

Exploiting the potential of bioreactors for creating spatial organisation in the soil microbiome: A strategy for increasing sustainable agricultural practices

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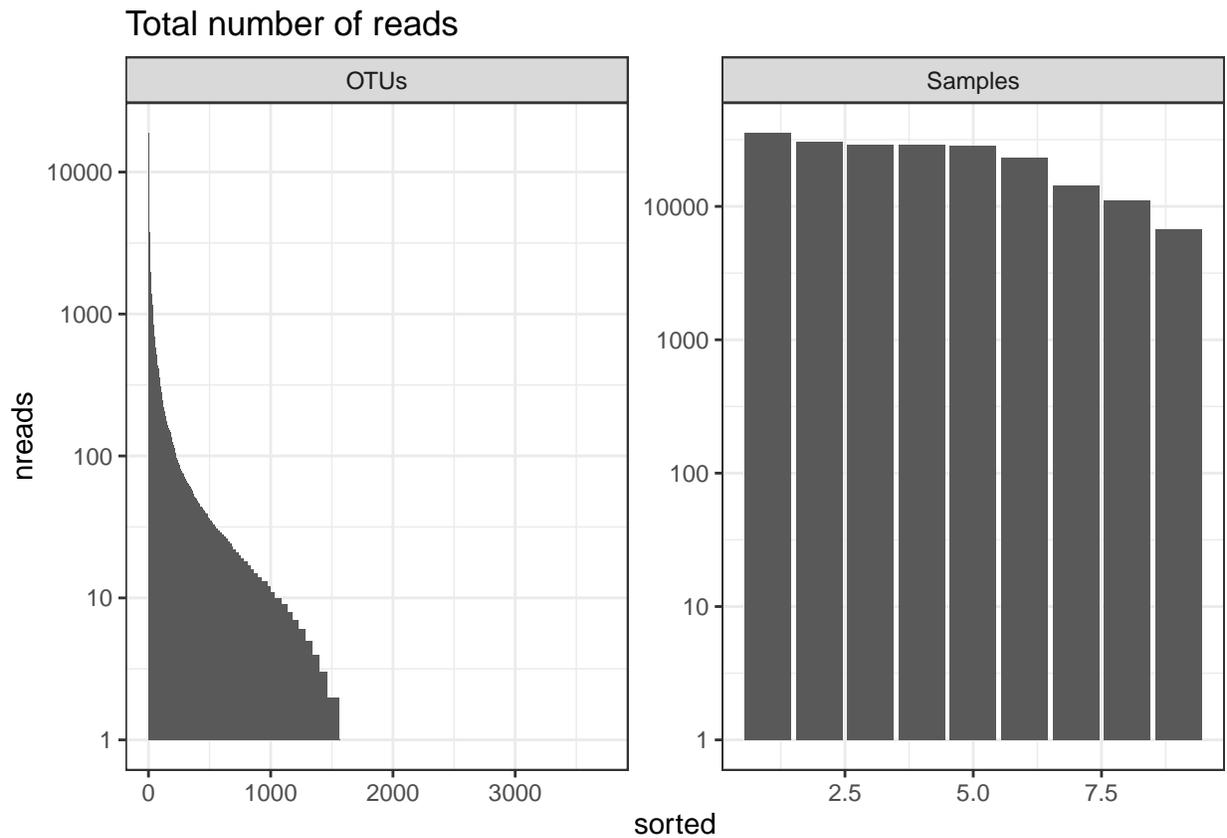


Figure S1: Number of reads obtained after bioinformatic processing shown as (a) number of reads per OTU (or amplicon sequence variants - ASVs) and (b) number of reads per sample.

