



Article Woven-Fiber Microfiltration (WFMF) and Ultraviolet Light Emitting Diodes (UV LEDs) for Treating Wastewater and Septic Tank Effluent

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Abstract: Decentralized wastewater treatment systems enable wastewater to be treated at the source for cleaner discharge into the environment, protecting public health while allowing for reuse for agricultural and other purposes. This study, conducted in Thailand, investigated a decentralized wastewater treatment system incorporating a physical and photochemical process. Domestic wastewater from a university campus and conventional septic tank effluent from a small community were filtered through a woven-fiber microfiltration (WFMF) membrane as pretreatment for ultraviolet (UV) disinfection. In domestic wastewater, WFMF reduced TSS (by 79.8%), turbidity (76.5%), COD (38.5%), and NO₃ (41.4%), meeting Thailand irrigation standards for every parameter except BOD. In septic tank effluent, it did not meet Thailand irrigation standards, but reduced TSS (by 77.9%), COD (37.6%), and TKN (13.5%). Bacteria (total coliform and Escherichia coli) and viruses (MS2 bacteriophage) passing through the membrane were disinfected by flow-through UV reactors containing either a low-pressure mercury lamp or light-emitting diodes (LEDs) emitting an average peak wavelength of 276 nm. Despite challenging and variable water quality conditions (2% < UVT < 88%), disinfection was predictable across water types and flow rates for both UV sources using combined variable modeling, which enabled us to estimate log inactivation of other microorganisms. Following UV disinfection, wastewater quality met the WHO standards for unrestricted irrigation.

Keywords: decentralized; domestic wastewater; septic tank effluent; woven membrane; UV validation; combined variable modelling; LMIC; MS2 bacteriophage

1. Introduction

By 2050, global water demand is projected to be 20–30% higher than current levels, given both population growth and socioeconomic development [1]. Agriculture accounts for the highest percentage of water withdrawals worldwide, contributing to roughly 70–85% of freshwater withdrawals in some developing economies [2]. This trend is predicted to continue for decades. Easing the burden on our freshwater resources requires increasing the use of reclaimed water, particularly for agriculture and irrigation; however, reclaimed water poses health risks to users [3].



Citation: Beck, S.E.; Suwan, P.; Rathnayeke, T.; Nguyen, T.M.H.; Huanambal-Sovero, V.A.; Boonyapalanant, B.; Hull, N.M.; Koottatep, T. Woven-Fiber Microfiltration (WFMF) and Ultraviolet Light Emitting Diodes (UV LEDs) for Treating Wastewater and Septic Tank Effluent. *Water* **2021**, *13*, 1564. https://doi.org/10.3390/ w13111564

Academic Editors: Simona Consoli and Giuseppe Luigi Cirelli

Received: 1 April 2021 Accepted: 27 May 2021 Published: 31 May 2021

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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). The WHO/UNICEF Joint Monitoring Programme characterizes sanitation facilities into five categories: (1) open defecation, (2) unimproved services such as pit latrines, (3) limited or shared facilities, (4) "basic" sanitation services, and (5) safely managed facilities where the excreta are treated and safely disposed of [4]. Basic sanitation facilities include flush and pour-flush toilets, septic tanks, and improved pit latrines where the waste is collected but not safely managed beyond the toilet [4]. An estimated 29% of the global population (2.1 billion people) fall into this category [4]; of them, many live in the Asia and Pacific region, including Thailand. Thailand is one of the few countries in Southeast Asia with near complete coverage of basic sanitation nationwide; however, the waste is not safely managed, leading to a high prevalence of disease [5]. Only approximately 21% of human waste is treated at a municipal facility [4–6]. The remainder is often discharged into the environment or nearby neighborhoods, flowing directly into receiving waters, contaminating local water supplies and affecting public and environmental health.

Wastewater is one of the primary point source contaminants polluting freshwater and marine environments as well as shallow groundwater sources. Contaminated surface water contributes to disease burden and, as a result, adversely impacts the economy [5,7]. Public and private wells are susceptible to contamination by enteric pathogens, including viruses, bacteria, and protozoa [8], and septic tanks have been identified as a source of that contamination [9–11]. Enteric viruses have been detected in wells as far as 32 m from the septic tank source of fecal pollution, thus, contamination poses a significant threat in areas with on-site septic systems without appropriate treatment [12]. Thailand has recognized this serious environmental problem and identified a need for wastewater collection and treatment while encouraging decentralized treatment for households and buildings [6]. Decentralized wastewater treatment systems enable wastewater to be treated at the source for cleaner discharge into the environment, while allowing for potential reuse for agricultural and other purposes.

Membrane technology plays a substantial role in decentralized wastewater treatment and reuse [13,14]; however, membranes are relatively costly and require well-trained operation, making application at a household level challenging. Woven-fiber microfiltration (WFMF) membranes have recently been introduced as a cost-effective, robust, and easy-tooperate filtration method [15]. WFMF textiles have been successful for point-of-use water treatment when combined with disinfectants such as sodium hypochlorite, bromochlor, and silver nanoparticles [16,17]. In addition, fouling removal for WFMF, which is one of the most important maintenance requirements in membrane operation, is possible by removing a cake layer without required chemical addition [17]. Thus, WFMF membranes have proven effective in less populated areas and in decentralized systems [18,19].

Ultraviolet (UV) light is frequently used for pathogen inactivation in water and wastewater treatment [20–22]. UV light effectively inactivates viruses, bacteria, and cysts by penetrating cell walls and damaging DNA or RNA without chemical addition. Traditional UV lamps are low-cost and accessible in developing economies, but contain toxic mercury vapor. UV LEDs are more expensive, but also mercury-free. They are significantly smaller, lighter, and more durable than traditional lamps, and require less power. In the Middle East, traditional UV lamps have been used for inactivating enteric pathogens remaining after treatment by constructed wetlands [23]. In the Mediterranean, UV LEDs were used to inactivate fecal bioindicators present after centralized wastewater treatment using activated sludge [24]. In our previous work in Southeast Asia, we investigated a household or building-scale UV LED reactor for disinfecting pretreated domestic wastewater for agriculture reuse [25]. In that study, the wastewater was pretreated with conventional slow-sand filtration and inclined settling, which was low-cost but high in footprint. In this study, we evaluated a system with a smaller footprint and shorter operating time.

