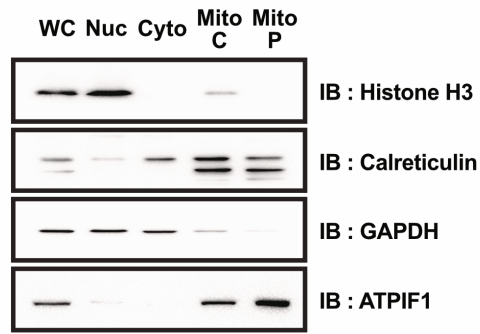
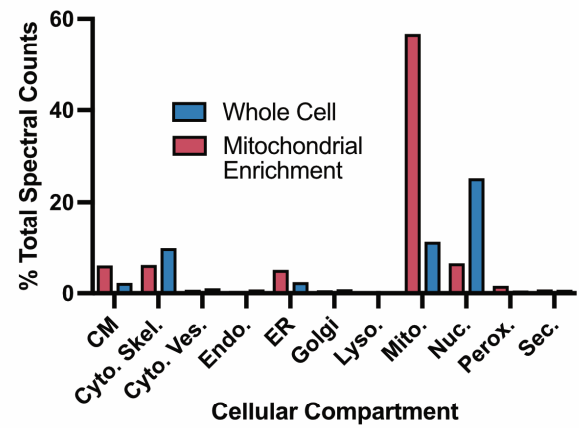


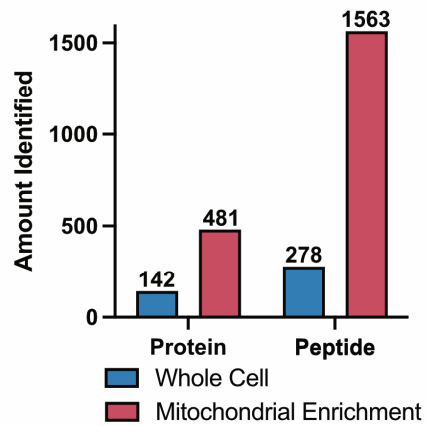
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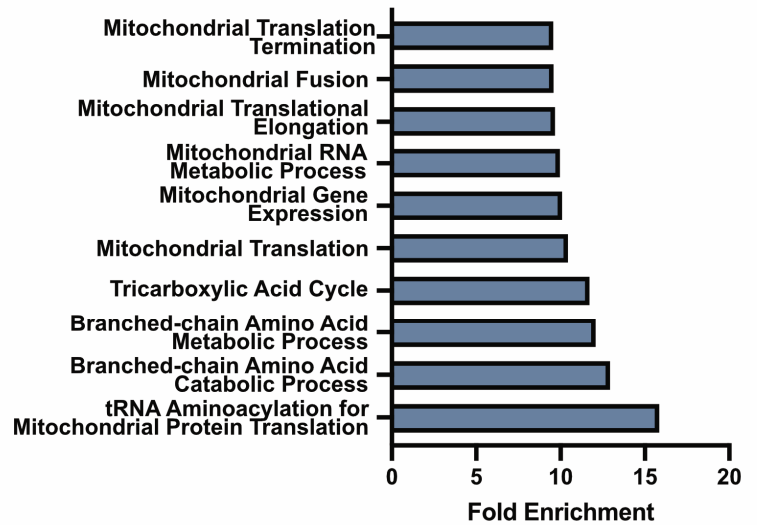
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**D**



