



Opinion Subspecific Nomenclature of *Giardia duodenalis* in the Light of a Compared Population Genomics of Pathogens

Michel Tibayrenc 问

Maladies Infectieuses et Vecteurs Écologie, Génétique, Évolution et Contrôle, MIVEGEC (IRD 224-CNRS 5290-UM1-UM2), Institut de Recherche pour le Développement, CEDEX 5, 34394 Montpellier, France; michel.tibayrenc@ird.fr

Abstract: Genetic and genomic data have long recognized that the species *Giardia duodenalis* is subdivided into at least eight genetic clusters that have been named "assemblages" by specialists in the field. Some of these assemblages have been given the status of species, with Linnean binames. In the framework of the predominant clonal evolution model (PCE), we have shown that, from an evolutionary point of view, *G. duodenalis* assemblages are equatable to "near-clades", that is to say: clades whose discreteness is somewhat clouded by occasional genetic exchange, but remain discrete and stable in space and time. The implications of this evolutionary status for the species described within *G. duodenalis* are discussed in light of the most recent genetic and genomic studies. The pattern of this species' subspecific genetic variability and genetic clustering appears to be very similar to the ones of various parasitic, fungal and bacteria species. This underlines the relevance of a compared population genomics of pathogenic species allowed by the broad framework of the PCE model.

Keywords: clonality; genetic recombination; parasite; speciation; taxonomy

1. Introduction

Giardia duodenalis is a cosmopolitan parasite that causes waterborne infections of the digestive tract called Giardiasis. It infects humans and many other mammalian species, including dogs, cats, rats, beavers and cattle [1]. The medical and veterinary relevance of this parasite is considerable since it can be the cause of serious health problems. Moreover, it is highly transmissible, which explains why molecular epidemiology* (* = terms and abbreviations defined in the Appendix A) approaches have been developed for it for a very long time.

2. A Brief Reminder on G. duodenalis Genetic Variability

Genetic analysis of *G. duodenalis* is complicated by the fact that this parasite features two nuclei. Their genomes are similar but not identical. This leads to a phenomenon termed "allelic sequence heterozygosity" (ASH).

Since the 80s, *G. duodenalis* has been the target of isoenzyme*/Multilocus Enzyme Electrophoresis* (MLEE*) pioneering studies [2]. These studies have led to suspect that *G. duodenalis* is a complex of species [2]. Further studies were based on Multilocus Sequence Typing* (MLST*) routinely involving three housekeeping genes, namely β -Giardin (*Bg*), Glutamate Dehydrogenase (*Gdh*) and Triose Phosphate Isomerase (*Tpi*). They all have shown that this species is subdivided into at least eight genetic clusters, labeled A to H, that have been named "assemblages". These assemblages feature some host specificity [1], although it is not absolute. A and B are zoonoses* and are found in humans and many mammal species. C and D are found in canids; E is isolated from hoofed animals; F is specific to cats; G is to rodents; H is to sea mammals [3,4]. More sophisticated MLST approaches and genomic studies based on whole genome sequencing* (WGS*) and single nucleotide polymorphism* (SNP*) [5] have confirmed the validity of *G. duodenalis* assemblages and have revealed additional genetic clusters within them, termed subassemblages [4].



Citation: Tibayrenc, M. Subspecific Nomenclature of *Giardia duodenalis* in the Light of a Compared Population Genomics of Pathogens. *Pathogens* **2023**, *12*, 249. https://doi.org/ 10.3390/pathogens12020249

Academic Editor: Sébastien Besteiro

Received: 30 December 2022 Revised: 30 January 2023 Accepted: 30 January 2023 Published: 3 February 2023



Copyright: © 2023 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

3. G. duodenalis Mode of Reproduction and Population Structure

In the framework of the "clonal theory of parasitic protozoa", we have proposed [6,7] that *G. duodenalis* has a clonal population structure, like many other parasitic species. The evidence was based on the observation of repeated ubiquitous Multilocus Genotypes* (MLGs*) at frequencies that do not fit panmictic* expectations, the presence of statistically significant linkage disequilibrium* and the lack of recombinant genotypes. This clonal pattern has been corroborated by other authors [8], with comparable arguments.

