

Article

## Reassessing Domain Architecture Evolution of Metazoan Proteins: The Contribution of Different Evolutionary Mechanisms

Alinda Nagy and Laszlo Patthy \*

Institute of Enzymology, Biological Research Center, Hungarian Academy of Sciences, Budapest H-1113, Hungary; E-Mail: [nagya@enzim.hu](mailto:nagya@enzim.hu)

\* Author to whom correspondence should be addressed; E-Mail: [patthy@enzim.hu](mailto:patthy@enzim.hu); Tel.: +361-2-093-537; Fax: +361-4-665-465.

Received: 30 June 2011; in revised form: 13 July 2011 / Accepted: 2 August 2011 /

Published: 5 August 2011

---

**Abstract:** In the accompanying papers we have shown that sequence errors of public databases and confusion of paralogs and epaktologs (proteins that are related only through the independent acquisition of the same domain types) significantly distort the picture that emerges from comparison of the domain architecture (DA) of multidomain Metazoan proteins since they introduce a strong bias in favor of terminal over internal DA change. The issue of whether terminal or internal DA changes occur with greater probability has very important implications for the DA evolution of multidomain proteins since gene fusion can add domains only at terminal positions, whereas domain-shuffling is capable of inserting domains both at internal and terminal positions. As a corollary, overestimation of terminal DA changes may be misinterpreted as evidence for a dominant role of gene fusion in DA evolution. In this manuscript we show that in several recent studies of DA evolution of Metazoa the authors used databases that are significantly contaminated with incomplete, abnormal and mispredicted sequences (e.g., UniProtKB/TrEMBL, EnSEMBL) and/or the authors failed to separate paralogs and epaktologs, explaining why these studies concluded that the major mechanism for gains of new domains in metazoan proteins is gene fusion. In contrast with the latter conclusion, our studies on high quality orthologous and paralogous Swiss-Prot sequences confirm that shuffling of mobile domains had a major role in the evolution of multidomain proteins of Metazoa and especially those formed in early vertebrates.

**Keywords:** epaktologs; evolution of domain architecture; exon-shuffling; gene fusion; gene prediction error; mobility of domains; promiscuity of domains; versatility of domains

---

## 1. Introduction

### 1.1. Expected impact of Different Genetic Mechanisms on the Spectrum of Changes in Domain Architecture

**Unequal crossing-over.** It is generally recognized that one of the major genetic mechanisms responsible for changing DA of multidomain proteins is unequal crossing-over that can lead to tandem duplication of domains as well as to deletion of tandem duplicated domains (e.g., ABC  $\leftrightarrow$  ABBC). All domain-types may be duplicated/deleted by this mechanism in both prokaryotes and eukaryotes but the rate of duplication/deletion may be significantly increased by intronic recombination, explaining why mobile modules (which frequently participate in exon-shuffling) are also prone to undergo tandem duplication [1].

**Gene fusion.** It is widely accepted that fusion of neighboring genes is another major mechanism for DA change in both prokaryotes and eukaryotes. The most plausible pathway for gene fusion in animals is through co-transcription and alternative splicing of neighboring genes, followed by fixation of genomic changes that favor fusion over separate transcription of the constituent genes. This view is supported by the observation that transcripts frequently span two adjacent, tandem genes [2,3]. Typically, such chimeric transcripts begin at the promoter of the upstream gene and end at the termination point of the downstream gene, the intergenic region being spliced out of the transcript by alternative splicing in which the 5' splice site of an intron of the upstream genes is joined to the 3' splice site of an intron of the downstream gene. As pointed out earlier, such cotranscription and intergenic alternative splicing of tandem genes may have played a significant role in the evolution of multidomain proteins of eukaryotes [4].

In the case of gene fusion, the DA of the resulting chimeric gene is dictated by the relative position of the neighboring genes in the species where gene fusion occurs in as much as the upstream gene provides the N-terminal domain(s), whereas the downstream gene provides the C-terminal domain(s) of the chimeric protein. As a corollary, the degree of freedom with which genes (domains) may be combined by gene fusion depends on the rate of genomic rearrangements during evolution.

It must be emphasized that fusion of genes (e.g., when genes with DAs A and B are fused  $A + B \rightarrow AB$ ) leads to an N-terminal DA change from the perspective of the downstream gene (gene with DA = B;  $B \rightarrow AB$ ) and C-terminal change from the perspective of the upstream gene (gene with DA = A;  $A \rightarrow AB$ ), therefore the frequencies of N-terminal and C-terminal DA changes due to gene fusion are expected to be similar.

**Gene fission.** Most studies agree that the process when a gene encoding a multidomain protein is 'split' in a way such that its constituent domains are expressed separately ( $AB \rightarrow A + B$ ) is much rarer than gene fusion. [5,6]. A plausible explanation for the infrequency of gene fission relative to gene fusion is that it is much easier to lose the 5' termination signal of the upstream gene and the *cis* regulatory regions of the downstream gene (as in gene fusion, see above) than to gain both of

these (as in gene fission). Since the balance of gene fusion/fission is thus tilted in favor of fusion, it is frequently assumed that gene fusion is the main driver of the evolution of more complex multidomain proteins.

We wish to emphasize that fission of genes (resulting in DA change of the type  $AB \rightarrow A + B$ ) involves an N-terminal change from the perspective of one of the resulting genes (gene with  $DA=B$ ;  $AB \rightarrow B$ ) and C-terminal change from the perspective of the other gene (gene with  $DA=A$ ;  $AB \rightarrow A$ ), therefore the frequencies of N-terminal and C-terminal DA changes due to gene fission are also expected to be similar. It should be noted, however, that the term gene fission is sometimes (e.g., [7]) used in a different sense: loss of terminal domains (e.g.,  $AB \rightarrow A$  or  $AB \rightarrow B$ ).

**Point mutations changing the boundaries of the Open Reading Frame.** In principle, changing signals for translation initiation and termination may favor loss over gain of terminal domains, since the use of a novel translation initiation site upstream of the original site (converting 5'-untranslated region to translated region) or conversion of the original stop codon to a sense codon (converting the 3'-untranslated region to translated region) is unlikely to result in the gain of a new domain since the 5'- and 3'-untranslated regions were not selected to encode folded domains. Conversely, the gain of a novel translation initiation site downstream of the original one may lead to the loss of an N-terminal domain (e.g.,  $AB \rightarrow B$ ) or the gain of a novel translation termination site upstream of the original one may lead to the loss of C-terminal domains of a protein (e.g.,  $AB \rightarrow A$ ). As a corollary, changing signals for translation initiation and termination would favor loss over gain of terminal domains. As to the relative probabilities of loss of N-terminal or C-terminal domains: we must take into account the fact that nonsense-mediated decay (NMD) may detect premature stop codons [8] and may prevent the formation of C-terminally truncated proteins and thus act against the loss of C-terminal domains.

**Exon-shuffling and other ways of domain-shuffling.** It is generally agreed that shuffling of symmetrical class 1-1 domains (domains flanked by phase 1 introns) by intronic recombination contributed significantly to the evolution of multidomain proteins of Metazoa but it is also clear that intronic recombination is not an absolute prerequisite of domain-shuffling [1,9–14].

Analysis of a large number of cases where the evolutionary history of the DA change involving class 1-1 domains could be reliably reconstructed revealed that—as a rule—exons/exons-sets encoding class 1-1 domains are inserted in pre-existing phase 1 introns of the recipient gene [13]. The resulting DA change may be classified as N-terminal, C-terminal or internal DA change, depending on the position of the intron where the class 1-1 domain was inserted. For example, in the collection of examples discussed in the accompanying papers the class 1-1 TSP1 domains were inserted internally in the case of the thrombospondin family (see TSP2\_HUMAN), the class 1-1 Laminin EGF domains and SEA-domains were inserted internally during evolution of agrins [15], the class 1-1 FN1 domain was inserted at an N-terminal position during evolution of plasminogen activators (see TPA\_HUMAN; [8]), the class 1-1 kringle-domain was inserted internally in thrombin during evolution of blood coagulation proteins (see THRB\_HUMAN; [8]), the class 1-1 FN2 domain was inserted internally during evolution of the MMP and SE1L families (see MMP2\_HUMAN and SE1L1\_HUMAN).

The same genomic features that are essential for exon-shuffling (introns of identical phase at the boundaries of the domain that is shuffled) also facilitate the loss of domains acquired by exon-shuffling through fixation of exon skipping [16].

A survey of the various genetic mechanisms that may change the DA of proteins thus suggests that unequal crossing-over, gene-fusion and domain-shuffling are the ‘creative’ mechanisms that may increase the complexity of the DA of multidomain proteins. Conversely, the DA complexity of multidomain proteins may be decreased by unequal crossing-over, gene fission, point mutations that change the boundaries of the open reading frame and fixation of exon-skipping.

It must be emphasized that a major difference between gene fusion and domain-shuffling is that the former may alter DAs only at the termini (e.g.,  $A + B \rightarrow AB$  or  $AB + C \rightarrow ABC$  or  $A + BC \rightarrow ABC$ ) whereas exon-shuffling does not have this requirement: it may add domains both internally (e.g.,  $AB + C \rightarrow ACB$ ) or at the termini (e.g.,  $AB + C \rightarrow ABC$  or  $AB + C \rightarrow CAB$ ). As a corollary, the relative frequency of DA change in internal positions versus N-terminal and C-terminal positions may be used to assess the relative contribution of gene fusion and domain-shuffling to DA evolution.

