

## **Supplementary material for:**

# **Prevalence of *Borrelia burgdorferi* in Ixodidae tick around Asia: a systematic review and meta-analysis**

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## Supplementary Material 1: Assessment of risk of bias

Items
<p><b>Was the study's target population a close representation of the population in relation to relevant variables?</b></p> <p>0. No or Unclear</p> <p>1. Clear description of infection diagnosis: Polymerase Chain Reaction (PCR), immunofluorescent antibody assay (IFA), Direct Immunofluorescent Assay (DFA), Spirochete Culture, and so on</p>
<p><b>Was the sampling frame a true or close representation of the target population?</b></p> <p>0. No or Unclear</p> <p>1. Yes</p>
<p><b>Was some form of random selection used to select the sample, OR was a census undertaken?</b></p> <p>0. No or Unclear</p> <p>1. Yes</p>
<p><b>Was the likelihood of nonresponse bias minimal?</b></p> <p>0. No or Unclear</p> <p>1. Yes</p>
<p><b>Were data collected directly from the subjects (as opposed to a proxy)?</b></p> <p>0. No: proxy, databases, medical reports only</p> <p>1. Directly from the subjects</p>
<p><b>Was an acceptable case definition of positive infection used in the study?</b></p> <p>0. Unclear</p> <p>1. Laboratory-based</p>
<p><b>Was the study method that diagnosed Positive <i>B.burgdorferi</i> infection shown to have validity and reliability?</b></p> <p>0. No description of the method</p> <p>1. Yes: Polymerase Chain Reaction (PCR), immunofluorescent antibody assay (IFA), Direct Immunofluorescent Assay (DFA), Spirochete Culture, and so on</p>
<p><b>Was the same mode of data collection used for all subjects?</b></p> <p>0. No or Unclear</p> <p>1. Yes</p>
<p><b>Was the length of the shortest prevalence period for the parameter of interest appropriate?</b></p> <p>0. No or Unclear</p> <p>1. Yes</p>
<p><b>Were the numerator(s) and denominator(s) for the parameter of interest appropriate and clearly reported?</b></p> <p>0. No or Unclear</p> <p>1. Yes</p>

Study	Authors	Score	Low risk	Moderate risk	High risk
1	Gao Y (2021)	10	+	-	-
2	Seo HJ (2021)	8	-	+	-
3	Wang Q (2021)	10	+	-	-
4	Seo MG (2020)	8	-	+	-
5	Kim SY (2020)	9	+	-	-
6	Nakayama S (2020)	8	-	+	-
7	Seto J (2020)	10	+	-	-
8	Orkun Ö (2020)	9	+	-	-
9	Kaenkan W (2020)	9	+	-	-
10	Fischer T (2020)	10	+	-	-
11	Ghafar A (2020)	9	+	-	-
12	Rar V (2019)	9	+	-	-
13	Pukhovskaya NM (2019)	8	-	+	-
14	Kim CM (2019)	10	+	-	-
15	Duan CJ (2019)	9	+	-	-
16	Jiang BG (2018)	10	+	-	-
17	Hagen RM (2018)	9	+	-	-
18	Narankhajid M (2018)	8	-	+	-
19	Zheng WQ (2018)	8	-	+	-
20	Khoo JJ (2018)	8	-	+	-
21	Ye NN (2018)	9	+	-	-
22	Qin LX (2018)	7	-	+	-
23	Yang Y (2018)	9	+	-	-
24	Tang F (2018)	9	+	-	-
25	Liu XY (2017)	10	+	-	-
26	Karasartova D (2017)	10	+	-	-
27	Kuo CC (2017)	7	-	+	-
28	Pan YP (2017)	8	-	+	-
29	Li HY (2017)	10	+	-	-
30	Sakakibara K (2016)	9	+	-	-
31	Khasnatinov MA (2016)	9	+	-	-
32	Yu PF (2016)	9	+	-	-
33	Yang Y (2016)	8	-	+	-
34	Zang F (2016)	10	+	-	-
35	Han H (2016)	9	+	-	-
36	Yang J (2016)	9	+	-	-
37	Hong XK (2016)	9	+	-	-
38	Liu L (2015)	9	+	-	-
39	Chen C (2015)	8	-	+	-
40	Han H (2015)	10	+	-	-
41	Wang XR (2015)	9	+	-	-
42	Chen Z (2014)	10	+	-	-
43	Takano A (2014)	10	+	-	-
44	Lee K (2014)	10	+	-	-
45	Hou J (2014)	8	-	+	-

46	Masuzawa T (2014)	9	+	-	-
47	Orkun O (2014)	8	-	+	-
48	Wang W (2013)	9	+	-	-
49	Sun X (2013)	9	+	-	-
50	Wang XL (2013)	8	-	+	-
51	Murase Y (2012)	9	+	-	-
52	Chao LL (2012)	8	-	+	-
53	Scholz HC (2012)	9	+	-	-
54	Li YP (2012)	10	+	-	-
55	Ahantarig A (2011)	10	+	-	-
56	Sen E (2011)	8	-	+	-
57	Yang Y (2011)	9	+	-	-
58	Huang CM (2010)	8	-	+	-
59	Chao LL (2010)	8	-	+	-
60	Geng Z (2010)	8	-	+	-
61	Niu QL (2010)	10	+	-	-
62	Chao LL (2009)	7	-	+	-
63	Wang CS (2009)	8	-	+	-
64	Chu CY (2008)	9	+	-	-
65	He H (2007)	9	+	-	-
66	Gao LY (2007)	9	+	-	-
67	Chu CY (2006)	10	+	-	-
68	Huang HM (2006)	9	+	-	-
69	Hiroko H (2005)	9	+	-	-
70	Zhao QM (2005)	10	+	-	-
71	Güner ES (2003)	9	+	-	-
72	Cao WC (2003)	10	+	-	-
73	Wang ZX (2003)	7	-	+	-
74	Takada N (2001)	9	+	-	-
75	Ishiguro F (2000)	8	-	+	-
76	Hua MT (2000)	9	+	-	-
77	Tian W (1998)	10	+	-	-
78	Takada N (1998)	9	+	-	-
79	Wan KL (1998)	8	-	+	-
80	Seo MG (2020)	8	-	+	-
81	Miyamoto K (1997)	9	+	-	-
82	Sato Y (1996)	10	+	-	-
83	NAKAO M (1992)	8	-	+	-
84	Takada N (1994)	9	+	-	-
85	Mo YM (1994)	7	-	+	-
86	NAKAO M (1992)	9	+	-	-
87	Miyamoto K (1992)	7	-	+	-
88	Du Y (1992)	9	+	-	-
89	Zhag QN (1991)	8	-	+	-
90	Ai CX (1990)	9	+	-	-
91	Li SQ (1990)	9	+	-	-

## Supplementary Material 2: PRISMA checklist

Section/topic	#	Checklist item	Reported on page #
<b>TITLE</b>			
Title	1	Identify the report as a systematic review, meta-analysis, or both.	Title
<b>ABSTRACT</b>			
Structured summary	2	Provide a structured summary including, as applicable: background; objectives; data sources; study eligibility criteria, participants, and interventions; study appraisal and synthesis methods; results; limitations; conclusions and implications of key findings; systematic review registration number.	Abstract
<b>INTRODUCTION</b>			
Rationale	3	Describe the rationale for the review in the context of what is already known.	Introduction, paragraph 1-3
Objectives	4	Provide an explicit statement of questions being addressed with reference to participants, interventions, comparisons, outcomes, and study design (PICOS).	Introduction, paragraph 4
<b>METHODS</b>			
Protocol and registration	5	Indicate if a review protocol exists, if and where it can be accessed (e.g., Web address), and, if available, provide registration information including registration number.	Methods, paragraph 1-8
Eligibility criteria	6	Specify study characteristics (e.g., PICOS, length of follow-up) and report characteristics (e.g., years considered, language, publication status) used as criteria for eligibility, giving rationale.	Methods, paragraph 2
Information sources	7	Describe all information sources (e.g., databases with dates of coverage, contact with study authors to identify additional studies) in the search and date last searched.	Methods, paragraph 1
Search	8	Present full electronic search strategy for at least one database, including any limits used, such that it could be repeated.	Methods, paragraph 1
Study selection	9	State the process for selecting studies (i.e., screening, eligibility, included in systematic review, and, if applicable, included in the meta-analysis).	Methods, paragraph 2
Data collection process	10	Describe method of data extraction from reports (e.g., piloted forms, independently, in duplicate) and any processes for obtaining and confirming data from investigators.	Methods, paragraph 2-3
Data items	11	List and define all variables for which data were sought (e.g., PICOS, funding sources) and any assumptions and simplifications made.	Methods, paragraph 3
Risk of bias in individual studies	12	Describe methods used for assessing risk of bias of individual studies (including specification of whether this was done at the study or outcome level), and how this information is to be used in any data synthesis.	Methods, paragraph 4
Summary measures	13	State the principal summary measures (e.g., risk ratio, difference in means).	Methods, paragraph 4-6
Synthesis of results	14	Describe the methods of handling data and combining results of studies, if done, including measures of consistency (e.g., $I^2$ ) for each meta-analysis.	Methods, paragraph 4-6

Risk of bias across studies	15	Specify any assessment of risk of bias that may affect the cumulative evidence (e.g., publication bias, selective reporting within studies).	<b>Methods, paragraph 4</b>
Additional analyses	16	Describe methods of additional analyses (e.g., sensitivity or subgroup analyses, meta-regression), if done, indicating which were pre-specified.	<b>Methods, paragraph 4</b>
<b>RESULTS</b>			
Study selection	17	Give numbers of studies screened, assessed for eligibility, and included in the review, with reasons for exclusions at each stage, ideally with a flow diagram.	<b>Results, paragraph 1 Figure 1</b>
Study characteristics	18	For each study, present characteristics for which data were extracted (e.g., study size, PICOS, follow-up period) and provide the citations.	<b>Results, paragraph 2 Table 1</b>
Risk of bias within studies	19	Present data on risk of bias of each study and, if available, any outcome level assessment (see item 12).	<b>Results, paragraph 2</b>
Results of individual studies	20	For all outcomes considered (benefits or harms), present, for each study: (a) simple summary data for each intervention group (b) effect estimates and confidence intervals, ideally with a forest plot.	<b>Results, paragraph 3 Figure 2</b>
Synthesis of results	21	Present results of each meta-analysis done, including confidence intervals and measures of consistency.	<b>Results, paragraph 2-4 Table 1</b>
Risk of bias across studies	22	Present results of any assessment of risk of bias across studies (see Item 15).	<b>Results, paragraph 7 Figure 1-3 Table 1</b>
Additional analysis	23	Give results of additional analyses, if done (e.g., sensitivity or subgroup analyses, meta-regression [see Item 16]).	<b>Supplementary</b>
<b>DISCUSSION</b>			
Summary of evidence	24	Summarize the main findings including the strength of evidence for each main outcome; consider their relevance to key groups (e.g., healthcare providers, users, and policy makers).	<b>Discussion, paragraph 1-6</b>
Limitations	25	Discuss limitations at study and outcome level (e.g., risk of bias), and at review-level (e.g., incomplete retrieval of identified research, reporting bias).	<b>Discussion, paragraph 6</b>
Conclusions	26	Provide a general interpretation of the results in the context of other evidence, and implications for future research.	<b>Discussion, paragraph 6</b>
<b>FUNDING</b>			
Funding	27	Describe sources of funding for the systematic review and other support (e.g., supply of data); role of funders for the systematic review.	<b>Acknowledgments</b>