The definition of clonality proposed by us [6,7] limits itself to strongly restrained genetic recombination*. It does not deal with any precise cytological or mating processes and only concerns population structure. It, therefore, includes not only mitotic propagation ("classic" clonality) but also self-fertilization*, strong homogamy*/inbreeding* and several cases of parthenogenesis*. This definition of clonality is shared by many, if not most, scientists working on pathogen population structure (see many references and Table 2 in [9]). Conversely, recombination is equated with sexuality by some authors [10]. However, other authors privilege alternative definitions of clonality. Rougeron et al. [11] restrain clonality to the sole mitotic propagation ("classic" clonality with genetic monomorphism.

4. G. duodenalis and the Predominant Clonal Evolution* (PCE*) Model

The advent of high-resolution genetic and genomic surveys (WGS, SNP) has allowed us to refine the clonal evolution model [13]. We have proposed that predominant clonal evolution concerns all the cases where the impact of genetic exchange is unable to erase the predominant clonality pattern, the most typical manifestation of it being the presence of a persistent phylogenetic* signal at all evolutionary scales. We have forged the term "near-clade*" (NC*; [13]) to designate the phylogenetic clusters observed in many pathogen species. The term "clade*" is not appropriate, since it concerns evolutionary lineages that are strictly separated from each other, whereas it is probable that occasional genetic exchange occurs within many pathogen species, and even, among some species. Now the NCs feature some remarkable properties: despite the possible occurrence of occasional mating, they are discrete and highly stable in space and time, within the whole ecogeographical range of the species considered. We have proposed that the *G. duodenalis* assemblages perfectly fit the definition of NC [14]. One should emphasize that the concept of NC is compatible with some limited genetic exchange, provided that it does not disrupt the prevalent pattern of detectable phylogenetic signal and of Multigene Bifurcating Trees (MGBTs) from macro- to micro-evolutionary levels. Such MGBTs are indeed considered by many authors as the most reliable manifestation of restrained recombination and predominant clonality [15–19]. Species phylogenies should be based on multigene surveys. The analysis of isolated genes may not reflect the true evolutionary story of the species under survey [20]. When G. duodenalis is concerned, discrepancies between phylogenies inferred from different genes have been considered as evidence for genetic exchange [10,21]. However, (i) If such incongruences are due to genetic exchange, they say nothing about the frequency of these exchanges; (ii) they may have many possible causes but genetic exchange [14], for example, homoplasies, markers having different molecular clocks* [22], or undergoing different selective pressures, or different evolutionary tendencies. Another possible cause of phylogenetic discrepancies is the presence of mixed genotypes in the same isolate [23].

The existence of gene flow among and within the assemblages still is in debate. Ankarklev et al. [4] postulate that genetic recombination is "widespread" within assemblage A, but also, between A and E. Cooper et al. [21] infer that there is "evidence for nonclonal evolution" within the subassemblage A2, a hypothesis that is based on incongruence between trees of individual loci. Thompson and Ash [24] hypothesize that occasional recombination could be observed when the transmission is more frequent. Tsui et al. [5] consider that recombination is observed in *G.duodenalis*, but it does not affect the interpretation of phylogenetic data.

However, if genetic exchange obtains in *G. duodenalis*, it does not affect either the discreteness or stability of the assemblages, in accordance with the statement by Tsui et al. [5]. These properties of the assemblages (NCs) have been corroborated by many independent studies relying on different tools and samplings [3,4,10,14,25–31].

5. Challenging the PCE Model: Pseudo-Speciation vs. Russian Doll Model*

Ramírez and Llewellyn have claimed [32,33] that the clonal model of pathogens is "artefactual" for the following reasons. These authors have postulated that restrained recombination in pathogens is verified only among the main genetic subdivisions (=NCs) of the species under study, whereas genetic exchange is much more frequent within each of these main clusters. This model is very similar to the model of pseudo-speciation proposed by Maynard Smith et al. [34] (see Figure 1).

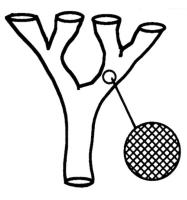


Figure 1. Model of pseudo-speciation: gene flow is inhibited among the main genetic clusters (NCs) of the species under study, whereas it is not within each of these genetic clusters (after Maynard Smith et al. [34]) (Copyright (1993) National Academy of Sciences, U.S.A).

We have designed [35] the Russian doll model, that challenges Ramírez and Lewellyn's proposals [32,33]. It postulates that restrained recombination is not limited to the main genetic subdivisions (NCs) of the species considered. It is also verified within these main NCs. (Figure 2).

