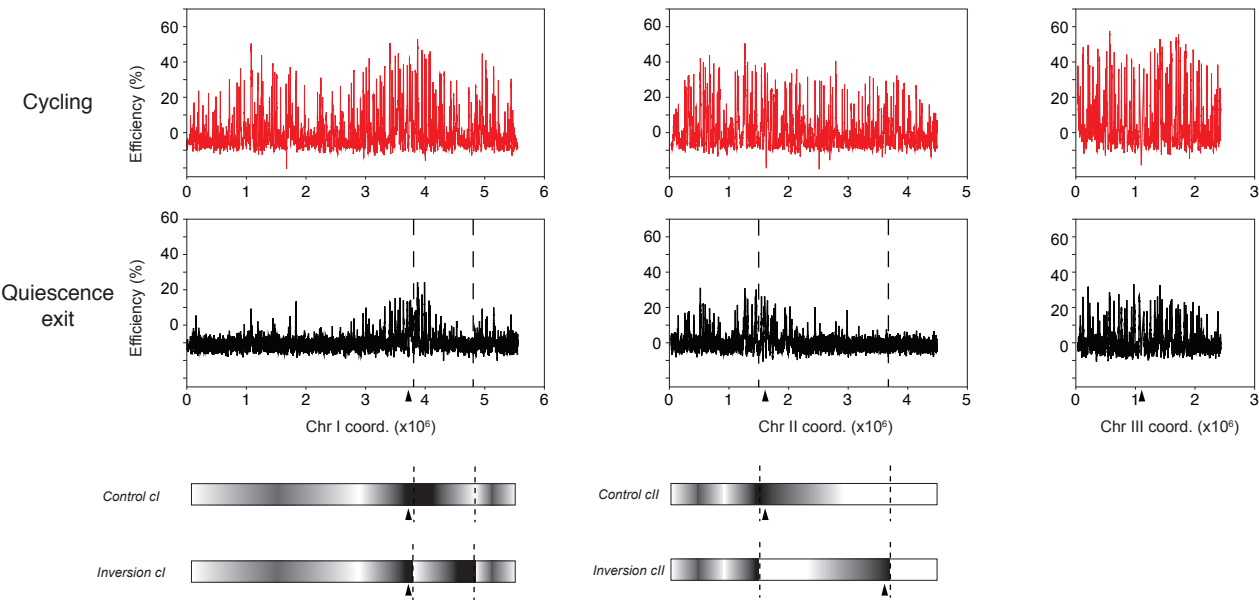


Figure S1

A



B

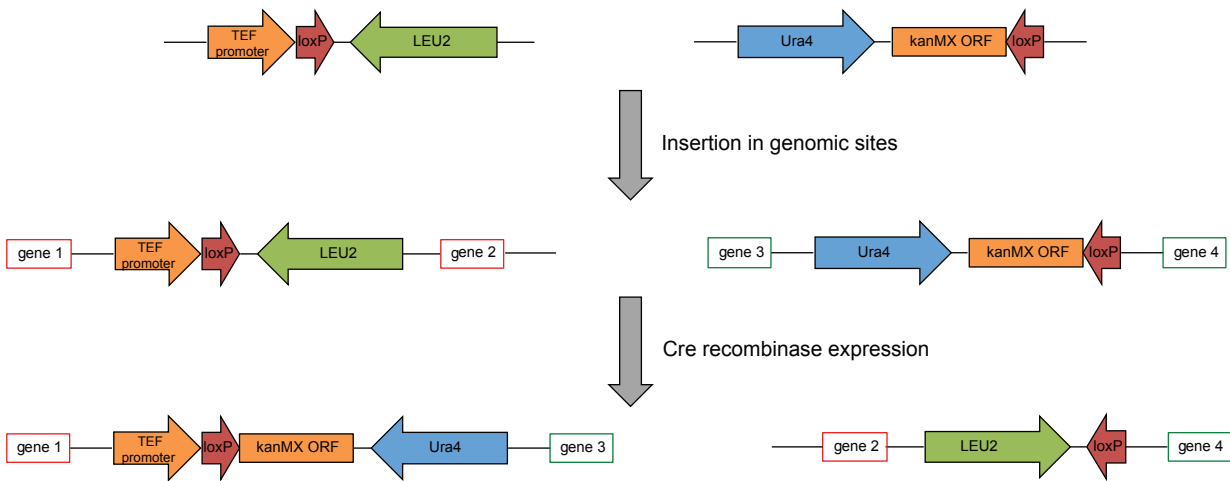
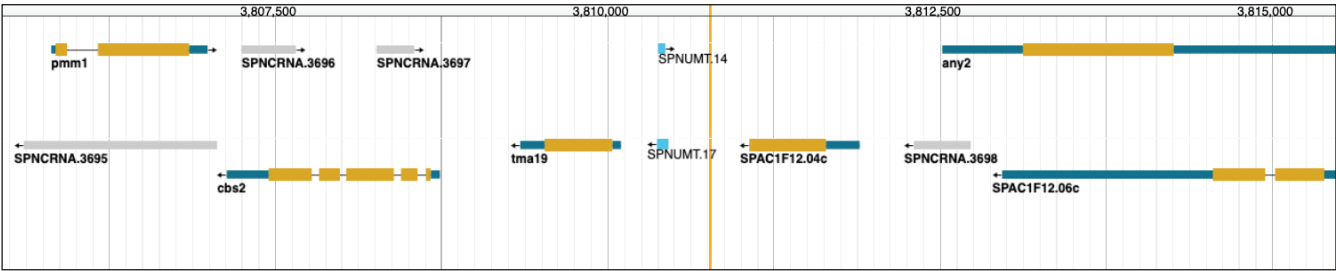