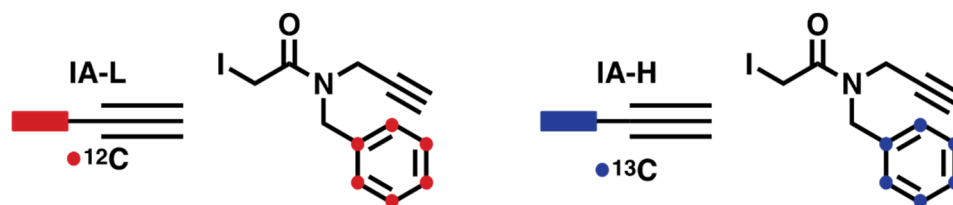
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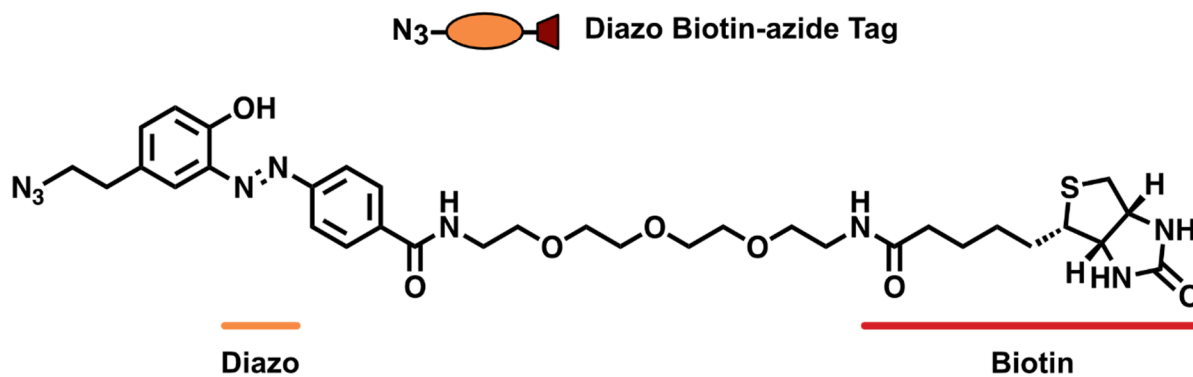
## **Figure S1. Evaluation of mitochondrial isolation by differential centrifugation**

- (A) Differential centrifugation workflow for mitochondrial isolation from whole cells.
- (B) Western blot analysis of isolated subcellular fractions during mitochondrial enrichment by differential centrifugation using organelle markers including Histone H3 (nuclear), Calreticulin (endoplasmic reticulum), GAPDH (cytosol), and ATP1F1 (mitochondria). “WC” depicts the whole cell fraction, “Nuc” depicts the nuclear fraction, and “Cyto” depicts the cytosolic fraction. “Mito-C” depicts the crude mitochondrial fraction and “Mito-P” depicts the pure mitochondrial fraction.
- (C) Bar graph of the percentage of total spectral counts belonging to proteins with specified UniProt sub-cellular localizations in a whole cell (blue) and isolated mitochondria (red) samples. Annotated compartments include cell membrane (CM), cytoskeleton (Cyto. Skel.), endosome (Endo.), endoplasmic reticulum (ER), golgi apparatus (Golgi), lysosome (Lyso.), mitochondria (Mito.), nucleus (Nuc.), peroxisome (Perox.), and secreted (Sec.).
- (D) Bar graph of the total annotated mitochondrial proteins and peptides identified from whole cell (blue) or isolated mitochondria (red).
- (E) Gene ontology analysis of the enriched biological processes in the Mito-P sample.