### Supplementary Material 3: Meta-analysis included articles

- [1] Gao Y, Lv XL, Han SZ, et al. First detection of *Borrelia miyamotoi* infections in ticks and humans from the northeast of Inner Mongolia, China. 2021. Acta Trop. 2021, 217:105857.
- [2] Seo HJ, Noh J, Kim HC, et al. Molecular Detection and Phylogenetic Analysis of *Anaplasma* and *Borrelia* Species in Ticks Collected from Migratory Birds at Heuksan, Hong, and Nan Islands, Republic of Korea. Vector Borne Zoonotic Dis. 2021, 21(1):20-31.
- [3] Wang Q, Pan YS, Jiang BG, et al. Prevalence of Multiple Tick-Borne Pathogens in Various Tick Vectors in Northeastern China. Vector Borne Zoonotic Dis. 2021;21(3):162-171.
- [4] Seo MG, Kwon OD, Kwak D. Molecular Identification of *Borrelia afzelii* from Ticks Parasitizing Domestic and Wild Animals in South Korea. Microorganisms. 2020;8(5):649.
- [5] Kim SY, Kim TK, Kim TY, et al. Geographical Distribution of *Borrelia burgdorferi* sensu lato in Ticks Collected from Wild Rodents in the Republic of Korea. Pathogens. 2020;9(11):866.
- [6] Nakayama S, Kobayashi T, Nakamura A, et al. Detection of *Borrelia* DNA in Tick Species Collected from Vegetation and Wild Animals in Fukuoka, Japan. Jpn J Infect Dis. 2020, 73(1):61-64.
- [7] Seto J, Tanaka S, Kawabata H, et al. Detection of tick-borne pathogens in ticks from dogs and cats in Yamagata Prefecture, Japan, 2018. Jpn J Infect Dis. 2021, 74(2):122-128.
- [8] Orkun Ö, Çakmak A, Nalbantoğlu S, et al. Turkey tick news: A molecular investigation into the presence of tick-borne pathogens in host-seeking ticks in Anatolia; Initial evidence of putative vectors and pathogens, and footsteps of a secretly rising vector tick, *Haemaphysalis parva*. Ticks Tick Borne Dis. 2020, 11(3):101373.
- [9] Kaenkan W, Nooma W, Chelong IA, et al. Reptile-associated *Borrelia spp.* In Amblyomma ticks, Thailand. Ticks Tick Borne Dis. 2020, 11(1):101315.
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- [11] Ghafar A, Cabezas-Cruz A, Galon C, et al. Bovine ticks harbour a diverse array of microorganisms in Pakistan. Parasit Vectors. 2020;13(1):1.
- [12] Rar V, Livanova N, Sabitova Y, et al. *Ixodes persulcatus/pavlovskyi* natural hybrids in Siberia: Occurrence in sympatric areas and infection by a wide range of tick-transmitted agents. Ticks Tick Borne Dis. 2019, 10(6):101254.
- [13] Pukhovskaya NM, Morozova OV, Vysochina NP, et al. Prevalence of *Borrelia burgdorferi* sensu lato and *Borrelia miyamotoi* in ixodid ticks in the Far East of Russia. Int J Parasitol Parasites Wildl. 2019, 8:192-202.
- [14] Kim CM, Seo JW, Kim DM, et al. Detection of *Borrelia miyamotoi* in *Ixodes nipponensis* in Korea. PLoS One. 2019, 14(7): e0220465.
- [15] Duan CJ, Guo Y, Hou XX, et al. Investigation of vector and host infection of Lyme disease in Jinghong, Yunnan. Dis Surv. 2019, 34 (3):246-250.
- [16] Jiang BG, Jia N, Jiang JF, et al. *Borrelia miyamotoi* Infections in Humans and

- Ticks, Northeastern China. *Emerg Infect Dis.* 2018, 24(2):236-241.
- [17] Hagen RM, Frickmann H, Ehlers J, et al. Presence of *Borrelia spp.* DNA in ticks, but absence of *Borrelia spp.* and of *Leptospira spp.* DNA in blood of fever patients in Madagascar. *Acta Trop.* 2018, 177:127-134.
- [18] Narankhajid M, Yeruult C, Gurbadam A, et al. Some aspects on tick species in Mongolia and their potential role in the transmission of equine piroplasms, *Anaplasma phagocytophilum* and *Borrelia burgdorferi* L. *Parasitol Res.* 2018, 117(11):3557-3566.
- [19] Zheng WQ, Xuan XN, Fu RL, et al. Tick-Borne Pathogens in Ixodid Ticks from Poyang Lake Region, Southeastern China. *Korean J Parasitol.* 2018, 56(6):589-596.
- [20] Khoo JJ, Ishak SN, Lim FS, et al. Detection of a *Borrelia sp.* from *Ixodes granulatus* ticks collected from rodents in Malaysia. *J Med Entomol.* 2018, 55(6):1642-1647.
- [21] Ye NN, Pei XC, Sun SH. Investigation of the ticks in border trade regions of Heihe. *Chin J Zoon.* 2018, 34(8):761-767.
- [22] Qin LX, Z Q. Investigation on the Infection of *Borrelia Burgdorferi* Carried *Dermacentor sp.* in Yichangerkuang and Xinghuo Pasture in Hailun City of Heilongjiang Province. *Nutr Dis.* 2018, (8):53-57.
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- [24] Tang F, Zhou L, Jiang LF, et al. Investigation on *Borrelia burgdorferi* in ticks collected in Qiqian region of Inner Mongolia. *Infect Dis Info.* 2018, 31(1):31-33.
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- [26] Karasartova D, Gureser AS, Gokce T, et al. Bacterial and protozoal pathogens found in ticks collected from humans in Corum province of Turkey. *PLoS Negl Trop Dis.* 2018, 12(4): e0006395.
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- [28] Pan YP, Yang JF, Niu QL, et al. Study on *Borrelia burgdorferi sensu lato* and spotted fever group *Rickettsia* in *Ixodes persulcatus* in Heilongjiang Province. *Chin Vet Sci.* 2017, 47(1):31-37.
- [29] Li HY, Zhao SS, Zhang L, et al. Molecular investigation on the genera of *Anaplasma* and *Borrelia* in north region of Xinjiang. *J Shihezi Univ: Nat Sci.* 2017, 35(1):108-112.
- [30] Sakakibara K, Şen E, Sato K, et al. Detection and Characterization of the Emerging Relapsing Fever Pathogen, *Borrelia miyamotoi*, from the *Ixodes ricinus* Tick in the Rural Trakya (Thrace) Region of Northwestern Turkey. *Vector Borne Zoonotic Dis.* 2016, 16(12):797-799.
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- [32] Yu PF, Niu QL, Liu ZJ, et al. Molecular epidemiological surveillance to assess emergence and re-emergence of tick-borne infections in tick samples from China



- evaluated by nested PCRs. *Acta Trop.* 2016, 158:181-188.
- [33] Yang Y, Wang J, Wang JC, et al. Investigation on tick-borne pathogens in Ganqimaodu port areas on the borders between China and Mongolia, 2012-2013. *Chin Front Health Quar.* 2016, 39(5):330-332, 347.
- [34] Zang F, Wang W, Li L. *Borrelia Burgdorferi* Infection and Genotypes in Ticks and Rodents Collected in Qinghai Province. *J Prev Med Chin PLA.* 2016, 34(6):799-802.
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- [37] Hong XK, Liu Y, Xu BL, et al. A study on ticks carrying pathogens at the border of China and Korea. *Port Health Control.* 2016, 21 (4):57-59.
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- [49] Sun X, Zhang GL, Liu XM, et al. Investigation of tick species and tick-borne pathogens in Hoxud county of Xinjiang Uyghur Autonomous Region, China. *Chin J*

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# Supplementary Material 4: Study characteristic

Study	Authors	Event	No	Country	Method	Phase	Capture month	Genus	No.infection/No.examined
1	Gao Y (2021)	10	774	China	PCR	-	4-9	264 <i>I. persulcatus</i> , 127 <i>D. nuttalli</i> , 78 <i>D. silvarum</i> , 275 <i>H. longicornis</i>	7/264 <i>I. persulcatus</i> , 1/127 <i>D. nuttalli</i> , 1/78 <i>D. silvarum</i> , 1/275 <i>H. longicornis</i>
2	Seo HJ (2021)	15	380	South Korea	PCR	adult, larva, nymph	-	40 <i>I. nipponensis</i> , 230 <i>I. turdus</i> , 74 <i>H. flava</i> , 2 <i>H. formosensis</i> , 29 <i>H. longicornis</i> , 4 <i>H. phasiana</i> , 1 <i>A. testudinarium</i>	0/40 <i>I. nipponensis</i> , 14/230 <i>I. turdus</i> , 1/74 <i>H. flava</i> , 0/2 <i>H. formosensis</i> , 0/29 <i>H. longicornis</i> , 0/4 <i>H. phasiana</i> , 0/1 <i>A. testudinarium</i>
3	Wang Q (2021)	18	250	China	PCR	adult, nymph	4-6	120 <i>D. silvarum</i> , 21 <i>D. nuttalli</i> , 47 <i>H. concinna</i> , 38 <i>H. japonica</i> , 42 <i>I. persulcatus</i>	1/120 <i>D. silvarum</i> , 0/21 <i>D. nuttalli</i> , 2/47 <i>H. concinna</i> , 2/38 <i>H. japonica</i> , 13/42 <i>I. persulcatus</i>
4	Seo MG (2020)	6	329	South Korea	PCR	adult, nymph	5-9	14 <i>I. nipponensis</i> , 282 <i>H. longicornis</i> , 33 <i>H. flava</i>	2/14 <i>I. nipponensis</i> , 3/282 <i>H. longicornis</i> , 1/33 <i>H. flava</i>
5	Kim SY (2020)	248	738	South Korea	PCR	larva, nymph	3-5, 9-11	193 <i>I. nipponensis</i> , 96 <i>I. angustus</i> , 440 <i>I. spp.</i> , 9 <i>H. longicornis</i>	109/193 <i>I. nipponensis</i> , 3/96 <i>I. angustus</i> , 135/440 <i>I. spp.</i> , 1/9 <i>H. longicornis</i>
6	Nakayama S (2020)	2	561	Japan	PCR	adult, larva, nymph	1-12	3 <i>I. turdus</i> , 5 <i>I. ovatus</i> , 22 <i>A. testudinarium</i> , 213 <i>H. flava</i> , 78 <i>H. formosensis</i> , 24 <i>H. kitaokai</i> , 137 <i>H. longicornis</i> , 15 <i>H. hystricis</i> , 28 <i>H. megaspinosus</i>	1/3 <i>I. turdus</i> , 0/5 <i>I. ovatus</i> , 0/22 <i>A. testudinarium</i> , 0/213 <i>H. flava</i> , 0/78 <i>H. formosensis</i> , 0/24 <i>H. kitaokai</i> , 0/137 <i>H. longicornis</i> , 0/15 <i>H. hystricis</i> , 1/28 <i>H. megaspinosus</i>
7	Seto J (2020)	12	164	Japan	PCR	adult	3-7	9 <i>I. persulcatus</i> , 95 <i>I. ovatus</i> , 4 <i>I. tanuki</i> , 37 <i>I. nipponensis</i> , 7 <i>H. longicornis</i> , 10 <i>H. flava</i> , 2 <i>H. japonica</i>	1/9 <i>I. persulcatus</i> , 11/95 <i>I. ovatus</i> , 0/37 <i>I. nipponensis</i> , 0/4 <i>I. tanuki</i> , 0/7 <i>H. longicornis</i> , 0/10 <i>H. flava</i> , 0/2 <i>H. japonica</i>
8	Orkun Ö (2020)	3	1019	Turkey	PCR	adult, nymph	-	30 <i>I. ricinus</i> , 739 <i>H. parva</i> , 16 <i>H. sulcata</i> , 3 <i>H. punctata</i> , 105 <i>H. marginatum</i> , 11 <i>H. excavatum</i> , 9 <i>H. aegyptium</i> , 41 <i>D. marginatus</i> , 2 <i>D. reticulatus</i> , 9 <i>Rh. turanicus</i>	3/30 <i>I. ricinus</i> , 0/739 <i>H. parva</i> , 0/16 <i>H. sulcata</i> , 0/3 <i>H. punctata</i> , 0/105 <i>H. marginatum</i> , 0/11 <i>H. excavatum</i> , 0/9 <i>H. aegyptium</i> , 0/41 <i>D. marginatus</i> , 0/2 <i>D. reticulatus</i> , 0/9 <i>Rh. turanicus</i>