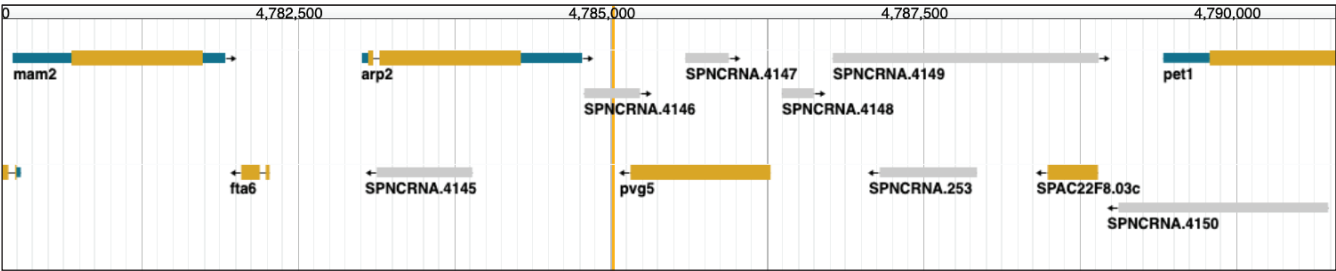
Figure S1

C

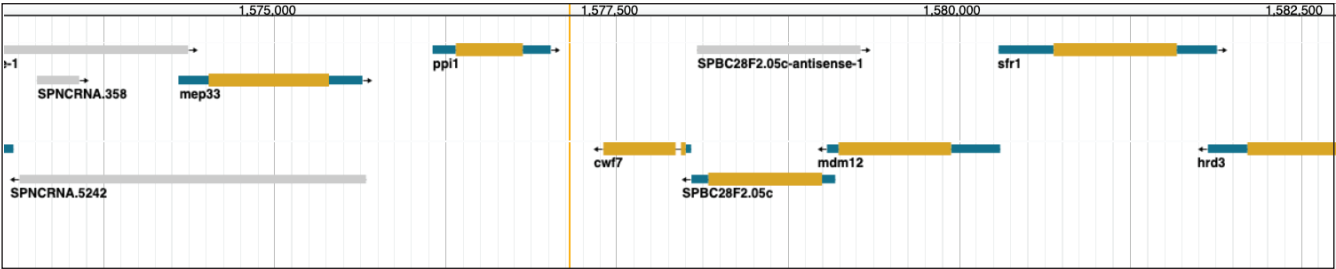
ChrI: 3810766



ChrI: 4785016



ChrII: 1577150



ChrII: 3694900

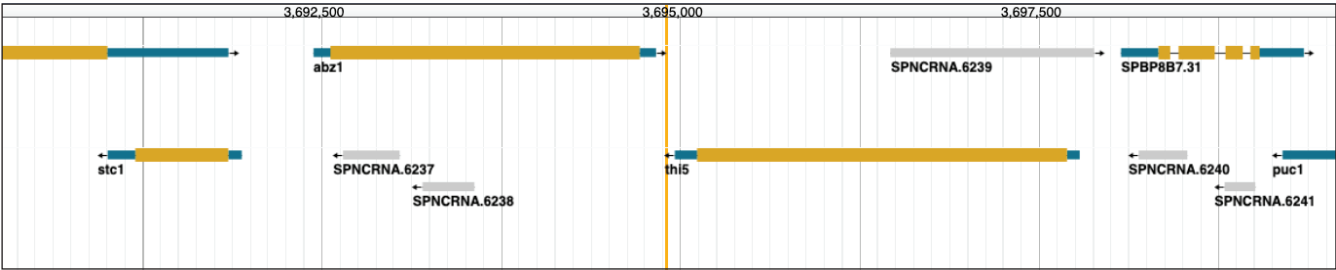
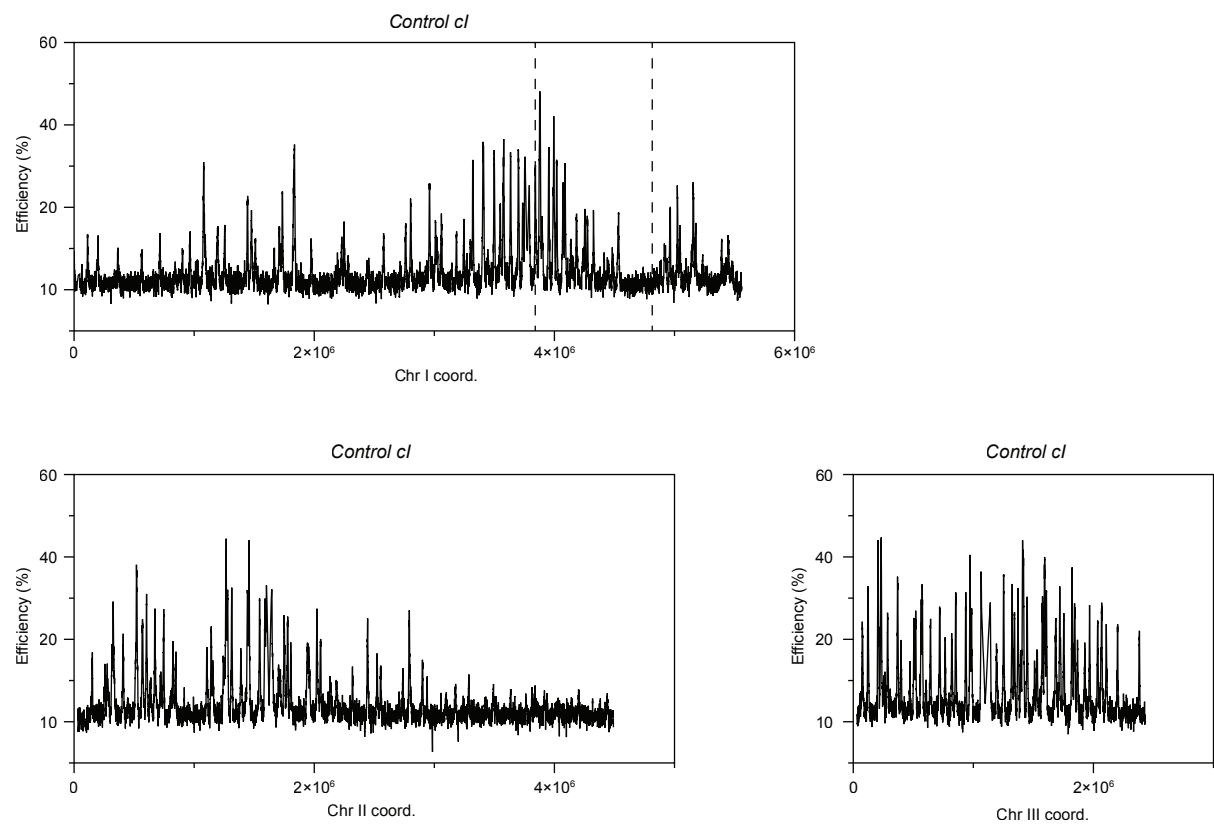


