



# Review H<sub>2</sub>S Donors with Cytoprotective Effects in Models of MI/R Injury and Chemotherapy-Induced Cardiotoxicity

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Abstract: Hydrogen sulfide (H<sub>2</sub>S) is an endogenous signaling molecule that greatly influences several important (patho)physiological processes related to cardiovascular health and disease, including vasodilation, angiogenesis, inflammation, and cellular redox homeostasis. Consequently, H<sub>2</sub>S supplementation is an emerging area of interest, especially for the treatment of cardiovascular-related diseases. To fully unlock the medicinal properties of hydrogen sulfide, however, the development and refinement of H<sub>2</sub>S releasing compounds (or donors) are required to augment its bioavailability and to better mimic its natural enzymatic production. Categorizing donors by the biological stimulus that triggers their H<sub>2</sub>S release, this review highlights the fundamental chemistry and releasing mechanisms of a range of H<sub>2</sub>S donors that have exhibited promising protective effects in models of myocardial ischemia-reperfusion (MI/R) injury and cancer chemotherapy-induced cardiotoxicity, specifically. Thus, in addition to serving as important investigative tools that further advance our knowledge and understanding of H<sub>2</sub>S chemical biology, the compounds highlighted in this review have the potential to serve as vital therapeutic agents for the treatment (or prevention) of various cardiomyopathies.

**Keywords:** hydrogen sulfide; H<sub>2</sub>S donors; cardioprotection; MI/R injury; chemotherapy-induced cardiotoxicity; H<sub>2</sub>S codrugs

# 1. Introduction

Hydrogen sulfide (H<sub>2</sub>S) is a malodorous, toxic, and flammable gas that was once disregarded as a mere environmental and industrial pollutant [1–3]. Landmark studies near the turn of the 20th century [4–6], however, revealed that H<sub>2</sub>S is also a biologically active gas that is expressed in mammalian systems, primarily via the enzymatic metabolism of cysteine and homocysteine [7]. From these reports, a paradigm shift ensued, and today H<sub>2</sub>S is regarded as the third gasotransmitter, alongside nitric oxide (NO) and carbon monoxide (CO) [8–11].

 $H_2S$  is soluble in water (~80 mM at 37 °C [12]) and exhibits weak acidity that gives rise to an equilibrium between its diprotic ( $H_2S$ ) and hydrosulfide ( $HS^-$ ) forms in an aqueous environment. With a p $K_{a1}$  of 6.98 [12], its  $HS^-$  form dominates at physiological pH and begets its high reactivity and strong nucleophilic character under biologically relevant conditions.

In its diprotic form, its lipophilicity, low molecular weight, and gaseous nature enable  $H_2S$  to easily traverse the lipid bilayer, allowing it to act on intracellular targets that mediate numerous physiological and pathophysiological processes within the human body [13–16]. Its proven ability to reduce oxidative stress and inflammation [17–19], induce vasodilation [6], and promote angiogenesis [20] underscores the positive influence of  $H_2S$  on the cardiovascular system, specifically. Not surprisingly, small molecule donors that improve the exogenous delivery and bioavailability of  $H_2S$  are currently being investigated with great enthusiasm as potential cardioprotective agents [21].



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**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). This review will summarize the structure, reactivity, and mode of delivery for H<sub>2</sub>S donors that have displayed promising cardioprotective effects in myocardial ischemia-reperfusion (MI/R) injury and cancer chemotherapeutic-induced cardiotoxicity models, in particular. Thus, the compounds reported on herein not only represent important investigative tools for probing the chemical biology of hydrogen sulfide but may also serve to unlock its vast therapeutic potential for the treatment of cardiovascular-related diseases.

#### 2. H<sub>2</sub>S Biosynthesis and Metabolism

In mammals, both enzymatic and nonenzymatic pathways are involved in H<sub>2</sub>S biosynthesis, with the former being the principal route towards its formation. The use of enzymes provides strict spatiotemporal control over the production of H<sub>2</sub>S, resulting in concentration variances in specific tissues and cellular compartments, and in response to certain physiological and pathophysiological events. The three enzymes primarily responsible for H<sub>2</sub>S biosynthesis are cystathionine  $\beta$ -synthase (CBS) [22], cystathionine  $\gamma$ -lyase (CSE) [23], and 3-mercaptopyruvate sulfurtransferase (3-MST) [24].

CBS and CSE are ubiquitous enzymes of the transsulfuration pathway that facilitate the conversion of homocysteine to cysteine via the intermediate cystathionine (Figure 1) [25,26]. Both are pyridoxal 5'-phosphate (PLP)-dependent enzymes that are primarily located in the cytosol and generate H<sub>2</sub>S via the direct desulfhydration of cysteine and homocysteine. In addition to being primarily responsible for H<sub>2</sub>S biosynthesis in the brain and central nervous system, CBS is amply expressed in the ileum, kidneys, liver, and uterus [5,27,28]. CSE, on the other hand, exhibits low expression levels in the central nervous system but is the principal H<sub>2</sub>S-producing enzyme of the cardiovascular system [29].



