

**Table S1.** Number of metagenome-assembled genomes (**MAGs**) recovered (n=1806) per species and strains identified (n=177).

Taxonomic identification	Number of MAGs	Taxonomic identification	Number of MAGs
<i>Escherichia coli</i>	140	<i>Lachnospiraceae bacterium KM106-2</i>	7
<i>Faecalibacterium prausnitzii</i>	116	<i>Phascolarctobacterium faecium</i>	7
<i>Ruminococcus gnavus</i>	114	<i>Prevotella dentalis</i>	7
<i>Collinsella aerofaciens</i>	71	<i>Ruminococcus albus</i>	7
<i>Bifidobacterium longum</i>	70	<i>Alistipes onderdonkii</i>	6
<i>Veillonella parvula</i>	62	<i>Bacteroides vulgatus</i>	6
<i>Akkermansia muciniphila</i>	59	<i>Clostridium saccharolyticum</i>	6
<i>Lachnospiraceae bacterium GAM79</i>	50	<i>Haemophilus parainfluenzae</i>	6
<i>Ruthenibacterium lactatiformans</i>	46	<i>Lactococcus lactis</i>	6
<i>Bifidobacterium bifidum</i>	39	<i>Muribaculum sp. TLL-A4</i>	6
<i>Bifidobacterium breve</i>	38	<i>Roseburia hominis</i>	6
<i>Blautia sp. SC05B48</i>	38	<i>Veillonella rodentium</i>	6
<i>Clostridium bolteae</i>	36	<i>Alistipes communis</i>	5
<i>Bifidobacterium pseudocatenulatum</i>	35	<i>Alistipes shahii</i>	5
<i>Faecalitalea cylindroides</i>	35	<i>Bacteroides fragilis</i>	5
<i>Ruminococcus sp. JE7A12</i>	32	<i>Eggerthella lenta</i>	5
<i>Parabacteroides distasonis</i>	27	<i>Streptococcus thermophilus</i>	5
<i>Ruminococcus bicirculans</i>	26	<i>Turicibacter sp. H121</i>	5
<i>Lachnoclostridium sp. YL32</i>	25	<i>Absiella sp. 9CBEGH2</i>	4
<i>Longibaculum sp. KGMB06250</i>	25	<i>Barnesiella viscericola</i>	4
<i>Absiella argi</i>	23	<i>Butyrivibrio fibrisolvens</i>	4
<i>Anaerostipes hadrus</i>	22	<i>Clostridioides difficile</i>	4
<i>Prevotella ruminicola</i>	22	<i>Enterococcus avium</i>	4
<i>Flavonifractor plautii</i>	21	<i>Paraprevotella xylaniphila</i>	4
<i>Lachnospira eligens</i>	21	<i>Sutterella faecalis</i>	4
<i>Lachnospiraceae bacterium</i>	20	<i>Burkholderiales bacterium YL45</i>	3
<i>Caproiciproducens sp. NJN-50</i>	19	<i>Clostridium butyricum</i>	3
<i>Veillonella atypica</i>	19	<i>Clostridium cellulosi</i>	3
<i>Veillonella dispar</i>	18	<i>Dialister pneumosintes</i>	3
<i>Bifidobacterium adolescentis</i>	17	<i>Flintibacter sp. KGMB00164</i>	3
<i>Clostridium sp. enoides</i>	17	<i>Klebsiella aerogenes</i>	3
<i>Ruminococcus chamanellensis</i>	17	<i>Lachnoclostridium phocaeense</i>	3
<i>Blautia sp. N6H1-15</i>	16	<i>Ligilactobacillus ruminis</i>	3
<i>Lachnospiraceae bacterium Choco86</i>	16	<i>Megamonas funiformis</i>	3
<i>Enterococcus faecalis</i>	15	<i>Methanobrevibacter smithii</i>	3
<i>Erysipelotrichaceae bacterium GAM147</i>	15	<i>Prevotella intermedia</i>	3
<i>Clostridium perfringens</i>	14	<i>Actinomyces pacaensis</i>	2
<i>Megasphaera elsdenii</i>	13	<i>Alistipes dispar</i>	2
<i>Blautia producta</i>	13	<i>Alistipes finegoldii</i>	2
<i>Bacteroides uniformis</i>	12	<i>Bacillus sp. SB49</i>	2
<i>Klebsiella pneumoniae</i>	12	<i>Bifidobacterium dentium</i>	2
<i>Lacticaseibacillus rhamnosus</i>	11	<i>Brachyspira pilosicoli</i>	2
<i>Streptococcus sp. HSISS3</i>	11	<i>Citrobacter freundii</i>	2
<i>Roseburia intestinalis</i>	9	<i>Citrobacter koseri</i>	2
<i>Streptococcus salivarius</i>	9	<i>Clostridium hylemoniae</i>	2
<i>Bacteroides thetaiotomicron</i>	8	<i>Clostridium scindens</i>	2
<i>Enterococcus sp. HSIEG1</i>	8	<i>Clostridium sp. CT4</i>	2
<i>Oscillibacter valericigenes</i>	8	<i>Enterobacter hormaechei</i>	2
<i>Phascolarctobacterium succinatutens</i>	8	<i>Eubacterium limosum</i>	2
<i>Streptococcus lutetiensis</i>	8	<i>Faecalibaculum rodentium</i>	2