Figure S2

A



B

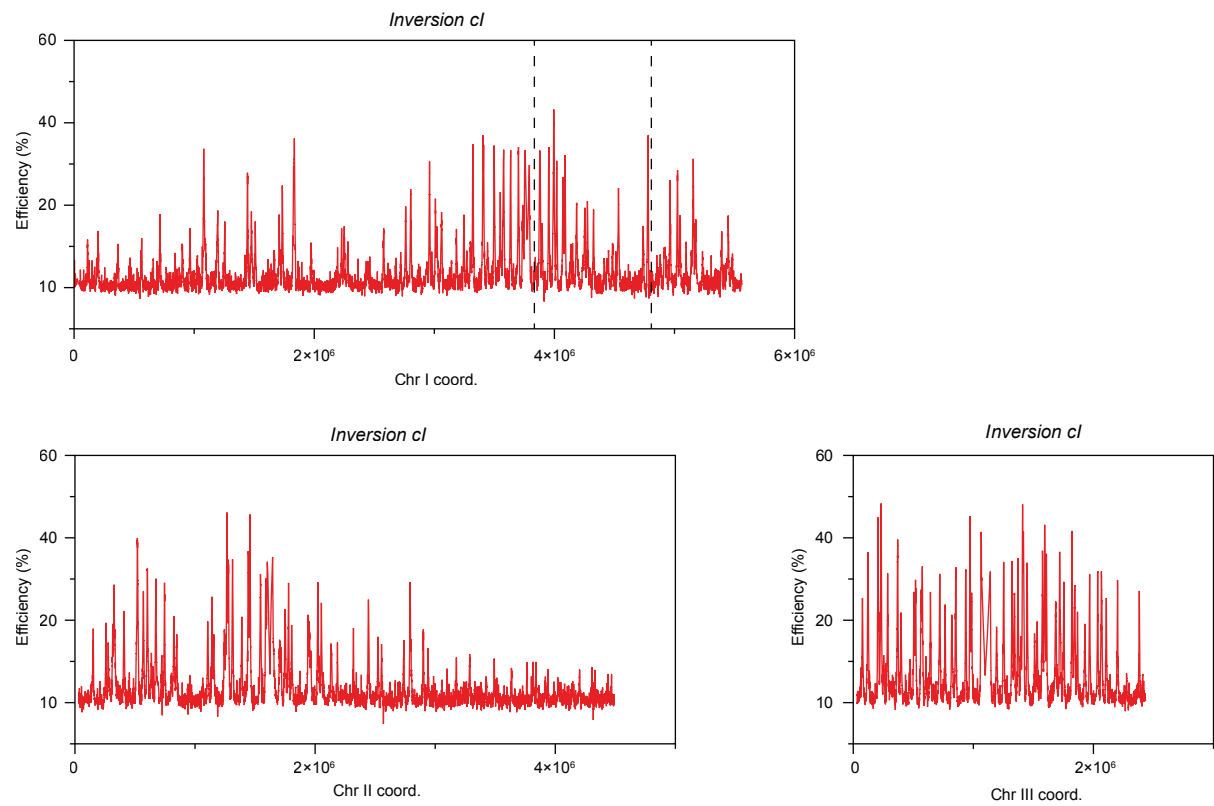


Figure S2

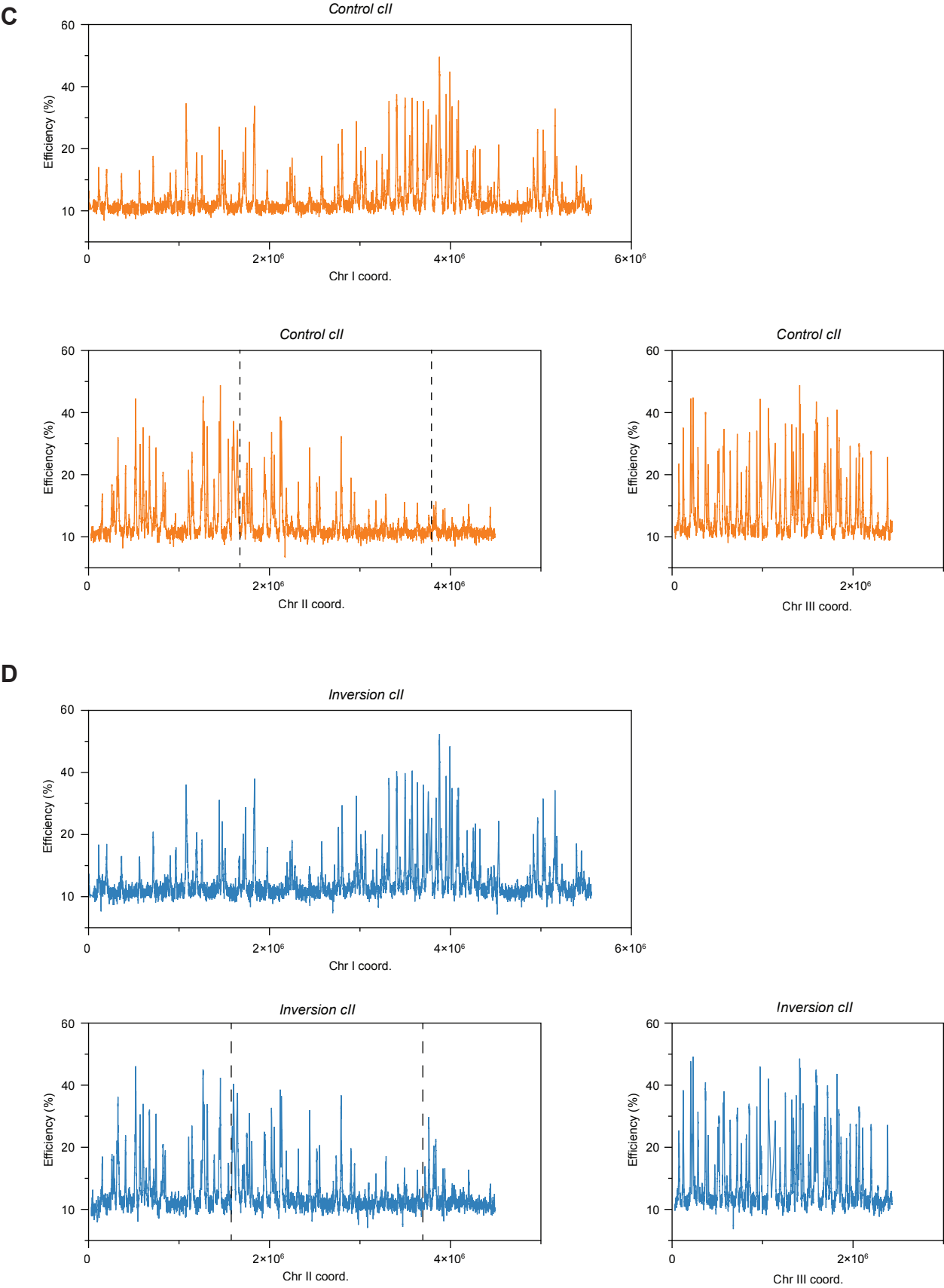
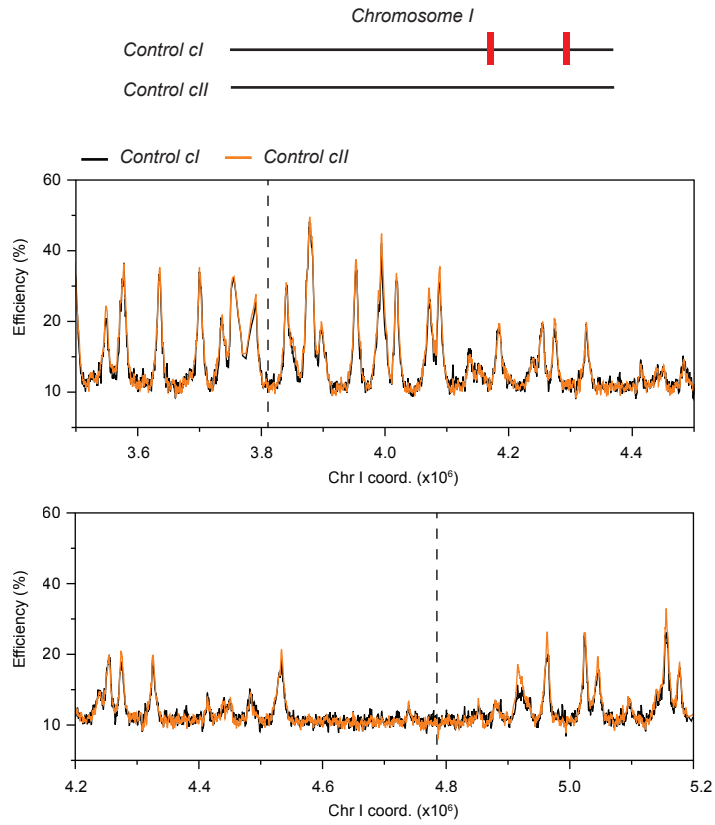
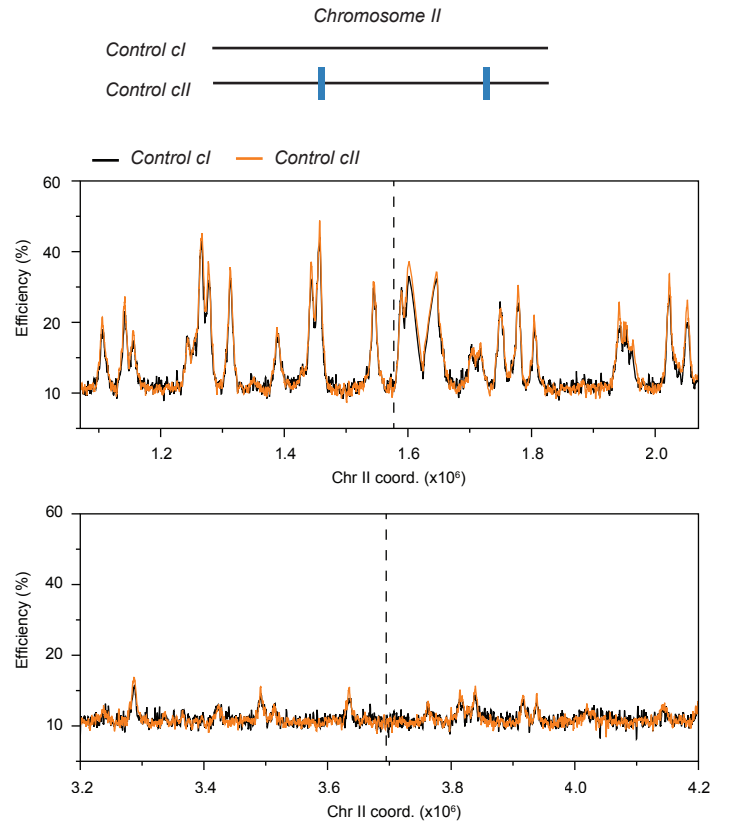