A



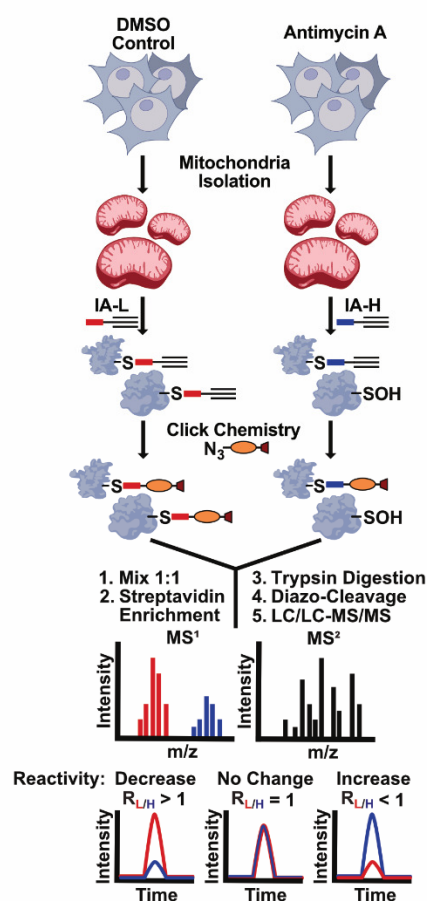
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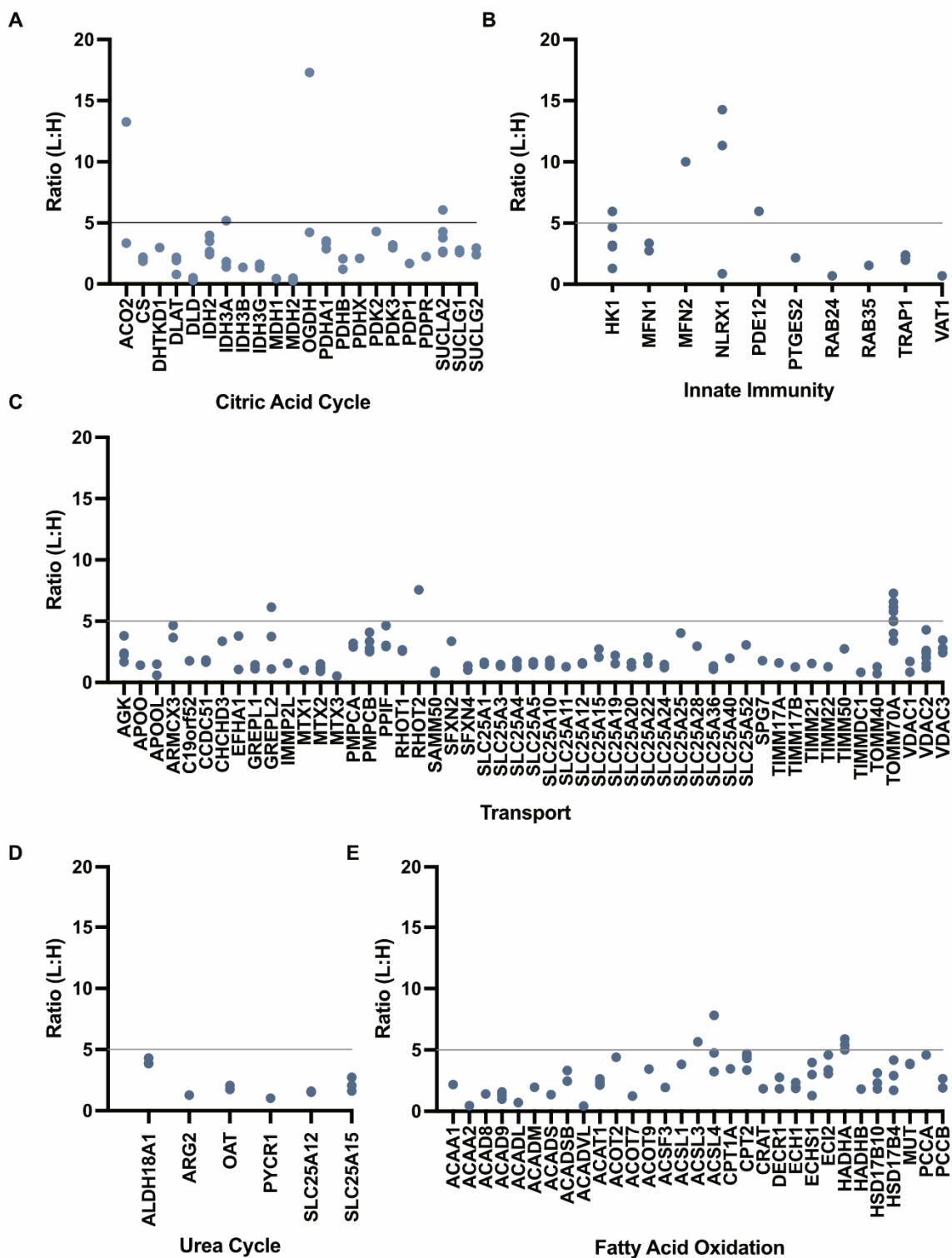
**Figure S2. Probe structures used for MS experiments.**

(A) Isotopically coded alkylating reagents, light (IA-L,  $^{12}\text{C}$ ) and heavy (IA-H,  $^{13}\text{C}$ ) iodoacetamide alkyne.

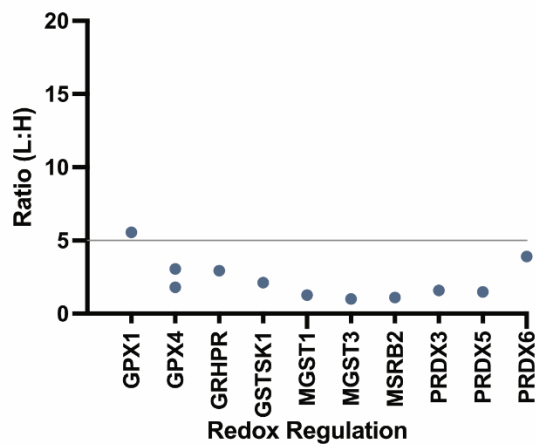
(B) Structure of cleavable diazo biotin-azide tag.



