



Article An Exploratory Application of Multilayer Networks and Pathway Analysis in Pharmacogenomics

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Abstract: Over the years, network analysis has become a promising strategy for analysing complex system, i.e., systems composed of a large number of interacting elements. In particular, multilayer networks have emerged as a powerful framework for modelling and analysing complex systems with multiple types of interactions. Network analysis can be applied to pharmacogenomics to gain insights into the interactions between genes, drugs, and diseases. By integrating network analysis techniques with pharmacogenomic data, the goal consists of uncovering complex relationships and identifying key genes to use in pathway enrichment analysis to figure out biological pathways involved in drug response and adverse reactions. In this study, we modelled omics, disease, and drug data together through multilayer network representation. Then, we mined the multilayer network with a community detection algorithm to obtain the top communities. After that, we used the identified list of genes from the communities to perform pathway enrichment analysis (PEA) to figure out the biological function affected by the selected genes. The results show that the genes forming the top community have multiple roles through different pathways.

Keywords: pharmacogenomics; network analysis; multilayer networks; community detection; pathway enrichment analysis

1. Introduction

Network analysis is a branch of network science that deals with the study of complex networks. To investigate complex relationships, network analysis adopts theories and methods typical of several research areas [1]. Networks and network analysis methods are a keystone in computational biology and bioinformatics and are increasingly being used to study biological and clinical data in an integrated way. In detail, network analysis consists of a collection of techniques with a shared methodological perspective, which allows to depiction of relations among entities and to analysis of the structures that emerge from the recurrence of these relations. The basic assumption is that better explanations of different phenomena are yielded by the analysis of the relations among entities. A classical network analysis method is represented by community detection [2]. Community detection is one of the most popular research areas in various complex systems, such as biology, sociology, medicine, and transportation systems [3,4]. The reason for this is that the community structures, defined as groups of nodes that are more densely connected than the rest of the network, represent significant characteristics for understanding the functionalities and organizations of complex systems modelled as a network [2]. It is expected that the communities play significant roles in the structure-function relationship. For example, in



Citation: Milano, M.; Agapito, G.; Cannataro, M. An Exploratory Application of Multilayer Networks and Pathway Analysis in Pharmacogenomics. *Genes* 2023, 14, 1915. https://doi.org/10.3390/ genes14101915

Academic Editor: Cinzia Ciccacci

Received: 4 September 2023 Revised: 26 September 2023 Accepted: 5 October 2023 Published: 7 October 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). biological networks such as protein-protein interaction (PPI) networks, the communities represent proteins involved in a similar function; in neuroscience, the communities detected in brain networks mean regions of interest (ROI) that are active during tasks. In social networks, communities can be groups of friends or colleagues. In the World Wide Web, communities represent web pages sharing the same topic [5]. Thus, the discovery of communities in these systems has become an interesting approach to figuring out how network structure relates to system behaviours. In recent years, network analysis has become an essential tool in pharmacogenomics [6,7]. By providing a powerful framework to model data, network analysis has allowed researchers to analyse and interpret complex interactions between genes, proteins, and drugs in the pharmacogenomics field. This enables uncovering underlying biological mechanisms, identifying potential drug targets and biomarkers, facilitating drug repurposing efforts, and enabling personalized medicine approaches. In particular, network analysis can identify potential drug targets by constructing biological networks that integrate various data sources, such as protein-protein interactions, gene expression data, and pathway information. Furthermore, by analysing the network topology and identifying key nodes or modules, it is possible to pinpoint genes or proteins that play crucial roles in disease pathways or drug responses [8–10]. This information can guide the development of targeted therapies. Also, network analysis can aid in the discovery of genetic biomarkers that predict drug response or adverse reactions. By integrating genomic and clinical data, researchers can construct networks that capture the relationships between genetic variations, clinical phenotypes, and drug response [11]. Network-based approaches can identify modules or subnetworks that are highly associated with specific drug responses, enabling the discovery of potential biomarkers for personalized medicine. Also, network analysis can uncover the interconnected pathways and biological processes affected by genetic variations or drug treatments. By mapping genetic variants onto biological networks, pathways that are significantly enriched for these variants can be identified [12]. This knowledge helps in understanding the molecular mechanisms underlying drug response and identifying potential targets for intervention.

