

Review

Epigenetic Perspective of Immunotherapy for Cancers

Sunita Keshari ^{1,†}, Praveen Barrodia ^{2,†} and Anand Kamal Singh ^{2,*} 

¹ Department of Immunology, The University of Texas MD Anderson Cancer Center, Houston, TX 77054, USA

² Department of Genomic Medicine, The University of Texas MD Anderson Cancer Center, Houston, TX 77054, USA

* Correspondence: aksingh1@mdanderson.org

† These authors contributed equally to this work.

Abstract: Immunotherapy has brought new hope for cancer patients in recent times. However, despite the promising success of immunotherapy, there is still a need to address major challenges including heterogeneity in response among patients, the reoccurrence of the disease, and iRAEs (immune-related adverse effects). The first critical step towards solving these issues is understanding the epigenomic events that play a significant role in the regulation of specific biomolecules in the context of the immune population present in the tumor immune microenvironment (TIME) during various treatments and responses. A prominent advantage of this step is that it would enable researchers to harness the reversibility of epigenetic modifications for their druggability. Therefore, we reviewed the crucial studies in which varying epigenomic events were captured with immunology set-ups. Finally, we discuss the therapeutic possibilities of their utilization for the betterment of immunotherapy in terms of diagnosis, progression, and cure for cancer patients.

Keywords: cancer; immunotherapy; epigenetics; immune checkpoint drugs; epigenetic drugs

1. Introduction

Immunotherapy (a type of cancer treatment that relies on empowering the body's own immune cells to fight cancer) has expanded to include: i—immune checkpoint/ligand inhibitors (CTLA-4, PD-1, PD-L1/L2, TIM3, and TIGIT) [1–12]; ii—adoptive T-cell transfer therapy (CAR-T, TCR-T, TIL and NK cell) [13–17]; iii—cancer vaccines (T-vec, BCG and Sipuleucel-T) [18,19]; and iv—immunomodulators (thalidomide, lenalidomide and pomalidomide) [20–23]. Immunotherapy has revolutionized cancer treatment, providing significant clinical benefits to patients with different types of cancers. However, only a small subset of patients benefit from immunotherapy, which highlights limitations of this therapy. Major limitations include the low response rate evidenced by primary/acquired resistance and iRAEs [24–27]. These limitations can be attributed to epigenetic changes acquired by the TIME that play an imperative role in the development of intra/inter tumor heterogeneity by favoring the evolution of transcriptionally distinct clonal populations of cancer cells, which ultimately aid tumor progression and development [28,29].

Epigenetic aberrations are considered hallmarks of cancer development and progression [30]. In the TIME, cancer cells escape immune-mediated cell death by utilizing epigenetic mechanisms to escape host immune recognition and immunogenicity [31,32]. In the tumor microenvironment (TME), in addition to cancer cells, immune cells also undergo various epigenetic modifications that alter their effector cytokine expression, cancer immunosurveillance, immune-checkpoint molecule expression, and tumor-associated antigen presentation with MHC molecules [33,34]. Additionally, epigenetic modulators such as DNA methyltransferase inhibitors (DNMTis) and histone deacetylase inhibitors (HDACis) can re-program the TIME to increase the susceptibility of tumor cells to cytotoxic T-cell-mediated killing, leading to enhanced anti-tumor immune responses [35,36]. Moreover, unlike genetic alterations, epigenetic modifiers can be pharmacologically altered to revert the changes acquired during cancer initiation and progression [37–40].



Citation: Keshari, S.; Barrodia, P.; Singh, A.K. Epigenetic Perspective of Immunotherapy for Cancers. *Cells* **2023**, *12*, 365. <https://doi.org/10.3390/cells12030365>

Academic Editor: Ali R. Jazirehi

Received: 16 November 2022

Revised: 12 January 2023

Accepted: 17 January 2023

Published: 19 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

An improved understanding of epigenetic events related to immunotherapy resistance would be helpful in designing potential combination strategies for immunotherapy. Multiple factors including constitutive PD-L1 expression in cancer cells, a lack of tumor antigens, defective antigen presentation and processing machinery, the exhaustion of infiltrated T cells, and the presence of an immunosuppressive population—such as Tregs, myeloid-derived suppressor cells (MDSCs), and tumor-associated macrophages (TAMs)—could contribute to acquired resistance to immunotherapy [41–43] for TAMs [44].

Therefore, combination therapies involving epigenetic drugs/targets and immunotherapy can serve as improved therapeutic strategies for cancer management by boosting anti-tumor immunity.

2. Epigenetic Modifiers

Epigenetic modification involves a broad range of heritable and reversible changes in gene expression without altering DNA sequences [45,46]. Epigenomic modifications regulate transcription via the modulation of chromatin through the following mechanisms: (1) the post-translational modifications (PTMs) of histone proteins, (2) CpG methylation/demethylation, (3) ATP-dependent nucleosomal repositioning, (4) histone variant exchange, and (5) the action of noncoding RNAs (such as micro RNAs) and (6) chromatin loops [45,47]. Histone tail chemical modifications such as acetylation, methylation, and DNA methylation, which are heritable marks and crucial for the accurate transmission of chromatin states and subsequent gene expression, are the most studied epigenetic modifications [48]. Importantly, these epigenetic modifications are profoundly altered in tumor generation and progression [49]. One important unanswered question is which hierarchical order of events leads to altered gene expression during cancer development. The enzymes involved in epigenetic modifications include DNA methyltransferases (DNMTs), DNA demethylases, histone methyltransferases (HMTs), histone demethylases (HDMs), histone acetyltransferases (HAT), and histone deacetylases (HDACs). DNMT inhibitors (DNMTis) and HDAC inhibitors (HDACis) are the most common epigenetic modulators in clinical use; they, along with the immune modulators, have been identified to regulate the function of immune cells in multiple tumor types (Table 1).