For example, if we assume that gene fusion is the dominant mechanism responsible for DA change we expect that the rates of DA change at the N-terminal and C-terminal ends of proteins significantly exceeds that observed at internal positions. Conversely, if we assume that domain-shuffling was the dominant mechanism responsible for DA change we expect that the rates of DA change at N-terminal, C-terminal and internal positions are roughly similar. It should be noted, however, that this analysis would be meaningless if we disregard the fact that in the case of one-domain  $\leftrightarrow$  two domain transitions (type 1 transitions that account for the majority of DA changes), domain architecture change can only be classified as terminal (e.g.,  $A \leftrightarrow AB$  or  $A \leftrightarrow BA$ ).

### *1.2. Expected impact of Different Genetic Mechanisms on the Versatility, Promiscuity and Mobility of Domains*

Domains of multidomains proteins are frequently labeled with epithets to express some aspects of their role in the evolution of different domain architectures. The most frequently used terms are ‘versatile domains’, ‘promiscuous domains’, ‘mobile domains’ and sometimes these terms are used as if they were synonyms [17].

The generally accepted definition of domain promiscuity/versatility is that domains are promiscuous/versatile if they are present in many different domain architectures [18–20]. Typically, the degree of promiscuity of a domain is defined as the number of distinct architectures in which it is present or the number of domain-types associated with it. For example, the Pfam A domains ‘trypsin’ and ‘pkinase’ have high promiscuity/versatility scores since they combine with a large variety of domains to form a rich repertoire of domain architectures. It should be noted, however, that the terms promiscuity/versatility have no implications as to the genetic mechanism that alters DA.

In contrast with this, the term ‘domain mobility’ is intended to reflect the frequency with which a domain is shuffled, *i.e.*, moved from one local environment (within a gene) to a new local environment (of another gene). Accordingly, the mobility of a domain is related to but not equivalent with its versatility/promiscuity: mobile domains are necessarily versatile, but versatile domains are not necessarily mobile. For example, domain A is involved in a mobility event if it is inserted into a new environment of a recipient gene (e.g., encoding a protein with DA XYZ to give protein with DA XYAZ) but the XY and Z domains of the recipient gene are not since they did not move from their original environment: the mobility score of domain A is increased by one count, but the mobility

scores of X, Y and Z are unaffected. The situation is quite different for versatility/promiscuity scores: the DA change of XYZ → XYAZ will equally affect the versatility/promiscuity score of all four domains involved. This point may be illustrated by the cases of the trypsin domains of regulatory proteases such as TPA\_HUMAN, THRB\_HUMAN or NTR\_HUMAN: these vertebrate-specific multidomain architectures arose by shuffling of mobile domains (e.g., class 1-1 kringle-, FN1-, SRCR-domains) whereas, in these DA changes, their common trypsin domain served as recipients for the mobile domains.

Despite the practical problems associated with genome-scale reconstruction of such events some prototypical studies may serve to illustrate the importance of the distinction of versatility and mobility of domains. Analysis of the evolutionary history of proteases of the blood coagulation and fibrinolytic cascade revealed that during evolution of these paralogous proteins of the trypsin-family a variety of mobile modules (e.g., kringle-, EGF-, FN1- and FN2-modules) were inserted into the ‘recipient’ genes [9,21]. In terms of versatility, each acquisition of a novel mobile module increases the versatility/promiscuity score of the trypsin-like protease domain of the recipient genes, even though its mobility score is unaffected.

In our definition domain-shuffling (domain mobility) is restricted to cases where the partners involved in DA changes are non-equivalent; the domain is moved from one genome location (a donor gene) to a recipient gene. This definition excludes gene fusion as a mobility event since both partners can be considered donor and acceptor. Conversely, domain insertion (e.g., by exon-shuffling) is a mobility event; since the roles of the recipient gene and donor genes are non-equivalent and can be clearly distinguished.

### *1.3. Expected Impact of Sequence Errors on Conclusions About Domain Architecture Evolution of Metazoan Proteins*

As emphasized in an accompanying paper (Nagy, Szláma, Szarka, Trexler, Bányai, Patthy, Reassessing Domain Architecture Evolution of Metazoan Proteins: Major Impact of Gene Prediction Errors), reliable analysis of DA evolution of multidomain proteins requires that the protein sequences compared are valid, correct and complete. It must be pointed out that many authors realized that gene annotation errors may cause some problems in the analysis of DA of proteins but most studies implicitly assumed that such errors may be neglected in genome-scale analyses and that they occur at random thus they do not obscure the general tendencies of DA evolution. We have shown in an accompanying paper that neither of these assumptions is justified (Nagy, Szláma, Szarka, Trexler, Bányai, Patthy, Reassessing Domain Architecture Evolution of Metazoan Proteins: Major Impact of Gene Prediction Errors). First, in the case of most Metazoan species the contribution of gene prediction errors to domain architecture differences of orthologous and paralogous proteins is comparable/greater than those of true gene rearrangements. Second, the accuracy of gene prediction itself has a strong positional bias, in as much as it is most reliable for internal exons and least reliable for N-terminal exons [22], thus, errors in gene prediction do not merely increase the rate of DA differences at random: they introduce a strong positional bias in favor of apparent terminal DA changes.

As discussed above, neither unequal crossing-over, nor gene fusion nor domain-shuffling are expected to favor N-terminal versus C-terminal DA change, thus we have no obvious genetic

explanation for the observation in genome-scale studies that DA changes are preferred at the N-terminal end [5]. It seemed more likely that this observation reflects the fact that the majority of erroneous (incomplete, mispredicted) sequences present in databases such as TrEMBL, EnsEMBL differ from the correct sequence more frequently at the N-terminal end than the C-terminal end. By analyzing type 1 transitions (one domain  $\leftrightarrow$  two domain transitions), type 2 transitions, (two-domain  $\leftrightarrow$  three domain transitions) and type 3 transitions (N-domain  $\leftrightarrow$  N + 1-domain transitions, where N is greater than 2), separately we have shown that in the case of high quality Swiss-Prot proteins of Metazoa the probability of DA change is similar at internal and terminal positions. In contrast with this, in the case of TrEMBL (where a significant proportion of the sequences is incomplete, incorrect or aberrant), RefSeq, EnsEMBL and NCBI's GNOMON predicted sequences (that frequently contain mispredicted sequences) the apparent rate of terminal changes were significantly increased relative to internal changes.

Our findings thus cautioned that earlier proteome-scale studies that neglected the contribution of sequence errors may have led to erroneous conclusions about the evolution of novel domain architectures of multidomain proteins. Our observation on high quality Swiss-Prot sequences, that the contribution of internal DA alterations increased in vertebrates (Nagy, Banyai and Patthy, Reassessing Domain Architecture Evolution of Metazoan Proteins: Major Impact of Errors Caused by Confusing Paralogs and Epaktologs), is consistent with our suggestions that exon-shuffling played a major role in shaping the DA of multidomain proteins unique to vertebrates [12].

#### *1.4. Expected Impact of Confusing Epaktologs and Paralogs on Conclusions about Domain Architecture Evolution of Metazoan Proteins*

We have demonstrated that contamination of protein families with epaktologs (proteins that are related only through the independent acquisition of the same domain types) increases the apparent rate of DA change and introduces a strong bias in DA differences in as much as it increases the proportion of terminal over internal DA differences (Nagy, Banyai, Patthy, Reassessing Domain Architecture Evolution of Metazoan Proteins: Major Impact of Errors Caused by Confusing Paralogs and Epaktologs). These findings cautioned that earlier studies based on analysis of datasets of protein families that were contaminated with epaktologs may have led to some erroneous conclusions about the evolution of novel domain architectures of multidomain proteins.

## **2. Results and Discussion**

In view of our observation that sequence errors and confusion of epaktologs with other types of homologs significantly distorts the evolutionary history of the DA of multidomain proteins, it is important to re-examine the conclusions of earlier studies that neglected the influence of these errors. As emphasized in the accompanying papers, these errors not only increase the apparent rate of DA change but they also introduce a strong positional bias in favor of terminal over internal DA changes.

In the case of bacterial genomes (where it is justified to neglect misprediction) it was convincingly shown that terminal changes are significantly (more than 10-fold) more frequent than internal ones [23]. Interestingly, studies that analyzed datasets of Archean, Bacterial, Eukaryotic proteins have noted a similar degree of bias in favor of terminal over internal DA changes for eukaryotes and prokaryotes,

leading several authors to conclude that this bias is also valid for eukaryotic organisms [5,6,24–27]. Since there are major differences in the organization of genomes/genes of prokaryotes and higher eukaryotes such as Metazoa one would expect that these differences have some impact on DA evolution. Indeed, there is a general consensus that the rate of formation of new DAs is significantly higher in Metazoa than in prokaryotes or other eukaryotes [25] so it is even more surprising that this increase in the rate of DA evolution (that is generally attributed to an increased role of exon-shuffling in the Metazoan lineage [12]) is not reflected in a shift in favor of internal DA changes.