By analysing the interactions between drugs, genes, and diseases in a network context, potential off-target effects or repurposed drugs for different indications can be revealed [13]. Finally, network analysis can contribute to personalized medicine approaches by integrating patient-specific genetic and clinical data into networks.

Recently, the need to investigate more complicated frameworks than the classical networks has led to the introduction of a multilayer approach as an extension of graph theory. The reason for this is that many real networks cannot be exhaustively explained with a classical network approach, but need more complex structures [14,15]. The introduction of multilayer networks provides a more comprehensive and realistic representation of complex systems where multiple types of relationships coexist. It allows to analyse and understand the dynamics and behaviour of interconnected entities in a more nuanced manner [16].

Multilayer network analysis enables the study of various properties and phenomena that are not easily captured by traditional network analysis approaches. It allows for the examination of interdependencies, correlations, and patterns that emerge across different layers. This can provide insights into how different layers influence each other, the resilience of the system, the spread of information or diseases, and the identification of key nodes or communities in the network.

Starting from these considerations, in this work, we aim to present an application of network analysis in pharmacogenomics to demonstrate how network analysis methods are able to extract hidden relationships and to discover novel knowledge, i.e., identifying key genes in biological pathways involved in drug response and adverse reactions. For this aim, we built a biological multilayer network comprising genes, drugs, diseases, and their associations extracted from a public database. Then, we analysed the multilayer network by applying a community detection algorithm, enabling the identification of essential genes from gene–disease–drug communities. After that, we used the identified list of genes

from the communities to perform pathway enrichment analysis (PEA) to figure out the biological function affected by the selected genes. In particular, the identified genes are detached from their biological context, making it impossible to know in which biological mechanisms and functions they are involved. To understand which biological mechanisms are affected by these communities of essential genes, it is mandatory to link each gene to the opportune biological reference context by means of a pathway enrichment analysis (PEA). PEA links genes and groups of genes to the influenced biological pathways responsible for disease development, adverse drug reactions, as well as the different overall survival rates of patients treated with the same drugs. Thus, the new knowledge allows the development of new treatments that are more effective than drug repositioning strategies, in addition to realizing more adequate drugs for reducing or, even better, eliminating the onset of possible adverse drug reactions.

2. Background on Multilayer Networks

Multilayer networks have emerged as a powerful framework for modelling and analysing complex systems with multiple types of interactions. Unlike classical networks, which only consider one type of relationship between nodes, multilayer networks present the interdependencies between the entities of a system and the interacting layers [17]; see Figure 1 for a complete example.



Figure 1. Examples of classical and multilayer networks. The figure shows two toy examples of a classical biological network (**a**) and a multilayer network (**b**). The first example (**a**) reports three different networks, i.e., a gene–gene interaction network, disease–disease interaction network, and drug–drug interaction network. In the second example, each of those networks represent a distinct layer in the multilayer network. The nodes of the multilayer network are the genes, the diseases, and the drugs, all discriminated by belonging to the respective layer. The *intra*-edges represent the gene–gene, drug–drug, and disease–disease associations, while the *inter*-edges are the gene–disease, gene–drug, and disease–drug associations.

Formally, a multilayer network can be traced back to a set of nodes, edges, and layers that take into account the physical and functional relationships between them [16,18].

In particular, each layer in a multilayer network represents a specific aspect or type of relationship between nodes. For example, in a social network, different layers can represent friendships, professional connections, or family relationships. Each layer can have its own set of nodes and edges, and there can be connections between nodes across different layers.

