

Article

Differential Effects of *ABCG5/G8* Gene Region Variants on Lipid Profile, Blood Pressure Status, and Gallstone Disease History in Taiwan

Ming-Sheng Teng ^{1,†}, Kuan-Hung Yeh ^{2,3,†}, Lung-An Hsu ⁴, Hsin-Hua Chou ^{2,3}, Leay-Kiaw Er ^{3,5}, Semon Wu ⁶ and Yu-Lin Ko ^{1,2,3,*}

¹ Department of Research, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei City 23142, Taiwan

² Cardiovascular Center and Division of Cardiology, Department of Internal Medicine, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei City 23142, Taiwan

³ School of Medicine, Tzu Chi University, Hualien 97004, Taiwan

⁴ The First Cardiovascular Division, Department of Internal Medicine, Chang Gung Memorial Hospital and Chang Gung University College of Medicine, Taoyuan 33305, Taiwan

⁵ The Division of Endocrinology and Metabolism, Department of Internal Medicine, Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, New Taipei City 23142, Taiwan

⁶ Department of Life Science, Chinese Culture University, Taipei 11114, Taiwan

* Correspondence: yulinkotw@yahoo.com.tw; Tel.: +886-2-6628-9779 (ext. 5355); Fax: +886-2-6628-9009

† These authors contributed equally to this work.

Abstract: *ABCG5* and *ABCG8* are two key adenosine triphosphate-binding cassette (ABC) proteins that regulate whole-body sterol trafficking. This study aimed to elucidate the association between *ABCG5/G8* gene region variants and lipid profile, cardiometabolic traits, and gallstone disease history in Taiwan. A total of 1494 Taiwan Biobank participants with whole-genome sequencing data and 117,679 participants with Axiom Genome-Wide CHB Array data were enrolled for analysis. Using genotype–phenotype and stepwise linear regression analyses, we found independent associations of four Asian-specific *ABCG5* variants, rs119480069, rs199984328, rs560839317, and rs748096191, with total, low-density lipoprotein (LDL), and non-high-density lipoprotein (HDL) cholesterol levels (all $p \leq 0.0002$). Four other variants, which were in nearly complete linkage disequilibrium, exhibited genome-wide significant associations with gallstone disease history, and the *ABCG8* rs11887534 variant showed a trend of superiority for gallstone disease history in a nested logistic regression model ($p = 0.074$). Through regional association analysis of various other cardiometabolic traits, two variants of the *PLEKHH2*, approximately 50 kb from the *ABCG5/G8* region, exhibited significant associations with blood pressure status ($p < 10^{-6}$). In conclusion, differential effects of *ABCG5/G8* region variants were noted for lipid profile, blood pressure status, and gallstone disease history in Taiwan. These results indicate the crucial role of individualized assessment of *ABCG5/G8* variants for different cardiometabolic phenotypes.

Keywords: *ABCG5*; *ABCG8*; genetic variants; gallstone disease; lipid profile; differential effect



Citation: Teng, M.-S.; Yeh, K.-H.; Hsu, L.-A.; Chou, H.-H.; Er, L.-K.; Wu, S.; Ko, Y.-L. Differential Effects of *ABCG5/G8* Gene Region Variants on Lipid Profile, Blood Pressure Status, and Gallstone Disease History in Taiwan. *Genes* **2023**, *14*, 754. <https://doi.org/10.3390/genes14030754>

Academic Editors: Albert Jeltsch and Marc Via

Received: 20 February 2023

Revised: 15 March 2023

Accepted: 16 March 2023

Published: 20 March 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The adenosine triphosphate (ATP)-binding cassette (ABC) family contains more than 40 ABC transporters in seven subfamilies (ABCA to ABCG) and is one of the largest transporter families. This family couples ATP binding, hydrolysis, and phosphate release to accomplish translocation of diverse substrates across membranes [1,2]. With their involvement in endothelial dysfunction, cholesterol homeostasis, regulation of blood pressure, vascular inflammation, and platelet aggregation, ABC transporters are crucial in the pathogenesis of atherosclerotic vascular diseases [3,4]. *ABCG5* and *ABCG8* are two functional ABC proteins that mediate the efflux of xenosterols from hepatocytes and enterocytes

and prevent xenosterol from accumulating in the body [3,5]. ABCG5 and ABCG8 are half-transporter heterodimers that affect bile cholesterol excretion and intestinal cholesterol absorption rates [6]. The *ABCG5* and *ABCG8* genes (*ABCG5/G8*) are highly expressed in the livers and small intestines of both humans and mice [7–10]. In humans, mutations of *ABCG5/G8* may cause autosomal recessive sitosterolemia [11–13]. In animals, as determined through quantitative trait locus linkage analysis, *Abcg5/g8* has been identified as the mouse gallstone gene, *Lith9* [14–18]. In mice, low biliary cholesterol concentrations may develop through the disruption of *ABCG5*, *ABCG8*, or both [19–21]. Conversely, a more than fivefold increase in biliary cholesterol levels can be caused by *ABCG5* and *ABCG8* overexpression [22]. In analyses of human and animal models, *ABCG5/G8* have also been shown to affect various cardiometabolic traits and disorders, such as lipid and glucose metabolism, blood pressure control, metabolic syndrome, and fatty liver disease [23–30].

ABCG5 and *ABCG8* (*ABCG5/G8*) are coregulated at the transcription level through their sharing of a common bidirectional promoter and their location next to each other on chromosome 2p21 [12]. Exome sequence analysis of 60,706 individuals of diverse ancestries revealed 33 and 36 exome sequences predicted loss-of-function variants for *ABCG5* and *ABCG8*, respectively (<https://gnomad.broadinstitute.org/> v3.1.2, accessed on 18 October 2022) [31]. By contrast, to date, 769 and 978 missense variants have been catalogued in the database of SNP according to the PUBMed website ([PUBMed.gov](https://pubmed.ncbi.nlm.nih.gov/)) for *ABCG5* and *ABCG8*, respectively. Most of the missense variants are predicted to be benign, whereas the majority of dysfunctional alleles in selected likely pathogenic *ABCG5/G8* missense mutants are dysfunctional due to their inability to heterodimerize ABCG5 and ABCG8 and traffic beyond the endoplasmic reticulum [32]. Although familial sitosterolemia is a rare Mendelian recessive disorder, with the affected individuals typically having homozygous loss-of-function variants in the *ABCG5/G8* genes, heterozygous *ABCG5* gene deficiency has been shown to be associated with increased sitosterol and LDL cholesterol levels and increased risk of coronary artery disease [33]. Elevation of sitosterol serum concentrations due to *ABCG5/G8* mutations also showed risk-increasing causal relationships with a detrimental effect on coronary atherosclerosis [34]. These results suggested the critical role of elucidating novel *ABCG5/G8* mutations in preventive medicine.

Ethnic genetic heterogeneity for *ABCG5/G8* variants has been widely reported [11,12,33–37], and the role and differential effects of *ABCG5/G8* variants on lipid profile, cardiometabolic traits, and gallstone disease history in Asian populations have not been fully elucidated. The evolution of geographically dispersed populations is affected by factors such as the founder effect and evolutionary selection, resulting in genetic drift and ethnic heterogeneity in genetic architectures [38]. The Taiwan Biobank (TWB) is a population-based cohort study sponsored by the Taiwanese government and has enrolled more than 150,000 individuals aged between 30 and 70 years without a history of cancer [39,40]. By combining both regional association analysis and candidate variant approaches, we have previously shown the crucial role of ethnicity-specific variants on genetic determinants of lipid profiles [41–43]. In this study, we investigated the associations of *ABCG5/G8* variants with lipid profile, cardiometabolic traits, and gallstone disease history in Taiwanese individuals who were participants in the TWB.

2. Subjects and Methods

2.1. TWB Population-Based Cohort Study

The cohort of TWB participants for the current study was composed of 129,542 participants who had Axiom Genome-Wide CHB Array data and were recruited in centers across Taiwan between 2008 and 2020. In total, 11,863 participants were excluded according to the following criteria: quality control (QC) for the array data with identity by descent score >0.187 to remove cryptic relatedness (7216) and fasting for <6 h (4647). We also performed ultrafast whole-genome secondary analysis of 1478 TWB participants who had whole-genome sequencing (WGS) data, using the Illumina sequencing platform to search for candidate variants within the coding and promoter regions of *ABCG5/G8* [44]. The flowchart of participant recruitment is shown in Figure 1. Participants with a history of

hyperlipidemia (8799) were excluded from the analyses of lipid profile. In a questionnaire, participants were asked whether they or either of their parents had ever received a clinical diagnosis of gallstone disease. Ethical approval was granted by the institutional review boards of Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation (approval number: 08-XD-005) and the Ethics and Governance Council of the TWB (approval number: TWBR11011-02). All participants provided written informed consent.

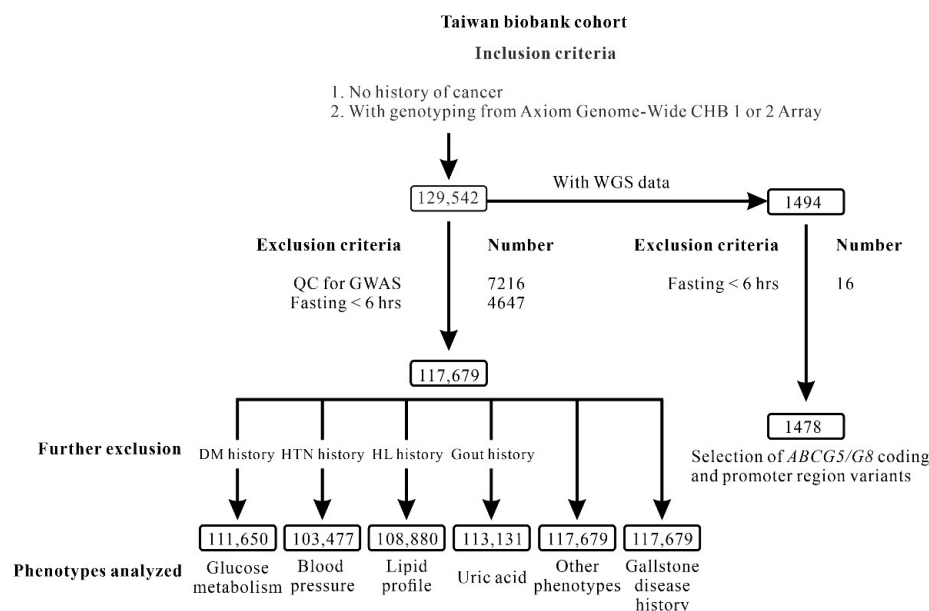


Figure 1. Study flowchart of inclusion and exclusion criteria used to screen Taiwan Biobank project participants. The number of participants analyzed were shown in each box.

