

Automated cell treatment for competence and transformation of *Escherichia coli* in high throughput

Sebastian Hans, Mathias Gimpel, Florian Glauche, Peter Neubauer and M. Nicolas Cruz-Bournazou *


















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























Source Code S1: Hamilton Script Overview

	Method
1	 Assignment 'main_runID' = '325'
2	 HSL code. main_current_date_str = TimGetFormattedDate("%Y-%m-%d") + " " + TimGetFormattedTime("%H %M %S");
3	 Assignment 'main_cultivation_dilution' = '10'
4	 Assignment 'main_cultivation_cycletime_min' = '60'
5	 Assignment 'main_incubation_onIce' = '1800'
6	 Assignment 'main_Operator_Number' = "491775724287"
7	 Grouping Konstanten
50	 Grouping Initialize
61	 Grouping Cultivation
138	 Grouping Filtrieren
150	 Grouping Resuspendiren
159	 Grouping Incubation on ice
164	 Grouping Transformation
174	 Grouping Incubation on Ice & Prepare fresh media plate
181	 Grouping Heatshock
189	 Grouping Cultivation2
198	 Grouping Ausplätieren
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Source Code S2: Hamilton Script detail




















		Method
1	X=0	Assignment main_runid' = '325'
2		HSL code main_current_date_str = TimeGetFormattedDate("%Y-%m-%d") + " _" + TimeGetFormattedTime("%H_%M_%S");
3	X=0	Assignment main_cultivation_dilution' = '10'
4	X=0	Assignment main_cultivation_cycletime_min' = '60'
5	X=0	Assignment main_incubation_once' = '1800'
6	X=0	Assignment main_Operator_Number' = "'491775724287'"
7		Grouping Konstanten
8	X=0	Assignment main_Gen5_PlateCounter' = '2'
9	X=0	Assignment main_PreCulture_KryoVol' = '5'
10		Comment <main_PreCulture_IncubationTime in Hours>
11		Array: Declare / Set Size Set array 'main_PreCulture_IncubationTime' to empty size.
12	X=0	Assignment main_PreCulture_dilution' = '10'
13	X=0	Assignment main_cultivationVol' = '170'
14		HSL code // Berechne PreCult Media Volume main_PreCulture_MediaVol = main_cultivationVol - main_PreCulture_KryoVol; // Teile PreCult Cultivation Time in 3 Intervalle ein ($1\frac{1}{3}, 5\frac{1}{3}, 5\frac{1}{3}$) // Umrechnen der Kultivierungszeit von Stunden in Sekunden main_PreCulture_IncubationTime.AddAsLast(3600); main_PreCulture_IncubationTime.AddAsLast(3600); main_PreCulture_IncubationTime.AddAsLast(3600); main_PreCulture_IncubationTime.AddAsLast(3600); main_PreCulture_IncubationTime.AddAsLast(3600); main_PreCulture_IncubationTime.AddAsLast(3600); main_PreCulture_IncubationTime.AddAsLast(3600); main_PreCulture_IncubationTime.AddAsLast(3600); main_PreCulture_IncubationTime.Temp = 0; main_PreCulture_LoopNum = main_PreCulture_IncubationTime.GetSize(); // Rechne Verdünnungen für die ODMessung aus main_PreCulture_VolSample = main_cultivationVol / main_PreCulture_dilution; main_PreCulture_VolInACl = main_cultivationVol - main_PreCulture_VolSample;
15	X=0	Assignment main_reader_RowName' = "OD600"
16		Array: Declare / Set Size Set array 'main_reader_PreCultureValues_1' to empty size.
17		Array: Declare / Set Size Set array 'main_reader_PreCultureValues_2' to empty size.
18		Array: Declare / Set Size Set array 'main_reader_PreCultureValues_3' to empty size.
19	X=0	Assignment main_OD600_threshold' = '0,041'
20	X=0	Assignment main_OD600_correctionFactor' = '2'