**Figure 1.** Enzymatic and nonenzymatic production of  $H_2S$  in mammalian systems. PLP: pyridoxal 5'-phosphate; CBS: cystathionine  $\beta$ -synthase; CSE: cystathionine  $\gamma$ -lyase; CAT: cysteine amino-transferase; 3-MST: 3-mercaptopyruvate sulfurtransferase; NADPH: nicotinamide adenine dinucleotide phosphate.

Unlike CBS and CSE, 3-MST is a PLP-independent enzyme that is chiefly expressed in mitochondria and produces H<sub>2</sub>S from the indirect desulfhydration of cysteine [30]. As depicted in Figure 1, in this pathway, cysteine must first be transformed into 3-mercaptopyruvate

(3MP) via the enzyme cysteine aminotransferase (CAT). Then, using 3MP as a substrate, 3-MST transfers a sulfur atom onto itself forming a hydropersulfide (3-MST-SSH). In the presence of reductants, 3MST-SSH is reduced, releasing  $H_2S$  in the process.

In addition to the enzymatic routes outlined above, nonenzymatic pathways also contribute to the endogenous production of  $H_2S$  in mammals. In general, sulfane sulfur and other reactive sulfur species (RSS), including hydropersulfides (RSSH), polysulfides (RSS<sub>n</sub>R), and thiosulfate ( $S_2O_3^{2-}$ ), serve as effective  $H_2S$  precursors in the presence of glutathione and other reductants (Figure 1) [31–33]. To this end, processes that increase the production of nicotinamide adenine dinucleotide phosphate (NADPH), which facilitates the recycling of oxidized glutathione back to its reduced form, have been shown to enhance this nonenzymatic pathway and promote  $H_2S$  biosynthesis [8].

While less is known about the metabolism and removal of  $H_2S$  from mammalian systems, the primary pathways are believed to involve mitochondrial oxidation [34,35], cytosolic methylation [36], hemoglobin and metalloprotein binding [37], expiration via the lungs [38], and its storage in proteins as bound sulfane sulfur [39]. The majority of  $H_2S$  is ultimately excreted via the kidneys in the form of sulfate ( $SO_4^{2-}$ ) [40]. This oxidation of  $H_2S$  occurs in mitochondria and is facilitated by the enzymes sulfide quinone reductase (SQR) and rhodanese. This metabolic process also accentuates the biological activity of  $H_2S$  and its ability to stimulate oxidative phosphorylation and ATP production through its donation of electrons to the mitochondrial electron transport chain through SQR and mitochondrial complex II [41,42].

#### 3. H<sub>2</sub>S Bioactivity and Its Attenuation of Myocardial Ischemia-Reperfusion Injury

In addition to serving as a mitochondrial protectant and stimulator of mitochondrial bioenergetics, endogenous  $H_2S$  has been shown to play a key role in several other physiological and pathophysiological processes [14–16,43–47]. The cardiovascular system, in particular, appears to be positively influenced by  $H_2S$  given its involvement in vasodilation and blood pressure regulation [6,48]; its antioxidative [19], anti-inflammatory [17,49], and cytoprotective properties [50,51]; and its ability to promote angiogenesis [20]. Additionally, recent evidence suggests that the co-release of  $H_2S$  (via the transsulfuration pathway) and adenosine (via the methionine cycle) may protect the myocardium from injury [52,53]. For these reasons it is theorized that the exogenous delivery of  $H_2S$  may hold therapeutic value for the prevention and treatment of various cardiovascular-related diseases [54,55], including myocardial ischemia-reperfusion (MI/R) injury [56–62] (Figure 2).



**Figure 2.** Molecular mechanisms that underscore the cardioprotective effects of H<sub>2</sub>S, including its ability to combat MI/R injury.

Myocardial ischemia occurs when blood flow to the heart is restricted due to the buildup of plaque in a coronary artery. If left unchecked, this may lead to myocardial infarction, or heart attack, which is the leading cause of death worldwide [63,64]. To repair myocardial structural damage and prevent ischemic progression, reperfusion therapy is typically employed. This rapid return of blood to ischemic tissue, however, often leads to (MI/R) injury caused by inflammation and oxidative damage [65,66]. Increased levels of reactive oxygen species (ROS), coupled with an overwhelmed antioxidant defense, play a major role in reperfusion injury and can exacerbate cardiac damage that occurs during ischemia [67].

Intracellular calcium overload, a hallmark of reperfusion injury, stimulates the translocation of CSE from the cytosol to mitochondria, which elevates the production of H<sub>2</sub>S within that subcellular space [68]. This innate response of the human body is produced in an effort to preserve mitochondrial function and protect the myocardium from oxidative damage, highlighting the potential for therapeutic intervention with H<sub>2</sub>S delivery. Indeed, recent studies have highlighted the protective effects of exogenous hydrogen sulfide during MI/R. One of the earliest examples in vitro was a study conducted by Johansen and co-workers [69]. Using an isolated perfused heart assay with rats, preconditioning with 1  $\mu$ M NaHS (an H<sub>2</sub>S equivalent in buffer) 10 min prior to coronary occlusion and up until 10 min post reperfusion, they observed a 20% reduction in infarct size. Pretreatment with Glibenclamide (K<sub>ATP</sub> blocker) nullified the effect of exogenous H<sub>2</sub>S, which supports its involvement in K<sub>ATP</sub> channel opening as a primary mechanism of alleviation. Later studies have shown that H<sub>2</sub>S promotes the persulfidation (protein-SSH) of Cys43 of the K<sub>ATP</sub> protein, resulting in channel opening, an influx of K<sup>+</sup>, and vascular smooth muscle relaxation.

