



Article

Conyza sumatrensis Resistant to Paraquat, Glyphosate and Chlorimuron: Confirmation and Monitoring the First Case of Multiple Resistance in Paraguay

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Received: 8 October 2020; Accepted: 19 November 2020; Published: 26 November 2020



Abstract: *Conyza sumatrensis* was reported to be associated with 20 cases of herbicide resistance worldwide, with a recent report of multiple drug resistance to paraquat, glyphosate, and chlorimuron in Brazil. In Paraguay, there were no reports of cases of resistance for this species; however, in 2017, researchers began identifying biotypes with resistance to paraquat, glyphosate, and chlorimuron, which is the focus of the present study. The goal of this study was to investigate the case of multiple resistance of *C. sumatrensis* to paraquat, glyphosate, and chlorimuron and to monitor the resistant biotypes in the departments of Canindeyú and Alto Paraná. Seeds were collected from sites where plants survived after herbicide application in the 2017/18 and 2018/19 seasons. After screening, biotypes were selected for the construction of dose–response curves. A resistance factor (RF) of 6.79 was observed for 50% control (C_{50}) and 3.92 for 50% growth reduction (C_{50}) for the application of paraquat. An RF of 12.32 was found for C_{50} and 4.15 for C_{50} and 10.96 for C_{50} . This confirms the application of chlorimuron, an RF of 11.32 was found for C_{50} and 10.96 for C_{50} . This confirms the multiple resistance of the *C. sumatrensis* biotype to paraquat, glyphosate, and chlorimuron. Population monitoring indicated the presence of *C. sumatrensis* with multiple resistance in departments of Canindeyú and Alto Paraná, Paraguay.

Keywords: ALS inhibitors; EPSPs inhibitors; herbicides; herbicide-resistance; South America; Sumatran fleabane; photosystem I inhibitor; weeds

1. Introduction

The selection of herbicide-resistant weed biotypes is one of the major problems in agriculture today. Herbicide resistant biotypes of hairy fleabane (*Conyza bonariensis* L. Cronquist), horseweed (*Conyza canadensis* L. Cronquist) and Sumatran fleabane (*Conyza sumatrensis* (Retz.) E. Walker) were previously reported. There are currently 105 herbicide-resistance cases for the three *Conyza* spp., which includes resistance to 5-enolpyruvylshikimate-3-phosphate synthase (EPSPs) inhibitors, acetolactate synthase (ALS) inhibitors, synthetic auxins, and photosystem I inhibitors, among others [1].

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The weed *Conyza* spp. is among the most problematic in soybean crops in Paraguay. Regarding the impact of this species on crops, Trezzi et al. [2] indicated that 2.7 plants of *Conyza* spp. M⁻² can reduce the soybean yield by 50%. The species has an annual life cycle, herbaceous size, and high seed production and is found in several agricultural environments, such as grain crops [3,4]. *Conyza sumatrensis* is believed to be originally from the subtropical region of South America, with dispersion to Europe, America, and Asia [5,6]. A single plant can produce more than 200 thousand seeds, which germinate mainly from fall to early spring [7].

Among the factors that lead to the selection of herbicide-resistant weed biotypes, there is the use of the same herbicides, or different herbicides but with the same mode of action, in which strong selection pressure results in the selection of resistant biotypes [8–10]. For soybean crops in Paraguay, one of the most common management techniques for *Conyza* spp. is the application, in the off-season, of glyphosate + 2,4-D with paraquat in sequence, in some cases with the application of diclosulam at soybean pre-emergence. In post-emergence, the application of glyphosate alone or in mixtures with ALS-inhibiting herbicides may be used.

One of the main tools for delaying the selection of resistant weed biotypes, as well as managing plants with cases of resistance, is the diversification of management practices, with an emphasis on the rotation and combination of herbicides integrated with non-chemical measures. In this context, monitoring resistant weed populations allows for the identification of the evolution and dispersion of resistance cases, which consequently provides important information for decision making for weed control [11–13].

Conyza sumatrensis presents 20 cases of herbicide resistance worldwide, and seven in Brazil [1], with a recent report of multiple resistance to glyphosate, chlorimuron, and paraquat [14]. In Paraguay, there were no reports of cases of resistance for this species; however, in 2017, researchers began focusing on identifying biotypes with resistance to glyphosate (an EPSPs inhibitor), chlorimuron (an ALS inhibitor), and paraquat (a photosystem I inhibitor) [1]. This was reported to the International Herbicide-Resistant Weed Database, and registered in it, and is the focus of the present study. Thus, the aim of this study was to investigate the case of triple resistance of *C. sumatrensis* to the herbicides glyphosate, chlorimuron, and paraquat, and to monitor resistant biotypes mainly in the departments of Canindeyú and Alto Paraná, in Paraguay.