We suggest that the absence of this shift is due to the fact that in most studies the high proportion of incomplete, abnormal or mispredicted sequences of higher eukaryotes increased the rate of terminal vs. internal changes and this was taken as evidence for gene fusion. This point may be illustrated by the work of Weiner *et al.* [24]. These authors have analyzed the whole SwissProt/TrEMBL set of proteins and concluded that DA changes occur most frequently at termini which in turn led the authors to conclude that “these results have further supported the emerging view that, by and large, the modular evolution of proteins is dominated by two major types of events: fusion, on the one hand, and deletion and fission on the other”. Buljan and Bateman [27] have also studied domain architecture evolution in animal gene families represented in UniProt (Swiss-Prot plus TrEMBL) database and these authors have also concluded that gain and loss of domains is preferred at protein termini. As we have pointed out in an accompanying paper (Nagy, Szláma, Szarka, Trexler, Bányai, Patthy, Reassessing Domain Architecture Evolution of Metazoan Proteins: Major Impact of Gene Prediction Errors), as a consequence of the large proportion of incomplete sequences in the TrEMBL section of UniProtKB the DA of these erroneous sequences differ from those of the correct sequences at the termini thus falsifying the positional distribution of DA changes during protein evolution. Accordingly, the conclusions drawn from DA analysis of datasets dominated by TrEMBL sequences seem to be unjustified.

In their genome-scale studies on DA evolution, Ekman *et al.* [25] have used the Ensembl proteomes for the eukaryotic genomes. They also concluded that most events involve a single domain, which is inserted at the N or C termini, implying that gene fusion is the dominant mechanism for DA change. As we have pointed out, as a consequence of the large proportion of mispredicted sequences in the Ensembl proteomes the DAs of these erroneous sequences differ from those of the correct sequences at the termini thus falsifying the positional distribution of DA changes during protein evolution. Accordingly, the conclusions drawn from DA analysis of Ensembl sequences seem to be unjustified.

Despite the fact that the use of erroneous sequences and other types of methodological errors casts doubt on the results of such analyses the view that the major genetic operations leading to novel arrangements are fusion of existing genes and terminal loss of domains is gaining popularity since it is propagated in several recent review papers [26,28].

Realizing the danger of confusing gene- and protein-annotation errors with true changes of DA, in a recent paper Buljan *et al.* [29] have chosen the strategy that instead of genome-scale analysis of DA evolution of datasets (of dubious quality) they analyzed a limited set of cases that they considered as high confidence domain gain events in Metazoa. Based on these studies they have reached the same conclusion as in their earlier genome-scale studies: “the major mechanism for gains of new domains in metazoan proteins is likely to be gene fusion through joining of exons from adjacent genes”. The

authors have also concluded that “insertion of exons into ancestral introns through intronic recombination are, in contrast to previous expectations, only minor contributors to domain gains” and have accounted for less than 10% of high confidence domain gain events. Buljan *et al.* also noted that the DA change occurs more frequently at the N-terminal than the C-terminal end. In a Research Highlight commentary of the work of Buljan *et al.* Marsh and Teichmann [30] have concluded that “although recombination between introns has been speculated to be one of the main mechanisms behind the diverse domain rearrangements observed in complex eukaryotes, it seems to have made a fairly limited contribution to the domain gain events”.

Since these conclusions contradict our data and the widely accepted view that exon-shuffling played a major role in DA evolution of proteins unique to Metazoa we have carefully examined the evidence presented in the paper of Buljan *et al.* [29]. Our analyses identified four major types of problems with the analysis of Buljan *et al.* that explain this contradiction.

The first and most important problem is that the set of ‘high-confidence domain gain events’ does not properly represent the spectrum of DA changes (e.g., noteworthy absence of numerous, well documented, high-confidence domain gain events). The authors acknowledge that the limited set (330 cases) of ‘high-confidence domain gain events’ may not properly represent the whole spectrum of DA changes: “even though we do not expect that the final set of high-confidence domain gains is biased towards any of the mechanisms, the total number of gain events in the set is relatively small and this could introduce apparent dominance of one mechanism over another”. To check this possibility they use a larger number of ‘medium-confidence’ domain gains (849 cases) to “test whether a larger set of domain gains would support the observed distribution of characteristics of gained domains”. Although they note that the “major difference between the two sets was in the number of middle domains coded by one exon” they dismiss this warning sign (pointing to the role of exon-shuffling) by saying that “we believe that this is largely due to false domain gain”.

The problems with the data set of Buljan *et al.* are probably best illustrated by the fact that many of the best-known, well-documented cases of domain-gains are missing from the list (Table S1 in [29]). If we check the presence/absence of the examples discussed in the accompanying papers we find that the majority are missing. For example, TreeFam tree TF329901 that presents many well-documented, exon-shuffling mediated cases of DA rearrangements of plasminogen-related proteins, urokinase, tissue-plasminogen activator (see TPA\_HUMAN), hyaluron-binding protein, blood coagulation factor 12, hepatocyte-growth factor activator etc. is missing from the list of high-confidence domain gain events. Similarly, trees TF315428, (containing MMP2/MMP9, the common ancestor of which is known to have gained FN2 domains by exon-shuffling; see MMP2\_HUMAN), TF324917 (containing TSP1/TSP2 (the common ancestor of which is known to have gained TSP1 domains by exon-shuffling; see TSP2\_HUMAN), TF315257 (containing sel-1 homologs where SE1L1 proteins are known to have gained an FN2 domain by exon-shuffling; see SE1L1\_HUMAN; TF326548 (containing agrins that are known to have gained a SEA domain by exon-shuffling in the chordate lineage; see AGRIN\_HUMAN), TF317274 (containing amyloid A4 precursor, known to have gained a Kunitz domain in the vertebrate lineage by exon-shuffling, see A4\_HUMAN) are also missing from the list of ‘high-confidence domain gain events’.

Another important problem with the analysis of Buljan *et al.* is that the conclusions are sometimes erroneous as a consequence of the fact that TreeFam sometimes confuses paralogs and epaktologs.

As discussed in an accompanying paper (Nagy, Banyai and Patthy, Reassessing Domain Architecture Evolution of Metazoan Proteins: Major Impact of Errors Caused by Confusing Paralogs and Epaktologs), this type of error is encountered most frequently in the case of epaktologous proteins that contain tandem repeats of the same domain-type. For example, in tree TF326157 (Family Name: Complement factor H-related protein precursor) complement factor H and factor XIIIb that arose in vertebrates (consisting of tandem sushi domains) are claimed to belong to the same family as some invertebrate proteins (e.g., CG10186, Q8INW2\_DROME) simply because they also contain multiple tandem sushi domains.

As a consequence of the contamination of some TreeFam trees with epaktologs the evolutionary history of the DA of proteins and the conclusions drawn from this history may be erroneous. The case of TF329295 is especially instructive. The common feature of proteins listed in this tree is that they contain tandem SRCR domains, including CD5-, CD6-, SRCRL-, C163A-, DMBT1- and NETR-related sequences, thus the tree implicitly assumes that they are all descendants of a common ancestor with multiple SRCR domains. Consequently, the tree implies that additional domains that are present only on some branches of the tree (*zona\_pellucida* domain in DMBT1-related sequences and trypsin in NETR-related sequences) (see Figure 2/c of [31]) were gained in the corresponding trees. As a consequence, in the list of ‘High confidence domain gain events’ (Table S1 of [29]) the authors claim that in the TF329295 family a trypsin domain (CL0124 Peptidase\_PA, trypsin) has been gained in vertebrates. As representative transcript with the gained domain they give ENST00000296498 (protein ID ENSP00000296498 equivalent with neurotrypsin, NETR\_HUMAN). The basis of this interpretation is that the tree TF329295 implies that the closest paralogs of neurotrypsin are CD5-, CD6-, DMBT-, WC11-related proteins *etc.* that all contain multiple SRCR domains. In contrast with this interpretation, neurotrypsins are paralogs of the plasminogen-activator branch of trypsin-like proteins (based on the evolutionary affiliations of its kringle and protease domains) and the present DA of neurotrypsin evolved through the gain of internal mobile SRCR domains by an ancestral protease and not through the gain of a terminal trypsin domain by an ancestral CD5-like protein. Similarly, in the list of ‘High confidence domain gain events’ in TreeFam family tree TF329295 ENSMUST00000084509 (ENSMUSP00000081556, corresponding to DMBT1\_MOUSE) is listed as an example of domain gain, claiming that this protein arose from an ancestor with multiple SRCR domains by gaining a terminal PF00100 (*Zona\_pellucida*) domain, rather than from an ancestor with a *Zona\_pellucida* domain through acquisition of mobile SRCR domains.

A third major source of errors is that the authors rely on EnSEMBL sequences, therefore the analysis inherits the problems (presence of erroneous sequences) of this database. For example, when we analyzed the dataset by blasting the EnSEMBL entries against UniProtKB we identified several cases where the corresponding UniProtKB entry was annotated as ‘no protein product’ or ‘retired entry’.