This study evaluated a cost-effective, user-friendly, and relatively fast treatment process involving a woven-fiber microfiltration (WFMF) membrane to filter domestic wastewater followed by UV disinfection to disinfect the permeate. With an effective pore size of 1–3 μ m [26], the WFMF was capable of removing *Ascaris lumbricoides* eggs (50 mm)

and *Giardia* cysts (10 μ m), whereas bacteria (1–2 μ m), viruses, and *Cryptosporidum* oocysts (3 μ m), which are small enough to pass through the filter pores, were inactivated by exposure to UV light. Standard mercury lamps and UV LEDs were investigated. This work is targeted at developing and evaluating a water reuse process for addressing water scarcity and contamination in low to middle income areas. The objective was to determine if disinfected permeate could meet the Thailand standards for irrigation and WHO guidelines for building effluent water quality [27–29].

2. Materials and Methods

2.1. Wastewater Sources

This research evaluated a combined WFMF and UV LED system with two wastewater sources: domestic wastewater and conventional septic tank effluent. Domestic wastewater was collected from a collection tank at the Asian Institute of Technology (AIT) campus (Khlong Luang, Pathum Thani Province, Thailand) as in Nguyen et al. [25]. It was produced by a residential population of 2000–2500 along with a transient commercial and academic population, combining blackwater from toilets and greywater from kitchens and sinks. Conventional septic tank effluent was pumped from a septic tank serving a toilet used by 30–50 people/day (Phra Pradaeng, Samut Prakan Province, Thailand). For both wastewater sources, 200 L were collected and analyzed weekly for at least four weeks.

2.2. Wastewater Quality Characterization

Wastewater quality parameters were monitored weekly, following the Standard Methods for the Examination of Water and Wastewater [30]. These include pH, temperature, electrical conductivity, total suspended solids (TSS), biological oxygen demand (BOD), chemical oxygen demand (COD), total Kjehdahl nitrogen (TKN), ammonia nitrogen (NH₃-N), nitrate nitrogen (NO₃-N), and total phosphorous (TP). Microbiological analysis also followed the Standard Methods for detecting fecal indicator bacteria, *Escherichia coli*, and total coliforms, which were measured as mean probable number (MPN) per 100 mL [30]. *Ascaris lumbricoides* ova were selected as an indicator of helminth eggs [31]. Regarding physicochemical parameters, removal efficiency (R, %) was calculated as R = $100 \times (C_{in} - C_{out})/C_{in}$, where C_{in} and C_{out} represent the inlet and outlet concentrations to the WFMF system, respectively, which were collected at the same time. UV absorbance (UVA, cm⁻¹) was measured using a Hitachi U-2900 spectrophotometer (Tokyo, Japan). UV transmittance (UVT, %) for a given wavelength, λ , was calculated as $UVT_{\lambda} = 100 \times 10^{-a(\lambda)}$, where $a(\lambda)$ is the UVA at each wavelength.

2.3. Woven-Fiber Microfiltration System

The wastewater effluents were filtered through a woven-fiber microfiltration (WFMF) membrane system as a physical barrier to remove particles and suspended solids prior to photochemical disinfection with ultraviolet (UV) light. The polyester woven-fiber material was supplied by Dr. Lingam Pillay from Stellenbosch University. The material has a nominal pore size of $1-3 \ \mu m$ [26].

The flat sheet WFMF membranes were fabricated in the environmental engineering laboratory at Asian Institute of Technology, Thailand. The WFMF membrane module used in this study (Figure 1) was designed with five double-sided flat sheet woven-fiber textile modules (21.0×29.7 cm), corresponding to a total surface area of 1 m². Each membrane module contained a 3-layer structure of woven-fiber membranes and PVC spacer; the layers and the permeate flow collection channel were joined together with adhesive to the PVC frame. The membrane tank was fabricated with PVC materials to have a conical shape to help settle the accumulated sludge and remove solids. Total tank volume was 130 L, with a sludge cone volume of 23.5 L.

The system was operated in batch mode, in dead-end, outside-in configuration with the submerged membrane module operated by negative pressure as the driving force. The electrical control system was designed to have an automated operation. Suction pressure was created by changing the speed of a rotary-style peristaltic suction pump (Model 77200-60, Master Flex, Wertheim, Germany), shown as P₂ in Figure 1. A pressure transducer (Model 579225-010, Trafag, Bubikon, Switzerland) and data logger (Model-EL-USB-4, Lascar Electronics, Wiltshire, UK) were used to record pressure data. The peristaltic suction pump was connected to a timer (Omron twin timer- H3CR-F8) and operated with intermittent running intervals (OFF 1 min, ON 6 min). A level controller system (Omron 61F-G) was introduced to the membrane tank to maintain a consistent water level inside the tank by connecting with the feed pump (P₁).



Figure 1. Schematic of the woven-fiber microfiltration (WFMF) system.

The system was operated continuously at $8 L/m^2/h$ with a goal to reduce the turbidity below 5 NTU (nephelometric turbidity units) and to reduce the total suspended solids below 30 mg/L as pretreatment for UV disinfection [32–34]. The permeate water from the WFMF system was introduced to the UV reactors for disinfection.

Although cleaning was not necessary during these WFMF experiments, the membrane cleaning process is typically conducted when transmembrane pressure (TMP) reaches -60 kPa. The process includes a physical, spray-brush method to remove colloids, followed by a chemical cleaning process for organic and inorganic fouling removal. Membranes are immersed in a solution of 0.5 M NaOH and 0.03% NaOCl for 8 h to remove organic fouling constituents. Once the base cleaning is carried out, the membrane is immersed in a solution of 0.01 % of HCl for 8 h to remove inorganic constituents prior to tap water cleaning and use.

2.4. Flow-Through UV Reactors

Approximately 25–30 L of WFMF permeate was collected for disinfection with one of two flow-through UV disinfection systems, a low-pressure (LP) UV lamp or UV LEDs, both of which were constructed in-house at the Asian Institute of Technology. The LP UV reactor (Figure 2) contained a 20 W mercury vapor lamp, emitting monochromatic UV irradiation at 254 nm (TOKIVA G20T8 by CCS SUCCESSPRODUCT Company Limited, Thailand), which was encased inside a welded stainless-steel casing with a small gasket. The reactor was 58.8 cm in length with an inner diameter of 4.6 cm surrounding the 2.5 cm diameter lamp for a maximum path length within the reactor of 1.05 cm. The lamp was connected to a ballast with voltage adapter as shown (Figure 2), supplying a constant voltage of 58 V at 0.36 mA. Water flowed outside the light source with a volume of 0.688 L up to the reactor's maximum flow rate of 1.8 L/min.

