



Review

Peroxiredoxins as Potential Targets for Cardiovascular Disease

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Abstract: Increased oxidative stress (OS) is considered a common etiology in the pathogenesis of cardiovascular disease (CVD). Therefore, the precise regulation of reactive oxygen species (ROS) in cardiovascular cells is essential to maintain normal physiological functions. Numerous regulators of cellular homeostasis are reportedly influenced by ROS. Hydrogen peroxide (H_2O_2) , as an endogenous ROS in aerobic cells, is a toxic substance that can induce OS. However, many studies conducted over the past two decades have provided substantial evidence that H_2O_2 acts as a diffusible intracellular signaling messenger. Antioxidant enzymes, including superoxide dismutases, catalase, glutathione peroxidases, and peroxiredoxins (Prdxs), maintain the balance of ROS levels against augmentation of ROS production during the pathogenesis of CVD. Especially, Prdxs are regulatory sensors of transduced intracellular signals. The intracellular abundance of Prdxs that specifically react with H_2O_2 act as regulatory proteins. In this review, we focus on the role of Prdxs in the regulation of ROS-induced pathological changes in the development of CVD.

Keywords: antioxidant enzymes; aneurysm; atherosclerosis; cardiovascular diseases; hydrogen peroxide; oxidative stress; peroxiredoxins; reactive oxygen species



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1. Introduction

1.1. Oxidative Stress (OS) and Cardiovascular Disease (CVD)

CVD is caused by various heterogeneous pathophysiological mechanisms, although increased OS is considered a potential common etiology [1-3]. Several factors control fluctuations in the concentrations of reactive oxygen species (ROS), including the sources of ROS production and antioxidant enzymes, as the precise regulation of ROS is essential to maintain normal physiological function of cardiovascular cells [4–6]. The regulation of cell differentiation and growth, as well as intracellular signaling molecules, such as phosphatases and kinases, are reportedly influenced by ROS to control cellular homeostasis [7–12]. However, an overabundance of ROS due to an imbalance between the production and removal of ROS damages intracellular molecules, including DNA, lipids, and proteins, leading to cellular dysfunctions and cell death via necrosis or apoptosis [13,14]. In cells, mitochondria and enzyme systems involving nicotinamide adenine dinucleotide phosphate (NADPH) oxidases (NOXs) and xanthine oxidases are the main drivers of ROS production [15–18]. ROS constitute both nonradicals, including hydrogen peroxide (H₂O₂), hypochlorous acid, and ozone, and radicals, including superoxide, hydroxyl radicals, and peroxyl radicals [19]. Therefore, the actions of ROS differ according to the cell type, source, location, and concentration, which leads to a variety of physiological and pathophysiological functions [19].

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Atherosclerosis is a chronic vascular inflammatory disease associated with the development of CVD, which is a leading cause of mortality and morbidity worldwide [20]. Atherogenic stimuli, including dyslipidemia, hypertension, and smoking, lead to endothelial dysfunction and structural alterations to the arterial walls and trigger the accumulation of circulating apolipoprotein B (apoB)-containing lipoproteins, mainly low-density lipoproteins (LDLs), in the intima region of the arterial wall [21,22]. Structural alterations to the aorta expose subendothelial proteoglycans, which contribute to the binding of apoB to LDLs under the endothelial layer [22]. ROS modify LDL particles, such as oxidized LDLs (oxLDLs), in the intima, which induces the expression of adhesion molecules, the release of chemokines by endothelial cells (ECs), and the recruitment of immune cells into the intima [3,23]. Monocytes recruited into the intima are differentiated into macrophages, which engulf oxLDLs through scavenger receptors and create inflammatory conditions in lesions by expressing various proinflammatory cytokines and the production of ROS and costimulatory molecules that activate other immune cells [23,24]. The excess engulfment of oxLDLs triggers the transformation of macrophages into foam cells, resulting in apoptotic cell death. The apoptosis of foam cells leads to the accumulation of cellular debris and cholesterol crystals that contribute to the development of the necrotic core, thereby inducing plaque destabilization [25]. Plaque rupture or erosion causes occlusive thrombosis, leading to acute cardiovascular events, which underlie mortality due to atherosclerotic CVD [24,26]. A schematic overview of ROS-mediated pathophysiology in the development of atherosclerosis is presented in Figure 1.

Xanthine oxidase NADPH oxidase Mitochondria

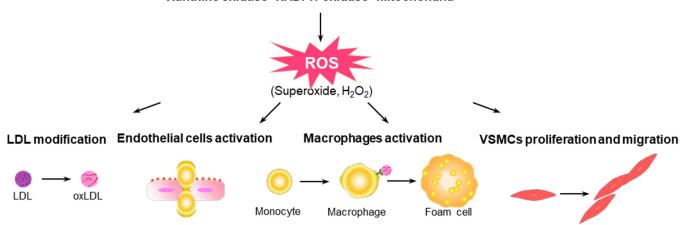


Figure 1. Potential roles of ROS in the progression of atherosclerosis. ROS modify LDL particles, such as oxLDLs, in the intima, which induce the expression of adhesion molecules, the release of chemokines from ECs, and the recruitment of immune cells into the intima. Monocytes recruited into the intima differentiate into macrophages that engulf oxLDLs through scavenger receptors, and macrophages create inflammatory conditions in lesions by expressing various proinflammatory cytokines, ROS, and costimulatory molecules that activate other immune cells. Excess engulfment of oxLDLs triggers the transformation of macrophages into foam cells. ROS is closely involved in the regulation of the growth, migration, extracellular matrix production, and inflammatory gene expression in VSMCs.

Numerous studies have demonstrated an important role of ROS in the development of atherosclerosis. NOX subunit gp91phox in macrophages and NOX4 in nonphagocytic vascular cells are upregulated in human atherosclerosis, and p47phox-deficient vascular smooth muscle cells (VSMCs) produce relatively lower levels of superoxide and undergo growth factor-induced proliferation [27,28]. NOX2-mediated superoxide production reduces the bioavailability of nitric oxide (NO) and augments the development of early atherosclerotic plaque formation in apolipoprotein E knockout (*ApoE*-/-) mice [29]. The deficiency of glutathione peroxidase (Gpx)-1 exacerbates atherosclerosis and loss of heme oxygenase-1 (HO-1) accelerates the development of atherosclerosis and vascular remodeling in *ApoE*-/- mice [30,31].

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The prevalence of abdominal aortic aneurysm (AAA) ranges from 2% to 8% in men older than 65 years and the mortality rate associated with ruptured AAA varies between 85% and 90% [20,32]. The development of AAA is associated with degraded plasticity of the tunica media with the activation of various proteases and inflammation caused by the accumulation of immune cells, angiogenesis, and necrosis [33]. ROS-induced OS is closely associated with inflammatory processes and the activation of matrix metalloproteinases (MMPs), which promote proteolytic degradation of structural proteins in the pathogenesis of AAA [34]. A schematic overview of ROS-mediated pathogenesis of AAA is presented in Figure 2.

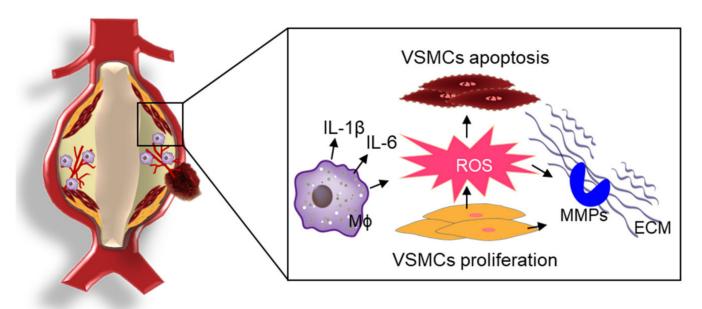


Figure 2. ROS-mediated the pathophysiology of AAA. In the development of AAA, ROS-induced OS is closely associated with inflammatory processes, the proliferation/migration/apoptosis of VSMCs, and the activation of MMPs for proteolytic degradation of structural proteins in the pathogenesis of AAA.

The superoxide levels and lipid peroxidation are increased and the expression of the NOX subunits p47phox and p22phox are upregulated in the aneurysmal aorta as compared with the normal aorta of human patients [35]. Inducible NO synthase in human patients with AAA, but not normal human abdominal aortas, is identified by in situ hybridization and immunohistochemical analyses [36]. The deficiency of p47phox in *ApoE*-/- mice attenuates angiotensin II (Ang II)-induced AAA formation with decreased OS, inflammation, and MMP-2 activity [37]. NOX1 deficiency reportedly prevents Ang II-induced aortic dissection with an increased expression of the tissue inhibitor of metalloproteinase-1 in mice as compared with the wild-type controls [38].