These networks provide a realistic representation of complex real-world systems, finding their way into practical applications in various domains, such as social network analysis, transport organization, biological systems, and technological networks [15,19]. Formally, a multilayer network graph may be described as a tuple $G_{ml} = V_L$, E_{intra_L} , E_{inter_LxL} , where G_{ml} is multilayer graph, V_L , E_{intra_L} is a set of nodes belonging to each layer, E_{inter_LxL} is a set of edges belonging to each layer, and $L = \{0, 1, ..., l\}$ is a set of layers. For each layer

k, we have a graph V_k , E_{intra_k} (intralayer edges), and for each pair of layers, *k*, *h*, we have a set of edges (interlayer edges) *Einter*_vxk connecting nodes of the layers v and k [20].

Examples of multilayer networks come from many different fields, from social network analysis to biological networks. For instance, Figure 1 represents an example of a biological multilayer network representing the interplay among diseases, genes, and drugs. Multilayer networks provide a powerful tool for analysing complex genetic and clinical data in pharmacogenomics, enabling better predictions of drug response, identification of drug targets, and acceleration of drug discovery and development processes. Multilayer networks can be used for general data analysis in pharmacogenomics. They can integrate and analyse large-scale genomic, transcriptomic, and proteomic data to uncover hidden relationships between genes, proteins, and drug responses. This can lead to a better understanding of the underlying mechanisms of drug response and aid in the development of personalized treatment strategies.

Furthermore, multilayer networks can be used to predict how individuals will respond to specific drugs based on their genetic information. By training the network on a dataset of patients' genetic profiles and corresponding drug responses, it can learn complex patterns and make predictions for new patients. This can help identify individuals who are likely to experience adverse drug reactions or those who are more likely to respond positively to a particular medication. Multilayer networks can help identify genetic markers associated with the risk of adverse drug reactions (ADRs). By integrating genetic and clinical data, the network can identify patterns that link specific genetic variations to ADRs. This information can be used to develop personalized medicine approaches, where patients at higher risk of ADRs can be identified and alternative treatment options can be explored. Multilayer networks can aid in the identification of potential drug targets by analysing genetic data and can assist in drug discovery and repurposing efforts by analysing genetic data and identifying potential drug candidates. By integrating information from various sources such as gene expression, protein-protein interactions, and biological pathways, the network can identify key genes or proteins that play a crucial role in disease development or drug response.

3. Material and Methods

In order to apply multilayer network formalism and pathway enrichment analysis, with the goal to improve knowledge in the pharmacogenomics field, we design a methodology that comprises four steps:

- The building of a biological multilayer network comprising genes, drugs, diseases, and their associations extracted from the BioSNAP database;
- The analysis of the multilayer network by applying a community detection algorithm;
- The identification of essential genes from gene-disease-drug communities;
- Performing pathway enrichment analysis (PEA) to figure out the biological function affected by the selected genes.

Figure 2 summarize all steps.



Figure 2. Methodology workflow.

3.1. Case Study

We considered the following datasets from the Stanford Biomedical Network Dataset Collection (BioSNAP) [21]:

- 1. *Drug–Drug Interaction (DrDrI)* network of interactions between drugs, approved by the U.S. Food and Drug Administration (FDA): 1514 nodes and 48,514 edges.
- 2. *Disease–Disease (DD)* network of interactions between 6878 inherited nodes and 6877 inherited edges.
- 3. *Gene–Gene (GG)* network of interactions between in 25,825 inherited nodes and 208,836,746 inherited edges. The nodes are given by NCBI Entrez Gene IDs.
- 4. *Disease–Drug Association (DDrA)* network, a set of curated relationships between diseases and drugs: 5535 disease nodes, 1662 drug nodes, and 466,656 edges. The diseases are given by DOIDs, i.e., Disease Ontology terms.
- 5. *Gene–Disease (GDA) Association* network, a set of relationships between genes and disease: 7294 gene nodes, 519 disease nodes, and 21,357 edges.
- 6. *Gene–Drug Interaction (GDrI)* network, a set of relationships between genes and drugs: 3648 gene nodes, 284 drug nodes, and 18,690 edges.