9	Kaenkan W (2020)	20	127	Thailand	PCR	adult, nymph	-	28 <i>A. helvolum</i> , 62 <i>A. varanense</i> , 31 <i>A. geoemydae</i> , 6 <i>A. spp</i>	1/28 <i>A. helvolum</i> , 17/62 <i>A. varanense</i> , 2/31 <i>A. geoemydae</i> , 0/6 <i>A. spp</i>
10	Fischer T (2020)	1	310	Mongolia	PCR	adult	5-6	310 <i>D. nuttalli</i>	1/310 <i>D. nuttalli</i>
11	Ghafar A (2020)	15	234	Pakistan	PCR	-	9-11	212 <i>Hy. Anatolicum</i> , 3 <i>Hy. Hussaini</i> , 1 <i>Hy. Scupense</i> , 17 <i>Rh. Microplus</i> , 1 <i>Rh. annulatus</i>	14/212 <i>Hy. Anatolicum</i> , 0/3 <i>Hy. Hussaini</i> , 0/1 <i>Hy. Scupense</i> , 1/17 <i>Rh. Microplus</i> , 0/1 <i>Rh. annulatus</i>
12	Rar V (2019)	742	1822	Russia (Siberia)	PCR	-	5-6	668 <i>I. persulcatus</i> , 1154 <i>I. pavlovskyi</i>	252/668 <i>I. persulcatus</i> , 490/1154 <i>I. pavlovskyi</i>
13	Pukhovskaya NM (2019)	605	2158	Russia (Siberia)	PCR	-	-	1599 <i>I. persulcatus</i> , 481 <i>H. concinna</i> , 49 <i>H. japonica</i> , 29 <i>D. silvarum</i>	582/1599 <i>I. persulcatus</i> , 14/481 <i>H. concinna</i> , 7/49 <i>H. japonica</i> , 2/29 <i>D. silvarum</i>
14	Kim CM (2019)	5	484	South Korea	PCR	adult, larva, nymph	4-9	25 <i>I. nipponensis</i> , 417 <i>H. longicornis</i> , 42 <i>H. flava</i>	5/25 <i>I. nipponensis</i> , 0/417 <i>H. longicornis</i> , 0/42 <i>H. flava</i>
15	Duan CJ (2019)	3	724	China	other	-	3	724 <i>R(B). microplus</i>	3/724 <i>R(B). microplus</i>
16	Jiang BG (2018)	20	692	China	PCR	adult	-	627 <i>I. persulcatus</i> , 36 <i>H. concinna</i> , 29 <i>D. silvarum</i>	19/627 <i>I. persulcatus</i> , 1/36 <i>H. concinna</i> , 0/29 <i>D. silvarum</i>
17	Hagen RM (2018)	23	403	South Korea	PCR	adult, nymph	-	255 <i>A. variegatum</i> , 148 <i>R. microplus</i>	21/255 <i>A. variegatum</i> , 2/148 <i>R. microplus</i>
18	Narankhajid M (2018)	9	360	Mongolia	PCR	adult	-	16 <i>I. persulcatus</i> , 402 <i>D. nuttalli</i> , 44 <i>D. marginatus</i> , 6 <i>D. silvarum</i> , 60 <i>H. asiaticum</i>	2/7 <i>I. persulcatus</i> , 6/272 <i>D. nuttalli</i> , 0/35 <i>D. marginatus</i> , 0/2 <i>D. silvarum</i> , 1/44 <i>H. asiaticum</i>
19	Zheng WQ (2018)	3	311	China	PCR	adult, larva, nymph	-	16 <i>I. granulatus</i> , 1 <i>I. acuminatus</i> , 4 <i>I. sinensis</i> , 173 <i>H. longicornis</i> , 21 <i>H. flava</i> , 77 <i>R. microplus</i> , 5 <i>H. phasiana</i> , 9 <i>R. sanguineus</i> , 3 <i>R. haemaphysaloides</i> , 1 <i>A. testudinarium</i> , 1 <i>D. auratus</i>	3/16 <i>I. granulatus</i> , 0/1 <i>I. acuminatus</i> , 0/4 <i>I. sinensis</i> , 0/173 <i>H. longicornis</i> , 0/21 <i>H. flava</i> , 0/77 <i>R. microplus</i> , 0/5 <i>H. phasiana</i> , 0/9 <i>R. sanguineus</i> , 0/3 <i>R. haemaphysaloides</i> , 0/1 <i>A. testudinarium</i> , 0/1 <i>D. auratus</i>
20	Khoo JJ (2018)	72	156	Malaysia	PCR	-	1-12	122 <i>I. granulatus</i> , 24 <i>D. spp</i> , 3 <i>A. spp</i>	72/122 <i>I. granulatus</i> , 0/24 <i>D. spp</i> , 0/3 <i>A. spp</i>
21	Ye NN (2018)	0	1111	China	PCR	-	4-10	1054 <i>D. silvarum</i> , 236 <i>H. concinna</i> , 53 <i>I. persulcatus</i>	0/1054 <i>D. silvarum</i> , 0/236 <i>H. concinna</i> , 0/53 <i>I. persulcatus</i>

22	Qin LX (2018)	51	96	China	PCR	-	-	96 <i>D. spp</i>	51/96 <i>D. spp</i>
23	Yang Y (2018)	20	388	China	PCR	-	5-7	267 <i>I. persulcatus</i> , 559 <i>D. nuttalli</i>	0/125 <i>I. persulcatus</i> , 20/263 <i>D. nuttalli</i>
24	Tang F (2018)	86	320	China	PCR	-	4-6	293 <i>I. persulcatus</i> , 22 <i>D. silvarum</i> , 5 <i>H. concinna</i>	85/293 <i>I. persulcatus</i> , 1/22 <i>D. silvarum</i> , 0/5 <i>H. concinna</i>
25	Liu XY (2017)	0	500	China	PCR	adult	1-12	500 <i>H. longicornis</i>	0/500 <i>H. longicornis</i>
26	Karasartova D (2017)	1	322	Turkey	PCR	-	-	4 <i>I. ricinus</i> , 164 <i>H. marginatum</i> , 5 <i>H. excavatum</i> , 1 <i>H. aegyptium</i> , 41 <i>H. Parva</i> , 6 <i>H. Punctata</i> , 1 <i>H.</i> <i>Sulcata</i> , 46 <i>H. spp.</i> , 34 <i>R. turanicus</i> , 3 <i>R. bursa</i> , 17 <i>D. marginatus</i> 42 <i>H. doenitzi</i> , 1 <i>H. flava</i> , 2 <i>H.</i> <i>formosensis</i> , 12 <i>H. hystricis</i> , 21 <i>H.</i> <i>ornithophila</i> , 17 <i>H. wellingtoni</i> , 17 <i>I. columnae</i> , 6 <i>A.</i> <i>spp</i> 5 <i>I. granulatus</i> , 1 <i>I. nipponensis</i> , 4 <i>I.</i> <i>turdus</i> , 1 <i>R. haemaphysaloides</i> , 4 <i>I. spp.</i> , 6 <i>H. spp.</i> ,	1/4 <i>I. ricinus</i> , 0/164 <i>H. marginatum</i> , 0/5 <i>H. excavatum</i> , 0/1 <i>H. aegyptium</i> , 0/41 <i>H. Parva</i> , 0/6 <i>H. Punctata</i> , 0/1 <i>H.</i> <i>Sulcata</i> , 0/46 <i>H. spp.</i> , 0/34 <i>R. turanicus</i> , 0/3 <i>R. bursa</i> , 0/17 <i>D. marginatus</i> 3/4 <i>I. granulatus</i> , 2/5 <i>I. turdus</i> , 0/24 <i>H.</i> <i>doenitzi</i> , 0/1 <i>H. flava</i> , 0/1 <i>H.</i> <i>formosensis</i> , 0/9 <i>H. hystricis</i> , 0/6 <i>H.</i> <i>ornithophila</i> , 0/8 <i>H. wellingtoni</i> , 0/17 <i>I. columnae</i> , 0/1 <i>I. nipponensis</i> , 0/1 <i>R. haemaphysaloides</i> , 0/3 <i>H. spp.</i> , 0/5 <i>A. spp.</i>
27	Kuo CC (2017)	5	85	China	PCR	adult, larva, nymph	1-12	17 <i>H. wellingtoni</i> , 17 <i>I. columnae</i> , 6 <i>A.</i> <i>spp</i> 5 <i>I. granulatus</i> , 1 <i>I. nipponensis</i> , 4 <i>I.</i> <i>turdus</i> , 1 <i>R. haemaphysaloides</i> , 4 <i>I. spp.</i> , 6 <i>H. spp.</i> ,	3/4 <i>I. granulatus</i> , 2/5 <i>I. turdus</i> , 0/24 <i>H.</i> <i>doenitzi</i> , 0/1 <i>H. flava</i> , 0/1 <i>H.</i> <i>formosensis</i> , 0/9 <i>H. hystricis</i> , 0/6 <i>H.</i> <i>ornithophila</i> , 0/8 <i>H. wellingtoni</i> , 0/17 <i>I. columnae</i> , 0/1 <i>I. nipponensis</i> , 0/1 <i>R. haemaphysaloides</i> , 0/3 <i>H. spp.</i> , 0/5 <i>A. spp.</i>
28	Pan YP (2017)	5	25	China	PCR	adult	4-5	25 <i>I. persulcatus</i>	5/25 <i>I. persulcatus</i>
29	Li HY (2017)	97	546	China	PCR	-	4-5	210 <i>H. asiaticum</i> , 209 <i>H. punctate</i> , 61 <i>D. nuttalli</i> , 40 <i>D. marginatus</i> , 26 <i>R. turanicus</i>	-
30	Sakakibara K (2016)	1	248	Turkey	PCR	adult	6	248 <i>I. ricinus</i>	1/248 <i>I. ricinus</i>
31	Khasnatino v MA (2016)	90	445	Russia (Siberia)	PCR	adult	-	445 <i>I. persulcatus</i>	0/445 <i>I. persulcatus</i>
32	Yu PF (2016)	156	849	China	PCR	-	-	34 <i>H. bispinosa</i> , 165 <i>D. silvarum</i> ,	11/ 34 <i>H. bispinosa</i> , 3/165 <i>D. silvarum</i> ,