1.2. ROS-Mediated Pathophysiological Changes in Vascular Cells

OS and inflammation mainly induce dysfunction of ECs, resulting in the augmentation of vasoconstrictor and prothrombotic factors [39]. ECs modulate the vascular tone, cellular adhesion, thrombosis, smooth muscle cell proliferation, and vessel inflammation through the production of NO, prostaglandins, hyperpolarization factors, and contracting factors [40–42]. Therefore, the dysfunction of ECs is closely associated with an increased risk for the development of CVD and other diseases [39,43]. Endothelial nitric oxide synthase (NOS)-derived NO controls the contraction of vascular smooth muscle cells through the regulation of soluble guanylyl cyclase and cyclic guanosine monophosphate and inhibits leukocyte adhesion and platelet aggregation [42]. Cardiovascular risk factors, such as smoking, hypercholesterolemia, hypertension, and hyperglycemia, activate oxidative enzyme systems, including NOXs and uncoupled NOS, leading to the inactivation of

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NO by elevating superoxide levels [39,44]. The inactivation of NO induces an imbalance between EC-derived vasodilators and vasoconstrictors, resulting in endothelial dysfunction [42]. Inflammation also contributes to OS-mediated endothelial dysfunction in several human diseases, including CVD [45,46]. Proinflammatory stimuli, including oxLDL, tumor necrosis factor-alpha (TNF-α), interleukin (IL)-1β), IL-6, and monocyte chemoattractant protein-1 (MCP-1), increase the production of ROS and induce endothelial dysfunction through a variety of signaling pathways involved in activation of the nuclear factor-kappa B (NF-κB) and mitogen-activated protein kinase (MAPK) pathways [45,47–49]. The TNF- α induced expression of vascular adhesion molecule-1 and intracellular adhesion molecule-1 in aortic ECs is inhibited by antioxidants and inhibitors of NF-κB and MAPK [50–52]. Inhibition of the oxidation of LDLs increases the expression of MCP-1 and adhesion molecules in ECs through the ROS- and NF-kB-dependent activation of lectin-like oxLDL receptor-1 [53,54]. In addition, OS and proinflammatory stimuli induce the apoptosis of ECs, resulting in atherogenesis and thrombosis, which are inhibited by superoxide dismutase (SOD), catalase, N-acetylcysteine (NAC), and vitamins [55,56]. Angiogenesis is also involved in the pathology of atherogenesis [57]. ROS regulates the migration, proliferation, and tubulogenesis of ECs by controlling intracellular signaling and angiogenic growth factor expression [43,57]. Taken together, these observations suggest that ROS is a critical factor for dysfunction and pathological signaling pathways in ECs in CVD (Figure 3).

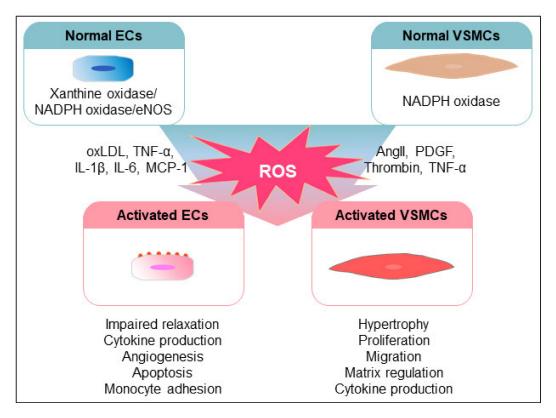


Figure 3. Modulation of the vascular cells by ROS. Various extracellular stimuli increase the production of ROS through ROS-generated enzyme systems in vascular cells. oxLDLs and proinflammatory cytokines induce the augmentation of ROS, which leads to the dysfunction of ECs, including impaired vasorelaxation, cytokine production, angiogenesis, apoptosis, and immune cell adhesion. Additionally, VSMCs are activated by various stimuli, such as AngII, PDGF, thrombin, and TNF- α , leading to alterations of the cellular functions, including proliferation, migration, extracellular matrix formation, and cytokine production, through increased ROS production.

ROS is closely involved in the regulation of the growth, migration, extracellular matrix production, and inflammatory gene expression in VSMCs [6]. The switching of VSMCs from the "contractile phenotype" to the "synthetic phenotype" is common in hyperten-

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sion, atherosclerosis, aortic aneurysm, and restenosis after balloon angioplasty [58,59]. Phenotypic switching is a prerequisite for the proliferation and migration of VSMCs in the pathogenesis of CVD. Various factors, such as Ang II, platelet-derived growth factor (PDGF), and thrombin, induce ROS-dependent proliferation and the migration of VSMCs. Ang II induces the hypertrophy of VSMCs through the generation of ROS, which is inhibited by catalase and antisense p22phox [60,61]. PDGF and thrombin also induce the H₂O₂-dependent proliferation of VSMCs, which is inhibited by catalase, NAC, or a treatment with diphenyleneiodonium chloride [62,63]. The overexpression of catalase and peroxiredoxin (Prdx) 2 and inhibition of antioxidant production inhibit the PDGF-induced migration of VSMCs, implying that the PDGF-induced migration of VSMCs is dependent on ROS generation [11,62,64]. In addition, the production of the extracellular matrix and activation of inflammatory genes in VSMCs are regulated in a ROS-dependent manner. MMPs secreted from VSMCs are activated by ROS, and the NF-κB-mediated expression of inflammatory genes in VSMC is ROS-sensitive. The Ang II-induced expression of MCP-1 and IL-6 in VSMCs is also regulated in a ROS-dependent manner [65,66]. In addition, the TNF-α-induced MCP-1 expression in VSMCs is considered a redox-sensitive pathway [67]. Therefore, ROS-mediated alterations of the functions of VSMCs contribute to the progression of CVD (Figure 3).

Several antioxidant enzymes, including catalase, Gpxs, Prdxs, and SODs, are responsible for maintaining ROS levels against augmentation of ROS production during the pathogenesis of CVD. In this review, we focus on the role of Prdxs on the regulation of ROS-induced pathological changes in the development of CVD.

2. Prdxs in CVD

2.1. Overview of Prdxs

Prdxs, which consist of six Prdx isoforms (Prdx1-6) in mammalian cells, are conserved abundant cysteine (Cys)-based peroxidases involved in various biological functions [68,69]. Prdx1, 2, and 6 are localized in the cytosol and nucleus, while Prdx3 is restricted to the mitochondria, and Prdx5 is localized in the cytosol, mitochondria, and peroxisomes [70]. Prdxs are classified into typical 2-Cys, atypical 2-Cys, and 1-Cys Prdx subfamilies that possess diverse amino acid sequences, although all Prdxs have one conserved cysteine residue as the peroxidatic cysteine (C_P) [68,71,72]. Typical 2-Cys Prdxs (Prdx1–4) contain two conserved cysteine residues (C_P and a resolving Cys, C_R) and consist of a homodimer formed via a disulfide bond between C_P and C_R in the chain of another subunit [68,72]. Atypical 2-Cys Prdx (Prdx5) contains an intrasubunit disulfide bond between C_P and C_R in the identical subunit, which is distinct from typical 2-Cys Prdxs [68,72]. In the 1-Cys Prdx (Prdx6), C_P forms a disulfide bond with the C_R of another protein or small thiol molecules because of the lack of C_R in the 1-Cys Prdx [68,72]. A schematic of the catalytic mechanisms of Prdx isoforms is presented in Figure 4. By surrounding residues in the active site of Prdxs, the catalytic reactivity (rate constant, 10⁶–10⁸ M⁻¹s⁻¹) of C_P for H₂O₂ is markedly augmented by up to one million-fold greater than other thiol proteins [73]. H₂O₂, as an endogenous form of ROS in aerobic cells, is considered a toxic substance that induces OS. However, many studies conducted over the past two decades have provided substantial evidence that H₂O₂ acts as a diffusible intracellular signaling messenger when it is transiently produced through the activation of cell surface receptors [74,75]. Prdxs rapidly react with H₂O₂, peroxynitrite, and other hydroperoxides but not with other electrophiles, indicating the target-specific reactivity of Prdxs [73,76]. Intracellular Prdxs that specifically react with H₂O₂ act as regulatory sensors during the transduction of intracellular signals [68,72].

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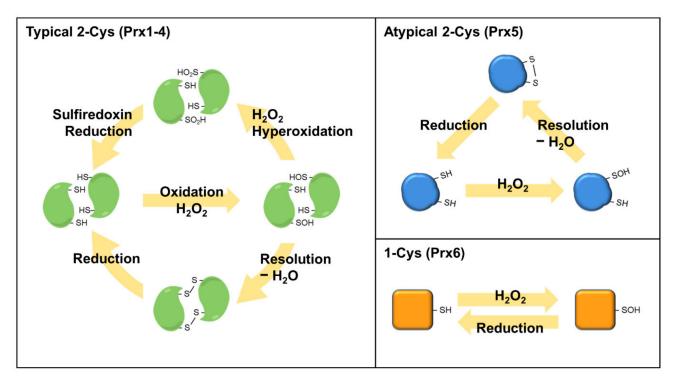


Figure 4. Reaction mechanisms of Prdx isoforms. The peroxidatic cysteine residue of Prdxs reacts with H_2O_2 to release H_2O_3 and form a sulfenic acid (SOH) intermediate. SOH is resolved with a second monomer to form an intermolecular disulfide bond in typical 2-Cys Prdxs (Prdx1–4). Atypical 2-Cys Prdx (Prdx5) forms an intra-subunit disulfide in the identical subunit, which is distinct from typical 2-Cys Prdxs. The 1-Cys Prdx (Prdx6) react with H_2O_2 to form a SOH intermediate, which forms a disulfide bond with other proteins or small thiol molecules to reduce the peroxidatic cysteine.