Table 1. Clinical trials of epigenetic agents combined with immune checkpoint inhibitors for cancer therapy.

NCT Identifier	Malignant Conditions	Therapeutics (Single or Combined)	Start Year	Status
NCT05089370	●Malignant Melanoma	●Combination Product: Oral Decitabine/Cedazuridine (DNMT inhibitor) in Combination with Nivolumab (PD-1 inhibitor)	2022	Recruiting
NCT04705818	●Advanced Solid Tumor ●Advanced Colorectal Carcinoma ●Advanced Soft tissue Sarcoma ●Advanced Pancreatic Adenocarcinoma ●Adult Solid Tumor ●Sarcomas ●Melanomas ●Germ Cell Tumors	●Drug: Durvalumab (PD-L1 inhibitor) ●Drug: Tazemetostat (EZH2 inhibitor)	2021	Recruiting
NCT04648826	●Epithelial Malignancies (Excluding Lung and Renal Cell Carcinomas) ●Pulmonary Metastases	●Drug: Bintrafusp alfa (bifunctional fusion protein composed of the extracellular domain of the TGF-receptor II fused to an IgG1 antibody blocking PD-L1) ●Drug: Azacytidine (DNMT1 inhibitor)	2021	Withdrawn

Table 1. Cont.

NCT Identifier	Malignant Conditions	Therapeutics (Single or Combined)	Start Year	Status
NCT04190056	<ul style="list-style-type: none"> •Anatomic Stage IV Breast Cancer AJCC v8 •Prognostic Stage IV Breast Cancer AJCC v8 •Castration-Resistant Prostate Carcinoma •Metastatic Prostate Adenocarcinoma 	<ul style="list-style-type: none"> •Biological: Pembrolizumab (PD-1 inhibitor) •Drug: Tamoxifen (antiestrogen) •Drug: Vorinostat (HDAC inhibitor) 	2021	Recruiting
NCT04471974	<ul style="list-style-type: none"> •Metastatic Prostate Small Cell Carcinoma •Stage IV Prostate Cancer AJCC v8 •Stage IVA Prostate Cancer AJCC v8 •Stage IVB Prostate Cancer AJCC v8 	<ul style="list-style-type: none"> •Drug: ZEN-3694 (BET bromodomain inhibitor) •Drug: Enzalutamide (nonsteroidal antiandrogen (NSAA) medication) •Biological: Pembrolizumab (PD-1 inhibitor) 	2021	Recruiting
NCT04708470	<ul style="list-style-type: none"> •Cancer •Solid Tumor •Metastatic Checkpoint Refractory HPV-Associated Malignancies •Microsatellite Stable Colon Cancer (MSS) 	<ul style="list-style-type: none"> •Drug: Bintrafusp Alfa (bifunctional fusion protein composed of the extracellular domain of the TGF-receptor II fused to an IgG1 antibody blocking PD-L1) •Drug: NHS-IL12 (tumor-targeting immunocytokine) •Drug: Entinostat (HDAC inhibitor) •Drug: Romidepsin (HDAC inhibitor) •Drug: Azacitidine (DNMT inhibitor) •Drug: Nab-Paclitaxel (stops cancer cells from separating into two new cells) 	2021	Recruiting
NCT04257448	<ul style="list-style-type: none"> •Pancreas Cancer •Pancreatic Adenocarcinoma •Pancreatic Ductal Adenocarcinoma 	<ul style="list-style-type: none"> •Drug: Gemcitabine (induces interferon signaling) •Drug: Durvalumab (PD-L1 inhibitor) •Drug: Lenalidomide capsule (potent molecular analog of thalidomide) •Drug: Decitabine (DNMT inhibitor) + TQB2450 injection (PD-1 inhibitor) 	2020	Recruiting
NCT04611711	<ul style="list-style-type: none"> •Patients With Digestive System Tumors Resistant to PD-1 Inhibitors 	<ul style="list-style-type: none"> •Drug: Decitabine (DNMT inhibitor) + TQB2450 injection (PD-1 inhibitor) + Anlotinib (VEGFR inhibitor) 	2020	Not yet recruiting
NCT04553393	<ul style="list-style-type: none"> •Refractory or Relapsed Aggressive r/r BNHL With Huge Tumor Burden 	<ul style="list-style-type: none"> •Drug: Chidamide (HDAC inhibitor) •Drug: Decitabine (DNMT inhibitor) •Biological: Decitabine-Primed Tandem CAR19/20-Engineered T Cells 	2020	Recruiting
NCT04407741	<ul style="list-style-type: none"> •Solid Tumor •Lymphoma 	<ul style="list-style-type: none"> •Drug: SHR2554 (EZH2 inhibitor) •Drug: SHR1701 (PD-1 and TGF-β inhibitor) 	2020	Recruiting
NCT04414969	<ul style="list-style-type: none"> •Immune Checkpoint Inhibitor •Chemotherapy Effect •Epigenetic Disorder •NK/T-Cell Lymphoma of Nasal Cavity 	<ul style="list-style-type: none"> •Drug: Anti-PD-1 antibody + Peg-Asparaginase + Chidamide (HDAC inhibitor) 	2020	Recruiting
NCT04250246	<ul style="list-style-type: none"> •Melanoma •Non-Small Cell Lung Cancer 	<ul style="list-style-type: none"> •Drug: Ipilimumab (CTLA-4 antibody) •Biological: Nivolumab (PD-1 inhibitor) •Drug: Guadecitabine (DNMT inhibitor) 	2020	Not yet recruiting
NCT04277442	<ul style="list-style-type: none"> •Acute Myeloid Leukemia 	<ul style="list-style-type: none"> •Drug: Decitabine (DNMT inhibitor) •Biological: Nivolumab (PD-1 inhibitor) •Drug: Venetoclax (Bcl-2 inhibitor) 	2020	Suspended

Table 1. Cont.