Sometimes the authors draw conclusions based on analysis of abnormal transcripts. In Additional file 8 of [29] the authors illustrate their conclusions by highlighting some examples of domain gains by joining of exons from adjacent genes. Here the authors discuss the case of CELSR3 (Treefam tree TF323983) containing Cadherin EGF LAG seven-pass G-type receptor (CESLR) precursor genes as an example of DA change in the vertebrate lineage. The authors conclude that “one branch of the family, containing vertebrate genes, has gained the Sulfate transport and STAS domains in addition to the

ancestral cadherin, EGF and other extracellular domains” and suggest that “ the gain occurred after the other vertebrates diverged from fish and homologues without the gained domains are present in all animals.” A closer examination of this case, however, questions the validity of the conclusion that a gene fusion has occurred. The single experimental evidence supporting this claim is that a cDNA (Accession AY714129) was cloned that arose as a result of co-transcription of the closely linked genes for the cadherin EGF LAG seven-pass G-type receptor 3-like protein CELR3\_HUMAN and the gene for sulfate transport protein S26A6\_HUMAN. When we searched the human EST database with the ‘fusion’ region we identified numerous ESTs for the separate transcription of the two genes but no EST supporting the fusion. These results indicate that a rare event of co-transcription and alternative splicing gave rise to transcript AY714129 but this should not be confused with genomic rearrangement leading to DA change. It is worth noting that the transcript AY714129 is also abnormal in the sense that the protein product (Accession AAU94938) lacks the cleavable signal peptide of CELR3\_HUMAN, so the putative protein (Q5Y190\_HUMAN) is unlikely to be integrated normally in the plasma membrane (Figure S1). The fact that two predicted “low quality protein sequences” (XP\_002808368.1 of *Macaca mulatta* and XP\_002926069.1 of *Ailuropoda melanoleuca*) also arose by (*in silico*) fusion of the orthologous tandem genes CELR3 and S26A6 cannot be used to support gene fusion; it just illustrates the danger of error-propagation.

Analysis of the case CELR3/S26A6 ‘fusion’ thus has several important messages. First, since co-transcription of tandem genes is quite general [32], there is a significant probability that in cDNA and EST databases one will find transcripts derived from independent genes through co-transcription and alternative splicing [2,3]. If the existence of such transcripts (fusion at the transcript level) is erroneously equated with fusion at the gene level, the contribution of gene fusion to DA change will be significantly overestimated. Second, the influence of such errors in the interpretation of the data of DA evolution becomes more exaggerated as a result of error-propagation in gene predictions.

As an example of erroneous conclusion based on errors in gene prediction we may mention the case of TreeFam family TF351422 that the authors highlight as an example of a domain gain after segmental duplication and exon joining. This TreeFam family contains only primate sequences and it is claimed that after a gene duplication event one branch of the family has gained the PTEN\_C2 (PF10409) domain (Additional file 8, of [29]). A closer look at the evidence presented here, however, raises doubts about the validity of the scenario proposed by the authors. As representative transcript the authors give ENST00000381866 (protein ID ENSP00000371290 equivalent with Swiss-Prot entry YM016\_HUMAN). Although there is experimental support for the existence of the transcript ENST00000381866 containing the PTEN\_C2 domain, the other sequences of this branch of TreeFam family TF351422 (implied to represent the ‘acceptor’ state) are very short predicted sequences whose existence is not supported by ESTs. It is noteworthy that the corresponding entries Q5T6R3\_HUMAN and Q49A76\_HUMAN have been deleted recently from the UniProtKB database.

As another example, we may mention the case of TreeFam family TF330855 (MSR1/SCARA5, Macrophage receptor family). In the list of ‘High confidence domain gain events’ (Table S1 of [29]) the authors claim that in this family the branch of Macrophage scavenger receptor types I and II acquired the PF03523 (Macscav\_rec) domain only in mammals. As representative transcript with the gained domain they give ENST00000262101 (protein ID ENSP00000262101, equivalent with Swiss-Prot entry MSRE\_HUMAN). The only non-mammalian Macrophage scavenger receptor types I and II

in TreeFam TF330855 (that serves as a representative of the state before the gain of domain PF03523) is SCARA5\_XENTR (ENSXETT00000037776, ENSXETP00000037776). The predicted protein ENSXETP00000037776, however, is clearly truncated at the N-terminal end (its first amino acid is an Asn) so this cannot be taken as evidence for a domain gain in mammals (Figure S2).

In summary, although one could argue that it could have been worth paying the price of small-scale analysis in order to avoid errors, a closer examination of the cases of ‘high-confidence’ DA changes indicates that the analysis of Buljan *et al.* failed to achieve this goal. First, the limited set of ‘high-confidence domain gain events’ does not properly represent the spectrum of DA changes. Second, the dataset relies on EnsEMBL therefore the analysis inherits the problems associated with errors in gene prediction (mispredicted or non-existent genes). Third, the authors rely on TreeFam where many trees contain (in addition to orthologs and paralogs) epaktologs.

### 3. Conclusions

We have shown that earlier conclusions that DA changes of Metazoan multidomain proteins occur preferentially at termini (and that the major mechanism for gains of new domains in metazoan proteins is gene fusion) are unwarranted since they reflect the fact that the databases used in these studies (e.g., UniProtKB/TrEMBL, EnsEMBL) were significantly contaminated with incomplete, abnormal and mispredicted sequences and that the authors failed to separate paralogs and epaktologs. Our studies that eliminated these problems (use of high quality Swiss-Prot sequences, separation of paralogs from epaktologs) confirmed that shuffling of mobile domains had a major role in the evolution of multidomain proteins of Metazoa and especially those formed in early vertebrates.

### Acknowledgements

This research was supported by the grant TECH\_09\_A1-FixPred9 from the National Office for Research and Technology of Hungary.

### References

1. Patthy, L. Modular exchange principles in proteins. *Curr. Opin. Struct. Biol.* **1991**, *1*, 351–361.
2. Akiva, P.; Toporik, A.; Edelheit, S.; Peretz, Y.; Diber, A.; Shemesh, R.; Novik, A.; Sorek, R. Transcription-mediated gene fusion in the human genome. *Genome Res.* **2006**, *16*, 30–36.
3. Parra, G.; Reymond, A.; Dabbouseh, N.; Dermitzakis, E.T.; Castelo, R.; Thomson, T.M.; Antonarakis, S.E.; Guigo, R. Tandem chimerism as a means to increase protein complexity in the human genome. *Genome Res.* **2006**, *16*, 37–44.
4. Magrangeas, F.; Pitiot, G.; Dubois, S.; Bragado-Nilsson, E.; Cherel, M.; Jobert, S.; Lebeau, B.; Boisteau, O.; Lethe, B.; Mallet, J.; *et al.* Cotranscription and intergenic splicing of human galactose-1-phosphate uridylyltransferase and interleukin-11 receptor alpha-chain genes generate a fusion mRNA in normal cells. Implication for the production of multidomain proteins during evolution. *J. Biol. Chem.* **1998**, *273*, 16005–16010.
5. Björklund, A.K.; Ekman, D.; Light, S.; Frey-Skött, J.; Elofsson, A. Domain rearrangements in protein evolution. *J. Mol. Biol.* **2005**, *353*, 911–923.

6. Kummerfeld, S.K.; Teichmann, S.A. Relative rates of gene fusion and fission in multi-domain proteins. *Trends Genet.* **2005**, *21*, 25–30.
7. Fong, J.H.; Geer, L.Y.; Panchenko, A.R.; Bryant, S.H. Modeling the evolution of protein domain architectures using maximum parsimony. *J. Mol. Biol.* **2007**, *366*, 307–315.
8. Behm-Ansmant, I.; Kashima, I.; Rehwinkel, J.; Saulière, J.; Wittkopp, N.; Izaurralde, E. mRNA quality control: An ancient machinery recognizes and degrades mRNAs with nonsense codons. *FEBS Lett.* **2007**, *581*, 2845–2853.
9. Patthy, L. Evolution of the proteases of blood coagulation and fibrinolysis by assembly from modules. *Cell* **1985**, *41*, 657–663.
10. Patthy, L. Intron-dependent evolution: Preferred types of exons and introns. *FEBS Lett.* **1987**, *214*, 1–7.
11. Patthy, L. Exon shuffling and other ways of module exchange. *Matrix Biol.* **1996**, *15*, 301–310.
12. Patthy, L. Genome evolution and the evolution of exon-shuffling—A review. *Gene* **1999**, *238*, 103–114.
13. Patthy, L. *Protein Evolution*; Blackwell Publishing Ltd: Oxford, UK, 1999.
14. Patthy, L. Modular assembly of genes and the evolution of new functions. *Genetica* **2003**, *118*, 217–231.
15. Bányai, L.; Sonderegger, P.; Patthy, L. Agrin binds BMP2, BMP4 and TGFβ1. *PLoS One* **2010**, *5*, e10758.
16. Patthy, L. Alternative Splicing: Evolution. In *Encyclopedia of Life Sciences (ELS)*; John Wiley and Sons, Ltd.: Chichester, UK, 2008.
17. Basu, M.K.; Poliakov, E.; Rogozin, I.B. Domain mobility in proteins: Functional and evolutionary implications. *Brief. Bioinform.* **2009**, *10*, 205–216.
18. Song, N.; Joseph, J.M.; Davis, G.B.; Durand, D. Sequence similarity network reveals common ancestry of multidomain proteins. *PLoS Comput. Biol.* **2008**, *4*, e1000063.
19. Basu, M.K.; Carmel, L.; Rogozin, I.B.; Koonin, E.V. Evolution of protein domain promiscuity in eukaryotes. *Genome Res.* **2008**, *18*, 449–461.
20. Chothia, C.; Gough, J. Genomic and structural aspects of protein evolution. *Biochem. J.* **2009**, *419*, 15–28.
21. Patthy, L. Evolutionary assembly of blood coagulation proteins. *Semin. Thromb. Hemost.* **1990**, *16*, 245–259.
22. Bernal, A.; Crammer, K.; Hatzigeorgiou, A.; Pereira, F. Global discriminative learning for higher-accuracy computational gene prediction. *PLoS Comput. Biol.* **2007**, *3*, e54.
23. Pasek, S.; Risler, J.L.; Brézellec, P. Gene fusion/fission is a major contributor to evolution of multi-domain bacterial proteins. *Bioinformatics* **2006**, *22*, 1418–1423.
24. Weiner, J., 3rd; Beaussart, F.; Bornberg-Bauer, E. Domain deletions and substitutions in the modular protein evolution. *FEBS J.* **2006**, *273*, 2037–2047.
25. Ekman, D.; Björklund, A.K.; Elofsson, A. Quantification of the elevated rate of domain rearrangements in metazoa. *J. Mol. Biol.* **2007**, *372*, 1337–1348.
26. Moore, A.D.; Björklund, A.K.; Ekman, D.; Bornberg-Bauer, E.; Elofsson, A. Arrangements in the modular evolution of proteins. *Trends Biochem. Sci.* **2008**, *33*, 444–451.