		Method
21		X=0 Assignment 'main_cultivation_VolPreCulture' = '10'
22		HSL code main_cultivation_VolMedium = main_cultivationVol - main_cultivation_VolPreCulture;
23		X=0 Assignment 'main_cultivation_VolSample' = '20'
24		HSL code main_cultivation_VolNaCl = 200 - main_cultivation_VolSample;
25		X=0 Assignment 'main_cultivation_slot' = '4'
26		X=0 Assignment 'main_cultivation_temp' = '37'
27		X=0 Assignment 'main_cultivation_speed' = '1000'
28		X=0 Assignment 'main_reader_path2resultsFolder' = "C:\Dokumente und Einstellungen\Wpeter Neubauer\Eigene Dateien\Sebastian\Robot"
29		X=0 Assignment 'main_reader_path2experimentFolder' = "C:\Programme\BioTek\Gen5 1.09\Experiments\Sebastian\Robot"
30		HSL code Shell("C:\WINDOWS\system32\cmd.exe /C mkdir %~ + main_reader_path2resultsFolder + "%~ + main_current_date_str + "%", 2, 1) Shell("C:\WINDOWS\system32\cmd.exe /C mkdir %~ + main_reader_path2experimentFolder + "%~ + main_current_date_str + "%", 2, 1); Shell("C:\WINDOWS\system32\cmd.exe /C copy %~ + main_reader_path2experimentFolder + "%~ + _default_OD600.xpt" + " %~ + main_reader_path2experimentFolder + "%~ + main_current_date_str + "%~ + Assignment 'main_cultivation_abort' = '0'
31		X=0 Assignment 'main_filter_VolFromCultur' = '180'
32		X=0 Assignment 'main_filter_VolFromCultur' = '180'
33		HSL code // Rechne Cydetime in Sec main_cultivation_cydetime_sec = main_cultivation_cydetime_min * 60; // Ermittle wieviele Stämme vorhanden sind main_numStrains = ML_STAR_Res_Kryos.GetTotal();
34		X=0 Assignment 'main_pump_ID' = '1'
35		X=0 Assignment 'main_pump_COMP ort' = '3'
36		X=0 Assignment 'main_pump_dPressure' = '200'
37		X=0 Assignment 'main_pump_fresholdPressure' = '10'
38		X=0 Assignment 'main_filter_duration' = '20'
39		X=0 Assignment 'main_wash_Volume' = '200'
40		X=0 Assignment 'main_wash_cycles' = '3'
41		X=0 Assignment 'main_resuspend_Vol' = '150'
42		X=0 Assignment 'main_incubation_time1' = '7200'
43		X=0 Assignment 'main_dha_Vol' = '2'

	Method
44	 X=0 Assignment 'main_incubation_heatShock' = '120'
45	 X=0 Assignment 'main_cultivation2_Vol' = '150'
46	 HSL code. main_cultivation2_VolKultur = 200 - main_cultivation2_Vol;
47	 X=0 Assignment 'main_cultivation2_time' = '3600'
48	 X=0 Assignment 'main_plating_vol' = '200'
49	 Grouping
50	 Grouping Initialize
51	 If, Else (main_runID is NOT equal to 0)
52	 iLAB_connect of BVTlib_iLab_connector iLAB_connect()
53	 getBioreactorIDs of BVTlib_iLab_connector main_biolD_array = getBioreactorIDs(main_runID)
54	 X=0 Assignment 'main_biolD' = 'main_biolD_array[1]'
55	 End If
56	 HSL code. main_StatusLTU = MessageBox("Is it Plate of the LTU on the site of the Hamilton?" "Status LTU": 4) if (main_StatusLTU == 6) { Position_Tecan(); }
57	 Initialize (Single Step) on ML_STAR Always initialize: Off 3 return value(s)
58	 Initialize_fame of unifiedlibrary UNIFIEDLIBRARY::fame_transport(ML_STAR, main_cultivation_temp, 2, main_cultivation_speed, ML_STAR_Plate_Cultivation)
59	 BVSInitialize of HSLVacuuBrandPump HSLStarBVSlib::BVSInitialize(main_pump_ID, main_pump_COMPort)
60	 Grouping
61	 Grouping Cultivation
62	 Comment <Prepair PreCulture Plate>
63	 Sub_01_AddFromContainer_sterilTips of KompetenteZellen_v0.0.1 Sub_01_AddFromContainer_sterilTips(ML_STAR_Res_Medium, ML_STAR_Plate_Preculture, main_PreCulture_MediaVol)
64	 Sub_02_AddFromSequence of KompetenteZellen_v0.0.1 Sub_02_AddFromSequence(ML_STAR_Res_Kryos, ML_STAR_Plate_Preculture, main_PreCulture_KryoVol)
65	 Comment <Start Preculture>
66	 fame_transport of unifiedlibrary UNIFIEDLIBRARY::fame_transport(2, ML_STAR_Plate_Preculture, main_cultivation_slot, main_cultivation_temp, main_cultivation_speed, ML_STAR)
67	 Timer: Start Start timer 'main_timer_preculture', set to relative time: '0' [s]



