NCT Identifier	Malignant Conditions	Therapeutics (Single or Combined)	Start Year	Status
NCT03812796	<ul style="list-style-type: none"> •Cancer •GI Cancer 	<ul style="list-style-type: none"> •Drug: Domatinostat (HDAC inhibitor) •Drug: Avelumab (PD-1 inhibitor) 	2019	Unknown status
NCT03765229	<ul style="list-style-type: none"> •Melanoma 	<ul style="list-style-type: none"> •Drug: Entinostat (HDAC inhibitor) •Drug: Pembrolizumab (PD-1 inhibitor) 	2019	Recruiting
NCT03854474	<ul style="list-style-type: none"> •Locally Advanced Urothelial Carcinoma •Metastatic Urothelial Carcinoma •Stage III Bladder Cancer AJCC v8 •Stage IIIA Bladder Cancer AJCC v8 •Stage IIIB Bladder Cancer AJCC v8 •Stage IV Bladder Cancer AJCC v8 •Stage IVA Bladder Cancer AJCC v8 •Stage IVB Bladder Cancer AJCC v8 •Non-Small-Cell Lung Carcinoma 	<ul style="list-style-type: none"> •Biological: Pembrolizumab (PD-1 inhibitor) •Drug: Tazemetostat (EZH2 inhibitor) 	2019	Recruiting
NCT03233724	<ul style="list-style-type: none"> •Lung Cancer •Non-Small Cell Lung Cancer •Esophageal Carcinoma •Malignant Pleural Mesotheliomas •Childhood Solid Tumor •Childhood Lymphoma 	<ul style="list-style-type: none"> •Drug: Decitabine (DNMT inhibitor) •Drug: Tetrahydrouridine (inhibitor of cytidine deaminase) •Drug: Pembrolizumab (PD-1 inhibitor) 	2018	Recruiting
NCT03445858	<ul style="list-style-type: none"> •Relapsed Cancer •Refractory Cancer •Adult Solid Tumor •Adult Lymphoma 	<ul style="list-style-type: none"> •Drug: Pembrolizumab (PD-1 inhibitor) •Drug: Decitabine (DNMT inhibitor) 	2018	Active, not recruiting
NCT03161223	<ul style="list-style-type: none"> •T-Cell Lymphoma 	<ul style="list-style-type: none"> •Drug: Durvalumab (PD-L1 inhibitor) •Drug: Pralatrexate (dihydrofolate reductase inhibitor) •Drug: Romidepsin (HDAC inhibitor) •Drug: 5-Azacitidine (Methyltransferase inhibitor) •Drug: Nivolumab (PD-1 inhibitor) 	2018	Recruiting
NCT02664181	<ul style="list-style-type: none"> •Lung Cancer •Non-Small Cell Lung Cancer 	<ul style="list-style-type: none"> •Drug: Oral decitabine (DNMT inhibitor) •Drug: Tetrahydrouridine (inhibitor of cytidine deaminase) 	2017	Active, not recruiting
NCT03206047	<ul style="list-style-type: none"> •Platinum-Resistant Fallopian Tube Carcinoma •Platinum-Resistant Ovarian Carcinoma •Platinum-Resistant Primary Peritoneal Carcinoma •Recurrent Fallopian Tube Carcinoma •Recurrent Ovarian Carcinoma •Recurrent Primary Peritoneal Carcinoma 	<ul style="list-style-type: none"> •Drug: Atezolizumab (PD-L1 inhibitor) •Biological: DEC-205/NYESO-1 Fusion Protein CDX-1401 (vaccine that may help the immune system specifically target and kill cancer cells) •Drug: Guadecitabine (DNMT inhibitor) •Drug: Poly ICLC (induces immunohematopoietic cells) 	2017	Active, not recruiting

Table 1. Cont.