27. Buljan, M.; Bateman, A. The evolution of protein domain families. *Biochem Soc Trans.* **2009**, *37*, 751–755.
28. Bornberg-Bauer, E.; Huylmans, A.K.; Sikosek, T. How do new proteins arise? *Curr. Opin. Struct. Biol.* **2010**, *20*, 1–7.
29. Buljan, M.; Frankish, A.; Bateman, A. Quantifying the mechanisms of domain gain in animal proteins. *Genome Biol.* **2010**, *11*, R74.
30. Marsh, J.A.; Teichmann, S.A. How do proteins gain new domains? *Genome Biol.* **2010**, *11*, 126.
31. Nagy, A.; Bányai, L.; Patthy, L. Reassessing Domain Architecture Evolution of Metazoan Proteins: Major Impact of Errors Caused by Confusing Paralogs and Epaktologs. *Genes* **2011**, *2*, 516–561.
32. ENCODE Project Consortium. Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project. *Nature* **2007**, *447*, 799–816.

### Supplementary Materials

**Figure S1.** Alignment of the fusion product (q5y190\_human) with the products of the distinct tandem genes for celr3\_human and s26a6\_human found on chromosome 3. Note that the fusion protein lacks a cleavable signal peptide characteristic of a type I transmembrane protein (the signal peptide of celr3\_human is underlined).

	1		50
celr3_human	~~~~~	~~~~~	~~~~~
q5y190_human	~MGPPSKRLS	EVEAATFHRK	PMGGKYGVLV GDALLTAYCQ KARISHSTPS
s26a6_human	~~~~~	~~~~~	~~~~~
	51		100
celr3_human	~~~~~	~~~~~	<u>.MMARRPPWR</u> GLGERSTPIL LLLLLSLFPL
q5y190_human	QSYENAHGHQ	NFRPKSGTRF	RTSGQEGGTR DLEGQKKAGR RSWDSALEGG
s26a6_human	~~~~~	~~~~~	~~~~~
	101		150
celr3_human	SQEELGGGGH	QGWDPLAAT	TGPRAHIGGG ALALCPSSG VREDGGPGLG
q5y190_human	SQEELGGGGH	QGWDPLAAT	TGPRAHIGGG ALALCPSSG VREDGGPGLG
s26a6_human	~~~~~	~~~~~	~~~~~
	151		200
celr3_human	VREPIFVGLR	GRRQSARNSR	GPPEQPNEEL GIEHGVQPLG SRERETGQGP
q5y190_human	VREPIFVGLR	GRRQSARNSR	GPPEQPNEEL GIEHGVQPLG SRERETGQGP
s26a6_human	~~~~~	~~~~~	~~~~~
	201		250
celr3_human	GSVLYWRPEV	SSCGRTGPLQ	RGSLSPGALS SGVPGSGNSS PLPSDFLIRH
q5y190_human	GSVLYWRPEV	SSCGRTGPLQ	RGSLSPGALS SGVPGSGNSS PLPSDFLIRH
s26a6_human	~~~~~	~~~~~	~~~~~
	251		300
celr3_human	HGPKPVSSQR	NAGTGSRKRK	GTARCCGELW ATGSKGQGER ATTSGAERTA
q5y190_human	HGPKPVSSQR	NAGTGSRKRK	GTARCCGELW ATGSKGQGER ATTSGAERTA
s26a6_human	~~~~~	~~~~~	~~~~~
	301		350
celr3_human	PRRNCLPGAS	GSGPELDSAP	RTARTAPASG SAPRESRTAP EPAPKRMRSR
q5y190_human	PRRNCLPGAS	GSGPELDSAP	RTARTAPASG SAPRESRTAP EPAPKRMRSR
s26a6_human	~~~~~	~~~~~	~~~~~

Figure S1. Cont.

	351				400
celr3_human	GLFRCRFLPQ	RPGPRPPGLP	ARPEARKVTS	ANRARFRRRAA	NRHPQFFPQYN
q5y190_human	GLFRCRFLPQ	RPGPRPPGLP	ARPEARKVTS	ANRARFRRRAA	NRHPQFFPQYN
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	401				450
celr3_human	YQTLVPENEA	AGTAVLRVVA	QDPDAGEAGR	LVYSLAALMN	SRSLELFSID
q5y190_human	YQTLVPENEA	AGTAVLRVVA	QDPDAGEAGR	LVYSLAALMN	SRSLELFSID
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	451				500
celr3_human	PQSGLIRTAA	ALDRESMERH	YLRVTAQDHG	SPRLSATTMV	AVTVADRNDH
q5y190_human	PQSGLIRTAA	ALDRESMERH	YLRVTAQDHG	SPRLSATTMV	AVTVADRNDH
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	501				550
celr3_human	SPVFEQAQYR	ETLRENVEEG	YPILQLRATD	GDAPPNANLR	YRFVGPPAAR
q5y190_human	SPVFEQAQYR	ETLRENVEEG	YPILQLRATD	GDAPPNANLR	YRFVGPPAAR
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	551				600
celr3_human	AAAAAAFEID	PRSGLISTSG	RVDREHMESY	ELVVEASDQG	QEPGPRSATV
q5y190_human	AAAAAAFEID	PRSGLISTSG	RVDREHMESY	ELVVEASDQG	QEPGPRSATV
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	601				650
celr3_human	RVHITVLDEN	DNAPQFSEKR	YVAQVREDVR	PHTVVLRVTA	TDRDKDANGL
q5y190_human	RVHITVLDEN	DNAPQFSEKR	YVAQVREDVR	PHTVVLRVTA	TDRDKDANGL
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	651				700
celr3_human	VHYNIISGNS	RGHFAIDSLT	GEIQVVAPLD	FEAEREYALR	IRAQDAGRPP
q5y190_human	VHYNIISGNS	RGHFAIDSLT	GEIQVVAPLD	FEAEREYALR	IRAQDAGRPP
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	701				750
celr3_human	LSNNTGLASI	QVVDINDHIP	IFVSTPFQVS	VLENAPLGHS	VIHIQAVDAD
q5y190_human	LSNNTGLASI	QVVDINDHIP	IFVSTPFQVS	VLENAPLGHS	VIHIQAVDAD
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	751				800
celr3_human	HGENARLEYS	LTGVAPDTPF	VINSATGWVS	VSGPLDRESV	EHYFFGVEAR
q5y190_human	HGENARLEYS	LTGVAPDTPF	VINSATGWVS	VSGPLDRESV	EHYFFGVEAR
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	801				850
celr3_human	DHGSPPLSAS	ASVTVTVLDV	NDNRPEFTMK	EYHLRLNEDA	AVGTSVVSVT
q5y190_human	DHGSPPLSAS	ASVTVTVLDV	NDNRPEFTMK	EYHLRLNEDA	AVGTSVVSVT
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	851				900
celr3_human	AVDRDANSAI	SYQITGGNTR	NRFAISTQGG	VGLVTLALPL	DYKQERYFKL
q5y190_human	AVDRDANSAI	SYQITGGNTR	NRFAISTQGG	VGLVTLALPL	DYKQERYFKL
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~

Figure S1. Cont.