Figure S3

A



B



C

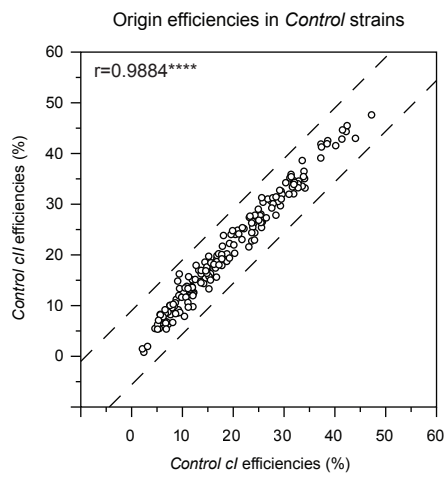


Figure S4

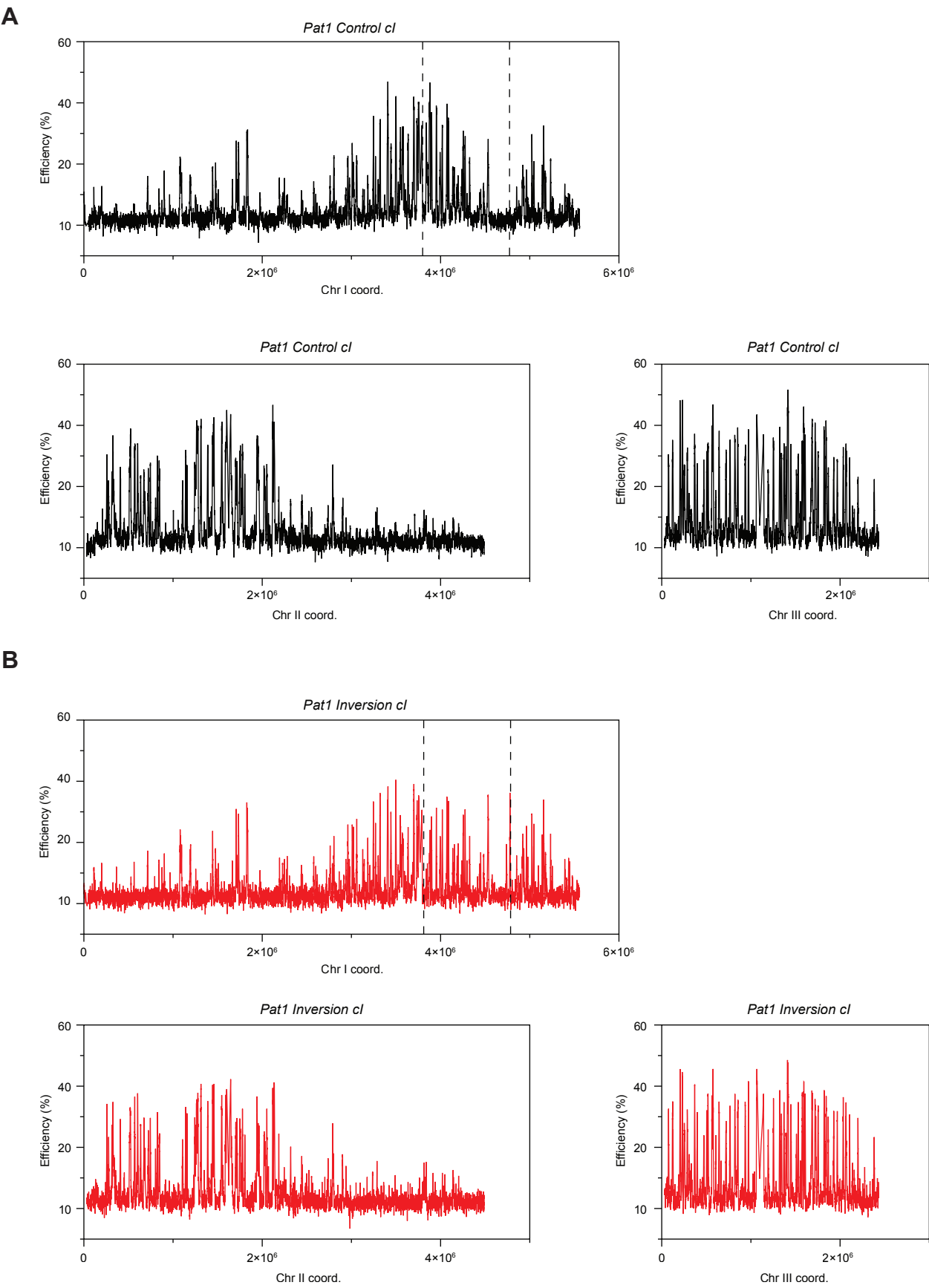
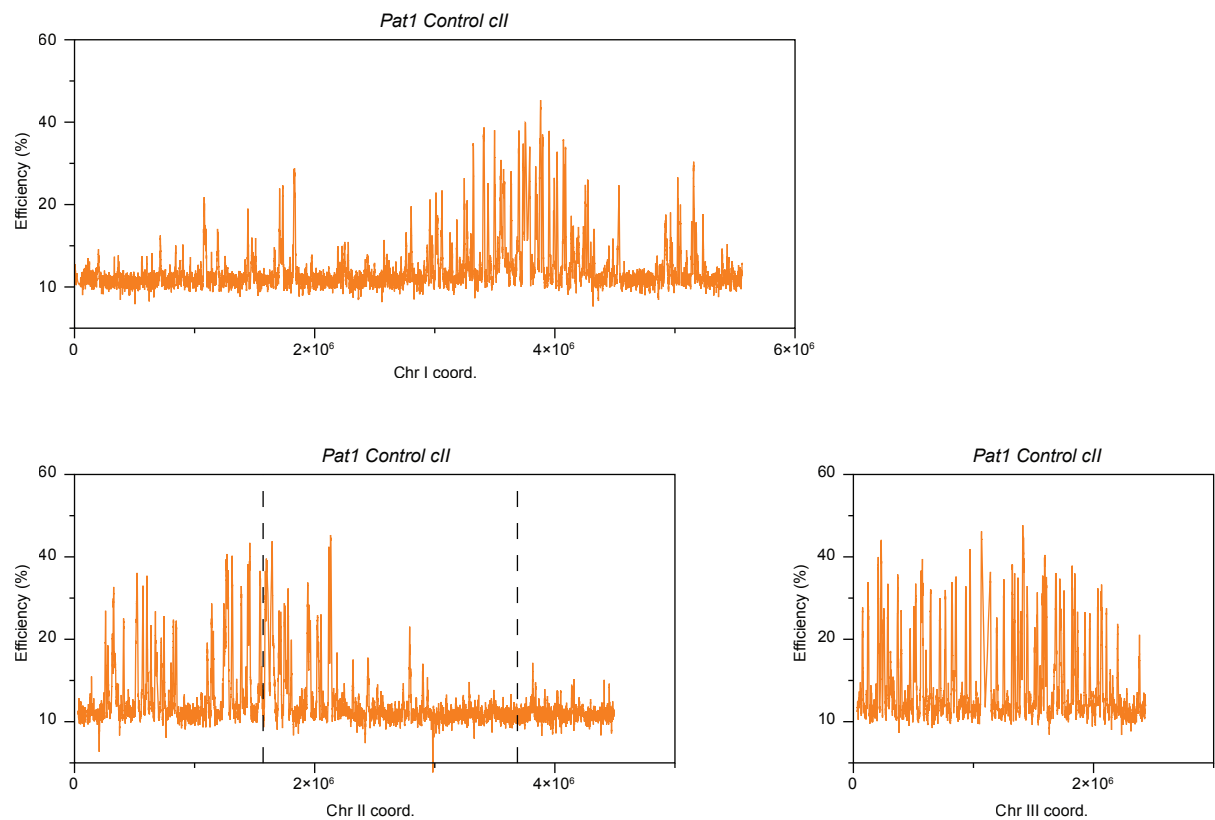