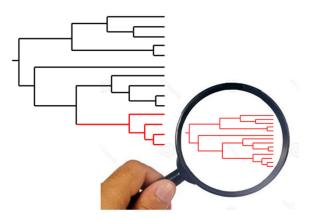


Figure 2. Russian doll model. When population genetic tests are practiced with adequate markers of suitable resolution within each of the NCs that subdivide the species under study (large tree, **left**), they show a miniature picture of the whole species, with the main PCE features, namely, LD and lesser NCs (small tree, **right**). This means that PCE is also verified at these low evolutionary levels.

Such Russian doll patterns (RDPs) have been observed in many species of pathogenic bacteria, yeasts and fungi, and parasitic protozoa (for a recent survey, see [36]). It, therefore, appears that this evolutionary pattern is widespread in pathogenic microorganisms and that the proposal in [32,33] is not corroborated in the species surveyed by us.

When *G. duodenalis is* concerned, subclustering patterns (=lesser NCs), that is to say: RDPs have been repeatedly recorded within the assemblages (=main NCs). Subassemblages or subtypes (=lesser NCs) within assemblage A have been corroborated by several studies and have been termed A I, II and III [4,29,37]. RDPs within assemblage B are less clear [27,29]. However, substructuring within B has been observed too. Lesser NCs within assemblage B have been termed B III and IV [4,37]. Figure 1 in Cacciò et al. [37] clearly shows RDPs within assemblages A and B. Perez Faria et al. [3] have evidenced additional RDPs within the subassemblages AI and II (their Figure 1). Sprong et al. [38] also identified RDPs within AI, II and III, as well as within BIII and IV (Figures 1 and 2). However, bootstrap support within BIII and IV was weak. Tsui et al. [5], with genomic data (WGS and SNPs), fully confirmed the discreteness of assemblages A and B and the subclusteruring (RDPs) within them. However, the intra-assemblage pattern was different between A and B. AI (termed A1 in their article) was considered "highly clonal" (close to monomorphic according to their definition), while the subclustering within B was clearer and strongly linked (although not 100%) to geographical location (Figure 2 and Figure 3). Lastly, Woschke et al. [39], with the MLST protocol proposed in [4], fully confirmed RDPs within both assemblages A and B.

RDPs within *G. duodenalis* assemblages have been therefore fully corroborated by many authors relying on various techniques and different sampling strategies. This ubiquity of MGBTs down to low evolutionary levels does not rule out the possibility of limited genetic exchange within the assemblages. However, it does indicate that PCE is verified within them, which makes it possible to reject for this species the pseudo-speciation model proposed in [32,33].

6. Are G. duodenalis Assemblages Equatable to Species?

This is a long-open debate among the specialists in the field. The need to design a robust terminology that permits scientific exchanges is compelling [24]. However, Latin binames are not a panacea. The description of new species should be based on solid criteria. It should be recalled that the description of new species in pathogenic microorganisms is "largely used as a label of convenience" and that there is no universal bacterial species concept [12].

Several authors have proposed to consider assemblages A and B as different species, respectively termed *G. duodenalis* and *G. enterica* [24,26]. It has been also proposed to term assemblages C/D, *G. canis*, assemblage E, *G. bovis*, assemblage F, *G. cati*, and assemblage G, *G. simondi* [24].

However, the specialists in the field seem to be undecided about the use of the biological species concept* (BSC*), which states that different species are defined by the lack of genetic exchange among them [40]. More generally, the criteria on which assemblages should be described as species are unclear. Andersson [41] states that genomics* and experimental recombination in *Giardia* and other parasites will make it possible to explore possible signatures of sexual recombination and to describe distinct biological species. In other words, if sex occurs among assemblages, they would not deserve the status of distinct species, which deals with the BSC [40]. Other authors [1,21,23,26] have similar views. Birky's opinion [25] is that one lacks clear criteria for describing new species within *G.duodenalis*. Capewell et al. [27] propose that a higher genetic resolution would make it possible to know whether the assemblages are "true species", without making it clear which criteria are needed for describing "true" species.