The UV LED reactor (Figure 3) contained 4 UV LED arrays emitting polychromatic irradiation near 280 nm (TDS-UV280J16-C-A/H, TDS LIGHT Company Limited, China). Each array contained 16 diodes arranged in a 4×4 pattern and measuring 14×14 mm in size. The arrays were mounted on four sides, forming a square outside of a cylindrical quartz sleeve (1.8 cm diameter \times 20 cm length). In contrast to the LP UV system, water flowing inside the sleeve (0.051 L) was irradiated by LEDs on the outside with a path length of 0.9 cm at flow rates up to 0.1 L/min. Together, the UV LED emission spectra (Figure 3) had an average peak of 276.2 nm, a weighted average wavelength of 278.1 nm, and an average full width at half maximum (FWHM) of 10.6 nm. Emission spectra were measured at the National Institute of Metrology Thailand (NIMT) using a diode array spectroradiometer (CAS140CT-154) with non-optical density filter and cosine receptor probe (EOP-146, Instrument Systems, Germany). The arrays were wired in parallel at a current of 80 mA and a voltage of 21–24 DC.

The two UV light sources in this study were not meant for direct comparison, rather this was the state of their development at the time of experimentation (2016). Operating in the germicidal UV-B and UV-C range, both light sources induce damage to RNA and DNA through direct photolysis, causing microorganism inactivation [35]. Although fouling and scaling were not an issue during these experiments, fouling and scaling of UV systems is reversible through citric acid circulation [25].



Figure 2. Photo and schematic of the flow-through low-pressure UV reactor constructed in-house in Thailand.

2.5. UV Fluence Determination

UV fluence (mJ/cm²), or dose, is calculated as the product of irradiance (mW/cm²) and time (s); therefore, the doses were changed by varying the exposure time for batch experiments, or by varying flow rate for flow-through experiments. The UV fluences applied by the flow-through reactors were calculated using biodosimetry with MS2 bacteriophage as described previously [25,35,36]. Bacteriophages such as MS2 are common indicators of

fecal contamination [37]. Virus inactivation studies often use MS2 as a surrogate for enteric viruses, RNA viruses, and other pathogens to allow a comparison between conventional and emerging water and wastewater treatment technologies [38–40]. MS2 has been used in water reclamation studies and is frequently used for validating small-scale and large-scale UV reactors [36,41].

Although the wastewater and septic tank effluent contained background concentrations of 0 to $10^{2.2}$ PFU/mL MS2, additional bacteriophage was spiked into the WFMF filtration permeates at 10^{6} – 10^{8} PFU/mL as a surrogate microorganism for the disinfection study. The MS2 bacteriophage (ATCC 15597-B1, American Type Culture Collection, Manassas, VA, USA) was propagated and enumerated with *E. coli* Famp (ATCC 700891) as the bacterial host, following the USEPA double agar layer (DAL) method [25,42] with experimental and analytical duplicates. Log inactivation of MS2 was calculated as the ratio of log₁₀ concentration of MS2 in the water sample before and after UV irradiation.



Figure 3. Photo, schematic, and emission spectra of the flow-through UV LED reactor constructed in Thailand.

For both flow-through reactors, the UV-induced log reduction of MS2 was measured at each flow rate for each water matrix tested. In parallel, quasi-collimated beam experiments were conducted with a batch reactor to determine the dose response of the microorganism in each wastewater matrix. Given the log reduction of MS2, the dose response curves were then used as a standard curve to back-calculate the reduction equivalent dose (RED) delivered by the flow-through reactors at each flow rate. Multiple flow rates were used for each experiment, corresponding to multiple UV doses. To evaluate the disinfection performance of the UV LEDs and the LP UV lamp for the flow-through reactors, the same wastewaters and septic tank samples spiked with MS2 were used for the batch tests in continuously stirred 5 mL samples (6 mm depth). Batch experiments with the UV LED batch reactor were conducted in parallel with experiments involving the flow-through UV LED reactor; batch experiments with the LP UV reactor were conducted in parallel with experiments using the flow-through LP UV reactor.

UV LED batch reactor tests incorporated the batch reactor described previously [25] with a peak wavelength emission at 277 nm and a full width at half maximum (FWHM) of 10 nm. As shown in Figure 3, the spectral emission of the batch reactor was exactly aligned with two of the arrays in the flow-through reactor and deviated from the other two arrays by approximately 1 nm and 3 nm at peak wavelength. This UV LED batch reactor was fully characterized for our previous study; the average irradiance across the surface of the water sample was determined using biodosimetry with MS2 as described previously [25]. Briefly, the published dose response of MS2 (suspended in phosphate buffered saline, PBS) to UV LEDs with the same wavelength emission (peak = 276.6 nm; FWHM = 9.8 nm, inactivation rate constant $k_{\rm D} = 0.052 \, {\rm cm}^2/{\rm mJ}$) was used to back-calculate the average irradiance across the surface of the water sample [25,43]. The average UV LED irradiance across the surface of each water sample measured through biodosimetry was 0.208 mW/cm² \pm 0.013 (sample size, *n* = 3). This value was then confirmed by off-site radiometry at the National Institute of Metrology Thailand (0.212 mW/cm²) after adjusting for the Petri factor (0.876), reflection factor, sensor factor, and divergence factor [36]. Petri factor was sufficient over a coefficient of variation in this scenario because the irradiance peaked at the center of the dish (Figure S1) [44]. The value for average irradiance across the surface of the water sample was then used together with the water factor and the DNA absorbance spectrum to calculate the average germicidal irradiance throughout each water sample, as is conventional for polychromatic UV systems [45].