2. Material and Methods

2.1. Seed Collection

Seeds were collected in sites where *C. sumatrensis* plants survived after herbicide burndown application in pre-sowing in the 2017/18 and 2018/19 growing seasons, in 33 agricultural areas located in the departments of Canindeyú and Alto Paraná, Paraguay. The geographical coordinates, biotype identification, and infested crops are listed in Table 1. Among these sites, there are two locations with possibly susceptible plants that served as a comparison control.

The sampling sites were chosen according to reports of control failures as sites with possible cases of resistance. Our seed collection followed the methodology proposed by Burgos et al. [15]. For each site, seeds were collected from 5–10 plants, with the same characteristics, pooled into a single sample per site (with at least 1000 physiologically mature seeds per sample).

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Table 1. Collection sites of *Conyza sumatrensis* populations with suspected resistance to herbicides. Canindeyú and Alto Paraná, Paraguay, 2017/18 and 2018/19 seasons.

Site	Department	Latitude	Longitude	Crop
01	Canindeyú	24°10′30″ S 54°41′33″ W		Soybean
02	Canindeyú	24°15′28″ S	54°47′30″ W	Soybean
03	Canindeyú	24°14′42″ S	54°52′25" W	Soybean
04	Canindeyú	24°13′18″ S	54°52′09" W	Soybean
05	Canindeyú	24°20′09″ S	54°49′41″ W	Soybean
06	Canindeyú	24°22′23″ S	54°50′26″ W	Soybean
07	Canindeyú	24°23′59″ S	54°50′54" W	Soybean
08	Canindeyú	24°34′53″ S	54°51′39″ W	Soybean
09	Alto Paraná	24°41′04″ S	54°52′11″ W	Soybean
10	Alto Paraná	25°00′01″ S	54°52′59" W	Soybean
11	Alto Paraná	25°03′13″ S	54°55′00″ W	Soybean
12	Alto Paraná	25°08′01″ S	54°58′02″ W	Soybean
13	Alto Paraná	25°10′38″ S	54°56′42″ W	Soybean
14	Alto Paraná	25°37′56″ S	54°58′16″ W	Soybean
15	Alto Paraná	25°36′42″ S	54°58′55″ W	Soybean
16	Alto Paraná	25°54′16″ S	55°07′03″ W	Soybean
17	Alto Paraná	25°00′14″ S	54°56′50″ W	Soybean
18	Canindeyú	24°04′26″ S	54°27′05" W	Soybean
19	Canindeyú	24°06′33″ S	54°31′46″ W	Soybean
20	Canindeyú	24°07′01″ S	54°34′59″ W	Soybean
21	Canindeyú	24°33′15″ S	54°46′47″ W	Soybean
22	Alto Paraná	25°02′23″ S	54°54′18″ W	Soybean
23	Canindeyú	24°21′03″ S	55°02′29″ W	Soybean
24	Canindeyú	24°04′08″ S	54°49′33″ W	Soybean
25	Canindeyú	24°15′57″ S	54°43′34″ W	Soybean
26	Canindeyú	24°20′19″ S	55°00′56″ W	Soybean
27	Canindeyú	24°03′34″ S	55°00′20″ W	Soybean
28	Canindeyú	24°09′26″ S	54°52′21″ W	Oat
29	Canindeyú	24°10′39″ S	54°53′20″ W	Oat
30	Canindeyú	24°12′00″ S	54°56′02″ W	Oat
31	Alto Paraná	25°45′04″ S	55°04′39″ W	Oat
32	Canindeyú	24°11′60″ S	54°56′10″ W	Chia
33	Canindeyú	24°08′58″ S	54°51′24″ W	Pasture

2.2. Screening

In a greenhouse, with daily irrigation, in the municipality of Katueté, Canindeyú, Paraguay $(24^{\circ}09'28.7'' \text{ S}, 54^{\circ}52'10.4'' \text{ W})$, about 100 seeds were sown in a plastic tray filled with substrate potting mix, for each sampling site, from October to November 2018. After germination, seedlings were transplanted into 800 mL plastic pots and filled with substrate potting mix with one seedling per pot. A completely randomized design with eight replications was used for each herbicide applied. Six herbicides, at the average recommended dose for the control of *C. sumatrensis* at the stage of 6–8 true leaves, were applied to plants, in addition to the control (no application) (Table 2), for each sampling site. The application took place at the stage of 6–8 true leaves, using a series 110.02 (TeeJet Technologies, Wheaton, IL, USA) CO_2 backpack sprayer pressurized at a constant pressure of 2 kgf cm⁻², with a bar with four fan nozzles, positioned at 50 cm from the target and at a speed of 1 m s⁻¹, providing a total spray volume of 200 L ha⁻¹.

Plant control was evaluated at 28 days after application (DAA), and visual scores were assigned to each experimental unit, where 0 represents no damage and 100% indicates total plant death [16]. The results were presented descriptively. After screening, plants from certain populations were selected to be grown alone to generate F_1 seeds, which were used for the construction of dose–response curves. The generation of F_1 is important to attest to the inheritance of the resistance character of populations.