It is proposed here not to base the description of assemblages as species on the BSC, which is clearly inappropriate for pathogenic microorganisms that undergo only occasional mating. PCE approach does confirm that *G. duodenalis* assemblages are genetically discrete and highly stable in space and time. These properties are more than enough to apply the phylogenetic species concept* (PSC*; [42]) to them. In the case of *G.duodenalis*, as is the case for other pathogens [36], the PSC should be applied in a flexible way, since the genetic separation among assemblages, according to the NC concept, may not be total.

According to the Russian doll model, the number of lesser NCs in *G.duodenalis*, like for other pathogen species, may be virtually unlimited. Available data for *G. duodenalis* is not sufficient to confirm it, but for other species, RDPs and tiny NCs are observed down to extreme microevolutionary* scales (time scales of historical times, and even, recent year: "measurably evolving pathogens" [43]). This is the case for example of the parasite *Leishmania donovani* and the bacteria *Escherichia coli* and *Staphylococcus aureus* [36]. It would therefore be misleading to design new species on the only criterion of individual genetic clustering, which could lead to countless new species. It is recommendable to design pathogen species based on the PSC only in the case where the new taxa have medical or epidemiological relevance. It is up to the specialists in the concerned field to decide whether it is the case. This is what *G. duodenalis* specialists have done with the assemblages. In their case, the BSC, which is poorly handled in micropathogens, should be abandoned in favor of the PSC, since *G. duodenalis* assemblages exhibit both genetic discreteness and epidemiological relevance.

7. PCE, Speciation and Aneuploidy* in G. duodenalis

It has been inferred [44] that an uploidy is widespread in *G.duodenalis*, as is the case for the parasitic species *Leishmania* [45] and *Trypanosoma cruzi* [46]. This fact is highly relevant for population genetics and, therefore, for the problem of speciation in *G. duodenalis*. Indeed, some tests proposed to detect genetic exchange in parasites' natural populations are based on the hypothesis that these parasites are diploid [11]. Aneuploidy makes therefore the interpretation of these tests debatable [13]. Such tests should hence be used quite cautiously for exploring the problem of genetic exchange in *G. duodenalis*. In relation to this, it can be recalled that the BSC [40], based on genetic isolation among putative species, is ill-adapted for exploring speciation in pathogenic microorganisms.

8. Conclusion: Towards a Compared Population Genomics of Pathogenic Microorganisms

In the last 20 years, our knowledge of *G. duodenalis* molecular epidemiology, population genetics and evolution has made significant progress. Extensive comparison with various other pathogen species, including bacteria, parasitic protozoa and fungi [36] shows that *G. duodenalis* population structure is not a special case, and, on the contrary, exhibits striking similarities with that of other pathogens. This underlines the relevance of a compared population genomics of pathogenic species allowed by the broad framework of the PCE model.

The existence of at least eight "assemblages" that can be equated to NCs is firmly established, as well as their stability in space and time. The question of genetic exchange among and within the assemblages needs further exploration, however, it is well ascertained that such possible gene flow does not erase the integrity of the assemblages/NCs in the long run.

It is important to insist on the fact that the new species described within *G. duodenalis*, such as *G. enterica*, *G. bovis*, *G. simondi*, etc., [24] are not equatable to biological species [40] like for example horses and donkeys. From an evolutionary point of view, they are NCs. It is probably the same for other *Giardia* species such as *G. microti* or *G. muris*.

In the near future, genomic studies should be actively developed to better address the problems of this parasite's drug resistance and pathogenicity and to further explore the *G. duodenalis* population structure at a microevolutionary level, for knowing whether *G. duodenalis* pertains to the category of the so-called "measurably evolving pathogens" [43].

Funding: This research received no external funding.

Acknowledgments: We thank Jenny Telleria (IRD, Montpellier, France) for designing Figure 2.

Conflicts of Interest: The author declares no conflict of interest.