								213 <i>R. (B.) microplus</i> , 125 <i>H. qinghaiensis</i> , 186 <i>R. sanguineus</i> , 84 <i>H. longicornis</i> , 42 <i>I. persulcatus</i>	18/213 <i>R. (B.) microplus</i> , 18/42 <i>I. persulcatus</i> , 3/125 <i>H. qinghaiensis</i> , 65/84 <i>H. longicornis</i> , 38/186 <i>R. sanguineus</i>
33	Yang Y (2016)	0	178	China	PCR	-	-	178 <i>H. asiaticum</i>	0/178 <i>H. asiaticum</i>
34	Zang F (2016)	97	618	China	PCR	adult	3-4	249 <i>D. nuttalli</i> , 152 <i>H. garhwalensis</i> , 217 <i>H. qinghaiensis</i>	28/249 <i>D. nuttalli</i> , 22/152 <i>H. garhwalensis</i> , 47/217 <i>H. qinghaiensis</i>
35	Han H (2016)	3	232	China	PCR	-	-	69 <i>H. nepalensis</i> , 5 <i>H. formosensis</i> , 86 <i>R. microplus</i> , 72 unidentified	3/86 <i>R. microplus</i> , 0/69 <i>H. nepalensis</i> , 0/5 <i>H. formosensis</i> , 0/72 unidentified
36	Yang J (2016)	0	257	China	PCR	-	4-6	106 <i>H. concinna</i> , 84 <i>I. persulcatus</i> , 66 <i>D. silvarum</i> , 1 <i>H. japonica</i>	0/106 <i>H. concinna</i> , 0/84 <i>I. persulcatus</i> , 0/66 <i>D. silvarum</i> , 0/1 <i>H. japonica</i>
37	Hong XK (2016)	2	90	China	PCR	-	3-10	-	-
38	Liu L (2015)	0	292	China	PCR	adult	5-6	113 <i>I. persulcatus</i> , 179 <i>D. nuttalli</i>	0/113 <i>I. persulcatus</i> , 0/179 <i>D. nuttalli</i>
39	Cheng C (2015)	4	44	China	PCR	-	4-6	8 <i>I. persulcatus</i> , 34 <i>H. concinna</i> , 2 <i>D. silvarum</i>	3/8 <i>I. persulcatus</i> , 1/34 <i>H. concinna</i> , 0/2 <i>D. silvarum</i>
40	Han H (2015)	12	643	China	PCR	-	5	573 <i>H. concinna</i> , 70 <i>I. persulcatus</i>	1/573 <i>H. concinna</i> , 11/70 <i>I. persulcatus</i>
41	Wang XR(2015)	53	227	China	other	-	3-8	117 <i>H. qinghaiensis</i> , 110 <i>D. nuttalli</i>	28/117 <i>H. qinghaiensis</i> , 25/110 <i>D. nuttalli</i>
42	Chen Z (2014)	0	308	China	PCR	adult, larva, nymph	4-12	298 <i>H. longicornis</i> , 10 <i>R. microplus</i>	0/298 <i>H. longicornis</i> , 0/10 <i>R. microplus</i>
43	Takano A (2014)	77	4325	Japan	PCR	adult	4-7	3532 <i>I. persulcatus</i> , 117 <i>I. pavlovskyi</i> , 676 <i>I. ovatus</i>	71/3532 <i>I. persulcatus</i> , 5/117 <i>I. pavlovskyi</i> , 1/676 <i>I. ovatus</i>
44	Lee K (2014)	6	670	Japan	PCR	adult, nymph	5-9	670 <i>H. spp.</i>	6/670 <i>H. spp.</i>
45	Hou J (2014)	21	447	China	PCR	adult	1-12	212 <i>H. longicornis</i> , 20 <i>R. haemaphysaloides</i> , 29 <i>A. testudinarium</i> , 94 <i>I. sinensis</i> , 20 <i>B. microplus</i> , 19 <i>H. yeni</i> , 9 <i>D. taiwanensis</i> , 5 <i>H. hystricis</i> ,	11/212 <i>H. longicornis</i> , 0/20 <i>R. haemaphysaloides</i> , 1/29 <i>A. testudinarium</i> , 1/94 <i>I. sinensis</i> , 1/20 <i>B. microplus</i> , 0/19 <i>H. yeni</i> , 0/9 <i>D. taiwanensis</i> , 0/5 <i>H. hystricis</i> ,



								1 <i>H. asiaticum</i> , 36 <i>I. granulatus</i> , 2 <i>I. ovatus</i>	0/1 <i>H. asiaticum</i> , 7/36 <i>I. granulatus</i> , 0/2 <i>I. ovatus</i>
46	Masuzawa T (2014)	71	129	Mongolia	PCR	-	6	129 <i>I. persulcatus</i>	71/129 <i>I. persulcatus</i>
47	Orkun O (2014)	6	169	Turkey	PCR	-	1-12	17 <i>H. excavatum</i> , 30 <i>H. marginatum</i> , 6 <i>Hy. spp.</i> , 35 <i>H. Parva</i> , 28 <i>R. turanicus</i> , 25 <i>D. marginatus</i> , 16 <i>H. aegyptium</i> , 3 <i>H. punctata</i> , 3 <i>R. bursa</i> , 3 <i>R. sanguineus</i> , 2 <i>I. ricinus</i> , 1 <i>H. spp.</i>	1/17 <i>H. excavatum</i> , 1/30 <i>H. marginatum</i> , 2/6 <i>Hy. spp.</i> , 2/35 <i>H. Parva</i> , 0/28 <i>R. turanicus</i> , 0/25 <i>D. marginatus</i> , 0/16 <i>H. aegyptium</i> , 0/3 <i>H. punctata</i> , 0/3 <i>R. bursa</i> , 0/3 <i>R. sanguineus</i> , 0/2 <i>I. ricinus</i> , 0/1 <i>H. spp.</i>
48	Wang W (2013)	0	197	China	other	-	-	58 <i>I. persulcatus</i> , 139 <i>H. longicornis</i>	0/58 <i>I. persulcatus</i> , 0/139 <i>H. longicornis</i>
49	Sun X (2013)	18	367	China	PCR	-	4	174 <i>H. asiaticum</i> , 138 <i>R. pumilio</i> , 14 <i>R. turanicus</i> , 41 <i>D. marginatus</i>	3/174 <i>H. asiaticum</i> , 15/138 <i>R. pumilio</i> , 0/14 <i>R. turanicus</i> , 0/41 <i>D. marginatus</i>
50	Wang XL (2013)	229	542	China	PCR	-	-	-	-
51	Murase Y (2012)	119	338	Japan	PCR	-	4	284 <i>I. persulcatus</i> , 41 <i>I. ovatus</i> , 13 <i>H. longicornis</i>	98/284 <i>I. persulcatus</i> , 8/41 <i>I. ovatus</i> , 1/13 <i>H. longicornis</i>
52	Chao LL (2012)	123	261	China	PCR	adult, larva	-	261 <i>I. granulatus</i>	123/ 261 <i>I. granulatus</i>
53	Scholz HC (2012)	91	372	Mongolia	PCR	adult	5	372 <i>I. persulcatus</i>	91/372 <i>I. persulcatus</i>
54	Li YP (2012)	304	2460	China	PCR	-	4-6	-	-
55	Ahantari A (2011)	0	166	Thailand	PCR	adult, larva, nymph	-	166 <i>H. spp.</i>	0/166 <i>H. spp.</i>
56	Sen E (2011)	10	24	Turkey	PCR	-	6	24 <i>I. ricinus</i>	10/24 <i>I. ricinus</i>
57	Yang Y (2011)	5	162	China	PCR	-	5	162 <i>H. asiaticum</i>	5/162 <i>H. asiaticum</i>
58	Huang CM (2010)	23	113	China	PCR	adult, larva, nymph	-	113 <i>I. granulatus</i>	23/113 <i>I. granulatus</i>
59	Chao LL (2010)	27	147	China	PCR	adult	-	147 <i>I. granulatus</i>	27/147 <i>I. granulatus</i>