Additionally, sulfide salts have been used to demonstrate the protective effects of  $H_2S$  against MI/R injury in vivo. In an early study by Sivarajah et al. [70], mice were exposed to 25 min of regional myocardial ischemia and 2 h of subsequent reperfusion. When NaHS (3 mg/kg) was delivered 15 min prior to ischemia, a 26% reduction in infarct size was reported in comparison to the vehicle control. Subsequently, Elrod and co-workers investigated the impact of exogenous  $H_2S$  being delivered at the time of reperfusion rather than prior to the ischemic event [71]. In their study, mice were subjected to 30 min of left coronary artery ischemia followed by a 24 h period of reperfusion in the presence of Na<sub>2</sub>S (50 µg/kg). Remarkably, they observed a 72% reduction in infarct size under these conditions.

While sulfide salts, such as NaHS and Na<sub>2</sub>S, serve as convenient  $H_2S$  precursors, their addition to buffered solutions results in a rapid surge in  $H_2S$  concentration, followed by a swift decline due to the instability and transient nature of hydrogen sulfide [72]. Moreover, these characteristics poorly mimic the slow and steady enzymatic production of  $H_2S$ , which often leads to adverse side effects when sulfide salts are employed. For these reasons, small molecule donors designed to release  $H_2S$  in a controlled fashion, and under biologically relevant conditions, have been sought to harness the medicinal properties of  $H_2S$  [73–77].

In the ensuing section, we will highlight examples of small molecule donors that better mimic the natural biosynthesis of H<sub>2</sub>S and exhibit promising cardioprotective effects, especially against myocardial ischemia-reperfusion injury.

### 4. H<sub>2</sub>S Donors That Protect against Myocardial Ischemia-Reperfusion Injury

Hydrogen sulfide donors with success at protecting against MI/R injury are highlighted in Table 1 and arranged by their mechanism for  $H_2S$  release. In this section, the  $H_2S$ releasing mechanism of each donor will be detailed, and their resulting therapeutic effects in various MI/R injury models will be summarized. \_

H <sub>2</sub> S Donor	Release Mechanism	Preclinical Studies
MeO - HeO	Hydrolysis-Triggered	In vivo rat and diabetic mice models of MI/R injury
R S-S DTTs	Hydrolysis-Triggered	Rat isolated perfused heart model of MI/R injury
S Ph−P−O HN R O JK Donors	pH-Triggered	H9c2 cardiomyocyte model of H/R injury and in vivo murine model of MI/R injury
$R_1 \underbrace{\downarrow}_{O}^{S} \underbrace{N}_{H} \underbrace{R_2}_{R_2}$	Thiol-Triggered	In vivo murine model of MI/R injury
R <sub>1</sub> S <sub>S</sub> R <sub>2</sub> Acyl Perthiols	Thiol-Triggered	In vivo murine model of MI/R injury
Allyl Thioesters	Thiol-Triggered	In vivo murine model of MI/R injury
$\begin{array}{c} R_{1} \\ N_{1} \\ N_{2} \\ H_{2} \\ \end{array} \xrightarrow{N_{n}} N_{0} \\ N_{0} \\ S_{S} \\ S_{n} \\ \mathsf$	Thiol-Triggered	Mouse isolated perfused heart model of MI/R injury
N=C=S Ar <sup>′</sup> Aryl Isothiocyanates	Thiol-Triggered	Rat isolated perfused heart model and in vivo murine model of MI/R Injury
Ar NH <sub>2</sub> Arylthioamides	Thiol-Triggered	In vivo rabbit model of MI/R injury
$R_{2} \xrightarrow{O} R_{1}$ R <sub>1</sub> S S R <sub>1</sub> Hydroxymethyl Persulfide Prodrugs	Enzyme-Triggered	in vivo murine model of MI/R injury
HSDs	ROS-Triggered	H9c2 cardiomyocyte model of H/R injury and in vivo murine model of MI/R injury

Table 1. Synthetic H<sub>2</sub>S donors with documented protective effects against MI/R injury.

# 4.1. Hydrolysis-Triggered Donors

Morpholin-4-ium 4-methoxyphenyl (morpholino) phosphinodithioate (GYY4137) is the first and most-researched H<sub>2</sub>S donor ever developed [78,79]. It was accessed by treating Lawesson's reagent with morpholine to impart high water solubility (~30 mg/mL at pH 7.4), which facilitates its use in biological studies. The proposed H<sub>2</sub>S releasing mechanism for GYY4137 is depicted in Figure 3. From detailed mechanistic work carried out by Alexander and co-workers [80], a two-step hydrolysis was put forth, which ultimately yields an arylphosphonate and 2 equiv of H<sub>2</sub>S. The second hydrolysis step, however, was deemed to be too slow to be responsible for any of its observed biological activity, suggesting that GYY4137 primarily undergoes a single hydrolytic P–S bond cleavage event in water to release 1 equiv of H<sub>2</sub>S.