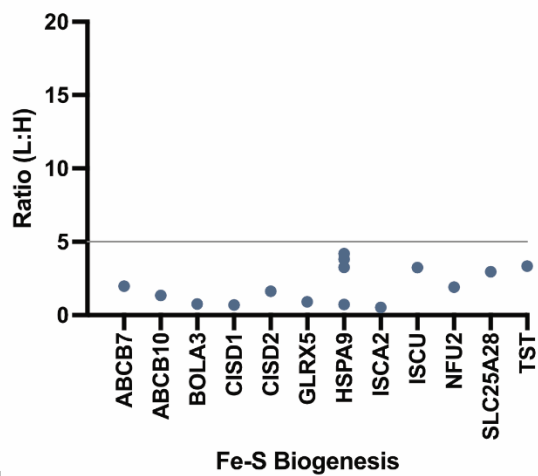
**Figure S3.** Workflow for quantifying redox-dependent changes in cysteine reactivity in isolated mitochondria using IsoTOP-ABPP. Labeling of thiols from DMSO control or Antimycin A (AMA) treated samples with IA-L or IA-H, respectively, appending cleavable biotin-azide tags, enrichment of labeled cysteine containing proteins, trypsin digestion, and isolation of labeled cysteine-containing peptides for quantitative MS to identify and quantify cysteine reactivity changes due to oxidation.