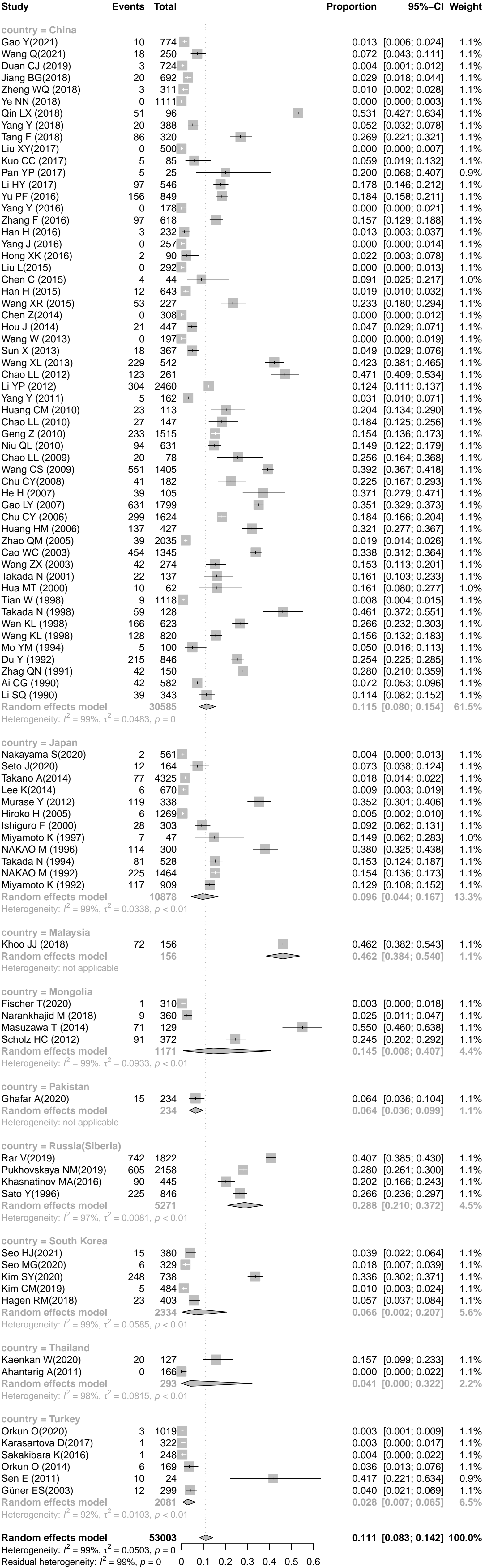
60	Geng Z (2010)	233	1515	China	other	-	-	-	-
61	Niu QL (2010)	94	631	China	PCR	-	4-7	264 <i>D. spp.</i> , 173 <i>H. spp.</i> , 185 <i>R. Curtice</i> , 45 <i>R. spp.</i>	10/264 <i>D.spp</i> , 11/173 <i>H. spp.</i> , 59/185 <i>R. Curtice</i> , 14/45 <i>R. spp.</i>
62	Chao LL (2009)	20	78	China	PCR	adult	-	78 <i>I. granulatus</i>	20/78 <i>I. granulatus</i>
63	Wang CS(2009)	551	1405	China	other	-	-	1239 <i>I. persulcatus</i> , 110 <i>H. concinna</i>	444/1239 <i>I. persulcatus</i> , 9/110 <i>H. concinna</i>
64	Chu CY (2008)	41	182	China	PCR	adult	10-12	4 <i>I. vespertlionis</i> , 1 <i>I. hyatti</i> , 2 <i>H. concinna</i> , 114 <i>H. longicornis</i> , 40 <i>H. warburconi</i> , 21 <i>R. microplus</i>	0/4 <i>I. vespertlionis</i> , 0/1 <i>I. hyatti</i> , 1/2 <i>H. concinna</i> , 32/114 <i>H. longicornis</i> , 5/40 <i>H. warburconi</i> , 3/21 <i>R. microplus</i>
65	He H (2007)	39	105	China	other	-	-	2065 <i>I. persulcatus</i> , 2438 <i>D. silvarum</i> , 1848 <i>H. concinna</i> , 203 <i>H. japonica</i>	-
66	Gao LY (2007)	631	1799	China	other	-	-	1799 <i>I. persulcatus</i> ,	631/1799 <i>I. persulcatus</i>
67	Chu CY (2006)	299	1624	China	PCR	-	5-7	1336 <i>I. persulcatus</i> , 144 <i>D. silvarum</i> , 144 <i>H. concinna</i>	293/1336 <i>I. persulcatus</i> , 6/144 <i>D. silvarum</i> , 0/144 <i>H. concinna</i>
68	Huang HM (2006)	137	427	China	PCR	-	4-5	100 <i>I.persulcatus</i> , 327 <i>D.silvarum</i>	36/100 <i>I.persulcatus</i> , 101/327 <i>D.silvarum</i>
69	Hiroko H (2005)	6	1269	Japan	PCR	-	-	-	-
70	Zhao QM (2005)	39	2035	China	PCR	-	4-10	-	-
71	Güner ES(2003)	12	299	Turkey	other	-	5	299 <i>I. ricinus</i>	12/299 <i>I. ricinus</i>
72	Cao WC (2003)	454	1345	China	PCR	-	6-8	1345 <i>I. persulcatus</i>	454/1345 <i>I. persulcatus</i>
73	Wang ZX (2003)	42	274	China	other	-	1-12	-	-
74	Takada N (2001)	22	137	China	PCR	adult, larva, nymph	5	66 <i>I. persulcatus</i>	26/66 <i>I. persulcatus</i>
75	Ishiguro F (2000)	28	303	Japan	PCR	adult, larva, nymph	1-5, 9-12	343 <i>H. flava</i> , 6 <i>H. longicornis</i> , 1 <i>I. columnae</i> , 9 <i>I. persulcatus</i> ,	0/2 <i>H. flava</i> , 0/2 <i>H. longicornis</i> , 0/1 <i>H. phasiana</i> , 0/11 <i>I. turdus</i>

								1 <i>H. phasiana</i> , 15 <i>I. turdus</i> , 2 unidentified	
76	Hua MT (2000)	10	62	China	other	-	5	1543 <i>D. silvarum</i> , 1099 <i>D. niveus</i> , 383 <i>D. nuttalli</i> , 58 <i>D. marginatus</i> , 3 <i>H. concinna</i> , 2 <i>H. campanulata</i> , 1 <i>H. detritum</i> , 1 <i>I. persulcatus</i>	6/32 <i>D. silvarum</i> , 4/30 <i>D. niveus</i>
77	Tian W (1998)	9	1118	China	PCR	adult, larva, nymph	5-9	947 <i>H. japonicum</i> and <i>H. longicornis</i> , 158 <i>I. persulcatus</i> , 13 <i>D. silvarum</i> 21 <i>I. nipponensis</i> , 287 <i>I. persulcatus</i> , 1 <i>I. pavlovskyi</i> , 42 <i>H. concinna</i> , 7 <i>H.</i>	4/947 <i>H. japonicum</i> and <i>H. longicornis</i> , 5/158 <i>I. persulcatus</i> , 0/13 <i>D. silvarum</i>
78	Takada N (1998)	59	128	China	PCR	adult	5	<i>flava</i> , 15 <i>H. douglasi</i> , 45 <i>H. longicornis</i> , 6 <i>H. megaspinosa</i> , 2 <i>H. punctata</i> , 59 <i>H.</i> <i>spp</i> , 12 <i>D. silvarum</i> , 7 <i>D. silvarum</i>	-
79	Wan KL (1998)	166	623	China	other	-	-	-	-
80	Wan KL (1998)	128	820	China	other	-	-	-	-
81	Miyamoto K (1997)	7	47	Japan	PCR	adult, larva, nymph	10	113 <i>I. persulcatus</i>	7/47 <i>I. persulcatus</i>
82	Sato Y (1996)	225	846	Russia (Siberia)	other	adult	5-6	846 <i>I. persulcatus</i>	225/846 <i>I. persulcatus</i>
83	NAKAO M (1996)	114	300	Japan	PCR	adult	5-7	300 <i>I. persulcatus</i>	112/300 <i>I. persulcatus</i>
84	Takada N (1994)	81	528	Japan	other	adult, larva	4-7	1 <i>A. testudinarium</i> , 8 <i>D. taiwanensis</i> , 40 <i>H. flava</i> , 6 <i>H. japonica</i> , 23 <i>H. kitaokai</i> , 7 <i>H. longicornis</i> , 12 <i>H. megaspinosa</i> , 2 <i>I. angustus</i> , 58 <i>I. monospinosus</i> , 6 <i>I. nipponensis</i> , 193 <i>I. ovatus</i> , 85 <i>I. persulcatus</i> , 2 <i>I. spp.</i>	37/193 <i>I. ovatus</i> , 11/85 <i>I. persulcatus</i> , 0/1 <i>A. testudinarium</i> , 0/8 <i>D.</i> <i>taiwanensis</i> , 1/40 <i>H. flava</i> , 0/6 <i>H. japonica</i> , 0/23 <i>H.</i> <i>kitaokai</i> , 0/12 <i>H. megaspinosa</i> , 0/58 <i>I.</i> <i>monospinosus</i>
85	Mo YM (1994)	5	100	China	other	-	-	308 <i>R(B). microplus</i> , 136 <i>H. hystricis</i> , 40 <i>H. bispinosa</i>	0/40 <i>R(B). microplus</i> , 2/40 <i>H. hystricis</i> , 3/20 <i>H. bispinosa</i>
86	NAKAO M (1992)	225	1464	Japan	other	adult	1-12	777 <i>I. persukatus</i> , 687 <i>I. ovatus</i>	129/777 <i>I. persukatus</i> , 162/687 <i>I. ovatus</i>
87	Miyamoto K (1992)	117	909	Japan	other	adult, larva	4-8	273 <i>I. ovatus</i> , 792 <i>I. persulcatus</i>	41/273 <i>I. ovatus</i> , 130/792 <i>I. persulcatus</i>

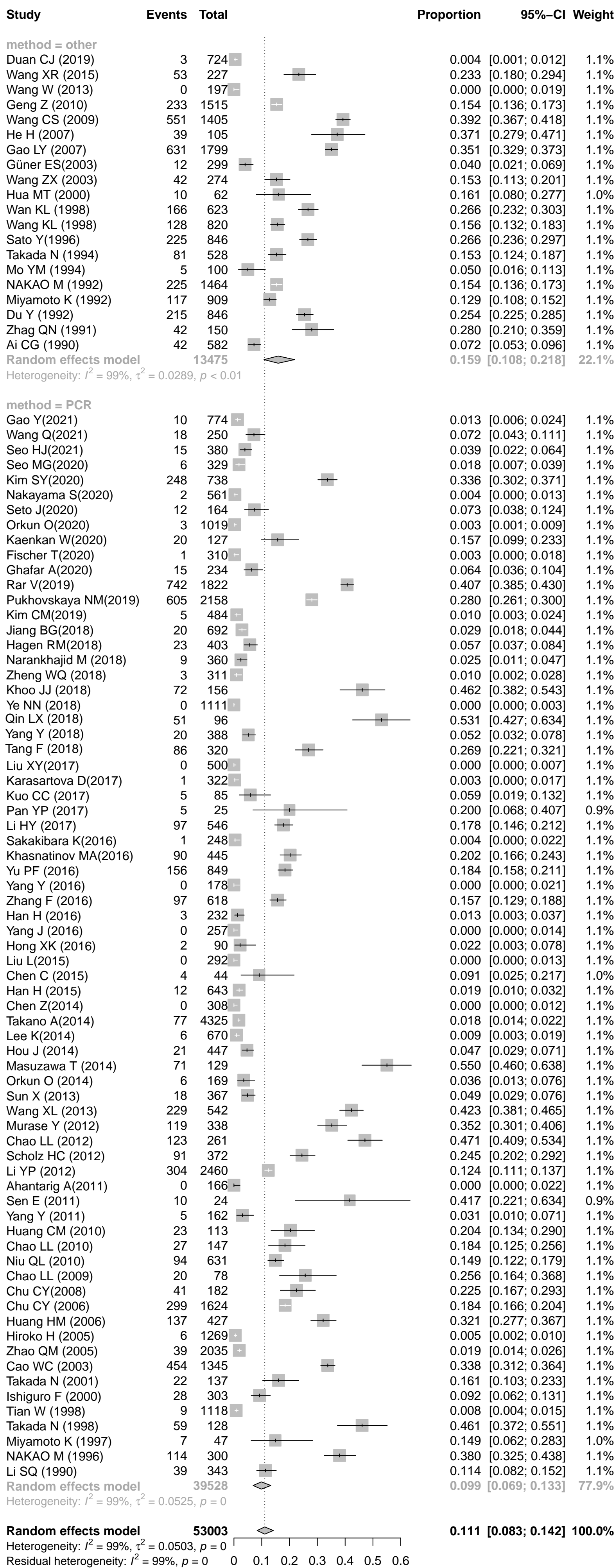
88	Du Y (1992)	215	846	China	other	-	4-7	766 <i>I. persulcatus</i> , 62 <i>H. japonica</i> , 18 <i>D. silvarum</i>	215/766 <i>I. persulcatus</i> , 0/62 <i>H.</i> <i>japonica</i> , 18 <i>D. silvarum</i>
89	Zhag QN (1991)	42	150	China	other	adult	4-7	150 <i>I. persulcatus</i> , 105 <i>D. marginatus</i> , 45 <i>H. spp.</i>	42/150 <i>I. persulcatus</i> , 0/105 <i>D.</i> <i>marginatus</i> , 0/45 <i>H. spp</i>
90	Ai CX (1990)	42	582	China	other	adult	4-8	567 <i>I. persulcatus</i> , 15 <i>H. concinna</i>	-
91	Li SQ (1990)	39	343	China	PCR	adult, larva, nymph	4-7	-	-