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Herbicide	Group	Dose ¹	Commercial Product ²
2,4-D	O—synthetic auxins	1005	DMA [®] 6
paraquat	D—photosystem I inhibitors	400	Tecnoquat [®] SL
glyphosate	G—EPSPs inhibitors	720	Roundup Full [®] II
chlorimuron	B—ALS inhibitors	20	Poker® 75 WG
saflufenacil	E—PPO inhibitors	35	Heat [®]
glufosinate	H—GS inhibitors	500	Finale [®]
control (without application)	-	-	-

Table 2. Herbicides applied to *C. sumatrensis* plants for each site.

2.3. Dose–Response Curves

The same screening procedures were followed for sowing, seedling transplantation, and herbicide application, at the same location. The biotype whose F_1 seeds were collected and investigated for resistance came from sampling site 27 (24°03′34″S 55°00′20″W), and the susceptible biotype, also from the F_1 generation, came from site 33 (24°08′58″S 54°51′24″W). The herbicides applied were paraquat (0, 50, 100, 200, 400, 800, 1600, and 3200 g active ingredient (ai) ha⁻¹ (Tecnoquat SL, Tecnomyl S.A., Asunción, Paraguay) combined with 0.1% (v/v) non-ionic adhesive spreader; glyphosate (0; 90; 180; 360; 720; 1440; 2880 and 5760 g acid equivalent (ae) ha⁻¹ (Roundup Full II, Monsanto Paraguay S.A., Asunción, Paraguay); and chlorimuron (0, 2.5, 5, 10, 20, 40, 80, and 160 g ai ha⁻¹ (Poker 75 WG, Glymax Paraguay S.A., Hernandarias, Paraguay) combined with 0.5% (v/v) mineral oil. The doses used represent the dose recommended in the package insert for each herbicide, in proportions of 0, $^1/_8$, $^1/_4$, $^1/_2$, 1, 2, 4, and 8X the recommendation. A completely randomized design was used with four replications, for each herbicide dose. Each repetition consisted of a 0.8 L plastic pot, with one plant per pot.

The application took place at the stage of 6–8 true leaves, via a backpack sprayer pressurized with CO_2 , with a constant pressure of 2 kgf cm⁻², with a bar with four fan nozzles, series 110.02 (TeeJet Technologies, Wheaton, IL, USA) positioned at 50 cm from the target, and at a speed of 1 m s⁻¹, providing a total spray volume of 200 L ha⁻¹.

The plant control was evaluated at 28 DAA; visual scores were assigned to each experimental unit, where 0 indicates no damage and 100% indicates total plant death [16]. Dry mass evaluation was carried out at 28 DAA of the herbicides. Plants were cut at the ground level, placed in paper bags, dried in an oven at 70 °C for four days (to reach constant mass), and then measured.

2.4. Statistical Analysis

After screening for the generation of heritability (F_1), selection of biotypes, and realization of dose–response curves, the data of the evaluations from 28 DAA were subjected to analysis of variance and regression ($p \le 0.05$) and adjusted for the nonlinear logistic regression model proposed by Streibig [17]:

$$y = a/[1 + (x/b)^c],$$
 (1)

where y is the response variable (percentage control or shoot dry mass); x is the herbicide dose (g ha^{-1}); and a, b, and c are the estimated parameters of the equation, so that a is the amplitude between the maximum and the minimum point of the variable, b is the dose that provides 50% response, and c is the slope of the curve around b.

The non-linear logistic model provides an estimate of parameter C_{50} (50% control) or GR_{50} (50% growth reduction). Thus, we opted for mathematical calculation using the inverse equation of

¹ Doses in g ae ha⁻¹, for glyphosate and 2,4-D. For the others, in g ai ha⁻¹. Recommended average dose for the control of *C. sumatrensis* at the stage of 6–8 true leaves. ² DMA[®] 6, Dow AgroSciences Paraguay S.A., Asunción, Paraguay; Tecnoquat[®] SL, Tecnomyl S.A., Asunción, Paraguay; Roundup Full[®] II, Monsanto Paraguay S.A., Asunción, Paraguay; Poker[®] 75 WG, Glymax Paraguay S.A., Hernandarias, Paraguay; Heat[®], BASF Paraguay S.A., Asunción, Paraguay; Finale[®], BASF Paraguay S.A. Asunción, Paraguay

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Streibig [17], allowing the calculation of C_{50} , as proposed by Souza et al. [18]. The models used to obtain C_{50} were the same as those used by Takano et al. [19], Takano et al. [20], and Albrecht et al. [14].

$$x = b(|a/y - 1|)^{(1/c)}.$$
 (2)

Based on the values of C_{50} and GR_{50} , we calculated the resistance factor (RF = C_{50} or GR_{50} of the resistant biotype/ C_{50} or GR_{50} of the susceptible biotype). The resistance factor expresses the number of times that the dose required to control 50% resistant biotypes is greater than the dose controlling 50% susceptible biotypes [15,21].