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	Method
68	 Cleaning_0050_Tips of SM_SH_Cleaning SM_SH_CLEANING::cleaning_0050_Tips(ML_STAR, ML_STAR_Tips_0050_1, ML_STAR_Res_ETOH, ML_STAR_Res_ETOH_Lid, ML_STAR_Res_ETOH_Lid_Storage, sendWhatsApp of SM_Gluocse
69	 SM_GLUOCSE::sendWhatsApp(main_Operator_Number, "First Timer is started ...")
70	 Loop 'main_PreCulture_LoopNum' times 'main_PreCulture_Loop' used as loop counter variable
71	 Grouping Timer
72	 Timer: Wait for Wait for timer 'main_timer_preculture', showtimer display, is stoppable
73	 Timer: Start Start timer 'main_timer_preculture', set to relative time: 'main_PreCulture_IncubationTime[main_PreCulture_Loop] [s]
74	 sendWhatsApp of SM_Gluocse SM_GLUOCSE::sendWhatsApp(main_Operator_Number, "Next Timer is started ...")
75	 Grouping
76	 Comment <Take newPlate>
77	 1000ul Channel CO-RE Grip Get Plate (Single Step) on ML_STAR Transport mode: (0) Plate only, Sequence: ML_STAR_Plate_Storage, Sequence counting: (1) Automatic, Channel to be used: 8 3 return value(s)
78	 1000ul Channel CO-RE Grip Place Plate (Single Step) on ML_STAR Transport mode: (0) Plate only, Sequence: ML_STAR_Plate_working, Sequence counting: (0) Manually, Eject tool: (1) Yes 3 return value(s)
79	 Sub_01_AddFromContainer of KompetenteZellen_v0.0.1 Sub_01_AddFromContainer(ML_STAR_Res_NaCl, ML_STAR_Plate_working, main_PreCulture_VolNaCl)
80	 Comment <Hole Platte aus dem Inkubator>
81	 fame_transport of unifiedlibrary UNIFIEDLIBRARY::fame_transport(1, ML_STAR_Plate_FromIncubator, main_cultivation_slot, main_cultivation_temp, main_cultivation_speed, ML_STAR)
82	 Comment <Messe OD>
83	 Sub_02_AddFromSequence_96 of KompetenteZellen_v1.1.0 Sub_02_AddFromSequence_96(ML_STAR_Plate_FromIncubator, ML_STAR_Plate_working, main_PreCulture_VolSample)
84	 Comment <Lege Platte zurück in den Inkubator>
85	 fame_transport of unifiedlibrary UNIFIEDLIBRARY::fame_transport(2, ML_STAR_Plate_FromIncubator, main_cultivation_slot, main_cultivation_temp, main_cultivation_speed, ML_STAR)
86	 Grouping ReaderMessung und Daten erreichen

