2.2. Clinical and Laboratory Examinations

Data on baseline characteristics, including age, sex, body mass index (BMI), and smoking status, were collected. Total, low-density lipoprotein (LDL) and high-density lipoprotein (HDL) cholesterol levels and triglyceride levels were measured using colorimetric assays (Hitachi LST008, Automatic Clinical Chemistry Analyzer, Hitachi, Naka, Japan). Non-HDL and remnant cholesterol levels were calculated by subtracting HDL from total cholesterol levels and by subtracting HDL and LDL from total cholesterol levels, respectively [45]. The definition of metabolic syndrome is shown in the Supplementary Materials, and other metabolic traits are shown in Supplementary Table S7 and as previously reported [41,42].

2.3. Regional Association Analysis

The genotyping in regional association analysis was performed using the Axiom Genome-Wide CHB Array data, and the data were analyzed after the exclusion criteria were applied (Figure 1). Imputation of the GWAS data was performed using East Asian populations of the 1000 Genomes Project Phase 3 as a reference panel and conducted using SHAPEIT (version 2, Oxford, UK, https://mathgen.stats.ox.ac.uk/genetics_software/shapeit/shapeit.html, accessed on 2 December 2020) and IMPUTE2 (version 2, Oxford, UK, http://mathgen.stats.ox.ac.uk/impute/impute_v2.html, accessed on 2 December 2020). The imputed results were validated through comparison with the WGS data from 137 independent samples [46]. After imputation, SNPs were filtered for QC with IMPUTE2 imputation quality scores of >0.3, and insertion and deletion mutations were removed using VCFtools (version 0.1, <https://vcftools.github.io/index.html>, accessed on 2 December 2020). For subsequent analyses, all samples were enrolled if the SNP missing call rate was <3%, the minor allele frequency was >0.01, and the Hardy–Weinberg equilibrium showed $p > 10^{-6}$. A total of 117,679 participants

were finally included for regional association analysis with 311 SNPs within the *ABCG5/G8* region ranging at positions between 43.93 and 44.21 Mb.

2.4. Statistical Analysis

We used Kolmogorov–Smirnov test to test normality of continuous variables in SPSS. Because all the variables we analyzed are skewed, we presented continuous variables as median and interquartile range. Categorical data are presented as percentage and number. In genotype–phenotype association studies, after adjustment for age, sex, BMI, and smoking status, a general linear regression model was used to evaluate the genetic effect of *ABCG5/G8* region variants on studied phenotypes. A logistic regression model was used to evaluate the effects of *ABCG5/G8* region variants on the risk of categorical phenotypes [expressed as odds ratios (ORs) with 95% confidence intervals (95% CIs)]. Nested logistic regression was used to cluster *ABCG5/G8* region variants to assess their correlations [47]. A stepwise multiple linear regression model was used to evaluate the influence of *ABCG5/G8* region variants on lipid profile. The calculations above were conducted using SPSS (version 22; SPSS, Chicago, IL, USA). Genome-wide significance association was defined as a significance level of $p < 5 \times 10^{-8}$. For Bonferroni correction, according to a total of 311 variants and 31 traits analyzed in regional associational analysis, the significance was indicated by $p < 5.0 \times 10^{-6}$, calculated as $0.05/(311 \times 31)$; whereas, according to a total of 12 rare variants and 8 traits analyzed in genotype–phenotype association analysis, a more liberal threshold of $p < 5.0 \times 10^{-4}$, calculated as $0.05/12 \times 8$ with rare mutations, was used. The PLINK software package (version 1.07, Shaun Purcell, Cambridge, MA, USA, <https://zzz.bwh.harvard.edu/plink/>, accessed on 14 August 2021) was used for the regional association analysis and subsequent conditional analysis. LDmatrix (<https://analysistools.nci.nih.gov/LDlink/?tab=ldmatrix>, accessed on 19 April 2021) was used to calculate linkage disequilibrium (LD).

3. Results

3.1. Baseline Characteristics of TWB Participants According to Gallstone Disease History

Baseline characteristics, lipid profile, and family history of gallstone disease of 117,679 participants with whole-genome genotyping array data with and without a history of gallstone disease are summarized in Table 1. In logistic regression analysis, It was found that participants with a history of gallstone disease were older and more likely to be male and have a family history of gallstone disease, higher BMI, waist circumference, waist hip ratio and total, LDL, and non-HDL cholesterol levels and lower HDL cholesterol level.

Table 1. Baseline characteristics of Taiwan Biobank participants: according to gallstone disease histories.

Clinical and Laboratory Parameters *	Total	Gallstone Disease History	
		with	without
Number (%)	117,679	5359 (4.55%)	112,320 (95.45%)
Anthropology			
Age (years)	51.0 (40.0–59.0)	56.0 (48.0–62.0)	50.0 (40.0–59.0) ***
Sex (male vs. female)	42,462/75,217	2025/3334	40,437/71,883 *
Waist circumference (cm)	83.0 (76.0–90.0)	85.0 (79.0–92.0)	82.5 (76.0–89.5) ***
Waist–hip ratio	0.87 (0.82–0.91)	0.89 (0.84–0.93)	0.87 (0.82–0.91) ***
Body mass index (kg/m ²)	23.8 (21.6–26.3)	24.5 (22.3–27.0)	23.7 (21.5–26.3) ***
Lipid profile			
Total cholesterol (mg/dL)	193.0 (171.0–217.0)	193.0 (172.0–215.0)	193.0 (171.0–217.0) ***
LDL cholesterol (mg/dL)	119.0 (99.0–141.0)	120.0 (100.0–140.0)	119.0 (99.0–141.0) ***
Non-HDL cholesterol (mg/dL)	138.0 (116.0–162.0)	140.0 (118.0–162.0)	138.0 (116.0–162.0) ***
HDL cholesterol (mg/dL)	53.0 (45.0–63.0)	52.0 (44.0–61.0)	53.0 (45.0–63.0) ***
Triglyceride (mg/dL)	91.0 (64.0–133.0)	99.0 (71.0–140.0)	90.0 (64.0–133.0)
Remnant cholesterol (mg/dL)	16.0 (11.0–23.0)	18.0 (12.0–25.0)	16.0 (11.0–23.0)
History of gallstone disease			
Gallstone disease (%)	4.55% (5359)	100% (5359)	0% (0) ***
Family history of gallstone disease (%)	7.54% (8869)	12.09% (648)	7.32% (8221) ***

HDL: high-density lipoprotein, LDL: low-density lipoprotein, se: standard error. * Participant recruitment for analysis is shown in Figure 1. Level presented as median (interquartile range) or percentage (number). Logistic regression, * for $p < 0.05$, and *** for $p < 0.0001$, adjusted for sex, age, BMI, and current smoking. Age: adjusted for sex, BMI, and current smoking status. Sex: adjusted for age, BMI, and current smoking. BMI: adjusted for sex, age, and current smoking status.

3.2. Selection of Candidate ABCG5/G8 Region Variants

To investigate the associations of *ABCG5/G8* variants with cardiometabolic traits and the risks of metabolic syndrome and gallstone disease, we first selected candidate variants within the coding and promoter region of *ABCG5/G8* among 1478 TWB participants with WGS data. A total of 50 exonic, upstream, and 5'UTR variants were selected, among which 22 were Asian-specific, 30 were nonsynonymous, 12 were synonymous, 3 were nonsense, and 5 were located between *ABCG5* and *ABCG8* and at the promoter region of *ABCG5* and *ABCG8* (Supplementary Table S1). Among these selected variants, twelve of them were available on the Axiom Genome-Wide CHB Array and were enrolled for genotype–phenotype association analysis. Eight were nonsynonymous mutations: rs6756629 (p.R50C), rs748096191 (p.E146D), rs148186696 (p.R253H), rs119480069 (p.R389H), rs536081800 (p.R446Q), and rs199984328 (p.H510N) from *ABCG5* and rs11887534 (p.D19H) and rs750352877 (p.N160S) from *ABCG8*. Two were synonymous mutations: *ABCG5* rs767751451 (p.A181A) and *ABCG8* rs56132765 (p.V151V). Two were located between *ABCG5* and *ABCG8* and at the promoter region of *ABCG5*: rs560839317 and rs189132480. The locations and characteristics of the 12 variants are shown in Supplementary Tables S1 and S2.

3.3. Genotype–Phenotype Association Analysis of ABCG5/G8 Variants with Lipid Profile and Gallstone Disease History

We further investigated the associations of the *ABCG5/G8* variants with the lipid profile and gallstone disease history of participants with GWAS Array data (Table 2 and Supplementary Table S3). By linear regression analysis, genome-wide significant associations were noted between rs199984328, rs119480069, and rs560839317 genotypes and total, LDL, and non-HDL cholesterol levels. The rs748096191 genotype also showed significant association with LDL, and non-HDL cholesterol levels and borderline significant association with total cholesterol level after Bonferroni correction ($p = 2.00 \times 10^{-4}$, $p = 2.68 \times 10^{-4}$, and $p = 8.20 \times 10^{-4}$, respectively). By logistic regression analysis, significant associations were also noted between rs6756629, rs11887534, and rs56132765 genotypes and the risk of gallstone disease ($p = 4.18 \times 10^{-7}$, $p = 2.08 \times 10^{-7}$, $p = 6.08 \times 10^{-7}$, respectively) and a trend of association with family history of gallstone disease ($p = 0.0056$, $p = 0.0061$, $p = 0.0161$, respectively).

Table 2. Phenotype–genotype associations of *ABCG5* and *ABCG8* exonic mutations and promoter variants.