	901				950
celr3_human	VLASDRALH	DHCYVHINIT	DANTHRPVFQ	SAHYSVSVNE	DRPMGSTIVV
q5y190_human	VLASDRALH	DHCYVHINIT	DANTHRPVFQ	SAHYSVSVNE	DRPMGSTIVV
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	951				1000
celr3_human	ISASDDDVGE	NARITYLLED	NLPQFRIDAD	SGAITLQAPL	DYEDQVTTYTL
q5y190_human	ISASDDDVGE	NARITYLLED	NLPQFRIDAD	SGAITLQAPL	DYEDQVTTYTL
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1001				1050
celr3_human	AITARDNGIP	QKADTTYVEV	MVNDVNDNAP	QFVASHYTGL	VSEDAPPFTS
q5y190_human	AITARDNGIP	QKADTTYVEV	MVNDVNDNAP	QFVASHYTGL	VSEDAPPFTS
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1051				1100
celr3_human	VLQISATDRD	AHANGRVQYT	FQNGEDGDGD	FTIEPTSGIV	RTVRRLDREA
q5y190_human	VLQISATDRD	AHANGRVQYT	FQNGEDGDGD	FTIEPTSGIV	RTVRRLDREA
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1101				1150
celr3_human	VSVYELTAYA	VDRGVPPLRT	PVSIQVMVQD	VNDNAPVFPA	EEFEVRVKEN
q5y190_human	VSVYELTAYA	VDRGVPPLRT	PVSIQVMVQD	VNDNAPVFPA	EEFEVRVKEN
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1151				1200
celr3_human	SIVGSVVAQI	TAVDPDEGPN	AHIMYQIVEG	NIPELFQMDI	FSGELTALID
q5y190_human	SIVGSVVAQI	TAVDPDEGPN	AHIMYQIVEG	NIPELFQMDI	FSGELTALID
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1201				1250
celr3_human	LDYEARQEYV	IVVQATSAPL	VSRATVHVRL	VDQNDNSPVL	NNFQILFNKY
q5y190_human	LDYEARQEYV	IVVQATSAPL	VSRATVHVRL	VDQNDNSPVL	NNFQILFNKY
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1251				1300
celr3_human	VSNRSDTFPS	GIIGRIPAYD	PDVSDHLFYS	FERGNELQLL	VVNQTSGELR
q5y190_human	VSNRSDTFPS	GIIGRIPAYD	PDVSDHLFYS	FERGNELQLL	VVNQTSGELR
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1301				1350
celr3_human	LSRKLNNRP	LVASMLVTVT	DGLHSVTAQC	VLRVVIITEE	LLANSLTVRL
q5y190_human	LSRKLNNRP	LVASMLVTVT	DGLHSVTAQC	VLRVVIITEE	LLANSLTVRL
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1351				1400
celr3_human	ENMWQERFLS	PLLGRFLEGV	AAVLATPAED	VFIFNIQNDT	DVGGTVLNVS
q5y190_human	ENMWQERFLS	PLLGRFLEGV	AAVLATPAED	VFIFNIQNDT	DVGGTVLNVS
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1401				1450
celr3_human	FSALAPRGAG	AGAAGPWFSS	EELQEQLYVR	RAALAARSL	DVLPFDDNVC
q5y190_human	FSALAPRGAG	AGAAGPWFSS	EELQEQLYVR	RAALAARSL	DVLPFDDNVC
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~

Figure S1. Cont.

	1451				1500
celr3_human	LREPCENYMK	CVSVLRFDSS	APFLASASTL	FRPIQPIAGL	RCRCPPGFTG
q5y190_human	LREPCENYMK	CVSVLRFDSS	APFLASASTL	FRPIQPIAGL	RCRCPPGFTG
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1501				1550
celr3_human	DFCETELDLC	YSNPCRNGGA	CARREGGYTC	VCRPRFTGED	CELDTEAGRC
q5y190_human	DFCETELDLC	YSNPCRNGGA	CARREGGYTC	VCRPRFTGED	CELDTEAGRC
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1551				1600
celr3_human	VPGVCRNGGT	CTDAPNGGFR	CQCPAGGAFE	GPRCEVAARS	FPPSSFVMFR
q5y190_human	VPGVCRNGGT	CTDAPNGGFR	CQCPAGGAFE	GPRCEVAARS	FPPSSFVMFR
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1601				1650
celr3_human	GLRQRFHLLT	SLSFATVQQS	GLLFYNGRLN	EKHDFLALEL	VAGQVRLTYS
q5y190_human	GLRQRFHLLT	SLSFATVQQS	GLLFYNGRLN	EKHDFLALEL	VAGQVRLTYS
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1651				1700
celr3_human	TGESNTVVSP	TVPGGLSDGQ	WHTVHLRYYN	KPRTDALGGA	QGPSKDKVAV
q5y190_human	TGESNTVVSP	TVPGGLSDGQ	WHTVHLRYYN	KPRTDALGGA	QGPSKDKVAV
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1701				1750
celr3_human	LSVDDCDVAV	ALQFGAEIGN	YSCAAAGVQT	SSKKSLLDTG	PLLLGGVPNL
q5y190_human	LSVDDCDVAV	ALQFGAEIGN	YSCAAAGVQT	SSKKSLLDTG	PLLLGGVPNL
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1751				1800
celr3_human	PENFPVSHKD	FIGCMRDLHI	DGRRVDMAAF	VANNGTMAGC	QAKLHFCDSDG
q5y190_human	PENFPVSHKD	FIGCMRDLHI	DGRRVDMAAF	VANNGTMAGC	QAKLHFCDSDG
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1801				1850
celr3_human	PCKNSGFCSE	RWGSFSCDCP	VGFGGKDCQL	TMAHPHHFRG	NGTLSWNFGS
q5y190_human	PCKNSGFCSE	RWGSFSCDCP	VGFGGKDCQL	TMAHPHHFRG	NGTLSWNFGS
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1851				1900
celr3_human	DMAVSVPWYL	GLAFRTRATQ	GVLMQVQAGP	HSTLLCQLDR	GLLSVTVTRG
q5y190_human	DMAVSVPWYL	GLAFRTRATQ	GVLMQVQAGP	HSTLLCQLDR	GLLSVTVTRG
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1901				1950
celr3_human	SGRASHLLLD	QVTVSDGRWH	DLRLELQEEP	GRRRGHHVLM	VSLDFSLFQD
q5y190_human	SGRASHLLLD	QVTVSDGRWH	DLRLELQEEP	GRRRGHHVLM	VSLDFSLFQD
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	1951				2000
celr3_human	TMAVGSELQG	LKVKQLHVGG	LPPGSAEEAP	QGLVGCIQGV	WLGSTPSGSP
q5y190_human	TMAVGSELQG	LKVKQLHVGG	LPPGSAEEAP	QGLVGCIQGV	WLGSTPSGSP
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2001				2050
celr3_human	ALLPPSHRVN	AEPGCVVTNA	CASGPCPPHA	DCRDLWQTFS	CTCQPGYYGP
q5y190_human	ALLPPSHRVN	AEPGCVVTNA	CASGPCPPHA	DCRDLWQTFS	CTCQPGYYGP
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2051				2100
celr3_human	GCVDACLNP	CQNQGSCRHL	PGAPHGYTCD	CVGGYFGHHC	EHRMDQQCPR
q5y190_human	GCVDACLNP	CQNQGSCRHL	PGAPHGYTCD	CVGGYFGHHC	EHRMDQQCPR
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~

Figure S1. Cont.

	2101				2150
celr3_human	GWGWSPTCGP	CNCDVHKGFD	PNCNKTNGQC	HCKEFHYRPR	GSDSCLPCDC
q5y190_human	GWGWSPTCGP	CNCDVHKGFD	PNCNKTNGQC	HCKEFHYRPR	GSDSCLPCDC
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2151				2200
celr3_human	YPVGSTSRSC	APHSGQCPCR	PGALGRQCNS	CDSPFPAEVTA	SGCRVLYDAC
q5y190_human	YPVGSTSRSC	APHSGQCPCR	PGALGRQCNS	CDSPFPAEVTA	SGCRVLYDAC
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2201				2250
celr3_human	PKSLRSGVWW	PQTKFGVLAT	VPCPRGALGA	AVRLCDEAQQ	WLEPDLFNCT
q5y190_human	PKSLRSGVWW	PQTKFGVLAT	VPCPRGALGA	AVRLCDEAQQ	WLEPDLFNCT
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2251				2300
celr3_human	SPAFRELSLL	LDGLELNKTA	LDTMEAKKLA	QRLREVTGHT	DHYFSQDVRV
q5y190_human	SPAFRELSLL	LDGLELNKTA	LDTMEAKKLA	QRLREVTGHT	DHYFSQDVRV
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2301				2350
celr3_human	TARLLAHLA	FESHQQGFGL	TATQDAHFNE	NLLWAGSALL	APETGDLWAA
q5y190_human	TARLLAHLA	FESHQQGFGL	TATQDAHFNE	NLLWAGSALL	APETGDLWAA
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2351				2400
celr3_human	LGQRAPGGSP	GSAGLVRHLE	EYAATLARNM	ELTYLNPML	VTPNIMLSID
q5y190_human	LGQRAPGGSP	GSAGLVRHLE	EYAATLARNM	ELTYLNPML	VTPNIMLSID
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2401				2450
celr3_human	RMEHPSSPRG	ARRYPRYHSN	LFRGQDAWDP	HTHVLLPSQS	PRPSPSEVLP
q5y190_human	RMEHPSSPRG	ARRYPRYHSN	LFRGQDAWDP	HTHVLLPSQS	PRPSPSEVLP
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2451				2500
celr3_human	TSSSIENSTT	SSVPPPAPP	EPEPGISIII	LLVYRTLGL	LPAQFQAERR
q5y190_human	TSSSIENSTT	SSVPPPAPP	EPEPGISIII	LLVYRTLGL	LPAQFQAERR
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2501				2550
celr3_human	GARLPQNPVM	NSPVVSVAVF	HGRNFLRGIL	ESPISLEFRL	LQTANRSKAI
q5y190_human	GARLPQNPVM	NSPVVSVAVF	HGRNFLRGIL	ESPISLEFRL	LQTANRSKAI
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2551				2600
celr3_human	CVQWDPPGLA	EQHGVWTARD	CELVHRNGSH	ARCRCSTRGT	FGVLMDASPR
q5y190_human	CVQWDPPGLA	EQHGVWTARD	CELVHRNGSH	ARCRCSTRGT	FGVLMDASPR
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2601				2650
celr3_human	ERLEGDLELL	AVFTHVVAV	SVAALVLTAA	ILLSLRSLKS	NVRGIHANVA
q5y190_human	ERLEGDLELL	AVFTHVVAV	SVAALVLTAA	ILLSLRSLKS	NVRGIHANVA
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2651				2700
celr3_human	AALGVAELLF	LLGIHRTHNQ	.....	.....	.....LVC
q5y190_human	AALGVAELLF	LLGIHRTHNQ	VQDQGGQTCV	LMTLLAQEAW	GQNSGSELVC
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~

Figure S1. Cont.