2.2. Prdx1

Prdx1 is a typical 2-Cys Prdx that is expressed in the nucleus and cytosol of several cell types. Recent studies have elucidated various functional roles of Prdx1 in CVD. One found that laminar shear stress upregulates Prdx1, which functionally decreases the shear stress-dependent H₂O₂ levels and maintains the redox balance in ECs [77]. It is well-known that endogenous H₂O₂ formation induces the release of cytokines by activated ECs and inflammation, leading to atherosclerosis [78]. By scavenging H₂O₂, Prdx1 suppresses the excessive activation of ECs and consequent vascular inflammation, while Prdx1 deficiency induces early atherosclerosis in ApoE^{-/-} mice [79]. Moreover, another report suggested that lipophagy, the selective autophagic degradation of lipid in macrophages, is another possible protective role of Prdx1 in atherosclerosis [80]. Among the various antioxidant enzymes in macrophages, Prdx1 is highly expressed and is a major contributor to the maintenance of lipophagic flux and cholesterol homeostasis by regulating excessive H₂O₂ in macrophages and reducing foam cell formation and atherosclerosis [80]. While the mechanism underlying the role of Prdx1 in AAA is unknown, human studies have shown that the serum levels of Prdx1, which are increased in AAA patients [81], are positively correlated with the biomarkers plasmin-antiplasmin and myeloperoxidase and the diameter of AAA, suggesting that Prdx1 may be used as biomarker of AAA [81]. The overexpression of Prdx1 in cardiomyocytes of mice prevents transverse aortic constriction (TAC)-induced cardiac hypertrophy and heart failure, with decreased pressure overload-induced cardiac inflammation and OS via activation of the nuclear factor-erythroid 2-related factor 2/HO-1 signaling pathway [82]. In addition, the cardiomyocyte-specific expression of Prdx1 prevents doxorubicin-induced cardiotoxicity by reducing the OS and apoptosis [83]. Collectively, these results suggest that Prdx1 would be a useful target for the treatment of CVD.

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2.3. Prdx2

Prdx2, a 2-Cys Prdx, is abundantly expressed in most cells and tissues. Although Prdx2 shares most of the features of typical 2-Cys Prdx with Prdx1, the expression patterns of Prdx2 in vascular cells appear to be relevant to its role in CVD. Prdx2 can suppress PDGF signaling by inhibiting protein tyrosine phosphatase in VSMCs and protects against PDGF-dependent neointimal thickening, which is a key feature of vascular remodeling [11]. The migration and proliferation of VSMCs during atherogenesis contribute to fibrous plaque formation, and PDGF is a strong growth factor that stimulates these pathologic processes [84]. The current evidence suggests that Prdx2 has a beneficial effect in CVD. One report demonstrated that Prdx2 is highly expressed in both vascular and immune cells, while Prdx2 deficiency accelerates atherosclerotic plaque formation and immune cell infiltration due to plaque progression, as compared with other types of antioxidant enzymes, such as Gpx1 and catalase [52]. Prdx2 expression is increased in the aneurysmal aorta of both human AAA patients and a mouse model of AAA, but the expression is relatively low in healthy humans and normal control mice, suggesting that Prdx2 is a potential biomarker of the progression of AAA [85]. In addition, Prdx2 deficiency induces the progression of AAA and most of the features of AAA tissue, including the OS, immune cell infiltration, MMP activation, elastin degradation, and aortic dilation [85]. Furthermore, Prdx2 negatively regulates H₂O₂ generation and thrombosis formation by platelets and VSMCs [86,87]. These findings suggest that Prdx2 may serve as a potential target for the treatment of CVD.

2.4. Prdx3

Prdx3 is a typical 2-Cys Prdx isoform that is exclusively localized in the mitochondria and contributes to the elimination of intracellular ROS, which is generated as a byproduct during energy production and cellular respiration. Mitochondria play central roles in various metabolic processes, including the production of cellular ATP through oxidative phosphorylation and the regulation of the intrinsic apoptosis pathway [88]. The increased mitochondrial production of ROS leads to mitochondrial DNA damage and mitochondrial dysfunction. To date, several kinds of intracellular antioxidant enzymes have been identified, especially Prdx3, which is a major antioxidant scavenger that can remove almost 90% of mitochondrial H_2O_2 to maintain mitochondrial homeostasis [89,90].

Prdx3 has protective roles in various cell types and ameliorates inflammation, which is highly associated with the pathogenesis of CVD. A recent study reported that the overexpression of Prdx3 protects the heart against left ventricular remodeling and failure after myocardial infarction (MI) [91]. The hallmarks of MI, such as reduced left ventricular cavity dilatation and dysfunction, myocyte hypertrophy, interstitial fibrosis, and apoptosis of the non-infarcted myocardium, are reduced in mice overexpressing Prdx3 [91]. Further, the redox state of Prdx3 significantly changes during ischemia-reperfusion heart injury [92]. The beneficial effects of Prdx-3 include maintaining cardiac function by the attenuation of the mitochondrial OS [92]. Although several studies have reported the protective roles of Prdx isoforms against atherosclerosis, the role of Prdx3 in the pathogenesis of atherosclerosis remains unclear. However, a possible mechanism of Prdx3 in atherosclerosis may be associated with mitophagy [93,94]. Mitophagy is a critical mechanism of cells to maintain metabolic homeostasis by eliminating damaged or long-lived mitochondria via autophagic degradation and has been linked with the pathogenesis of numerous human diseases, including cancers, neurodegenerative disorders, and CVD [95,96]. Although alterations to the expression of Prdx3 in mitophagy appear to be relevant, current data are insufficient to identify potential mechanisms in atherosclerosis. Nonetheless, recent studies have implicated potential roles of Prdx3 in various cell types associated with atherosclerosis [97,98].

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2.5. Prdx4

Prdx4 is localized in the endoplasmic reticulum (ER), which is an important metabolic organelle involved in lipid and protein synthesis and transport by regulating the correct folding of proteins. The ER redox environment dictates the fate of entering proteins and the level of redox signaling mediators modulates the level of the ROS. Impairment of the ER leads to the cellular accumulation of ROS, and increased ER stress by ROS can induce apoptosis. Therefore, Prdx4 catabolizes H₂O₂ within the ER to provide a cytoprotective effect against the OS, acceleration of proper protein folding, and prevention of ER stress [99–102]. Prdx4 is a secretable peroxidase and, thus, a potential candidate biomarker. Prdx4 has recently been identified in the circulation of both healthy individuals and patients with peripheral artery disease (PAD) [103,104]. The overexpression of human Prdx4 in ApoE^{-/-} mice highly attenuated the development of atherosclerosis by limiting the infiltration of T-lymphocytes, reducing the OS, and ameliorating necrosis [105]. In ApoE^{-/-} mice with transplanted bone marrow, hematopoietic Prdx4 overexpression was sufficient to suppress the progression of atherosclerosis [105]. Therefore, Prdx4 is a potential target for treatment of CVD in the context of ER stress-induced apoptosis in the progression of atherosclerotic plaque formation [106].

2.6. Prdx5

Among the six Prdx isoforms, Prdx5 is the most recently discovered. Prdx5 is an atypical 2-Cys Prdx in mammals and is widely localized to the mitochondria, peroxisomes, cytosol, and nucleus. Although Prdx5 has unique features, such as localization in peroxisomes and expression as a short form and long form, which has a mitochondria targeting sequence, it has been studied less than other Prdxs [107]. Recent studies have found that Prdx5 prevents various pathological conditions by regulating the OS [108]. Prdx5 deficiency induces a susceptibility to high-fat diet-induced obesity and associated metabolic disorders [109]. Moreover, the depletion of Prdx5 significantly augmented kidney inflammation after renal ischemia-reperfusion injury [110]. Notably, extracellular Prdx5 is the strongest contributor among all Prdx isoforms to induce the expression of inflammatory cytokines through Toll-like receptor (TLR) 2 and TLR4 after a postischemic brain injury via damage-associated molecular patterns [111]. Additionally, Prdx5 can specifically bind with TLR4, and this interaction can modulate the release of proinflammatory cytokines and cell stiffening, which could be highly relevant in the inflammatory response [112]. However, several studies have indicated that intracellular Prdx5 is involved in the inhibition of inflammatory responses [113–116]. Prdx5 is constitutively expressed in human cartilage and is upregulated in osteoarthritic cartilage, suggesting that Prdx5 may have a protective function against OS in human cartilage [113]. In primary macrophages, Prdx5 expression is induced by inflammatory stimuli, such as lipopolysaccharides and interferon- γ , suggesting that Prdx5 plays a defensive role in activated macrophages [114]. However, further studies are needed to clarify the role of Prdx5 in the pathogenesis of CVD.