3. Results

For all sampling sites, 100% control of *C. sumatrensis* plants was found with the application of saflufenacil and glufosinate; for 2,4-D, the results were close to 100%. For paraquat, \leq 50% control was observed in 19 out of the 33 sampling sites; for glyphosate, in 12 sites; for chlorimuron, in 7. The population of four sampling sites (13, 18, 25, and 27) had control \leq 50% for the application of glyphosate, chlorimuron, and paraquat, simultaneously. In 12 sites, a control \geq 86% was verified for paraquat, with only 2 sites for glyphosate, and only 2 for chlorimuron (Table 3).

Table 3. Control of the *C. sumatrensis* populations at 28 days after herbicide application.

Site	Paraquat	Glyphosate	Chlorimuron	2,4-D	Glufosinate	Saflufenacil	No Application	
%%								
1	100	65	70	100	100	100	0	
2	90	70	80	100	100	100	0	
3	70	65	65	98	100	100	0	
4	45	70	65	100	100	100	0	
5	100	60	75	100	100	100	0	
6	95	75	60	100	100	100	0	
7	25	65	65	100	100	100	0	
8	30	60	65	100	100	100	0	
9	60	45	45	95	100	100	0	
10	15	40	60	100	100	100	0	
11	95	50	40	100	100	100	0	
12	20	55	55	95	100	100	0	
13	20	40	50	100	100	100	0	
14	15	45	70	100	100	100	0	
15	15	60	65	100	100	100	0	
16	95	60	60	100	100	100	0	
17	100	65	60	100	100	100	0	
18	30	35	45	100	100	100	0	
19	95	65	40	98	100	100	0	
20	100	50	70	100	100	100	0	
21	20	55	55	100	100	100	0	
22	20	55	55	95	100	100	0	
23	40	50	60	98	100	100	0	
24	50	60	60	100	100	100	0	
25	20	50	50	100	100	100	0	
26	100	100	100	100	100	100	0	
27	15	30	40	95	100	100	0	
28	25	60	60	100	100	100	0	
29	40	40	65	100	100	100	0	
30	30	55	65	95	100	100	0	
31	25	45	55	98	100	100	0	
32	100	50	70	100	100	100	0	
33	100	100	100	100	100	100	0	

Site 27 (resistant to paraquat, glyphosate, and chlorimuron) and site 33 (susceptible) used for dose-response curves.

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Points with \leq 50% control were plotted in red, from 51% to 85% in yellow, \geq 86% in green, highlighting site 27 (R)—resistant to paraquat, glyphosate, and chlorimuron—and site 33 (S)—susceptible to herbicides. The proximity of collection points of the resistant and susceptible biotypes is presented in Figure 1.

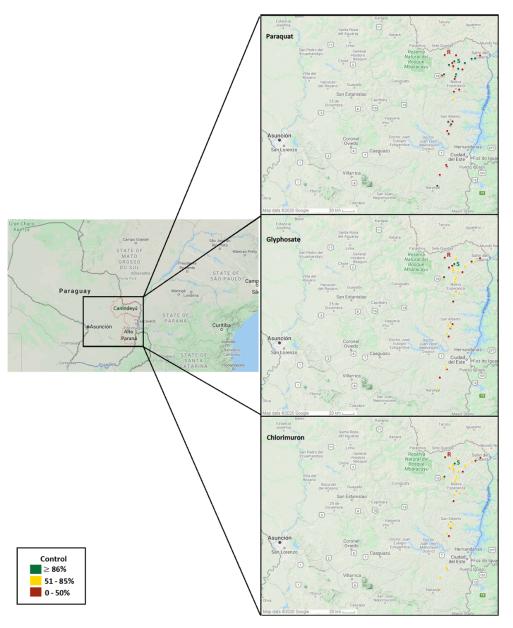


Figure 1. The geographic distribution of *C. sumatrensis* collection points with suspected resistance to herbicides, with respective effectiveness in the control. Canindeyú and Alto Paraná, Paraguay, 2017/18, season. R: site 27—resistant to paraquat, glyphosate, and chlorimuron; S: site 33—susceptible.

According to the results of the screening, the biotype from site 27 was selected to investigate the possible case of resistance to herbicides. An RF of 6.79 was observed for C_{50} (Figure 2A) and 3.92 for GR_{50} (Figure 2B), for the application of paraquat. The ineffectiveness in controlling *C. sumatrensis* under the application of glyphosate was also verified; for C_{50} and GR_{50} , RF was 12.32 (Figure 2C) and 4.15 (Figure 2D), respectively. For chlorimuron, RF was 11.32 for C_{50} (Figure 2E) and 10.96 for GR_{50} (Figure 2F). This confirmed the triple resistance of the *C. sumatrensis* biotype (site 27) to the herbicides paraquat, glyphosate, and chlorimuron (Table 4).