Genetic Variants	Genotypes			β	SE	p Value *
<i>ABCG5</i> rs199984328 (108,563)	GG (107,083)	GT (1473)	TT (7)			
Total cholesterol# (mg/dL)	193.0 (171.0–217.0)	198.0 (175.0–224.0)	206.0 (190.0–212.0)	0.0116	0.0020	4.92×10^{-9}
LDL cholesterol# (mg/dL)	119.0 (99.0–140.0)	125.0 (104.0–146.0)	133.0 (125.0–134.0)	0.0186	0.0030	3.35×10^{-10}
Non-HDL cholesterol# (mg/dL)	138.0 (116.0–162.0)	143.0 (120.0–169.0)	153.0 (146.0–166.0)	0.0171	0.0027	1.71×10^{-10}
HDL cholesterol# (mg/dL)	53.0 (45.0–63.0)	53.0 (45.0–63.0)	46.0 (41.0–54.0)	−0.0020	0.0024	0.4072
Triglyceride# (mg/dL)	91.0 (64.0–133.0)	92.0 (63.0–137.0)	103.0 (71.0–175.0)	0.0102	0.0055	0.0655
Remnant cholesterol# (mg/dL)	16.0 (11.0–23.0)	16.0 (12.0–23.0)	19.0 (15.0–31.0)	0.0090	0.0068	0.1839
Gallstone disease (%)	4.55% (5267)	3.88% (63)	12.5% (1)	−0.1375	0.1274	0.2805
Family history of gallstone disease (%)	7.54% (8727)	7.08% (115)	0.00% (0)	−0.0772	0.0969	0.4257
<i>ABCG5</i> rs119480069 (108,808)	CC (108,166)	CT (641)	TT (1)			
Total cholesterol# (mg/dL)	193.0 (171.0–217.0)	204.0 (179.0–230.0)	233.0	0.0261	0.003	5.02×10^{-18}
LDL cholesterol# (mg/dL)	119.0 (99.0–140.0)	130.0 (107.0–153.0)	169.0	0.0398	0.0045	9.72×10^{-19}
Non-HDL cholesterol# (mg/dL)	138.0 (116.0–162.0)	150.0 (126.0–176.0)	195.0	0.0375	0.0041	3.22×10^{-20}
HDL cholesterol# (mg/dL)	53.0 (45.0–63.0)	53.0 (45.0–63.0)	38.0	−0.0025	0.0036	0.4899
Triglyceride# (mg/dL)	91.0 (64.0–133.0)	92.0 (66.0–135.5)	171.0	0.0137	0.0084	0.1033
Remnant cholesterol# (mg/dL)	16.0 (11.0–23.0)	17.0 (12.0–24.0)	26.0	0.0227	0.0103	0.0272
Gallstone disease (%)	4.56% (5329)	3.90% (28)	0.00% (0)	−0.1560	0.1941	0.4215

Table 2. Cont.

Genetic Variants		Genotypes		β	SE	p Value *
Family history of gallstone disease (%)	7.54% (8811)	7.66% (55)	0.00% (0)	0.0159	0.1406	0.9102
ABCG5 rs748096191 (108,793)	CC (108,673)	CG (120)	GG (0)			
Total cholesterol# (mg/dL)	193.0 (171.0–217.0)	204.0 (179.0–231.8)	–	0.0237	0.0071	8.20×10^{-4}
LDL cholesterol# (mg/dL)	119.0 (99.0–141.0)	130.5 (108.3–160.0)	–	0.0388	0.0104	2.00×10^{-4}
Non-HDL cholesterol# (mg/dL)	138.0 (116.0–162.0)	150.5 (122.0–183.8)	–	0.0343	0.0094	2.68×10^{-4}
HDL cholesterol# (mg/dL)	53.0 (45.0–63.0)	54.5 (46.0–62.8)	–	0.0039	0.0084	0.6443
Triglyceride# (mg/dL)	91.0 (64.0–133.0)	98.0 (67.3–142.8)	–	0.0316	0.0195	0.1054
Remnant cholesterol# (mg/dL)	16.0 (11.0–23.0)	16.0 (10.0–24.8)	–	−0.0060	0.0247	0.8083
Gallstone disease (%)	4.56% (5351)	3.82% (5)	–	−0.1757	0.4586	0.7017
Family history of gallstone disease (%)	7.54% (8851)	6.11% (8)	–	−0.2267	0.3651	0.5347
ABCG5 rs6756629 (108,862)	GG (105,597)	GA (3236)	AA (29)			
Total cholesterol# (mg/dL)	193.0 (171.0–217.0)	193.0 (170.0–217.0)	207.0 (191.0–229.5)	−0.0009	0.0013	0.4793
LDL cholesterol# (mg/dL)	119.0 (99.0–141.0)	118.0 (99.0–140.8)	130.0 (109.0–145.5)	−0.0019	0.002	0.3352
Non-HDL cholesterol# (mg/dL)	138.0 (116.0–162.0)	137.0 (115.0–161.0)	153.0 (137.0–172.5)	−0.0012	0.0018	0.5145
HDL cholesterol# (mg/dL)	53.0 (45.0–63.0)	53.0 (45.0–63.0)	54.0 (45.5–62.0)	−0.0005	0.0016	0.7504
Triglyceride# (mg/dL)	91.0 (64.0–133.0)	91.0 (63.0–133.0)	99.0 (76.5–169.0)	0.0037	0.0037	0.3231
Remnant cholesterol# (mg/dL)	16.0 (11.0–23.0)	16.0 (12.0–23.0)	21.0 (14.5–34.0)	0.0041	0.0046	0.3726
Gallstone disease (%)	4.50% (5134)	6.29% (220)	9.68% (3)	0.3535	0.0698	4.18×10^{-7}
Family history of gallstone disease (%)	7.50% (8558)	8.75% (306)	9.68% (3)	0.1656	0.0597	0.0056
ABCG5 rs560839317 (108,565)	GG (108,289)	GA (276)	GG (0)			
Total cholesterol# (mg/dL)	193.0 (171.0–217.0)	202.0 (180.0–229.8)	–	0.0222	0.0046	1.43×10^{-6}
LDL cholesterol# (mg/dL)	119.0 (99.0–141.0)	128.5 (108.0–151.8)	–	0.0339	0.0069	8.59×10^{-7}
Non-HDL cholesterol# (mg/dL)	138.0 (116.0–162.0)	146.0 (125.0–173.0)	–	0.0335	0.0062	6.97×10^{-8}
HDL cholesterol# (mg/dL)	53.0 (45.0–63.0)	52.0 (44.0–62.0)	–	−0.0081	0.0055	0.1376
Triglyceride# (mg/dL)	91.0 (64.0–133.0)	94.5 (69.3–147.0)	–	0.0344	0.0128	0.0073
Remnant cholesterol# (mg/dL)	16.0 (11.0–23.0)	17.0 (12.0–24.0)	–	0.0197	0.0156	0.2067
Gallstone disease (%)	4.56% (5331)	4.17% (13)	–	−0.0948	0.2853	0.7398
Family history of gallstone disease (%)	7.53% (8814)	8.97% (28)	–	0.1915	0.1984	0.3345
ABCG5/ABCG8 rs11887534 (108,880)	GG (105,602)	GC (3249)	CC (29)			
Total cholesterol# (mg/dL)	193.0 (171.0–217.0)	193.0 (170.0–217.0)	207.0 (189.5–229.5)	−0.0011	0.0013	0.3962
LDL cholesterol# (mg/dL)	119.0 (99.0–141.0)	118.0 (99.0–140.0)	128.0 (104.5–145.5)	−0.0023	0.0020	0.2401
Non-HDL cholesterol# (mg/dL)	138.0 (116.0–162.0)	137.0 (115.0–161.0)	153.0 (135.0–172.0)	−0.0015	0.0018	0.4000
HDL cholesterol# (mg/dL)	53.0 (45.0–63.0)	54.0 (45.0–63.0)	54.0 (45.5–62.5)	−0.0003	0.0016	0.8396
Triglyceride# (mg/dL)	91.0 (64.0–133.0)	91.0 (63.0–133.0)	99.0 (75.5–169.0)	0.0036	0.0037	0.3334
Remnant cholesterol# (mg/dL)	16.0 (11.0–23.0)	16.0 (12.0–23.0)	21.0 (14.5–34.0)	0.0045	0.0046	0.3197
Gallstone disease (%)	4.50% (5133)	6.34% (223)	9.68% (3)	0.3604	0.0694	2.08×10^{-7}
Family history of gallstone disease (%)	7.50% (8558)	8.79% (309)	6.45% (2)	0.1636	0.0596	0.0061
ABCG8 rs56132765 (108,805)	GG (105,499)	GA (3277)	AA (29)			
Total cholesterol# (mg/dL)	193.0 (171.0–217.0)	193.0 (170.0–217.0)	206.0 (189.5–221.5)	−0.0007	0.0013	0.5854
LDL cholesterol# (mg/dL)	119.0 (99.0–141.0)	119.0 (99.0–141.0)	128.0 (104.5–142.5)	−0.0016	0.0020	0.4120
Non-HDL cholesterol# (mg/dL)	138.0 (116.0–162.0)	137.0 (115.0–161.0)	149.0 (135.0–171.0)	−0.0010	0.0018	0.5685
HDL cholesterol# (mg/dL)	53.0 (45.0–63.0)	54.0 (45.0–63.0)	52.0 (43.5–61.5)	−0.0003	0.0016	0.8632
Triglyceride# (mg/dL)	91.0 (64.0–133.0)	90.0 (63.0–132.0)	99.0 (75.5–176.5)	0.0030	0.0037	0.4264
Remnant cholesterol# (mg/dL)	16.0 (11.0–23.0)	16.0 (12.0–23.0)	21.0 (14.0–34.0)	0.0032	0.0045	0.4747
Gallstone disease (%)	4.50% (5129)	6.26% (222)	9.68% (3)	0.3470	0.0696	6.08×10^{-7}
Family history of gallstone disease (%)	7.50% (8554)	8.57% (304)	9.68% (3)	0.1440	0.0599	0.0161

Data presented as median (interquartile range). Abbreviation as in Table 1. Number of the participants shown in brackets after the genotypes. # Participants were analyzed after the exclusion of those with a history of hyperlipidemia. * p value: adjusted for age, sex, BMI, and current smoking.

3.4. Regional Association Analysis for the Associations of *ABCG5* Region Variants with Lipid Profile and Gallstone Disease History

Regional association analyses in participants with GWAS Array data were performed to determine the peak association of genetic variants around the *ABCG5*/*G8* region with lipid profile and gallstone disease history. Our data revealed that the lead SNPs with genome-wide significance were rs75832441 for total, LDL, and non-HDL cholesterol levels and rs115445558 for history of gallstone disease (Figure 2).