	2701				2750
celr3_human	TAVAILLHYF	FLSTFAWLFV	QGLHLYRMQV	EPRNVDRGAM	RFYHALGWGV
q5y190_human	TAVAILLHYF	FLSTFAWLFV	QGLHLYRMQV	EPRNVDRGAM	RFYHALGWGV
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2751				2800
celr3_human	PAVLLGLAVG	LDPEGYGNPD	FCWISVHEPL	IWSFAGPVVL	VIVMNGTMFL
q5y190_human	PAVLLGLAVG	LDPEGYGNPD	FCWISVHEPL	IWSFAGPVVL	VIVMNGTMFL
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2801				2850
celr3_human	LAARTSCSTG	QREAKKTSAL	.TLRSSFLLL	LLVSASWLFV	LLAVNHSILA
q5y190_human	LAARTSCSTG	QREAKKTSAL	RTLRSSFLLL	LLVSASWLFV	LLAVNHSILA
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2851				2900
celr3_human	FHYLHAGLCG	LQGLAVLLLF	CVLNADARAA	WMPACLGRKA	APEEARPAPG
q5y190_human	FHYLHAGLCG	LQGLAVLLLF	CVLNADARAA	WMPACLGRKA	APEEARPAPG
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2901				2950
celr3_human	LGPGAYNNTA	LFEESGLIRI	TLGASTVSSV	SSARSGRTQD	QDSQRGRSYL
q5y190_human	LGPGAYNNTA	LFEESGLIRI	TLGASTVSSV	SSARSGRTQD	QDSQRGRSYL
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	2951				3000
celr3_human	RDNVLRHGS	AADHTDHSLO	AHAGPTDLDV	AMFHRDAGAD	SDSDSDLSLE
q5y190_human	RDNVLRHGS	AADHTDHSLO	AHAGPTDLDV	AMFHRDAGAD	SDSDSDLSLE
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	3001				3050
celr3_human	EERSLSIPSS	ESEDNGRTRG	RFQRPLCRAA	QSERLLTHPK	DVDGNDLLSY
q5y190_human	EERSLSIPSS	ESEDNGRTRG	RFQRPLCRAA	QSERLLTHPK	DVDGNDLLSY
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	3051				3100
celr3_human	WPALGECEAA	PCALQTWGSE	RRLGLDTSKD	AANNNQPDPA	LTSGDETSLG
q5y190_human	WPALGECEAA	PCALQTWGSE	RRLGLDTSKD	AANNNQPDPA	LTSGDETSLG
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	3101				3150
celr3_human	RAQRQRKGIL	KNRLQYPLVP	QTRGAPELSW	CRAATLGHRA	VPAASYGRIY
q5y190_human	RAQRQRKGIL	KNRLQYPLVP	QTRGAPELSW	CRAATLGHRA	VPAASYGRIY
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	3151				3200
celr3_human	AGGGTGSLSQ	PASRYSSREQ	LDLLLRQLS	RERLEEAPAP	VLRPLSRPGS
q5y190_human	AGGGTGSLSQ	PASRYSSREQ	LDLLLRQLS	RERLEEAPAP	VLRPLSRPGS
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	3201				3250
celr3_human	QECMDAAPGR	LEPKDRGSTL	PRRQPPRDYP	GAMAGRFGSR	DALDLGAPRE
q5y190_human	QECMDAAPGR	LEPKDRGSTL	PRRQPPRDYP	GAMAGRFGSR	DALDLGAPRE
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	3251				3300
celr3_human	WLSTLPPPRR	TRDLDPQPPP	LPLSPQRQLS	RDPLLPSRPL	DSLRSRNSR
q5y190_human	WLSTLPPPRR	TRDLDPQPPP	LPLSPQRQLS	RDPLLPSRPL	DSLRSRNSR
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~

Figure S1. Cont.

	3301				3350
celr3_human	EQLDQVPSRH	PSREALGPLP	QLLRAREDSV	SGPSHGPSTE	QLDILSSILA
q5y190_human	EQLDQVPSRH	PSREALGPLP	QLLRAREDSV	SGPSHGPSTE	QLDILSSILA
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
	3351				3400
celr3_human	SFNSSALSSV	QSSSTPLGPH	TTATPSATAS	VLGPSTPRSA	TSHSISELSP
q5y190_human	SFNSSALSSV	QSSSTPLGPH	TTATPSATAS	VLGPSTPRSA	TSHSISELSP
s26a6_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~MGLA.
	3401				3450
celr3_human	DSEVPRSEGH	S~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	DSE.PRDTQA	LLSATQAMD	RRRDYHMERP	LLNQEHLLEL	GRWGSAPRTH
s26a6_human	DASGPRDTQA	LLSATQAMD	RRRDYHMERP	LLNQEHLLEL	GRWGSAPRTH
	3451				3500
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	QWRTWLQCSR	ARAYALLLQH	LPVLVWLPY	PVRDWLLGDL	LSGLSVAIMQ
s26a6_human	QWRTWLQCSR	ARAYALLLQH	LPVLVWLPY	PVRDWLLGDL	LSGLSVAIMQ
	3501				3550
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	LPQGLAYALL	AGLPPVFGLY	SSFYPVFIYF	LFGTSRHISV	ESLCVPGPVD
s26a6_human	LPQGLAYALL	AGLPPVFGLY	SSFYPVFIYF	LFGTSRHISV	.....
	3551				3600
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	TGTFAVMSVM	VGSVTESLAP	QALNDSMINE	TARDAARVQV	ASTLSVLVGL
s26a6_human	.GTFAVMSVM	VGSVTESLAP	QALNDSMINE	TARDAARVQV	ASTLSVLVGL
	3601				3650
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	FQVGLGLIHF	GFVVTYLSEP	LVRGYTTAAA	VQVFVSQLKY	VFGLHLSSH
s26a6_human	FQVGLGLIHF	GFVVTYLSEP	LVRGYTTAAA	VQVFVSQLKY	VFGLHLSSH
	3651				3700
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	GPLSLIYTVL	EVCWKLPQSK	VGTVVTAAVA	GVVLVVVKLL	NDKLQQQLPM
s26a6_human	GPLSLIYTVL	EVCWKLPQSK	VGTVVTAAVA	GVVLVVVKLL	NDKLQQQLPM
	3701				3750
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	PIPGELLTLI	GATGISYGMG	LKHRFEVDVV	GNIPAGLVPP	VAPNTQLFSK
s26a6_human	PIPGELLTLI	GATGISYGMG	LKHRFEVDVV	GNIPAGLVPP	VAPNTQLFSK
	3751				3800
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	LVGSAFTIAV	VGFAIAISLG	KIFALRHGYR	VDSNQELVAL	GLSNLIGGIF
s26a6_human	LVGSAFTIAV	VGFAIAISLG	KIFALRHGYR	VDSNQELVAL	GLSNLIGGIF
	3801				3850
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	QCFFVSCSMS	RSLVQESTGG	NSQVAGAISS	LFILLIIVKL	GELFHDLPKA
s26a6_human	QCFFVSCSMS	RSLVQESTGG	NSQVAGAISS	LFILLIIVKL	GELFHDLPKA
	3851				3900
celr3_human	~~~~~	~~~~~	~~~~~	~~~~~	~~~~~
q5y190_human	VLAIIIVNL	KGMLRQLSDM	RSLWKANRAD	LLIWLVTFTA	TILLNLDLGL
s26a6_human	VLAIIIVNL	KGMLRQLSDM	RSLWKANRAD	LLIWLVTFTA	TILLNLDLGL