Appendix A

| Aneuploidy | When: in a given species, different chromosomes may have different copy numbers. |
|---|---|
| Biological species concept (BSC) | A model in which different species are defined by the fact that they cannot produce fertile offspring, such as horses and donkeys. |
| Clade | Evolutionary lineage identified by phylogenetic analysis. A clade is monophyletic (it has only one ancestor) and is genetically isolated (which means that it evolves independently) from other clades. |
| Genetic recombination | Exchange of genetic material occurring at two or more different genetic loci between different individuals, which leads to the generation of offspring with combinations of genetic characteristics that differ from those observed in either parent. |
| Genomics | Discipline that explores the whole genome, whereas genetics considers individual genes or sets of genes. |
| Homogamy | The tendency of an organism to mate with individuals that are genetically very similar or identical to itself. |
| Inbreeding | See homogamy |
| Isoenzymes | Protein extracts of given biological samples are separated by electrophoresis. The gel is then processed with a histochemical reaction involving the specific substrate of a given enzyme. This enzyme's zone of activity is then specifically stained on the gel. From one sample to another, migration differences can appear for this same enzyme. These different electrophoretic forms of a single enzyme are referred to as isoenzymes or isozymes. These electrophoretic forms are a reflection of sequence differences in the genes that command the synthesis of these proteins. They are therefore a reliable reflection of the genetic polymorphism of the organism under study. |
| Linkage disequilibrium | Nonrandom association of genotypes recorded at different loci: contrary to what happens in sexual organisms, these genotypes are not mixed up by genetic recombination, which makes it possible to predict with a high probability a given genotype at locus B when one knows the genotype at locus A is known, whereas this is not possible in sexual organisms. Linkage disequilibrium is a manifestation of restrained genetic recombination. |
| Microevolution | Classically: evolution at the level of a given population. More specifically in the present article: spans of time that could go down to recent years or at least historical times, a definition that fits better the epidemiology of microbial pathogens and the testing of the Russian doll model (see text). |
| Molecular clock | In its strict, original sense (more correctly called the DNA clock hypothesis), the concept that the rate of nucleotide substitutions in DNA remains constant. In a broader sense, it is simply how fast the genomic part that codes for the variability of a given marker evolves. This speed is driven by the rate of substitution/mutation. It may be regular or irregular. |
| Molecular epidemiology | An approach that relies on various molecular techniques for the epidemiological tracking of pathogen strains. |
| Multilocus enzyme electrophoresis (MLEE) | Isoenzyme analysis based on the analysis of a broad range of enzyme systems. Each enzyme system corresponds to one or several genetic loci. Strains that share the same MLEE profile are referred to as zymodemes (parasitology) or electrophoretic types (bacteriology). |
| Multilocus genotype (MLG) | The combined genotype of a given strain or a given individual is established with several genetic loci. |
| Multilocus Sequence Typing (MLST) | Method of pathogen characterization based on the sequencing of several housekeeping genes. |
| Near-Clade [§] (NC [§]) | Discrete, stable and widespread phylogenetic lineages whose discreteness can be somewhat clouded by an occasional genetic exchange. Different from true clades, an NC can have more than one ancestor (hybrid NC). |
| Panmixia, panmictic | A type of genetic structure in which genetic exchange occurs at random in a given population. Panmictic expectations are the verification of this state by various population genetics tests. |
| Parthenogenesis | Mode of reproduction of some metazoa (insects, amphibians, fishes, reptiles) performed without the genetic contribution of a mating partner. |
| Phylogenetic species concept (PSC) [42] | A taxonomic nomenclature in which species correspond to distinct phylogenetic lineages. |

| Phylogeny, phylogenetic | Evolutionary relationships between organisms or genes or molecules often visualized as dendrograms of trees that are computed through various software. |
|--|---|
| Predominant clonal evolution [§] : (PCE [§]) | A population. structure in which the impact of genetic recombination is not sufficient to disrupt a pattern of predominant clonality, the main manifestation of it being a stable and persistent phylogenetic signal with Multigene Bifurcating Trees (MGBTs) observed at all evolutionary scales. |
| Russian doll pattern [§] (RDP [§]), Russian doll model [§] | When the population structure of the species under study at lower evolutionary scales (within the main NCs) looks like a miniature of the whole species' population structure, with widespread clonal MLGs, LD and lesser NCs. This indicates that PCE is observed at these lower evolutionary scales also. |
| Selfing/self-fertilization | Fertilization of an organism by itself, hence by a genotype that is identical to itself. |
| Single nucleotide polymorphisms (SNP) | Polymorphisms of one-letter variations in the DNA sequence. |
| Whole genome sequencing (WGS) | The sequencing of the complete genome, including noncoding sequences. |
| Zoonosis | A disease that can be transmitted to humans from animals. |
| | § = terms coined by us, specifically linked to the PCE model, used only in this article, not proposed for |

[§] = terms coined by us, specifically linked to the PCE model, used only in this article, not proposed for general use.