Figure S4

C



D

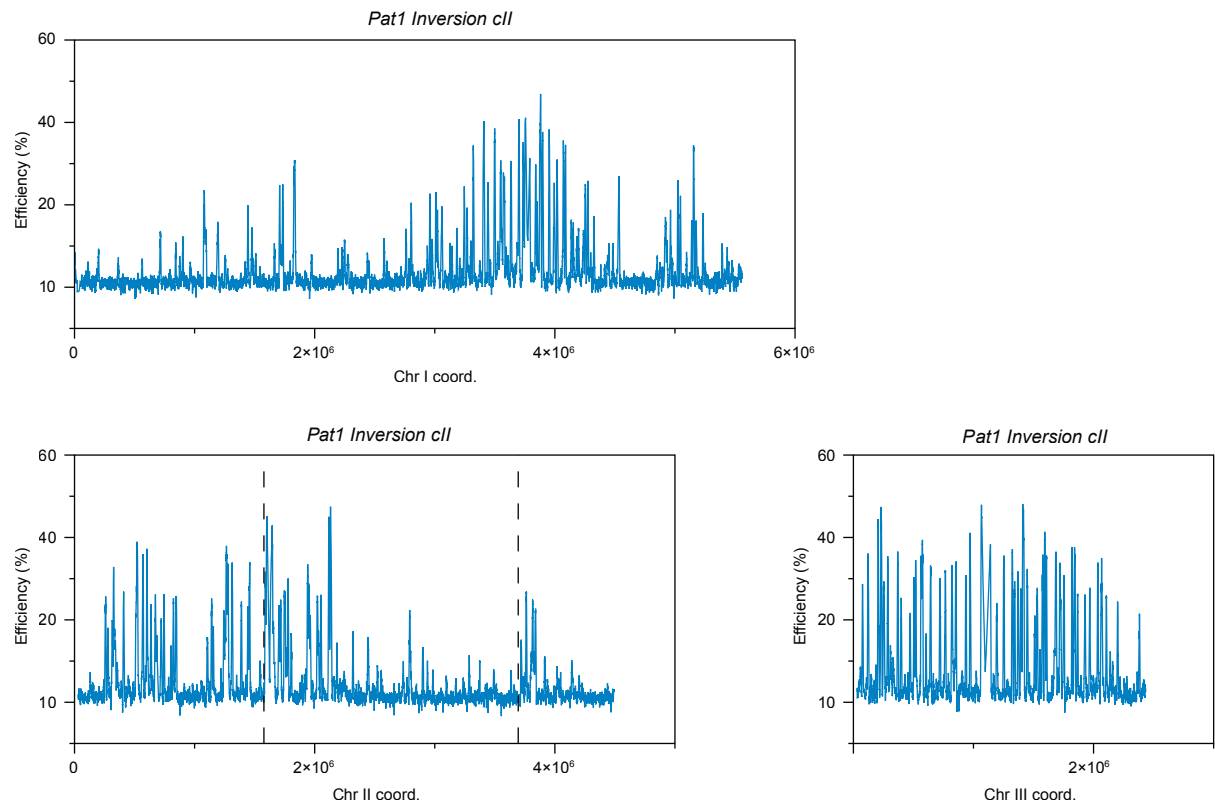


Figure S5

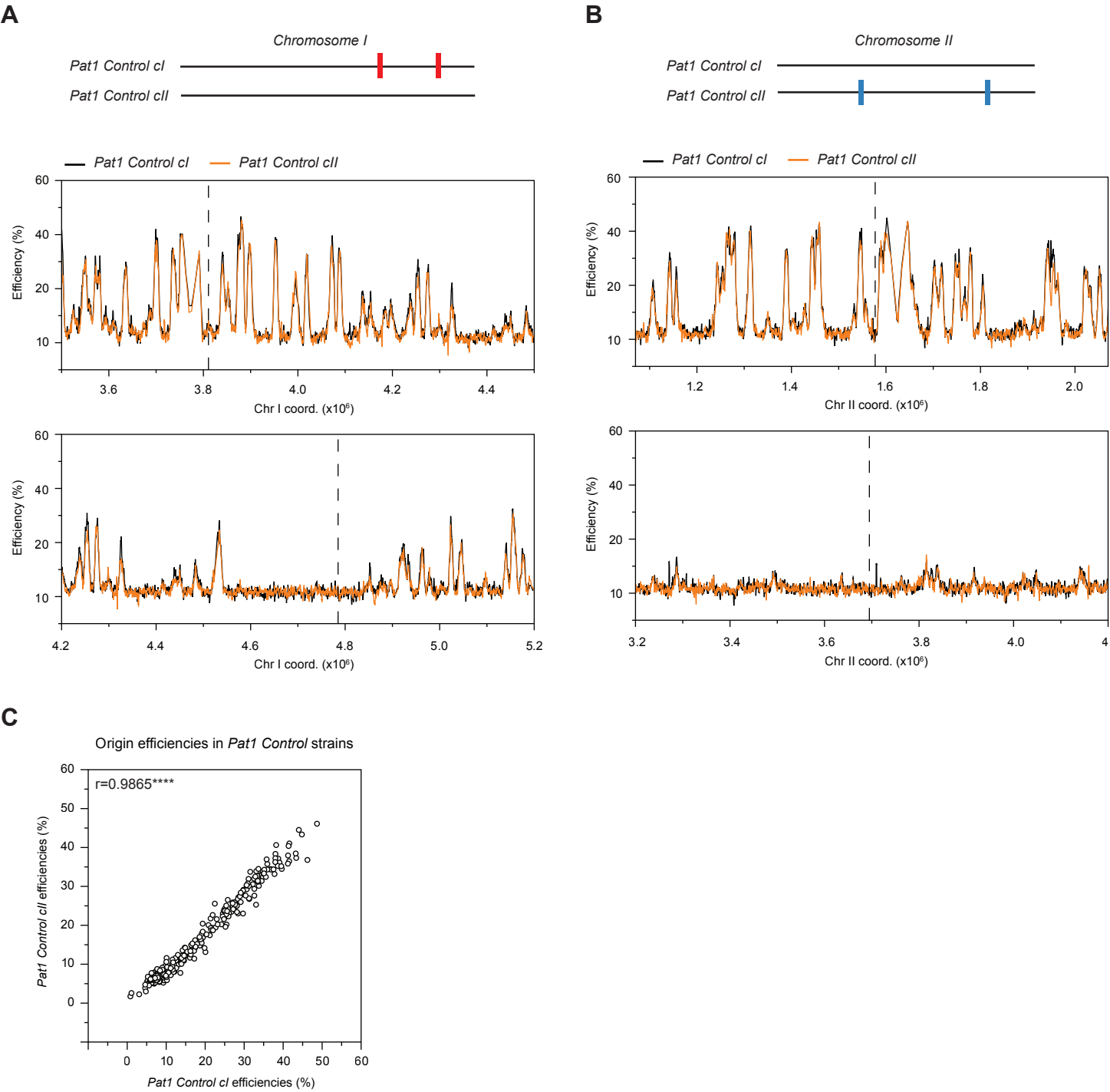
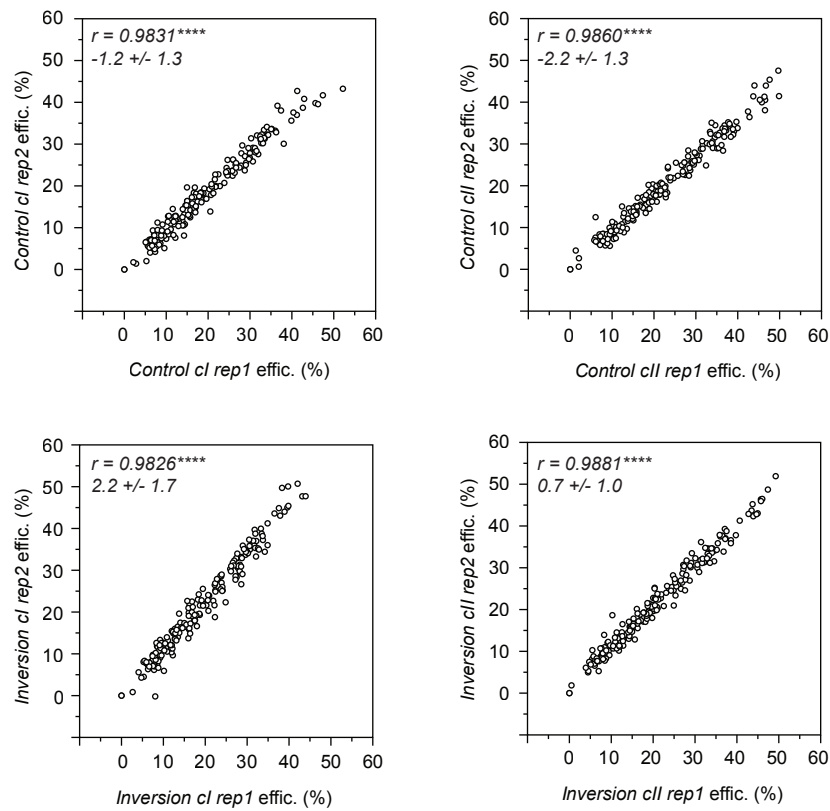