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	Method
87	 <pre> HSL code //Erstelle Path zum Experiment main_reader_path2experiment = main_reader_path2experimentFolder + "W" + main_current_date_str + "W" + main_current_date_str + "_OD600.xpt"; //Erstelle Path zur Exceldatei mit den Ergebnissen main_reader_path2results = main_reader_path2resultsFolder + "W" + main_current_date_str; main_reader_filenameResults = main_current_date_str + "_OD600_Preculture_" + IStr(main_PreCulture_Loop); main_reader_TabName = "Plate" + IStr(main_Gen5_PlateCounter) + "- Data"; main_Gen5_PlateCounter++; main_reader_filenameReturn = "C:\Dokumente und Einstellungen\Walter Neubauer\Meine Dateien\Sebastian\RobotW" + main_current_date_str + "W" + main_reader_filenameResults + ".xls"; Readermessung of unifiedlibrary UNIFIEDLIBRARY:Readermessung(ML_STAR, HxGen5, ML_STAR_Plate_working, main_reader_path2experiment, main_reader_path2results, main_reader_filenameResults, main_cultivation_dilution, main_cultivation_start, main_reader_filenameReturn) Comment <Werte OD aus> readRow of BVTlib_ExcelImport main_reader_results = readRow(main_reader_filenameReturn, main_reader_TabName, main_reader_RowName) subArray of BVTlib_ExcelImport main_reader_resultsSub = subArray(main_reader_results, 1, main_numStrains) subtractArray of BVTlib_ExcelImport main_reader_resultsSub = subtractArray(main_reader_resultsSub, main_OD600_threshold) multiplyArray of BVTlib_ExcelImport main_reader_resultsSub = multiplyArray(main_reader_resultsSub, main_PreCulture_dilution) multiplyArray of BVTlib_ExcelImport main_reader_PreCultureValues = multiplyArray(main_reader_resultsSub, main_OD600_correctionFactor) HSL code main_TimeStamp_IIab = TimGetFormattedDate("%Y-%m-%d") + " " + TimGetFormattedTime("%H:%M:%S"); If, Else (main_runID is NOT equal to 0) sendData_long of BVTlib_IIab_connector sendData_long(main_runID, main_biolD, "OD600", main_reader_PreCultureValues, 1, main_TimeStamp_IIab, main_PreCulture_dilution) End If arrayMinValue of BVTlib_ExcelImport main_ODmin = arrayMinValue(main_reader_PreCultureValues) If, Else (main_ODmin is greater than OR equal to 0,75) Loop: Break End If </pre>
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	Method
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	Method
151	HSL code. Trace("Resuspend - Loop"); ML_STAR_Plate_Filter.SetCurrentPosition(1); main_resuspend_LoopCounter = main_numStrains/8;
152	Loop 'main_resuspend_LoopCounter' times 'main_Loop_resuspend' used as loop counter variable
153	1000µl Channel Aspirate on ML_STAR Sequence: ML_STAR_Res_CaCl2, Volume [µl]: main_resuspend_Vol 0 return value(s)
154	1000µl Channel Dispense on ML_STAR Sequence: ML_STAR_Plate_Filter, Volume [µl]: main_resuspend_Vol 0 return value(s)
155	1000µl Channel Aspirate on ML_STAR Sequence: ML_STAR_Plate_Filter, Volume [µl]: main_resuspend_Vol 0 return value(s)
156	1000µl Channel Dispense on ML_STAR Sequence: ML_STAR_Plate_PCR, Volume [µl]: Remaining volume inclusive blowout air 0 return value(s)
157	End Loop
158	Grouping
159	Grouping Incubation on ice
160	Timer: Start Start timer 'main_timer_incubation', set to relative time: 'main_incubation_time1' [s]
161	iSWAP Transport on ML_STAR Transport labware from 'ML_STAR_Plate_FromIncubator' to 'ML_STAR_platewaste' 1 return value(s)
162	Timer: Wait for Wait for timer 'main_timer_incubation', show timer display, is stoppable timer.
163	Grouping
164	Grouping Transformation
165	HSL code. Trace("Starting Loop - Transformation"); ML_STAR_Plate_PCR.SetCurrentPosition(1); Sub05_TransportLid of KompetentZellen_v0.0.1 Sub05_TransportLid(ML_STAR_Res_Plasmid_Lid, ML_STAR_LidStorage)
166	Loop 'main_resuspend_LoopCounter' times 'main_Loop_DNA' used as loop counter variable
168	1000µl Channel Aspirate on ML_STAR Sequence: ML_STAR_Res_Plasmid, Volume [µl]: main_dna_Vol 0 return value(s)
169	1000µl Channel Dispense on ML_STAR Sequence: ML_STAR_Plate_PCR, Volume [µl]: Remaining volume inclusive blowout air 0 return value(s)
170	End Loop
171	Sub05_TransportLid of KompetentZellen_v0.0.1 Sub05_TransportLid(ML_STAR_LidStorage, ML_STAR_Res_Plasmid_Lid)
172	HSL code. Trace("Transformation done");
173	Grouping

