Effects of microbiological fermentation on glucosinolates in rapeseed meal.

Source	Glucosinolates Content (μmol/g)	Treatment	Degradation Ratio	Reference
Canola meal	9.31	Aspergillus sojae and Aspergillus icuum	30%	Olukomaiya et al. [75]
Rapeseed meal	16.45	Aspergillus niger	43.07%	Shi et al. [7]
Rapeseed meal	23.79	Aspergillus niger	30.6%	Tie et al. [76]
Rapeseed press cake	32.1	Rhizupus	15.9%	Lucke et al. [77]
Rapeseed meal	64.6	Lactobacillus delbrueckii and Bacillus subtilis	94.62%	Zhang et al. [72]
Rapeseed meal	203.7	Bacillus subtilis and Actinomucor elegans	45.26%	Hao et al. [78]

4. Use of Dietary Rapeseed Meal in Pig Nutrition

4.1. Growth Performance and Meat Quality

Due to its high content and well-balanced essential amino acids, RSM is a good protein resource of diets for monogastric animals. Recent research has shown that a small amount of RSM could be used in swine diets without detrimental effects on growth performance and meat quality (Table 4). Do et al. demonstrated that a diet containing 8% RSM has no adverse effects on the growth performance of weaning pigs, which is similar to those reported by Shi et al., who found that adding 10% RSM in the diet has no adverse effects on production performance of finishing pig [7,80]. It has been reported that feeding pigs with diets containing RSM doesn't affect the pork meat quality, but it may decrease the weight gain [81,82]. But, Grabez et al. reported addition of RSM to finishing pig diets increased feed conversation ratio, improved meat coloring, increased sweet tasting metabolites and improved the flavor attributes of meat [83]. These differences in these results might be explained by the different RSM breeds and processing treatments, as well as the different breeds and life-stages of experimental pigs.

Animal	Source	Results	References
Weanling pig	Rapeseed meal	No adverse effects on the growth performance with up to 8% rapeseed meal	Do et al. [80]
Weaned pig	<i>Brassica napus</i> and <i>Brassica</i> <i>juncea</i> canola meal	No difference in feed intake, BWG and FCR	Landero et al. [84]
Growing pig	Rapeseed meal fermented by Aspergillus niger	No adverse effects on performance, when replaced with rapeseed meal up to 10%	Shi et al. [7]
Growing pig	Canola/double low rapeseed meal/expeller	No difference in growth performance with rapeseed meal up to 5%	Hansen et al. [85]
Growing-finishing pig	Rapeseed meal	No adverse effects on performance, when rapeseed meal was provided up to 9%	Choi et al. [81]
Finishing pig	Extracted rapeseed meal and legume plant	No adverse effects on pork quality; Reduce fatness; daily BWG↓	Zmudzinska et al. [82]
Growing-finishing pig	Comercial expeller pressed rapeseed	ADG↓; generally no difference in meat quality with rapeseed meal up to 20%	Skugor et al. [86]
Growing-finishing pig	Rapeseed meal	FCR↑; glucose level, lightness and yellowness of meat↓; oxidative stress↓; free amino acids, sweet tasting metabolites and flavor attributes↑	Grabez et al. [83]
Growing-finishing pig	Rapeseed meal	FCR↑; total MUFA↑, SFA and PUFA↓ in the steak cuts↓; modified the microbial balance in the digestive tract	Skoufos et al. [87]

Table 4. The effects of rapeseed meal on growth performance and meat quality of pig.

ADG = average daily gain; BWG = body weight gain; FCR = feed conversation ratio; MUFA = monounsaturated fatty acid; SFA = saturated fatty acid; PUFA = polyunsaturated fatty acid.

4.2. Reproduction of Sows

Pregnant sows are usually subject to strict feeding restrictions, which unfortunately cause various problems such as constipation and malnutrition [88]. The reproductive performance of sows fed diets supplemented with RSM are included in Table 5. Grela et al. found that the addition of 4-9% share of a fermented RSM component in gestation and lactation sow diet, respectively, improved the production parameters (litter size and litter weight), changed the microbiological composition of the digestive tract contents in pregnant gilts and reduce the severity of diarrhoea and mortality of the offspring [89]. This may be due to the fact that the fermentation process enriches the diet with enzymes, vitamins and short-chain fatty acids, thereby stimulating the gut environments of pig. Meanwhile, RSM contains crude fiber and adding an appropriate amount of fiber to the diet can prevent constipation and increase satiety of the sow. Quiniou et al. reported that sows fed diets with 10% RSM over three reproductive cycles, piglet weight at birth and litter weight gain were not affected [90]. Nevertheless, researchers also found that the addition of 4–9% fermented RSM in pig diet could increase litter size and weight, simulate the immune system (increase LYM counts and IgG titres in the blood plasma), and improve the nutrient digestibility [89,91,92]. These enhancements in pigs may be attributed to fermentation, which could reduce the content of anti-nutrients, improve the gut bacteria structure and promote the nutrients absorption of sows. Taken together, RSM has the potential to improve the reproductive performance of sows, but the processing treatment and applied RSM level in feed need to maintain considerable flexibility.