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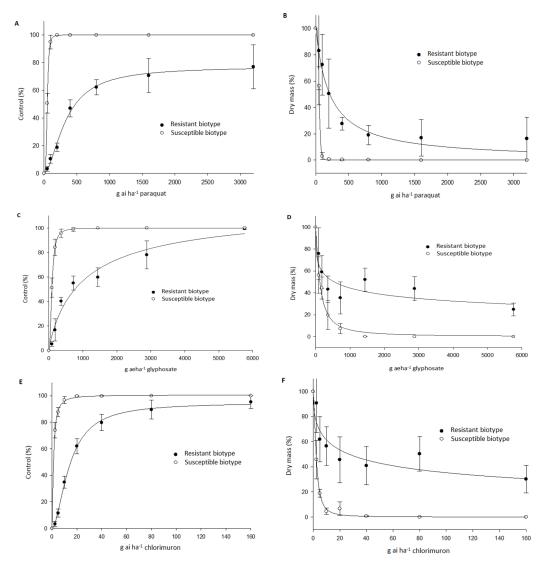


Figure 2. Control (**A**) and dry mass (**B**) of *C. sumatrensis* at 28 days after paraquat application. Control (**C**) and dry mass (**D**) of *C. sumatrensis* at 28 days after glyphosate application. Control (**E**) and dry mass (**F**) of *C. sumatrensis* (%) at 28 days after chlorimuron application. Site 27 (resistant to paraquat, glyphosate, and chlorimuron) and site 33 (susceptible). Bars shows the standard deviation (SD), n = 4.

Table 4. The required dose of herbicides for C_{50} (50% control) or GR_{50} (50% growth reduction) and resistance factor (RF) for *C. sumatrensis*.

	Paraquat		Glyphosate		Chlorimuron	
Biotype	C ₅₀	GR_{50}	C ₅₀	GR_{50}	C ₅₀	GR_{50}
	g ha ⁻¹					
Susceptible (site 33)	49.65	52.46	87.85	126.10	1.25	2.26
Resistant (site 27)	337.19	205.94	1082.36	523.35	14.16	24.78
RF	6.79	3.92	12.32	4.15	11.32	10.96

Dose in g ai ha⁻¹ for paraquat and chlorimuron, in g ae ha⁻¹ for glyphosate.

4. Discussion

The low efficiency of paraquat, glyphosate, and chlorimuron was observed in most areas where C. sum at rensis seeds were collected. Control of $\geq 86\%$ was observed in only two sites, for the three herbicides simultaneously. The identification of biotypes resistant to the three herbicides demonstrated the low effectiveness of these herbicides in controlling C. sum at rensis in a large area. The low effectiveness

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of these herbicides against C. sumatrensis was been reported in Brazil, including in states bordering Paraguay (Paraná and Mato Grosso do Sul). This low efficacy was confirmed by the cases of simple and multiple resistance to paraquat, glyphosate, and chlorimuron [14,22,23]. Albrecht et al. [14] showed multiple resistance to paraquat, glyphosate, and chlorimuron with RF for the C_{50} of 7.43, 3.58, and 14.35 and for the GR_{50} of 2.65, 2.79, and 11.31, respectively. In the present study, we observed RF for the C_{50} of 6.79, 12.32, and 11.32 and for the GR_{50} of 3.92, 4.15, and 10.96, respectively, for paraquat, glyphosate, and chlorimuron—that is, with RF close to paraquat and chlorimuron in the comparison between these biotypes. A higher RF was found for glyphosate in the biotype identified in Paraguay in this study.

In contrast, the herbicides saflufenacil and glufosinate were effective in controlling *C. sumatrensis* in all sampling sites, and the herbicide 2,4-D also showed good control; however, 2,4-D and other synthetic auxins are the subject of other specific studies due to the rapid necrosis, as studied in Brazil [24]. This reinforces the need to use different herbicides to control weeds, focusing not only on management, but also on preventing the selection of new resistant biotypes. Other studies demonstrated the effectiveness of these herbicides in the control of species of the genus *Conyza* [25–29]—in most situations, in combination with other herbicides, including products with confirmed resistance.

The combination and rotation of herbicides with different mechanisms of action are reinforced by several studies as essential in preventing the selection of new cases, in the effective management of already resistant cases, and in expanding the spectrum of action of the herbicidal treatment [30–32]. In addition, non-chemical measures, such as cover crops, should be highlighted. For example, vetch and barley crop residues were effective in suppressing *C. canadensis* [33], and black oat and wheat in suppressing *C. bonariensis* [34]. The importance of monitoring the populations of resistant weeds is therefore emphasized, which allows for the identification of the evolution and dispersion of cases of resistance, which consequently provides subsidies for decision-making for the effective management of weeds [35,36]. This study highlights the importance of and identifies the levels of effectiveness of herbicides in the region where the biotype was recorded.