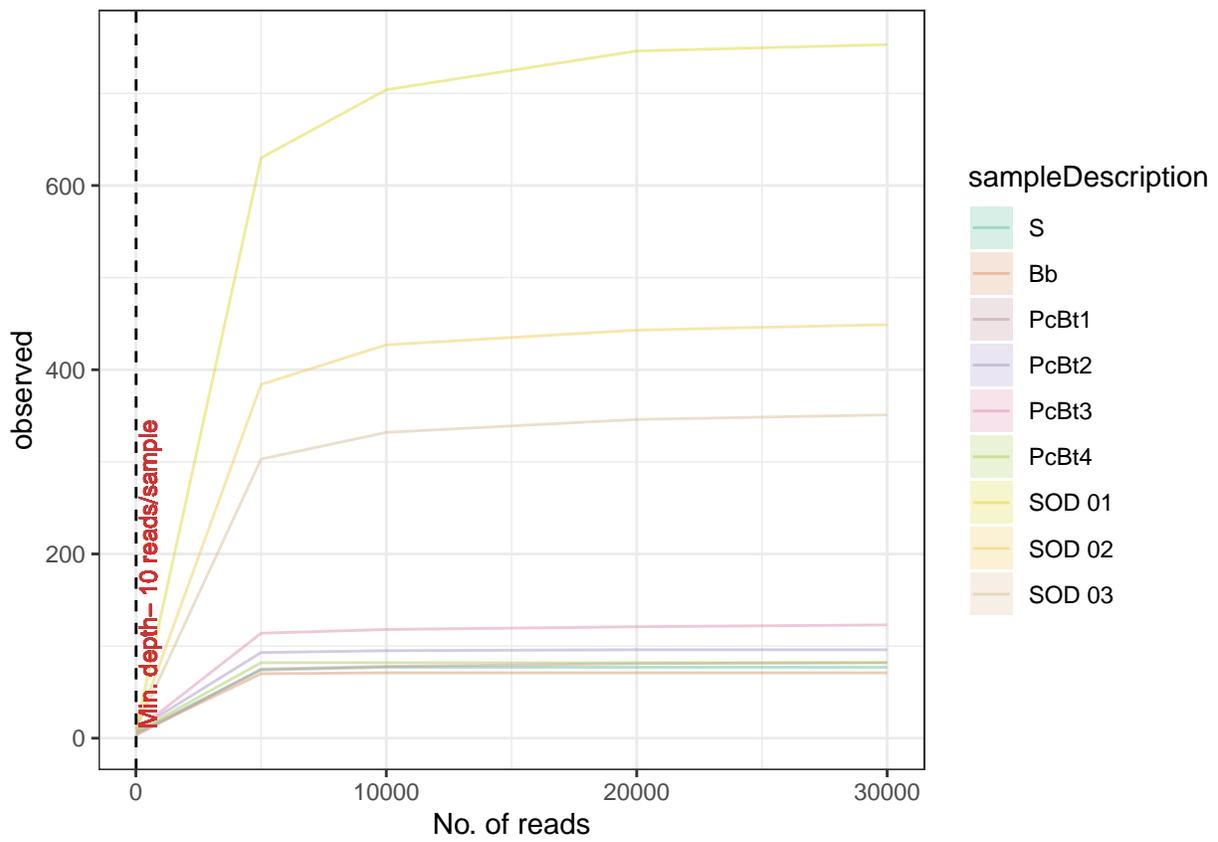


Figure S2: Rarefaction of final taxonomic data from microbiome analysis

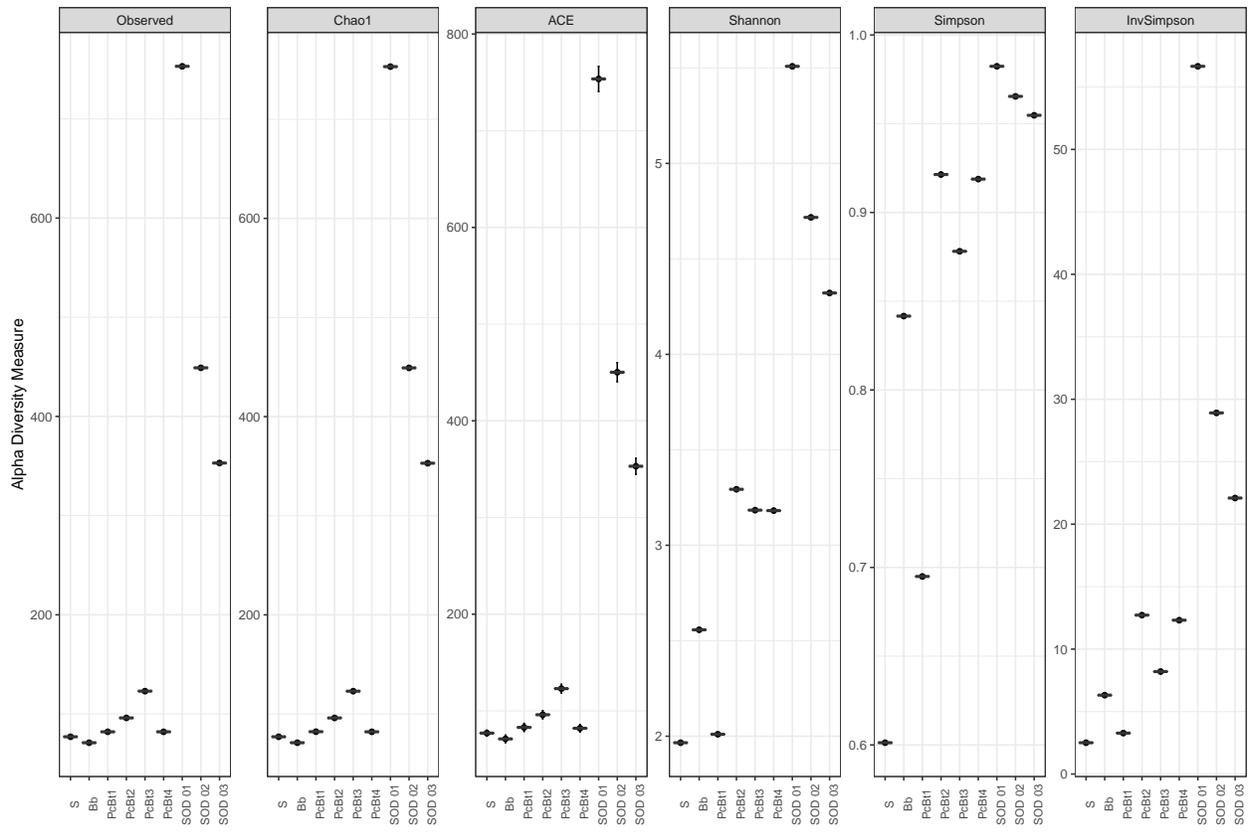


Figure S3: Alpha diversity analysis using Observed, Chao1, ACE, Shannon, Simpson and Inverse Simpson metrics

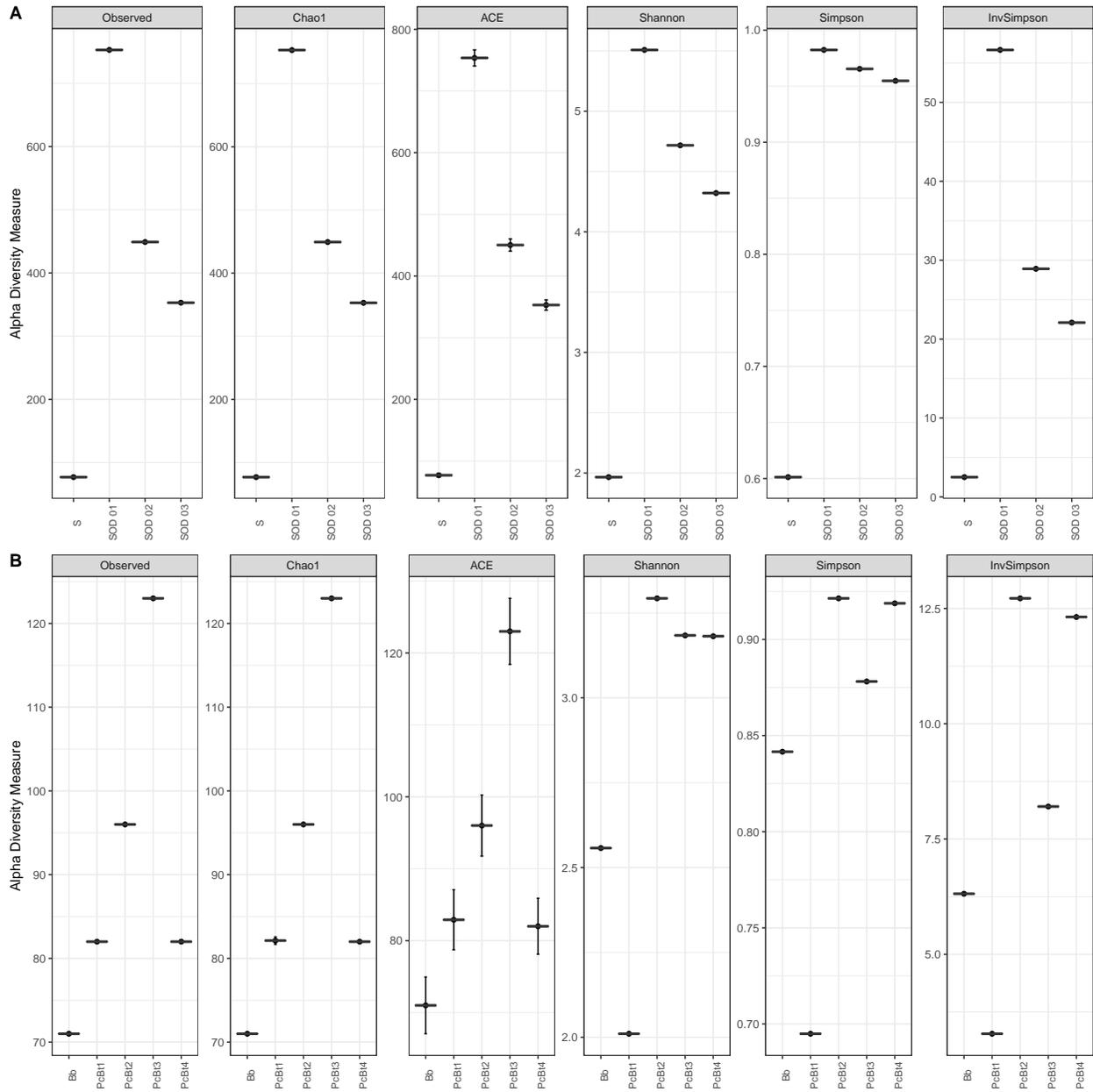


Figure S4: Alpha diversity analysis using Observed, Chao1, ACE, Shannon, Simpson and Inverse Simpson metrics for subset data