Table 5. The effects of rapeseed meal on reproduction performance of sow.

Sources	Appending Proportion (%)	Main Results	References
Rapeseed meal	10%	Not affect piglet weight at birth or weaning, survival and litter weight gain	Quiniou et al. [90]
Rapeseed meal	12%	No detrimental effects on reproductive performance and growth their progeny	Park et al. [93]
Rapeseed meal	6%	No detrimental effects on growth and production Gut lactic acid bacteria [†] ; sow body weight and	Bowland and Hardin [94]
Canola meal	30%	plasma urea nitrogen↓; No adverse effects on milk composition and nutrient digestibility	Velayudhan et al. [59]
Rapeseed press cake	8~14%	Body weight of piglet↑; piglet growth rate↑;	Hanczakowska et al. [95]
Fermented rapeseed meal	4~9%	Stimulate immune and antioxidant system;	Czech et al. [92]
Fermented rapeseed meal	4~9%	Litter size and litter weight↑; nutrient digestibility↑; maleficent bacteria↓	Grela et al. [89]
Fermented rapeseed meal	4~9%	Plasma content of Ht, Hb, RBC and mineral↑; plasma content of total cholesterol and triacylglycerols↓; liver enzyme activity↓	Czech et al. [91]

Ht = haematocrit; Hb = haemoglobin; RBC = erythrocyte count.

5. Conclusions and Future Perspective

Although considerable progress (heat treatment, microbial fermentation, and enzymolysis) has been taken in recent years, some questions still remain unclear.

It is difficult to improve the effectiveness of fermentation such as finding the most suitable temperature and time and optimum ratio of bacteria/enzyme.

Modern methods for processing RSM are not yet perfect.

The metabolic characteristics of probiotics in the fermentation process need more exploration. To better answer these practical questions, more studies are still needed to be done to fully utilize RSM in swine diets. **Author Contributions:** H.C., X.L., Q.X., L.T. and N.L.: Literature collection, H.C. and F.Z. Writing-Original draft preparation. X.J., J.C. and J.W.: Writing—Reviewing and Editing. B.T. and X.M.: Funding acquisition. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by the National Key R&D Program of China (2021YFD1301004), the Hunan Provincial Natural Science Foundation of China (2021JJ30318), the "Open Project Program of Key Laboratory of Feed Biotechnology, the Ministry of Agriculture and Rural Affairs of the People's Republic of China", and the China Agriculture Research System of MOF and MARA, Earmarked Fund for China Agriculture Research System (CARS).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: We declare that there is no conflict of interest.