2.7. Prdx6

Prdx6 has a unique 1-Cys structure but similar functional and structural properties with other Prdx isoforms. Prdx6 has peroxidase activity via a single conserved Cys residue, short-chain organic fatty acids, and phospholipid hydroperoxides by using glutathione instead of thioredoxin as a physiological reductant. Moreover, Prdx6 is a bifunctional enzyme with glutathione peroxidase and phospholipase A2 (PLA2) activities, which characteristically responds to various extracellular stimuli [117,118]. Several recent studies have provided evidence of the role of Prdx6 in CVD. As a possible link between Prdx6 and atherosclerosis, as mentioned above, Prdx6 possesses PLA2 activity. Previous studies have demonstrated that members of the PLA2 family generate various active lipid metabolites that promote OS and inflammatory cytokine production, leading to the acceleration of inflammatory metabolic diseases, such as atherosclerosis and hyperlipidemia [119]. Additionally, Prdx6 has an important role in heart recovery following ischemia–reperfusion injury

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and can protect against phospholipid peroxidation-mediated membrane damage [120,121]. Prdx6-deficient mice are more susceptible to ischemia–reperfusion injury, with a reduced recovery of left ventricular function, increased MI area, and augmented apoptotic cell death [120]. The overexpression of Prdx6 in ECs prevents AngII-induced inflammation, OS, and endothelial dysfunction via inactivation of the p38 MAPK and c-Jun N-terminal kinase pathways [122]. In addition, a significant increase in plasma levels of Prdx6 has been positively correlated with endothelial dysfunction in diabetic patients with PAD [104]. Although Prdx6 has been extensively studied, more detailed studies are necessary to elucidate the role of Prdx6 in the pathogenesis of atherosclerosis. Prdx6 is reported to play a minor athero-protective role in mice with a mixed B6; 129 background [123]. Prdx6 deficiency induces higher plasma levels of lipid hydroperoxides and the oxidation of LDLs, whereas Prdx6 in transgenic mice was insufficient to protect against atherogenesis [123,124].

3. Conclusions and Future Perspectives

OS, which is a common feature of CVD, is thought to occur by an imbalance in the production and removal of ROS in cardiovascular cells. Various experimental studies of dietary antioxidants and antioxidant enzymes to block the abundance of OS in CVD revealed an efficient inverse correlation. However, dietary antioxidant therapies for patients with CVD have failed in many randomized trials. To overcome the limitation of dietary antioxidants, the development of antioxidant enzymes, such as Prdxs, is a potential therapeutic strategy against CVD. Prdxs are a family of abundant thiol-dependent peroxidases, consisting of six isoforms in mammalian cells. As sensor proteins, Prdxs are appropriate antioxidant enzymes for the regulation of OS and H₂O₂-mediated intracellular signaling. Various experimental studies have confirmed the potential of Prdxs as a therapeutic strategy against CVD (Figure 5). Therefore, the development of derivatives or mimetics of the catalytic activity of Prdxs offers great promise for antioxidant therapy in CVD. The evolution of biotechnologies in the pharmaceutical industry could provide unique derivatives or mimetics for each Prdx isoform to develop precise medications matched to the pathophysiology of CVD.

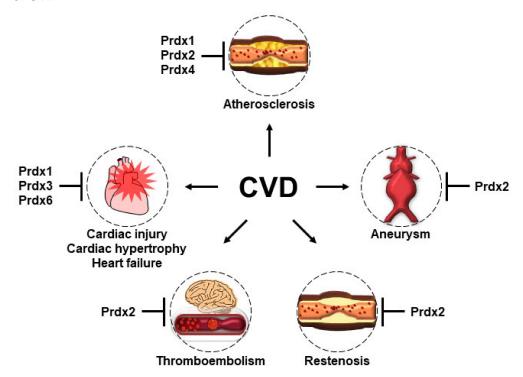


Figure 5. The protective roles of Prdxs in the pathogenesis of CVD. Prdx 1, 2, and 4 play protective roles in the development of atherosclerosis, although through different mechanisms. Prdx1, which is

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highly expressed in macrophages, maintains lipophagic flux and cholesterol homeostasis and reduces the foam cell formation and atherosclerosis. Prdx2 deficiency accelerates atherosclerotic plaque formation and immune cell infiltration. The overexpression of human Prdx4 attenuates the development of atherosclerosis by limiting the infiltration of T-lymphocytes, reducing OS, and ameliorating necrosis. Prdx2 deficiency promotes the progression and most features of AAA. Prdx2 suppresses PDGF signaling by inhibiting the protein tyrosine phosphatase in VSMCs and protects against PDGF-dependent neointimal thickening. Prdx2 negatively regulates H₂O₂ generation and thrombosis formation by platelets and VSMCs. The overexpression of Prdx1 in the cardiomyocytes of mice prevents TAC-induced cardiac hypertrophy and heart failure. The overexpression of Prdx3 protects the heart against left ventricular remodeling and failure after MI. Prdx6-deficient mice have increased susceptibility to ischemia–reperfusion injury.

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Abbreviations

EC Endothelial cellOS Oxidative stressCVD Cardiovascular diseaseROS Reactive oxygen species

NADPH Nicotinamide adenine dinucleotide phosphate

NOX NADPH oxidase
ApoB Apolipoprotein B
LDL Low-density lipoprotein

oxLDL oxidized LDL

VSMCs Vascular smooth muscle cells

NO Nitric oxide

ApoE^{-/-} Apolipoprotein E knockout Gpx Glutathione peroxidase HO-1 Heme oxygenase-1

AAA Abdominal aortic aneurysm MMPs Matrix metalloproteinases

Ang II Angiotensin II

TNF- α Tumor necrosis factor-alpha

IL Interleukin

MCP-1 Monocyte chemoattractant protein-1

NF-κB Nuclear factor-kappa B

MAPK Mitogen-activated protein kinase

SOD Superoxide dismutase NAC N-acetylcysteine

PDGF Platelet-derived growth factor

Prdx Peroxiredoxin

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Cys Cysteine

C_P Peroxidatic cysteine

C_R Resolving Cys

TAC Transverse aortic constriction

MI Myocardial infarction ER Endoplasmic reticulum PAD Peripheral artery disease

TLR Toll-like receptor PLA2 Phospholipase A2

References

1. Tsutsui, H.; Kinugawa, S.; Matsushima, S. Oxidative stress and heart failure. *Am. J. Physiol. Heart Circ. Physiol.* **2011**, 301, H2181–H2190. [CrossRef]