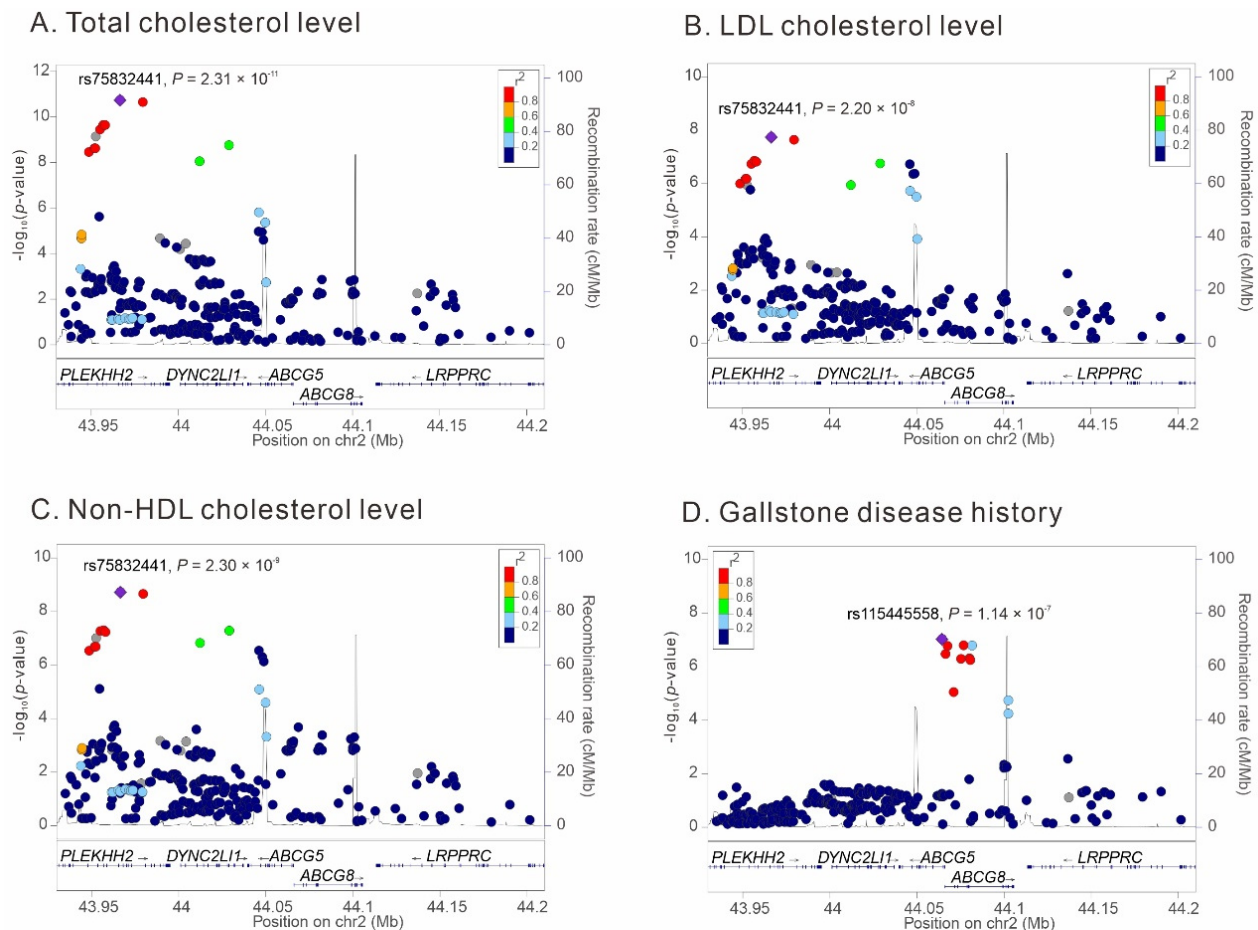
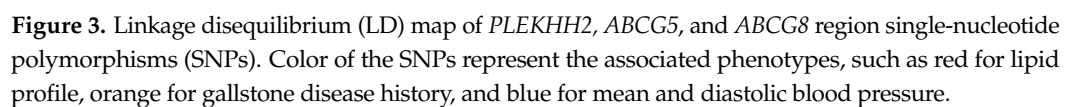


Figure 2. Regional association analysis of *ABCG5* and *ABCG8* region variants, (A) total cholesterol level, (B) LDL cholesterol level, (C) Non-HDL cholesterol level, and (D) gallstone disease history in Taiwan Biobank participants who had Axiom Genome-Wide CHB Array data. The lead SNPs with genome-wide significance were rs75832441 for total, LDL, and non-HDL cholesterol levels and rs115445558 for history of gallstone disease.

3.5. Linkage Disequilibrium (LD) between the Selected *ABCG5* and *ABCG8* Variants and Lead SNPs

Because both the selected *ABCG5*/*G8* variants and the lead SNPs were associated to various degrees with lipid profile and gallstone disease history, we further tested the LD between all these variants (Figure 3). In population genetics, LD is the non-random association of alleles at different loci in a given population. Our data revealed nearly complete LD (r^2 between 0.9503 and 0.9974) between four variants (rs6756629, rs11887534, rs56132765, and rs115445558) that are significantly associated with history of gallstone disease. Moderate LD was noted between rs75832441 and rs199984328, and both were in moderate-to-weak LD with the four variants associated with the risk of gallstone disease (r^2 between 0.1578 and 0.2344). The LDs of all other variant associations were weak ($r^2 \leq 0.0001$).



Because a total of five SNPs were found to be significantly associated with total, LDL, and non-HDL cholesterol levels, we tested the multicollinearity between the variables in the stepwise multiple linear regression model. Since the *ABCG5* rs75832441 had the lowest tolerance (0.319, that is <0.4) for all three phenotypes analyzed (Supplementary Table S4), we removed *ABCG5* rs75832441 from the model (Supplementary Table S4). Further, principal component analysis was performed according to the method previously reported [46]. Stepwise linear regression analysis for lipid profile with age, sex, BMI, smoking status, and four *ABCG5/G8* variants showed independent associations between rs119480069, rs199984328, rs560839317, and rs748096191 and total, LDL, and non-HDL cholesterol levels, which contributed to 0.07%, 0.03%, 0.02% and 0.01%, respectively for total and non-HDL cholesterol levels and 0.07%, 0.04%, 0.02% and 0.01%, respectively for LDL cholesterol levels. Together, these four variants contributed to 0.13%, 0.14%, and 0.13% for total, LDL, and non-HDL cholesterol levels, respectively (Table 3).

Table 3. Stepwise linear regression analysis: lipid profile.

	Total Cholesterol# (mg/dL)				LDL Cholesterol# (mg/dL)				Non-HDL Cholesterol# (mg/dL)			
	β	SE	R ²	p Value	β	SE	R ²	p Value	β	SE	R ²	p Value
Age (years)	0.0013	0.00002	0.0340	$<10^{-307}$	0.0014	0.00003	0.0167	$<10^{-307}$	0.0019	0.00003	0.0352	$<10^{-307}$
Sex (male vs. female)	0.0139	0.0005	0.0047	8.35×10^{-170}	-	-	-	-	-0.0054	0.0008	0.0008	2.17×10^{-12}
Body mass index (kg/m ²)	0.0015	0.0001	0.0053	4.15×10^{-130}	0.0049	0.0001	0.0256	$<10^{-307}$	0.006	0.0001	0.0484	$<10^{-307}$
Current smoking status (%)	-	-	-	-	-0.0028	0.0009	0.0001	0.0017	0.0031	0.0009	0.0001	0.0006
ABCG5 rs119480069 (CC vs. CT vs. TT)	0.0261	0.003	0.0007	6.89×10^{-18}	0.0398	0.0045	0.0007	1.66×10^{-18}	0.0371	0.0041	0.0007	1.25×10^{-19}
ABCG5 rs199984328 (GG vs. GT vs. TT)	0.0117	0.002	0.0003	5.08×10^{-9}	0.0188	0.003	0.0004	3.22×10^{-10}	0.0173	0.0027	0.0003	1.34×10^{-10}
ABCG5 rs560839317 (GG vs. GA vs. AA)	0.0226	0.0046	0.0002	9.94×10^{-7}	0.0346	0.0069	0.0002	5.60×10^{-7}	0.034	0.0063	0.0003	5.45×10^{-8}
ABCG5 rs748096191 (CC vs. CG vs. GG)	0.0268	0.007	0.0001	0.0001	0.0398	0.0105	0.0001	0.0001	0.0355	0.0094	0.0001	0.0002

Abbreviations as Table 1. # Participants were analyzed after the exclusion of those with a history of hyperlipidemia.

3.7. Nested Logistic Regression and Subgroup Analysis for History of Gallstone Disease

Among four *ABCG5/G8* variants associated with gallstone disease history, only two were nonsynonymous [*ABCG5* R50C (rs6756629) and *ABCG8* D19H (rs11887534)]. Using nested logistic regression analysis by including *ABCG5* R50C as a mandatory explanatory variable to statistically evaluate the causative role of individual variants in the disease association region, we found that the *ABCG8* D19H variant showed a trend of significant improvement for model fit ($p = 0.074$) (Table 4). This result is consistent with that reported by von Kampen, et al. [48], who suggested that the *ABCG8* D19H variant is the major causative variant in the region. Due to previous reports of a stronger association between the *ABCG8* D19H variant and gallstone disease history in female patients and younger individuals [36,49], we further tested the associations in different age and sex subgroups with interaction analysis. Our data showed no evidence of interaction between the age subgroups and sex categories on the associations (Supplementary Table S5).

Table 4. Nested logistic regression analysis of *ABCG5* R50C and *ABCG8* D19H mutations with gallstone disease histories.

	R50C			D19H			Nested Models	
	β	p Value	OR (95%CI)	β	p Value	OR (95%CI)	R50C D19H	D19H R50C
Gallstone disease (%)	0.3535	4.18×10^{-7}	1.42 (1.24–1.63)	0.3604	2.08×10^{-7}	1.43 (1.25–1.64)	0.127	0.074

3.8. Regional Association Analysis and Genotype–Phenotype Analysis for the Association between *ABCG5/G8* Region Variants and Cardiometabolic Traits

Several human and animal studies have linked *ABCG5/G8* variants and expression with cardiometabolic traits, such as insulin sensitivity, glycemic control, blood pressure status, and fatty liver disease [23–30]. We tested whether *ABCG5/G8* variants in TWB participants were also associated with various cardiometabolic traits and metabolic syndrome (Supplementary Table S6). Our data revealed that, with the exception of total, LDL, and non-HDL cholesterol levels, none of the other study phenotypes reached a genome-wide significant association under either the regional association analysis or candidate genotype–phenotype association analysis (Supplementary Figure S1, Supplementary Tables S8–S14).