Figure 3. Hydrolysis-triggered H<sub>2</sub>S release from GYY4137.

In stark contrast to sulfide salts, GYY4137 is recognized for its ability to provide the slow and continuous release of H<sub>2</sub>S for up to a week after its introduction to water. In its first reported study, GYY4137 was shown to relax rat aortic rings due to its activation of vascular smooth muscle  $K_{ATP}$  channels [78]. Moreover, unlike sulfide salts whose effects were brief, GYY4137 was found to be a far more potent vasorelaxant, presumably due to its sustained release of H<sub>2</sub>S and extended interaction with aortic rings. Perhaps not surprisingly, GYY4137 has also exhibited protective effects against MI/R injury [81–83]. Beyond its activation of vascular smooth muscle  $K_{ATP}$  channels [78,84], additional mechanisms have been invoked which include the ability of GYY4137 to attenuate oxidative stress and apoptosis through increased Bcl-2 expression and its activation of the Nrf2 signaling pathway [81,82].

Aside from GYY4137, 1,2-dithiole-3-thiones (DTTs) represent another important  $H_2S$  donating scaffold that operates via chemical hydrolysis (Figure 4) [85]. Although detailed mechanistic studies have yet to be carried out, conventional wisdom suggests that, in water, DTTs are converted into their corresponding 1,2-dithiole-3-one structure with the concurrent liberation of  $H_2S$ .



Figure 4. Hydrolysis-triggered H<sub>2</sub>S release from DTTs.

ADT and ADT-OH are the most common among this donor class, and their biological properties have been assessed in numerous disease models [86–91]. Perhaps most notably, several interesting H<sub>2</sub>S donor hybrids have been obtained by coupling ADT-OH through its phenol onto other therapeutically useful drugs [92], yielding compounds such as MADTOH and ACS14 (Figure 5).



Figure 5. Chemical structures of DTTs and donor hybrids with protective effects against MI/R injury.

Impressive drug synergism was observed with MADTOH, a monastrol-H<sub>2</sub>S-releasing hybrid, as increased inhibitory effects against L-type calcium channels were observed with this compound in comparison to both monastrol and ADT-OH alone [93]. L-type calcium channel blockers hold promise as an effective therapy for several cardiovascular disorders, including myocardial ischemia [94]. Thus, hybrid molecules, such as MADTOH, may be especially advantageous in treating MI/R injury and warrant further studies.

Along those lines, ACS14 is an  $H_2$ S-releasing, nonsteroidal anti-inflammatory hybrid that combines aspirin and donor ADT-OH. Originally reported on in 2009 [95], ACS14 was first developed in an effort to reduce the gastric toxicity of aspirin by combatting redox imbalance through its release of  $H_2$ S and subsequent increase in heme oxygenase-1 expression. Since this initial report, the cardioprotective effects of ACS14 have also been highlighted in later studies, including its ability to reduce MI/R injury in buthionine sulfoximine-treated rats [96,97].

Similarly, AP39 is an ADT-OH conjugate with impressive therapeutic effects in cardiovascular disease models (Figure 5) [98,99]. By combining ADT-OH with a triphenylphosphonium moiety through an ester linkage, AP39 effectively targets mitochondria, which significantly improves its potency. This was first established in a study aimed at assessing its effects on mitochondrial bioenergetics, which noted that only nanomolar concentrations of AP39 were required to observe stimulatory effects whereas micromolar doses of other  $H_2S$  donors are typically required to evoke similar results. The selective delivery of  $H_2S$  to mitochondria may also heighten its cardioprotective qualities. Indeed, later studies have showcased the ability of AP39 to protect myocardium from ischemia-reperfusion injury by significantly attenuating mitochondrial ROS production and through its stabilization of mitochondrial membrane potentials [100–102].

#### 4.2. pH-Triggered Donors

JK donors are a class of pH-triggered,  $H_2S$ -releasing compounds developed by Xian and co-workers [103]. By appending different amino acids, a series of phosphorothioatebased donors were accessed that undergo an intramolecular cyclization reaction that liberates  $H_2S$  with high efficiency in weakly acidic (pH 5–6) environments (Figure 6). This pathway, however, appears to be inoperable under neutral to slightly basic conditions (pH 7–8), which provides greater spatiotemporal control over their delivery of hydrogen sulfide. These observations are likely to stem from the fact that under weakly acidic conditions, the phosphorothiol moiety is protonated and functions as a good leaving group, while the carboxylate component still resides in its deprotonated, nucleophilic form.



Figure 6. pH-triggered H<sub>2</sub>S release from JK donors.

