

**Table S1.** Bacterial strains and plasmids used in this study

Bacterial strain or plasmid	Relevant characteristics	Reference or source
<i>E. coli</i> strain		
DH5 $\alpha$	<i>F</i> <sup>-</sup> $\lambda$ : $\phi$ 80dLacZ $\Delta$ M15 $\Delta$ ( <i>lacZYA-argF</i> )U169 <i>recA1 endA1 hsdR17</i> (rK <sup>-</sup> mK <sup>+</sup> ) <i>supE44 thi-1 gyrA relA1 thi pro hsdR<sup>-</sup> hsdM<sup>+</sup> recA</i> [chr::RP4-2-Tc::Mu-Km::Tn7]	Takara bio, Kusatsu, Japan
S17-1		Schafer et al. (1994) [29]
<i>P. cannabina</i> pv. <i>alisalensis</i> ( <i>Pcal</i> )		
Isolate KB211	Wild type, Rif <sup>r</sup>	Nagano vegetable and ornamental crops experiment station
<i>hexR</i> mutant	<i>Pcal hexR</i> mutant, Rif <sup>r</sup> , Km <sup>r</sup>	Sakata et al. (2019) [20]
<i>hexR</i> -complemented	<i>Pcal hexR</i> mutant containing pDSKG- <i>hexR</i> , Rif <sup>r</sup> , Km <sup>r</sup> , Gn <sup>r</sup>	This work
<i>P. syringae</i> pv. <i>tomato</i> ( <i>Pst</i> )		
Isolate DC3000	Wild type, Rif <sup>r</sup>	From Dr. Katagiri
$\Delta$ <i>hexR</i>	<i>Pst hexR</i> mutant, Rif <sup>r</sup>	This study
Plasmid		
pGEM-T Easy	Cloning vector, Amp <sup>r</sup>	Promega, Tokyo, japan
pK18 <i>mobsacB</i>	Small mobilizable vector, Km <sup>r</sup> , sucrose sensitive ( <i>sacB</i> )	Schafer et al. (1994) [29]
pDSKG- <i>hexR</i>	The vector containing constitutive <i>psbA</i> promoter and <i>hexR</i> gene inserted into pDSKG, Gn <sup>r</sup>	This work

Amp<sup>r</sup> ampicillin resistance, Gn<sup>r</sup> gentamicin resistance, Km<sup>r</sup> kanamycin resistance, Rif<sup>r</sup> rifampicin resistance

**Table S2.** Primer sets used in this study

Gene	Primer name	Primer usage	Primer sequence	Reference
<i>Pst hexR</i>	PSPTO_1299-1	Deletion mutagenesis of <i>Pst hexR</i>	GGCCAGTGCCTTGAGCACTT	This study
<i>Pst hexR</i>	PSPTO_1299-2	Deletion mutagenesis of <i>Pst hexR</i>	TGCAACGCTTCTTCCGATCC	This study
<i>Pst hexR</i>	PSPTO_1299-3	Deletion mutagenesis of <i>Pst hexR</i>	CGGGATCCAGGTTTCTTACGCGGTCCAT	This study
<i>Pst hexR</i>	PSPTO_1299-4	Deletion mutagenesis of <i>Pst hexR</i>	CGGGATCCGATCGAGGATCAGGGCTGA	This study
<i>Pcal hrpL</i>	hrpL-FW	Expression analysis on <i>Pcal</i>	CATGCCAGTAAACCGCAAAC	Sakata et al. (2021) [36]
<i>Pcal hrpL</i>	hrpL-RV	Expression analysis on <i>Pcal</i>	GTCTTCCCAGCTTTCCTGATAA	Sakata et al. (2021) [36]
<i>Pcal avrPto</i>	avrPto-FW	Expression analysis on <i>Pcal</i>	CATCGATACCTGACCAACGATAC	This study
<i>Pcal avrPto</i>	avrPto-RV	Expression analysis on <i>Pcal</i>	CTCAGTCAGCTTGGTGCTAAT	This study
<i>Pcal hopM1</i>	hopM1-FW	Expression analysis on <i>Pcal</i>	GGTGCCGATGAAGGCTATT	This study
<i>Pcal hopM1</i>	hopM1-RV	Expression analysis on <i>Pcal</i>	TTTCGACGGACGCTTTGT	This study
<i>Pcal avrE1</i>	avrE1-FW	Expression analysis on <i>Pcal</i>	GACAATCAGGGCAGGCTTTA	This study
<i>Pcal avrE1</i>	avrE1-RV	Expression analysis on <i>Pcal</i>	TGACCTGCCAAGGTGTAATG	This study
<i>Pcal corR</i>	corR-FW	Expression analysis on <i>Pcal</i>	GGATCGAACGCTGGCAGATA	Sakata et al. (2021) [36]
<i>Pcal corR</i>	corR-RV	Expression analysis on <i>Pcal</i>	GTCTGCTCATGAGTCGCTT	Sakata et al. (2021) [36]
<i>Pcal cfl</i>	cfl-FW	Expression analysis on <i>Pcal</i>	GAACTGGTGGCGTTGTACTAT	Sakata et al. (2021) [36]
<i>Pcal cfl</i>	cfl-RV	Expression analysis on <i>Pcal</i>	GTGGAGCAGATGCTCAATTTTC	Sakata et al. (2021) [36]
<i>Pcal cmaA</i>	cmaA-FW	Expression analysis on <i>Pcal</i>	AAAGCCTACCGCCGATTT	Sakata et al. (2021) [36]
<i>Pcal cmaA</i>	cmaA-RV	Expression analysis on <i>Pcal</i>	CGTCTGGAGCTGTTGATAAGT	Sakata et al. (2021) [36]
<i>Pcal oprF</i>	oprF-FW	Expression analysis on <i>Pcal</i>	GGCTTGGCCATTGGTACTAT	Sakata et al. (2021) [36]
<i>Pcal oprF</i>	oprF-RV	Expression analysis on <i>Pcal</i>	GCGCTGTCGTAATACTCTTTCT	Sakata et al. (2021) [36]
<i>Pcal recA</i>	recA-FW	Expression analysis on <i>Pcal</i>	TCTCTACGGCAAGGGTATCT	Sakata et al. (2021) [36]
<i>Pcal recA</i>	recA-RV	Expression analysis on <i>Pcal</i>	GCTTTACCCTGACCGATCTT	Sakata et al. (2021) [36]

BamHI digestion sites are underlined.