References

- Lasek-Nesselquist, E.; Welch, D.M.; Thompston, R.C.A.; Steuart, R.F.; Sogin, M.L. Genetic Exchange Within and Between Assemblages of *Giardia duodenalis*. J. Euk. Microbiol. 2009, 56, 504–518. [CrossRef] [PubMed]
- Andrews, R.H.; Adams, M.; Boreham, P.F.L.; Mayrhofer, G.; Meloni, B.P. *Giardia intestinalis*: Electrophoretic evidence for a species complex. *Int. J. Parasitol.* 1989, 19, 183–190. [CrossRef] [PubMed]
- Perez Faria, C.; Zanini, G.M.; Dias, G.S.; da Silva, S.; do Céu Sousa, M. New multilocus genotypes of *Giardia lamblia* human isolates. *Infect. Genet. Evol.* 2017, 54, 128–137. [CrossRef] [PubMed]
- Ankarklev, J.; Lebbad, M.; Einarsson, E.; Franzén, O.; Ahola, H.; Troell, K.; Svärd, S.G. A novel high-resolution multilocus sequence typing of *Giardia intestinalis* Assemblage A isolates reveals zoonotic transmission, clonal outbreaks and recombination. *Infect. Genet. Evol.* 2018, 60, 7–16. [CrossRef] [PubMed]
- Tsui, C.K.M.; Miller, R.; Uyaguari-Diaz, M.; Tang, P.; Chauve, C.; Hsiao, W.; Isaac-Renton, J.; Prystajeckya, N. Beaver Fever: Whole-Genome Characterization of Waterborne Outbreak and Sporadic Isolates to Study the Zoonotic Transmission of Giardiasis. *mSphere* 2018, 3, e00090-18. [CrossRef]
- 6. Tibayrenc, M.; Kjellberg, F.; Ayala, F.J. A clonal theory of parasitic protozoa: The population structure of *Entamoeba*, *Giardia*, *Leishmania*, *Naegleria*, *Plasmodium*, *Trichomonas* and *Trypanosoma*, and its medical and taxonomical consequences. *Proc. Nat. Acad. Sci. USA* **1990**, *87*, 2414–2418. [CrossRef]
- 7. Tibayrenc, M.; Kjellberg, F.; Arnaud, J.; Oury, B.; Brenière, S.F.; Dardé, M.L.; Ayala, F.J. Are eukaryotic microorganisms clonal or sexual? A population genetics vantage. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 5129–5133. [CrossRef]
- 8. Meloni, B.P.; Lymbery, A.J.; Thompson, R.C.A. Genetic characterization of isolates of *Giardia duodenalis* by enzyme electrophoresis: Implications for reproductive biology, population structure, taxonomy, and epidemiology. *J. Parasitol.* **1995**, *81*, 368–383. [CrossRef]
- 9. Tibayrenc, M.; Ayala, F.J. Is predominant clonal evolution a common evolutionary adaptation to parasitisms in parasitic protozoa, fungi, bacteria and viruses? *Adv. Parasitol.* **2017**, *96*, 243–325.
- 10. Cacciò, S.M.; Ryan, U. Molecular epidemiology of giardiasis. Mol. Biochem. Parasitol. 2008, 160, 75-80. [CrossRef]
- 11. Rougeron, V.; De Meeûs, T.; Hide, M.; Waleckx, E.; Bermudez, H.; Arevalo, J.; Llanos-Cuentas, A.; Dujardin, J.C.; De Doncker, S.; Le Ray, D.; et al. Extreme inbreeding in *Leishmania braziliensis*. *Proc. Nat. Acad. Sci. USA* **2009**, *106*, 10224–10229. [CrossRef]
- Hauck, S.; Maiden, M.C. Clonally evolving pathogenic bacteria. In *Molecular Mechanisms of Microbial Evolution, Grand Challenges in Biology and Biotechnology;* Rampelotto , P.H., Ed.; Springer International Publishing: Berlin/Heidelberg, Germany, 2018; pp. 307–325.
- 13. Tibayrenc, M.; Ayala, F.J. Reproductive clonality of pathogens: A perspective on pathogenic viruses, bacteria, fungi, and parasitic protozoa. *Proc. Nat. Acad. Sci. USA* **2012**, *109*, E3305–E3313. [CrossRef]
- 14. Tibayrenc, M.; Ayala, F.J. Cryptosporidium, Giardia, Cryptococcus, Pneumocystis genetic variability: Cryptic biological species or clonal near-clades? PLoS Pathog. 2014, 10, e1003908. [CrossRef]
- 15. Hanage, W.P. Not So Simple After All: Bacteria, Their Population Genetics, and Recombination. *Cold Spring Harb. Perspect. Biol.* **2016**, *8*, a018069. [CrossRef]
- 16. Maiden, M.C.J. Multilocus Sequence Typing of Bacteria. Ann. Rev. Microbiol. 2006, 60, 561–588. [CrossRef]
- Pearson, T.; Giffard, P.; Beckstrom-Sternberg, S.; Auerbach, R.; Hornstra, H.; Tuanyok, A.; Erin, P.; Price, E.P.; Glass, M.B.; Leadem, B.; et al. Phylogeographic reconstruction of a bacterial species with high levels of lateral gene transfer. *BMC Biol.* 2009, 7, 1–14. [CrossRef]
- 18. Pérez-Losada, M.; Browne, E.B.; Madsen, A.; Wirth, T.; Viscidi, R.P.; Crandall, K.A. Population genetics of microbial pathogens estimated from multilocus sequence typing (MLST) data. *Infect. Genet. Evol.* **2006**, *6*, 97–112. [CrossRef]