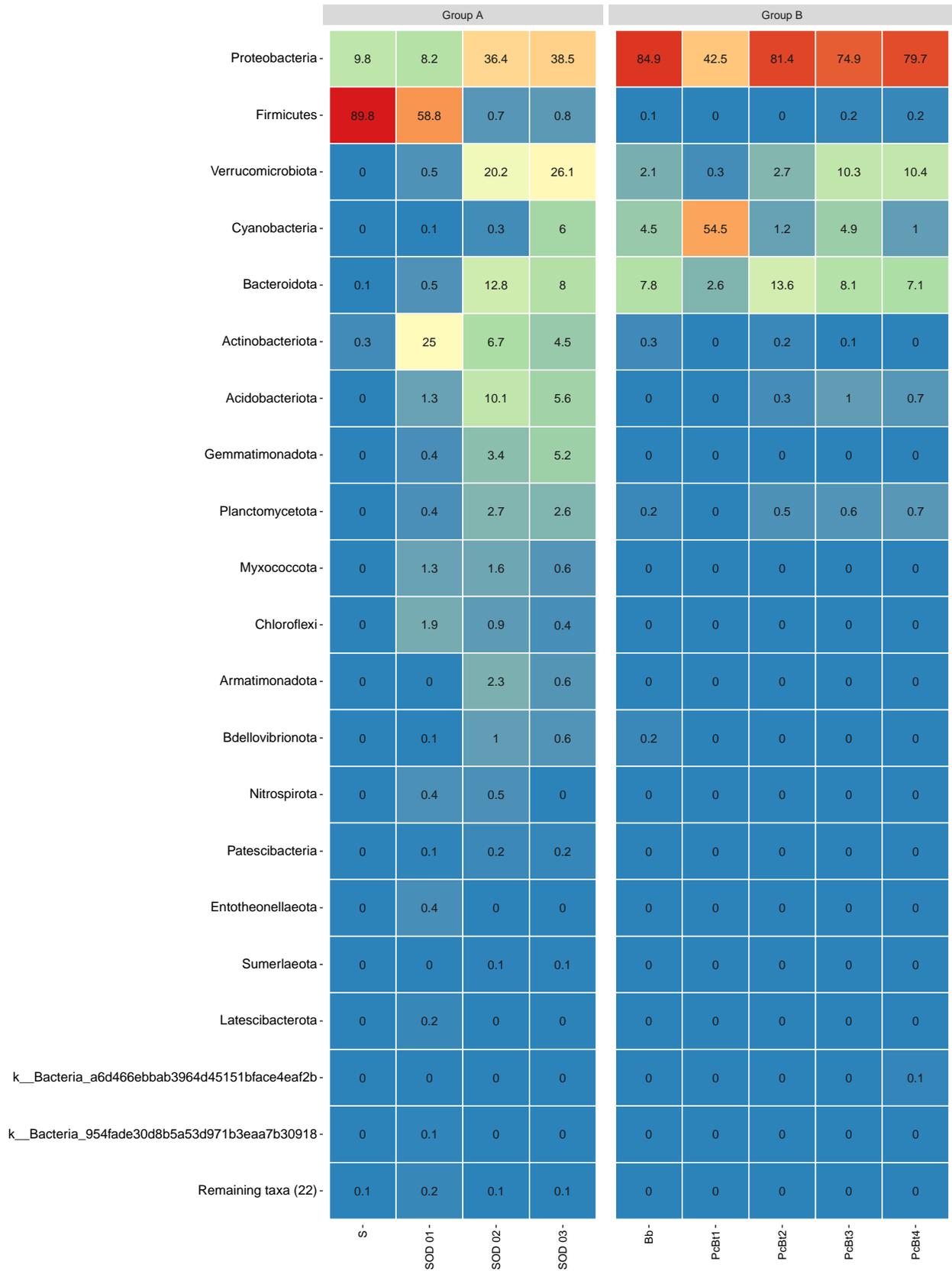


Figure S5: Heatmap showing relative abundance of top 20 phylum taxa identified from samples

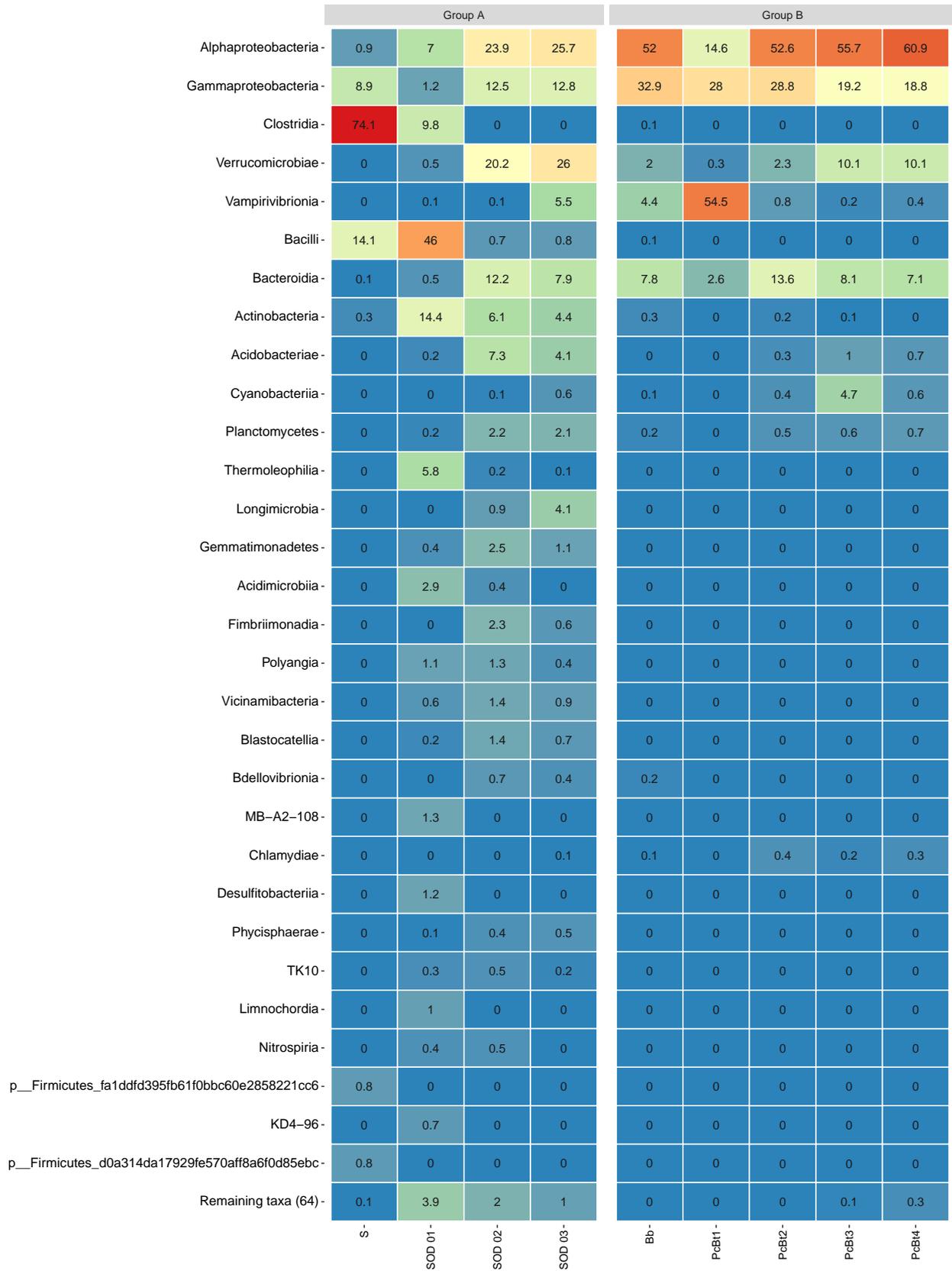


Figure S6: Heatmap showing relative abundance of top 30 class identified from samples



Figure S7: Heatmap showing relative abundance of top 40 order taxa identified from samples

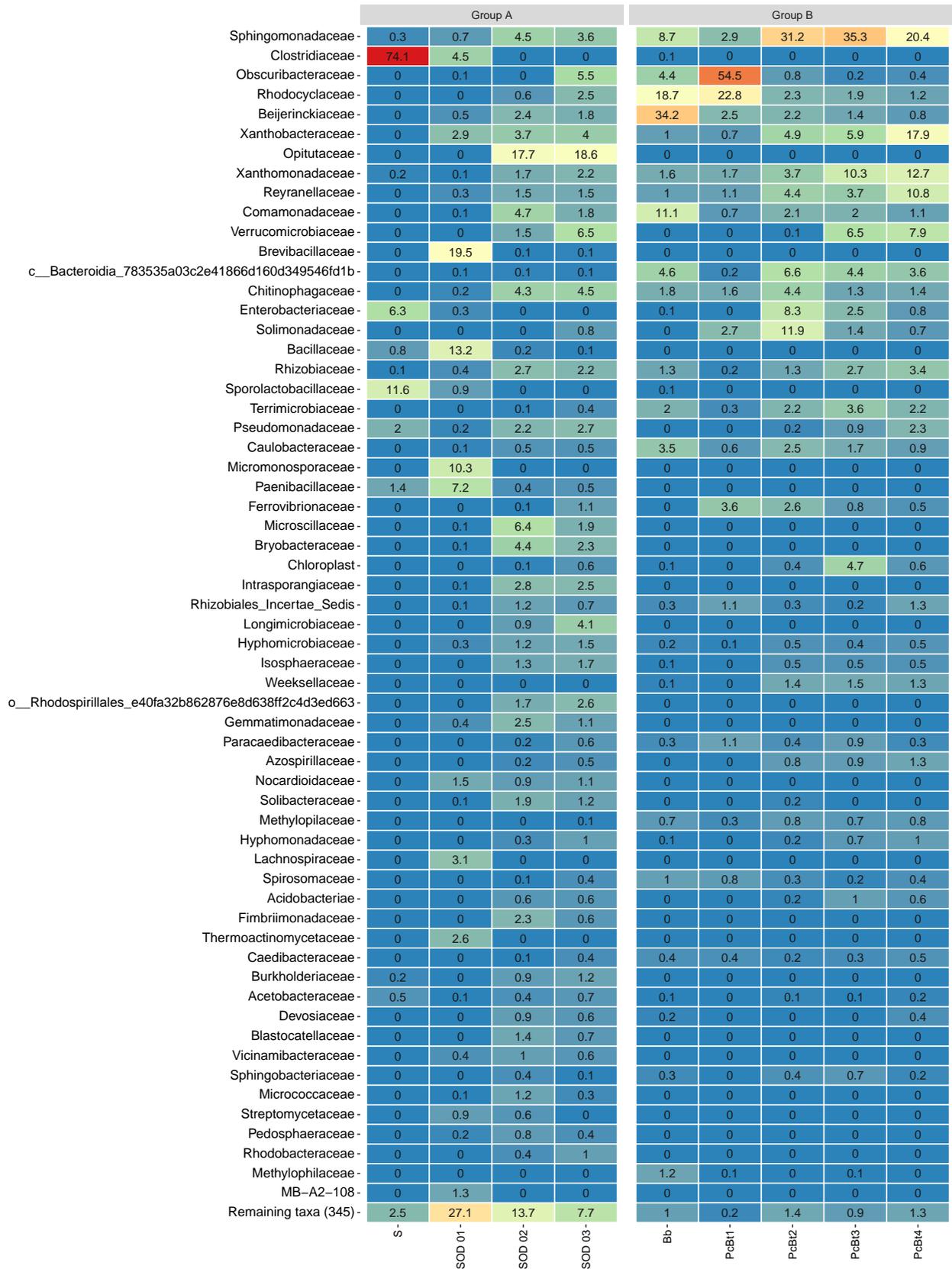


Figure S8: Heatmap showing relative abundance of top 60 family taxa identified from samples