Similarly, for the low-pressure (LP) UV lamp, the published dose response of MS2 (suspended in PBS) to LP UV ($k_D = 0.052 \text{ cm}^2/\text{mJ}$) was used to back-calculate the average irradiance from the LP UV lamp across the surface of the water sample [21]. The average LP UV irradiance across the surface of the water sample measured through biodosimetry was 2.437 mW/cm² ± 0.044 (n = 3). This was confirmed by off-site radiometry at NIMT (2.442 mW/cm²). The average irradiance across the surface of the water sample was then used with the water factor to calculate the average irradiance throughout each sample.

The flow-through UV LED reactor was also validated by varying UV transmission (UVT) using coffee as recommended previously [36,41,46]. MS2 experiments therefore involved MS2 suspended in PBS, wastewater or septic tank effluent, or coffee to vary the UVT.

2.6. Statistical Analysis

All experiments were conducted multiple times with analytical duplicates or triplicates. Paired *t*-tests were conducted for evaluating the statistically significant differences between influent and effluent data of AIT wastewater and conventional septic tank effluent treated by WFMF.

2.7. Combined Variable Approach to RED and Log Inactivation Modelling

Reduction equivalent dose (RED) and log inactivation were modeled for a single challenge microorganism based on the combined variable approach presented by Wright et al. [47] and Hull et al. [48] and described in Equation (1):

$$RED \text{ or } \log I = 10^{a} \times UVA^{b} \times \left(\frac{S/S_{0}}{Q}\right)^{c+d \times UVA+e \times UVA^{2}}$$
(1)

where UVA is the UV absorbance at 254 or 276 nm or the UVA weighted by relative lamp emission (RLE), as was the case with the polychromatic UV LEDs. S/S_0 is the ratio of measured UV intensity (S) over UV intensity for new lamps (S_0), which was assumed to equal 1 in these experiments. Q is the volumetric flow rate (L/min). Empirical coefficients (a–e) were determined in R v. 4.0.3 and RStudio v. 1.4.1103 by minimizing the sum of the squares of the difference between the measured and predicted values using the Gauss–Newton method for nonlinear least squares regression. The empirical coefficients (a–e) were determined to be statistically significant based on their *p*-value (p < 0.05). Coefficients that were not statistically significant were removed (i.e., set equal to zero) from the model in

a stepwise fashion, starting with the coefficient with the highest *p*-value [47]. The final log inactivation and RED models included only statistically significant coefficients. Following the findings from Hull et al. [48], log inactivation by UV LEDs was modeled as a function of RLE absorbance and flow rate. LP UV log inactivation was modeled as a function of UVA₂₅₄ and flow rate.

3. Results and Discussion

3.1. WFMF Membrane System Performance

Table 1 presents the quality of the wastewater and septic tank effluent used in these experiments before and after the WFMF membrane filtration pretreatment. WFMF removal performance for various parameters in the AIT campus domestic wastewater and septic effluent are given in Figure 4. Compared to the literature, for a typical composition of low-strength untreated domestic wastewater in the United States, AIT domestic wastewater, which comprises blackwater and greywater, is more dilute (by approximately half) for solids, BOD, and COD, but has almost double the nutrient content [49]. However, these values are in line with other wastewater systems in Southeast Asia [50]. Parameters from the septic tank effluent from Phra Pradaeng, including COD, BOD, and TKN, are also on par for conventional septic tanks in the region [50,51].

Table 1. Characteristics of AIT wastewater and conventional septic tank effluent before (raw) and after the WFMF pretreatment (permeate). Data presented as average \pm SD with sample size *n*. Parameters marked with a "-" indicate data that was not collected or lost.

Parameters (Units)	Analysis Method	Raw Wastewater	Wastewater WFMF Permeate	Raw Septic Tank Effluent	Septic Tank WFMF Permeate
рН	4500B-H+	7.29 ± 0.31 n = 15	7.46 ± 0.48 n = 15	7.32 ± 0.39 n = 14	-
Temp (°C)	-	29.0 ± 4.1 n = 15	29.3 ± 3.7 n = 15	29.4 ± 2.7 n = 14	-
Conductivity (µS/cm ²)	2510B	724 ± 49 $n = 13$	728 ± 54 $n = 13$	2795 ± 236 n = 14	-
TSS (mg/L)	2540D	65.7 ± 67 $n = 15$	8.6 ± 4 $n = 15$	128.8 ± 33 n = 21	56.5 ± 31 $n = 4$
Turbidity (NTU)	Turbidity (NTU) 2130B		5.9 ± 6.0 n = 14	-	-
BOD (mg/L) 5210B		51.9 ± 27.1 n = 7	31.3 ± 26.9 n = 7	302.8 ± 124 $n = 9$	-
COD (mg/L) 5220C		96.2 ± 41 $n = 16$	58.3 ± 32 $n = 16$	510.6 ± 167 n = 19	242.8 ± 77 $n = 4$
TKN (mg/L)	4500-NorgC	49.5 ± 32 $n = 8$	30.0 ± 3 $n = 8$	264.8 ± 20 n = 20	240.6 ± 8 $n = 2$
NH ₃ -N (mg/L) 4500-NH3 C		24.9 ± 8.7 $n = 9$	24.6 ± 7.9 $n = 9$	257.0 ± 53 n = 20	191.2 ± 63 $n = 4$
NO ₃ -N (mg/L) 8039-HR		3.9 ± 2.8 n = 7	$\begin{array}{c} 1.7 \pm 1.4 \\ n = 7 \end{array}$	5.4 ± 3.2 $n = 2$	3.4 ± 2.4 $n = 2$
TP (mg/L)	TP (mg/L) 4500P		18.0 ± 19 $n = 9$	368.7 ± 105 n = 2	320.8 ± 83 n = 2
A. lumbricoides ova (eggs/L)	[31]	$0 \\ n = 5$	$\begin{array}{c} 0\\ n=5 \end{array}$	-	-