The monitoring weed resistance cases is, therefore, an essential practice to understand, identify, and quantify the frequency of these plants in advance [37]. Thus, studies on resistance monitoring lead to increased research and, consequently, new techniques for the control of problematic plants, such as the use of pre-emergent herbicides to decrease the selection pressure [38–40].

In Paraguay, only four cases of herbicide-resistant weed biotypes have been officially reported, including the present study. In addition to this, *Euphorbia heterophylla* was found to be resistant to imazethapyr (an ALS inhibitor), and *Digitaria insularis* and *Bidens subalternans* were resistant to glyphosate [1]. This reinforces the importance of the present study, not only by identifying the first case of multiple resistance in the country, but also for monitoring the population of *C. sumatrensis* and investigating the effectiveness of herbicides. This provides important information for the management of this weed and for prevention of the selection of new resistant biotypes.

This population of *C. sumatrensis* meets all the criteria set to confirm a new case of resistance to paraquat, glyphosate, and chlorimuron, according to the criteria for confirming a new case of weed resistance to a herbicide of the Herbicide Resistance Action Committee (HRAC) [41]. These criteria include the definition of weed resistance; confirmation of the results obtained by scientifically based protocols; characterization of the heritability of weed resistance to the herbicide; demonstration of the practical impact in the field of weed resistance to the herbicide; and botanical identification of the weed species under analysis and not as a result of deliberate/artificial selection. This case was reported to the International Herbicide-Resistant Weed Database and is already registered [1].

5. Conclusions

Our results confirmed the multiple drug resistance of *C. sumatrensis* to the herbicides paraquat (a photosystem I inhibitor), glyphosate (an EPSPs inhibitor), and chlorimuron (an ALS inhibitor) as all the criteria set to prove new cases of resistance of weeds were met, thus scientifically demonstrating the first case of a weed with multiple resistance to herbicides in Paraguay.

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Population monitoring indicated the presence of *C. sumatrensis* plants with triple multiple resistance in the departments of Canindeyú and Alto Paraná, Paraguay, in most of the sampled sites. Further monitoring research on this weed species is ongoing in Paraguay, also covering the suspected resistance to 2,4-D and for other weed species, due to the scarcity of results in this country. Studies are underway with the objective of characterizing effective and sustainable alternatives for the control of this weed.

Author Contributions: Conceptualization and investigation, A.J.P.A., L.P.A., A.P., G.T., and F.S.A.; formal analysis, A.J.P.A., G.T., J.B.L., and M.T.Y.D.; data collection, A.J.P.A., G.T., and A.P.; writing—original draft, A.F.M.S.; writing—review and editing, all authors. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Heap, I. International Survey of Herbicide-Resistant Weeds. 2020. Available online: http://www.weedscience.org (accessed on 26 August 2020).
- 2. Trezzi, M.M.; Vidal, R.A.; Patel, F.; Miotto, E., Jr.; Debastiani, F.; Balbinot, A.A., Jr.; Mosquen, R. Impact of *Conyza bonariensis* density and establishment period on soyabean grain yield, yield components and economic threshold. *Weed Res.* **2015**, *55*, 34–41. [CrossRef]
- 3. Moreira, H.J.C.; Bragança, H.N.P. *Manual de Identificação de Plantas Infestantes: Cultivos de Verão*; FMC Agricultural Products: Campinas, Brazil, 2011.
- 4. Lorenzi, H. *Manual de Identificação e Controle de Plantas Daninhas: Plantio Direto e Convencional*, 7th ed.; Instituto Plantarum: Nova Odessa, Brazil, 2014.
- 5. Hao, J.H.; Qiang, S.; Liu, Q.Q.; Cao, F. Reproductive traits associated with invasiveness in *Conyza sumatrensis*. *J. Syst. Evol.* **2009**, 47, 245–254. [CrossRef]
- 6. Thebaud, C.; Finzi, A.C.; Affre, L.; Debussche, M.; Escarre, J. Assessing why two introduced *Conyza* differ in their ability to invade Mediterranean old fields. *Ecology* **1996**, 77, 791–804. [CrossRef]
- 7. Weaver, S.E. The biology of Canadian weeds. 115. *Conyza canadensis. Can. J. Plant Sci.* **2001**, *81*, 867–875. [CrossRef]
- 8. Riar, D.S.; Norsworthy, J.K.; Steckel, L.E.; Stephenson, D.O.; Eubank, T.W.; Bond, J.; Scott, R.C. Adoption of best management practices for herbicide-resistant weeds in midsouthern United States cotton, rice, and soybean. *Weed Technol.* **2013**, 27, 788–797. [CrossRef]
- 9. Green, J.M. The rise and future of glyphosate and glyphosate-resistant crops. *Pest Manag. Sci.* **2018**, 74, 1035–1039. [CrossRef]
- 10. Gage, K.L.; Krausz, R.F.; Walters, S.A. Emerging challenges for weed management in herbicide-resistant crops. *Agriculture* **2019**, *9*, 180. [CrossRef]
- 11. Lopez-Ovejero, R.F.; Takano, H.K.; Nicolai, M.; Ferreira, A.; Melo, M.S.; Cavenaghi, A.L.; Christoffoleti, P.J.; Oliveira, R.S., Jr. Frequency and dispersal of glyphosate-resistant sourgrass (*Digitaria insularis*) populations across Brazilian agricultural production areas. *Weed Sci.* **2017**, *65*, 285–294. [CrossRef]
- 12. Broster, J.C.; Pratley, J.E.; Ip, R.H.L.; Ang, L.; Seng, K.P. A quarter of a century of monitoring herbicide resistance in *Lolium rigidum* in Australia. *Crop Pasture Sci.* **2019**, *70*, 283–293. [CrossRef]
- 13. Lucio, F.R.; Kalsing, A.; Adegas, F.S.; Rossi, C.V.S.; Correia, N.M.; Gazziero, D.L.P.; Silva, A.F. Dispersal and frequency of glyphosate-resistant and glyphosate-tolerant weeds in soybean-producing edaphoclimatic microregions in Brazil. *Weed Technol.* **2019**, *33*, 217–231. [CrossRef]
- 14. Albrecht, A.J.P.; Pereira, V.G.C.; Souza, C.N.Z.; Zobiole, L.H.S.; Albrecht, L.P.; Adegas, F.S. Multiple resistance of *Conyza sumatrensis* to three mechanisms of action of herbicides. *Acta Sci. Agron.* **2020**, *42*, e42485. [CrossRef]
- 15. Burgos, N.R.; Tranel, P.J.; Streibig, J.C.; Davis, V.M.; Shaner, D.; Norsworthy, J.K.; Ritz, C. Confirmation of resistance to herbicides and evaluation of resistance levels. *Weed Sci.* **2013**, *61*, 4–20. [CrossRef]
- 16. Velini, D.E.; Osipe, R.; Gazziero, D.L.P. *Procedimentos Para Instalação, Avaliação e Análise de Experimentos Com Herbicidas*; SBCPD: Londrina, Brazil, 1995.