References

- 1. United States Department of Agriculture. *Oilseeds: World Markets and Trade;* United States Department of Agriculture: Washington, DC, USA, 2021; pp. 1–38.
- Wickramasuriya, S.S.; Yi, Y.J.; Yoo, J.; Kang, N.K.; Heo, J.M. A review of canola meal as an alternative feed ingredient for ducks. J. Anim. Sci. Technol. 2015, 57, 29. [CrossRef] [PubMed]
- Xiao, Z.L.; Pan, Y.Y.; Wang, C.; Li, X.C.; Lu, Y.Q.; Tian, Z.; Kuang, L.Q.; Wang, X.F.; Dun, X.L.; Wang, H.Z. Multi-functional development and utilization of rapeseed: Comprehensive analysis of the nutritional value of rapeseed sprouts. *Food* 2022, *11*, 778. [CrossRef] [PubMed]
- 4. National Research Council. *Nutrient Requirements of Swine*; National Research Council: Washington, DC, USA, 2012.
- 5. Wang, L.; Hu, Q.; Li, P.; Lai, C.; Li, D.; Zang, J.; Ni, S. Development and validation of equations for predicting the metabolizable energy value of double-low rapeseed cake for growing pigs. *Animals* **2021**, *11*, 1168. [CrossRef] [PubMed]
- 6. Wang, Y.; Sun, H.; Han, B.; Li, H.Y.; Liu, X.L. Improvement of nutritional value, molecular weight patterns (soluble peptides), free amino acid patterns, total phenolics and antioxidant activity of fermented extrusion pretreatment rapeseed meal with Bacillus subtilis YY-1 and Saccharomyces cerevisiae YY-2. *LWT* **2022**, *160*, 113280.
- Shi, C.; He, J.; Wang, J.; Yu, J.; Yu, B.; Mao, X.; Zheng, P.; Huang, Z.; Chen, D. Effects of Aspergillus niger fermented rapeseed meal on nutrient digestibility, growth performance and serum parameters in growing pigs. *Anim. Sci. J.* 2016, 87, 557–563. [CrossRef]
- 8. Oliveira, A.M.; Yu, P. Research progress and future study on physicochemical, nutritional, and structural characteristics of canola and rapeseed feedstocks and co-products from bio-oil processing and nutrient modeling evaluation methods. *Crit. Rev. Food Sci. Nutri.* **2022**, 1–7. [CrossRef]
- Radfar, M.; Rogiewicz, A.; Slominski, B.A. Chemical composition and nutritive value of canola-quality Brassica juncea meal for poultry and the effect of enzyme supplementation. *Anim. Feed Sci. Tech.* 2017, 225, 97–108. [CrossRef]
- 10. Slominski, B.A.; Jia, W.; Rogiewicz, A.; Nyachoti, C.M.; Hickling, D. Low-fiber canola. Part 1. Chemical and nutritive composition of the meal. *J. Agric. Food Chem.* **2012**, *60*, 12225–12230.
- 11. Mejicanos, G.A. Tail End Dehulling of Canola Meal Chemical Composition and Nutritive Value of Dehulled Meal for Brolier Chicken and Weaned Pigs; University of Manitoba: Winnipeg, MB, Canada, 2015.
- 12. Mosenthin, R.; Messerschmidt, U.; Sauer, N.; Carre, P.; Quinsac, A.; Schone, F. Effect of the desolventizing/toasting process on chemical composition and protein quality of rapeseed meal. *J. Anim. Sci. Biotechnol.* **2016**, *7*, 205–216.
- 13. Li, P.; Wang, F.; Wu, F.; Wang, J.; Liu, L.; Lai, C. Chemical composition, energy and amino acid digestibility in double-low rapeseed meal fed to growing pigs. *J. Anim. Sci. Biotechnol.* **2015**, *6*, 1–10.
- 14. Banaszkiexicz, T.; Nutritional Value of Soybean Meal. Soybean and Nutrition. 2011, pp. 1–20. Available online: www.intechopen. com/books/nutritional-value-of-soybean-meal (accessed on 20 April 2022).
- 15. Nega, T. Review on nutritional limitations and opportunities of using rapeseed meal and other rape seed byproducts in animal feeding. *J. Nutr. Health Food Engl.* **2018**, *8*, 43–48.
- 16. Khajali, F.; Slominski, B.A. Factors that affect the nutritive value of canola meal for poultry. *Poult. Sci.* **2012**, *91*, 2564–2575. [CrossRef] [PubMed]
- 17. Salazar-Villanea, S.; Bruininx, E.M.; Gruppen, H.; Hendriks, W.H.; Carre, P.; Quinsac, A.; van der Poel, A.F. Physical and chemical changes of rapeseed meal proteins during toasting and their effects on in vitro digestibility. *J. Anim. Sci. Biotechnol.* **2016**, *7*, 62. [CrossRef] [PubMed]
- Zhang, Z.Y.; Li, P.L.; Liu, L.; Zhang, S.; Li, J.T.; Zhang, L.X.; Li, D.F. Ether extract and acid detergent fibre but not glucosinolates are determinants of the digestible and metabolizable energy of rapeseed meal in growing pigs. *J. Appl. Anim. Res.* 2020, 48, 384–389. [CrossRef]
- 19. Adewole, D.I.; Rogiewicz, A.; Dyck, B.; Slominski, B.A. Chemical and nutritive characteristics of canola meal from Canadian processing facilities. *Anim. Feed Sci. Technol.* **2016**, *222*, 17–30. [CrossRef]