**Table S1.** Cont.

Taxonomic identification	Number of MAGs	Taxonomic identification	Number of MAGs
<i>Klebsiella michiganensis</i>	2	<i>Lentilactobacillus hilgardii</i>	1
<i>Klebsiella oxytoca</i>	2	<i>Limosilactobacillus reuteri</i>	1
<i>Muribaculaceae bacterium DSM</i>	2	<i>Lactobacillus terrae</i>	1
<i>Selenomonas sp. oral</i>	2	<i>Leuconostoc lactis</i>	1
<i>Serratia marcescens</i>	2	<i>Mageebacillus indolicus</i>	1
<i>Staphylococcus epidermidis</i>	2	<i>Megasphaera stantonii</i>	1
<i>Succinivibrio dextrinosolvens</i>	2	<i>Microbacterium aurum</i>	1
<i>Weissella confusa</i>	2	<i>Microbacterium hominis</i>	1
<i>Acetobacterium woodii</i>	1	<i>Neorhizobium galegae</i>	1
<i>Actinomyces sp. oral</i>	1	<i>Olsenella sp. GAM18</i>	1
<i>Alistipes megagutti</i>	1	<i>Paenibacillus sabinae</i>	1
<i>Aminipila butyrlica</i>	1	<i>Prevotella denticola</i>	1
<i>Anaerostipes rhamnosivorans</i>	1	<i>Pseudobutyryrivibrio xylanivorans</i>	1
<i>Bacillus megaterium</i>	1	<i>Rathayibacter sp. VKM</i>	1
<i>Bacillus thuringiensis</i>	1	<i>Rhizobium sp. NXC24</i>	1
<i>Bacteroides coprosuis</i>	1	<i>Serratia liquefaciens</i>	1
<i>Bacteroides ovatus</i>	1	<i>Streptococcus gallolyticus</i>	1
<i>Bacteroides salanitronis</i>	1	<i>Streptococcus sp. HSIM1</i>	1
<i>Bacteroides xylinosolvens</i>	1	<i>Streptococcus sp. HSISS2</i>	1
<i>Bifidobacterium catenulatum</i>	1	<i>Streptococcus sp. LPB0220</i>	1
<i>Bifidobacterium scardovii</i>	1	<i>Streptococcus vestibularis</i>	1
<i>Bradyrhizobium erythrophlei</i>	1	<i>Streptomyces qinzhouensis</i>	1
<i>Brevibacillus agri</i>	1	<i>Streptomyces sp. 769</i>	1
<i>Burkholderia cenocepacia</i>	1	<i>Streptomyces sp. S4.7</i>	1
<i>Burkholderia multivorans</i>	1	<i>Streptomyces sp. Sge12</i>	1
<i>Butyrivibrio hungatei</i>	1	<i>Sutterella megalosphaerooides</i>	1
<i>Butyrivibrio proteoclasticus</i>	1	<i>Virgibacillus phasianinus</i>	1
<i>Cellulomonas sp. Z28</i>	1		
<i>Citrobacter portucalensis</i>	1		
<i>Citrobacter sp. CFNIH10</i>	1		
<i>Clostridiales bacterium CCNA10</i>	1		
<i>Clostridium botulinum</i>	1		
<i>Clostridium saccharobyticum</i>	1		
<i>Clostridium sp. BNLI100</i>	1		
<i>Clostridium sp. JN-9</i>	1		
<i>Corynebacterium epidermidicanis</i>	1		
<i>Corynebacterium sp. 2184</i>	1		
<i>Corynebacterium stationis</i>	1		
<i>Cutibacterium avidum</i>	1		
<i>Dialister hominis</i>	1		
<i>Dialister massiliensis</i>	1		
<i>Eggerthella sp. YY7918</i>	1		
<i>Enterococcus casseliflavus</i>	1		
<i>Erysipelotrichaceae bacterium I46</i>	1		
<i>Ethanoligenens harbinense</i>	1		
<i>Finegoldia magna</i>	1		
<i>Intestinibaculum porci</i>	1		
<i>Klebsiella variicola</i>	1		
<i>Lachnospiraceae bacterium oral</i>	1		
<i>Companilactobacillus futsaii</i>	1		