Agriculture 2020, 10, 582 10 of 11

- 17. Streibig, J.C. Herbicide bioassay. Weed Res. 1988, 28, 479–484. [CrossRef]
- 18. Souza, A.P.; Ferreira, F.A.; Silva, A.A.; Cardoso, A.A.; Ruiz, H.A. Logistic equation use in studying the dose-response of glyphosate and imazapyr by using bioassays. *Planta Daninha* **2000**, *18*, 17–28. [CrossRef]
- 19. Takano, H.K.; Oliveira, R.S., Jr.; Constantin, J.; Braz, G.B.P.; Franchini, L.H.M.; Burgos, N.R. Multiple resistance to atrazine and imazethapyr in hairy beggarticks (*Bidens pilosa*). *Ciencia e Agrotecnologia* **2016**, 40, 548–554. [CrossRef]
- 20. Takano, H.K.; Oliveira, R.S., Jr.; Constantin, J.; Braz, G.B.P.; Gheno, E.A. Goosegrass resistant to glyphosate in Brazil. *Planta Daninha* **2017**, *35*, e017163071. [CrossRef]
- 21. Hall, L.M.; Stromme, K.M.; Horsman, G.P.; Devine, M.D. Resistance to acetolactate synthase inhibitors and quinclorac in a biotype of false cleavers (*Galium spurium*). Weed Sci. **1998**, 46, 390–396. [CrossRef]
- 22. Santos, G.; Oliveira, R.S., Jr.; Constantin, J.; Francischini, A.C.; Osipe, J.B. Multiple resistance of *Conyza sumatrensis* to chlorimuron-ethyl and to glyphosate. *Planta Daninha* **2014**, 32, 409–416. [CrossRef]
- 23. Zobiole, L.H.S.; Pereira, V.G.C.; Albrecht, A.J.P.; Rubin, R.S.; Adegas, F.S.; Albrecht, L.P. Paraquat resistance of Sumatran fleabane (*Conyza sumatrensis*). *Planta Daninha* **2019**, 37, e019183264. [CrossRef]
- 24. Queiroz, A.R.; Delatorre, C.A.; Lucio, F.R.; Rossi, C.V.S.; Zobiole, L.H.S.; Merotto, A., Jr. Rapid necrosis: A novel plant resistance mechanism to 2,4-D. *Weed Sci.* **2020**, *68*, 6–18. [CrossRef]
- 25. Waggoner, B.S.; Mueller, T.C.; Bond, J.A.; Steckel, L.E. Control of glyphosate-resistant horseweed (*Conyza canadensis*) with saflufenacil tank mixtures in no-till cotton. *Weed Technol.* **2011**, 25, 310–315. [CrossRef]
- 26. Dalazen, G.; Kruse, N.D.; Machado, S.L.D.; Balbinot, A. Synergism of the glyphosate and saflufenacil combination for controlling hairy fleabane. *Pesquisa Agropecuária Tropical* **2015**, 45, 249–256. [CrossRef]
- 27. Tahmasebi, B.K.; Alebrahim, M.T.; Roldán-Gómez, R.A.; Silveira, H.M.; Carvalho, L.B.; Alcántara de la Cruz, R.; De Prado, R. Effectiveness of alternative herbicides on three *Conyza* species from Europe with and without glyphosate resistance. *Crop Prot.* **2018**, *112*, 350–355. [CrossRef]
- 28. Flessner, M.L.; Pittman, K.B. Horseweed control with preplant herbicides after mechanical injury from small grain harvest. *Agron. J.* **2019**, *111*, 3274–3280. [CrossRef]
- 29. Albrecht, A.J.P.; Albrecht, L.P.; Silva, A.F.M.; Ramos, R.A.; Corrêa, N.B.; Carvalho, M.G.; Lorenzetti, J.B.; Danilussi, M.T.Y. Control of *Conyza* spp. with sequential application of glufosinate in soybean pre-sowing. *Ciencia Rural* **2020**, *50*, e20190868. [CrossRef]