NCT Identifier	Malignant Conditions	Therapeutics (Single or Combined)	Start Year	Status
NCT03250273	<ul style="list-style-type: none"> •Metastatic Cholangiocarcinoma •Cholangiocarcinoma •Pancreatic Cancer •Metastatic Pancreatic Cancer •Unresectable Pancreatic Cancer •Unresectable Cholangiocarcinoma 	<ul style="list-style-type: none"> •Drug: Entinostat (HDAC inhibitor) •Drug: Nivolumab (PD-1 inhibitor) 	2017	Completed
NCT02915523	<ul style="list-style-type: none"> •Epithelial Ovarian Cancer •Peritoneal Cancer •Fallopian Tube Cancer 	<ul style="list-style-type: none"> •Drug: Entinostat (HDAC inhibitor) •Drug: Avelumab (PD-1 inhibitor) 	2017	Unknown status
NCT03024437	<ul style="list-style-type: none"> •Metastatic Cancer •Renal Cancer 	<ul style="list-style-type: none"> •Drug: Atezolizumab (PD-L1 inhibitor) •Drug: Bevacizumab (VEGF inhibitor) •Drug: Entinostat (HDAC inhibitor) 	2017	Suspended
NCT02437136	Non-Small Cell Lung Cancer <ul style="list-style-type: none"> •Melanoma •Mismatch Repair-Proficient Colorectal Cancer 	<ul style="list-style-type: none"> •Drug: Entinostat (HDAC inhibitor) •Drug: Pembrolizumab (PD-1 inhibitor) 	2017	Active, not recruiting
NCT02959437	<ul style="list-style-type: none"> •Solid Tumors •Advanced Malignancies •Metastatic Cancer 	<ul style="list-style-type: none"> •Drug: Azacitidine (DNMT inhibitor) •Drug: Pembrolizumab (PD-1 inhibitor) •Drug: Epacadostat (indoleamine2,3-dioxygenase inhibitor) •Drug: INCB057643 (BET inhibitor) •Drug: INCB059872 (LSD1 inhibitor) 	2017	Terminated
NCT02816021	<ul style="list-style-type: none"> •Melanoma and Other Malignant Neoplasms of Skin •Metastatic Melanoma •Previously Treated Myelodysplastic Syndrome •Recurrent Acute Myeloid Leukemia •Recurrent Acute Myeloid Leukemia with Myelodysplasia Related Changes 	<ul style="list-style-type: none"> •Drug: Azacitidine (DNMT inhibitor) •Drug: Pembrolizumab (PD-1 inhibitor) 	2017	Active, not recruiting
NCT02890329	<ul style="list-style-type: none"> •Recurrent Myelodysplastic Syndrome •Refractory Acute Myeloid Leukemia •Refractory Myelodysplastic Syndrome •Secondary Acute Myeloid Leukemia •Secondary Myelodysplastic Syndrome 	<ul style="list-style-type: none"> •Drug: Decitabine (DNMT inhibitor) •Biological: Ipilimumab (CTLA-4 antibody) 	2017	Active, not recruiting
NCT03019003	<ul style="list-style-type: none"> •Head and Neck Cancer 	<ul style="list-style-type: none"> •Drug: Oral Decitabine (DNMT inhibitor) •Drug: Durvalumab (PD-L1 inhibitor) 	2017	Active, not recruiting
NCT03066648	<ul style="list-style-type: none"> •Leukemia •Myeloid Leukemia •Acute Myeloid Leukemia •Myelodysplastic Syndromes •Preleukemia •Bone Marrow Diseases •Hematologic Diseases •Chronic Myelomonocytic Leukemia 	<ul style="list-style-type: none"> •Drug: Decitabine (DNMT inhibitor) •Drug: PDR001 (PD-1 antibody) •Drug: MBG453 (Tim3 antibody) •Drug: Azacitidine (DNMT inhibitor) 	2017	Active, not recruiting

Table 1. Cont.

NCT Identifier	Malignant Conditions	Therapeutics (Single or Combined)	Start Year	Status
NCT02951156	•Diffuse Large B-cell Lymphoma	<ul style="list-style-type: none"> •Biological: Avelumab (PD-1 inhibitor) •Biological: Utomilumab (binds to CD-137 protein receptor and stimulates/increases the number of immune cells) •Biological: Rituximab (chimeric monoclonal antibody against the protein CD20) •Other: Azacitidine (DNMT inhibitor) •Drug: Bendamustine (chemotherapy medication) •Drug: Gemcitabine (induces interferon signaling) •Drug: Oxaliplatin (inhibits the synthesis of deoxyribonucleic acid (DNA)) 	2016	Terminated
NCT02900560	•Epithelial Ovarian Cancer	<ul style="list-style-type: none"> •Drug: CC-486 (hypomethylation of DNA) •Biological: Pembrolizumab (PD-1 inhibitor) •Drug: Oral CC-486 (hypomethylation of DNA) 	2016	Terminated
NCT02512172	•Colorectal Cancer	<ul style="list-style-type: none"> •Drug: Romidepsin (HDAC inhibitor) •Drug: MK-3475 (PD-1 inhibitor) 	2016	Completed
NCT02395627	Breast Neoplasms	<ul style="list-style-type: none"> •Drug: Tamoxifen (antiestrogens) •Drug: Vorinostat (HDAC inhibitor) •Drug: Pembrolizumab (PD-1 inhibitor) 	2015	Terminated
NCT02608437	•Metastatic Melanoma	<ul style="list-style-type: none"> •Drug: SGI-110 (DNA methylation inhibitor) •Drug: Ipilimumab (CTLA-4 antibody) 	2015	Unknown status
NCT02453620	<ul style="list-style-type: none"> •Breast Adenocarcinoma •Invasive Breast Carcinoma •Metastatic Breast Carcinoma •Metastatic Malignant Solid Neoplasm •Stage III Breast Cancer AJCC v7 •Stage IIIA Breast Cancer AJCC v7 •Stage IIIB Breast Cancer AJCC v7 •Stage IIIC Breast Cancer AJCC v7 •Stage IV Breast Cancer AJCC v6 and v7 •Unresectable Solid Neoplasm 	<ul style="list-style-type: none"> •Drug: Entinostat (HDAC inhibitor) •Biological: Ipilimumab (CTLA-4 antibody) •Biological: Nivolumab (PD-1 inhibitor) 	2015	Active, not recruiting
NCT02546986	•Non-Small-Cell Lung Carcinoma	<ul style="list-style-type: none"> •Drug: CC-486 (hypomethylation of DNA) •Drug: Pembrolizumab (PD-1 inhibitor) 	2015	Active, not recruiting

Table 1. Cont.