	Group A				Group B				
	S	SOD 01	SOD 02	SOD 03	Bb	PcBt1	PcBt2	PcBt3	PcBt4
Shingobium-	0.1	0.1	0.2	0.2	0.7	0.8	21.7	32.6	17.5
Clostridium_sensu_stricto_12-	68.2	3	0	0	0.1	0	0	0	0
Candidatus_Obscuribacter-	0	0.1	0	5.5	4.4	54.3	0.7	0.1	0.1
Zoogloea-	0	0	0.6	2.5	18.1	22.8	0	0	0
Beijerinckia-	0	0.2	0.1	0.9	33.4	2.5	2	1.3	0.8
Opiritatus-	0	0	17.2	18.3	0	0	0	0	0
Pseudoxanthomonas-	0	0	1.3	2	1.5	1.7	3.7	10.1	12.2
Xanthobacter-	0	0	1.3	2.4	0.5	0.3	4.2	5.2	17.9
Reyranella-	0	0.3	1.4	1.4	1	1.1	4.4	3.7	10.8
Brevibacillus-	0	19.5	0.1	0.1	0	0	0	0	0
c__Bacteroidia_783535a03c2e41866d160d349546fd1b-	0	0.1	0.1	0.1	4.6	0.2	6.6	4.4	3.6
Hydrocarboniphaga-	0	0	0	0.7	0	2.7	11.9	1.4	0.7
Novosphingobium-	0.1	0.1	0	0.3	7.4	1.1	6.7	0.7	0.4
Prostheobacter-	0	0	0.2	0.1	0	0	0.1	6.5	7.9
Bacillus-	0.8	12.3	0.2	0.1	0	0	0	0	0
Sporolactobacillus-	11.5	0.9	0	0	0.1	0	0	0	0
Shingomonas-	0	0.3	3.3	2.1	0	0.5	2.4	1.5	1.5
Terrimicrobium-	0	0	0.1	0.4	2	0.3	2.2	3.6	2.2
Pelomonas-	0	0	0	0	10.1	0.1	0	0	0
Ferrovibrio-	0	0	0.1	1.1	0	3.6	2.6	0.8	0.5
f__Chitinophagaceae_9d9e4ad8e3d7e910f6237bb14bcc4da-	0	0	0.5	1	0	0.3	3.9	1	1.4
Pseudomonas-	0	0	2.2	2.4	0	0	0.2	0.9	2.3
Roseimicrobium-	0	0	1.3	6.2	0	0	0	0	0
Bryobacter-	0	0.1	4.4	2.3	0	0	0	0	0
Clostridium_sensu_stricto_1-	5.5	1.2	0	0	0	0	0	0	0
Chloroplast-	0	0	0.1	0.6	0.1	0	0.4	4.7	0.6
f__Microscillaceae_d4802ec1ee4dc6981e399d101ce8a8d5-	0	0	5.3	1.2	0	0	0	0	0
f__Enterobacteriaceae_527a09644f077fbcef30effba12056ff-	0	0	0	0	0	0	3.4	1.1	0.8
Paenibacillus-	1.4	2.8	0.3	0.5	0	0	0	0	0
Longimicrobium-	0	0	0.9	4.1	0	0	0	0	0
Nordella-	0	0.1	1	0.6	0.3	1.1	0.3	0.2	1.3
Terrimonas-	0	0	0.8	0.9	1.8	1	0	0.1	0
Cloacibacterium-	0	0	0	0	0.1	0	1.4	1.5	1.3
o__Rhodospirillales_e40fa32b862876e8d638ff2c4d3ed663-	0	0	1.7	2.6	0	0	0	0	0
Phenyllobacterium-	0	0.1	0.2	0.3	3	0.1	0.1	0.1	0.2
Azonexus-	0	0	0	0	0.4	0	1.8	1.1	0.6
f__Isosphaeraceae_136d9eb41806e28a7dfc193424250524-	0	0	1.1	1.5	0.1	0	0.5	0.4	0.3
Gemmatimonas-	0	0.2	2.3	1.1	0	0	0	0	0
Brevundimonas-	0	0	0.2	0	0.3	0	1.1	1.1	0.8
f__Intrasporangiaceae_0cf152dc807b570c5f5ab1a7aea69901-	0	0	1.3	2.2	0	0	0	0	0
Candidatus_Finniella-	0	0	0	0.4	0.3	1.1	0.4	0.9	0.3
Shingopyxis-	0	0	0.2	0.3	0.6	0.5	0.4	0.5	1
Candidatus_Solibacter-	0	0.1	1.9	1.2	0	0	0.2	0	0
Methylophila-	0	0	0	0.1	0.7	0.3	0.8	0.7	0.8
Aminobacter-	0	0	0.2	0.1	0.6	0.1	0.4	0.8	1
f__Enterobacteriaceae_e8b77cf2f7dd1f17a7c7626f2904d7b3-	2.9	0.3	0	0	0	0	0	0	0
Hyphomonas-	0	0	0.3	0.9	0	0	0.2	0.7	1
f__Rhizobiaceae_90a634fef584638a5861ac4f26074d88-	0	0	0.2	0.2	0	0	0.6	0.9	1.1
Paludibaculum-	0	0	0.6	0.6	0	0	0.2	1	0.6
Fimbriimonadaceae-	0	0	2.3	0.6	0	0	0	0	0
Micromonospora-	0	2.9	0	0	0	0	0	0	0
Fibrisoma-	0	0	0	0.2	1	0.8	0.3	0.2	0.4
Anaerocolumna-	0	2.7	0	0	0	0	0	0	0
f__Enterobacteriaceae_97822797c63078d0e598100e24f71b7d-	0	0	0	0	0	0	1.7	1.1	0
Caulobacter-	0	0	0.1	0.2	0.2	0.5	1.2	0.4	0
Pedomicrobium-	0	0.3	1.1	1.2	0	0	0	0	0
Klebsiella-	2.4	0	0	0	0	0	0	0	0
Pseudacidovorax-	0	0	0	0	0	0	1	0.9	0.6
Hyphomicrobium-	0	0	0.2	0.3	0.2	0.1	0.5	0.4	0.5
f__Comamonadaceae_61f7050e1641b84a11835f862bb20726-	0	0	0	0.3	0.9	0.5	0.3	0.1	0.2
Candidatus_Nucleicultrix-	0	0	0.1	0.4	0.4	0.4	0.2	0.3	0.5
Azospirillum-	0	0	0.2	0.4	0	0	0	0.5	1
Flavitalea-	0	0	1.3	0.9	0	0	0	0	0
f__Enterobacteriaceae_15318ef23ab68840e3368e4d318f95ad-	0	0	0	0	0.1	0	1.7	0.3	0
f__Xanthobacteraceae_919ef4fa379e506a07d1d74ac3b7faba-	0	0	0	0.2	0.3	0.3	0.6	0.6	0
Azospira-	0	0	0	0	0.1	0	0.4	0.8	0.6
Cupriavidus-	0	0	0.9	1.1	0	0	0	0	0
Comamonas-	0	0	0	0	0	0	0.8	0.9	0.2
Flavhumibacter-	0	0	0.6	1.3	0	0	0	0	0
f__Comamonadaceae_15157002701d572334bc12450e742d75-	0	0	1.8	0	0	0	0	0	0
Shinella-	0	0.1	0.6	0.2	0	0	0	0.4	0.5
Devosia-	0	0	0.6	0.6	0.2	0	0	0	0.4
Vicinamibacteraceae-	0	0.4	0.8	0.5	0	0	0	0	0
Hydrogenophaga-	0	0	0.6	1.1	0	0	0	0	0
f__Pseudomonadaceae_5953c5b70f425e1d94a03e92aebcb6c7-	1.5	0.2	0	0	0	0	0	0	0
Nocardioides-	0	0.5	0.4	0.8	0	0	0	0	0
Microvirga-	0	0.2	1.2	0.2	0	0	0	0	0
Nubsella-	0	0	0.1	0	0.3	0	0.4	0.6	0.2
Roseomonas-	0	0	0.4	0.7	0.1	0	0.1	0.1	0.2
f__Rhizobiaceae_83e49f4390f396afb34415853f5470db-	0	0	0.5	0.5	0	0	0.1	0.2	0.2
Remaining taxa (743)-	5.3	50.7	33	18.6	4.2	0.9	4.8	2.6	3.3