**Table S2.** Number of metagenome-assembled genomes (**MAGs**) recovered per diet type. **GOS:** galacto-oligosaccharides, **FOS:** fructo-oligosaccharides.

Number of MAGs per diet type considering all species identified	
Taxonomic identification	Number of MAGs
Infant formula-fed (infant)	638
Fiber-rich (adult)	467
Infant formula-fed + GOS (infant)	281
Breast-fed (infant)	144
Infant formula-fed + FOS (infant)	106
Infant formula-fed + GOS + FOS (infant)	84
Breast-fed + GOS (infant)	75
Whole-milk-fed (infant)	8
Breast-fed + GOS + FOS (infant)	3

**Table S2.** Cont.

Taxonomic identification	Number of MAGs per diet type considering only <i>Bifidobacterium</i> species								
	Total	<i>B. adolescentis</i>	<i>B. bifidum</i>	<i>B. breve</i>	<i>B. catenulatum</i>	<i>B. dentium</i>	<i>B. longum</i>	<i>B. pseudocatenulatum</i>	<i>B. scardovii</i>
Infant formula-fed (infant)	64	0	8	9	1	0	32	13	1
Infant formula-fed + GOS (infant)	52	2	5	16	0	0	20	9	0
Breast-fed (infant)	31	0	11	7	0	0	9	4	0
Infant formula-fed + GOS + FOS (infant)	17	5	7	0	0	2	2	1	0
Infant formula-fed + FOS (infant)	12	2	0	3	0	0	3	4	0
Fiber-rich (adult)	11	8	0	0	0	0	2	1	0
Breast-fed + GOS (infant)	11	0	5	3	0	0	2	1	0
Whole-milk-fed (infant)	4	0	2	0	0	0	0	2	0
Breast-fed + GOS + FOS (infant)	1	0	1	0	0	0	0	0	0