NCT Identifier	Malignant Conditions	Therapeutics (Single or Combined)	Start Year	Status
NCT02397720	<ul style="list-style-type: none"> •Acute Bilineal Leukemia •Acute Biphenotypic Leukemia •Acute Myeloid Leukemia Arising from Previous Myelodysplastic Syndrome •Chronic Myelomonocytic Leukemia •Myelodysplastic Syndrome •Recurrent Acute Myeloid Leukemia •Refractory Acute Myeloid Leukemia •Secondary Acute Myeloid Leukemia •Therapy-Related Acute Myeloid Leukemia 	<ul style="list-style-type: none"> •Drug: Azacitidine (DNMT inhibitor) •Biological: Ipilimumab (CTLA-4 antibody) •Biological: Nivolumab (PD-1 inhibitor) 	2015	Recruiting
NCT02608268	<ul style="list-style-type: none"> •Advanced Malignancies 	<ul style="list-style-type: none"> •Drug: MBG453 (Tim3 antibody) •Drug: PDR001 (PD-1 antibody) •Drug: Decitabine (DNMT inhibitor) 	2015	Active, not recruiting
NCT01834248	<ul style="list-style-type: none"> •Acute Myeloid Leukemia •Alkylating Agent-Related Acute Myeloid Leukemia •Chronic Myelomonocytic Leukemia •Myelodysplastic Syndrome •Refractory Anemia with Excess Blasts 	<ul style="list-style-type: none"> •Biological: DEC-205/NYESO-1 Fusion Protein CDX-1401 (vaccine that may help the immune system specifically target and kill cancer cells) •Drug: Decitabine (DNMT inhibitor) •Drug: Poly ICLC (induces immunohematopoietic cells) 	2013	Completed
NCT01928576	<ul style="list-style-type: none"> •Non-Small Cell Lung Cancer •Epigenetic Therapy 	<ul style="list-style-type: none"> •Drug: Azacitidine (DNMT inhibitor) •Drug: Entinostat (HDAC inhibitor) •Drug: Nivolumab (PD-1 inhibitor) 	2013	Recruiting

3. Epigenetic Modifiers in T Cells

The functional differentiation of T cells, like short-lived effectors, long-term memory T cells, Treg, and other T-cell populations, is majorly influenced by epigenetic modifications. An increasing number of investigations support the crucial role of HATs, HDACs and HMTs in regulating the fate and function of T cells. The inhibition of HDAC1 and HDAC2 promote the differentiation of CD4⁺ T cells into cytotoxic CD4⁺ T cells [50,51]. HDAC3 is critical for the maturation of both CD4⁺ and CD8⁺ T cells and the production of TNF upon TCR/CD28 stimulation [52]. Enrichment in the central memory and stem cell memory phenotypes of T cells is regulated by H3K4me3 modification at specific gene promoters such as TCF7, LEF1, and KLF2. Interestingly, the upregulation of H3K4me3 and the downregulation of H3K27me3 at the Gnt1 locus were found to enhance the trafficking of memory T cells to tumor sites in an interleukin (IL)-15-dependent manner [53].

Scheer et al. reported that lysine methyltransferase Dot1l-dependent H3K79me2 is crucial for CD4⁺ T helper (Th) cell differentiation, as the loss of it was found to lead to the increased expression of Th-1-specific genes and the overproduction of IFN- γ at the expense of Th-2 cell development, advocating a central role for Dot1l in Th-2 cell lineage commitment and stability [54]. Another study investigated the role of menin, a major component of the trithorax group (TrxG) using Cd4-cre-driven conditional knockout (KO) mice; a deficiency in menin was shown to lead to the downregulation of Gata3 expression due to reduced levels of H3K9ac and H3K4me3 at the upstream regions of the Gata3 proximal promoter [55]. Interestingly, the suppression of histone H3K27 demethylases KDM6A (UTX) in mature Th-17 cells was found to reduce mitochondrial biogenesis, causing metabolic reprogramming and reducing the expression of key metabolic TFs, such as

PPRC1, which ultimately showed anti-inflammatory effects [56]. The results of these studies reinforce the role of epigenomic events in T-cell biology.

3.1. Epigenetic Modifiers in Immune Checkpoint Therapy

A critical balance between immune co-inhibitory and co-stimulatory signals in the TIME is maintained to restrict tumor development and progression (Figure 1) [57,58]. The epigenetically regulated aberrant expression of immune checkpoints (ICs), including PD-1, CTLA-4, TIM-3 (T-cell immunoglobulin and mucin-domain containing-3), LAG-3 (lymphocyte-activation gene 3), TIGIT (T-cell immunoreceptor with Ig and ITIM domains), VISTA (V-domain Ig suppressor of T-cell activation), CD276 (B7-H3), B7-H4 (VTCN1/B7x/B7S1/B7 homolog 40), IDO-1 (indoleamine 2,3-dioxygenase 1), CD161, CD38, CD93, and CD47 may result in the induction of an immune-suppressive environment, which helps tumor cells to evade immune destruction [12,59,60]. Targeting altered epigenetic modifications can significantly contribute to the reversal of the transcriptomic regulation of ICs and their ligands, which could help to re-establish potent host immunosurveillance mechanisms [61].

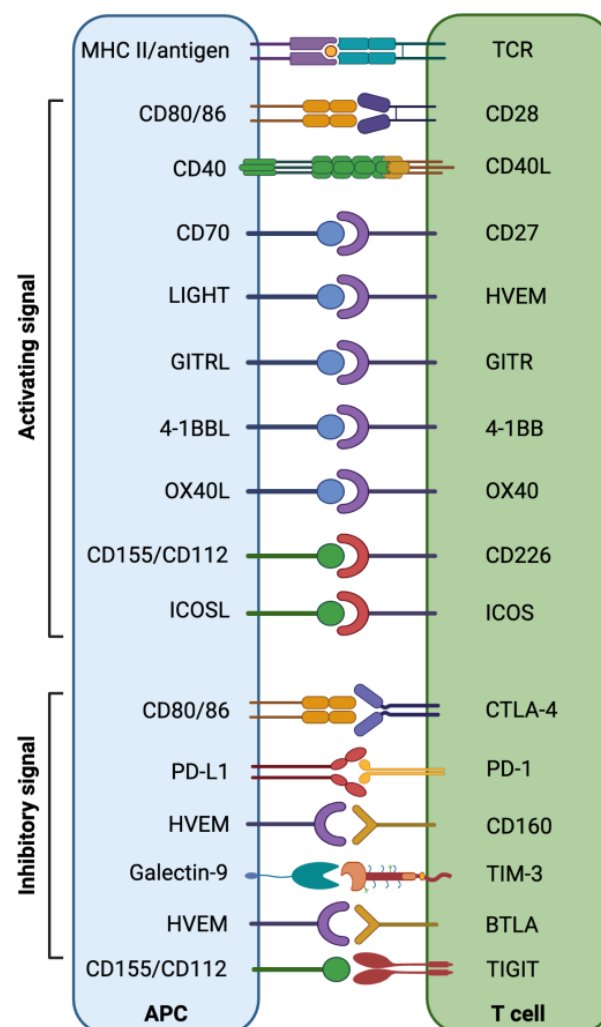


Figure 1. Interaction of co-stimulatory/inhibitory molecules between T cells and APCs/tumor cells provides an overview of the immune checkpoint/stimulatory molecules involved in the anti-tumor immune response.