Since numerous pathological conditions are known to lead to a reduction in pH (inflammation, cancer, and cardiovascular disorders), JK donors have the potential to selectively deliver  $H_2S$  under conditions in which a therapeutic benefit is likely to arise. In their original study, the authors successfully demonstrated that both JK-1 and JK-2 (Figure 7) could provide significant cardioprotection in both cellular and in vivo murine models of MI/R injury [103].



Figure 7. JK donors with established cardioprotective effects in MI/R injury models.

It is worth noting that additional donors of this type have been prepared by further modifying the amino acid substituent. Phosphorothioate 18 (Figure 7), for example, was recently accessed and found to protect H9c2 cardiomyocytes from hypoxia-reoxygenation (H/R) injury [104]. In addition, JK-1 was shown to exhibit low toxicity and good pharma-cokinetic properties, accentuating the fact that further structure–activity relationship (SAR) studies and additional therapeutic and preclinical profiling within this series is likely to be advantageous.

#### 4.3. Thiol-Triggered Donors

 $H_2S$  donors selectively responsive to biologically abundant thiols, such as cysteine and glutathione, have also exhibited promising cardioprotective effects. Figure 8 outlines specific compounds within this series that have displayed promising protective effects in MI/R injury models.



**Figure 8.** Thiol-triggered donors with established cardioprotective effects in H/R and MI/R injury models.

Among the first to be examined were a series of *N*-mercapto-based donors (NSHDs) developed by Zhao et al. (Figure 8) [105]. These compounds were shown to be stable in buffer and require the presence of cysteine to effectively deliver H<sub>2</sub>S in aqueous media. Specifically, within this donor class, NSHD-1, NSHD-2, and NSHD-6 demonstrated cytoprotective effects against H<sub>2</sub>O<sub>2</sub>-induced damage in H9c2 cardiomyocytes. Furthermore, NSHD-1 and NSHD-2 also exhibited potent cardioprotective effects in a murine model of MI/R injury.

Additionally, acyl perthiols, allyl thioesters, and perthiocarbamates are responsive to cellular thiols and have established cardioprotective effects in H9c2 cardiomyocytes and other MI/R injury models as a result of their  $H_2S$  release. In the case of acyl perthiols,

compounds 8a and 8l demonstrated notable reductions in infarct size relative to vehicletreated mice in a murine MI/R injury model [58]. Moreover, a significant reduction in circulating cardiac troponin I was observed in both 8a- and 8l-treated mice, which supports the involvement of an H<sub>2</sub>S-related mechanism in their cardioprotection. Within the allyl thioester series, 5e was shown to be the most potent donor in cardiomyocyte (H9c2) models of oxidative damage [106]. It also displayed protective qualities in an in vivo mouse model, reducing infarct size and cardiomyocyte apoptosis. Similarly, perthiocarbamate 7b showcased impressive cardioprotective effects in a Langendorff model of MI/R [107].

Although very electrophilic, isothiocyanates are another class of thiol-activated donors with promising cardioprotective characteristics. In Langendorff-perfused rat hearts, 4CPI was shown to improve post-ischemic recovery through its attenuation of oxidative stress and activation of mitoK<sub>ATP</sub> channels [108]. Through an extensive SAR study, 3-pyridyl-isothiocynante was identified as another potent donor within this series, exhibiting maximum myocardial protection in an in vivo rat model for acute myocardial infarction at a dose of just 20  $\mu$ g/kg and from its activation of mitoK<sub>ATP</sub> channels [109].

Arylthioamides are the final donor class that we will touch upon within this section. What makes arylthioamides distinct from the donors mentioned above is that their release of  $H_2S$  is extremely slow and inefficient, even with the addition of nucleophilic thiols [110]. Moreover, the release of  $H_2S$  from this scaffold proceeds through an unidentified mechanistic pathway. Nevertheless, two hybrid adenine-containing donors, arylthioamide 4 and 11, appear to show synergistic cardioprotective effects by activating the PKG/PLN pathway in ischemic myocardium [111].

Pathways for thiol-triggered release of H<sub>2</sub>S have been explored with these donors. Plausible mechanisms put forth by the authors, based on detailed mechanistic studies, the identification of reaction intermediates, and established organic reactivity, are presented below.

As depicted in Figure 9, NSHDs initially undergo a nucleophilic acyl substitution with cysteine to form a thioester and an *N*-mercapto (*N*-SH) species. Although this first step is reversible, the ensuing thioester undergoes a rapid *S*-to-*N*-acyl transfer that essentially renders this step irreversible. In the presence of excess cysteine, the *N*-mercapto species is transformed into a primary amide, forming cysteine persulfide in the process. Cysteine persulfide then reacts further with cysteine to generate cystine and free H<sub>2</sub>S. From a detailed SAR analysis, it was discovered that both electronic and steric effects at R<sub>1</sub> (but not R<sub>2</sub>) influence the rate of H<sub>2</sub>S release [105]. In general, NSHDs with smaller electron-withdrawing substituents at this position exhibited faster kinetics.



Figure 9. Thiol-triggered H<sub>2</sub>S release from NSHDs.