**Table S3.** Glycosidase activities of interest included in selected CAZy families.

CAZy family	Activities of interest included in the family
CBM32	Sialidase
CBM40	Sialidase
GH1	$\beta$ -galactosidase
GH2	$\beta$ -galactosidase
GH3	$\beta$ -N-acetylhexosaminidase
GH4	$\alpha$ -galactosidase
GH5 1	$\beta$ -N-acetylhexosaminidase
GH5 2	$\beta$ -N-acetylhexosaminidase
GH5 4	$\beta$ -N-acetylhexosaminidase
GH5 5	$\beta$ -N-acetylhexosaminidase
GH5 7	$\beta$ -N-acetylhexosaminidase
GH5 8	$\beta$ -N-acetylhexosaminidase
GH5 10	$\beta$ -N-acetylhexosaminidase
GH5 13	$\beta$ -N-acetylhexosaminidase
GH5 18	$\beta$ -N-acetylhexosaminidase
GH5 21	$\beta$ -N-acetylhexosaminidase
GH5 22	$\beta$ -N-acetylhexosaminidase
GH5 25	$\beta$ -N-acetylhexosaminidase
GH5 37	$\beta$ -N-acetylhexosaminidase
GH5 38	$\beta$ -N-acetylhexosaminidase
GH5 41	$\beta$ -N-acetylhexosaminidase
GH5 44	$\beta$ -N-acetylhexosaminidase
GH5 46	$\beta$ -N-acetylhexosaminidase
GH5 54	$\beta$ -N-acetylhexosaminidase
GH16	endo- $\beta$ -1,4-galactosidase
GH20	$\beta$ -hexosaminidase
GH29	$\alpha$ -L-fucosidase ; $\alpha$ -1,3/1,4-L-fucosidase
GH30	$\beta$ -fucosidase
GH30 1	$\beta$ -fucosidase
GH30 2	$\beta$ -fucosidase
GH30 3	$\beta$ -fucosidase
GH30 4	$\beta$ -fucosidase
GH30 5	$\beta$ -fucosidase
GH30 6	$\beta$ -fucosidase
GH30 8	$\beta$ -fucosidase
GH30 9	$\beta$ -fucosidase
GH31	$\alpha$ -galactosidase
GH32	invertase ; endo-inulinase ; $\beta$ -2,6-fructan 6-levanbiohydrolase ; endo-levanase ; exo-inulinase ; fructan $\beta$ --fructosidase/1-exohydrolase ; fructan $\beta$ --fructosidase/6-exohydrolase ; sucrose:sucrose 1-fructosyltransferase ; fructan:fructan 1-fructosyltransferase ; sucrose:fructan 6-fructosyltransferase ; fructan:fructan 6G-fructosyltransferase ; levan fructosyltransferase ; [retaining] sucrose:sucrose 6-fructosyltransferase ; cycloinulo-oligosaccharide fructanotransferase.
GH33	sialidase ; trans-sialidase
GH35	$\beta$ -galactosidase
GH36	$\alpha$ -galactosidase ; stachyose synthase ; raffinose synthase.
GH39	$\beta$ -galactosidase.
GH42	$\beta$ -galactosidase
GH58	endo-sialidase.
GH59	$\beta$ -galactosidase
GH68	levansucrase ; $\beta$ -fructofuranosidase ; inulosucrase
GH95	$\alpha$ -L-fucosidase ; $\alpha$ -1,2-L-fucosidase ; $\alpha$ -L-galactosidase
GH97	glucoamylase ; $\alpha$ -glucosidase ; $\alpha$ -galactosidase.
GH109	$\beta$ -N-acetylhexosaminidase
GH110	$\alpha$ -galactosidase
GH139	$\alpha$ -2-O-Me-L-fucosidase.
GH141	$\alpha$ -L-fucosidase
GH147	$\beta$ -galactosidase
GH151	$\alpha$ -L-fucosidase

**Table S4.** Sensitivity, specificity, precision, recall, F1 coefficients and balanced accuracy rates for artificial neural network (ANN) random forest (RF) and Generalized Linear Model Elastic-net (GLMNET) computed to classify metagenome-assembled genomes (MAGs) of *B. adolescentis*, *B. bifidum*, *B. breve*, *B. longum* and *B. pseudocatenulatum* based on their glycosidase activities.

<b>ANN</b>	<b>Sensitivity</b>	<b>Specificity</b>	<b>Precision</b>	<b>Recall</b>	<b>F1</b>	<b>Balanced accuracy</b>
<i>Bifidobacterium adolescentis</i>	0.63	1.00	1.00	0.63	0.77	0.81
<i>Bifidobacterium bifidum</i>	1.00	0.98	0.92	1.00	0.96	0.99
<i>Bifidobacterium breve</i>	1.00	0.98	0.91	1.00	0.95	0.99
<i>Bifidobacterium longum</i>	0.95	0.97	0.95	0.95	0.95	0.96
<i>Bifidobacterium pseudocatenulatum</i>	0.90	0.96	0.82	0.90	0.86	0.93

<b>RF</b>	<b>Sensitivity</b>	<b>Specificity</b>	<b>Precision</b>	<b>Recall</b>	<b>F1</b>	<b>Balanced accuracy</b>
<i>Bifidobacterium adolescentis</i>	0.71	1.00	1.00	0.71	0.83	0.86
<i>Bifidobacterium bifidum</i>	0.92	0.98	0.92	0.92	0.92	0.95
<i>Bifidobacterium breve</i>	1.00	0.98	0.91	1.00	0.95	0.99
<i>Bifidobacterium longum</i>	0.95	0.97	0.95	0.95	0.95	0.96
<i>Bifidobacterium pseudocatenulatum</i>	0.90	0.96	0.82	0.90	0.86	0.93