- Theodoridou, K.; Yu, P. Effect of processing conditions on the nutritive value of canola meal and presscake. Comparison of the yellow and brown-seeded canola meal with the brown-seeded canola presscake. J. Sci. Food Agric. 2013, 93, 1986–1995. [CrossRef]
- Bojanowska, M. Changes in chemical composition of rapeseed meal during storage, influencing nutritional value of its protein and lipid fractions. *J. Anim. Feed Sci.* 2017, 26, 157–164. [CrossRef]
- 22. Slominski, B.A.; Campbell, L.D. Non-starch Polysaccharides of Canola Meal: Qulification, Digestibility in Poutry and Potential Benefit of Dietary Enzyme Supplementation. *J. Sci. Food Agric.* **1990**, *53*, 175–184. [CrossRef]
- 23. Pustjens, A.M.; Vries, S.D.; Schols, H.A.; Gruppen, H.; Gerrits, W.J.; Kabel, M.A. Understanding carbohydrate structures fermented or resistant to fermentation in broilers fed rapeseed (*Brassica napus*) meal to evaluate the effect of acid treatment and enzyme addition. *Poult. Sci.* 2014, 93, 926–934. [CrossRef]
- 24. Chmielewska, A.; Kozlowska, M.; Rachwal, D.; Wnukowski, P.; Amarowicz, R.; Nebesny, E.; Rosicka-Kaczmarek, J. Canola/rapeseed protein—nutritional value, functionality and food application: A review. *Crit. Rev. Food Sci. Nutr.* 2020, *61*, 3836–3856. [CrossRef]
- 25. Carre, P.; Citeau, M.; Robin, G.; Estorges, M. Hull content and chemical composition of whole seeds, hulls and germs in cultivars of rapeseed (*Brassica napus*). OCL 2016, 23, A302. [CrossRef]
- 26. Beyzi, E.; Gunes, A.; Beyzi, S.B.; Konca, Y. Changes in fatty acid and mineral composition of rapeseed (*Brassica napus* ssp. *oleifera* L.) oil with seed sizes. *Ind. Crops Prod.* **2019**, 129, 10–14. [CrossRef]
- 27. Summer, J.D.; Bedford, M.; Spratt, D. Amino acid supplementation of canola meal. Can. J. Anim. Sci. 1989, 69, 469–475. [CrossRef]
- Szydlowska-Czerniak, A. Rapeseed and its products-sources of bioactive compounds: A review of their characteristics and analysis. Crit. Rev. Food Sci. Nutr. 2013, 53, 307–330. [CrossRef]
- 29. Vuorela, S.; Meyer, A.S.; Heinonen, M. Impact of isolation method on the antioxidant activity of rapeseed meal phenolics. *J. Agric. Food Chem.* **2004**, *52*, 8202–8207. [CrossRef]
- 30. Chen, S.; Andreasson, E. Update on glucosinolate metabolism and transport. Plant Physiol. Biochem. 2001, 39, 743–758. [CrossRef]
- Konkol, D.; Szmigiel, I.; Domzal-Kedzia, M.; Kulazynski, M.; Krasowska, A.; Opalinski, S.; Korczynski, M.; Lukaszewicz, M. Biotransformation of rapeseed meal leading to production of polymers, biosurfactants, and fodder. *Bioorganic Chem.* 2019, 93, 102865. [CrossRef]
- 32. Lee, J.W.; Woyengo, T.A. Growth performance, organ weights, and blood parameters of nursery pigs fed diets containing increasing levels of cold-pressed canola cake. *J. Anim. Sci.* **2018**, *96*, 4704–4712. [CrossRef]
- Goyal, A.; Tanwar, B.; Sihag, M.K.; Kumar, V.; Sharma, V.; Soni, S. Rapeseed Canola (Brassica napus) Seed. In Oilseeds: Health Attributes and Food Applications; Springer: Singapore, 2021; pp. 56–57.