Figure S6

A



B

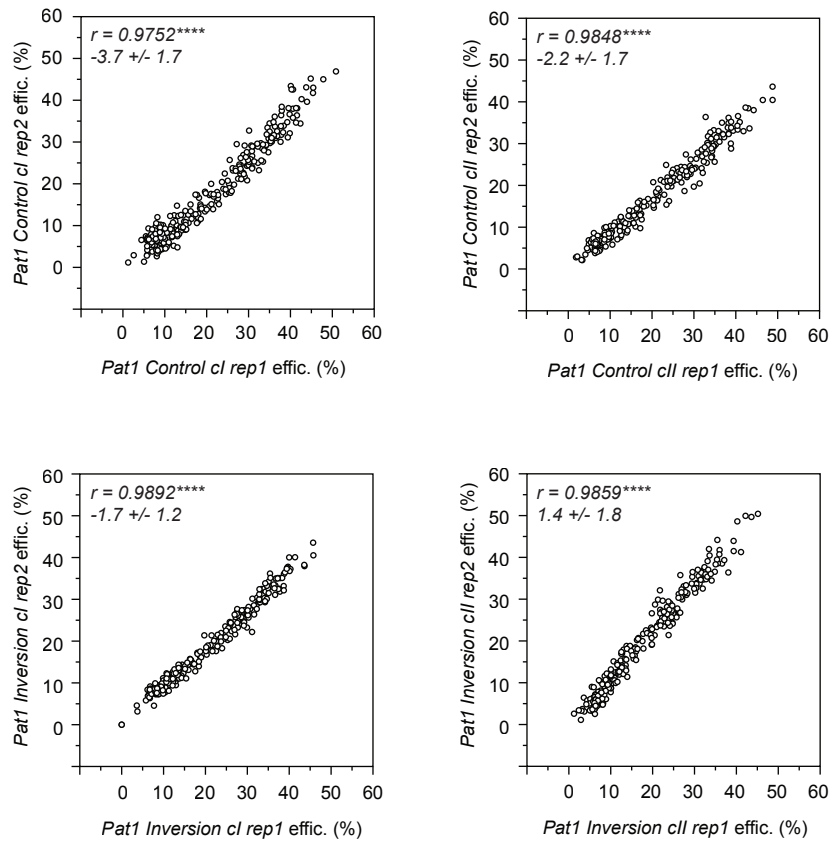
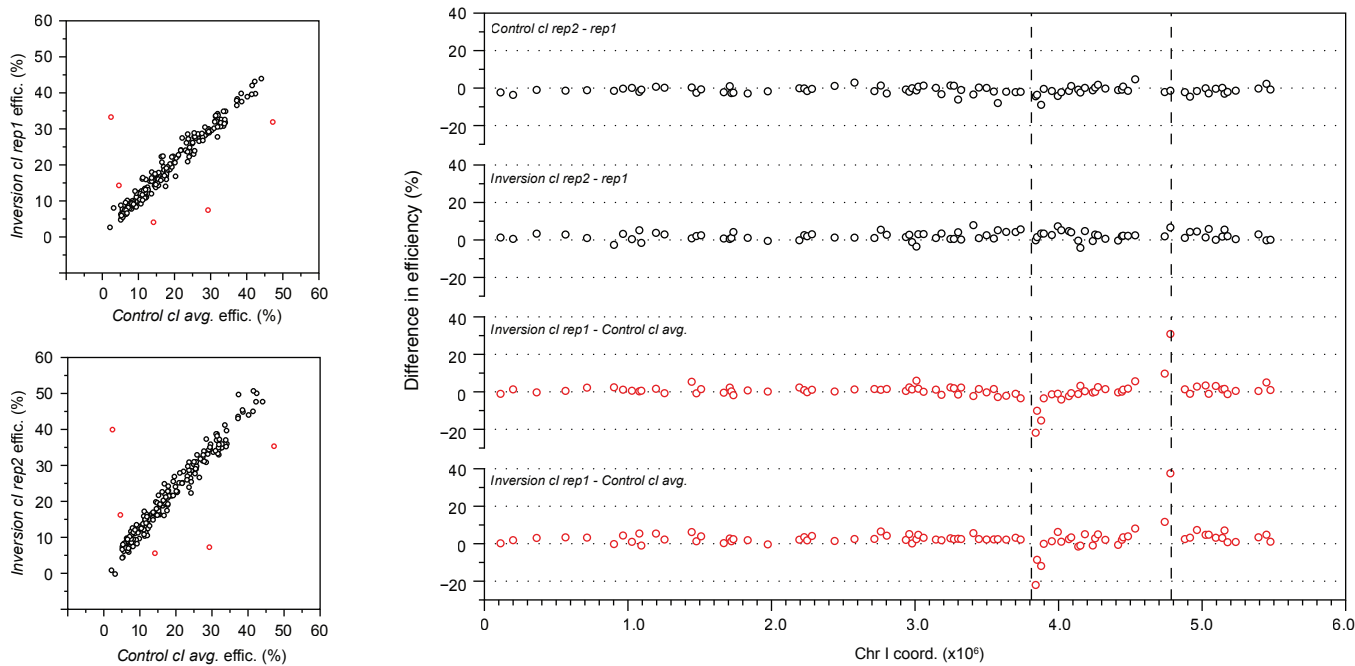