Parameters (Units)	Analysis Method	Raw Wastewater	Wastewater WFMF Permeate	Raw Septic Tank Effluent	Septic Tank WFMF Permeate
Total coliforms (MPN/100 mL)	9221C	$1.0 \times 10^{6} \pm 2.6 \times 10^{6} n = 9$	$3.3 imes 10^5 \pm 5.1 imes 10^5 n = 9$	$1.3 \times 10^{7} \pm 2.7 \times 10^{7} n = 17$	$1.6 imes 10^{6} \ \pm 1.6 imes 10^{6} \ n=2$
<i>E. coli</i> (MPN/100 mL)	9221F	$3.2 \times 10^{5} \pm 7.0 \times 10^{5} $ n = 9	$1.5 \times 10^{5} \pm 1.5 \times 10^{5} $ n = 9	$2.8 imes 10^{6} \ \pm 2.7 imes 10^{6} \ n = 17$	$3.5 \times 10^{5} \pm 5.5 \times 10^{3} n = 2$
MS2 coliphage (PFU/mL)	USEPA 1602	16 ± 44 $n = 11$	7 ± 17 $n = 11$	19 ± 13 $n = 4$	19 ± 25 $n = 4$
UVT ₂₅₄ (%)		33.1 ± 15 $n = 14$	52.5 ± 14 $n = 18$	3.2 ± 3.1 $n = 3$	3.5 ± 1.7 $n = 4$
UVT ₂₈₀ (%)		44.9 ± 12 $n = 3$	57.3 ± 17 $n = 8$	6.0 ± 5.1 $n = 3$	6.1 ± 2.9 $n = 4$

Table 1. Cont.



Figure 4. Box plots of WFMF removal efficiencies for treatment of AIT domestic wastewater and conventional septic tank effluent. Removal calculations included only influent and effluent data collected on the same day. Boxes and whiskers represent median and minimum to maximum values as well as 25th–75th percentiles. **** p < 0.0001, *** p < 0.001, ** p < 0.01, * $p \le 0.05$, no asterisk means no significant difference between influent and effluent data.

3.1.1. Domestic Wastewater

Overall, the WFMF performed well with particle removal and acted as an efficient pretreatment unit for UV treatment of domestic wastewater, as shown in Table 2. Its primary drawback was not meeting irrigation guidelines for BOD set by the Thai Pollution Control Department [27]. The WFMF process did not cause a significant difference in pH, temperature, or conductivity. The system, which had a pore size of 1–3 µm, separated the total suspended solid fraction, preventing most of the suspended solids content from passing through the membrane, resulting in a clearer permeate with fewer particles. For domestic wastewater, the WFMF reduced total suspended solid particles by 79.8% (\pm 13.9). This agreed well with a previous study on textile filters for wastewater treatment, which showed a suspended solids removal rate of 71% [52]. For domestic wastewater, the WFMF system generated product water quality with an average TSS of 8.6 mg/L, which accommodates the effluent quality limit of 30 mg/L for irrigation systems in Thailand (Table 2, [27,28]). For all 15 samples tested, the TSS limits of product water fell below 30 mg/L, which was also the suggested guideline for water reuse for processed food or non-food crops suggested by the USEPA (Table 2, [53]). Suspended solids removal was one of the primary objectives of the WFMF as pretreatment for UV disinfection and potential reuse applications.

	This	Study	Guidelines		
Parameters (Units)	Wastewater WFMF Effluent	Septic Tank WFMF Effluent	Thailand Irrigation Standard ¹	WHO Unrestricted Irrigation ²	USEPA ³ Processed Food or Non-Food Crops
pH	7.5	-	6.5–8.5		6.0–9.0
Conductivity (µmol/cm)	700	-	≤2000		
Turbidity (NTU)	5.9	_	_		≤2
TSS (mg/L)	8.6	56.5	≤ 30		≤30
BOD (mg/L)	31.3	-	≤ 20		≤ 30
COD (mg/L)	58.3	242.8	≤ 100		
TKN (mg/L)	30.0	240.6	≤35		
TP (mg/L)	18.0	320.8	-		
FC (CFU/100 mL)			-	≤ 1000	≤200
Total coliforms (MPN/100 mL)	1.0×10^{6}	1.1×10^{6}			
Helminth eggs (eggs/L)	0			≤1	

¹ [27,28]; ² [29]; ³ [53].

As solid particles were retained by the WFMF system, the turbidity of the domestic wastewater was reduced by an average of 76.5% (\pm 26.9). Turbidity can have a significant effect on light-based disinfection; therefore, removal is necessary for UV pretreatment. Domestic wastewater filtered through the WFMF reached turbidities as low as 1.6 NTU with an average of 5.9 NTU. Although this value was higher than our goal of 5 NTU, it was still within the range of turbidities for UV inactivation of wastewater [32,54]. Similar studies of WFMF treatment of contaminated surface water reached turbidities of 1.0 NTU [17,54].

For domestic wastewater, the WFMF reduced COD and BOD levels by an average of $38.5\% (\pm 17.3)$ and $47.8\% (\pm 22.6)$, respectively, through the filtration process. Considering TSS removal efficiencies, showing that solids were mostly retained in the membrane, it was presumed that COD transferring through was predominantly soluble. The product water contained an average COD of 58.3 mg/L, which falls under the 100 mg/L limit required by Thailand for irrigation (Table 2) [27,28]. BOD removal, however, did not meet the irrigation standards, with an average permeate quality of 31.3 mg/L, which was higher than the required Thailand irrigation and USEPA processed food crop limits of 20 mg/L and 30 mg/L, respectively (Table 2). This highlights the need for additional biological treatment of the wastewater.

Total Kjeldahl nitrogen (TKN) was not reduced significantly; however, it maintained a permeate concentration of 30.0 mg/L, which meets the Thailand effluent quality standards of 35 mg/L (Table 2). Ammonia nitrogen and total phosphorus were also not reduced significantly; however, nitrate nitrogen was reduced by an average of 41.4% (\pm 40.5). Previous studies involving WFMF membrane bioreactors showed statistically significant reduction of ammonia nitrogen, nitrite nitrogen, phosphate, and other nutrients; however, those studies involved biological membrane growth in which the biomass performed a nitrification step, whereas the WFMF in this study was primarily a physical removal process [26,48]. UVT₂₅₄ improved from an average of 33.1% (\pm 15) to an average of 52.5% (\pm 14) after WFMF filtration. Similarly, UVT₂₈₀ improved from an average of 44.9% (\pm 12) to 57.3% (\pm 17). UVT spectra of treated permeates are given in Figure 5.