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 $H_2S$  can be released from acyl perthiols through an initial thioester exchange reaction that liberates a persulfide [58] (Figure 10). The ensuing hydropersulfide can then undergo an additional thiol exchange reaction to form a disulfide while generating  $H_2S$ .



Figure 10. Thiol-triggered H<sub>2</sub>S release from acyl perthiols.

Similarly, allyl thioesters liberate  $H_2S$  by undergoing an initial thioester exchange reaction to generate an allylic thiol, which then oxidizes to form a diallyl disulfide (Figure 11). Diallyl disulfides are known  $H_2S$  donors that are likely to operate through a hydropersulfide intermediate [112–118].



Figure 11. Thiol-triggered H<sub>2</sub>S release from allyl thioesters.

Within this series of donors, perthiocarbamates are unique in their ability to generate  $H_2S$  from two distinct pathways: hydropersulfide formation and carbonyl sulfide (COS) liberation [107]. As outlined in Figure 12, the COS delivery pathway is initiated by a thiol–disulfide exchange reaction that yields an unstable carbamic thioacid that quickly decomposes and gives rise to COS. In the presence of the ubiquitous enzyme carbonic anhydrase (CA), COS is quickly transformed into  $H_2S$  [119]. Alternatively, perthiocarbamates can liberate  $H_2S$  from a hydropersulfide intermediate that is generated from an intramolecular cyclization reaction that bypasses the need for a specific stimulus to trigger the event.

The mechanism of  $H_2S$  liberation from isothiocyanates has been carefully investigated by Lin and co-workers [120]. As delineated in Figure 13, they propose that the reaction commences with the nucleophilic attack by cysteine to form a dithiocarbamate. This intermediate then undergoes an intramolecular cyclization that forms a 5-membered ring and assists in the elimination of  $H_2S$ .

## 4.4. Enzyme-Triggered Donors

Hydrogen sulfide donors that are selectively responsive to specific enzymes have also been developed. Those that have displayed promising protective effects in MI/R injury models are featured below.



Figure 12. Thiol-triggered  $H_2S$  release from perthiocarbamates.



Figure 13. Thiol-triggered H<sub>2</sub>S release from isothiocyanates.

Esterase enzymes are omnipresent in human cells and, as their name implies, catalyze the hydrolysis of esters [121]. Not surprisingly, H<sub>2</sub>S liberation from a donor that is initiated by esterase-catalyzed hydrolysis is a common approach [122–127]. In general, the molecular framework is designed in such a way that upon ester hydrolysis, the resultant alcohol undergoes a self-immolative step that results in the eventual release of H<sub>2</sub>S. Donor P2 (Figure 14) illustrates this approach, as an unstable hydroxymethyl persulfide is unveiled after esterase-catalyzed hydrolysis [128]. This intermediate quickly decomposes to generate acetaldehyde and a hydropersulfide, which serves as an effective H<sub>2</sub>S precursor under biological conditions. Donor P2 was used in a murine model of MI/R injury and displayed promising protective effects with a bell-shaped therapeutic profile.



**Figure 14.** Esterase-triggered H<sub>2</sub>S release from P2, a donor with cardioprotective effects in MI/R injury models.

Donors selectively responsive to the enzyme  $\beta$ -galactosidase have also displayed favorable cardioprotective effects. The NO-H<sub>2</sub>S donor hybrid depicted in Figure 15 is an

example of such a design [129]. In the presence of  $\beta$ -galactosidase, the glycosidic bonds in the molecule are cleaved, producing an unstable intermediate that further unravels to liberate H<sub>2</sub>S (via COS hydrolysis) and nitric oxide (NO). To underline its cardioprotective effects, this hybrid prodrug was used in a rat model of heart failure. In general, it was shown that administration of the NO-H<sub>2</sub>S donor hybrid noticeably improved cardiac function post myocardial infarction, and especially in comparison to NO or H<sub>2</sub>S treatment alone, highlighting the effectiveness of a hybrid approach.



**Figure 15.**  $\beta$ -galactosidase-triggered H<sub>2</sub>S release from an NO-H<sub>2</sub>S donor hybrid, a compound with cardioprotective effects in MI/R injury models.

## 4.5. ROS-Triggered Donors

 $H_2S$  donors selectively responsive to elevated levels of ROS have been shown to be especially advantageous at combatting oxidative stress-related diseases [130–134], including MI/R injury. Within the structural framework of these donors, an *O*- or *S*-alkyl thiocarbamate is often linked to an aryl boronate ester, which serves as an ROS-responsive trigger [135–137]. In the presence of ROS (especially hydrogen peroxide or peroxynitrite), the aryl boronate ester is quickly oxidized to an unstable phenol that undergoes a 1,6-elimination to provide  $H_2S$  through carbonic anhydrase catalyzed COS hydrolysis (Figure 16).