Figure S9: Heatmap showing relative abundance of top 80 genera identified from samples

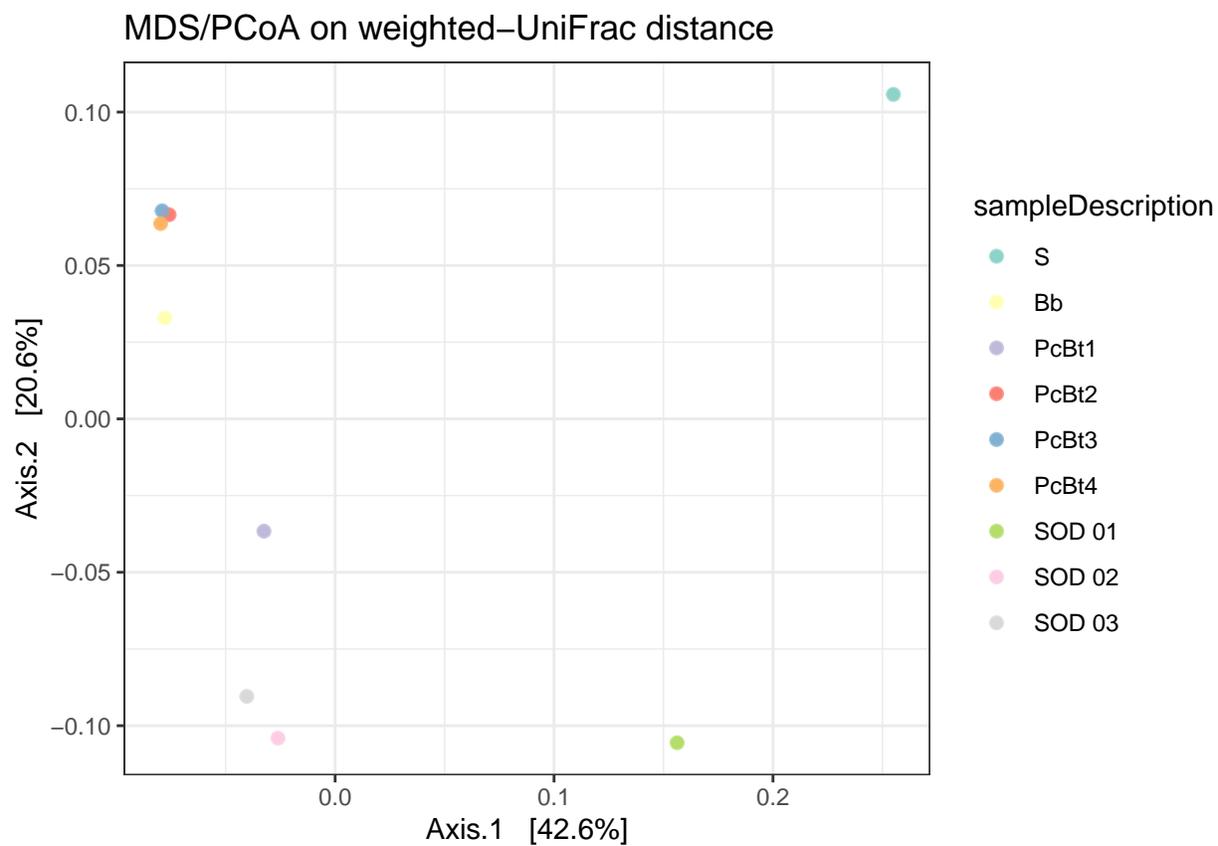


Figure S10: Principal coordinate analysis (multidimensional scaling) of distance matrix using weighted UniFrac metrics. Coordinates for axis 1 and 2 are shown in plot.

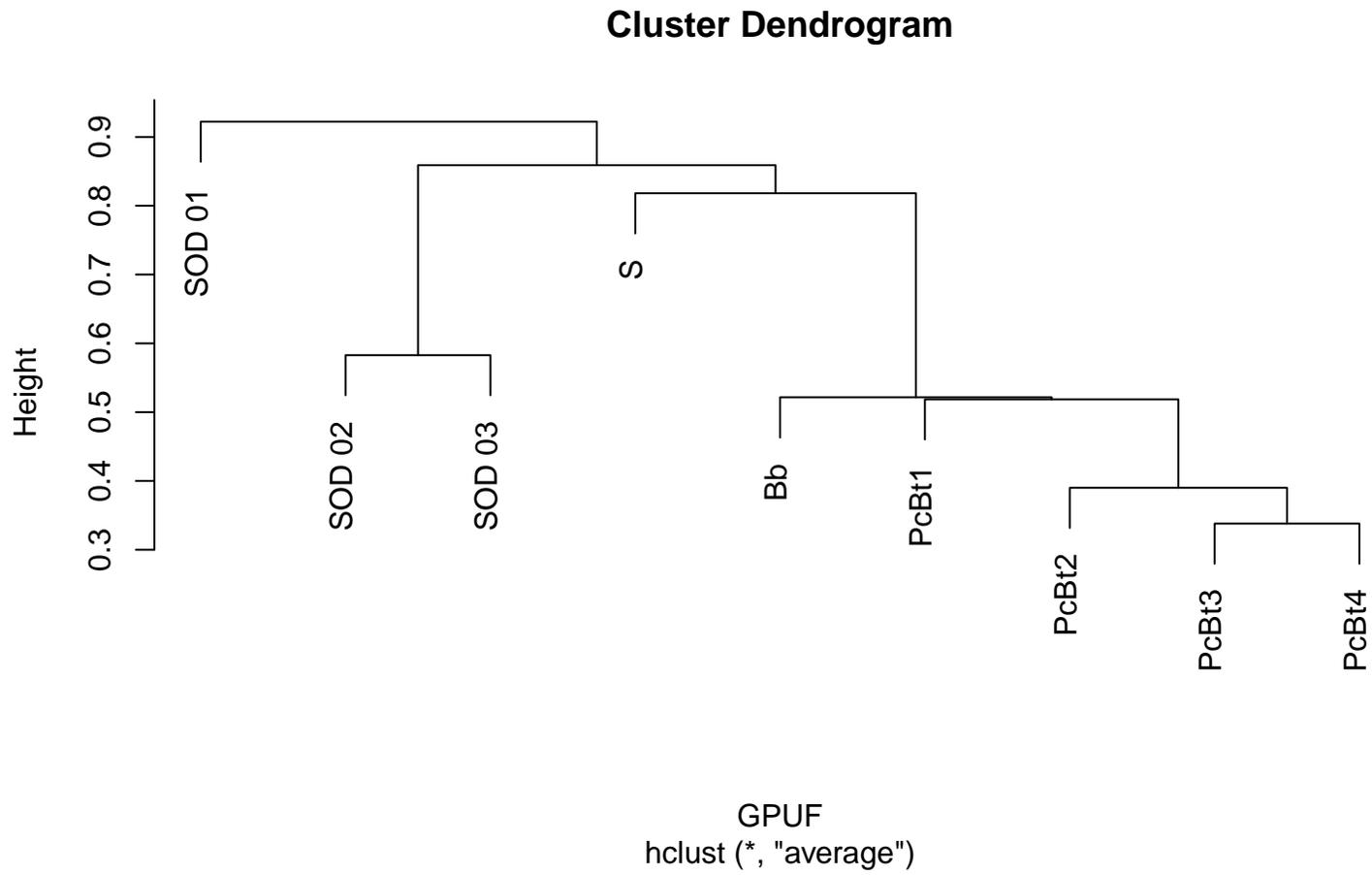


Figure S11: Hierarchical Clustering using UniFrac dissimilarity and UPGMA method.