From a microbiological perspective, the WFMF was not effective at removing pathogens or indicator organisms, hence the need for UV disinfection. *Ascaris lumbricoides* ova, which measure 40–75 microns in size [55] would have been filtered out by the 1–3 micron pore size; however, they were not detected in the WFMF influent due to prior settling. The WFMF did not cause a significant reduction in total coliforms or *E. coli*, which was surprising compared to two previous studies where removal of *E. coli* from natural or synthetic surface water for noncoated woven-fiber membranes was found to be 84–99.8% [16,17]. Those studies had higher turbidity in the influent, which could have caused a thicker cake layer formation, improving size exclusion and reducing bacteria.

MS2 bacteriophage was only present in small amounts in the influent, and its removal was also not statistically significant. A study on gravity-driven membranes indicated that biofilm growth on the membrane would have contributed an extra 2.0+ log removal of MS2 [56].

3.1.2. Septic Tank Effluent

As a pretreatment for UV, the WFMF also performed well for particle removal from the septic tank effluent, but the permeate wastewater quality did not meet the Thailand irrigation standard for any of the parameters (Table 2). When analyzing the WFMF removal performance for treating septic tank effluent, TSS was reduced by an average of 77.9% (\pm 13.3) to an average concentration of 56.5 mg/L, above the irrigation standard requirement of 30 mg/L (Table 2). The WFMF treatment achieved a COD removal of 37.6% (\pm 12.8) in the septic tank effluent to a concentration of 242.8 mg/L, above the required 100 mg/L limit. These results are similar to a study that characterized woven-fiber flat sheet membrane fouling in a membrane-based septic tank and resulted in 50–60% TSS and 50–65% COD removal [19].

Total Kjeldahl nitrogen (TKN) was reduced by 13.5% (\pm 4.0) to 240.6 mg/L; however, this was still well above the Thailand effluent quality standards of 35 mg/L (Table 2). This treatment process was operated as a physical process and not a biological one; therefore, high reduction in nutrients was not expected. Ammonia nitrogen, nitrate nitrogen, and total phosphorus all showed reductions that were not statistically significant. For these parameters, the low sample sizes may have contributed to the insignificance. The primary goal of the WFMF was to remove suspended solids and improve the UVT for enhanced disinfection. Nevertheless, UVT was improved more for domestic wastewater; and was not improved with statistical significance for septic tank effluent. It is believed that this was due to dissolved humic acids present in the septic effluent, which were too small to be removed by the WFMF. UVT spectra of the treated permeates are given in Figure 5.

The WFMF system achieved 85.8% reduction of total coliform and 90.6% reduction of *E. coli* rates with the septic tank effluent water prior to the UV disinfection process. However, with a small sample size, these values were not statistically significant. As with domestic wastewater, MS2 bacteriophage was present in small amounts in the influent, and its removal was also not statistically significant.





3.2. UV Disinfection System Performance

The primary purpose of the LP UV and UV LED reactors was to disinfect bacteria, viruses, and protozoan pathogens remaining in the domestic wastewater and septic tank permeate.

UV Dose Response Curves

Results from the batch reactor experiments using LP UV and UV LEDs to disinfect treated domestic wastewater and septic tank effluent are given in Figure 6. These dose response curves show that the experiments were repeatable across a wide range of UVT values. These curves were used to determine the reduction equivalent dose, RED, in the flow-through reactors, given the MS2 log reduction at a specific flow rate, through biodosimetry. The dose response did not exhibit linear inactivation kinetics as in our previous study of UV LED inactivation of treated wastewater effluent; however, the wastewater in this study had higher TSS and turbidity values [25]. Suspended particles shield microorganisms from inactivation in a phenomenon called *tailing*, which invalidates the first-order kinetics model [32,36,57]. The results from wastewater disinfection match the literature, with 2-log inactivation occurring at a UV dose of 33.3 mJ/cm² for the LP UV source and 42.6 mJ/cm^2 for the LED/276 nm source [42]. The LP UV results fall within the National Water Research Institute (NWRI) Guidelines for Drinking Water and Water Reuse. LED results are not yet included in the NWRI guidelines [40]. The results for MS2 inactivation in the septic tank effluent; however, are lower than the NWRI bounds, with 2-log inactivation from an LP UV dose occurring at 28.8 mJ/cm^2 and a LED/276 nm dose of 31.3 mJ/cm^2 . It should be noted that the guidelines were developed for water with higher UVT values. Nevertheless, the enhanced sensitivity of MS2 in the septic tank effluent could be due to a number of factors, including sorption or aggregation of viral particles, oxidation from the additional dissolved organic matter, and challenges related to the particulate samples with very low UV transmittance (UVT₂₅₄ < 6%) values. For example, the higher concentration of suspended solids and particulate matter in the septic effluent provided a substrate for viral particle sorption as well as a photosensitizer for oxidation, both of which may have contributed to additional log reduction of MS2. In addition, maintaining complete sample homogeneity during UV absorbance measurements (in the cuvette), exposure (in the stirred Petri dish), and sample enumeration is challenging. For this reason, among others, UV disinfection studies with unfiltered water samples at $UVT_{254} < 10\%$ are rare.



Figure 6. Dose response curves of MS2 bacteriophage in septic tank and domestic wastewater permeates. These collimated beam tests with the batch reactors were used to determine the UV dose in our flow-through reactors.

3.3. Flow-Through UV Reactor Disinfection Efficiency

For the LP UV flow-through reactor operating with wastewater and coffee at UVT_{254} values down to 37.8% and flow rates up to 1.5 L/min, all 10^{6} – 10^{7} PFU/mL of MS2 were inactivated, indicating an applied dose of at least 140 mJ/cm². *E. coli* and total coliform at concentrations lower than 10^{4} CFU/mL were also completely inactivated in all LP UV runs for domestic wastewater and septic effluent at flow rates as high as 1.8 L/min, which was the hydraulic limit for this reactor.