- 19. Posada, D.; Crandall, K.A.; Holmes, E.C. Recombination in evolutionary genomics. Ann. Rev. Genet. 2002, 36, 75–97. [CrossRef]
- 20. Nichols, R. Gene trees and species trees are not the same. *Trends Ecol. Evol.* **2001**, *16*, 358–364. [CrossRef]
- Cooper, M.A.; Adam, R.D.; Worobey, M.; Sterling, C.R. Population Genetics Provides Evidence for Recombination in *Giardia*. *Current Biol.* 2007, 17, 1984–1988. [CrossRef]
- 22. Wielinga, C.M.; Thompson, R.C.A. Comparative evaluation of *Giardia duodenalis* sequence data. *Parasitol.* **2007**, *134*, 1795–1821. [CrossRef] [PubMed]
- Feng, Y.; Xiao, L. Zoonotic Potential and Molecular Epidemiology of *Giardia* Species and Giardiasis. *Clin. Microbiol. Rev.* 2011, 24, 110–140. [CrossRef] [PubMed]
- 24. Thompson, R.C.A.; Ash, A. Molecular epidemiology of *Giardia* and *Cryptosporidium* infections. *Infect. Genet. Evol.* **2016**, 40, 315–323. [CrossRef] [PubMed]
- 25. Birky, C.W., Jr. Giardia Sex? Yes, but how and how much? Trends Parasitol. 2009, 263, 70–74. [CrossRef]
- Cacciò, S.M.; Sprong, H. *Giardia duodenalis*: Genetic recombination and its implications for taxonomy and molecular epidemiology. *Exp. Parasitol.* 2010, 124, 107–112. [CrossRef]
- 27. Capewell, P.; Krumrie, S.; Katzer, F.; Alexander, C.L.; Weir, W. Molecular Epidemiology of *Giardia* Infections in the Genomic Era. *Trends Parasitol.* **2021**, *37*, 142–153. [CrossRef]
- Higuera, A.; Muñoz, M.; Consuelo López, M.; Reyes, P.; Urbano, P.; Villalobos, O.; Ramírez, J.D. Development of a Multilocus Sequence Typing Scheme for *Giardia intestinalis*. *Genes* 2020, 11, 764. [CrossRef]
- Karim, R.; Wang, R.; Yu, F.; Li, T.; Dong, H.; Li, D.; Zhang, L.; Li, J.; Jian, F.; Zhang, S.; et al. Multi-locus analysis of *Giardia duodenalis* from nonhuman primates kept in zoos in China: Geographical segregation and host-adaptation of assemblage B isolates. *Infect. Genet. Evol.* 2015, *30*, 82–88. [CrossRef]
- Lebbad, M.; Petersson, I.; Karlsson, L.; Botero-Kleiven, S.; Jan, O.; Andersson, J.O.; Svenungsson, B.; Svärd, S.G. Multilocus Genotyping of Human *Giardia* Isolates Suggests Limited Zoonotic Transmission and Association between Assemblage B and Flatulence in Children. *PLoS Neglect. Trop. Dis.* 2011, 5, e1262. [CrossRef]
- 31. Monis, P.T.; Cacciò, S.M.; Thompson, R.C.A. Variation in *Giardia*: Towards a taxonomic revision of the genus. *Trends Parasitol.* **2009**, *25*, 93–100. [CrossRef]
- Ramírez, J.D.; Llewellyn, M.S. Reproductive clonality in protozoan pathogens—Truth or artefact? *Mol. Ecol.* 2014, 23, 4195–4202. [CrossRef]
- Ramírez, J.D.; Llewellyn, M.S. Response to Tibayrenc and Ayala: Reproductive clonality in protozoan pathogens—Truth or artefact? *Mol. Ecol.* 2015, 24, 5782–5784. [CrossRef]