DNMTis and HDACis have been shown to cause the upregulation of immune-signaling components and antigen presentation through the expression of ERVs (endogenous retro-

viral sequences), thereby improving tumor cell recognition [62]. Decitabine (DNMTi) upregulates the cancer testis antigen member *MAGE-1* via hypomethylation, thus increasing chances of its presentation through MHC molecules to effector immune cells [63]. Panobinostat (pan HDACi) has been found to significantly increase CD38 expression in multiple myeloma, so it has been utilized in the development of an effective combinatorial treatment with daratumumab [64]. Panobinostat also affects the PD-L1/PD1 axis via the upregulation of PD-L1 in melanoma cells, which can be then targeted with anti PD-L1 antibodies [65].

H3K9 lysine methyltransferase, SETDB1, has a critical role in the carcinogenesis of multiple tissue types through the transcriptional silencing of multiple genes at specific loci. The amplification and increased expression of SETDB1 in advanced clear renal cell carcinoma has been found to be associated with a poor response to anti-PD1 therapy [66]. Further exploring this information, the Bernstein lab identified that the reversal of the epigenetic silencing of *SETDB1* activates tumor immunogenicity through the hypomethylation of H3K9 in the transposable elements that reside in the MHC peptidome [67,68]. The results of these studies indicate the high potential of SETDB1 inhibitors such as mithramycin in combination with immune checkpoint blockade therapy (ICT) [69].

The H3K27Ac reader bromodomain and extra-terminal motif (BET) protein is overexpressed in various cancers and involved in the regulation of the (PD-1/PD-L1) immune checkpoint axis [36]. Accordingly, targeting BET with JQ1 inhibitors in combination with anti-PD1 therapy has been proven to be effective in ovarian and triple-negative breast cancer [36,70]. In another study, Adeegbe et al. showed that JQ1 treatment significantly lowered PD-L1 expression in tumor cells, which led to an increased tumor infiltration of cytotoxic T cells in a non-small cell lung cancer NSCLC xenograft, and a combination treatment of JQ1 with anti-PD-1 reduced tumor burden and resulted in an improved survival rate [71].

The promoter hypomethylation of LAG3 has been a major epigenetic regulator of mRNA expression in clear cell renal cell carcinoma (KIRC), which has a proven association with increased immune cell infiltration and an interferon- γ signature [72]. Beyond cancer, patients' aberrant histone methylation in chronic osteomyelitis is related to the higher expression of LAG3 in the T cells of peripheral blood [73]. Another negative stimulatory molecule, Tim-3, has been shown to be epigenetically regulated, so its increased expression inhibits the expansion of Th1 and Th17 responses via its binding to galectin-9, ultimately leading to immune exhaustion in the tumor microenvironment [74–76]. EZH2-H3K27me3/DNMT3A-DNA methylation regulates the expression of Tim-3 and galectin-9 in HPV18-associated cervical cancer [77]. Tim-3 and galectin-9 are overexpressed in cervical cancer cases, which is mediated through the hypomethylation of *HAVCR2* and *LGALS9* because of the lesser expression and recruitment of DNMT3A to their promoter regions. SUV39H1, a H3K9me3-specific histone methyltransferase, contributes to Tim-3 and galectin-9 regulation by upregulating the H3K9me3 level at the DNMT3A promoter region, hence downregulating its expression. Therefore, SUV39H1 can be utilized as a potential therapeutic target that can downregulate the immune checkpoint inhibitors Tim-3 and galectin-9 [78].

Another immune checkpoint, TIGIT, was found to be upregulated during T-cell and NK-cell exhaustion [9]. Moreover, TIGIT was reported to be regulated by promoter demethylation in melanoma, thus making it sensitive to anti PD-1 therapy [79].

3.2. Epigenetic Modifiers in Antigen Processing and Presentation

In a proper functioning immune system, T cells recognize tumor antigens based on the binding of a T-cell receptor (TCR) and a matching antigen packaged into major histocompatibility complex (MHC) proteins on APCs. Tumor cells escape immune recognition through multiple mechanisms such as alterations in antigen presentation and processing machinery (APM) or alterations in MHC class I molecules, which further impair their identification by CTLs (Figure 2).