The model for LP UV inactivation of MS2 in septic tank effluent (UVT₂₅₄ < 5.2%) is given in the Supplementary Info (Figure S2); with a high agreement for predicted versus observed values of MS2 log inactivation as a function of flow rate (slope = 0.9999, $R^2 = 0.9999$). As shown in Table 3, an average RED of 48.8 to 50.8 mJ/cm² was achieved at flow rates of 1.8 and 1.5 L/min for septic tank effluent. This dose was sufficient to completely inactivate *E. coli* and total coliforms remaining in the samples. Given the initial concentration of total coliforms (3.3×10^5 CFU/mL) and their dose response to LP UV light, REDs at these flow rates were more than high enough to meet the WHO irrigation guidelines for fecal coliforms of 1000 CFU/mL for WFMF-treated wastewater [29,37]. Microorganisms of concern for septic tanks, for which outbreaks have been associated with poor septic tank performance, include norovirus, rotavirus, hepatitis A, Salmonella spp., and E. coli [58]. Table 3 estimates the LP UV log-inactivation of these pathogens from these doses given this MS2 RED. These RED were also high enough for inactivation of *Giardia* spp. and *Cryptosporidium* spp., which are not linked to septic tank discharge, but require only 22 mJ/cm² for 4-log reduction [37,58]. Similarly, an LP UV dose of 40–50 mJ/cm² would completely inactivate SARS-CoV-2, the virus responsible for COVID-19, which requires only 3.7 mJ/cm² for 3-log reduction in aqueous solutions. However, water and wastewater are not considered transmission pathways of this virus [59,60].

	LP U	JV		Estimated Inactivation Credit				
Flow Rate (mL/min)	Exposure Time (s)	Log MS2 Inactivation (log ₁₀ \pm 1 SD)	RED (mJ/cm ²)	Norovirus ¹ (log ₁₀)	Rotavirus ² (log ₁₀)	Hepatitis A ² (log ₁₀)	Salmonella ² (log ₁₀)	
1500	28	3.2 ± 0.2	50.8 ± 3.8	6.0	>4.1	>5.4	>5.6	
1800	23	3.0 ± 0.3	48.8 ± 7.9	5.7	>4.1	>5.4	>5.6	

Table 3. Disinfection performance of flow-through LP UV inactivation of septic effluent (n = 4) for (UVT₂₅₄ < 5.2%). RED is the reduction equivalent dose determined from biodosimetry with the batch reactor.

¹ [61], ² [21].

The flow-through UV LED reactor, which had a lower irradiance than the LP UV reactor in this study, did not inactivate all MS2 bacteriophage in the spiked samples, as expected, which left more data for analysis. A model of the UV LED inactivation of MS2 bacteriophage as a function of flow rate and UVA-RLE for all water matrices tested was developed using a combined variable approach [47]. The modelled results are plotted along with the empirical data in Figure 7. Nonlinear regression was conducted to determine the coefficients of the mathematical model introduced in Equation (1), given below in Equation (2). The estimated coefficients for the UVC LED reactor were a = -0.38058, c = 0.48857, d = -0.30841, and e = 0.09897. Statistical significance (p < 0.05) was observed in coefficients a, c, d, and e; it was not observed in coefficient b. [47]. Statistical significance is not always reached for all coefficients of the combined variable approach; therefore, that term was removed (b = 0.0) as specified in the literature [47]. A close agreement was observed between the predicted and measured values (slope = 0.9889, $R^2 = 0.9889$, Figure 7), which indicates experimental repeatability across a wide range of UVT values.

$$\log I = 10^{-0.38058} \times \left(\frac{1}{Q}\right)^{0.48857 - 0.30841 \times UVA + 0.09897 \times UVA^2}$$
(2)

In comparison, Hull et al. [48] applied the combined variable approach for a UVC LED system challenged with MS2 bacteriophage in drinking water and obtained the following statistically significant (p < 0.05) coefficients: a = -0.1952, b = -0.25607, c = 0.65497, d = -7.99858, and e = 37.65489. Differences between the calculated coefficients may be explained by the UVT ranges used for model validation as well as the reactor design. Hull et al. utilized drinking water matrices ranging from 74.6 to 99.4% UVT, while the current study used real and synthetic wastewater with UVTs spanning from 2.0 to 88.2%. Regarding reactor design, the previous study used a spherical reactor with LEDs at one point source and a path length of up to 5.56 cm, whereas the current study used a cylindrical UV LED reactor with UV LED arrays on four sides, forming a box around the cylinder with a maximum path length of 0.9 cm [25,48].

A plot of estimated RED versus flow rate for the UV LED flow-through reactor is given in the Supplementary Info (Figure S3). With the combined variable model given in Equation (3), we could estimate the RED for a given flow rate and water matrix. With this RED, we could then estimate the log reduction of enteric pathogens, including adenovirus, coxsackievirus, and poliovirus, for the UV LED flow-through reactor. Given the published dose response data of these organisms to UV LEDs with the same wavelength emission (peak = 276.6 nm; FWHM = 9.8 nm), using Equation (3) below, we could estimate the inactivation credits (Table 4) [43,62].

$$RED = 10^{0.8716} \times \left(\frac{1}{Q}\right)^{0.4965 - 0.22554 \times UVA}$$
(3)

	UV LED		Estimated Inactivation Credit				
Flow Rate (mL/min)	UVT-RLE %	Estimated RED (mJ/cm ²)	MS2 ¹ (log ₁₀)	Adenovirus ² (log ₁₀)	Coxsackievirus ³ (log ₁₀)	Poliovirus ³ (log ₁₀)	
10	5	19.0	1.3	0.23	>4	>4	
10	30	42.5	2.1	0.80	>4	>4	
10	60	58.2	2.9	1.34	>4	>4	
10	90	69.8	3.7	1.83	>4	>4	
50	5	13.7	0.9	0.15	2.3	2.7	
50	30	23.1	1.2	0.31	>4	>4	
50	60	28.3	1.5	0.42	>4	>4	
50	90	31.9	1.7	0.51	>4	>4	

Table 4. Disinfection performance for the flow-through UV LED reactor estimated from Equations (2) and (3) for wastewater and septic tank effluent. RED is the reduction equivalent dose estimated from the combined variable model for a given flow rate and UVT. RLE = relative lamp equivalent.

¹ This study, ² [43], ³ [62].

It is important to note that the UV LED reactor is scalable, and the combined variable approach allows us to estimate the log reduction for a reactor with the same configuration for a given UV LED output, flow rate, and UV absorbance. For example, doubling the relative lamp output (S/So) and doubling the flow rate, Q, would generate the same log inactivation of a test microorganism for water with the same UV absorbance [47].