An advantage of donors that proceed through the  $COS/H_2S$  pathway is their concurrent release of an aryl amine (or aryl alcohol) which affords an easy opportunity to access self-reporting donors that can track their  $H_2S$  delivery via fluorescence spectroscopy and other imaging techniques [134,138–140]. HSD-B and HSD-R (Figure 17) serve as examples of this, due to there being a latent fluorescent reporter embedded within their *O*-alkyl thiocarbamate framework. Moreover, these compounds were rationally designed to target mitochondria, thanks to their lipophilicity and cationic charge, which are likely to contribute to their pronounced cardioprotective effects that have been observed in H/R injury models [139,140]. HSD-B, for example, was shown to provide protection in a H9c2 cellular model of H/R injury, while HSD-R exhibited anti-apoptotic (inhibition of pro-apoptotic genes, including Bid, Apaf-1, and P53), anti-inflammatory, and pro-angiogenic effects in a rat MI/R injury model.



**Figure 16.** General mechanism for H<sub>2</sub>S release from ROS-triggered, *O*-alkyl thiocarbamate-based donors.



Figure 17. ROS-triggered donors with cardioprotective effects in H/R and MI/R injury models.

#### 5. Chemotherapy-Induced Cardiotoxicity

Chemotherapy-induced cardiotoxicity is a serious complication that affects the longterm survival of cancer patients and often manifests itself several years to several decades after the completion of treatment [141,142]. By convention, chemotherapy-induced cardiotoxicity is sorted into two distinct categories: type I, which is more severe, dose-dependent, and triggered by anthracycline-based drugs [143–145], and type II, which is less severe, believed to be reversible upon the cessation of treatment, and associated with cisplatin, alkylating agents, antimetabolites, and other non-anthracycline-based chemotherapeutics [146].

Anthracyclines, such as doxorubicin (DOX) and daunorubicin, are among the most effective anticancer agents in clinical use [147]. Their planar anthraquinone tetracyclic structure allows them to insert between DNA base pairs and interfere with the enzyme topoisomerase II, which, in turn, prevents the DNA unwinding and replication that ultimately induces apoptosis in proliferating cancer cells [148]. However, the same chemical features that give rise to DNA intercalation also predispose anthracyclines to redox cycling that generates superfluous levels of ROS within a cellular environment and, specifically, in mitochondria [149,150]. With increased mitochondrial density and a relatively deficient antioxidant defense system in place, cardiomyocytes are especially susceptible to oxidative injury [151,152]. Therefore, while other mechanisms may be in play, the uncontrolled production of ROS is believed to be primarily responsible for the dose-dependent, irreversible heart damage that is observed with anthracycline-based chemotherapeutics [153–155].

Given the significance of anthracyclines in the fight against cancer, it comes as no surprise that new therapeutic strategies are being extensively explored in an effort to diminish their cardiotoxic side effects. To this end, it has been suggested that the co-administration of  $H_2S$ —with its impressive antioxidative, anti-inflammatory, and anti-apoptotic effects—may offer an effective solution [156]. This hypothesis was first explored by Su and co-workers in 2009, using a DOX-treated rat model [157]. Employing NaHS as an  $H_2S$  donor, the attenuation of DOX-induced mitochondrial injury was in fact observed, along with significant improvements in overall cardiac function. Subsequent investigations

have corroborated these initial findings, and the beneficial effects of  $H_2S$  are now wellestablished for combatting chemotherapy-induced cardiotoxicity of both type 1 and type 2 through various mechanisms (Figure 18) [158–162].



Figure 18. Protective mechanisms of H<sub>2</sub>S against chemotherapy-induced cardiotoxicity.

While the co-administration of  $H_2S$  appears to be a promising approach for reducing the cardiotoxic profile of drug molecules, efforts to improve absorption and target delivery have led to the emergence of a new codrug design in which a known  $H_2S$ -donating moiety is directly linked to a chemotherapeutic agent of interest. This strategy is akin to the ABT-OH donor hybrids discussed earlier (Section 4.1) and has proven to be especially beneficial for mitigating DOX-induced cardiotoxicity, in particular. Therefore, given their obvious translational potential and clever chemical design, these hybrid DOX molecules are detailed below.

#### 6. H<sub>2</sub>S Conjugated Codrugs That Combat Anthracycline-Induced Cardiotoxicity

Chegaev and co-workers were the first to synthesize and assess a series of H<sub>2</sub>S-releasing, DOX hybrid codrugs (termed H<sub>2</sub>S-DOXOs) [163]. To accomplish this, they appended known H<sub>2</sub>S-donating motifs via an ester bond at C-14 of DOX. As seen in Figure 19, the affixed H<sub>2</sub>S-donating moieties included DTT derivative (H<sub>2</sub>S-DOXOs 10–13), allyl sulfide (H<sub>2</sub>S-DOXO 14), allyl disulfide (H<sub>2</sub>S-DOXO 15), and an aryl thioamide (H<sub>2</sub>S-DOXO 16).

After verifying  $H_2S$  liberation from  $H_2S$ -DOXOs in cell culture media, an LDH assay was used to assess their cytotoxic effects in H9c2 cardiomyocytes in culture. Compared to DOX,  $H_2S$ -DOXOs 10–14 were found to be significantly less cytotoxic, and the addition of the  $H_2S$  scavenger hydroxocobalamin confirmed that their release of hydrogen sulfide was responsible for their reduced cardiotoxicity.