- 2. Baradaran, A.; Nasri, H.; Rafieian-Kopaei, M. Oxidative stress and hypertension: Possibility of hypertension therapy with antioxidants. *J. Res. Med. Sci.* **2014**, *19*, 358–367. [PubMed]
- 3. Kattoor, A.J.; Pothineni, N.V.K.; Palagiri, D.; Mehta, J.L. Oxidative stress in atherosclerosis. *Curr. Atheroscler. Rep.* **2017**, *19*, 42. [CrossRef]
- 4. Jones, D.P. Redefining oxidative stress. Antioxid. Redox Signal. 2006, 8, 1865–1879. [CrossRef]
- 5. Cervantes Gracia, K.; Llanas-Cornejo, D.; Husi, H. Cvd and oxidative stress. J. Clin. Med. 2017, 6, 22. [CrossRef]
- 6. Taniyama, Y.; Griendling, K.K. Reactive oxygen species in the vasculature: Molecular and cellular mechanisms. *Hypertension* **2003**, *42*, 1075–1081. [CrossRef]
- Mittal, C.K.; Murad, F. Activation of guanylate cyclase by superoxide dismutase and hydroxyl radical: A physiological regulator of guanosine 3',5'-monophosphate formation. Proc. Natl. Acad. Sci. USA 1977, 74, 4360–4364. [CrossRef]
- 8. Olguin-Albuerne, M.; Moran, J. Ros produced by nox2 control in vitro development of cerebellar granule neurons development. *ASN Neuro*. **2015**, *7*, 1759091415578712. [CrossRef] [PubMed]
- 9. Mandal, D.; Fu, P.; Levine, A.D. Redox regulation of il-13 signaling in intestinal epithelial cells: Usage of alternate pathways mediates distinct gene expression patterns. *Cell Signal.* **2010**, *22*, 1485–1494. [CrossRef] [PubMed]
- 10. Abimannan, T.; Peroumal, D.; Parida, J.R.; Barik, P.K.; Padhan, P.; Devadas, S. Oxidative stress modulates the cytokine response of differentiated th17 and th1 cells. *Free Radic. Biol. Med.* **2016**, *99*, 352–363. [CrossRef] [PubMed]
- 11. Choi, M.H.; Lee, I.K.; Kim, G.W.; Kim, B.U.; Han, Y.H.; Yu, D.Y.; Park, H.S.; Kim, K.Y.; Lee, J.S.; Choi, C.; et al. Regulation of pdgf signalling and vascular remodelling by peroxiredoxin ii. *Nature* **2005**, 435, 347–353.
- 12. Fujino, G.; Noguchi, T.; Matsuzawa, A.; Yamauchi, S.; Saitoh, M.; Takeda, K.; Ichijo, H. Thioredoxin and traf family proteins regulate reactive oxygen species-dependent activation of ask1 through reciprocal modulation of the n-terminal homophilic interaction of ask1. *Mol. Cell Biol.* **2007**, *27*, 8152–8163. [CrossRef]
- 13. Sies, H. Oxidative stress: A concept in redox biology and medicine. Redox Biol. 2015, 4, 180–183. [CrossRef] [PubMed]
- 14. Di Meo, S.; Reed, T.T.; Venditti, P.; Victor, V.M. Role of ros and rns sources in physiological and pathological conditions. *Oxid. Med. Cell Longev.* **2016**, 2016, 1245049. [CrossRef] [PubMed]
- 15. Dan Dunn, J.; Alvarez, L.A.; Zhang, X.; Soldati, T. Reactive oxygen species and mitochondria: A nexus of cellular homeostasis. *Redox Biol.* **2015**, *6*, 472–485. [CrossRef] [PubMed]
- 16. Konior, A.; Schramm, A.; Czesnikiewicz-Guzik, M.; Guzik, T.J. Nadph oxidases in vascular pathology. *Antioxid. Redox Signal.* **2014**, 20, 2794–2814. [CrossRef]
- 17. Amanso, A.M.; Griendling, K.K. Differential roles of nadph oxidases in vascular physiology and pathophysiology. *Front. BioSci.* **2012**, *4*, 1044–1064.
- 18. Nishino, T.; Okamoto, K.; Eger, B.T.; Pai, E.F.; Nishino, T. Mammalian xanthine oxidoreductase—Mechanism of transition from xanthine dehydrogenase to xanthine oxidase. *FEBS J.* **2008**, 275, 3278–3289. [CrossRef]
- 19. Holmstrom, K.M.; Finkel, T. Cellular mechanisms and physiological consequences of redox-dependent signalling. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 411–421.
- 20. Benjamin, E.J.; Virani, S.S.; Callaway, C.W.; Chamberlain, A.M.; Chang, A.R.; Cheng, S.; Chiuve, S.E.; Cushman, M.; Delling, F.N.; Deo, R.; et al. Heart disease and stroke statistics-2018 update: A report from the american heart association. *Circulation* **2018**, *137*, e67–e492. [CrossRef] [PubMed]
- 21. Maiolino, G.; Rossitto, G.; Caielli, P.; Bisogni, V.; Rossi, G.P.; Calo, L.A. The role of oxidized low-density lipoproteins in atherosclerosis: The myths and the facts. *Mediat. Inflamm.* **2013**, 2013, 714653. [CrossRef]
- 22. Kwon, G.P.; Schroeder, J.L.; Amar, M.J.; Remaley, A.T.; Balaban, R.S. Contribution of macromolecular structure to the retention of low-density lipoprotein at arterial branch points. *Circulation* **2008**, *117*, 2919–2927. [CrossRef] [PubMed]
- 23. Weber, C.; Noels, H. Atherosclerosis: Current pathogenesis and therapeutic options. Nat. Med. 2011, 17, 1410–1422. [CrossRef]
- 24. Sakakura, K.; Nakano, M.; Otsuka, F.; Ladich, E.; Kolodgie, F.D.; Virmani, R. Pathophysiology of atherosclerosis plaque progression. *Heart Lung Circ.* **2013**, 22, 399–411. [CrossRef]

Antioxidants 2021, 10, 1244 12 of 15

25. Yla-Herttuala, S.; Bentzon, J.F.; Daemen, M.; Falk, E.; Garcia-Garcia, H.M.; Herrmann, J.; Hoefer, I.; Jukema, J.W.; Krams, R.; Kwak, B.R.; et al. Stabilisation of atherosclerotic plaques. Position paper of the european society of cardiology (esc) working group on atherosclerosis and vascular biology. *Thromb. Haemost.* **2011**, *106*, 1–19.

- 26. Sugiyama, S.; Okada, Y.; Sukhova, G.K.; Virmani, R.; Heinecke, J.W.; Libby, P. Macrophage myeloperoxidase regulation by granulocyte macrophage colony-stimulating factor in human atherosclerosis and implications in acute coronary syndromes. *Am. J. Pathol.* **2001**, *158*, 879–891. [CrossRef]
- 27. Sorescu, D.; Weiss, D.; Lassegue, B.; Clempus, R.E.; Szocs, K.; Sorescu, G.P.; Valppu, L.; Quinn, M.T.; Lambeth, J.D.; Vega, J.D.; et al. Superoxide production and expression of nox family proteins in human atherosclerosis. *Circulation* **2002**, *105*, 1429–1435. [CrossRef]
- 28. Barry-Lane, P.A.; Patterson, C.; van der Merwe, M.; Hu, Z.; Holland, S.M.; Yeh, E.T.; Runge, M.S. P47phox is required for atherosclerotic lesion progression in apoe(-/-) mice. *J. Clin. Investig.* **2001**, *108*, 1513–1522. [CrossRef]
- 29. Judkins, C.P.; Diep, H.; Broughton, B.R.; Mast, A.E.; Hooker, E.U.; Miller, A.A.; Selemidis, S.; Dusting, G.J.; Sobey, C.G.; Drummond, G.R. Direct evidence of a role for nox2 in superoxide production, reduced nitric oxide bioavailability, and early atherosclerotic plaque formation in apoe-/- mice. *Am. J. Physiol. Heart Circ. Physiol.* 2010, 298, H24–H32. [CrossRef] [PubMed]
- 30. Torzewski, M.; Ochsenhirt, V.; Kleschyov, A.L.; Oelze, M.; Daiber, A.; Li, H.; Rossmann, H.; Tsimikas, S.; Reifenberg, K.; Cheng, F.; et al. Deficiency of glutathione peroxidase-1 accelerates the progression of atherosclerosis in apolipoprotein e-deficient mice. *Arterioscler. Thromb. Vasc. Biol.* **2007**, 27, 850–857. [CrossRef]
- 31. Yet, S.F.; Layne, M.D.; Liu, X.; Chen, Y.H.; Ith, B.; Sibinga, N.E.; Perrella, M.A. Absence of heme oxygenase-1 exacerbates atherosclerotic lesion formation and vascular remodeling. *FASEB J.* **2003**, *17*, 1759–1761. [CrossRef] [PubMed]
- 32. Kent, K.C. Clinical practice. Abdominal aortic aneurysms. N. Engl. J. Med. 2014, 371, 2101–2108. [CrossRef]
- 33. Sakalihasan, N.; Limet, R.; Defawe, O.D. Abdominal aortic aneurysm. Lancet 2005, 365, 1577-1589. [CrossRef]
- 34. Freestone, T.; Turner, R.J.; Coady, A.; Higman, D.J.; Greenhalgh, R.M.; Powell, J.T. Inflammation and matrix metalloproteinases in the enlarging abdominal aortic aneurysm. *Arterioscler. Thromb. Vasc. Biol.* **1995**, *15*, 1145–1151. [CrossRef] [PubMed]
- 35. Miller, F.J., Jr.; Sharp, W.J.; Fang, X.; Oberley, L.W.; Oberley, T.D.; Weintraub, N.L. Oxidative stress in human abdominal aortic aneurysms: A potential mediator of aneurysmal remodeling. *Arterioscler. Thromb. Vasc. Biol.* **2002**, 22, 560–565. [CrossRef]
- 36. Zhang, J.; Schmidt, J.; Ryschich, E.; Mueller-Schilling, M.; Schumacher, H.; Allenberg, J.R. Inducible nitric oxide synthase is present in human abdominal aortic aneurysm and promotes oxidative vascular injury. *J. Vasc. Surg.* 2003, 38, 360–367. [CrossRef]
- 37. Thomas, M.; Gavrila, D.; McCormick, M.L.; Miller, F.J., Jr.; Daugherty, A.; Cassis, L.A.; Dellsperger, K.C.; Weintraub, N.L. Deletion of p47phox attenuates angiotensin ii-induced abdominal aortic aneurysm formation in apolipoprotein e-deficient mice. *Circulation* 2006, 114, 404–413. [CrossRef] [PubMed]
- 38. Gavazzi, G.; Deffert, C.; Trocme, C.; Schappi, M.; Herrmann, F.R.; Krause, K.H. Nox1 deficiency protects from aortic dissection in response to angiotensin ii. *Hypertension* **2007**, *50*, 189–196. [CrossRef] [PubMed]
- 39. Hadi, H.A.; Carr, C.S.; Al Suwaidi, J. Endothelial dysfunction: Cardiovascular risk factors, therapy, and outcome. *Vasc. Health Risk Manag.* **2005**, *1*, 183–198. [PubMed]
- 40. Godo, S.; Shimokawa, H. Endothelial functions. Arterioscler. Thromb. Vasc. Biol. 2017, 37, e108–e114. [CrossRef]
- 41. Deanfield, J.E.; Halcox, J.P.; Rabelink, T.J. Endothelial function and dysfunction: Testing and clinical relevance. *Circulation* **2007**, 115, 1285–1295. [CrossRef]
- 42. Forstermann, U.; Munzel, T. Endothelial nitric oxide synthase in vascular disease: From marvel to menace. *Circulation* **2006**, *113*, 1708–1714. [CrossRef]
- 43. Maulik, N.; Das, D.K. Redox signaling in vascular angiogenesis. Free Radic. Biol. Med. 2002, 33, 1047–1060. [CrossRef]
- 44. Favero, G.; Paganelli, C.; Buffoli, B.; Rodella, L.F.; Rezzani, R. Endothelium and its alterations in cardiovascular diseases: Life style intervention. *Biomed. Res. Int.* **2014**, 2014, 801896. [CrossRef] [PubMed]
- 45. Pierce, G.L.; Lesniewski, L.A.; Lawson, B.R.; Beske, S.D.; Seals, D.R. Nuclear factor-{kappa}b activation contributes to vascular endothelial dysfunction via oxidative stress in overweight/obese middle-aged and older humans. *Circulation* **2009**, 119, 1284–1292. [CrossRef]
- 46. Glass, C.K.; Witztum, J.L. Atherosclerosis. The road ahead. Cell 2001, 104, 503–516. [CrossRef]
- 47. Vora, D.K.; Fang, Z.T.; Liva, S.M.; Tyner, T.R.; Parhami, F.; Watson, A.D.; Drake, T.A.; Territo, M.C.; Berliner, J.A. Induction of p-selectin by oxidized lipoproteins. Separate effects on synthesis and surface expression. *Circ. Res.* 1997, 80, 810–818. [CrossRef]
- 48. Takei, A.; Huang, Y.; Lopes-Virella, M.F. Expression of adhesion molecules by human endothelial cells exposed to oxidized low density lipoprotein. Influences of degree of oxidation and location of oxidized ldl. *Atherosclerosis* **2001**, *154*, 79–86. [CrossRef]
- 49. Khan, B.V.; Parthasarathy, S.S.; Alexander, R.W.; Medford, R.M. Modified low density lipoprotein and its constituents augment cytokine-activated vascular cell adhesion molecule-1 gene expression in human vascular endothelial cells. *J. Clin. Investig.* 1995, 95, 1262–1270. [CrossRef] [PubMed]
- 50. Marui, N.; Offermann, M.K.; Swerlick, R.; Kunsch, C.; Rosen, C.A.; Ahmad, M.; Alexander, R.W.; Medford, R.M. Vascular cell adhesion molecule-1 (vcam-1) gene transcription and expression are regulated through an antioxidant-sensitive mechanism in human vascular endothelial cells. *J. Clin. Investig.* 1993, 92, 1866–1874. [CrossRef]
- 51. Chappell, D.C.; Varner, S.E.; Nerem, R.M.; Medford, R.M.; Alexander, R.W. Oscillatory shear stress stimulates adhesion molecule expression in cultured human endothelium. *Circ. Res.* **1998**, *82*, 532–539. [CrossRef]