- 34. Landero, J.L.; Beltranena, E.; Zijlstra, R.T. Diet nutrient digestibility and growth performance of weaned pigs fed solvent-extracted Brassica juncea canola meal. *Anim. Feed Sci. Technol.* **2013**, *180*, 64–72. [CrossRef]
- 35. Zhou, Q.; Tang, H.; Jia, X.; Zheng, C.; Huang, F.H.; Zhang, M. Distribution of glucosinolate and pungent odors in rapeseed oils from raw and microwaved seeds. *Int. J. Food Prop.* **2018**, *21*, 2296–2308. [CrossRef]
- Lee, J.W.; Wang, S.; Huang, Y.; Seefeldt, T.; Donkor, A.; Logue, B.A.; Woyengo, T.A. Toxicity of canola-derived glucosinolates in pigs fed resistant starch-based diets. J. Anim. Sci. 2020, 98, 1–10. [CrossRef]
- 37. Tripathi, M.K.; Mishra, A.S. Glucosinolates in animal nutrition A review. Anim. Feed Sci. Technol. 2007, 132, skaa111. [CrossRef]
- Velayudhan, D.E.; Schuh, K.; Woyengo, T.A.; Sands, J.S.; Nyachoti, C.M. Effect of expeller extracted canola meal on growth performance, organ weights, and blood parameters of growing pigs. J. Anim. Sci. 2017, 95, 302–307. [CrossRef] [PubMed]
- Seneviratne, R.W.; Young, M.G.; Beltranena, E.; Goonewardene, L.A.; Newkirk, R.W.; Zijlstra, R.T. The nutritional value of expeller-pressed canola meal for grower-finisher pigs. J. Anim. Sci. 2010, 88, 2073–2083. [CrossRef]
- 40. Gresse, R.; Chaucheyras-Durand, F.; Fleury, M.A.; Van de Wiele, T.; Forano, E.; Blanquet-Diot, S. Gut Microbiota dysbiosis in postweaning piglets: Understanding the keys to health. *Trends Microbiol.* **2017**, *25*, 851–873. [CrossRef]
- 41. Moeser, A.J.; Pohl, C.S.; Rajput, M. Weaning stress and gastrointestinal barrier development: Implications for lifelong gut health in pigs. *Anim. Nutr.* **2017**, *3*, 313–321. [CrossRef] [PubMed]
- 42. Pluske, J.R.; Turpin, D.L.; Kim, J.C. Gastrointestinal tract (gut) health in the young pig. *Anim. Nutr.* 2018, 4, 187–196. [CrossRef] [PubMed]
- 43. Zhao, W.J.; Wang, Y.P.; Liu, S.Y.; Huang, J.J.; Zhai, Z.X.; He, C.; Ding, J.M.; Wang, J.; Wang, H.J.; Fan, W.B.; et al. The dynamic distribution of porcine microbiota across different ageds and gastrointestinal tract segments. *PLoS ONE* **2015**, *10*, e0117441.
- 44. Jacela, J.Y.; Derouchey, J.M.; Tokach, M.D.; Goodband, R.D.; Nelssen, J.L. Feed additives for swine: Fact sheets—flavors and mold inhibitors, mycotoxin binders, and antioxidants. *J. Swine Health Prod.* **2010**, *18*, 27–32. [CrossRef]
- 45. Nissar, J.; Ahad, T.; Naik, H.R.; Hussain, S.Z. A review phytic acid as antinutrient or nutraceutical. *J. Pharmacogn. Phytochem.* **2017**, *6*, 1554–1560.
- Hashmi, S.I.; Satwadhar, P.N.; Khotpal, R.R.; Deshpande, H.W.; Syed, K.A.; Vibhute, B.P. Rapeseed meal nutraceuticals. J. Oilseed Brassica 2010, 1, 43–54.
- 47. Tanwar, B.; Modgil, R.; Goyal, A. Antinutritional factors and hypocholesterolemic effect of wild apricot kernel (*Prunus armeniaca* L.) as affected by detoxification. *Food Funct.* **2018**, *9*, 2121–2135. [CrossRef] [PubMed]
- Woyengo, T.A.; Kiarie, E.; Nyachoti, C.M. Energy and amino acid utilization in expeller-extracted canola meal fed to growing pigs. J. Anim. Sci. 2010, 88, 1433–1441. [CrossRef] [PubMed]