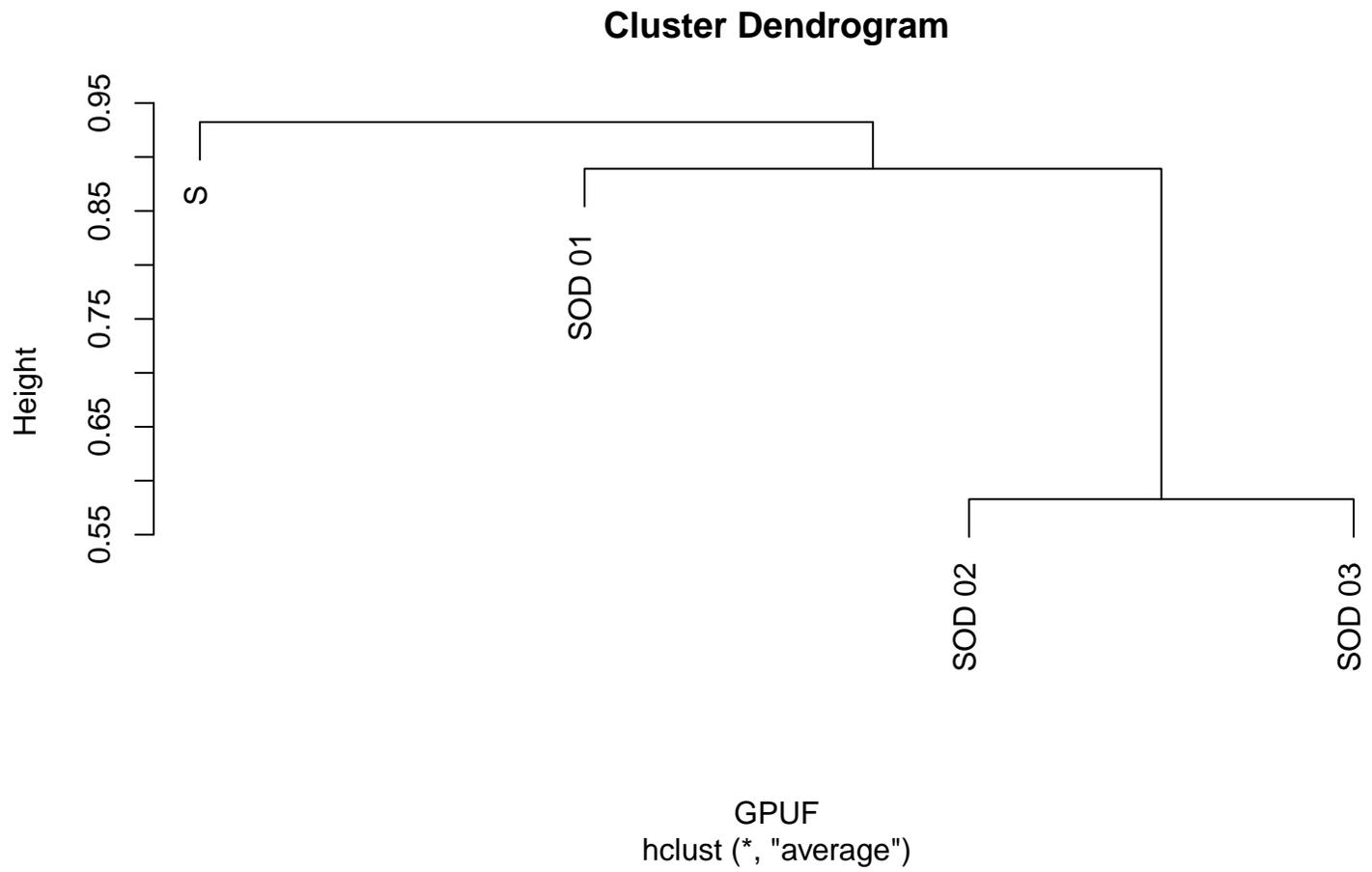


Figure S12: Hierarchical Clustering using UniFrac dissimilarity and UPGMA method of Group A sample group in the present study

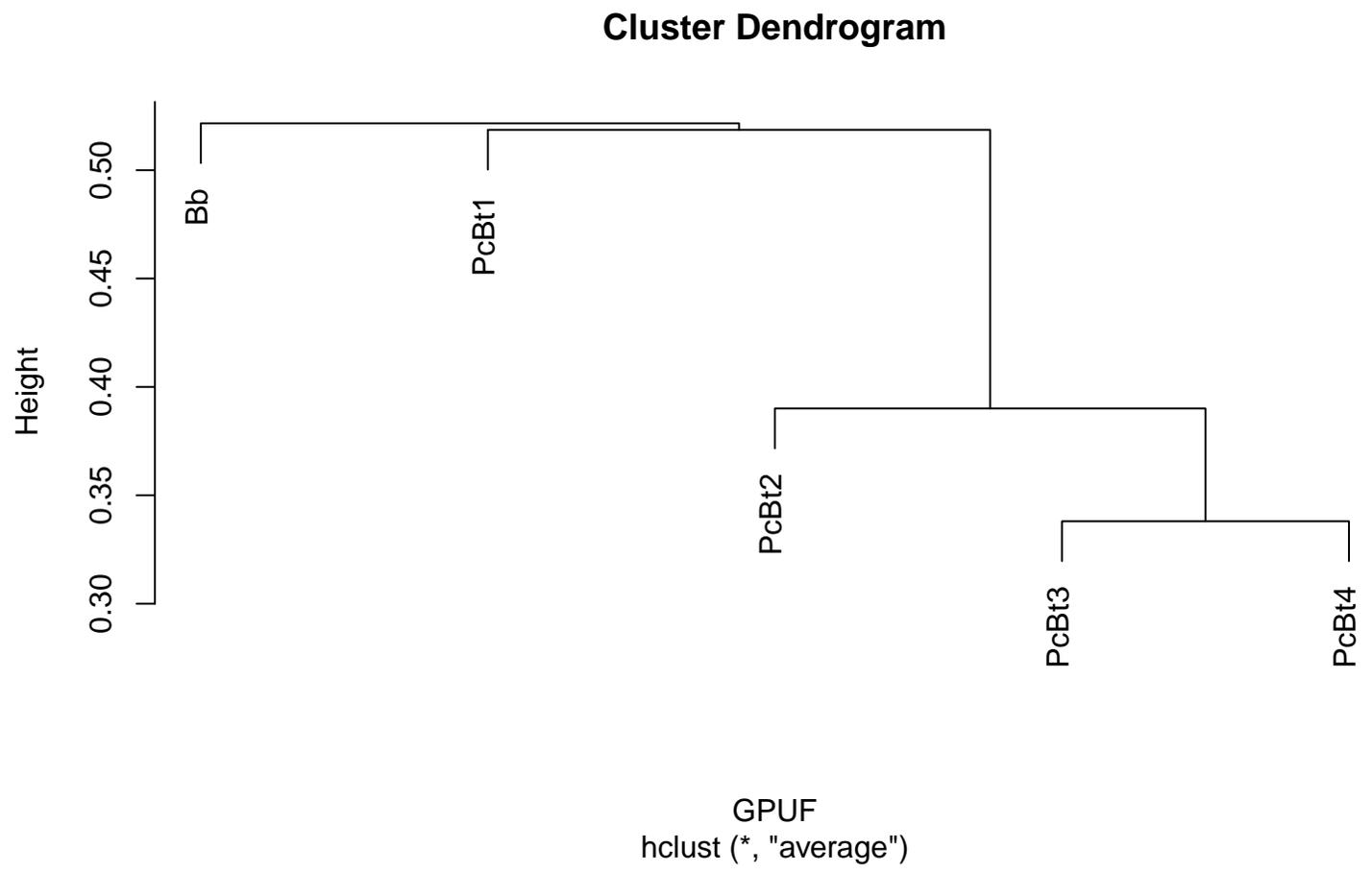


Figure S13: Hierarchical Clustering using UniFrac dissimilarity and UPGMA method of Group B sample group in the present study