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	Method
174	Grouping Incubation on ice & Prepair fresh media plate
175	HSL code Trace("Incubation on ice");
176	Timer: Start Start timer 'main_timer_incubationOnIce', set to relative time: 'main_incubation_onice' [s]
177	Sub_01_AddFromContainer_sterTips of KompetentZellen_v0.0.1 Sub_01_AddFromContainer_sterTips(ML_STAR_Res_Medium, ML_STAR_Plate_Cultivation_2, main_cultivation2_Vol)
178	Timer: Wait for Wait for timer 'main_timer_incubationOnIce', show timer display, is stoppable timer.
179	HSL code. Trace("Incubation on ice done");
180	Grouping
181	Grouping Heatshock
182	HSL code. Trace("Starting Heatshock");
183	iSWAP Transport on ML_STAR Transport labware from 'ML_STAR_Plate_PCR' to 'ML_STAR_Plate_PCR_heatshock' 1 return value(s)
184	Timer: Start Start timer 'main_timer_incubationHeatshock', set to relative time: 'main_incubation_heatShock' [s]
185	Timer: Wait for Wait for timer 'main_timer_incubationHeatshock', show timer display, is stoppable timer.
186	iSWAP Transport on ML_STAR Transport labware from 'ML_STAR_Plate_PCR_heatshock' to 'ML_STAR_Plate_PCR' 1 return value(s)
187	HSL code. Trace("Heatshock Done");
188	Grouping
189	Grouping Cultivation2
190	HSL code. Trace("Starting Cultivation 2");
191	Sub_02_AddFromSequence of KompetentZellen_v0.0.1 Sub_02_AddFromSequence(ML_STAR_Plate_PCR, ML_STAR_Plate_Cultivation_2, main_cultivation2_VolKultur)
192	fame_transport of unifiedlibrary UNIFIEDLIBRARY:fame_transport(2, ML_STAR_Plate_Cultivation_2, main_cultivation_slot, main_cultivation_temp, main_cultivation_speed, ML_STAR)
193	Timer: Start Start timer 'main_timer_cultivation2', set to relative time: 'main_cultivation2_time' [s]
194	Timer: Wait for Wait for timer 'main_timer_cultivation2', show timer display, is stoppable timer.
195	fame_transport of unifiedlibrary UNIFIEDLIBRARY:fame_transport(1, ML_STAR_Plate_FromIncubator, main_cultivation_slot, main_cultivation_temp, main_cultivation_speed, ML_STAR)
196	HSL code Trace("Cultivation 2 Done");
197	Grouping
198	Grouping Ausplattieren