However, we did identify several SNPs in the intron region of pleckstrin homology, MyTH4, and FERM domain containing the H2 (*PLEKHH2*) gene that showed significant associations ($p < 5 \times 10^{-7}$) with mean and diastolic blood pressure; the lead SNPs were rs7596913 and rs2060173, respectively.

4. Discussion

This study investigated the associations of *ABCG5/G8* variants with lipid profile, various cardiometabolic traits, and gallstone disease history in a Taiwanese cohort. Our data revealed that four Asian-specific, low-frequency or rare *ABCG5* variants, namely rs119480069, rs199984328, rs748096191, and rs560839317, were independently associated with total, LDL, and non-HDL cholesterol levels. To the best of our knowledge, rs119480069 (p.R389H) is the only variant to have previously been reported to be associated with LDL cholesterol levels [12,33,50,51]; the associations of the other three variants with LDL cholesterol levels are novel discoveries. In addition, all four studied *ABCG5/G8* variants that showed a significant association with gallstone disease history were in nearly complete LD with each other, and the most likely causative variant for the development of gallstone disease is *ABCG8* D19H (rs11887534), as was previously reported [48]. With the exception of the associations with mean and diastolic blood pressure of two variants at the intron region of *PLEKHH2*, a gene located very close to *ABCG5/G8*, associations with other metabolic traits were not found for the *ABCG5/G8* variants in our investigation. Further, in contrast to several studies of European populations [34,35,37], we found that the *ABCG8* D19H variant was not associated with lipid profile in our study population. Our data indicated differential associations of *ABCG5/G8* variants with lipid profile and gallstone disease history. These results also revealed the crucial role of individualized assessment of *ABCG5/G8* variants for different phenotypes in populations of different ethnicities.

4.1. *ABCG5/G8* Variants, Sitosterolemia, and Hypercholesterolemia

Although *ABCG5/G8* variants may increase total and LDL cholesterol levels, they are generally not considered to be the typical defective genes for familial hypercholesterolemia. Rather, *ABCG5/G8* variants can act as a component of an LDL cholesterol genetic risk score [52] and are considered LDL-cholesterol-altering accessory genes that mimic and worsen phenotypes of familial hypercholesterolemia [11,53–55]. Our study showed that *ABCG5* rs119480069, rs199984328, rs560839317, and rs748096191 were independently and positively associated with total, LDL, and non-HDL cholesterol levels and together contributed to 0.13% to 0.14% for total, LDL, and non-HDL cholesterol levels. All four variants are low frequency or rare in occurrence and are Asian-specific, and the increased LDL cholesterol levels from heterozygous to homozygous variants in rs119480069 and rs199984328 suggest a codominant inheritance model of these two variants. Williams, et al. [17] classified experimentally verified sitosterol variants into six classes; the R389H (rs119480069) variant was classified as a class II variant affecting maturation of *ABCG5/G8* heterodimers. The mechanism underlying the association between the other variants and cholesterol levels is still unknown; however, Graf, et al. [32] analyzed 13 sitosterolemia-causing *ABCG5/G8* mutations and found that all the mutations reduced G5/G8 heterodimer trafficking from the endoplasmic reticulum to the Golgi apparatus and that 10 of them prevented stable heterodimer formation between G5 and G8. Thus, further study is necessary to elucidate whether disruption of *ABCG5* and *ABCG8* heterodimerization or *ABCG5* trafficking to the cell surface is the molecular basis for the associations. Serum sitosterol levels have been associated with atherosclerotic cardiovascular disease; however, whether the associations are due to sitosterol levels or are secondary to total cholesterol levels remains controversial [33,34,56]. By analyzing nine sitosterolemia families, Nomura, et al. [33] observed that heterozygous carriers of a loss-of-function variant in *ABCG5*, but not in *ABCG8*, significantly increased LDL cholesterol and sitosterol levels and increased the risk of CAD twofold. Hypercholesterolemia in individuals with the *ABCG5/G8* mutations have also been shown to respond to ezetimibe treatment effectively [57]. Thus, the elucidation of

functional *ABCG5/G8* mutations is important in determining target drug therapy. Further prospective studies with measurement of serum sitosterol levels of the TWB participants may help to further elucidate the role of the atherogenic effects of sitosterols.

4.2. *ABCG5/G8 Variants That Increase the Risk of Gallstone Disease*

Cholesterol gallstone disease, which is secondary to bile supersaturated with cholesterol, is one of the most common digestive diseases in industrialized countries [58]. Gallstone is a disease influenced by genetic factors [59]. A higher prevalence of gallstone disease in identical twins and first-degree relatives in individuals with gallstone disease highlights the importance of searching for genes involved in biliary cholesterol secretion that are critical to gallstone formation, such as *ABCG5/G8* [60,61]. Previous studies have shown that the *ABCG8* D19H (rs11887534) variant is associated with gallstone disease history, cancer derived from biliary tract, lipid profile, and cardiovascular diseases [34,35,37,48,49,61–64]. The association may be affected by age and sex, being more prominent in individuals who are young and female and especially in those undergoing hormone treatment [36,49]. Krawczyk, et al. [65] further revealed that the *ABCG8* D19H variant increases the risk of early-onset gallstone formation in children. Meta-analysis of the association of various *ABCG5/G8* variants and gallstone disease showed a strong association of D19H polymorphism with gallstone disease. T400K and Y54C polymorphisms may also be associated with gallstone disease, though to a lesser extent [66]. Meta-analysis of GWAS that involved 8720 cases and 55,152 controls also showed four susceptible regions for gallstone disease, including *ABCG8*, *TM4SF4*, *SULT2A1*, and *CYP7A1*; the candidate variants for *ABCG8* were rs11887534 and rs4245791 [63]. In contrast to the *ABCG5* R50C (rs6756629) variant, the *ABCG8* D19H variant was shown to be associated with increased transport activity and decreased cholesterol absorption, which may increase the risk of gallstone disease [48]. As in our results, nested logistic regression analysis supported the superiority of the *ABCG8* D19H variant as a causative variant, as reported by von Kampen, et al. [48]. However, the *ABCG8* D19H variant is not responsible for cholesterol synthesis and ileal expression of *ABCG5*, *ABCG8*, and *NPCIL1* [67]. The *ABCG5* Q604E (rs6720173) genotypes have been associated with the risk of gallstone and gallbladder disease [36,68]; however, our data from the TWB participants showed no evidence of such an association (Supplementary Table S9). Furthermore, our data did not find interactions between age and sex on the association between the *ABCG8* D19H variant and the risk of gallstone disease.

4.3. *ABCG5/G8 Variants and Metabolic Traits*

In this study, we found lead SNPs of suggestive genome-wide significance for mean and diastolic blood pressure at the *PLEKHH2* intron region near the *ABCG5/G8* region. However, none of the *ABCG5/8* nonsynonymous mutations showed such strong associations with the blood pressure status. Plekhh2, encoded by the *PLEKHH2* gene, is an intracellular protein highly enriched in renal glomerular podocytes. Direct interactions between the FERM domain of the plekhh2 and the focal adhesion protein Hic-5 and actin stabilize the cortical actin cytoskeleton by attenuating actin depolymerization, and are involved in the podocyte foot processes [69]. A high expression of *PLEKHH2* also significantly increased the expression of proliferation- and invasion-related proteins and promoted cell proliferation, migration, and invasion [70]. *PLEKHH2* variants have been associated with diabetes nephropathy, coronary artery disease and venous thromboembolism, and have interacted with antihypertensive drugs for new-onset diabetes [71–74]. Further fine mapping may help to elucidate the causative gene/variant of the association. Previous genetic association studies have shown associations between *ABCG5/G8* polymorphisms and triglyceride, HDL, and VLDL cholesterol levels, insulin sensitivity, and metabolic syndrome [24–26]. Animal studies also showed that mice with *ABCG5/G8* deficiencies may cause hypertriglyceridemia via multiple metabolic pathways [27] and that sterol transportation via *ABCG5* and *ABCG8* opposes the development of fatty liver disease and loss of glycemic control independently of phytosterol accumulation [28]. Acceleration of *ABCG5/G8*-mediated biliary cholesterol secre-

tion showed restoration of glycemic control and alleviation of hypertriglyceridemia in obese db/db mice [29]. These results suggest that *ABCG5/G8* may be involved in the regulation of cardiometabolic traits and metabolic disorders. However, our data revealed that none of the other study phenotypes reached genome-wide significant association under either regional association analysis or candidate genotype–phenotype association analysis. In the future, further larger genetic association studies may be necessary to elucidate whether *ABCG5/G8* variants affect cardiometabolic traits in addition to the blood pressure status and total, LDL, and non-HDL cholesterol levels.

4.4. Ethnic Heterogeneity on Differential Associations for the Pleiotropic Effects of *ABCG5/G8* Variants

Ethnic heterogeneity on differential associations for the pleiotropic effects of *ABCG5/G8* variants has previously been reported. In an analysis of the genetic causes of sitosterolemia in 33 families from different ethnic populations, all six Japanese probands appeared to have mutations in *ABCG5* only [12]. Nomura, et al. (2020) [33] also revealed that seven of the nine Japanese sitosterolemia families have mutations on *ABCG5*. These results are similar to those reported in Chinese patients with sitosterolemia [51] and in 750 index familial hypercholesterolemia patients in a Taiwanese cohort [50]. By contrast, mutations in *ABCG8* were more commonly encountered in Caucasian populations with sitosterolemia [12,34,35]. These results indicate differential effects of *ABCG5/G8* variants and the crucial role of individualized assessment for *ABCG5/G8* variants on different phenotypes in different geographic areas.

4.5. Limitations

The present study has limitations. First, *ABCG5/G8* mutations resulted in autosomal recessive sitosterolemia; however, we did not measure sitosterol levels. Second, previous nutrigenetic studies have identified that dietary intervention may be involved in the association between *ABCG5/G8* variants and the interindividual variability of circulating cholesterol levels [75]. Further analysis of dietary effects may help to provide more personalized dietary recommendations. Third, survival bias in this investigation could not be avoided due to the cross-sectional study design. Fourth, Asian-specific variants were the most commonly used in this investigation; thus, our findings may not be applicable to other ethnic groups. Finally, our study lacked a second cohort to determine replicability. Further study with a larger sample size and longitudinal follow-up would strengthen the validity of our findings.