<b>GLMNET</b>	<b>Sensitivity</b>	<b>Specificity</b>	<b>Precision</b>	<b>Recall</b>	<b>F1</b>	<b>Balanced accuracy</b>
<i>Bifidobacterium adolescentis</i>	0.71	1.00	1.00	0.71	0.83	0.86
<i>Bifidobacterium bifidum</i>	0.92	0.98	0.92	0.92	0.92	0.95
<i>Bifidobacterium breve</i>	1.00	0.98	0.91	1.00	0.95	0.99
<i>Bifidobacterium longum</i>	0.95	0.97	0.95	0.95	0.95	0.96
<i>Bifidobacterium pseudocatenulatum</i>	0.90	0.96	0.82	0.90	0.86	0.93

**Table S5.** Variable importance coefficients of artificial neural network (ANN) computed to classify metagenome-assembled genomes (**MAGs**) of *B. adolescentis*, *B. bifidum*, *B. breve*, *B. longum* and *B. pseudocatenulatum* based on their glycosidase activities. Importance coefficients were determined by the sum of the product of raw input-hidden and hidden-output connection weights.

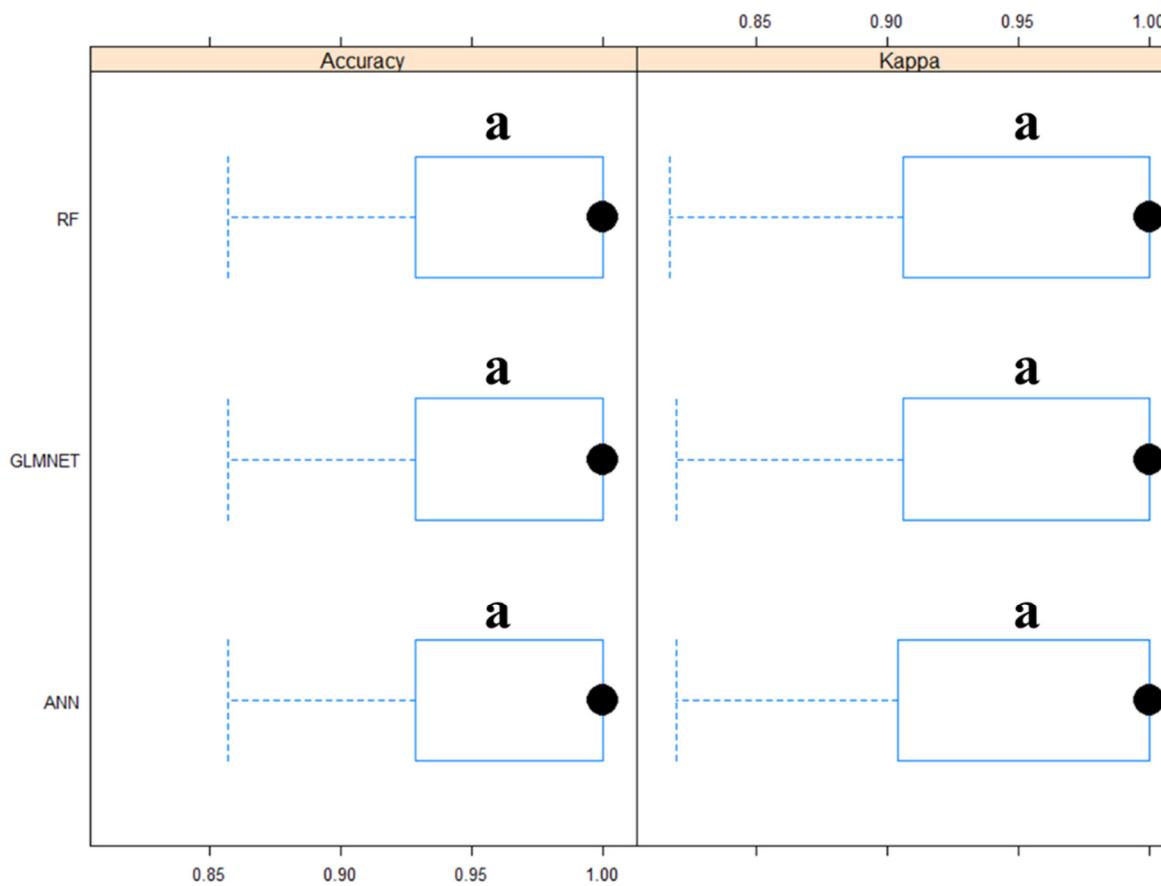
	Importance (%)		Importance (%)		Importance (%)		Importance (%)		Importance (%)
<i>B. adolescentis</i>		<i>B. bifidum</i>		<i>B. breve</i>		<i>B. longum</i>		<i>B. pseudocatenulatum</i>	
<b>GH2</b>	100.0	<b>GH110</b>	100.0	<b>GH5 18</b>	100.0	<b>GH30 5</b>	100.0	<b>GH5 44</b>	100.0
<b>GH30 2</b>	50.1	<b>GH20</b>	86.5	<b>GH1</b>	75.6	<b>GH5 18</b>	49.2	<b>GH20</b>	94.7
<b>GH32</b>	47.0	<b>CBM32</b>	82.3	<b>GH5 44</b>	29.6	<b>GH32</b>	22.3	<b>GH30 2</b>	60.7
<b>GH35</b>	26.3	<b>GH36</b>	57.6	<b>GH95</b>	10.0	<b>GH20</b>	20.2	<b>GH31</b>	31.5
<b>GH1</b>	21.3	<b>GH95</b>	51.7	<b>GH31</b>	9.4	<b>GH59</b>	12.9	<b>GH95</b>	10.4
<b>GH31</b>	17.7	<b>GH1</b>	34.1	<b>GH2</b>	7.9	<b>GH30 9</b>	12.9	<b>GH32</b>	6.0