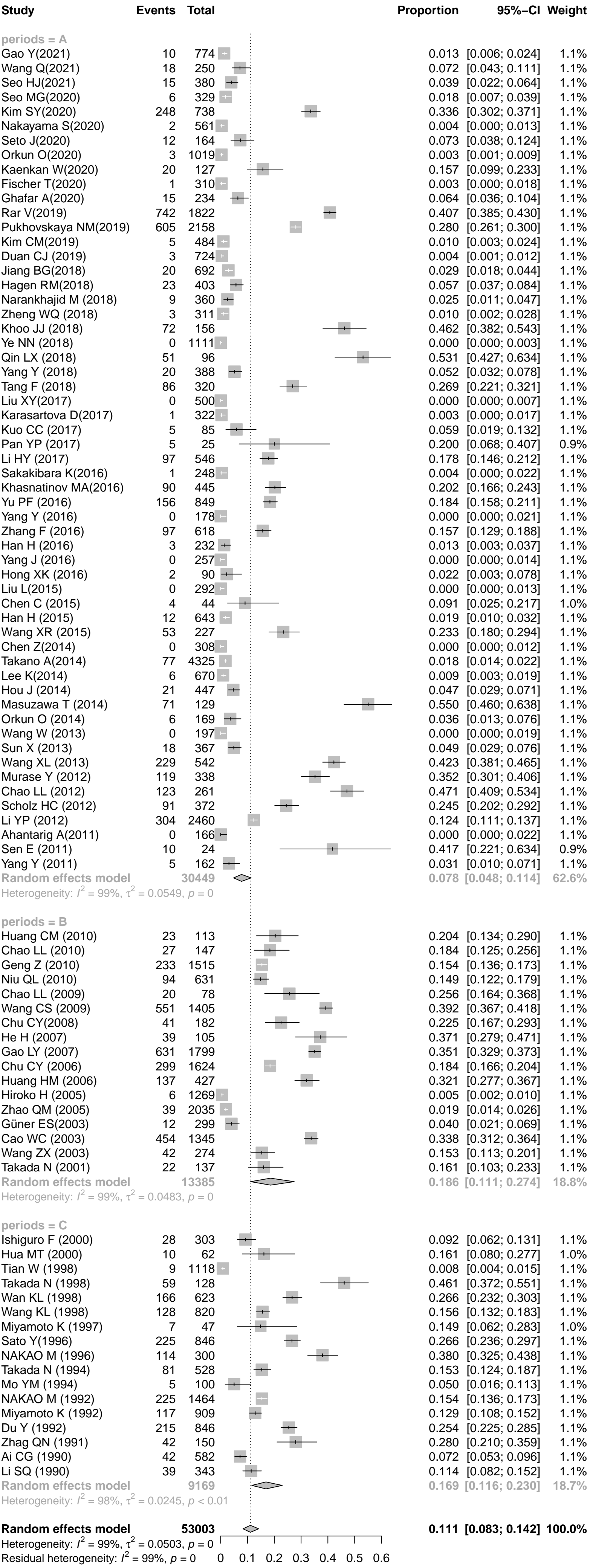
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Supplementary Material 5: Asia prevalence of B. burgdorferi in Ixodidae by subgroup