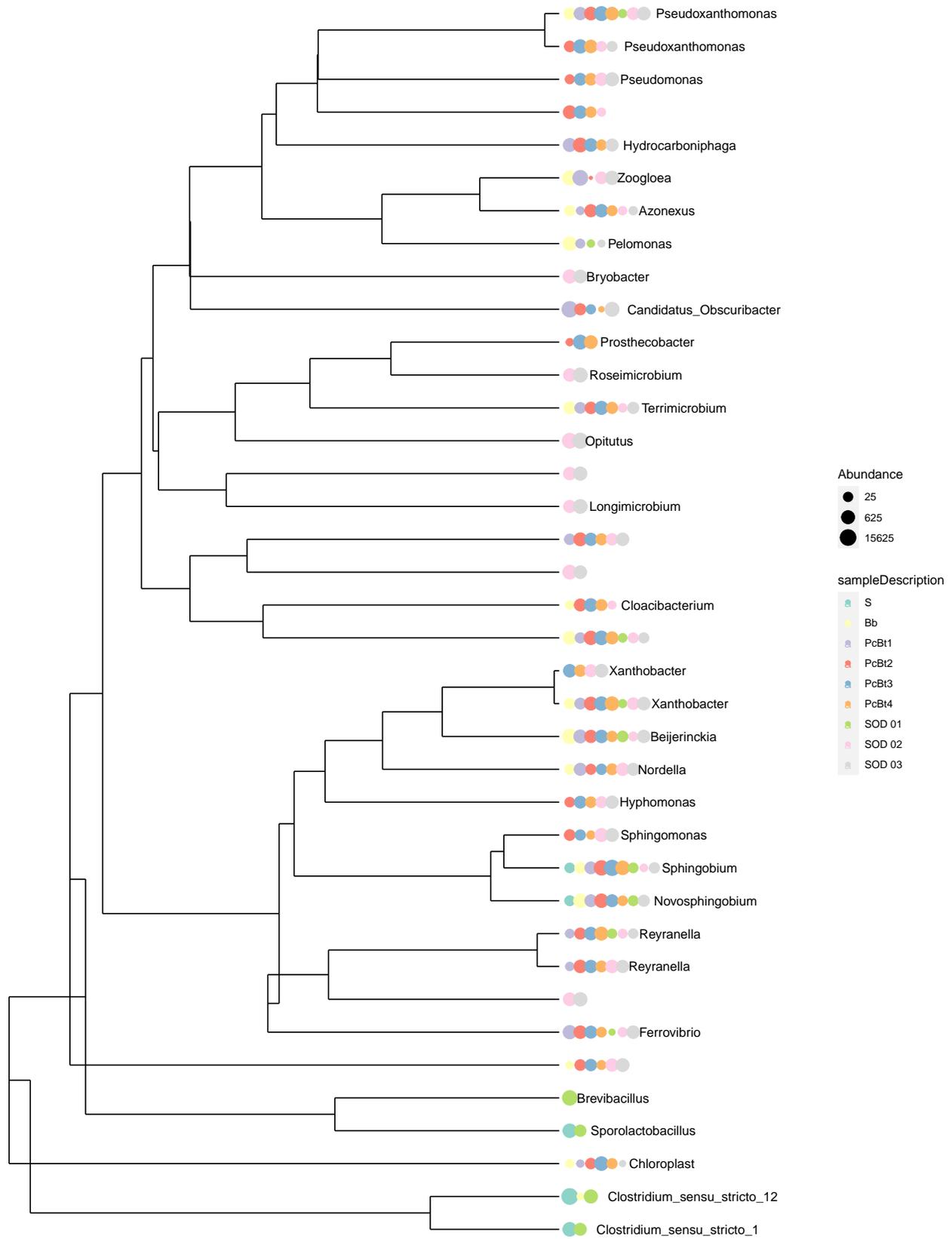
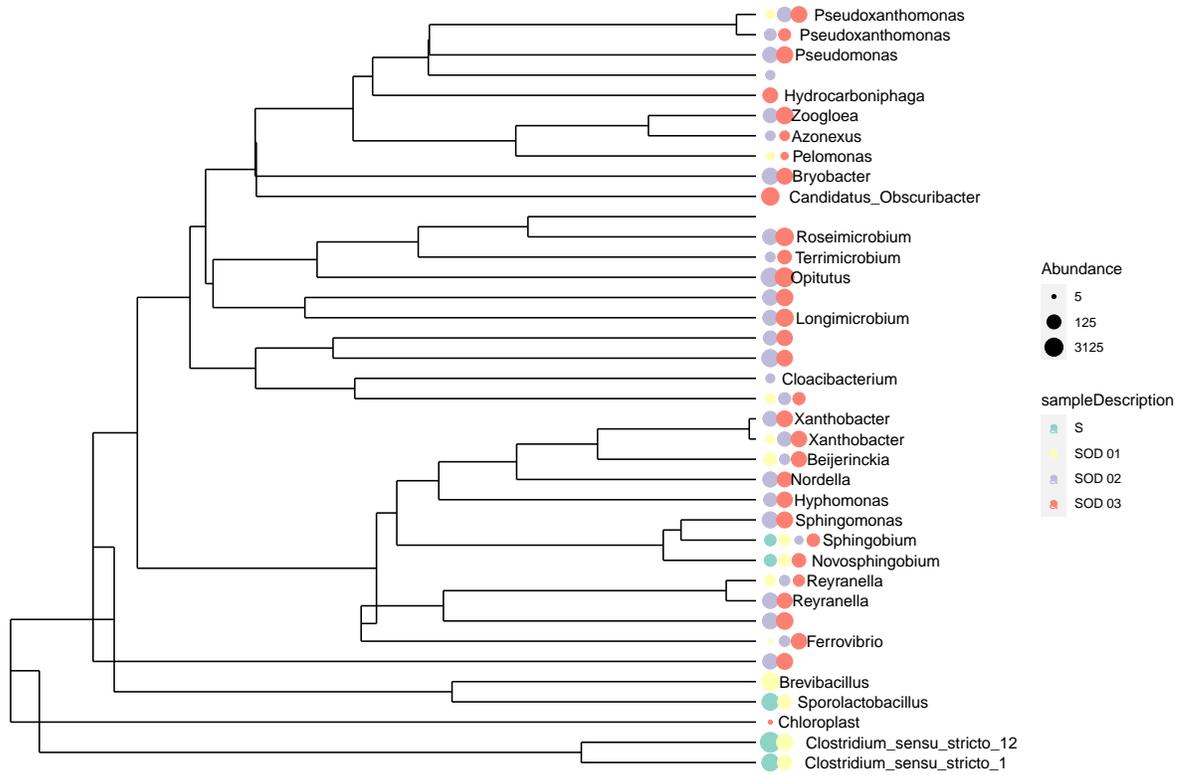


Figure S14: Phylogenetic tree using alignment and UPGMA method done in muscle64.

A



B

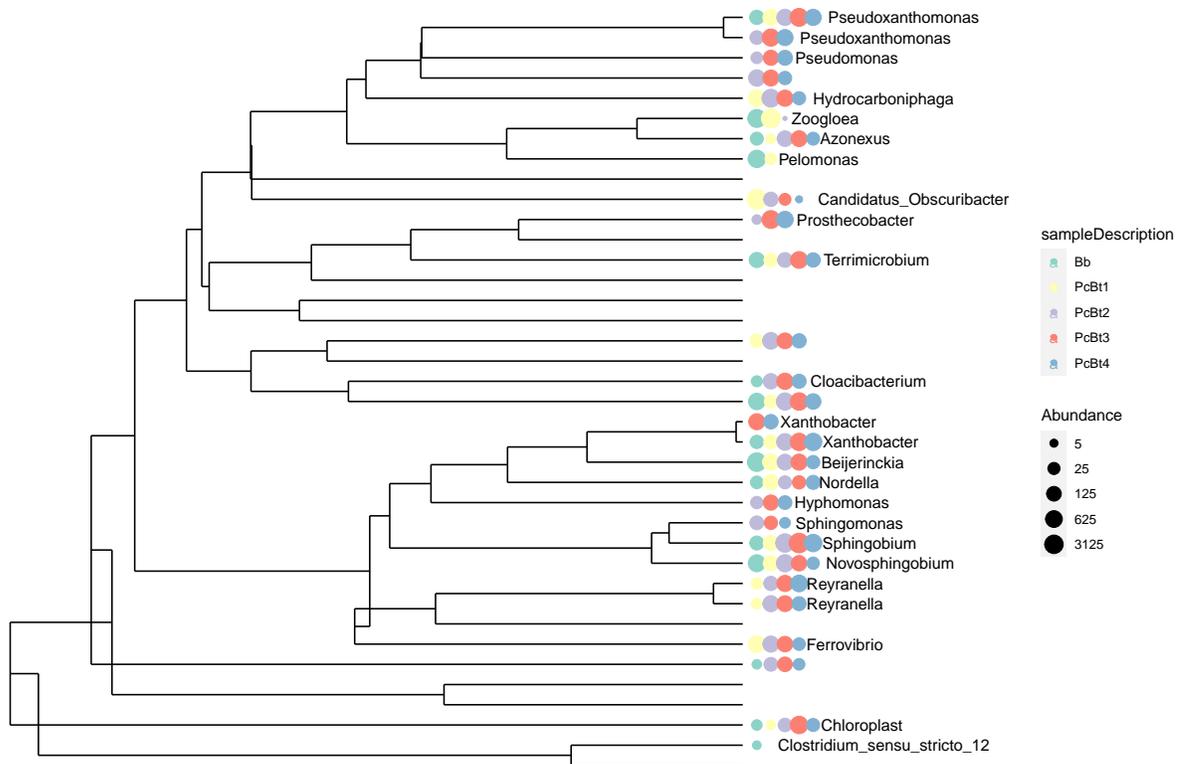


Figure S15: Phylogenetic tree using alignment and UPGMA method done in muscle64 of two sample groups in the present study