- Seneviratne, R.W.; Beltranena, E.; Newkirk, R.W.; Goonewardene, L.A.; Zijlstra, R.T. Processing conditions affect nutrient digestibility of cold-pressed canola cake for grower pigs. J. Anim. Sci. 2011, 89, 2452–2461. [CrossRef]
- 50. Toghyani, M.; Rodgers, N.; Barekatain, M.R.; Iji, P.A.; Swick, R.A. Apparent metabolizable energy value of expeller-extracted canola meal subjected to different processing conditions for growing broiler chickens. *Poult. Sci.* **2014**, *93*, 2227–2236. [CrossRef]
- 51. Ton, L.B.; Neik, T.X.; Batley, J. The Use of Genetic and Gene Technologies in Shaping Modern Rapeseed Cultivars (*Brassica napus* L.). *Genes* **2020**, *11*, 1116. [CrossRef]
- 52. Slominski, B.A.; Simbaya, J.; Campbell, L.D.; Rakow, G.; Guenter, W. Nutritive value for broilers of meals derived from newly. *Anim. Feed Sci. Technol.* **1999**, *79*, 249–262. [CrossRef]
- 53. Kracht, W.; Danicke, S.; Kluge, H.; Keller, K.; Matzke, W.; Hennig, U.; Schumann, W. Effect of dehulling of rapeseed on feed value and nutrient digestibility of rape products in pigs. *Arch. Anim. Nutr.* **2004**, *58*, 389–404. [CrossRef]
- Zhou, X.; Zijlstra, R.T.; Beltranena, E. Nutrient digestibility of solvent-extracted *Brassica napus* and Brassica juncea. *J. Anim. Sci.* 2015, 93, 217–228. [CrossRef]
- 55. Hansen, J.O.; Skrede, A.; Mydland, L.T.; Overland, M. Fractionation of rapeseed meal by milling, sieving and air classificationeffect on crude protein, amino acids and fiber content and digestibility. *Ani. Feed Sci. Technol.* **2017**, 230, 143–153. [CrossRef]
- Zhang, Z.B.; Kornegay, E.T.; Radcliffe, J.S.; Wilson, J.H.; Veit, H.P. Comparison of phytase from genetically engineered Aspergillus and canola in weanling pig diets. J. Anim. Sci. 2000, 78, 2868–2878. [CrossRef] [PubMed]
- 57. Kasprowicz-Potocka, M.; Zaworska-Zakrzewska, A.; Rutkowski, A. Effect of Phytase on Digestibility and Performance of Growing and Finishing Pigs Fed Diets with Lupins and Rapeseed Meal. J. Agric. Sci. Technol. A 2020, 10, 216–227. [CrossRef]
- Maison, T.; Liu, Y.; Stein, H.H. Apparent and standardized total tract digestibility by growing pigs of phosphorus in canola meal from North America and 00-rapeseed meal and 00-rapeseed expellers from Europe without and with microbial phytase. *J. Anim. Sci.* 2015, *93*, 3494–3502. [CrossRef]
- Velayudhan, D.E.; Hossain, M.M.; Regassa, A.; Nyachoti, C.M. Effect of canola meal inclusion as a major protein source in gestation and lactation sow diets with or without enzymes on reproductive performance, milk composition, fecal bacterial profile and nutrient digestibility. *Anim. Feed Sci. Technol.* 2018, 241, 141–150. [CrossRef]
- 60. Li, P.; Lyu, Z.Q.; Wang, L.; Huang, B.B.; Lai, C.H.; Miglior, F. Nutritive values of double-low rapeseed expellers and rapeseed meal with or without supplementation of multi-enzyme in pigs. *Can. J. Anim. Sci.* **2020**, *100*, 729–738. [CrossRef]
- 61. Torres-Pitarch, A.; McCormack, U.M.; Beattie, V.E.; Magowan, E.; Gardiner, G.E.; Pérez-Vendrell, A.M.; Torrallardona, D.; O'Doherty, J.V.; Lawlor, P.G. Effect of phytase, carbohydrase, and protease addition to a wheat distillers dried grains with solubles and rapeseed based diet on in vitro ileal digestibility, growth, and bone mineral density of grower-finisher pigs. *Livest. Sci.* 2018, 216, 94–99. [CrossRef]