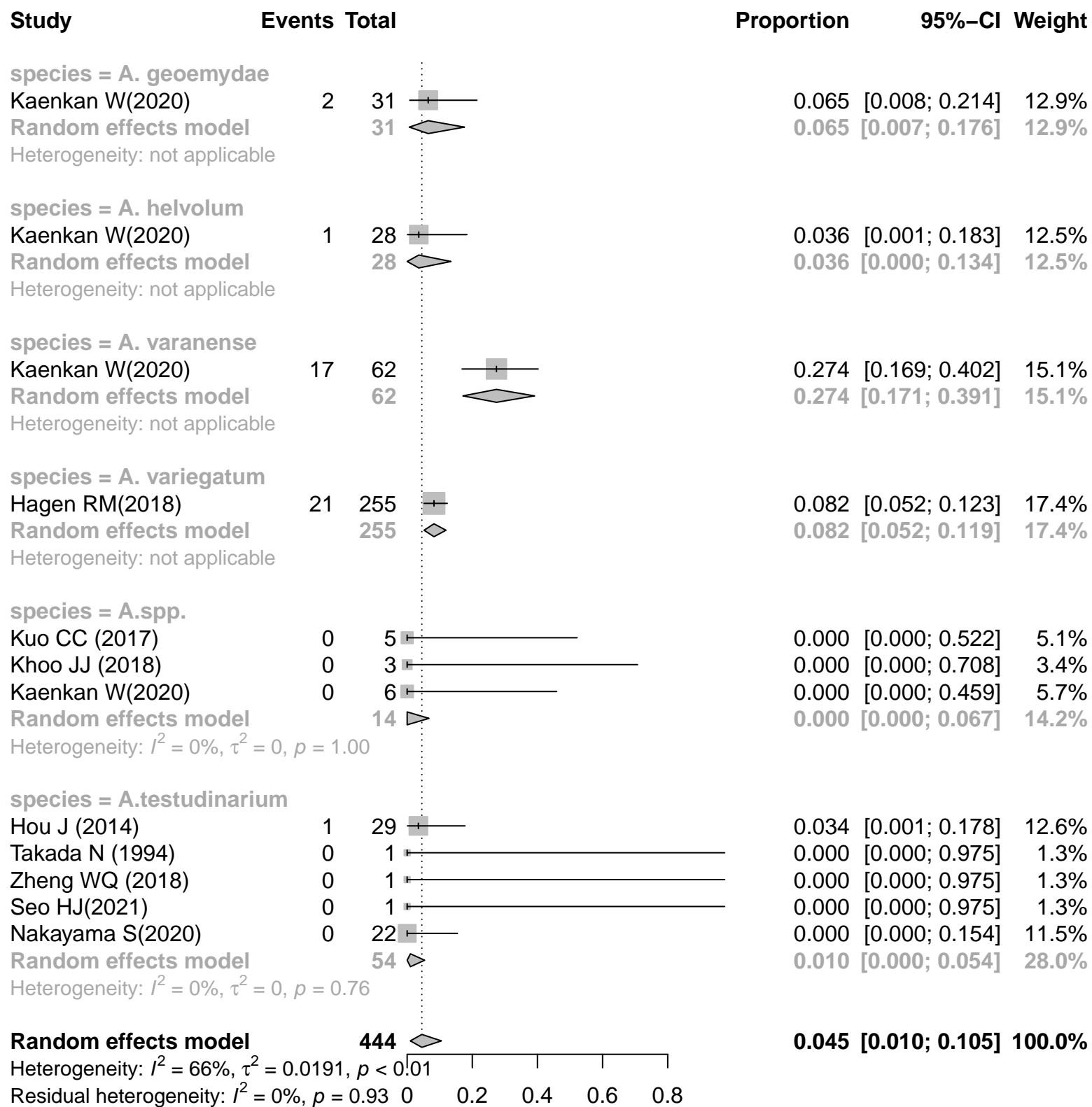


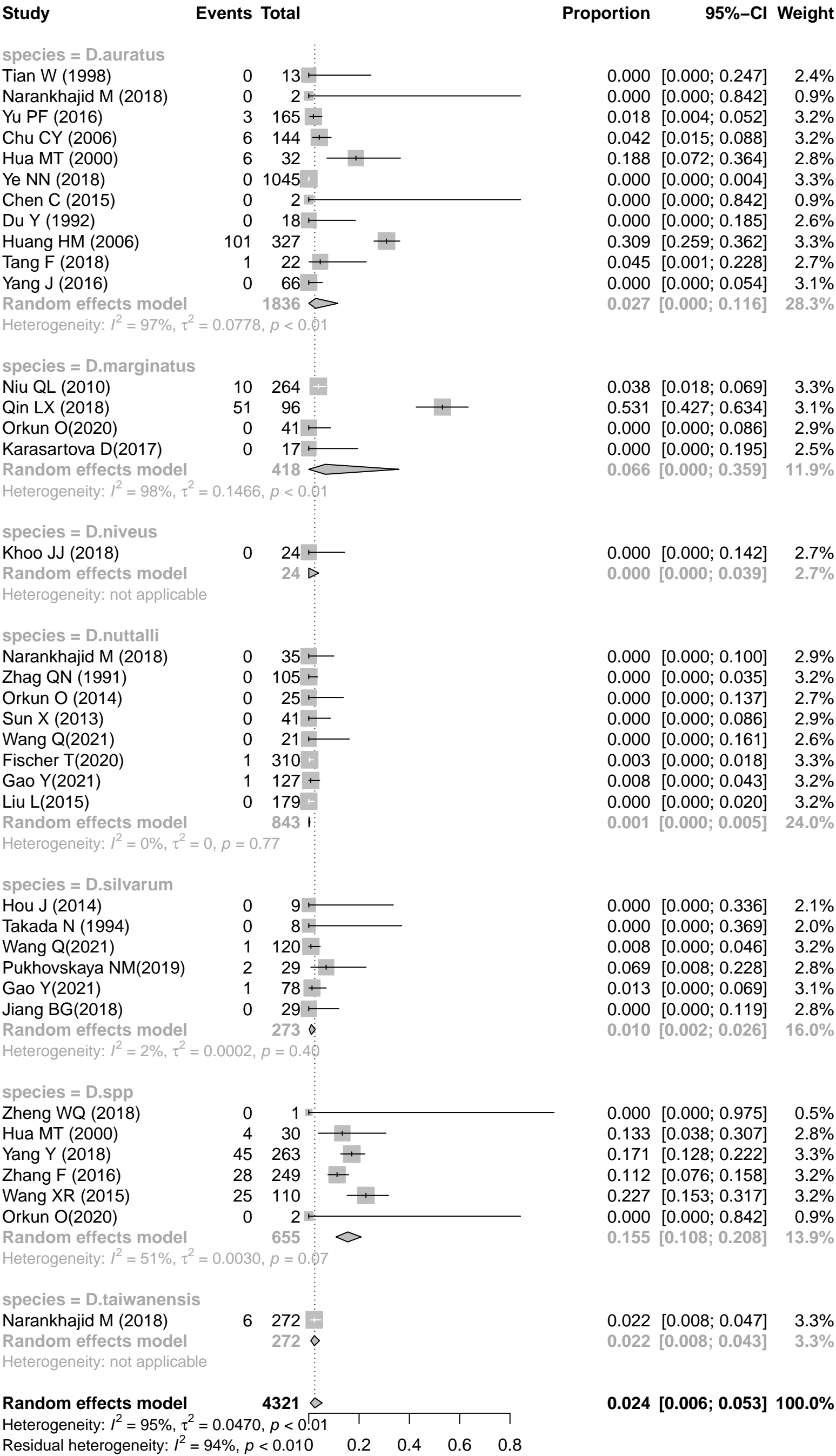
Study	Events	Total	Proportion	95%-CI	Weight
genus = Amblyomma					
Seo HJ(2021)	0	1	0.000	[0.000; 0.975]	0.1%
Nakayama S(2020)	0	22	0.000	[0.000; 0.154]	0.6%
Kaenkan W(2020)	20	127	0.157	[0.099; 0.233]	0.7%
Hagen RM(2018)	21	255	0.082	[0.052; 0.123]	0.7%
Zheng WQ (2018)	0	1	0.000	[0.000; 0.975]	0.1%
Khoo JJ (2018)	0	3	0.000	[0.000; 0.708]	0.3%
Kuo CC (2017)	0	5	0.000	[0.000; 0.522]	0.4%
Hou J (2014)	1	29	0.034	[0.001; 0.178]	0.6%
Takada N (1994)	0	1	0.000	[0.000; 0.975]	0.1%
Random effects model		444	0.048	[0.011; 0.108]	3.7%
Heterogeneity: $I^2 = 59\%$ , $\tau^2 = 0.0110$ , $p = 0.01$					
genus = Dermacentor					
Wang Q(2021)	26	141	0.184	[0.124; 0.258]	0.7%
Gao Y(2021)	2	205	0.010	[0.001; 0.035]	0.7%
Orkun O(2020)	0	43	0.000	[0.000; 0.082]	0.6%
Fischer T(2020)	1	310	0.003	[0.000; 0.018]	0.7%
Pukhovskaya NM(2019)	2	29	0.069	[0.008; 0.228]	0.6%
Jiang BG(2018)	0	29	0.000	[0.000; 0.119]	0.6%
Khoo JJ (2018)	0	24	0.000	[0.000; 0.142]	0.6%
Yang Y (2018)	20	263	0.076	[0.047; 0.115]	0.7%
Tang F (2018)	1	22	0.045	[0.001; 0.228]	0.6%
Ye NN (2018)	0	1045	0.000	[0.000; 0.004]	0.7%
Qin LX (2018)	51	96	0.531	[0.427; 0.634]	0.7%
Zheng WQ (2018)	0	1	0.000	[0.000; 0.975]	0.1%
Narankhajid M (2018)	6	309	0.019	[0.007; 0.042]	0.7%
Karasartova D(2017)	0	17	0.000	[0.000; 0.195]	0.6%
Zhang F (2016)	28	249	0.112	[0.076; 0.158]	0.7%
Yu PF (2016)	3	165	0.018	[0.004; 0.052]	0.7%
Yang J (2016)	0	66	0.000	[0.000; 0.054]	0.7%
Liu L(2015)	0	179	0.000	[0.000; 0.020]	0.7%
Wang XR (2015)	25	110	0.227	[0.153; 0.317]	0.7%
Chen C (2015)	0	2	0.000	[0.000; 0.842]	0.2%
Hou J (2014)	0	9	0.000	[0.000; 0.336]	0.5%
Orkun O (2014)	0	25	0.000	[0.000; 0.137]	0.6%
Sun X (2013)	0	41	0.000	[0.000; 0.086]	0.6%
Niu QL (2010)	10	264	0.038	[0.018; 0.069]	0.7%
Huang HM (2006)	101	327	0.309	[0.259; 0.362]	0.7%
Chu CY (2006)	6	144	0.042	[0.015; 0.088]	0.7%
Hua MT (2000)	10	62	0.161	[0.080; 0.277]	0.7%
Tian W (1998)	0	13	0.000	[0.000; 0.247]	0.5%
Takada N (1994)	0	8	0.000	[0.000; 0.369]	0.5%
Du Y (1992)	0	18	0.000	[0.000; 0.185]	0.6%
Zhag QN (1991)	0	105	0.000	[0.000; 0.035]	0.7%
Random effects model		4321	0.029	[0.008; 0.062]	19.0%
Heterogeneity: $I^2 = 96\%$ , $\tau^2 = 0.0459$ , $p < 0.01$					
genus = Haemaphysalis					
Gao Y(2021)	1	275	0.004	[0.000; 0.020]	0.7%
Seo HJ(2021)	1	109	0.009	[0.000; 0.050]	0.7%
Wang Q(2021)	4	85	0.047	[0.013; 0.116]	0.7%
Seo MG(2020)	4	315	0.013	[0.003; 0.032]	0.7%
Kim SY(2020)	1	9	0.111	[0.003; 0.482]	0.5%
Nakayama S(2020)	1	531	0.002	[0.000; 0.010]	0.7%
Seto J(2020)	0	19	0.000	[0.000; 0.176]	0.6%
Orkun O(2020)	0	937	0.000	[0.000; 0.004]	0.7%
Kim CM(2019)	0	459	0.000	[0.000; 0.008]	0.7%
Pukhovskaya NM(2019)	21	530	0.040	[0.025; 0.060]	0.7%
Tang F (2018)	0	5	0.000	[0.000; 0.522]	0.4%
Ye NN (2018)	0	236	0.000	[0.000; 0.016]	0.7%
Zheng WQ (2018)	0	199	0.000	[0.000; 0.018]	0.7%
Narankhajid M (2018)	1	44	0.023	[0.001; 0.120]	0.6%
Jiang BG(2018)	1	36	0.028	[0.001; 0.145]	0.6%
Kuo CC (2017)	0	52	0.000	[0.000; 0.068]	0.7%
Karasartova D(2017)	0	218	0.000	[0.000; 0.017]	0.7%
Liu XY(2017)	0	500	0.000	[0.000; 0.007]	0.7%
Han H (2016)	0	74	0.000	[0.000; 0.049]	0.7%
Zhang F (2016)	69	369	0.187	[0.149; 0.231]	0.7%
Yang Y (2016)	0	178	0.000	[0.000; 0.021]	0.7%
Yang J (2016)	0	107	0.000	[0.000; 0.034]	0.7%
Yu PF (2016)	79	243	0.325	[0.267; 0.388]	0.7%
Han H (2015)	1	573	0.002	[0.000; 0.010]	0.7%
Chen C (2015)	1	34	0.029	[0.001; 0.153]	0.6%
Wang XR (2015)	28	117	0.239	[0.165; 0.327]	0.7%
Orkun O (2014)	4	102	0.039	[0.011; 0.097]	0.7%
Lee K(2014)	6	670	0.009	[0.003; 0.019]	0.7%
Hou J (2014)	11	237	0.046	[0.023; 0.082]	0.7%
Chen Z(2014)	0	298	0.000	[0.000; 0.012]	0.7%
Sun X (2013)	3	174	0.017	[0.004; 0.050]	0.7%
Wang W (2013)	0	139	0.000	[0.000; 0.026]	0.7%
Murase Yusuke (2012)	1	13	0.077	[0.002; 0.360]	0.5%
Yang Y (2011)	5	162	0.031	[0.010; 0.071]	0.7%
Ahantarig A(2011)	0	166	0.000	[0.000; 0.022]	0.7%
Niu QL (2010)	11	173	0.064	[0.032; 0.111]	0.7%
Wang CS (2009)	9	110	0.082	[0.038; 0.150]	0.7%
Chu CY(2008)	38	156	0.244	[0.179; 0.319]	0.7%
Chu CY (2006)	0	144	0.000	[0.000; 0.025]	0.7%
Mo YM (1994)	5	60	0.083	[0.028; 0.184]	0.7%
Takada N (1994)	1	81	0.012	[0.000; 0.067]	0.7%
Du Y (1992)	0	62	0.000	[0.000; 0.058]	0.7%
Zhag QN (1991)	0	45	0.000	[0.000; 0.079]	0.6%
Random effects model		9046	0.017	[0.007; 0.033]	28.5%
Heterogeneity: $I^2 = 95\%$ , $\tau^2 = 0.0241$ , $p < 0.01$					
genus = Hyalomma					
Ghafar A(2020)	14	216	0.065	[0.036; 0.106]	0.7%
Karasartova D(2017)	0	46	0.000	[0.000; 0.077]	0.6%
Orkun O (2014)	2	6	0.333	[0.043; 0.777]	0.4%
Random effects model		268	0.052	[0.000; 0.207]	1.8%
Heterogeneity: $I^2 = 86\%$ , $\tau^2 = 0.0345$ , $p < 0.01$					
genus = Ixodes					
Gao Y(2021)	7	264	0.027	[0.011; 0.054]	0.7%
Seo HJ(2021)	14	270	0.052	[0.029; 0.085]	0.7%
Wang Q(2021)	13	42	0.310	[0.176; 0.471]	0.6%
Seo MG(2020)	2	14	0.143	[0.018; 0.428]	0.5%
Kim SY(2020)	247	729	0.339	[0.304; 0.374]	0.7%
Nakayama S(2020)	1	8	0.125	[0.003; 0.527]	0.5%
Seto J(2020)	12	145	0.083	[0.043; 0.140]	0.7%
Orkun O(2020)	3	30	0.100	[0.021; 0.265]	0.6%
Kim CM(2019)	5	25	0.200	[0.068; 0.407]	0.6%
Rar V(2019)	742	1822	0.407	[0.385; 0.430]	0.7%
Pukhovskaya NM(2019)	582	1599	0.364	[0.340; 0.388]	0.7%
Ye NN (2018)	0	53	0.000	[0.000; 0.067]	0.7%
Khoo JJ (2018)	72	122	0.590	[0.497; 0.678]	0.7%
Yang Y (2018)	0	125	0.000	[0.000; 0.029]	0.7%
Tang F (2018)	85	293	0.290	[0.239; 0.346]	0.7%
Zheng WQ (2018)	3	21	0.143	[0.030; 0.363]	0.6%
Narankhajid M (2018)	2	7	0.286	[0.037; 0.710]	0.5%
Jiang BG(2018)	19	627	0.030	[0.018; 0.047]	0.7%
Pan YP (2017)	5	25	0.200	[0.068; 0.407]	0.6%
Kuo CC (2017)	5	27	0.185	[0.063; 0.381]	0.6%
Karasartova D(2017)	1	4	0.250	[0.006; 0.806]	0.4%
Yu PF (2016)	18	42	0.429	[0.277; 0.590]	0.6%
Han H (2016)	0	72	0.000	[0.000; 0.050]	0.7%
Yang J (2016)	0	84	0.000	[0.000; 0.043]	0.7%
Sakakibara K(2016)	1	248	0.004	[0.000; 0.022]	0.7%
Khasnatinov MA(2016)	90	445	0.202	[0.166; 0.243]	0.7%
Han H (2015)	11	70	0.157	[0.081; 0.264]	0.7%
Chen C (2015)	3	8			