Antioxidants **2021**, 10, 1244 13 of 15

52. Park, J.G.; Yoo, J.Y.; Jeong, S.J.; Choi, J.H.; Lee, M.R.; Lee, M.N.; Hwa Lee, J.; Kim, H.C.; Jo, H.; Yu, D.Y.; et al. Peroxiredoxin 2 deficiency exacerbates atherosclerosis in apolipoprotein e-deficient mice. *Circ. Res.* **2011**, *109*, 739–749. [CrossRef]

- 53. Cushing, S.D.; Berliner, J.A.; Valente, A.J.; Territo, M.C.; Navab, M.; Parhami, F.; Gerrity, R.; Schwartz, C.J.; Fogelman, A.M. Minimally modified low density lipoprotein induces monocyte chemotactic protein 1 in human endothelial cells and smooth muscle cells. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 5134–5138. [CrossRef]
- 54. Chen, M.; Masaki, T.; Sawamura, T. Lox-1, the receptor for oxidized low-density lipoprotein identified from endothelial cells: Implications in endothelial dysfunction and atherosclerosis. *Pharmacol. Ther.* **2002**, *95*, 89–100. [CrossRef]
- 55. Dimmeler, S.; Zeiher, A.M. Reactive oxygen species and vascular cell apoptosis in response to angiotensin ii and pro-atherosclerotic factors. *Regul. Pept.* **2000**, *90*, 19–25. [CrossRef]
- 56. Li, A.E.; Ito, H.; Rovira, I.I.; Kim, K.S.; Takeda, K.; Yu, Z.Y.; Ferrans, V.J.; Finkel, T. A role for reactive oxygen species in endothelial cell anoikis. *Circ. Res.* **1999**, *85*, 304–310. [CrossRef] [PubMed]
- 57. Camare, C.; Pucelle, M.; Negre-Salvayre, A.; Salvayre, R. Angiogenesis in the atherosclerotic plaque. *Redox Biol.* **2017**, 12, 18–34. [CrossRef] [PubMed]
- 58. Bennett, M.R.; Sinha, S.; Owens, G.K. Vascular smooth muscle cells in atherosclerosis. Circ. Res. 2016, 118, 692–702. [CrossRef]
- 59. Petsophonsakul, P.; Furmanik, M.; Forsythe, R.; Dweck, M.; Schurink, G.W.; Natour, E.; Reutelingsperger, C.; Jacobs, M.; Mees, B.; Schurgers, L. Role of vascular smooth muscle cell phenotypic switching and calcification in aortic aneurysm formation. *Arterioscler. Thromb. Vasc. Biol.* **2019**, *39*, 1351–1368. [CrossRef]
- 60. Ushio-Fukai, M.; Zafari, A.M.; Fukui, T.; Ishizaka, N.; Griendling, K.K. P22phox is a critical component of the superoxide-generating nadh/nadph oxidase system and regulates angiotensin ii-induced hypertrophy in vascular smooth muscle cells. *J. Biol. Chem.* **1996**, *271*, 23317–23321. [CrossRef]
- 61. Zafari, A.M.; Ushio-Fukai, M.; Akers, M.; Yin, Q.; Shah, A.; Harrison, D.G.; Taylor, W.R.; Griendling, K.K. Role of nadh/nadph oxidase-derived h2o2 in angiotensin ii-induced vascular hypertrophy. *Hypertension* **1998**, 32, 488–495. [CrossRef]
- 62. Sundaresan, M.; Yu, Z.X.; Ferrans, V.J.; Irani, K.; Finkel, T. Requirement for generation of h2o2 for platelet-derived growth factor signal transduction. *Science* 1995, 270, 296–299. [CrossRef] [PubMed]
- 63. Patterson, C.; Ruef, J.; Madamanchi, N.R.; Barry-Lane, P.; Hu, Z.; Horaist, C.; Ballinger, C.A.; Brasier, A.R.; Bode, C.; Runge, M.S. Stimulation of a vascular smooth muscle cell nad(p)h oxidase by thrombin. Evidence that p47(phox) may participate in forming this oxidase in vitro and in vivo. *J. Biol. Chem.* 1999, 274, 19814–19822. [CrossRef]
- 64. San Martin, A.; Griendling, K.K. Redox control of vascular smooth muscle migration. *Antioxid. Redox Signal.* **2010**, 12, 625–640. [CrossRef] [PubMed]
- 65. Chen, X.L.; Tummala, P.E.; Olbrych, M.T.; Alexander, R.W.; Medford, R.M. Angiotensin ii induces monocyte chemoattractant protein-1 gene expression in rat vascular smooth muscle cells. *Circ. Res.* **1998**, *83*, 952–959. [CrossRef]
- 66. Han, Y.; Runge, M.S.; Brasier, A.R. Angiotensin ii induces interleukin-6 transcription in vascular smooth muscle cells through pleiotropic activation of nuclear factor-kappa b transcription factors. *Circ. Res.* **1999**, *84*, 695–703. [CrossRef]
- 67. De Keulenaer, G.W.; Ushio-Fukai, M.; Yin, Q.; Chung, A.B.; Lyons, P.R.; Ishizaka, N.; Rengarajan, K.; Taylor, W.R.; Alexander, R.W.; Griendling, K.K. Convergence of redox-sensitive and mitogen-activated protein kinase signaling pathways in tumor necrosis factor-alpha-mediated monocyte chemoattractant protein-1 induction in vascular smooth muscle cells. *Arterioscler. Thromb. Vasc. Biol.* 2000, 20, 385–391. [CrossRef]
- 68. Rhee, S.G.; Kang, S.W.; Chang, T.S.; Jeong, W.; Kim, K. Peroxiredoxin, a novel family of peroxidases. *IUBMB Life* **2001**, *52*, 35–41. [CrossRef]
- 69. Szeliga, M. Peroxiredoxins in neurodegenerative diseases. Antioxidants 2020, 9, 1203. [CrossRef]
- 70. Wood, Z.A.; Poole, L.B.; Karplus, P.A. Peroxiredoxin evolution and the regulation of hydrogen peroxide signaling. *Science* **2003**, 300, 650–653. [CrossRef]
- 71. Chae, H.Z.; Robison, K.; Poole, L.B.; Church, G.; Storz, G.; Rhee, S.G. Cloning and sequencing of thiol-specific antioxidant from mammalian brain: Alkyl hydroperoxide reductase and thiol-specific antioxidant define a large family of antioxidant enzymes. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 7017–7021. [CrossRef]
- 72. Wood, Z.A.; Schroder, E.; Robin Harris, J.; Poole, L.B. Structure, mechanism and regulation of peroxiredoxins. *Trends Biochem. Sci.* **2003**, *28*, 32–40. [CrossRef]
- 73. Peskin, A.V.; Low, F.M.; Paton, L.N.; Maghzal, G.J.; Hampton, M.B.; Winterbourn, C.C. The high reactivity of peroxiredoxin 2 with h(2)o(2) is not reflected in its reaction with other oxidants and thiol reagents. *J. Biol. Chem.* **2007**, *282*, 11885–11892. [CrossRef]
- 74. Ying, J.; Clavreul, N.; Sethuraman, M.; Adachi, T.; Cohen, R.A. Thiol oxidation in signaling and response to stress: Detection and quantification of physiological and pathophysiological thiol modifications. *Free Radic. Biol. Med.* **2007**, 43, 1099–1108. [CrossRef] [PubMed]
- 75. Rhee, S.G.; Bae, Y.S.; Lee, S.R.; Kwon, J. Hydrogen peroxide: A key messenger that modulates protein phosphorylation through cysteine oxidation. *Sci. STKE* **2000**, 2000, pe1. [CrossRef]
- 76. Hall, A.; Nelson, K.; Poole, L.B.; Karplus, P.A. Structure-based insights into the catalytic power and conformational dexterity of peroxiredoxins. *Antioxid. Redox Signal.* **2011**, *15*, 795–815. [CrossRef]
- 77. Mowbray, A.L.; Kang, D.H.; Rhee, S.G.; Kang, S.W.; Jo, H. Laminar shear stress up-regulates peroxiredoxins (prx) in endothelial cells: Prx 1 as a mechanosensitive antioxidant. *J. Biol. Chem.* **2008**, *283*, 1622–1627. [CrossRef]
- 78. Davignon, J.; Ganz, P. Role of endothelial dysfunction in atherosclerosis. Circulation 2004, 109, III27–III32. [CrossRef] [PubMed]