We build a multilayer network with three layers obtained from the DDI, DDr, and GG databases. Then, we add interlayer edges by considering the DDrA, GDA, and GDrI databases. Finally, the resulting multilayer network, that, for convenience, we called the GDD multilayer network, consisted of 52,640 nodes and 208,892,137 interactions, of which 506,703 *inter*edges exist. At first, we performed a topological analysis on the GDD multilayer network. The network analysis was performed using the multinet R package [22] (for complete details on multilayer network analysis, see [22]). Table 1 summarizes topological measures computed using GDD on the multilayer network for each layer. Table 2 summarizes topological measures computed using GDD on the multilayer network for layer comparison. The first part of the value indicates the type of comparison function (Jaccard, Coverage, Simple Matching, Russell Rao, Kulczynski, Hamann), and the second part indicates the configurations to which the comparison function is applied.

Network Measure	Layer 1	Layer 2	Layer 3
similarity between layer summaries	1.126	4.936	4.888644
layer min degree	1	1	1
layer max degree	89	443	696
layer mean degree	1.999	64.087	68.301
layer sd degree	3.239	68.339	90.953
layer skewness degree	11.373	1.5809	2.187
layer kurtosis degree	221.854	5.724	8.888
layer entropy.degree	1.126	4.936	4.888
layer CV degree	1.619	1.066	1.331
layer jarque bera degree	13,874	1098.965	4780.306

Table 1. Topological measures computed using GDD on the multilayer network for each layer.

Table 2. Topological measures computed using GDD on the multilayer network for layer comparison.

Network Measure	Value
Jaccard actors	0.268
Jaccard edges	0.2
Jaccard triangles	0.01
coverage actors	0.34
coverage edges	0.2
coverage triangle	0.1
sm actors	0.658
sm edges	0.999
sm triangles	1

Network Measure	Value
rr actors	0.061
rr edges	8.523 exp −5
rr triangles	$3.064 \exp{-0.9}$
Kulczynski2 actors	0.3374
Kulczynski2 edges	0.2
Kulczynski2 triangles	0.1
Hamann actors	0.317
Hamann edges	0.998
Hamann triangles	0.999

Table 3 summarizes the distribution dissimilarity computed using GDD on the multilayer network (notice that these are dissimilarity functions: 0 means the highest similarity) Table 4 summarizes the statistical degree correlations computed using GDD on the multilayer network.

Table 3. Distribution dissimilarity computed using GDD on the multilayer network.

Network Measure	Value
dissimilarity degree	0.232
KL degree	0.57
Jeffrey degree	0.949

Table 4. Statistical degree correlation computed using GDD on the multilayer network.

Network Measure	Value
Pearson degree	0.153
rho degree	0.220

3.2. Community Detection on GDD Multilayer Network

Once built, we analyse the GDD multilayer network by applying one of most useful exploratory technique for network analysis, i.e., community detection. Community detection is considered a first step in understanding network analysis and community structures, defined as groups of nodes that are more densely connected than the rest of the network, and represent significant characteristics for understanding the functionalities and organizations of complex systems modelled as networks. Thus, community extraction provides the identification of densely connected nodes within multilayer networks that play significant roles in the structure–function relationship. For this study, we selected Infomap [23] because, according to the literature, it outperforms other community detection methods for multilayer networks [24].

Then, we applied Infomap on the GDD multilayer network, obtaining 153 communities. Infomap extracted three typologies: (i) communities containing genes, diseases, and drugs; (ii) communities containing diseases and drugs; and (iii) communities containing genes. For our aims, we focus on the first typology of communities containing genes, diseases, and drugs. Then, we selected the top 10 communities, i.e., the communities comprising interlayer relations, for example, gene–drug and gene–disease relations. In Table 5, we reported the list of genes belonging to the top 10 communities.

Table 2. Cont.

Table 5. Top ten communities.