- 34. Maynard Smith, J.; Smith, N.H.; O'Rourke, M.; Spratt, B.G. How clonal are bacteria? *Proc. Nat. Acad. Sci. USA* **1993**, *90*, 4384–4388. [CrossRef]
- 35. Tibayrenc, M.; Ayala, F.J. How clonal are Trypanosoma and Leishmania? Trends Parasitol. 2013, 29, 264–269. [CrossRef]
- 36. Tibayrenc, M.; Ayala, F.J. Models in parasite and pathogen evolution: Genomic analysis reveals predominant clonality and progressive evolution at all evolutionary scales in parasitic protozoa, yeasts and bacteria. *Adv. Parasitol.* **2021**, *111*, 75–117.
- 37. Cacciò, S.M.; Beck, R.; Lalle, M.; Marinculic, A.; Pozio, E. Multilocus genotyping of *Giardia duodenalis* reveals striking differences between assemblages A and B. *Int. J. Parasitol.* **2008**, *38*, 1523–1531. [CrossRef]
- Sprong, H.; Cacciò, S.M.; van der Giessen, J.W.B.; on behalf of the ZOOPNET network and partners. Identification of Zoonotic Genotypes of Giardia duodenalis. *PLoS Neglect. Trop. Dis.* 2009, 3, e558. [CrossRef]
- Woschke, A.; Faber, M.; Stark, K.; Holtfreter, M.; Mockenhaupt, F.; Richter, J.; Regnath, T.; Sobottka, I.; Reiter-Owona, I.; Diefenbach, A.; et al. Suitability of current typing procedures to identify epidemiologically linked human *Giardia duodenalis* isolates. *PLoS Neglect. Trop. Dis.* 2021, 15, e0009277. [CrossRef]
- 40. Dobzhansky, T. Genetics and the Origin of Species; Columbia University Press: New York, NY, USA, 1937.
- 41. Andersson, J.O. Double peaks reveal rare diplomonad sex. *Trends Parasitol.* **2012**, *28*, 46–52. [CrossRef]
- Cracraft, J. Species concept and speciation analysis. In *Current Ornithology*; Notion d'Espèce, Phylogenetic Species Concept, PSC; Johnson, R.F., Ed.; Plenum Press: New York, NY, USA, 1983; pp. 159–187.
- 43. Biek, R.; Pybus, O.G.; Lloyd-Smith, J.O.; Didelot, X. Measurably evolving pathogens in the genomic era. *Trends Ecol. Evol.* **2015**, 30, 306–313. [CrossRef]
- 44. Tumová, P.; Uzlíková, M.; Jurczyk, T.; Nohýnková, E. Constitutive aneuploidy and genomic instability in the single-celled eukaryote *Giardia intestinalis*. *MicrobiologyOpen* **2016**, *5*, 560–574. [CrossRef] [PubMed]
- 45. Lachaud, L.; Bourgeois, N.; Kuk, N.; Morelle, C.; Crobu, L.; Merlin, G.; Bastien, P.; Pagès, M.; Sterkers, Y. Constitutive mosaic aneuploidy is a unique genetic feature widespread in the *Leishmania* genus. *Microbes Infect.* **2014**, *16*, 61–66. [CrossRef] [PubMed]
- Cruz-Saavedra, L.; Schwabl, P.; Vallejo, G.A.; Carranza, J.C.; Muñoz, M.; Patino, L.H.; Paniz-Mondolfi, A.; Llewellyn, M.S.; Ramírez, J.D. Genome plasticity driven by aneuploidy and loss of heterozygosity in *Trypanosoma cruzi*. *Microb. Genom.* 2022, *8*, 000843. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.