<b>GH3</b>	15.5	<b>GH42</b>	27.9	<b>GH59</b>	3.0	<b>GH42</b>	8.2	<b>GH35</b>	3.8
<b>GH36</b>	15.1	<b>GH59</b>	16.0	<b>GH30 9</b>	3.0	<b>GH31</b>	5.8	<b>GH3</b>	2.1
<b>GH42</b>	11.6	<b>GH30 9</b>	16.0	<b>GH32</b>	1.0	<b>GH3</b>	0.3	<b>GH5 4</b>	0.0
<b>GH110</b>	8.8	<b>GH3</b>	10.7	<b>GH5 4</b>	0.0	<b>GH5 4</b>	0.0	<b>GH42</b>	-7.1
<b>GH5 44</b>	5.9	<b>GH35</b>	5.6	<b>GH36</b>	-3.4	<b>GH35</b>	-3.0	<b>GH1</b>	-10.0
<b>CBM32</b>	4.0	<b>GH5 4</b>	0.0	<b>CBM32</b>	-13.8	<b>GH36</b>	-4.5	<b>GH30 9</b>	-11.6
<b>GH5 4</b>	0.0	<b>GH30 5</b>	-12.3	<b>GH3</b>	-17.3	<b>GH95</b>	-4.5	<b>GH59</b>	-11.6
<b>GH30 5</b>	-19.1	<b>GH2</b>	-22.6	<b>GH35</b>	-18.1	<b>GH110</b>	-4.6	<b>CBM32</b>	-19.8
<b>GH59</b>	-31.1	<b>GH30 2</b>	-57.1	<b>GH20</b>	-22.9	<b>CBM32</b>	-7.5	<b>GH110</b>	-24.0
<b>GH30 9</b>	-31.1	<b>GH5 18</b>	-63.8	<b>GH110</b>	-26.1	<b>GH2</b>	-16.3	<b>GH36</b>	-30.4
<b>GH95</b>	-61.5	<b>GH31</b>	-117.1	<b>GH42</b>	-26.7	<b>GH30 2</b>	-16.8	<b>GH2</b>	-38.1
<b>GH5 18</b>	-67.9	<b>GH32</b>	-139.4	<b>GH30 2</b>	-33.5	<b>GH5 44</b>	-37.4	<b>GH30 5</b>	-100.5
<b>GH20</b>	-219.0	<b>GH5 44</b>	-139.8	<b>GH30 5</b>	-38.9	<b>GH1</b>	-64.9	<b>GH5 18</b>	-108.6

**Table S6.** Variable importance coefficients of random forest (**RF**) computed to classify metagenome-assembled genomes (**MAGs**) of *B. adolescentis*, *B. bifidum*, *B. breve*, *B. longum* and *B. pseudocatenulatum* based on their glycosidase activities. Importance coefficients were determined by the permutation of the out of-bag-error.