- 62. Long, C.; Venema, K. Pretreatment of Rapeseed Meal Increases Its Recalcitrant Fiber Fermentation and Alters the Microbial Community in an in vitro Model of Swine Large Intestine. *Front. Microbiol.* **2020**, *11*, 58826. [CrossRef]
- 63. Velayudhan, D.E.; Hossain, M.M.; Stein, H.H.; Nyachoti, C.M. Standardized ileal digestibility of amino acids in canola meal fed to gestating and lactating sows1. *J. Anim. Sci.* 2019, *97*, 4219–4226. [CrossRef]
- 64. Czech, A.; Grela, E.R.; Kiesz, M. Dietary fermented rapeseed or/and soybean meal additives on performance and intestinal health of piglets. *Sci. Rep.* **2021**, *11*, 16952. [CrossRef]
- 65. Shi, C.; He, J.; Yu, J.; Yu, B.; Mao, X.; Zheng, P.; Huang, Z.; Chen, D. Amino acid, phosphorus, and energy digestibility of Aspergillus niger fermented rapeseed meal fed to growing pigs. *J. Anim. Sci.* **2015**, *93*, 2916–2925. [CrossRef]
- 66. Tomaszewska, E.; Muszyński, S.; Dobrowolski, P.; Kamiński, D.; Czech, A.; Grela, E.R.; Wiącek, D.; Tomczyk-Warunek, A. Dried fermented post-extraction rapeseed meal given to sows as an alternative protein source for soybean meal during pregnancy improves bone development of their offspring. *Livest. Sci.* 2019, 224, 60–68. [CrossRef]
- 67. Zhu, X.F.; Wang, L.Y.; Zhang, Z.; Ding, L.R.; Hang, S.Q. Combination of fiber-degrading enzymatic hydrolysis and lactobacilli fermentation enhances utilization of fiber and protein in rapeseed meal as revealed in simulated pig digestion and fermentation in vitro. *Anim. Feed Sci. Technol.* **2021**, *278*, 115001. [CrossRef]
- Rao, D.E.; Rao, K.V.; Reddy, T.P.; Reddy, V.D. Molecular characterization, physicochemical properties, known and potential applications of phytases: An overview. *Crit. Rev. Biotechnol.* 2009, 29, 182–198. [CrossRef] [PubMed]
- Rodrigues, I.M.; Carvalho, M.G.V.; Rocha, J.M. Increase of protein extraction yield from rapeseed meal through a pretreatment with phytase. J. Sci. Food Agric. 2017, 97, 2641–2646. [CrossRef]
- 70. Cheng, L.; Rosch, C.; Vries, S.D.; Schols, H.; Venema, K. Cellulase and alkaline treatment improve intestinal microbial degradation of recalcitrant fibers of rapeseed meal in pigs. J. Agric. Food Chem. 2020, 68, 11011–11025.
- 71. Sethy, K.; Mishra, S.K.; Mohanty, P.P.; Agarawal, J.; Meher, P.; Satapathy, D.; Satapathy, D.; Sahoo, J.K.; Panda, S.; Nayak, S.M. An Overview of Non-Starch Polysaccharide. *J. Anim. Nutr. Physiol.* **2015**, *1*, 17–22.
- 72. Zhang, Z.Y.; Wen, M.; Chang, Y.Q. Degradation of glucosinolates in rapeseed meal by Lactobacillus delbrueckii and Bacillus subtilis. *Grain Oil Sci. Technol.* **2020**, *3*, 70–76. [CrossRef]
- 73. Wang, Y.; Liu, J.; Wei, F.H.; Liu, X.L.; Yi, C.X.; Zhang, Y.G. Improvement of the nutritional value, sensory properties and bioavailability of rapeseed meal fermented with mixed microorganisms. *LWT* **2019**, *112*, 1082338. [CrossRef]
- 74. Vig, A.P.; Walia, A. Beneficial effects of Rhizopus oligosporus fermentation on reduction of glucosinolates, fibre and phytic acid in rapeseed (*Brassica napus*) meal. *Bioresour. Technol.* **2001**, *78*, 309–312.