5. Conclusions

This study, using a Taiwanese population-based genetic approach, confirmed the critical role of *ABCG5* variants in the *ABCG5/G8* region as the major determinants of LDL cholesterol levels, as has been confirmed in other Asian populations. Associations of *PLEKHH2* variants with blood pressure status is a novel finding that requires further confirmation. The association between the *ABCG8* D19H variant, a variant associated with gallstone disease, and lipid profile may depend on ethnicity. Our data indicate differential functional effects for each *ABCG5/G8* variant and the crucial role of individualized assessment for *ABCG5/G8* variants on different phenotypes and geographic areas. These results may also provide novel candidate *ABCG5* variants in determining target drug therapy and for preventive medicine in coronary atherosclerosis.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/genes14030754/s1>. Table S1: Selected *ABCG5* and *ABCG8* variants in Taiwan Biobank participants; data derived from the whole-genome sequence and Axiom Genome-Wide CHB Array plates with genotype imputation. Table S2: Lead single nucleotide polymorphisms (SNPs) for various phenotypes at the *ABCG5*, *ABCG8*, and *PLEKHH2* gene region; data derived from the Axiom Genome-Wide CHB Array plates with genotype imputation. Table S3: Phenotype–genotype associations of *ABCG5* and *ABCG8* exonic mutations and promoter variants with no significant association with lipid profile or gallstone disease history. Table S4: The multicollinearity

between the lipid levels in the stepwise multiple linear regression model. Table S5: Subgroup analysis between rs11887534 and gallstone disease history; age and sex categories. Table S6: Genotype-phenotype association analysis of *ABCG5* and *ABCG8* lead single nucleotide variants for lipid profile and gallstone disease history. Table S7: Baseline characteristics of Taiwan Biobank participants. Figure S1: Regional association analysis of *ABCG5* and *ABCG8* loci variants and lipid profile. Table S8: Association between *ABCG5* rs560839317 genotypes and clinical and laboratory parameters in Taiwan Biobank participants. Table S9: Association of the *ABCG5* rs6720173 genotype with metabolic and hematological phenotypes; Table S10: Association between *ABCG5* rs199984328 genotypes and clinical and laboratory parameters in Taiwan Biobank participants. Table S11: Association between *ABCG5* rs119480069 genotypes and clinical and laboratory parameters in Taiwan Biobank participants. Table S12: Association between *ABCG5* rs11887534 genotypes and clinical and laboratory parameters in Taiwan Biobank participants. Table S13: Association between *ABCG5* rs7596913 genotypes and clinical and laboratory parameters in Taiwan Biobank participants. Table S14: Association between *ABCG5* rs2060173 genotypes and clinical and laboratory parameters in Taiwan Biobank participants.

Author Contributions: Conceptualization, Y.-L.K. and M.-S.T.; methodology, Y.-L.K. and L.-A.H.; software and validation, M.-S.T. and L.-A.H.; formal analysis, M.-S.T., Y.-L.K. and K.-H.Y.; resources, Y.-L.K. and H.-H.C.; data curation, M.-S.T., K.-H.Y. and S.W.; writing—original draft preparation, M.-S.T. and K.-H.Y.; writing—review & editing, Y.-L.K., L.-K.E. and L.-A.H.; visualization, S.W. and H.-H.C.; project administration, Y.-L.K. and H.-H.C.; supervision, Y.-L.K.; funding acquisition, K.-H.Y., H.-H.C. and Y.-L.K. All authors have read and agreed to the published version of the manuscript.

Funding: This study was supported by grants from Buddhist Tzu Chi Medical Foundation (TCMF-EP 111-02) and Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation (TCRD-TPE-112-02) to Y.-L.K.; Buddhist Tzu Chi Medical Foundation (TCMF-A 112-02) and Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation (TCRD-TPE-112-31) to K.-H.Y.; and Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation (TCRD-TPE-109-RT-1) to H.-H.C.

Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki, and approved by the Institutional Review Board of Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation (approval number: 08-XD-005), and the Ethics and Governance Council of the Taiwan Biobank (approval number: TWBR11011-02).

Informed Consent Statement: Written informed consent was obtained from all subjects involved in the study.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Acknowledgments: We greatly appreciate technical support from the Core Laboratory of the Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation, and expert statistical analysis assistance from Tsung-Han Hsieh.

Conflicts of Interest: No potential conflict of interest relevant to this article was reported.

References

1. Liu, X. ABC Family Transporters. *Adv. Exp. Med. Biol.* **2019**, *1141*, 13–100. [[CrossRef](#)] [[PubMed](#)]
2. Thomas, C.; Tampé, R. Structural and Mechanistic Principles of ABC Transporters. *Annu. Rev. Biochem.* **2020**, *89*, 605–636. [[CrossRef](#)] [[PubMed](#)]
3. Plummer, A.M.; Culbertson, A.T.; Liao, M. The ABCs of Sterol Transport. *Annu. Rev. Physiol.* **2021**, *83*, 153–181. [[CrossRef](#)] [[PubMed](#)]
4. Schumacher, T.; Benndorf, R.A. ABC Transport Proteins in Cardiovascular Disease—A Brief Summary. *Molecules* **2017**, *22*, 589. [[CrossRef](#)]
5. Patel, S.B.; Graf, G.A.; Temel, R.E. ABCG5 and ABCG8: More than a defense against xenosterols. *J. Lipid Res.* **2018**, *59*, 1103–1113. [[CrossRef](#)]
6. Wang, D.Q. Regulation of intestinal cholesterol absorption. *Annu. Rev. Physiol.* **2007**, *69*, 221–248. [[CrossRef](#)]
7. Graf, G.A.; Li, W.P.; Gerard, R.D.; Gelissen, I.; White, A.; Cohen, J.C.; Hobbs, H.H. Coexpression of ATP-binding cassette proteins ABCG5 and ABCG8 permits their transport to the apical surface. *J. Clin. Invest.* **2002**, *110*, 659–669. [[CrossRef](#)]
8. Graf, G.A.; Yu, L.; Li, W.P.; Gerard, R.; Tuma, P.L.; Cohen, J.C.; Hobbs, H.H. ABCG5 and ABCG8 are obligate heterodimers for protein trafficking and biliary cholesterol excretion. *J. Biol. Chem.* **2003**, *278*, 48275–48282. [[CrossRef](#)]

9. Wang, H.H.; Liu, M.; Portincasa, P.; Wang, D.Q. Recent Advances in the Critical Role of the Sterol Efflux Transporters ABCG5/G8 in Health and Disease. *Adv. Exp. Med. Biol.* **2020**, *1276*, 105–136. [\[CrossRef\]](#)
10. Zhang, D.W.; Graf, G.A.; Gerard, R.D.; Cohen, J.C.; Hobbs, H.H. Functional asymmetry of nucleotide-binding domains in ABCG5 and ABCG8. *J. Biol. Chem.* **2006**, *281*, 4507–4516. [\[CrossRef\]](#)
11. Tada, H.; Kawashiri, M.A.; Nomura, A.; Teramoto, R.; Hosomichi, K.; Nohara, A.; Inazu, A.; Mabuchi, H.; Tajima, A.; Yamagishi, M. Oligogenic familial hypercholesterolemia, LDL cholesterol, and coronary artery disease. *J. Clin. Lipidol.* **2018**, *12*, 1436–1444. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Lu, K.; Lee, M.H.; Hazard, S.; Brooks-Wilson, A.; Hidaka, H.; Kojima, H.; Ose, L.; Stalenhoef, A.F.; Mietinnen, T.; Bjorkhem, I.; et al. Two genes that map to the STSL locus cause sitosterolemia: Genomic structure and spectrum of mutations involving sterolin-1 and sterolin-2, encoded by ABCG5 and ABCG8, respectively. *Am. J. Hum. Genet.* **2001**, *69*, 278–290. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Niu, D.M.; Chong, K.W.; Hsu, J.H.; Wu, T.J.; Yu, H.C.; Huang, C.H.; Lo, M.Y.; Kwok, C.F.; Kratz, L.E.; Ho, L.T. Clinical observations, molecular genetic analysis, and treatment of sitosterolemia in infants and children. *J. Inher. Metab. Dis.* **2010**, *33*, 437–443. [\[CrossRef\]](#) [\[PubMed\]](#)
14. Wang, D.Q.; Afdhal, N.H. Genetic analysis of cholesterol gallstone formation: Searching for Lith (gallstone) genes. *Curr. Gastroenterol. Rep.* **2004**, *6*, 140–150. [\[CrossRef\]](#)
15. Wang, H.H.; Portincasa, P.; Afdhal, N.H.; Wang, D.Q. Lith genes and genetic analysis of cholesterol gallstone formation. *Gastroenterol. Clin. N. Am.* **2010**, *39*, 185–207. [\[CrossRef\]](#)
16. Wang, T.Y.; Portincasa, P.; Liu, M.; Tso, P.; Wang, D.Q. Mouse models of gallstone disease. *Curr. Opin. Gastroenterol.* **2018**, *34*, 59–70. [\[CrossRef\]](#)
17. Williams, K.; Segard, A.; Graf, G.A. Sitosterolemia: Twenty Years of Discovery of the Function of ABCG5/ABCG8. *Int. J. Mol. Sci.* **2021**, *22*, 2641. [\[CrossRef\]](#)
18. Wittenburg, H.; Lyons, M.A.; Li, R.; Churchill, G.A.; Carey, M.C.; Paigen, B. FXR and ABCG5/ABCG8 as determinants of cholesterol gallstone formation from quantitative trait locus mapping in mice. *Gastroenterology* **2003**, *125*, 868–881. [\[CrossRef\]](#)
19. Plösch, T.; Bloks, V.W.; Terasawa, Y.; Berdy, S.; Siegler, K.; Van Der Sluijs, F.; Kema, I.P.; Groen, A.K.; Shan, B.; Kuipers, F.; et al. Sitosterolemia in ABC-transporter G5-deficient mice is aggravated on activation of the liver-X receptor. *Gastroenterology* **2004**, *126*, 290–300. [\[CrossRef\]](#)
20. Wang, H.H.; Patel, S.B.; Carey, M.C.; Wang, D.Q. Quantifying anomalous intestinal sterol uptake, lymphatic transport, and biliary secretion in *Abcg8*(^{-/-}) mice. *Hepatology* **2007**, *45*, 998–1006. [\[CrossRef\]](#)
21. Yu, L.; Hammer, R.E.; Li-Hawkins, J.; Von Bergmann, K.; Lutjohann, D.; Cohen, J.C.; Hobbs, H.H. Disruption of *Abcg5* and *Abcg8* in mice reveals their crucial role in biliary cholesterol secretion. *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 16237–16242. [\[CrossRef\]](#) [\[PubMed\]](#)
22. Yu, L.; Li-Hawkins, J.; Hammer, R.E.; Berge, K.E.; Horton, J.D.; Cohen, J.C.; Hobbs, H.H. Overexpression of ABCG5 and ABCG8 promotes biliary cholesterol secretion and reduces fractional absorption of dietary cholesterol. *J. Clin. Invest.* **2002**, *110*, 671–680. [\[CrossRef\]](#)
23. Chen, J.; Batta, A.; Zheng, S.; Fitzgibbon, W.R.; Ullian, M.E.; Yu, H.; Tso, P.; Salen, G.; Patel, S.B. The missense mutation in *Abcg5* gene in spontaneously hypertensive rats (SHR) segregates with phytosterolemia but not hypertension. *BMC Genet.* **2005**, *6*, 40. [\[CrossRef\]](#)
24. Chen, Z.C.; Shin, S.J.; Kuo, K.K.; Lin, K.D.; Yu, M.L.; Hsiao, P.J. Significant association of ABCG8:D19H gene polymorphism with hypercholesterolemia and insulin resistance. *J. Hum. Genet.* **2008**, *53*, 757–763. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Garcia-Rios, A.; Perez-Martinez, P.; Fuentes, F.; Mata, P.; Lopez-Miranda, J.; Alonso, R.; Rodriguez, F.; Garcia-Olvid, A.; Ruano, J.; Ordoas, J.M.; et al. Genetic variations at ABCG5/G8 genes modulate plasma lipids concentrations in patients with familial hypercholesterolemia. *Atherosclerosis* **2010**, *210*, 486–492. [\[CrossRef\]](#) [\[PubMed\]](#)
26. Gylling, H.; Hallikainen, M.; Pihlajamäki, J.; Agren, J.; Laakso, M.; Rajaratnam, R.A.; Rauramaa, R.; Miettinen, T.A. Polymorphisms in the ABCG5 and ABCG8 genes associate with cholesterol absorption and insulin sensitivity. *J. Lipid Res.* **2004**, *45*, 1660–1665. [\[CrossRef\]](#)
27. Méndez-González, J.; Julve, J.; Rotllan, N.; Llaverias, G.; Blanco-Vaca, F.; Escolà-Gil, J.C. ATP-binding cassette G5/G8 deficiency causes hypertriglyceridemia by affecting multiple metabolic pathways. *Biochim. Et Biophys. Acta* **2011**, *1811*, 1186–1193. [\[CrossRef\]](#)
28. Su, K.; Sabeva, N.S.; Liu, J.; Wang, Y.; Bhatnagar, S.; van der Westhuyzen, D.R.; Graf, G.A. The ABCG5 ABCG8 sterol transporter opposes the development of fatty liver disease and loss of glycemic control independently of phytosterol accumulation. *J. Biol. Chem.* **2012**, *287*, 28564–28575. [\[CrossRef\]](#)
29. Su, K.; Sabeva, N.S.; Wang, Y.; Liu, X.; Lester, J.D.; Liu, J.; Liang, S.; Graf, G.A. Acceleration of biliary cholesterol secretion restores glycemic control and alleviates hypertriglyceridemia in obese db/db mice. *Arterioscler. Thromb. Vasc. Biol.* **2014**, *34*, 26–33. [\[CrossRef\]](#)
30. Yu, H.; Pandit, B.; Klett, E.; Lee, M.H.; Lu, K.; Helou, K.; Ikeda, I.; Egashira, N.; Sato, M.; Klein, R.; et al. The rat STSL locus: Characterization, chromosomal assignment, and genetic variations in sitosterolemic hypertensive rats. *BMC Cardiovasc. Disord.* **2003**, *3*, 4. [\[CrossRef\]](#)
31. Lek, M.; Karczewski, K.J.; Minikel, E.V.; Samocha, K.E.; Banks, E.; Fennell, T.; O'Donnell-Luria, A.H.; Ware, J.S.; Hill, A.J.; Cummings, B.B.; et al. Analysis of protein-coding genetic variation in 60,706 humans. *Nature* **2016**, *536*, 285–291. [\[CrossRef\]](#)