Perhaps most notably, however,  $H_2S$ -DOXOs 10 and 11 simultaneously displayed impressive anticancer activity in human osteosarcoma cells (U-20S), even compared to the parent drug. Follow-up studies with  $H_2S$ -DOXO 10 indicated that the increased potency is likely to stem from their disruption of drug efflux by Pgp [164], which increases their cellular concentration. Thus, the appendage of an  $H_2S$  donor to DOX appears to impart several distinct advantages, including improved functional activity against multidrug-resistant cancers in addition to a reduced cardiotoxic profile.

Since this initial study, H<sub>2</sub>S-DOXO 10 (or Sdox) has undergone additional preclinical studies (Table 2) [165–167]. In a DOX-resistant prostate cancer mouse model, treatment with Sdox led to significantly reduced tumor volumes and improved safety. Conversely, DOX-treated mice exhibited reduced body weight and cardiotoxicity, which was assessed by measuring troponin plasma levels and left-ventricular-wall thickness.



**Figure 19.** H<sub>2</sub>S-donating, doxorubicin hybrid codrugs with protective effects against anthracyclineinduced cardiotoxicity.





In a similar fashion, Hu et al. recently reported on an  $H_2S$ -releasing, DOX hybrid codrug (c1, Figure 20) [168]. However, unlike  $H_2S$ -DOXOs, c1 is a prodrug that only liberates active DOX and  $H_2S$  under conditions of oxidative stress.



**Figure 20.** ROS-triggered H<sub>2</sub>S release from c1, an ROS-responsive DOX hybrid prodrug with reduced cardiotoxicity in rat cardiomyocytes in culture.

Elevated levels of ROS are found in most cancers for a variety of reasons [169]. Consequently, ROS-inducible anticancer prodrugs have emerged as a promising design strategy for improving the therapeutic index of anticancer chemotherapeutic agents [170–173]. Thus, the design of c1 represents a novel strategy that imparts both tumor-selective activation and H<sub>2</sub>S delivery in combination to further reduce the cardiotoxic side effects of DOX.

As highlighted in Figure 20, c1 utilizes an aryl boronate ester as an  $H_2O_2$ -selective trigger. Upon its oxidation by peroxide, the ensuing phenol undergoes a 1,6-elimination that releases both  $H_2S$  (by way of COS hydrolysis) and DOX. The authors confirmed this mechanism through LCMS studies and verified the selective release of both DOX and  $H_2S$  in response to  $H_2O_2$ .

The toxicity of c1 was assessed in rat cardiomyocytes in culture (Table 2). Using this model, c1 exhibited reduced cardiotoxicity compared to that of DOX. By enlisting an  $H_2O_2$ -activated DOX prodrug as a control, which provided  $CO_2$  release rather than COS, it was concluded that the protective effects of c1 are likely to stem from its co-release of  $H_2S$ . Cells treated with c1 also evinced significantly higher Nrf2 activation and heme oxygenase-1 expression compared to controls, providing a likely mechanism of cellular protection [174–177].

Notably, c1 also appeared to maintain the antitumor effects of DOX in a 4T1 mouse breast-cancer cell line. Therefore, while further preclinical profiling—especially in vivo—is required, the selective tumor activation and H<sub>2</sub>S liberation provided by c1 offer further promising options for overcoming DOX-derived cardiotoxicity in the clinic.

# 7. Conclusions

Once regarded as merely a toxic and foul-smelling gas,  $H_2S$  has more recently been recognized as a key signaling molecule and important endogenous mediator of numerous physiological and pathophysiological processes within mammalian systems. Its positive influence on the cardiovascular system, in particular, is rooted in its involvement in vasodilation (activation of  $K_{ATP}$  channels and the PI3K/Akt signaling pathway) [178–181], as well as its anti-inflammatory (inhibition of the p38 MAPK/NF- $\kappa$ B pathway) [182,183], antioxidative (activation of the Nrf2 signaling pathway) [59,168,184], and anti-apoptotic (suppression of pro-apoptotic genes Bid, Apaf-1, and p53) [140] properties, which have been extensively reviewed elsewhere in the literature [54,55,116,185].

Exogenous supplementation with H<sub>2</sub>S has been shown to vastly improve outcomes in various in vitro and in vivo cardiovascular disease models. In this review, its effectiveness at combating MI/R injury and chemotherapy-induced cardiotoxicity was explored, along with the fundamental chemistry and H<sub>2</sub>S releasing mechanism of the donor molecules that were utilized in these studies. The continued development and refinement of H<sub>2</sub>S-releasing compounds is critical to unlocking the translational therapeutic potential of hydrogen sulfide, by augmenting its delivery and bioavailability while better mimicking its natural and prolonged enzymatic production. Thus, the compounds reported on herein not only represent important investigative tools for probing the chemical biology of hydrogen sulfide but may also one day serve as important therapeutic agents for the treatment of MI/R injury and anthracycline-induced cardiotoxicity.

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