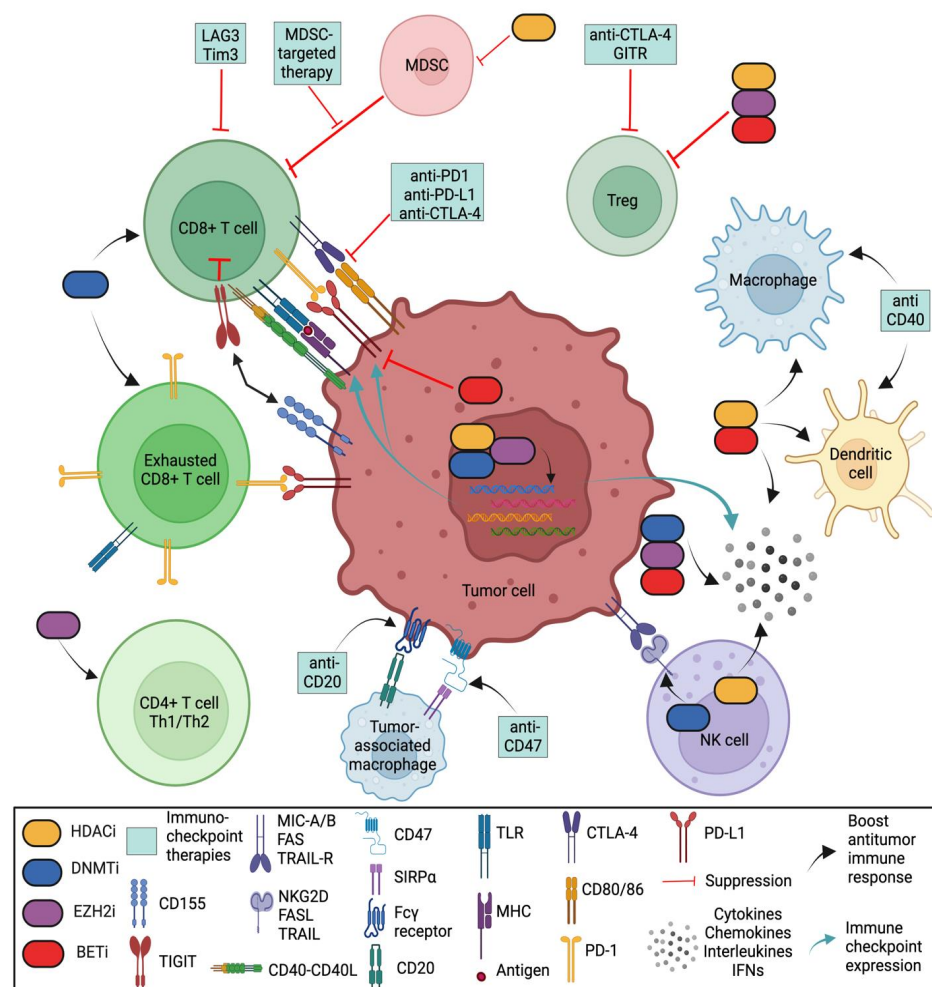


Figure 2. Role of various epigenetic modifiers in the tumor immune microenvironment. DNA methyltransferase inhibitors (DNMTis), histone deacetylase inhibitors (HDACis), an inhibitor of histone methylation on histone H3 at lysine 27 (EZH2i), and inhibitor of bromodomain and extra-terminal motif (BETi) shape the tumor-immune microenvironment by (i) increasing the number of CD8 and CD4 T cells; (ii) activating antigen processing and presentation machinery; (iii) decreasing the abundance of MDSCs and tumor-associated macrophages (TAMs); (iv) downregulating the immune checkpoint inhibitors Tim-3, Lag-3 and TIGIT; (v) upregulating immune checkpoint PD-L1 (by DNMTis, HDACis and EZH2i) and downregulating PD-L1 (by BETi); (vi) enhancing NK-mediated lysis (by HDACis) or decreasing NK cytotoxicity (by EZH2i); and (vii) upregulating inflammatory genes and pathways that control the secretion of interferons (IFNs), cytokines, and chemokines from tumor cells. (Regulatory T cells (Tregs) are a specialized subpopulation of T cells that act to suppress the immune response, thereby maintaining homeostasis and self-tolerance. It has been shown that Tregs are able to inhibit T-cell proliferation and cytokine production, as well as play a critical role in preventing autoimmunity. Tumor-associated macrophages (TAMs) are the key cells that create an immunosuppressive tumor microenvironment (TME) by producing cytokines, chemokines, and growth factors and by triggering the inhibitory immune checkpoint proteins release in T cells. Natural killer (NK) cells are effector lymphocytes of the innate immune system that control several types of tumors and microbial infections by limiting their spread and subsequent tissue damage. Cancer-associated fibroblasts (CAFs) are one of the most abundant and critical components of the tumor mesenchyme; they not only provide physical support for tumor cells but also play a key role in promoting and retarding tumorigenesis in a context-dependent manner. Recent studies have revealed their roles in immune evasion and poor responses to cancer immunotherapy).

An efficient cancer immunotherapy depends on the recognition of antigens loaded onto the MHC molecules of antigen-presenting cells by T cells in the TIME. The epigenomic regulatory factors that can influence the T-cell recognition of tumor antigens include: (1) the aberrant expression of genes involved in the processing or presentation of tumor antigens and (2) the aberrant expression of antigens. There is a subclass of cancer testis antigens (CTAs), including MAGE (melanoma-associated antigen), PRAME (preferentially expressed antigen of melanoma) and NY-ESO-1 (New York esophageal squamous cell carcinoma-1), which are controlled by DNA methylation and remain silenced in mature somatic cells but are demethylated and overexpressed in various cancers [80,81]. Guadecitabine (SGI-110) and decitabine, which are hypomethylating drugs, have been shown to upregulate/overexpress CTAs such as NY-ESO-1 in epithelial ovarian cancer cells and xenografts when used in combination with NY-ESO-1 vaccine and doxorubicin chemotherapy; T-cell responses to NY-ESO-1 have been observed in most studied patients [82,83].

Studies have evidenced that DNMTis and/or HDACis could alter the expression of MHC class I molecules in cancer cells such as neuroblastoma, cervical, and prostate cancer [84]. Furthermore, the expression of different components of the APM pathway such as TAP-1, TAP-2, LMP2, LMP7 and tapasin can be manipulated by both DNMTis and HDACis in different tumor types [85–87]. DNMTis and HDACis can regulate the expression of the costimulatory molecules ICAM-1, CD40, CD80, and CD86 [86,88,89].