Figure S1. Cont.

```

3901                                     3950
celr3_human ~~~~~
q5y190_human VVAVIFSLLL VVVRTQMPHY SVLGQVPD TD IYRDVAEYSE AKEVRGVKVF
s26a6_human VVAVIFSLLL VVVRTQMPHY SVLGQVPD TD IYRDVAEYSE AKEVRGVKVF

3951                                     4000
celr3_human ~~~~~
q5y190_human RSSATVYFAN AEFYSDALKQ RCGVDVDFLI SQKKKLLKKQ EQLK LKQLQK
s26a6_human RSSATVYFAN AEFYSDALKQ RCGVDVDFLI SQKKKLLKKQ EQLK LKQLQK

4001                                     4050
celr3_human ~~~~~
q5y190_human EEKLRKQAGP LLSACLA... ..PQQV SSGDKMEDAT
s26a6_human EEKLRKQAAS PKGASVSINV NTSLED MRSN NVEDCKMMQV SSGDKMEDAT

4051                                     4100
celr3_human ~~~~~
q5y190_human ANGQEDSKAP DGSTL KALGL PQPDFHSLIL DLGALS FVDT VCLKSLKNIF
s26a6_human ANGQEDSKAP DGSTL KALGL PQPDFHSLIL DLGALS FVDT VCLKSLKNIF

4101                                     4150
celr3_human ~~~~~
q5y190_human HDFREIEVEV YMAACHSPV V SQLEAGHF FD ASITKKHLFA SVHDAVTFAL
s26a6_human HDFREIEVEV YMAACHSPV V SQLEAGHF FD ASITKKHLFA SVHDAVTFAL

4151                                     4200
celr3_human ~~~~~
q5y190_human QHPRFVPD SP VSPSLAVSSD VKQLEPELLL RNNLLSGIPE KVQGSV GANG
s26a6_human QHPRFVPD SP VSVTRL~~~~ ~~~~~

4201
celr3_human ~~~~~
q5y190_human QSLEDTE
s26a6_human ~~~~~

```

Figure S2. Alignment of MSRE\_HUMAN, MSRE\_RABIT, MSRE\_BOVIN, MSRE\_MOUSE with SCARA5\_XENTR. Note that the predicted sequence SCARA5\_XENTR is incomplete: the N-terminal region where macrophage scavenger receptor types I and II contain a PF03523 motif is missing from this incomplete sequence.

```

1                                     50
msre_human ~~~~MEQWDH F.HNQ QEDTD SCSESVK FDA RSMTALLPPN PKNSPSLQEK
enst00000262101 ~~~~MEQWDH F.HNQ QEDTD SCSESVK FDA RSMTALLPPN PKNSPSLQEK
msre_rabit ~~~~MAQWDS F.TDQ QEDTD SCSESVK FDA RSNTALLPPN PKNGPPLQEK
msre_bovin ~~~~MAQWDD F.PDQ QEDTD SCTESVK FDA RSVTALLPPH PKNGPTLQER
msre_mouse MTKEMTENQR LCPHEREDAD CSSESVK FDA RSMTASLPHS TKNGPSVQEK
pf03523 ~~~~~
scara5_xentr ~~~~~

51                                     100
msre_human LKSFKAALIA LYLLVFAVLI PLIGIVAAQL LKWETKNCSV SSTNANDITQ
enst00000262101 LKSFKAALIA LYLLVFAVLI PLIGIVAAQL LKWETKNCSV SSTNANDITQ
msre_rabit LKSFKAALIA LYLLVFAVLI PIIAIMAAQL LKWEMKNCTV G SINANSVSS
msre_bovin MKSYKTALIT LYLLVFAVLI PIIGIVAAQL LKWETKNCTV GSVNA.DISP
msre_mouse LKSFKAALIA LYLLVFAVLI PVVGIVTAQL LNWEMKNCLV CSRNTSDTSQ
pf03523 ~~~~~
scara5_xentr ~~~~~

101                                    150
msre_human SLTGKGN DSE EEMRFQEVFM EHMSNMEKRI QHILDMEANL MDTEHFQNF S
enst00000262101 SLTGKGN DSE EEMRFQEVFM EHMSNMEKRI QHILDMEANL MDTEHFQNF S
msre_rabit SLLGRGN DSE HEVRFREVM EHISKMEKRI QYISDTEENL VDSEHFQNF S
msre_bovin SPEGKGN GSE DEMRFREAVM ERMSNMESRI QYLSDN EANL L DAKNFQNF S
msre_mouse GPMEKENTSN VEMRF.TIIM AHMKDMEERI QSISNSKADL IDTGRFQNF S
pf03523 ~~~~~~MEXRI QYxSDxEANL xDxxx FQNF S
scara5_xentr ~~~~~~MEXRI QYxSDxEANL xDxxx FQNF S

```

```

151
msre_human MTTDQRFNDI LLQLSTLFSS VQGHGNAIDE ISKSLISLNT TLLDLQLNIE
enst00000262101 MTTDQRFNDI LLQLSTLFSS VQGHGNAIDE ISKSLISLNT TLLDLQLNIE
msre_rabbit VTTDQRFADV LLQLSTLVPT VQGHGNAVDE ITRSLISLNT TLLDLHLHYVE
msre_bovin ITTDQRFNDV LFQLNSLLSS IQEHENIIGD ISKSLVGLNT TVLDDLQFSIE
msre_mouse MATDQRLNDI LLQLNSLILS VQEHGNSLDA ISKSLQSLNM TLLDVLQHTE
pf03523 xTTDQRxnDV LxQLNSLxSS xQEH~~~~~ ~~~~~~ ~~~~~~
scara5_xentr ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~

201
msre_human NLNGKIQENT FKQEEISKL EERVYNVSAE IMAMKEEQVH LEQEIKGEVK
enst00000262101 NLNGKIQENT FKQEEISKL EERVYNVSAE IMAMKEEQVH LEQEIKGEVK
msre_rabbit TLNVKIQENT LKQEEISKL KERVHNASAE IMSMKEEQVH LEQEIKREVH
msre_bovin TLNQRVQENA FKQEEEMRKL EERIYNASAE IKSLDEKQVY LEQEIKGEMK
msre_mouse TLHVRVREST AKQEDISKL EERVYKVS AE VQSVKEEQAH VEQEVKQEVH
pf03523 ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~
scara5_xentr ~~~~~~ ~~~~~~ ~~~~~~NTSTE IAGVRQECMT LEKEMKEEVK

251
msre_human VLNNITNDLR LKDWEHSQTL RNITLIQPPP GPPGEKGDGR PTGESGPRGF
enst00000262101 VLNNITNDLR LKDWEHSQTL RNITLIQPPP GPPGEKGDGR PTGESGPRGF
msre_rabbit VLNNITNDLR LKDWEHSQTL RNITLIQPPP GPPGEKGDGR PTGESGPPGV
msre_bovin LLNNITNDLR LKDWEHSQTL KNITLLQPPP GPPGEKGDGR PPGQNGIPGF
msre_mouse VLNNITNDLR LKDWEHSQTL KNITFIQPPP GPQGEKGDGR LTGQTGPPGA
pf03523 ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~
scara5_xentr TLNKITNDLR LKDWEKAFIL NNLTSIQPPP GPKGEKGDGT VKGDTGVSGF

301
msre_human PGPIGPPGLK GDRGAIGFPG SRGLPGYAGR PGNS...GPK GQKGEKGSNG
enst00000262101 PGPIGPPGLK GDRGAIGFPG SRGLPGYAGR PGNS...GPK GQKGEKGSNG
msre_rabbit PGPVGPGLK GDRGSIGFPG SRGYPGQSGK TGRTGYPPGPK GQKGEKGSNG
msre_bovin PGLIGTPGLK GDRGISGLPG VRGFPMPGK TGKPLNGQK GQKGEKGSNG
msre_mouse PGIRGIPGVK GDRGQIGFPG GRGNPGAPGK PGRSGSPGPK GQKGEKGSNG
pf03523 ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~
scara5_xentr QGIRGFPGSK GEKQNGYLG SKGAPGIP...VSIGLK GQKGEKGEDA

351
msre_human TLTPTFK... ..VRLVGGSG PHEGRVEILH SGQWGTICDD RWEVRVQVV
enst00000262101 TLTPTFK... ..VRLVGGSG PHEGRVEILH SGQWGTICDD RWEVRVQVV
msre_rabbit ILTPSAT... ..VRLVGGSG PHEGRVEILH NGQWGTVCDD HWELRAGVV
msre_bovin MQRQSNT... ..VRLVGGSG PHEGRVEIFH EGQWGTVCDD RWELRGLVV
msre_mouse GSTPLKT... ..VRLVGGSG AHEGRVEIFH QGQWGTICDD RWDIRAGVV
pf03523 ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~
scara5_xentr ATTPSRTTTT PLVRLVGGSS SLLGRVEVFH NGEWGTVCDD HWGLEDGKVV

401
msre_human CRSLGYPGVQ AVHKAHFGQ .GTGPIWLNE VFCFGRESSI EECKIRQWGT
enst00000262101 CRSLGYPGVQ AVHKAHFGQ .GTGPIWLNE VFCFGRESSI EECKIRQWGT
msre_rabbit CRSLGYRGVK SVHKKAYFGQ .GTGPIWLNE VPCLGMESSI EECKIRQWV
msre_bovin CRSLGYKGVQ SVHKRAYFGK .GTGPIWLNE VFCFGKESSI EECKIRQWV
msre_mouse CRSLGYQEV L AVHKRAHFGQ .GTGPIWLNE VMCFGRESSI ENCKINQWV
pf03523 ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~ ~~~~~~
scara5_xentr CKMLGYTGVA QVYVNAYFGQ AGLARIWMDD VQCNGIEKSI FECKFKGWGI

451 468
msre_human RACSHSEDAG VTCTL~~~
enst00000262101 RACSHSEDAG VTCTL~~~
msre_rabbit RVCSHGEDAG VTCTL~~~
msre_bovin RACSHDEDAG VTCTT~~~
msre_mouse LSCSHSEDAG VTCTS~~~
pf03523 ~~~~~~ ~~~~~~
scara5_xentr TNCRHTEDAG VRCVAYIP

```