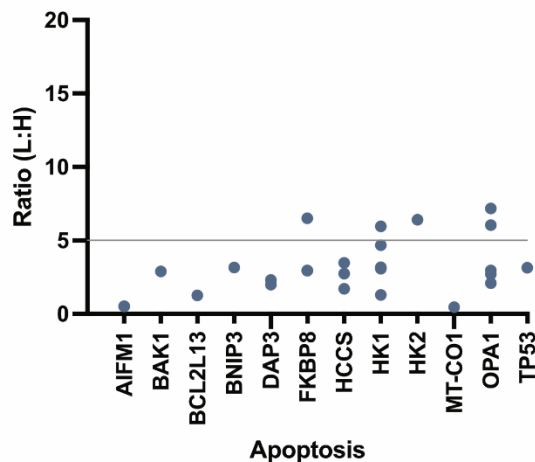
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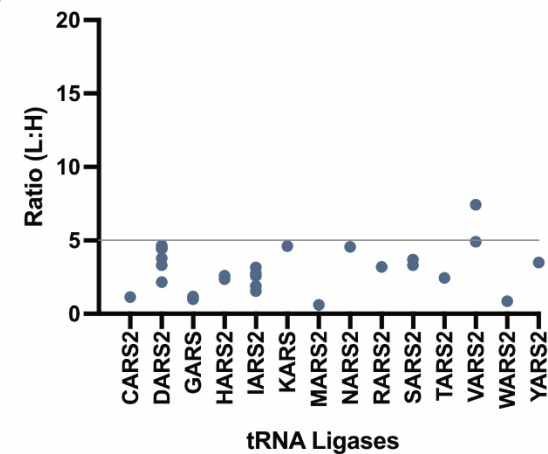
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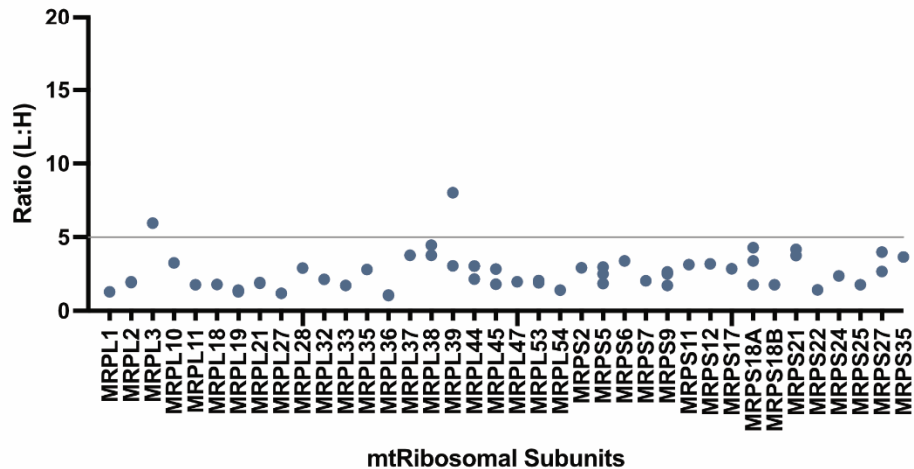
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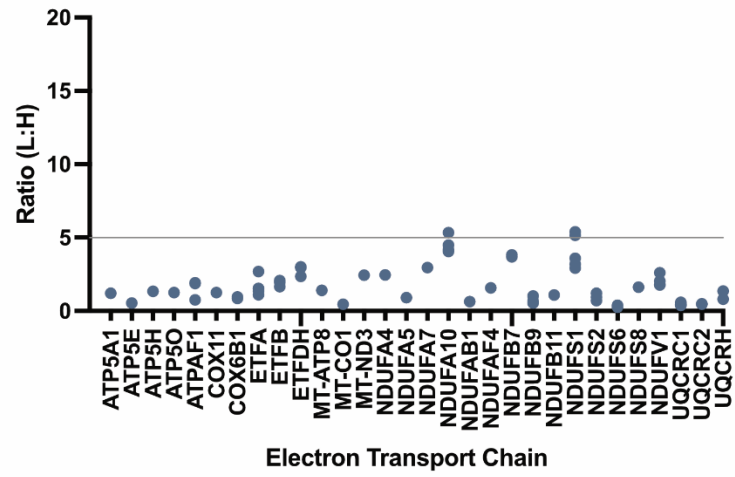
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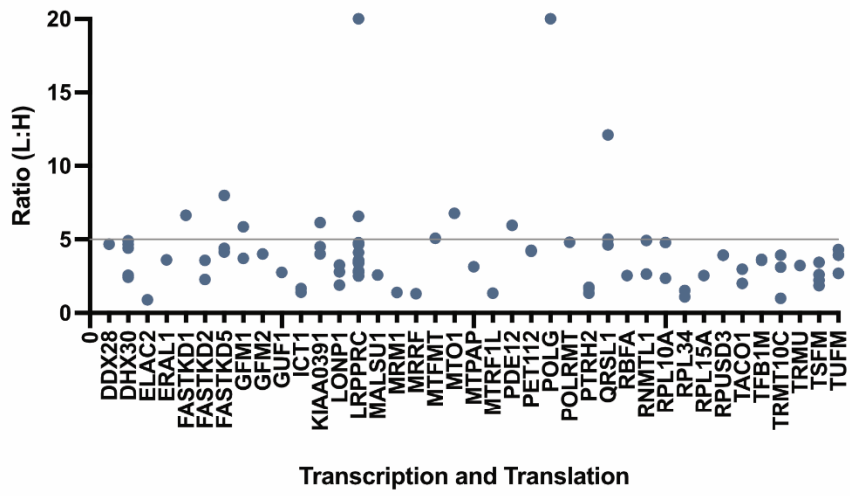
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K

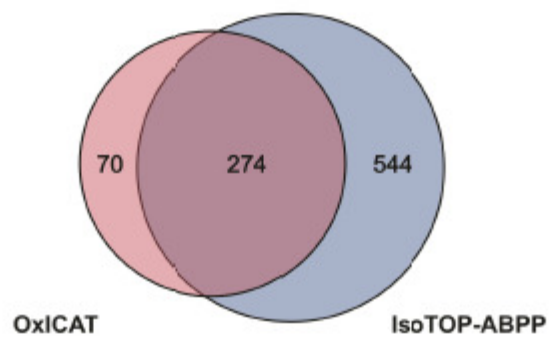


L



**Figure S4.** L:H ratios plotted from isoTOP-ABPP analysis of 10 mM peroxide treated Mito-P samples. Identified unique cysteine residues are plotted for proteins belonging to mitochondrial biological pathways and categories including the (A) citric acid cycle, (B) innate immunity, (C) transport, (D) urea cycle, (E) fatty acid oxidation, (F) redox regulation, (G) Fe-S biogenesis, (H) apoptosis, (I) tRNA ligases, (J) mitochondrial (mt)-ribosomal subunits, (K) electron transport chain, (L) transcription and translation.





**Figure S5.** Overlap between isoTOP-ABPP analysis and OxICAT analysis for all identified cysteines.