- 75. Olukomaiya, O.O.; Fernando, W.C.; Mereddy, R.; Li, X.H.; Sultanbawa, Y. Solid-state fermentation of canola meal with Aspergillus sojae, Aspergillus ficuum and their co-cultures: Effects on physicochemical, microbiological and functional properties. *LWT* **2020**, 127, 109362. [CrossRef]
- 76. Tie, Y.; Li, L.; Liu, J.; Liu, C.; Fu, J.; Xiao, X.; Wang, G.; Wang, J. Two-step biological approach for treatment of rapeseed meal. *J. Food Sci.* 2020, 85, 340–348. [CrossRef] [PubMed]
- 77. Lucke, F.K.; Fritz, V.; Tannhauser, K.; Arya, A. Controlled fermentation of rapeseed presscake by Rhizopus, and its effect on some components with relevance to human nutrition. *Food Res. Int.* **2019**, *120*, 726–732. [CrossRef] [PubMed]
- Hao, Y.; Wang, Z.; Zou, Y.; He, R.; Ju, X.; Yuan, J. Effect of static-state fermentation on volatile composition in rapeseed meal. J. Sci. Food Agric. 2020, 100, 2145–2152. [CrossRef] [PubMed]
- 79. Yusuf, H.A.; Piao, M.; Ma, T.; Huo, R.Y.; Tu, Y. Enhancing the Quality of Total Mixed Ration Containing Cottonseed or Rapeseed Meal by Optimization of Fermentation Conditions. *Fermentation* **2021**, *7*, 234. [CrossRef]
- Do, S.H.; Kim, B.O.; Fang, L.H.; You, D.H.; Hong, J.S.; Kim, Y.Y. Various levels of rapeseed meal in weaning pig diets from weaning to finishing periods. *Asian-Australas J. Anim. Sci.* 2017, *30*, 1292–1302. [CrossRef]
- Choi, H.B.; Jeong, J.H.; Kim, D.H.; Lee, Y.; Kwon, H.; Kim, Y.Y. Influence of rapeseed meal on growth performance, blood profiles, nutrient digestibility and economic benefit of growing-finishing pigs. *Asian-Australas J. Anim. Sci.* 2015, 28, 1345–1353. [CrossRef]
- Zmudzinska, A.; Bigorowski, B.; Banaszak, M.; Roslewska, A.; Adamski, M.; Hejdysz, M. The effect of diet based on legume seeds and rapeseed meal on pig performance and meat quality. *Animals* 2020, 10, 1084. [CrossRef]
- Grabez, V.; Egelandsdal, B.; Kjos, N.P.; Hakenasen, I.M.; Mydland, L.T.; Vik, J.O.; Hallenstvedt, E.; Devle, H.; Overland, M. Replacing soybean meal with rapeseed meal and faba beans in a growing-finishing pig diet: Effect on growth performance, meat quality and metabolite changes. *Meat Sci.* 2020, 166, 108134. [CrossRef]
- 84. Landero, J.L.; Wang, L.F.; Beltranena, E.; Bench, C.J.; Zijlstra, R.T. Feed preference of weaned pigs fed diets containing soybean meal, *Brassica napus* canola meal, or Brassica juncea canola meal. *J. Anim. Sci.* **2018**, *96*, 600–611. [CrossRef]
- Hansen, J.; Øverland, M.; Skrede, A.; Anderson, D.M.; Collins, S.A. A meta-analysis of the effects of dietary canola/double low rapeseed meal on growth performance of weanling and growing-finishing pigs. *Anim. Feed Sci. Technol.* 2020, 259, 114302. [CrossRef]
- Skugor, A.; Kjos, N.P.; Sundaram, A.Y.M.; Mydland, L.T.; Anestad, R.; Tauson, A.H.; Overland, M. Effects of long-term feeding of rapeseed meal on skeletal muscle transcriptome, production efficiency and meat quality traits in Norwegian Landrace growing-finishing pigs. *PLoS ONE* 2019, 14, e0220441. [CrossRef] [PubMed]
- Skoufos, I.; Tzora, A.; Giannenas, I.; Bonos, E.; Papagiannis, N.; Tsinas, A.; Christaki, E.; Florou-Paneri, P. Dieraty inclusion of rapeseed meal as soybean meal subtitute on growth performance, gut microbiota, oxidative stability and fatty acid profile in growing-fattening pigs. *Asian J. Anim. Vet. Adv.* 2016, 11, 89–97. [CrossRef]
- Tan, C.Q.; Wei, H.K.; Sun, H.Q.; Long, G.; Ao, J.T.; Jiang, S.W.; Peng, J. Effects of supplementing sow diets during two gestations with konjac flour and Saccharomyces boulardii on constipation in peripartal period, lactation feed intake and piglet performance. *Anim. Feed Sci. Technol.* 2015, 210, 254–262. [CrossRef]
- Grela, E.R.; Czech, A.; Kiesz, M.; Wlazlo, L.; Nowakowicz-Debek, B. A fermented rapeseed meal additive: Effects on production performance, nutrient digestibility, colostrum immunoglobulin content and microbial flora in sows. *Anim. Nutr.* 2019, *5*, 373–379. [CrossRef] [PubMed]
- Quiniou, N.; Quinsac, A.; Crepon, K.; Evrard, J.; Peyronnet, C.; Bourdillon, A.; Royer, E.; Etienne, M. Effects of feeding 10% rapeseed meal (*Brassica napus*) during gestation and lactation over three reproductive cycles on the performance of hyperprolific sows and their litters. *Can. J. Ani. Sci.* 2012, *92*, 513–524. [CrossRef]
- 91. Czech, A.; Grela, E.R.; Kiesz, M.; Kłys, S. Biochemical andhaematological blood parameters of sows and piglets fed a diet with a dried fermented rapeseed meal. *Ann. Anim. Sci.* 2020, 20, 535–550. [CrossRef]
- 92. Czech, A.; Stepniowska, A.; Kiesz, M. Effect of fermented rapeseed meal as a feed component on the redox and immune system of pregnant sows and their offspring. *Ann. Anim. Sci.* 2022, *1*, 201–219. [CrossRef]
- Park, C.S.; Helmbrecht, A.; Htoo, J.K.; Adeola, O. Comparison of digestibility of amino acids in full-fat soybean, soybean meal, and peanut flour between broiler chickens and pigs. J. Ani. Sci. 2017, 95, 3110–3111. [CrossRef]
- 94. Bowland, J.P.; Hardin, R.T. Rapeseed meal as apartial replacement for soybean meal in the diets of growing gilts and of sows for up to three reproduction cucles. *Can. J. Anim. Sci.* **1973**, *53*, 355–363. [CrossRef]
- 95. Hanczakowska, E.; Weglarzy, K.; Bereza, M. Effectiveness of Rapeseed Press Cake (RPC) in Sow Feeding in Two Reproduction Cycles. *Ann. Anim. Sci.* 2012, *12*, 95–104. [CrossRef]