Antioxidants **2021**, 10, 1244 14 of 15

79. Kisucka, J.; Chauhan, A.K.; Patten, I.S.; Yesilaltay, A.; Neumann, C.; Van Etten, R.A.; Krieger, M.; Wagner, D.D. Peroxiredoxin1 prevents excessive endothelial activation and early atherosclerosis. *Circ. Res.* **2008**, *103*, 598–605. [CrossRef] [PubMed]

- 80. Jeong, S.J.; Kim, S.; Park, J.G.; Jung, I.H.; Lee, M.N.; Jeon, S.; Kweon, H.Y.; Yu, D.Y.; Lee, S.H.; Jang, Y.; et al. Prdx1 (peroxiredoxin 1) deficiency reduces cholesterol efflux via impaired macrophage lipophagic flux. *Autophagy* **2018**, *14*, 120–133. [CrossRef]
- 81. Martinez-Pinna, R.; Ramos-Mozo, P.; Madrigal-Matute, J.; Blanco-Colio, L.M.; Lopez, J.A.; Calvo, E.; Camafeita, E.; Lindholt, J.S.; Meilhac, O.; Delbosc, S.; et al. Identification of peroxiredoxin-1 as a novel biomarker of abdominal aortic aneurysm. *Arterioscler. Thromb. Vasc. Biol.* **2011**, *31*, 935–943. [CrossRef]
- 82. Tang, C.; Yin, G.; Huang, C.; Wang, H.; Gao, J.; Luo, J.; Zhang, Z.; Wang, J.; Hong, J.; Chai, X. Peroxiredoxin-1 ameliorates pressure overload-induced cardiac hypertrophy and fibrosis. *Biomed. Pharmacother.* **2020**, 129, 110357. [CrossRef] [PubMed]
- 83. Jiang, L.; Gong, Y.; Hu, Y.; You, Y.; Wang, J.; Zhang, Z.; Wei, Z.; Tang, C. Peroxiredoxin-1 overexpression attenuates doxorubicin-induced cardiotoxicity by inhibiting oxidative stress and cardiomyocyte apoptosis. *Oxid Med. Cell Longev.* **2020**, 2020, 2405135. [CrossRef] [PubMed]
- 84. Basatemur, G.L.; Jorgensen, H.F.; Clarke, M.C.H.; Bennett, M.R.; Mallat, Z. Vascular smooth muscle cells in atherosclerosis. *Nat. Rev. Cardiol.* **2019**, *16*, 727–744. [CrossRef]
- 85. Jeong, S.J.; Cho, M.J.; Ko, N.Y.; Kim, S.; Jung, I.H.; Min, J.K.; Lee, S.H.; Park, J.G.; Oh, G.T. Deficiency of peroxiredoxin 2 exacerbates angiotensin ii-induced abdominal aortic aneurysm. *Exp. Mol. Med.* **2020**, *52*, 1587–1601. [CrossRef]
- 86. Jang, J.Y.; Wang, S.B.; Min, J.H.; Chae, Y.H.; Baek, J.Y.; Yu, D.Y.; Chang, T.S. Peroxiredoxin ii is an antioxidant enzyme that negatively regulates collagen-stimulated platelet function. *J. Biol. Chem.* **2015**, 290, 11432–11442. [CrossRef]
- 87. Li, W.; Febbraio, M.; Reddy, S.P.; Yu, D.Y.; Yamamoto, M.; Silverstein, R.L. Cd36 participates in a signaling pathway that regulates ros formation in murine vsmcs. *J. Clin. Investig.* **2010**, 120, 3996–4006. [CrossRef] [PubMed]
- 88. Osellame, L.D.; Blacker, T.S.; Duchen, M.R. Cellular and molecular mechanisms of mitochondrial function. *Best Pract. Res. Clin. Endocrinol. Metab.* **2012**, *26*, 711–723. [CrossRef]
- 89. Maharjan, S.; Oku, M.; Tsuda, M.; Hoseki, J.; Sakai, Y. Mitochondrial impairment triggers cytosolic oxidative stress and cell death following proteasome inhibition. *Sci. Rep.* **2014**, *4*, 5896. [CrossRef]
- 90. Mailloux, R.J. Mitochondrial antioxidants and the maintenance of cellular hydrogen peroxide levels. *Oxid. Med. Cell Longev.* **2018**, 2018, 7857251. [CrossRef]
- 91. Matsushima, S.; Ide, T.; Yamato, M.; Matsusaka, H.; Hattori, F.; Ikeuchi, M.; Kubota, T.; Sunagawa, K.; Hasegawa, Y.; Kurihara, T.; et al. Overexpression of mitochondrial peroxiredoxin-3 prevents left ventricular remodeling and failure after myocardial infarction in mice. *Circulation* 2006, 113, 1779–1786. [CrossRef] [PubMed]
- 92. Kumar, V.; Kitaeff, N.; Hampton, M.B.; Cannell, M.B.; Winterbourn, C.C. Reversible oxidation of mitochondrial peroxiredoxin 3 in mouse heart subjected to ischemia and reperfusion. *FEBS Lett.* **2009**, *583*, 997–1000. [CrossRef] [PubMed]
- 93. Bravo-San Pedro, J.M.; Kroemer, G.; Galluzzi, L. Autophagy and mitophagy in cardiovascular disease. *Circ. Res.* **2017**, *120*, 1812–1824. [CrossRef]
- 94. Poznyak, A.V.; Nikiforov, N.G.; Wu, W.K.; Kirichenko, T.V.; Orekhov, A.N. Autophagy and mitophagy as essential components of atherosclerosis. *Cells* **2021**, *10*, 443. [CrossRef]
- 95. Vasquez-Trincado, C.; Garcia-Carvajal, I.; Pennanen, C.; Parra, V.; Hill, J.A.; Rothermel, B.A.; Lavandero, S. Mitochondrial dynamics, mitophagy and cardiovascular disease. *J. Physiol.* **2016**, *594*, 509–525. [CrossRef]
- 96. Um, J.H.; Yun, J. Emerging role of mitophagy in human diseases and physiology. BMB Rep. 2017, 50, 299–307. [CrossRef]
- 97. Jiang, M.Y.; Han, Z.D.; Li, W.; Yue, F.; Ye, J.; Li, B.; Cai, Z.; Lu, J.M.; Dong, W.; Jiang, X.; et al. Mitochondrion-associated protein peroxiredoxin 3 promotes benign prostatic hyperplasia through autophagy suppression and pyroptosis activation. *Oncotarget* 2017, *8*, 80295–80302. [CrossRef]
- 98. Tian, Y.; Lv, W.; Lu, C.; Zhao, X.; Zhang, C.; Song, H. Lats2 promotes cardiomyocyte h9c2 cells apoptosis via the prx3-mfn2-mitophagy pathways. *J. Recept Signal. Transduct Res.* **2019**, 39, 470–478. [CrossRef]
- 99. Zito, E.; Melo, E.P.; Yang, Y.; Wahlander, A.; Neubert, T.A.; Ron, D. Oxidative protein folding by an endoplasmic reticulum-localized peroxiredoxin. *Mol. Cell* **2010**, *40*, 787–797. [CrossRef] [PubMed]
- 100. Braakman, I.; Bulleid, N.J. Protein folding and modification in the mammalian endoplasmic reticulum. *Annu. Rev. Biochem.* **2011**, 80, 71–99. [CrossRef] [PubMed]
- 101. Sato, Y.; Kojima, R.; Okumura, M.; Hagiwara, M.; Masui, S.; Maegawa, K.; Saiki, M.; Horibe, T.; Suzuki, M.; Inaba, K. Synergistic cooperation of pdi family members in peroxiredoxin 4-driven oxidative protein folding. *Sci. Rep.* **2013**, *3*, 2456. [CrossRef]
- 102. Zito, E. Prdx4, an endoplasmic reticulum-localized peroxiredoxin at the crossroads between enzymatic oxidative protein folding and nonenzymatic protein oxidation. *Antioxid. Redox Signal.* **2013**, *18*, 1666–1674. [CrossRef]
- 103. Abbasi, A.; Corpeleijn, E.; Postmus, D.; Gansevoort, R.T.; de Jong, P.E.; Gans, R.O.; Struck, J.; Schulte, J.; Hillege, H.L.; van der Harst, P.; et al. Peroxiredoxin 4, a novel circulating biomarker for oxidative stress and the risk of incident cardiovascular disease and all-cause mortality. *J. Am. Heart Assoc.* 2012, 1, e002956. [CrossRef]
- 104. El Eter, E.; Al Masri, A.; Habib, S.; Al Zamil, H.; Al Hersi, A.; Al Hussein, F.; Al Omran, M. Novel links among peroxiredoxins, endothelial dysfunction, and severity of atherosclerosis in type 2 diabetic patients with peripheral atherosclerotic disease. *Cell Stress Chaperones* **2014**, *19*, 173–181. [CrossRef] [PubMed]