32. Graf, G.A.; Cohen, J.C.; Hobbs, H.H. Missense mutations in ABCG5 and ABCG8 disrupt heterodimerization and trafficking. *J. Biol. Chem.* **2004**, *279*, 24881–24888. [[CrossRef](#)] [[PubMed](#)]
33. Nomura, A.; Emdin, C.A.; Won, H.H.; Peloso, G.M.; Natarajan, P.; Ardissino, D.; Danesh, J.; Schunkert, H.; Correa, A.; Bown, M.J.; et al. Heterozygous ABCG5 Gene Deficiency and Risk of Coronary Artery Disease. *Circ. Genom. Precis. Med.* **2020**, *13*, 417–423. [[CrossRef](#)]
34. Scholz, M.; Horn, K.; Pott, J.; Gross, A.; Kleber, M.E.; Delgado, G.E.; Mishra, P.P.; Kirsten, H.; Gieger, C.; Müller-Nurasyid, M.; et al. Genome-wide meta-analysis of phytosterols reveals five novel loci and a detrimental effect on coronary atherosclerosis. *Nat. Commun.* **2022**, *13*, 143. [[CrossRef](#)] [[PubMed](#)]
35. Helgadóttir, A.; Thorleifsson, G.; Alexandersson, K.F.; Tragante, V.; Thorsteinsdóttir, M.; Eiriksson, F.F.; Gretarsdóttir, S.; Björnsson, E.; Magnusson, O.; Sveinbjörnsson, G.; et al. Genetic variability in the absorption of dietary sterols affects the risk of coronary artery disease. *Eur. Heart J.* **2020**, *41*, 2618–2628. [[CrossRef](#)] [[PubMed](#)]
36. Kuo, K.K.; Shin, S.J.; Chen, Z.C.; Yang, Y.H.; Yang, J.F.; Hsiao, P.J. Significant association of ABCG5 604Q and ABCG8 D19H polymorphisms with gallstone disease. *Br. J. Surg.* **2008**, *95*, 1005–1011. [[CrossRef](#)]
37. Stender, S.; Frikke-Schmidt, R.; Nordestgaard, B.G.; Tybjaerg-Hansen, A. The ABCG5/8 cholesterol transporter and myocardial infarction versus gallstone disease. *J. Am. Coll. Cardiol.* **2014**, *63*, 2121–2128. [[CrossRef](#)]
38. Scheinfeldt, L.B.; Tishkoff, S.A. Recent human adaptation: Genomic approaches, interpretation and insights. *Nat. Reviews. Genet.* **2013**, *14*, 692–702. [[CrossRef](#)]
39. Chen, C.H.; Yang, J.H.; Chiang, C.W.K.; Hsiung, C.N.; Wu, P.E.; Chang, L.C.; Chu, H.W.; Chang, J.; Song, I.W.; Yang, S.L.; et al. Population structure of Han Chinese in the modern Taiwanese population based on 10,000 participants in the Taiwan Biobank project. *Hum. Mol. Genet.* **2016**, *25*, 5321–5331. [[CrossRef](#)]
40. Juang, J.J.; Lu, T.P.; Su, M.W.; Lin, C.W.; Yang, J.H.; Chu, H.W.; Chen, C.H.; Hsiao, Y.W.; Lee, C.Y.; Chiang, L.M.; et al. Rare variants discovery by extensive whole-genome sequencing of the Han Chinese population in Taiwan: Applications to cardiovascular medicine. *J. Adv. Res.* **2021**, *30*, 147–158. [[CrossRef](#)]
41. Hsu, L.A.; Teng, M.S.; Wu, S.; Chou, H.H.; Ko, Y.L. Common and Rare PCSK9 Variants Associated with Low-Density Lipoprotein Cholesterol Levels and the Risk of Diabetes Mellitus: A Mendelian Randomization Study. *Int. J. Mol. Sci.* **2022**, *23*, 418. [[CrossRef](#)] [[PubMed](#)]
42. Yeh, K.H.; Hsu, L.A.; Teng, M.S.; Wu, S.; Chou, H.H.; Ko, Y.L. Pleiotropic Effects of Common and Rare GCKR Exonic Mutations on Cardiometabolic Traits. *Genes* **2022**, *13*, 491. [[CrossRef](#)] [[PubMed](#)]
43. Yeh, K.H.; Wan, H.L.; Teng, M.S.; Chou, H.H.; Hsu, L.A.; Ko, Y.L. Genetic Variants at the APOE Locus Predict Cardiometabolic Traits and Metabolic Syndrome: A Taiwan Biobank Study. *Genes* **2022**, *13*, 1366. [[CrossRef](#)] [[PubMed](#)]
44. Raczy, C.; Petrovski, R.; Saunders, C.T.; Chorny, I.; Kruglyak, S.; Margulies, E.H.; Chuang, H.Y.; Källberg, M.; Kumar, S.A.; Liao, A.; et al. Isaac: Ultra-fast whole-genome secondary analysis on Illumina sequencing platforms. *Bioinformatics* **2013**, *29*, 2041–2043. [[CrossRef](#)]
45. Chait, A.; Ginsberg, H.N.; Vaisar, T.; Heinecke, J.W.; Goldberg, I.J.; Bornfeldt, K.E. Remnants of the Triglyceride-Rich Lipoproteins, Diabetes, and Cardiovascular Disease. *Diabetes* **2020**, *69*, 508–516. [[CrossRef](#)]
46. Wei, C.Y.; Yang, J.H.; Yeh, E.C.; Tsai, M.F.; Kao, H.J.; Lo, C.Z.; Chang, L.P.; Lin, W.J.; Hsieh, F.J.; Belsare, S.; et al. Genetic profiles of 103,106 individuals in the Taiwan Biobank provide insights into the health and history of Han Chinese. *NPJ Genom. Med.* **2021**, *6*, 10. [[CrossRef](#)]
47. Perez-Lopez, J.-B.; Novales, M.; Orro, A. Spatially correlated nested logit model for spatial location choice. *Transp. Res. Part B Methodol.* **2022**, *161*, 1–12. [[CrossRef](#)]
48. von Kampen, O.; Buch, S.; Nothnagel, M.; Azocar, L.; Molina, H.; Brosch, M.; Erhart, W.; von Schönfels, W.; Egberts, J.; Seeger, M.; et al. Genetic and functional identification of the likely causative variant for cholesterol gallstone disease at the ABCG5/8 lithogenic locus. *Hepatology* **2013**, *57*, 2407–2417. [[CrossRef](#)]
49. Liang, K.W.; Huang, H.H.; Wang, L.; Lu, W.Y.; Chou, Y.H.; Tantoh, D.M.; Nfor, O.N.; Chiu, N.Y.; Tyan, Y.S.; Liaw, Y.P. Risk of gallstones based on ABCG8 rs11887534 single nucleotide polymorphism among Taiwanese men and women. *BMC Gastroenterol.* **2021**, *21*, 468. [[CrossRef](#)]
50. Huang, C.C.; Niu, D.M.; Charng, M.J. Genetic Analysis in a Taiwanese Cohort of 750 Index Patients with Clinically Diagnosed Familial Hypercholesterolemia. *J. Atheroscler. Thromb.* **2022**, *29*, 639–653. [[CrossRef](#)]
51. Zhou, Z.; Su, X.; Cai, Y.; Ting, T.H.; Zhang, W.; Lin, Y.; Xu, A.; Mao, X.; Zeng, C.; Liu, L.; et al. Features of chinese patients with sitosterolemia. *Lipids Health Dis.* **2022**, *21*, 11. [[CrossRef](#)] [[PubMed](#)]
52. Futema, M.; Shah, S.; Cooper, J.A.; Li, K.; Whittall, R.A.; Sharifi, M.; Goldberg, O.; Drogari, E.; Mollaki, V.; Wiegman, A.; et al. Refinement of variant selection for the LDL cholesterol genetic risk score in the diagnosis of the polygenic form of clinical familial hypercholesterolemia and replication in samples from 6 countries. *Clin. Chem.* **2015**, *61*, 231–238. [[CrossRef](#)]
53. Lamiquiz-Moneo, I.; Baila-Rueda, L.; Bea, A.M.; Mateo-Gallego, R.; Pérez-Calahorra, S.; Marco-Benedí, V.; Martín-Navarro, A.; Ros, E.; Cofán, M.; Rodríguez-Rey, J.C.; et al. ABCG5/G8 gene is associated with hypercholesterolemias without mutation in candidate genes and noncholesterol sterols. *J. Clin. Lipidol.* **2017**, *11*, 1432–1440.e1434. [[CrossRef](#)] [[PubMed](#)]
54. Reeskamp, L.F.; Volta, A.; Zuurbier, L.; Defesche, J.C.; Hovingh, G.K.; Grefhorst, A. ABCG5 and ABCG8 genetic variants in familial hypercholesterolemia. *J. Clin. Lipidol.* **2020**, *14*, 207–217.e207. [[CrossRef](#)] [[PubMed](#)]