Figure S7

A



B

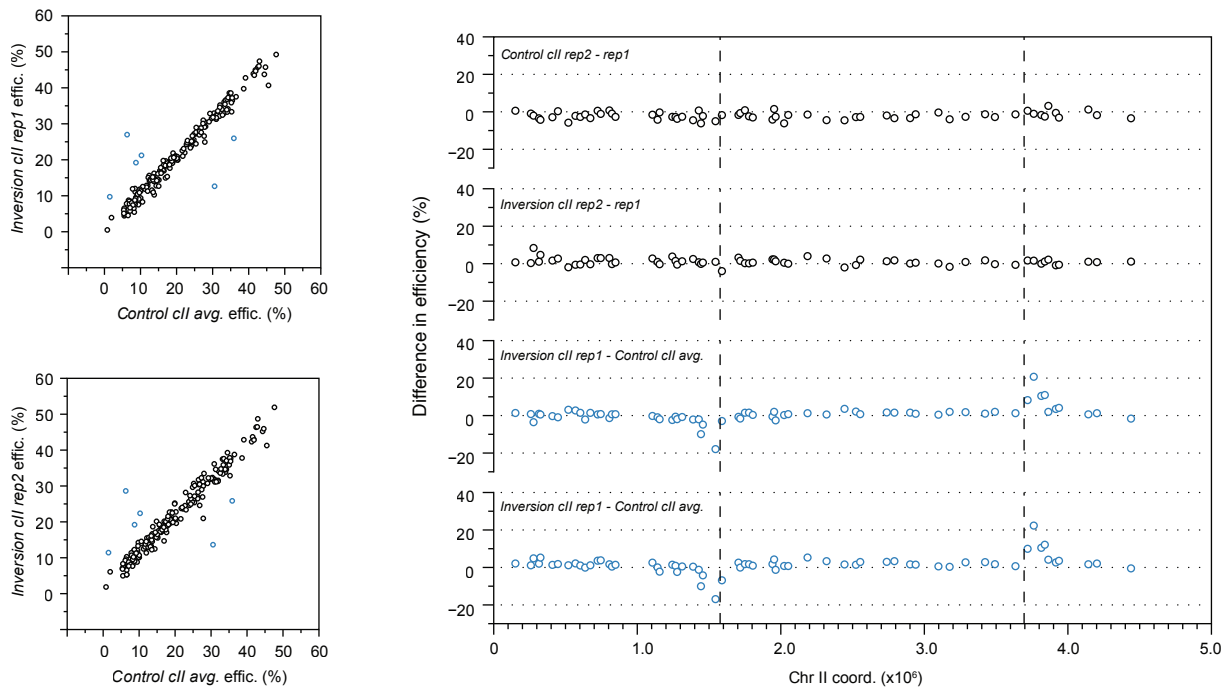
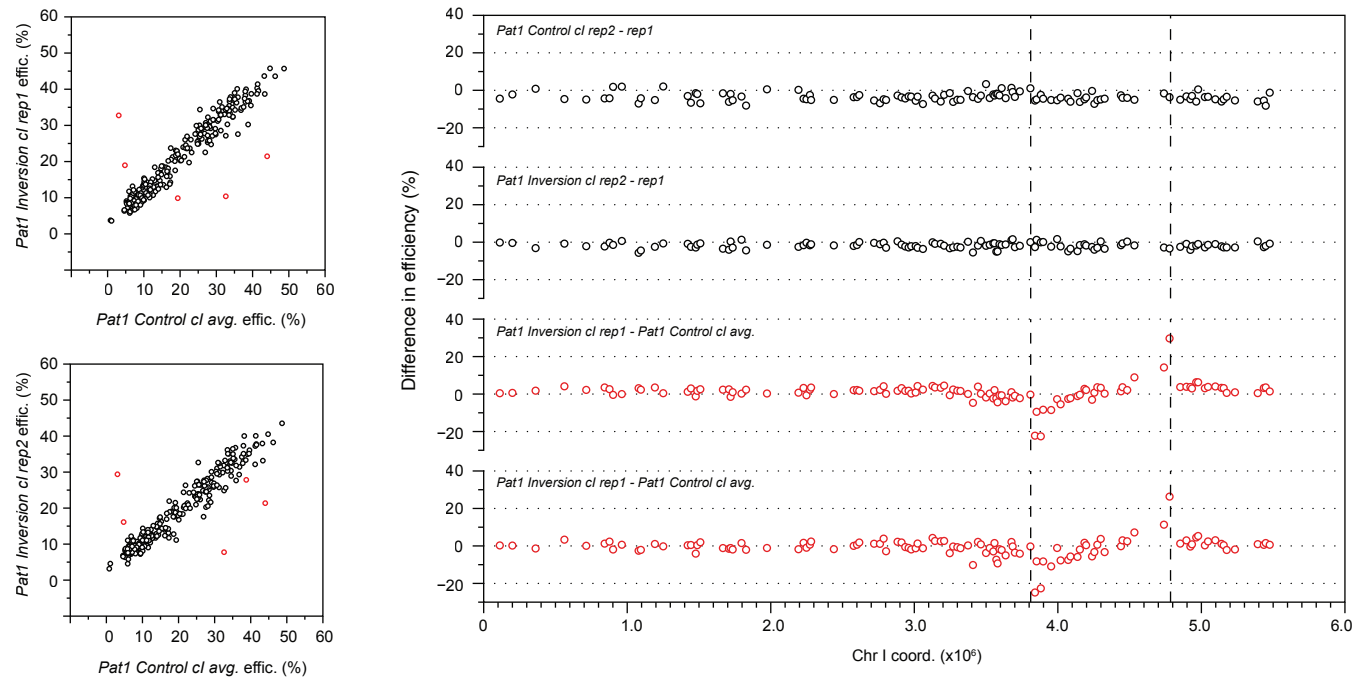
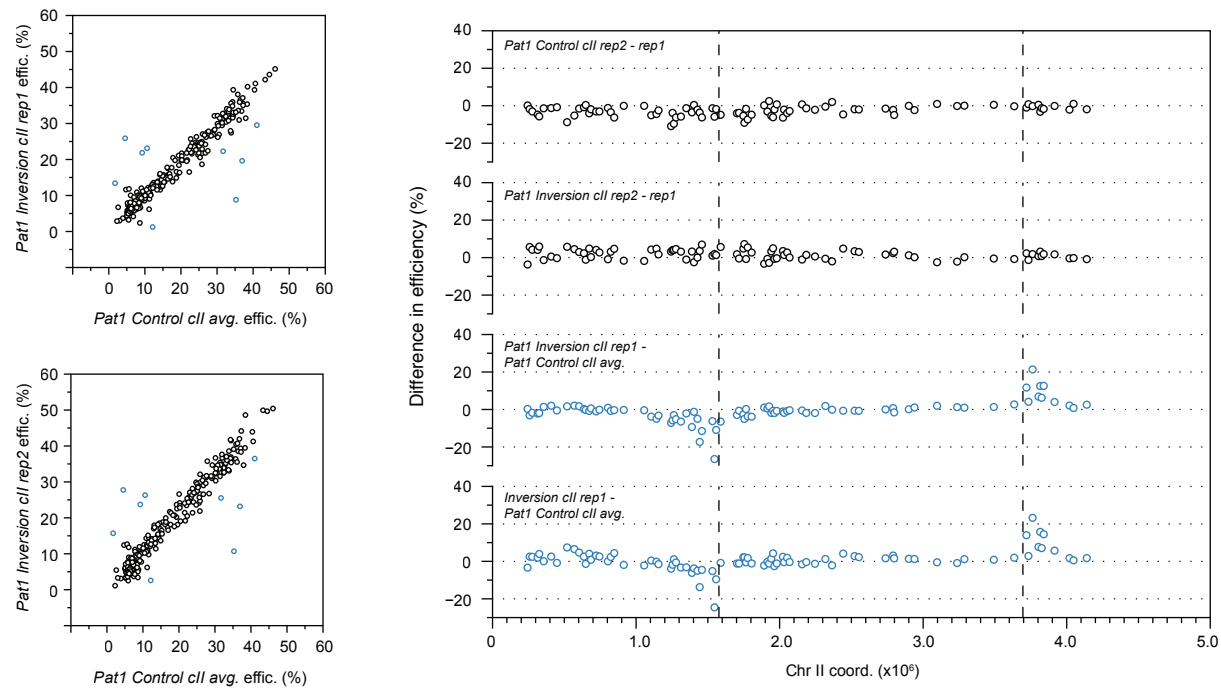