Community	Genes
1	P48169 Q15822 P08172 P08908 P25100 P28222 P19320 Q01959 O15399 Q9NYX4 P48051 P32297 P23560 P17787 P01189 P47870 P31645 P08173 O00264 P30939 P54219 P41145 Q9H3N8 P28223 P07550 P18825 P41143 Q12879 P28566 O60391 O14764 Q05586 P13945 P28476 P11229 P42263 O14732 P28472 P08912 O60858 Q8N910 Q13002 P30926 P28221 P08913 P21728 P34903 P14867 P48058 Q9UK17 Q16445 P23416 P46098 P17405 Q09470 P42262 P02708 P41595 P23415 P18507 P18505 P30049 P43681 O75311 Q8TCU5 Q9UGM1 Q96RJ0 Q9UHC3 P50406 Q15825 P02763 P28335 P19652 Q13224 P31644 P48549 Q99928 Q9NPC2 A5X5Y0 Q9GZZ6 P42261 P35368 P03886 P48167 P24046 P41146 Q9Y2I1 Q05901 Q99720 A8MPY1 P30536 P21918 Q9UN88 P35367 P20309 Q05940 P14416 P08588 O95264 P78334 O14649 P34969 Q9NZV8 Q8N1C3 P47898 P35348 P18089 P35372 P35462 Q14957 P23975 P47869 P36544 P23763 P21917 P30988 O00591 Q8WXA8 P20711 P98194 Q9Y5N1 Q70Z44 P13498 P30532 O94956 Q14542 P630271
2	Desited General Quick Operation Quick Quick Operation Quick Qui
3	DB01611 P17900 O60603 O75116 P01308 O00206 P17612 Q9NR96 P61925 Q9NYK1 P31749 Q04771 P43080 Q13464 Q9Y6Y9 P31751 P62942 P49841 Q08209
4	P14555 P02788 P04054 Q16706 P63208 Q9UKM7 DB03414 Q9NZK7
5	DB03880 Q9ULZ9 P09238 DB01197 Q9UKQ2 P39900 P08254 P09237 P14780 P03956 P51512 P08253 Q8N119 Q9UNA0 O60882 O60882 Q9Y5R2 Q9H306 P51511 P24347 Q9NPA2 P50281 P45452 P22894

Table 5. Cont.

Community	Genes
6	P01133 P04818 Q8NBP7 P05106 P54760 Q7LG56 Q99808 Q99062 P27707 P33151 P05091 P09871 O00142 P12318 P15309 Q16552 P32321 P06746 P12821 P30085 P26358 P62993 P00797 P31785 P14784 P02747 Q9NRF9 P56282 Q07864 P16220 P16066 P31994 Q05932 P08514 P00533 P36952 P00736 P24385 P19971 P04234 P32320 P12314 P12314 P02746 P20594 Q9UNI1 P01031 P05186 P78559 P01589 P15391 P31350 P08473 P24158 Q03393 P11836 P20701 O60493 Q9BYF1 P08246 Q16854 P23919 P22413 P19235 Q15303 P04183 O14788 P22102 P02745 Q92820 P31995 Q9NNW7 P31939 P08637 P0CG22 P04626 O75015 P00813 Q9H252 P23921 O00764 P00374 P04229 P0C0L5 P09884 P17342
7	P22303 P21964 Q92952 DB06218 P06276 Q86W47 Q9UQD0 P21397 P27338 Q16558 Q9NY46 P07686 Q9Y5Y9 Q9H2S1 Q9NPA1
8	TP61457 Q9Y619 P05089 P29475 P78540 Q8WY07 P50440 Q9BX12 P00439 P13716 P52569 Q15046 O95190 P35228 P01270 Q96A70 P00480 P04424 O43246 P30825 P29474 P54368 Q9UMX2 P20823 P35222
9	P35916 P20648 P49286 P46059 P10636 P16234 P06133 P09172 P11473 P33260 P16444 Q8TCC7 P02768 O15554 Q9Y6L6 Q9UM07 P24530 Q15858 Q9HCR9 Q9BQB6 P01023 P25021 P11712 P04629 P10635 O15111 P13569 P33402 Q9NPD5 Q16696 P31513 Q9Y5Y4 P54855 Q12809 P15538 Q6VVX0 Q9NSA0 P31639 P48039 Q9H244 P50225 P43116 Q15166 Q13956 Q969P6 Q92769 P05181 P08684 Q99250 P22309 Q4U2R8 P33261 Q9HB55 P09619 Q16853 P25024 Q9UI33 P19835 Q02928 P35499 P10632 P23219 O76074 P43115 DB00192 Q14524 Q16850 Q92753 P16662 P18440 P30711 P30556 Q9UQQ2 P18545 Q9UHC9 O43923 Q14123 P15382 P08174 Q08345 P04156 P09086 P15502
10	A 51X70 A 971YG A 921W8 A921W9 80FP48 B01712 B0(YP2