Antioxidants 2021, 10, 1244 15 of 15

105. Guo, X.; Yamada, S.; Tanimoto, A.; Ding, Y.; Wang, K.Y.; Shimajiri, S.; Murata, Y.; Kimura, S.; Tasaki, T.; Nabeshima, A.; et al. Overexpression of peroxiredoxin 4 attenuates atherosclerosis in apolipoprotein e knockout mice. *Antioxid. Redox Signal.* **2012**, 17, 1362–1375. [CrossRef] [PubMed]

- 106. Tabas, I. The role of endoplasmic reticulum stress in the progression of atherosclerosis. *Circ. Res.* **2010**, *107*, 839–850. [CrossRef] [PubMed]
- 107. Knoops, B.; Goemaere, J.; Van der Eecken, V.; Declercq, J.P. Peroxiredoxin 5: Structure, mechanism, and function of the mammalian atypical 2-cys peroxiredoxin. *Antioxid. Redox Signal.* **2011**, *15*, 817–829. [CrossRef] [PubMed]
- 108. Sim, J.; Park, J.; Woo, H.A.; Rhee, S.G. Maturation of mitochondrially targeted prx v involves a second cleavage by mitochondrial intermediate peptidase that is sensitive to inhibition by h2o2. *Antioxidants* **2021**, *10*, 346. [CrossRef]
- 109. Kim, M.H.; Park, S.J.; Kim, J.H.; Seong, J.B.; Kim, K.M.; Woo, H.A.; Lee, D.S. Peroxiredoxin 5 regulates adipogenesis-attenuating oxidative stress in obese mouse models induced by a high-fat diet. *Free Radic. Biol. Med.* **2018**, 123, 27–38. [CrossRef] [PubMed]
- 110. Park, J.; Lee, E.G.; Yi, H.J.; Kim, N.H.; Rhee, S.G.; Woo, H.A. Ablation of peroxiredoxin v exacerbates ischemia/reperfusion-induced kidney injury in mice. *Antioxidants* **2020**, *9*, 769. [CrossRef]
- 111. Shichita, T.; Hasegawa, E.; Kimura, A.; Morita, R.; Sakaguchi, R.; Takada, I.; Sekiya, T.; Ooboshi, H.; Kitazono, T.; Yanagawa, T.; et al. Peroxiredoxin family proteins are key initiators of post-ischemic inflammation in the brain. *Nat. Med.* **2012**, *18*, 911–917. [CrossRef]
- 112. Knoops, B.; Becker, S.; Poncin, M.A.; Glibert, J.; Derclaye, S.; Clippe, A.; Alsteens, D. Specific interactions measured by afm on living cells between peroxiredoxin-5 and tlr4: Relevance for mechanisms of innate immunity. *Cell Chem. Biol.* **2018**, 25, 550–559.e3. [CrossRef]
- 113. Wang, M.X.; Wei, A.; Yuan, J.; Trickett, A.; Knoops, B.; Murrell, G.A. Expression and regulation of peroxiredoxin 5 in human osteoarthritis. *FEBS Lett.* **2002**, *531*, 359–362. [CrossRef]
- 114. Abbas, K.; Breton, J.; Picot, C.R.; Quesniaux, V.; Bouton, C.; Drapier, J.C. Signaling events leading to peroxiredoxin 5 up-regulation in immunostimulated macrophages. *Free Radic. Biol. Med.* **2009**, 47, 794–802. [CrossRef]
- 115. Diet, A.; Abbas, K.; Bouton, C.; Guillon, B.; Tomasello, F.; Fourquet, S.; Toledano, M.B.; Drapier, J.C. Regulation of peroxiredoxins by nitric oxide in immunostimulated macrophages. *J. Biol. Chem.* **2007**, *282*, 36199–36205. [CrossRef]
- 116. Sun, H.N.; Kim, S.U.; Huang, S.M.; Kim, J.M.; Park, Y.H.; Kim, S.H.; Yang, H.Y.; Chung, K.J.; Lee, T.H.; Choi, H.S.; et al. Microglial peroxiredoxin v acts as an inducible anti-inflammatory antioxidant through cooperation with redox signaling cascades. *J. Neurochem.* **2010**, *114*, 39–50.
- 117. Fisher, A.B. Peroxiredoxin 6: A bifunctional enzyme with glutathione peroxidase and phospholipase a(2) activities. *Antioxid. Redox Signal.* **2011**, *15*, 831–844. [CrossRef] [PubMed]
- 118. Fisher, A.B.; Dodia, C.; Chatterjee, S. A peptide inhibitor of peroxiredoxin 6 phospholipase a2 activity significantly protects against lung injury in a mouse model of ventilator induced lung injury (vili). *Antioxidants* **2021**, *10*, 925. [CrossRef] [PubMed]
- 119. Hui, D.Y. Phospholipase a(2) enzymes in metabolic and cardiovascular diseases. *Curr. Opin. Lipidol.* **2012**, 23, 235–240. [CrossRef] [PubMed]
- 120. Nagy, N.; Malik, G.; Fisher, A.B.; Das, D.K. Targeted disruption of peroxiredoxin 6 gene renders the heart vulnerable to ischemia-reperfusion injury. *Am. J. Physiol. Heart Circ. Physiol.* **2006**, 291, H2636–H2640. [CrossRef] [PubMed]
- 121. Manevich, Y.; Sweitzer, T.; Pak, J.H.; Feinstein, S.I.; Muzykantov, V.; Fisher, A.B. 1-cys peroxiredoxin overexpression protects cells against phospholipid peroxidation-mediated membrane damage. *Proc. Natl. Acad. Sci. USA* 2002, 99, 11599–11604. [CrossRef] [PubMed]
- 122. Li, D.X.; Chen, W.; Jiang, Y.L.; Ni, J.Q.; Lu, L. Antioxidant protein peroxiredoxin 6 suppresses the vascular inflammation, oxidative stress and endothelial dysfunction in angiotensin ii-induced endotheliocyte. *Gen. Physiol. Biophys.* **2020**, *39*, 545–555. [CrossRef] [PubMed]
- 123. Wang, X.; Phelan, S.A.; Petros, C.; Taylor, E.F.; Ledinski, G.; Jurgens, G.; Forsman-Semb, K.; Paigen, B. Peroxiredoxin 6 deficiency and atherosclerosis susceptibility in mice: Significance of genetic background for assessing atherosclerosis. *Atherosclerosis* **2004**, 177, 61–70. [CrossRef] [PubMed]
- 124. Phelan, S.A.; Wang, X.; Wallbrandt, P.; Forsman-Semb, K.; Paigen, B. Overexpression of prdx6 reduces h2o2 but does not prevent diet-induced atherosclerosis in the aortic root. *Free Radic. Biol. Med.* **2003**, *35*, 1110–1120. [CrossRef]