- 30. São Miguel, A.S.C.; Pacheco, L.P.; Souza, E.D.; Silva, C.M.R.; Carvalho, I.C. Cover crops in the weed management in soybean culture. *Planta Daninha* **2018**, *36*, e018172534. [CrossRef]
- 31. Büchi, L.; Wendling, M.; Amossé, C.; Jeangros, B.; Charles, R. Cover crops to secure weed control strategies in a maize crop with reduced tillage. *Field Crop. Res.* **2020**, 247, 107583. [CrossRef]
- 32. Loddo, D.; Scarabel, L.; Sattin, M.; Pederzoli, A.; Morsiani, C.; Canestrale, R.; Tommasini, M.G. Combination of herbicide band application and inter-row cultivation provides sustainable weed control in maize. *Agronomy* **2020**, *10*, 20. [CrossRef]
- 33. Campiglia, E.; Radicetti, E.; Mancinelli, R. Cover crops and mulches influence weed management and weed flora composition in strip-tilled tomato (*Solanum lycopersicum*). Weed Res. **2015**, 55, 416–425. [CrossRef]
- 34. Guareschi, A.; Cechin, J.; Bianchi, M.A.; Kruse, N.D.; Piccinini, F.; Pinheiro, R.T.; Machado, S.L.O. Cover plants as a suppression and increasing tool to hairy fleabane control. *Revista Brasileira de Ciencias Agrarias* **2020**, *15*, e7522. [CrossRef]
- 35. Ye, R.; Huang, H.; Alexander, J.; Liu, W.; Millwood, R.J.; Wang, J.; Stewart, C.N. Field studies on dynamic pollen production, deposition, and dispersion of glyphosate-resistant horseweed (*Conyza canadensis*). *Weed Sci.* **2016**, *64*, 101–111. [CrossRef]
- 36. Zabala, D.; Carranza, N.; Darghan, A.; Plaza, G. Spatial distribution of multiple herbicide resistance in *Echinochloa colona* (L.) Link. *Chil. J. Agric. Res.* **2019**, *79*, 576–585. [CrossRef]
- 37. Schultz, J.L.; Chatham, L.A.; Riggins, C.W.; Tranel, P.J.; Bradley, K.W. Distribution of herbicide resistances and molecular mechanisms conferring resistance in Missouri waterhemp (*Amaranthus rudis* Sauer) populations. *Weed Sci.* 2015, 63, 336–345. [CrossRef]
- 38. Bailly, G.C.; Dale, R.P.; Archer, S.A.; Wright, D.J.; Kaundun, S.S. Role of residual herbicides for the management of multiple herbicide resistance to ACCase and ALS inhibitors in a black-grass population. *Crop Prot.* **2012**, 34, 96–103. [CrossRef]
- 39. Oliveira, M.C.; Feist, D.; Eskelsen, S.; Scott, J.E.; Knezevic, S.Z. Weed control in soybean with preemergence-and postemergence-applied herbicides. *Crop Forage Turfgrass Manag.* **2017**, *3*, 1–7. [CrossRef]

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40. Busi, R.; Powles, S.B.; Beckie, H.J.; Renton, M. Rotations and mixtures of soil-applied herbicides delay resistance. *Pest Manag. Sci.* **2020**, *76*, 487–496. [CrossRef]

41. (HRAC) Herbicide Resistance Action Committee. Critérios Para Relatos Oficiais Estatísticos de Biótipos de Plantas Daninhas Resistentes a Herbicidas. 2017. Available online: https://www.hrac-br.org (accessed on 26 November 2019).

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