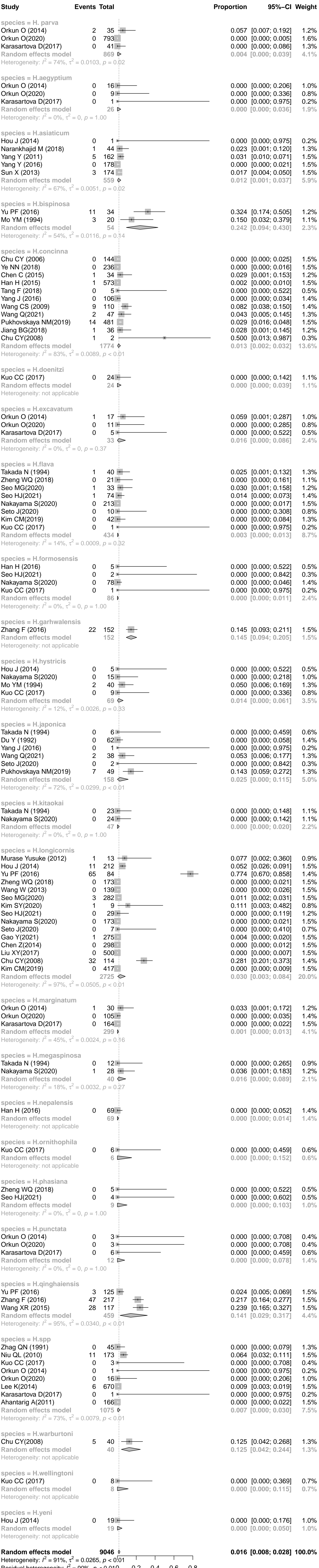


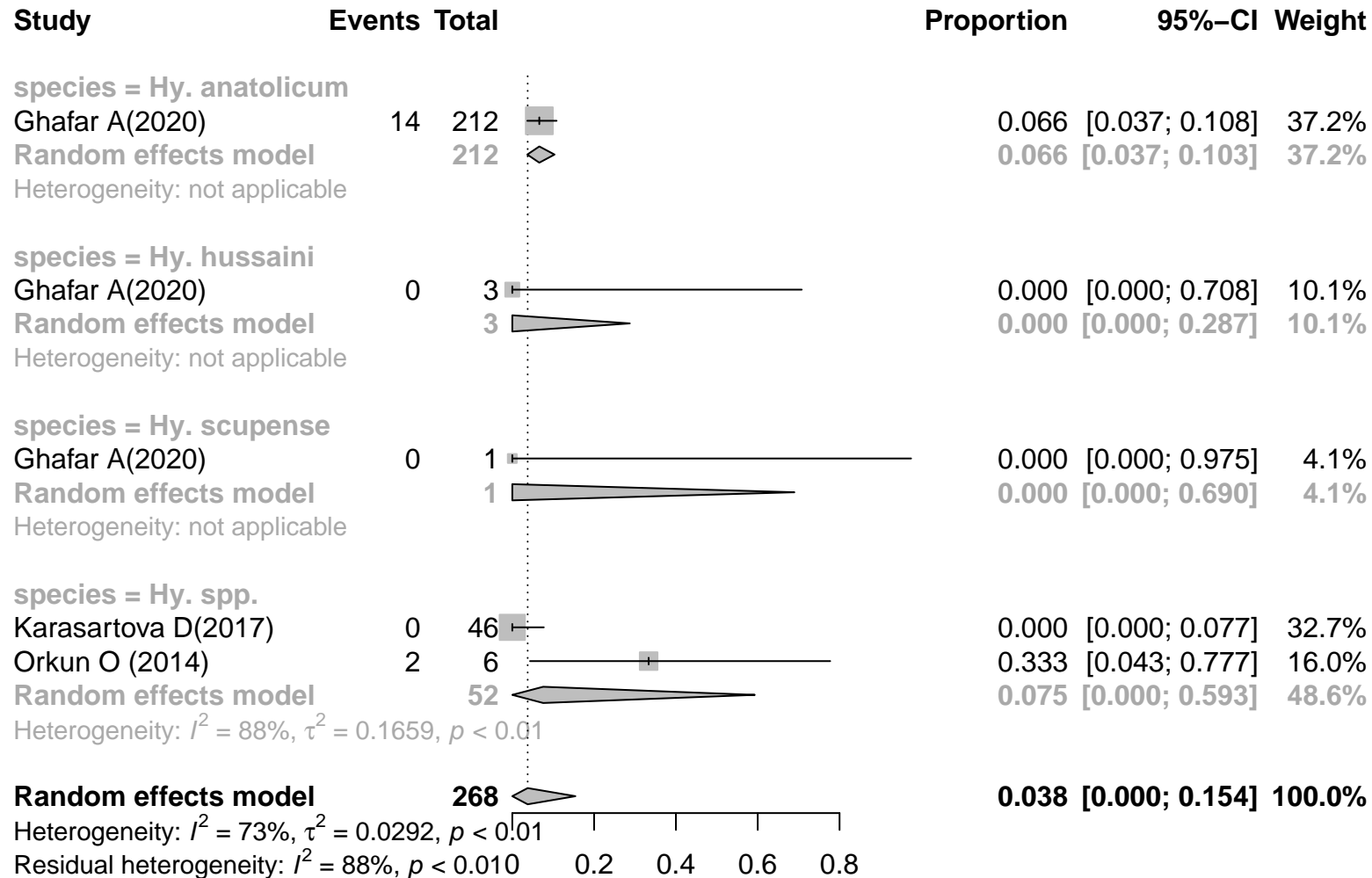


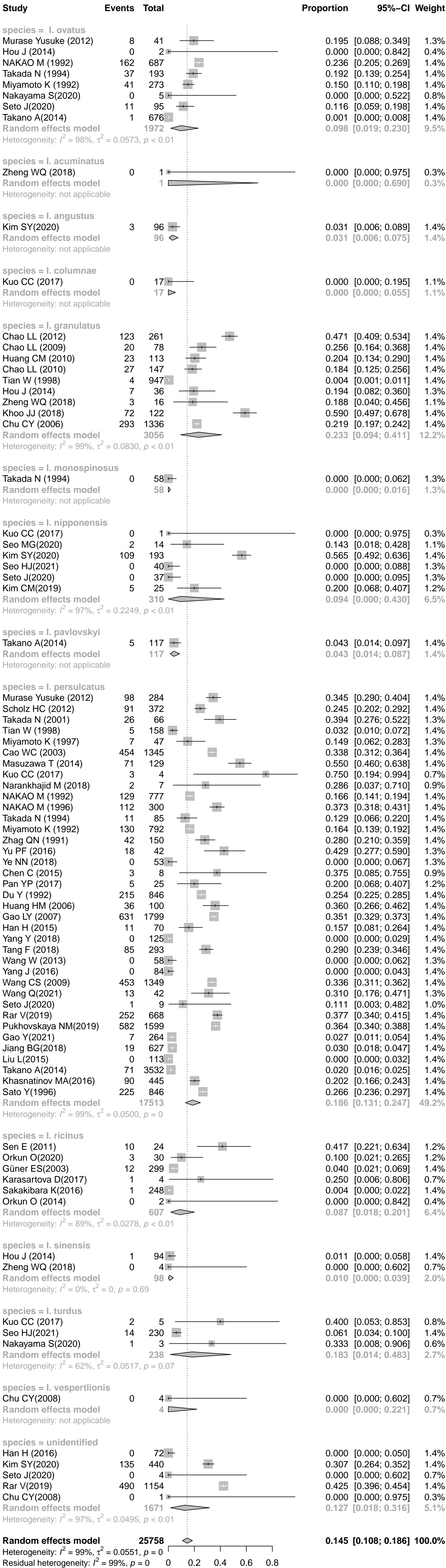


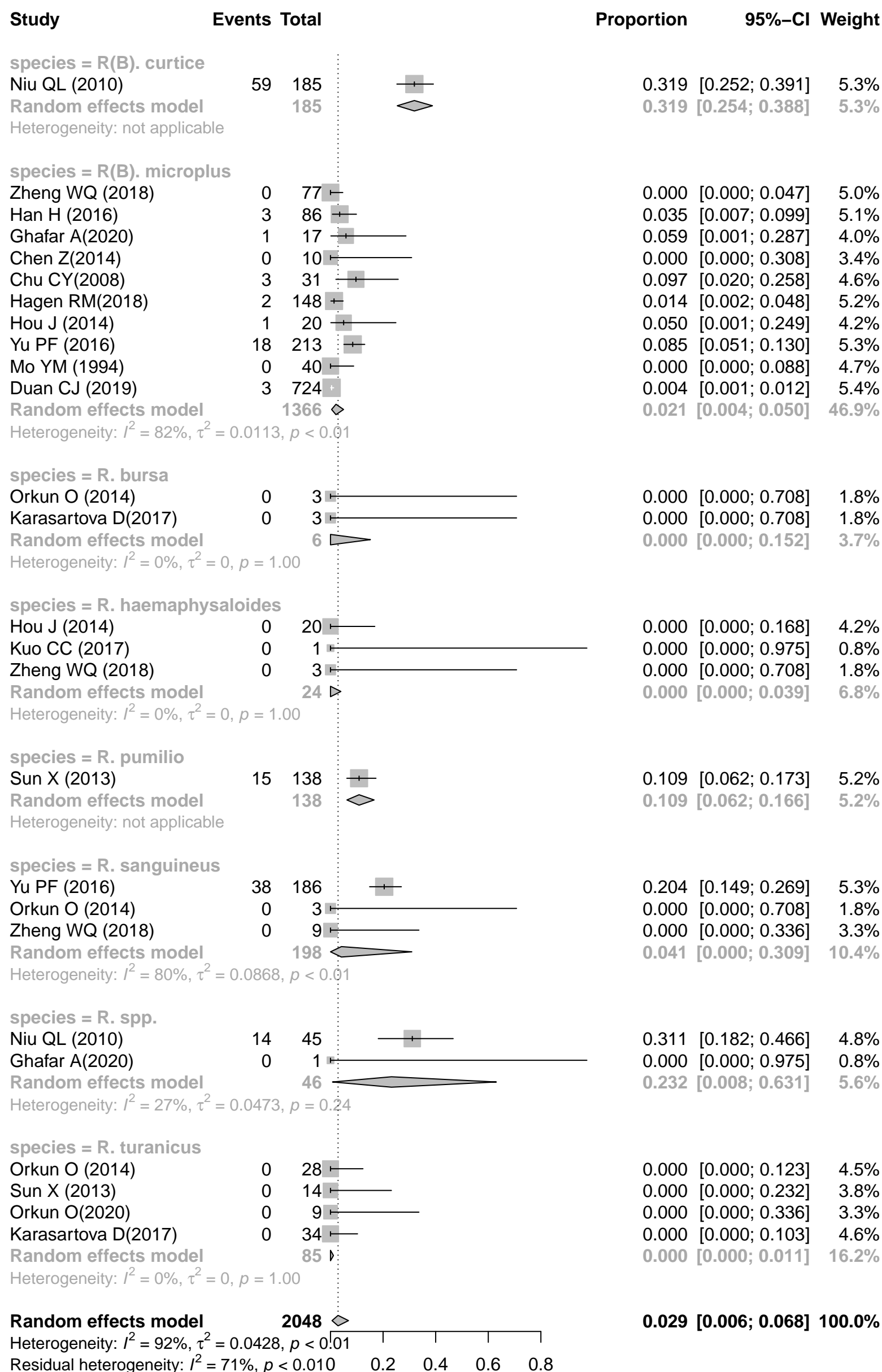












## Supplementary Material 6: Funnel plot

(A) Egg's plot of the Asia prevalence of *B. burgdorferi* s.l. in Ixodidae. (B) Funnel plot of the Asia prevalence of *B. burgdorferi* s.l. in Ixodidae.