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	Method
199	Sequence: Set Current Position current position of sequence 'ML_STAR_Plate_Cultivation_2' = '1'
200	HSL code. Trace("Start Plating"); main_jemp = 0; // Errechne wie viele Platten ausplattiert werden muessen main_platting_numPlates = main_numStrains%6; Assignment 'main_CyomatPos' = '148'
201	X=0
202	Loop 'main_platting_numPlates' times 'main_loop_platting' used as loop counter variable
203	Grouping Get Plate from cyomat
204	HSL code. sShellCmd = "C:\Dokumente und Einstellungen\Peter Neubauer\Eigene Dateien\Sebastian\Programms\Remote_Trick\ClientConsole.exe C:\Users\BVT-Administrator\Anaconda3\python.exe C:\Tecan_Scripts\Remoting\CyomatAgarplate.py -m 1 -s " + IStr(main_CyomatPos); Trace(sShellCmd); Shell(sShellCmd, 2, 1); Position_Hamilton of BVTlib_LTU Position_Hamilton()
205	iSWAP Transport on ML_STAR Transport labware from 'ML_STAR_move_Tecan' to 'ML_STAR_Plate_AgarTransport' 1 return value(s).
206	Grouping
207	Sequence: Set Current Position current position of sequence 'ML_STAR_LidAgar' = '1'
208	_Sub05_TransportLid_Agar of Kompetente Zellen_v0.0.1 _Sub05_TransportLid_Agar(ML_STAR_Plate_Agar_StorePlated_Lids, ML_STAR_LidAgar)
209	1000µl Channel CO-RE Grip Get Plate (Single Step) on ML_STAR Transport mode: (0) Plate only, Sequence: ML_STAR_Plate_Agar_StorePlated, Sequence counting: (0) Manually, Channel to be used: 8 3 return value(s).
210	1000µl Channel CO-RE Grip Place Plate (Single Step) on ML_STAR Transport mode: (0) Plate only, Sequence: ML_STAR_Plate_Agar, Sequence counting: (0) Manually, Eject tool: (0) No 3 return value(s).
211	1000µl Channel Aspirate on ML_STAR Sequence: ML_STAR_Res_Medium, Volume [µl]: 190 0 return value(s).
212	1000µl Channel Aspirate on ML_STAR Sequence: ML_STAR_Res_Medium, Volume [µl]: 10 0 return value(s).
213	1000µl Channel Aspirate on ML_STAR Sequence: ML_STAR_Plate_FromIncubator, Volume [µl]: 50 0 return value(s).
214	1000µl Channel Dispense on ML_STAR Sequence: ML_STAR_Plate_Agar, Volume [µl]: Remaining volume inclusive blowout air 0 return value(s).
215	Sub_05_Shaking of Kompetente Zellen_v0.0.1 Sub_05_Shaking()
216	Sequence: Set Current Position current position of sequence 'ML_STAR_Plate_Agar' = '1'
217	

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	Method
218	1000µl Channel CO-RE Grip Get Plate (Single Step) on ML_STAR Transport mode: (0) Plate only, Sequence: ML_STAR_Plate_Agar, Sequence counting: (0) Manually, Channel to be used: 8 3 return value(s).
219	1000µl Channel CO-RE Grip Place Plate (Single Step) on ML_STAR Transport mode: (0) Plate only, Sequence: ML_STAR_Plate_Agar_StorePlated, Sequence counting: (0) Manually, Eject tool: (0) No 3 return value(s).
220	Sequence: Set Current Position current position of sequence 'ML_STAR_LidAgar' = '1'
221	_Sub05_TransportLid_Agar of Kompetente Zellen_v1.1.0 _Sub05_TransportLid_Agar(ML_STAR_LidAgar, ML_STAR_Plate_Agar_StorePlated_Lids)
222	Grouping Transfer Agarplate to Cyomat
223	iSWAP Transport on ML_STAR Transport labware from 'ML_STAR_Plate_AgarTransport' to 'ML_STAR_move_Tecan' 1 return value(s).
224	Position_Tecan of BVTlib_LTU Position_Tecan()
225	HSL code. sShellCmd = "C:\Dokumente und Einstellungen\Peter Neubauer\Eigene Dateien\Sebastian\Programms\Remote_Trick\ClientConsole.exe C:\Users\BVT-Administrator\Anaconda3\python.exe C:\Tecan_Scripts\Remoting\CyomatAgarplate.py -m 0 -s " + IStr(main_CyomatPos); Trace(sShellCmd); Shell(sShellCmd, 2, 1); Assignment with Calculation 'main_CyomatPos' = 'main_CyomatPos' + '1'
226	X=i+1
227	Grouping
228	End Loop
229	HSL code. Trace("Plating done");
230	Grouping
231	

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Figure S3: Tecan LHS



Figure 1: Second used Liquid handling station (LHS). Plates are stored at the incubator on the right site of the LHS. On a command of the Hamilton LHS a plate is moved from the incubator to the Transfer unit. A second command was used to return the plate from the transfer unit into the incubator.