3.3. Pathway Enrichment Analysis

Pathway enrichment analysis (PEA) helps researchers comprehend the biological meaning of gene lists obtained from high-throughput experiments, such as RNA sequencing, genome-wide association studies, or proteomics. These experiments identify genes, including proteins and metabolites, that differ between the conditions of interest. However, this gene list alone is insufficient to understand the biological differences between these conditions. Therefore, PEA assists researchers in interpreting large gene lists and developing hypotheses about the underlying biology [25].

To identify the biological mechanisms and/or functions affected by the identified communities of genes, we used the Reactome pathway database [26]. In particular, we describe the enrichment performed using the communities with identifier 10 using the software tool BiopaxParser (BiP-v.1) [27].

Table 6 reports the enriched pathways using the list of proteins belonging to community 10.

Table 6. The first 10 enriched pathways, in order of statistical relevance, obtained using the gene list of community 10 as input data.

	Pathway Name	p Value	FDR Correction	Bonferroni Correction
(1)	Olfactory Signaling Pathway	1.25E-07	1.54E-04	1.54E-04
(2)	Metabolism of proteins	1.06E-05	0.007	0.013
(3)	Post-translational protein modification	1.16E-05	0.005	0.014
(4)	Leishmania parasite growth and survival	6.71E-05	0.021	0.082
(5)	Anti-inflammatory response favouring Leishmania parasite infection	6.71E-05	0.016	0.082
(6)	Signaling by Rho GTPases, Miro GTPases and RHOBTB3	6.83E-05	0.014	0.084
(7)	Signaling by Rho GTPases	7.20E-05	0.013	0.089
(8)	TCF dependent signaling in response to WNT	9.41E-05	0.014	0.116
(9)	Signaling by WNT	1.17E-04	0.016	0.143
(10) Transcriptional regulation by RUNX1	1.20E-04	0.015	0.148

Next, we used BiP to know which pathways are influenced for each input gene. Table 7 presents the relation between genes and affected pathways. Inside community 10, a total of 36 genes are a member of layer 5, e.g., the disease–gene layer, and layer 6, e.g., the drug–gene layer, revealing the multiple roles of a gene through different pathways. Analysing Table 7, it is worth noting that the activity of the *metabolism of proteins* pathway, a well-known pathway related to adverse or normal drug responses, as well as to disease progression or decline, is regulated by the interactions of more multilayer genes, namely *P43088*, *P15170*, *P18509*, *P05546*, *Q9Y277*, *Q02817*, *Q13285*, *O75976*, and *P15328*, reinforcing the benefits of the multilayer formalism to represent complex networks.

Table 7. The 20 multilayer genes and their affected biological pathways.

Gene Name	ID	Pathways
СҮТН3	O43739	Vesicle-mediated transport; Membrane trafficking; Intra-golgi and retrograde golgi-to-ER traffic
GSPT1	P15170	Translation; Metabolism of RNA; Metabolism of proteins
NDUFA9	Q16795	Respiratory electron transport; Respiratory electron transport, ATP synthesis via chemiosmotic coupling, and heat production by uncoupling proteins; Complex I biogenesis
ADCYAP1	P18509	Signalling pathways; Metabolism of proteins
SERPIND1	P05546	Haemostasis; Metabolism of proteins; Post-translational protein modification
SNRPA	P09012	mRNA splicing; pre-mRNA splicing; Metabolism of RNA; Processing of capped intron-containing pre-mRNA
VDAC3	Q9Y277	Metabolism of proteins; Post-translational protein modification
FDX1	P10109	Diseases of metabolism; Disease