55. Tada, H.; Okada, H.; Nomura, A.; Yashiro, S.; Nohara, A.; Ishigaki, Y.; Takamura, M.; Kawashiri, M.A. Rare and Deleterious Mutations in ABCG5/ABCG8 Genes Contribute to Mimicking and Worsening of Familial Hypercholesterolemia Phenotype. *Circ. J. Off. J. Jpn. Circ. Soc.* **2019**, *83*, 1917–1924. [\[CrossRef\]](#)
56. Teupser, D.; Baber, R.; Ceglarek, U.; Scholz, M.; Illig, T.; Gieger, C.; Holdt, L.M.; Leichtle, A.; Greiser, K.H.; Huster, D.; et al. Genetic regulation of serum phytosterol levels and risk of coronary artery disease. *Circulation. Cardiovasc. Genet.* **2010**, *3*, 331–339. [\[CrossRef\]](#)
57. Tada, H.; Okada, H.; Nomura, A.; Takamura, M.; Kawashiri, M.A. Beneficial effect of ezetimibe-atorvastatin combination therapy in patients with a mutation in ABCG5 or ABCG8 gene. *Lipids Health Dis.* **2020**, *19*, 3. [\[CrossRef\]](#)
58. Shaffer, E.A. Gallstone disease: Epidemiology of gallbladder stone disease. *Best Pract. Res. Clin. Gastroenterol.* **2006**, *20*, 981–996. [\[CrossRef\]](#)
59. Rebholz, C.; Krawczyk, M.; Lammert, F. Genetics of gallstone disease. *Eur. J. Clin. Investig.* **2018**, *48*, e12935. [\[CrossRef\]](#)
60. Katsika, D.; Grijibovski, A.; Einarsson, C.; Lammert, F.; Lichtenstein, P.; Marschall, H.U. Genetic and environmental influences on symptomatic gallstone disease: A Swedish study of 43,141 twin pairs. *Hepatology* **2005**, *41*, 1138–1143. [\[CrossRef\]](#)
61. Katsika, D.; Magnusson, P.; Krawczyk, M.; Grünhage, F.; Lichtenstein, P.; Einarsson, C.; Lammert, F.; Marschall, H.U. Gallstone disease in Swedish twins: Risk is associated with ABCG8 D19H genotype. *J. Intern. Med.* **2010**, *268*, 279–285. [\[CrossRef\]](#)
62. Grünhage, F.; Acalovschi, M.; Tirziu, S.; Walier, M.; Wienker, T.F.; Ciocan, A.; Mosteanu, O.; Sauerbruch, T.; Lammert, F. Increased gallstone risk in humans conferred by common variant of hepatic ATP-binding cassette transporter for cholesterol. *Hepatology* **2007**, *46*, 793–801. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Joshi, A.D.; Andersson, C.; Buch, S.; Stender, S.; Noordam, R.; Weng, L.C.; Weeke, P.E.; Auer, P.L.; Boehm, B.; Chen, C.; et al. Four Susceptibility Loci for Gallstone Disease Identified in a Meta-analysis of Genome-Wide Association Studies. *Gastroenterology* **2016**, *151*, 351–363.e328. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Srivastava, A.; Tulsyan, S.; Pandey, S.N.; Choudhuri, G.; Mittal, B. Single nucleotide polymorphism in the ABCG8 transporter gene is associated with gallbladder cancer susceptibility. *Liver Int. Off. J. Int. Assoc. Study Liver* **2009**, *29*, 831–837. [\[CrossRef\]](#)
65. Krawczyk, M.; Niewiadomska, O.; Jankowska, I.; Jankowski, K.; Więckowski, S.; Lebensztejn, D.; Więcek, S.; Gozdowska, J.; Kułaga, Z.; Weber, S.N.; et al. Common variant p.D19H of the hepatobiliary sterol transporter ABCG8 increases the risk of gallstones in children. *Liver Int. Off. J. Int. Assoc. Study Liver* **2022**, *42*, 1585–1592. [\[CrossRef\]](#)
66. Jiang, Z.Y.; Cai, Q.; Chen, E.Z. Association of three common single nucleotide polymorphisms of ATP binding cassette G8 gene with gallstone disease: A meta-analysis. *PLoS ONE* **2014**, *9*, e87200. [\[CrossRef\]](#)
67. Renner, O.; Lütjohann, D.; Richter, D.; Strohmeyer, A.; Schimmel, S.; Müller, O.; Stange, E.F.; Harsch, S. Role of the ABCG8 19H risk allele in cholesterol absorption and gallstone disease. *BMC Gastroenterol.* **2013**, *13*, 30. [\[CrossRef\]](#)
68. Rodriguez, S.; Gaunt, T.R.; Guo, Y.; Zheng, J.; Barnes, M.R.; Tang, W.; Danish, F.; Johnson, A.; Castillo, B.A.; Li, Y.R.; et al. Lipids, obesity and gallbladder disease in women: Insights from genetic studies using the cardiovascular gene-centric 50K SNP array. *Eur. J. Hum. Genet. EJHG* **2016**, *24*, 106–112. [\[CrossRef\]](#)
69. Perisic, L.; Lal, M.; Hulkko, J.; Hultenby, K.; Önfelt, B.; Sun, Y.; Dunér, F.; Patrakka, J.; Betsholtz, C.; Uhlen, M.; et al. Plekhh2, a novel podocyte protein downregulated in human focal segmental glomerulosclerosis, is involved in matrix adhesion and actin dynamics. *Kidney Int.* **2012**, *82*, 1071–1083. [\[CrossRef\]](#)
70. Wang, R.; Wang, S.; Li, Z.; Luo, Y.; Zhao, Y.; Han, Q.; Rong, X.Z.; Guo, Y.X.; Liu, Y. PLEKHH2 binds β -arrestin1 through its FERM domain, activates FAK/PI3K/AKT phosphorylation, and promotes the malignant phenotype of non-small cell lung cancer. *Cell Death Dis.* **2022**, *13*, 858. [\[CrossRef\]](#)
71. Greene, C.N.; Keong, L.M.; Cordovado, S.K.; Mueller, P.W. Sequence variants in the PLEKHH2 region are associated with diabetic nephropathy in the GoKinD study population. *Hum. Genet.* **2008**, *124*, 255–262. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Chang, S.W.; McDonough, C.W.; Gong, Y.; Johnson, T.A.; Tsunoda, T.; Gamazon, E.R.; Perera, M.A.; Takahashi, A.; Tanaka, T.; Kubo, M.; et al. Genome-wide association study identifies pharmacogenomic loci linked with specific antihypertensive drug treatment and new-onset diabetes. *Pharm. J.* **2018**, *18*, 106–112. [\[CrossRef\]](#) [\[PubMed\]](#)
73. Cunha, M.L.R.; Meijers, J.C.M.; Rosendaal, F.R.; Vlieg, A.V.H.; Reitsma, P.H.; Middeldorp, S. Whole exome sequencing in thrombophilic pedigrees to identify genetic risk factors for venous thromboembolism. *PLoS ONE* **2017**, *12*, e0187699. [\[CrossRef\]](#) [\[PubMed\]](#)
74. van der Harst, P.; Verweij, N. Identification of 64 Novel Genetic Loci Provides an Expanded View on the Genetic Architecture of Coronary Artery Disease. *Circ. Res.* **2018**, *122*, 433–443. [\[CrossRef\]](#)
75. Abdullah, M.M.H.; Vazquez-Vidal, I.; Baer, D.J.; House, J.D.; Jones, P.J.H.; Desmarchelier, C. Common Genetic Variations Involved in the Inter-Individual Variability of Circulating Cholesterol Concentrations in Response to Diets: A Narrative Review of Recent Evidence. *Nutrients* **2021**, *13*, 695. [\[CrossRef\]](#) [\[PubMed\]](#)

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.