Figure S7

C



D



Supplementary Figure legends

Figure S1. Strategy for generating chromosomal rearrangements. A) Replication profile of a wild-type haploid strain in vegetative growth (Top) or exiting quiescence (Middle). Data are from Fig. 1C and 1D in Wu and Nurse, 2014. x-axis: chromosome coordinates, y-axis: origin efficiencies. Dashed lines represent the locations of the *loxP* insertions (ChrI: 3810766 and 4785016; ChrII: 1577150 and 3694900). Bottom: Schematic of the engineered inversions. Origin efficiencies are indicated by the gradient; darker colors correspond to higher efficiencies. The objective was to alter the chromosome context by disrupting and juxtaposing distinct efficiency domains. The diagram shows how the replication program would appear after the simple inversion of a genomic fragment, in the case that it has no effects on DNA replication. Triangles indicate the positions of centromeres. B) Experimental design for generating chromosomal inversions. Two cassettes were constructed: one contains a *loxP* site flanked by the TEF promoter and the *LEU2* gene from *S. cerevisiae*; a second contains *loxP*, the *ura4* gene from *S. pombe*, and the kanMX ORF. These cassettes were amplified and inserted in distinct loci by homologous recombination. Transient expression of Cre recombinase promotes recombination between the *loxP* sites. Strains with productive rearrangements place the TEF promoter 5' to the kanMX ORF, resulting in resistance to G418. C) Sites of *loxP* insertion in the indicated positions on chromosomes I and II. Images are taken from PomBase [1].

Figure S2. Replication profiles of the first post-quiescence S phase in different genetic backgrounds. A-D) Genome-wide origin efficiencies of *Control cI* (A, black), *Inversion cI* (B, red), *Control cII* (C, orange), and *Inversion cII* (D, blue). To allow direct comparison with the *Control* strains, the data for *Inversion cI* and *Inversion cII* are plotted using the original, pre-rearrangement coordinates. Dashed lines: inversion endpoints. x-axis: chromosome coordinates, y-axis: origin efficiencies.

Figure S3. Origin usage in the *Control* strains. A-B) Top: Schematic of the *loxP* sites present in the indicated chromosomes in the *Control* strains (red boxes: sites in *Control cI*; blue boxes: sites in *Control cII*). Bottom panels: Detailed view of the replication profiles surrounding the inversion endpoints (dashed lines) in *Control cI* (black) and *Control cII* (orange). x-axis: chromosome coordinates, y-axis: origin efficiencies. C) Comparison of origin efficiencies in *Control cII* (y-axis) vs. *Control cI* (x-axis). Each point represents an origin. The Pearson's correlation coefficient (r) indicates a strong correlation between efficiencies in the two backgrounds. ****: p -value < 0.0001.

Figure S4. Replication profiles of pre-meiotic S phase in the different genetic backgrounds. Genome-wide origin efficiencies of *PatI Control cI* (A, black), *PatI Inversion cI* (B, red), *PatI Control cII* (C, orange), and *PatI Inversion cII* (D, blue). To allow direct comparison with the *PatI Control* strains, the data for *PatI Inversion cI* and *cII* are plotted using the original, pre-rearrangement coordinates. Dashed lines: inversion endpoints. x-axis: chromosome coordinates, y-axis: origin efficiencies.

Figure S5. Replication profiles of the *PatI Control* strains. A-B) Top: Schematic of the *loxP* sites present in the indicated chromosomes in the *PatI Control* strains (red boxes: sites in *PatI Control cI*; blue boxes: sites in *PatI Control cII*). Bottom panels: Detailed view of the replication profiles surrounding the inversion endpoints (dashed lines) in *PatI Control cI* (black) and *PatI Control cII* (orange). x-axis: chromosome coordinates, y-axis: origin efficiencies. C) Comparison of origin efficiencies in *PatI Control cII* (y-axis) vs. *PatI Control cI* (x-axis). Each point represents an origin. The Pearson's correlation coefficient (r) indicates a strong correlation between efficiencies in the two backgrounds. *****: p -value < 0.0001.

Figure S6. Comparisons of origin efficiencies from repeat experiments for each strain and condition. A-B) Graphs comparing the two biological repeats of origin mapping assays performed for each strain in a given condition in this study. The Pearson's correlation coefficient (r) for each comparison is displayed. *****: p < 0.00001. The median \pm median absolute deviation is also shown for each experiment. x- and y-axis: origin efficiencies of the indicated repeat (referred to as rep1 and rep2). These results show the high level of reproducibility of our datasets.

Figure S7. Reproducibility of the changes in origin efficiencies induced by chromosomal inversions. A-B) Post-quiesce S phase; C-D) Pre-meiotic S phase. Left panels: Comparisons of the origin efficiencies for individual repeats of a chromosomal inversion (y-axis) with the origin efficiencies determined from the average of the two repeats of the corresponding control strain (x-axis, data as in Figs. S3C and S5C). Each point represents an origin; those in red (A, *Inversion cI*; C, *PatI Inversion cI*) or in blue (B, *Inversion cII*; D, *PatI Inversion cII*) represent origins that were identified to show significant changes in Figures 3 and 6. Right panels: Changes in replication initiation are reproducibly detected near the endpoints of chromosomal inversions. Only chromosomes harboring the inversions are shown. Dashed lines indicate rearrangement endpoints. To allow direct comparison with the *Control* strains, the *Inversion* strains are plotted using the original, pre-translocation coordinates. x-axis: chromosome coordinates; y-axis: origin efficiencies. Black circles: Differences in origin efficiencies between repeats of the indicated strains. Data from Figure S6 are plotted according to their chromosomal positions. Red and blue circles: Differences in origin efficiencies between individual repeats of the indicated inversion strain compared to the origin efficiencies determined from the average of the two repeats of the corresponding control strain. Data from the left panels in A-D are plotted according to their chromosomal positions.

References

1. Harris, M. A.; Rutherford, K. M.; Hayles, J.; Lock, A.; Bähler, J.; Oliver, S. G.; Mata, J.; Wood, V. Fission stories: using PomBase to understand *Schizosaccharomyces pombe* biology. *Genetics* **2022**, 220.