<i>B. adolescentis</i>	Importance		Importance		Importance		Importance		Importance	
	(%)	<i>B. bifidum</i>	(%)	<i>B. breve</i>	(%)	<i>B. longum</i>	(%)	<i>B. pseudocatenulatum</i>	(%)	
<b>GH20</b>	100.0	<b>GH32</b>	100.0	<b>GH5 18</b>	100.0	<b>GH30 5</b>	100.0	<b>GH5 18</b>	100.0	
<b>GH5 18</b>	41.2	<b>GH5 44</b>	94.6	<b>GH1</b>	89.1	<b>GH1</b>	75.6	<b>GH30 5</b>	79.8	
<b>GH30 5</b>	31.2	<b>GH5 18</b>	68.4	<b>GH30 5</b>	49.8	<b>GH5 18</b>	51.2	<b>GH5 44</b>	69.7	
<b>GH110</b>	18.6	<b>GH31</b>	56.9	<b>GH95</b>	31.0	<b>GH110</b>	33.4	<b>GH20</b>	68.4	
<b>GH95</b>	16.5	<b>GH30 5</b>	56.0	<b>GH110</b>	25.9	<b>GH32</b>	30.1	<b>GH110</b>	36.1	
<b>GH2</b>	14.9	<b>GH20</b>	42.9	<b>GH5 44</b>	24.6	<b>GH95</b>	27.6	<b>GH30 2</b>	23.1	
<b>GH32</b>	6.0	<b>GH110</b>	40.4	<b>GH32</b>	20.5	<b>GH30 2</b>	23.2	<b>GH31</b>	14.4	
<b>CBM32</b>	4.2	<b>GH30 2</b>	26.9	<b>GH30 2</b>	14.4	<b>GH5 44</b>	22.6	<b>CBM32</b>	10.5	
<b>GH31</b>	4.2	<b>GH35</b>	4.7	<b>GH2</b>	9.5	<b>GH20</b>	11.5	<b>GH36</b>	5.9	
<b>GH35</b>	2.5	<b>CBM32</b>	2.7	<b>CBM32</b>	6.3	<b>GH36</b>	10.3	<b>GH42</b>	4.2	
<b>GH3</b>	0.0	<b>GH1</b>	1.4	<b>GH35</b>	4.1	<b>GH35</b>	8.3	<b>GH30 9</b>	4.2	
<b>GH5 4</b>	0.0	<b>GH5 4</b>	0.0	<b>GH5 4</b>	0.0	<b>CBM32</b>	6.8	<b>GH3</b>	0.0	
<b>GH30 9</b>	0.0	<b>GH30 9</b>	0.0	<b>GH42</b>	0.0	<b>GH30 9</b>	5.2	<b>GH5 4</b>	0.0	
<b>GH42</b>	0.0	<b>GH42</b>	0.0	<b>GH36</b>	-1.7	<b>GH2</b>	5.0	<b>GH59</b>	0.0	
<b>GH59</b>	0.0	<b>GH59</b>	0.0	<b>GH3</b>	-2.1	<b>GH3</b>	3.3	<b>GH1</b>	-0.1	
<b>GH5 44</b>	-0.4	<b>GH2</b>	-1.7	<b>GH20</b>	-2.2	<b>GH59</b>	2.3	<b>GH32</b>	-3.1	
<b>GH36</b>	-0.5	<b>GH3</b>	-9.3	<b>GH31</b>	-7.4	<b>GH5 4</b>	0.0	<b>GH2</b>	-6.4	
<b>GH30 2</b>	-3.9	<b>GH95</b>	-9.6	<b>GH30 9</b>	-8.8	<b>GH42</b>	0.0	<b>GH95</b>	-11.2	
<b>GH1</b>	-9.3	<b>GH36</b>	-10.2	<b>GH59</b>	-9.2	<b>GH31</b>	-0.7	<b>GH35</b>	-16.8	

**Table S7.** Variable importance coefficients of Generalized Linear Model Elastic-net (**GLMNET**) computed to classify metagenome-assembled genomes (**MAGs**) of *B. adolescentis*, *B. bifidum*, *B. breve*, *B. longum* and *B. pseudocatenulatum* based on their glycosidase activities. Importance coefficients were determined by calculating the area under the ROC (Receiver Operating Characteristic) curve.