Gene Name	ID	Pathways
NR5A1	Q13285	Post-translational protein modification; Gene expression (Transcription); Metabolism of proteins
MUC2	Q02817	Disease; Diseases of metabolism; Defective C1GALT1C1 causes TNPS; Termination of O-glycan biosynthesis; O-linked glycosylation; O-linked glycosylation of mucins; Metabolism of proteins; Post-translational protein modification
NDUFA7	095182	Respiratory electron transport; Respiratory electron transport, ATP synthesis via chemiosmotic coupling, and heat production by uncoupling proteins; Complex I biogenesis
LIMK1	P53667	Thrombin signalling through proteinase activated receptors (PARs); GPVI-mediated activation cascade; G alpha (12/13) signalling events; Signalling Pathways; Disease; Signalling by Rho GTPases, Miro GTPases, and RHOBTB3
NDUFC1	O43677	Respiratory electron transport; Respiratory electron transport, ATP synthesis via chemiosmotic coupling, and heat production by uncoupling proteins; Complex I biogenesis
CPD	075976	Signalling Pathways; Metabolism of proteins; Signalling by Rho GTPases, Miro GTPases, and RHOBTB3; Signalling by Rho GTPases; RHO GTPase cycle
РССВ	P05166	Metabolism of vitamins and cofactors; Biotin transport and metabolism
FOLR1	P15328	Metabolism of proteins; Post-translational protein modification; Vesicle-mediated transport; Membrane trafficking; Asparagine N-linked glycosylation; Transport to the golgi and subsequent modification; ER-to-golgi anterograde transport; COPI-mediated anterograde transport

Table 7. Cont.

3.4. Results and Discussion

Pharmacogenomics is a complex field where the drug response of the living organism is due to the interactions of several different biological entities like genes, enzymes, and small and large molecules that cooperate in a synchronized fashion to accomplish the task. Multilayer network representation allows for more comprehensive and realistic modelling of these heterogeneous interactions than traditional ones [28]. In addition, multilayer networks enable the identification of multilayer communities that are a bunch of genes more densely connected among them and the correlations through the different layers: information that can be used to perform PEA to comprehend the affected underlying biological mechanisms.

Performing PEA using the detected gene communities from layers 10 enriches several biological pathways, as reported in Table 6. Analysing the content of Table 6, it is worth noting that the enriched pathways present multiple intertwinements among them, some of which are more explicit than others. We conducted a literature search to explore the possible connections between the results of the protein enrichment analysis. According to Bhardwaj et al. [29], leishmania alters various signalling pathways to survive, which is in line with the other enriched pathways 6, 7, 8, 9. Additionally, Kaiser [30] found that cyclic nucleotides such as cAMP and cGMP are crucial for parasitic proliferation and regulate functions such as auditory and olfactory senses [31]. Also, Rho GTPases play a role in host-pathogen interaction by controlling innate and adaptive immune responses. Pathway 1 in Table 6 is another signalling pathway that leishmania affects, as described in [31]. Moreover, Schlessinger et al. [32] explain the vital role of the mediator of Rho GTPases in the WNT signalling pathway. Finally, Kikuchi et al. [33] describe the regulation of WNT signalling pathways through post/translation modifications, while Li et al. [34] provide details on the role of RUNX1 in promoting tumour metastasis by activating WNT.

Table 7 clearly displays the association between genes and pathways, emphasizing that a single gene can be involved in multiple pathways. The enrichment was calculated by implicitly incorporating topological and structural network properties, resulting in improved enrichment outcomes, as opposed to using more general genes, as described in [27].