Source Code S4 MATLAB Source Code

```
function DiluteForChemostat(iRunID)
sim = 0;
%% Ziel OD:
fTragetOD = 0.8;
iCultivationVol = 170; % [µL]

%% Connect to ilab
if ispc % check if running on Windows
    ilab = actxserver('BVT_iLabDriver.ilab_net_class');
% %    ilab = actxserver('ILAB_COM_BVT.ILAB_COM_BVT');
else
    ilab = iLab_driver_universal;
end
ilab.SQL_Close;
ilab.SQL_Connect;

%    ilab.run_id = iRunID;
%    ilab = ilab.get_bioID(iRunID);
%    iExpID = ilab.getExpIDByBioID();
%    iProfID = ilab.getProfIDByBioID();
iBioID = ilab.getBioreactorIDs(iRunID);
iBioID = iBioID(1);
iExpID = ilab.getExperimentIDs(iBioID);
iProfID = ilab.getProfilIDs(iBioID);
iNumOfExperiments = length(iExpID);

%% Calling Measurements
fOD600 = ilab.getExperimentMeasurements(iExpID,'OD600');
fTime = fOD600(:,1)/3600;
fOD600 = fOD600(:,2:end);

iSizeOfData = size(fOD600);
iNumOfMeasurements = iSizeOfData(1);

%% get last Setpoints to compute the last dilution factor
fDilutionOld = ones(1,length(iExpID));
if iNumOfMeasurements > 1
    for ci = 1:iNumOfExperiments
        fDilutionOld(ci) = (iCultivationVol-ilab.SetpointGetCurrent(iProfID(ci),
'Puls_Medium'))/iCultivationVol;
    end
    % Berechne vorletzte OD neu:
    fOD600(end-1,:) = fOD600(end-1,:).*fDilutionOld;
end

%% create timestamp:
iTime = ilab.getCultivationTime(iRunID);

%% Compute µ / OD set
% Erstelle Array's
fODNext = zeros(1,length(iExpID));
fDilution = ones(1,length(iExpID));
fVolRemain = zeros(1,length(iExpID));
fVolAdd = zeros(1,length(iExpID));
fVolRemove = zeros(1,length(iExpID));

fMu = zeros(iNumOfMeasurements-1, length(iExpID));
if ~(iNumOfMeasurements > 1) % if no µ values can be calculated
    for ci = 1:length(iExpID)
        fVolAdd(ci) = 20;
        fVolRemove(ci) = 0;
        if ~sim
            ilab.SetpointSet(int32(iProfID(ci)), 'Puls_Medium', iTime, fVolAdd(ci));
            ilab.SetpointSet(int32(iProfID(ci)), 'Probe_Volume', iTime,
fVolRemove(ci));
        end
    end
else % normal process ...
    for ci = 2:iNumOfMeasurements
        for cj = 1:length(iExpID)
```



```

        fMu(ci, cj) = (log(fOD600(ci,cj))-log(fOD600(ci-1,cj))) / (fTime(ci)-
fTime(ci-1));
    end
end
for ci = 1:length(iExpID)
    fODNext(ci) = fTragetOD / exp(fMu(end,ci) * (fTime(end)-fTime(end-1)));
    if fODNext(ci)~=0
        fDilution(ci) = fODNext(ci) / fOD600(end, ci);
    end
    fVolRemain(ci) = fDilution(ci) * iCultivationVol;
    if fVolRemain(ci) > (iCultivationVol - 20)
        fVolRemain(ci) = (iCultivationVol - 20);
    elseif fVolRemain(ci) < 45
        fVolRemain(ci) = 45;
    end
    % Compute Volumes
    fVolRemove(ci) = (iCultivationVol - 20) - fVolRemain(ci);
    fVolAdd(ci) = iCultivationVol - fVolRemain(ci);
    %% Write data to database
    if ~sim
        ilab.SetpointSet(int32(iProfID(ci)), 'Puls_Medium', iTime, fVolAdd(ci));
        ilab.SetpointSet(int32(iProfID(ci)), 'Probe_Volume', iTime,
fVolRemove(ci));
    end
end
end
end
end

```