<i>B. adolescentis</i>	Importance (%)	<i>B. bifidum</i>	Importance (%)	<i>B. breve</i>	Importance (%)	<i>B. longum</i>	Importance (%)	<i>B. pseudocatenulatum</i>	Importance (%)
<b>GH20</b>	100.0	<b>GH32</b>	100.0	<b>GH5 18</b>	100.0	<b>GH30 5</b>	100.0	<b>GH5 18</b>	100.0
<b>GH35</b>	28.4	<b>GH110</b>	84.0	<b>GH1</b>	70.1	<b>GH1</b>	53.4	<b>GH20</b>	96.8
<b>GH30 2</b>	25.2	<b>GH5 44</b>	74.2	<b>GH20</b>	22.0	<b>GH5 18</b>	44.0	<b>GH30 2</b>	74.1
<b>GH2</b>	23.0	<b>GH31</b>	65.6	<b>GH5 44</b>	18.5	<b>GH30 2</b>	25.6	<b>GH5 44</b>	73.7
<b>GH5 18</b>	18.6	<b>CBM32</b>	50.9	<b>GH95</b>	16.1	<b>GH5 44</b>	16.3	<b>GH30 5</b>	31.9
<b>GH95</b>	8.0	<b>GH42</b>	18.1	<b>GH30 2</b>	2.9	<b>GH2</b>	9.3	<b>GH36</b>	16.8
<b>CBM32</b>	0.0	<b>GH20</b>	8.0	<b>CBM32</b>	0.0	<b>GH30 9</b>	6.1	<b>GH31</b>	13.5
<b>GH1</b>	0.0	<b>GH1</b>	0.0	<b>GH2</b>	0.0	<b>GH59</b>	5.9	<b>GH2</b>	13.0
<b>GH3</b>	0.0	<b>GH2</b>	0.0	<b>GH3</b>	0.0	<b>GH20</b>	1.2	<b>CBM32</b>	0.0
<b>GH5 4</b>	0.0	<b>GH3</b>	0.0	<b>GH5 4</b>	0.0	<b>GH32</b>	1.0	<b>GH1</b>	0.0
<b>GH5 44</b>	0.0	<b>GH5 4</b>	0.0	<b>GH30 5</b>	0.0	<b>CBM32</b>	0.0	<b>GH3</b>	0.0
<b>GH30 5</b>	0.0	<b>GH5 18</b>	0.0	<b>GH30 9</b>	0.0	<b>GH3</b>	0.0	<b>GH5 4</b>	0.0
<b>GH30 9</b>	0.0	<b>GH30 2</b>	0.0	<b>GH31</b>	0.0	<b>GH5 4</b>	0.0	<b>GH30 9</b>	0.0
<b>GH31</b>	0.0	<b>GH30 5</b>	0.0	<b>GH32</b>	0.0	<b>GH31</b>	0.0	<b>GH32</b>	0.0
<b>GH32</b>	0.0	<b>GH30 9</b>	0.0	<b>GH35</b>	0.0	<b>GH35</b>	0.0	<b>GH35</b>	0.0
<b>GH36</b>	0.0	<b>GH35</b>	0.0	<b>GH36</b>	0.0	<b>GH36</b>	0.0	<b>GH42</b>	0.0
<b>GH42</b>	0.0	<b>GH36</b>	0.0	<b>GH42</b>	0.0	<b>GH42</b>	0.0	<b>GH59</b>	0.0
<b>GH59</b>	0.0	<b>GH59</b>	0.0	<b>GH59</b>	0.0	<b>GH95</b>	0.0	<b>GH95</b>	0.0
<b>GH110</b>	0.0	<b>GH95</b>	0.0	<b>GH110</b>	0.0	<b>GH110</b>	0.0	<b>GH110</b>	0.0



**Figure S1.** Comparative account of different three algorithms used for classification of metagenome-assembled genomes (MAGs) of *B. adolescentis*, *B. bifidum*, *B. breve*, *B. longum* and *B. pseudocatenulatum* based on their glycosidase activities: random forest (**RF**), Generalized Linear Model Elastic-net (**GLMNET**) and artificial neural network (**ANN**). Differences between models were calculated via their resampling distributions (number of resamples = 10). <sup>a</sup> No statistically significant ( $p > 0.05$ ) differences between models were found.