In Figure 3, community 1 is represented as a network. The green nodes with red labels in the network correspond to the genes listed in Table 7. These genes play a crucial role in the network's connectivity and are known as **hub genes** in the literature. For instance, if we

remove *O95182*, *P15328*, and *P09012*, the network loses its complete connectivity. To learn more about the role of the three hub proteins, we searched the Reactome database. We used the Reactome web pathway browser and found out that the three hub proteins are a part of the metabolism pathway. Specifically, all three proteins have an impact on the citric acid (TCA) cycle and respiratory electron transport pathway. Moreover, proteins P09012 and O95182 regulate respiratory electron transport, ATP synthesis via chemiosmotic coupling, and heat production by uncoupling proteins pathway. This highlights the importance of multilayer modelling and enables the selection of more relevant genes from the network for performing PEA.



Figure 3. Interactions among genes in community 2. The figure displays a network diagram depicting the interactions among the genes that belong to community 2.

In addition, Table 7 includes some genes that are not part of community 1, featured in Figure 4.

If we rely solely on a traditional network representation, we may overlook crucial information, such as the fact that gene *P18509* does not belong to community 1. However, through a multilayer representation, we can observe that gene *P18509* interacts with *P10109*, as illustrated in Figure 3.

The reason for this is that both genes affect the same category of pathway related to the metabolism. In conclusion, the use of multilayer networks to represent interactions among heterogeneous data is a novel approach, especially in the field of omics. A multilayer approach can help researchers capture more information and obtain a more accurate understanding of gene interactions. In the literature, Shang et al. in [28], propose a multilayer network representation learning method for predicting drug-target interactions. This method integrates information from different networks, reduces onise, and learns the feature vectors of drugs and targets, overcoming the challenges of integrating multiple data types and managing network noise. Using a multilayer network to infer new relationships among genes, diseases, and drugs is at its early stage and is a continuously developing field. This limits the possibility of validating the proposed method by comparing it with existing methodologies. Using the proposed method, we discovered potential new relationships between leishmania and different signalling pathways: results possible only through multilayer representation. This could help researchers to identify drugs targeting specific biological functions affected by the enriched pathways. Investigating leishmania is particularly important in the context of travel medicine. Berman reviewed several aspects



of diagnosis and treatment for leishmania in [35]. With our method, we could determine which drugs could contrast the damage caused by leishmania infection.

Figure 4. Interaction among genes in community 1. The figure displays a network diagram depicting the interactions among the genes that belong to community 1. In the network, green nodes with red labels indicate the genes listed in Table 7.

4. Conclusions

In this work, we explore the application of network analysis in the pharmacogenomics field. In particular, we used multilayer network representation to model the interaction among genes, drugs, diseases, and their associations. Then, we analysed the network by applying a community detection algorithm to discover the top communities. Finally, we used the identified list of genes from the communities to perform pathway enrichment analysis (PEA) to figure out the biological function affected by the selected genes. The results demonstrate that the genes forming the communities extracted from the multilayer network regulate the activity of the *protein metabolism* pathway related to adverse or normal drug response, as well as the progression or decline of disease, demonstrating the advantages of multilayer formalism to represent pharmacogenomic domains.

Author Contributions: Conceptualization, M.M. and G.A.; methodology, M.M. and G.A.; software, M.M. and G.A.; data curation, M.M. and G.A.; writing—original draft preparation, M.M., G.A. and M.C.; writing—review and editing, M.M., G.A. and M.C.; funding acquisition, M.C. All authors have read and agreed to the published version of the manuscript.

Funding: This work was funded by Next Generation EU, Italian NRRP, Mission 4, Component 2, Investment 1.5, call for the creation and strengthening of 'Innovation Ecosystems', building 'Territorial R&D Leaders' (Directorial Decree n. 2021/3277), and project Tech4You—Technologies for climate change adaptation and quality of life improvement, n. ECS0000009. This work reflects only the authors' views and opinions; neither the Ministry for University and Research nor the European Commission can be considered responsible for them.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

The following abbreviations are used in this manuscript:

PPI	Protein–Protein Interaction
ROI	Regions Of Interest
ADRs	Adverse Drug Reactions
PEA	Pathway Enrichment Analysis
GDD	Gene–Drug–Disease
BiP	BiopaxParser

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