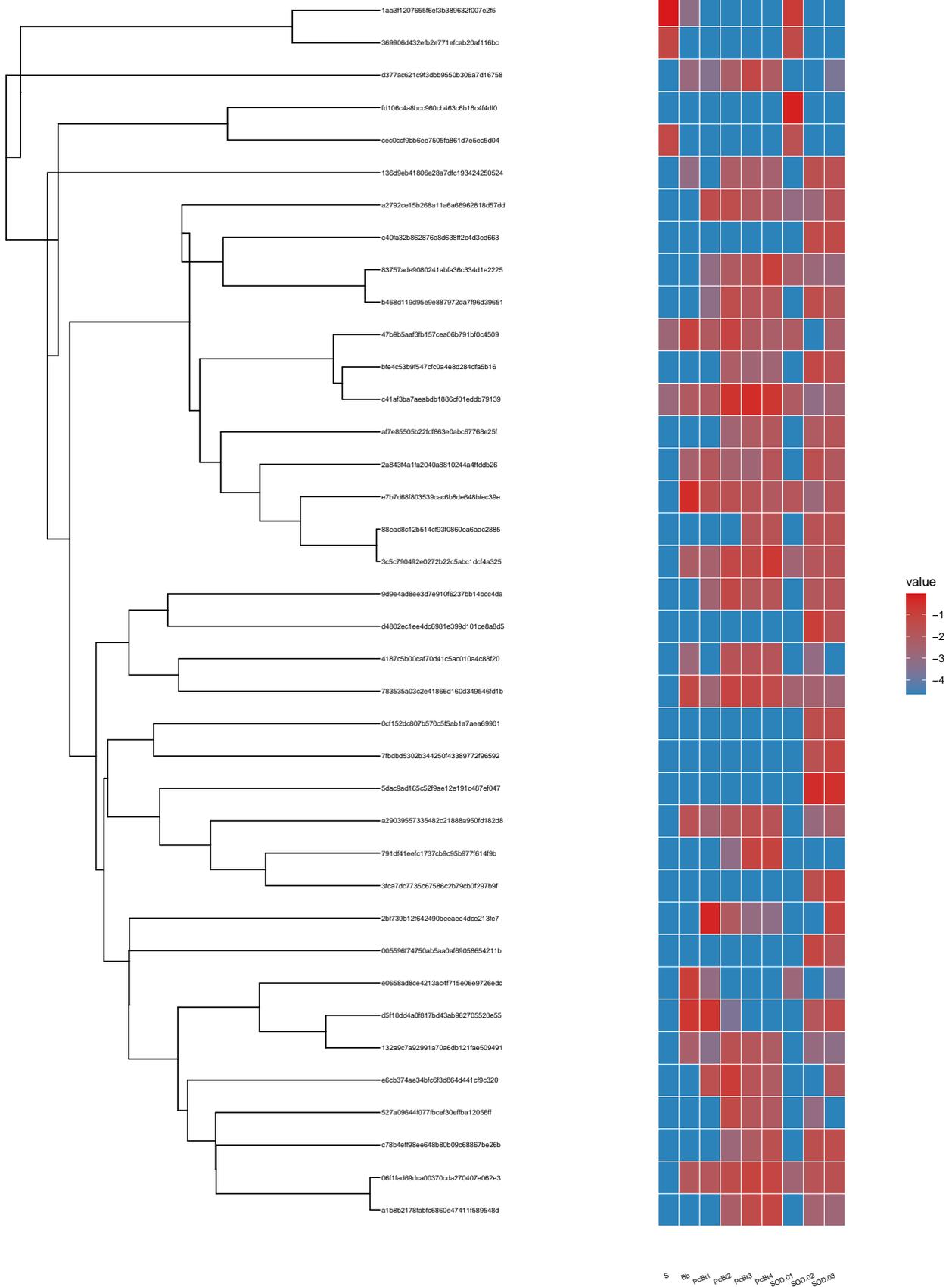


Figure S16: Combined heat map and phylogenetic tree of ASVs with ≥ 1000 total reads ($n = 38$) from samples.

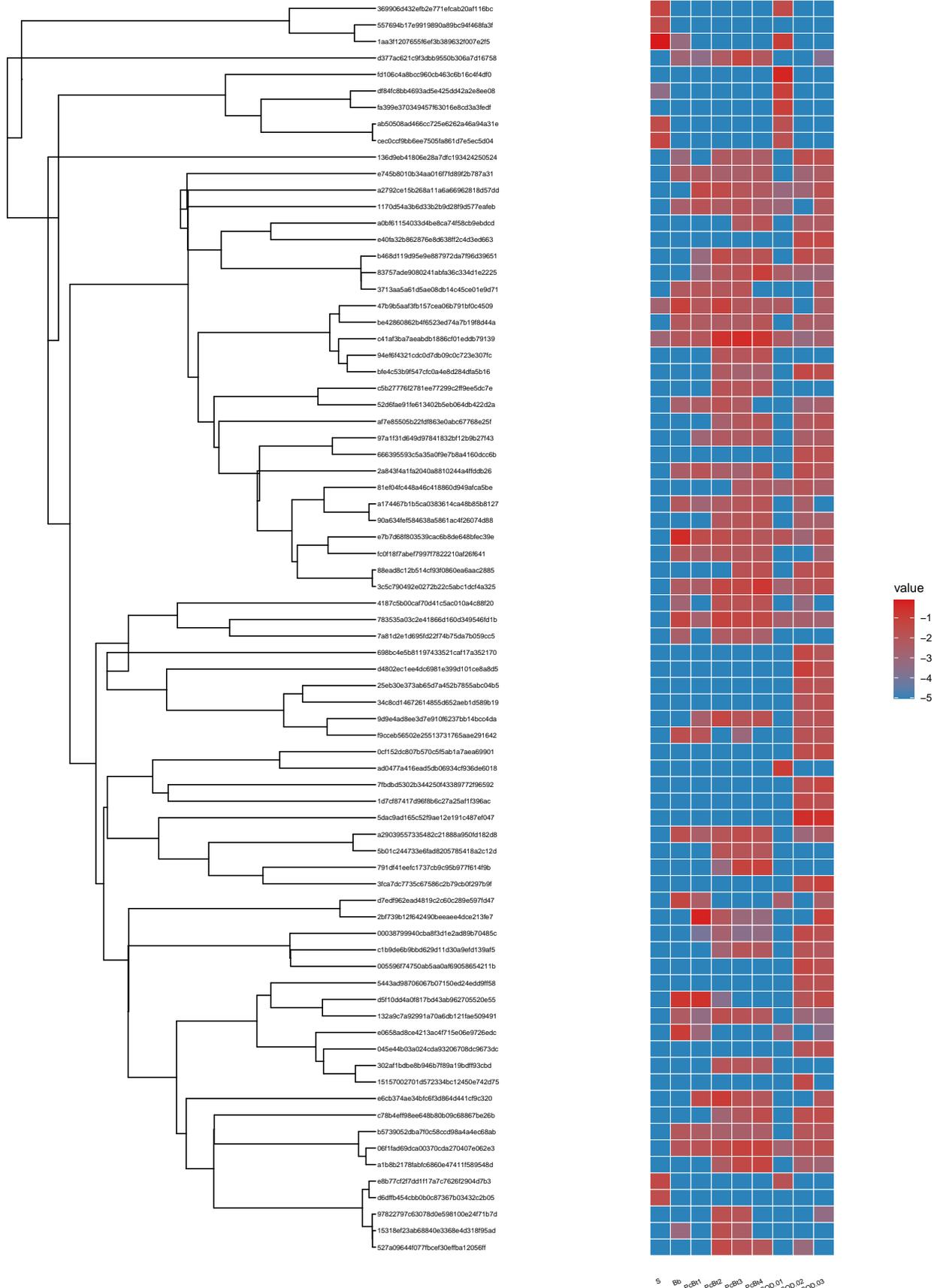


Figure S17: Combined heat map and phylogenetic tree of ASVs with ≥ 500 total reads ($n = 76$) from samples.