Figure 7. Measured (dots) and predicted (solid lines) MS2 log inactivation of MS2 coliphage as a function of the UV LED system flow rate and UVT-RLE. Values predicted using a combined variable approach Equation (2) as a function of UVA-RLE (presented as UVT-RLE) and flow rate with model coefficients a = -0.38058, b = 0.0, c = 0.48857, d = -0.30841, and e = 0.09897. Coefficients a, c, d, and e were statistically significant (p < 0.05), whereas b was not. Inset: Predicted MS2 log inactivation versus observed MS2 log inactivation for the UV LED reactor (276 nm) for wastewater, septic tank effluent, and coffee validation. RLE = relative lamp emission; UVT = ultraviolet transmittance.

While other studies have modeled UV inactivation of viruses or viral surrogates in flow-through UV LED systems [63], this study was, to our knowledge, the first to apply combined variable monitoring for validation of a small-scale UV LED disinfection system

used for wastewater treatment across a wide range of UVT values. This work showed that despite challenging and variable water quality conditions (2% < UVT < 88%), the disinfection was predictable across water types and flow rates. This approach enables us to optimize future reactor designs for broad applications.

WFMF is a low-cost filtration method, and UV LEDs have the major advantage of operating with low power consumption [16,20]. From a sustainability standpoint, this system could be further improved by operating both processes from remote or photovoltaic power sources, as has been done previously [64–66].

4. Conclusions

Global water demand requires increasing the use of reclaimed water, particularly for agriculture and irrigation to ease the burden on freshwater resources. However, reclaimed water poses health risks to users, and reuse must be done safely to protect public and environmental health. By making use of precious water and nutrient resources in a safe manner, wastewater reuse aligns with Sustainable Development Goal #6: To Ensure Availability and Sustainable Management of Water and Sanitation for All.

This study developed and evaluated a decentralized wastewater treatment system with a small footprint and short operating time, combining physical and photochemical processes for treating wastewater at the source for potential reuse in agriculture. WFMF with UV LEDs can be used as a final polishing step for wastewater treatment systems. The research novelty is in incorporating UV disinfection into decentralized wastewater treatment processes, which has the environmental and economic benefit of avoiding chemical disinfection prior to irrigation and discharge, while protecting public health. UV LEDs enable a paradigm shift for water and wastewater treatment processes by offering treatment efficacy with innovative, robust, and compact designs [67]. This study is one of few to investigate UV LEDs for water reuse, as a proof of concept and performance as LEDs trend toward becoming a viable option for wastewater disinfection.

For domestic wastewater from a university campus, WFMF reduced TSS (by 79.8%), turbidity (by 76.5%), COD (by 38.5%), BOD (by 47.8%), and NO₃ (by 41.4%). UVT at 254 nm improved by 19.4%, and UVT at 280 nm by 12.4%. The treatment process met the Thailand irrigation standards for all parameters tested except BOD. For conventional septic tank effluent from a small community, WFMF did not meet Thailand irrigation standards, but it reduced TSS (by 77.9%), COD (by 37.6%), and TKN (by 13.5%). For both wastewater sources, removal of microbial parameters, including total coliforms, *E. coli*, and MS2, was not statistically significant, requiring an additional disinfection step.

Following UV disinfection by LP UV and UV LEDs emitting at 276 nm, wastewater quality met the WHO standards for unrestricted irrigation. Using biodosimetry with MS2 bacteriophage, the flow-through low-pressure UV system, manufactured on-site, achieved a very high reduction equivalent dose (RED), greater than 140 mJ/cm² for the wastewater effluent. For the septic tank effluent (UVT₂₅₄ < 5.2%), it achieved an RED of 50.8 and 48.8 mJ/cm^2 for flow rates of 1.5 and 1.8 L/min, respectively. The UV LED reactor, which was also manufactured on-site, achieved up to 3.5-log reduction of MS2 in wastewater at a low flow rate of 0.01 L/min; however, the system can be scaled up by incorporating more LED arrays. For UV LED inactivation of septic tank effluent, up to 1.5-log reduction of MS2 was attained. This study is one of few to apply combined variable monitoring for validation of a small-scale UV LED disinfection system, and one of the first to cover a broad range of water qualities. The combined variable approach is scalable, allowing us to estimate the log reduction for a reactor with the same configuration for a given UV output, flow rate, and UV absorbance. This work can be used to predict the performance of other wastewaters and help inform system design and application.

Supplementary Materials: The following are available online at https://www.mdpi.com/article/1 0.3390/w13111564/s1, Figure S1: Irradiance surface profile of the UV LED batch reactor, Figure S2: Validation of predicted vs observed MS2 inactivation by the flow-through LP UV reactor in septic

effluent, Figure S3: Measured (dots) and predicted (solid lines) RED as a function of the UV LED system flow rate and UVT-RLE.

Author Contributions: S.E.B. and T.K. acquired funding; S.E.B., P.S., T.R., T.K. conceived the experiments; P.S., T.R. and B.B. designed and conducted the experiments; P.S., T.R., T.M.H.N., V.A.H.-S., N.M.H. and S.E.B. analyzed the data. P.S., T.M.H.N., T.R., V.A.H.-S. and S.E.B. prepared the original manuscript draft. T.K. reviewed and edited the manuscript. All English-speaking authors have read and agreed to the published version of the manuscript.

Funding: The research: including equipment, supplies, and personnel, were supported by a grant from the Bill & Melinda Gates Foundation in Seattle, Washington (grant OPP1029022). S. Beck was funded by the United States Department of State through a grant from the Fulbright Program (Fulbright ID 34143124). P. Suwan was supported financially by the Royal Thai Government (RTG) and Asian Institute of Technology (AIT) fellowships.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data available by contacting the corresponding author.

Acknowledgments: The authors thank the Naturally Acceptable and Technologically Sustainable (NATS) Staff for analyzing wastewater parameters and Panupong Boonyanun, senior technician in Environmental Engineering and Management Ambient Laboratory (AIT), for technical support in setting up the experimental system. We thank Chettiyappan Visvanathan for support with the woven-fiber textile and Traci Brooks for assistance with the combined variable modelling. We thank Pollawat Jamparuang at the National Institute of Metrology Thailand (NIMT) for conducting UV measurements and the Thailand Institute of Scientific and Technological Research (TISTR) for storing bacteria and virus stocks.

Conflicts of Interest: The authors declare no conflict of interest.

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