Histone methyltransferase SETDB1, which maintains heterochromatin (H3K9me3), plays crucial roles in the carcinogenesis of multiple tissue types through the transcriptional silencing of multiple genes [90]. Accordingly, the inhibition of SETDB1 was found to enhance specific cytotoxic T-cell responses against tumors via the activation of immunostimulatory genes, the encoding of retroviral antigens, and the generation of neoantigen MHC-I peptides, thus suggesting that SETDB1 has high potential to synergize with ICT [91]. The HDAC-1/3 inhibitor entinostat, upon combination with a PD-1 axis blockade, was found to lead to the complete remission of tumors, the expansion of neoantigen-specific T cells, and the induction of long-term immunologic memory in immune-competent bladder cancer mouse models [92].

3.3. Epigenetic Modifiers in Tumor-Infiltrating Immunosuppressive Cells

Tumor-infiltrating immunosuppressive cells such as myeloid-derived suppressor cells (MDSCs), tumor-associated macrophages (TAMs), regulatory T cells (Tregs), and cancer-associated fibroblasts (CAFs) inhibit T cells' effector functionality and anti-tumor responses, which lead to the immune escape of tumors. The presence of an immunosuppressive cell population in the TIME could be a major contributory factor in ineffective ICTs [93]. HDACis have antitumor effects in that they reduce the number of MDSCs through various mechanisms of action such as CG-745, a class I–IIb HDACi that induces the infiltration of lymphocytes by increased antigen presentation and that decreases the amount of MDSCs by decreasing the polarization of M2 macrophages in tumors [35]. Valproic acid (VPA), a class-I HDACi, attenuates the immunosuppressive function of MDSCs by downregulating the expression of retinoblastoma 1 (Rb1), toll-like receptor 4 (TLR4), programmed cell death 1 ligand (PD-L1), and interleukin-4 receptor-alpha (IL-4Ra)/arginase [94]. Moreover, the combinatorial treatment of VPA and anti-PD-1 antibodies was found to repress the growth of B16F10 and EL4 tumor models by impairing tumor-infiltrating M2-MDSC accumulation in the tumor microenvironment compared with their individual therapies [95]. Thus, treatment with epigenetic modifiers inhibits MDSC accumulation, thereby augmenting immune checkpoint inhibitors for successful cancer treatment. Vorinostat (suberoylanilide hydroxamic acid, SAHA), a class I–II–IV HDACi, was shown to have anti-tumor potential for a 4T1 mammary mice model in which it decreased MDSC accumulation in the spleen, blood, and tumor while promoting the activation and function of CD8+ T cells [96].

Tregs play significant roles in inducing variety of immune responses, as determined by the expression of Foxp3, a transcription factor in natural Tregs (nTregs) in the thymus [97,98]. Extrinsic molecular signals including IL-2 and TCR, along with a network

of transcription factors, are critical for regulating the expression of Foxp3 through epigenomic modulation, which ultimately determines a Treg's phenotypic plasticity [99,100]. Epigenetic modifiers such as DNMT1 and DNMT3b are differentially bound to Foxp3 promoter and enhancer sites in nTregs compared with extrinsically induced Tregs. Importantly, DNMTs demethylate and activate the Foxp3 promoter and enhancer elements to induce Foxp3 expression and subsequently enable the induction of Foxp3-dependent, Treg-restricted sets of genes [101]. Demethylation in synergy with TGF- β transforms naive T cells into Tregs with high Foxp3 expression and potent, stable suppressive function [102].

Foxp3 expression in Treg cells was found to be significantly upregulated upon treatment with trichostatin-A (TSA), a HDACi [103]. Moreover, the CTLA4, PD-1, GITR and IL-10 genes are reportedly upregulated by TSA [104]. Ohkura et al. reported that Treg maturation, Treg-specific gene expression, and Treg-specific immunosuppressive activity involve epigenetic regulation through genome-wide CpG DNA hypomethylation pattern [105]. In other study, Wang et al. showed that the inhibition of EZH2, a histone-lysine N-methyltransferase enzyme, resulted in Treg-mediated pro-inflammatory activities in the TME, supporting the idea of the generation of an effector T-cell-mediated anti-tumor immune response [106].

3.4. Epigenetic Modifiers in Inflammatory Cytokines and Chemokines

The pro-/anti-tumorigenic effect of inflammatory cytokines and chemokines, such as TNF- α , IL-1, IL-6, and IFN- γ , has been well-established in tumor malignancies; however there is little evidence that their aberrant expression is regulated through various epigenetic mechanisms in cancer development [107,108]. IFN- γ is a pleiotropic cytokine associated with the induction of reactions in T lymphocytes, which contributes to the enhancement of an immune response against malignant cells. The downregulation of IFN- γ mediated by hypermethylation has been observed in lung and cervical cancer [109,110]. Interestingly, IFN- γ is suppressed in the presence of E6 (a human papillomavirus (HPV) protein), suggesting the involvement of E6 in IFN- γ de novo methylation followed by transcriptional silencing [111]. One of the earliest studies in humans showed that epigenetic modifications occurring in the IFN- γ , IL-4, and IL-13 genes regulate the differentiation of CD4 T cells into Th1 and Th2 cell lineages. The IFN- γ promoter is demethylated during differentiation into Th1 cells [112], and the demethylation of several specific CpG dinucleotides occurs in the IL-4 and IL-13 genes during Th2 differentiation [113]. Most importantly, epigenetic histone marks are major determinants of Th1/Th2 cell fate.

In addition to their role in development and inflammatory responses, chemokines and their receptors also play critical roles in neoplastic transformations, cancer progression, and angiogenesis. CXCL14 (also known as BRAK), a member of the chemokine family, acts as a chemoattractant and stimulates the trafficking of natural killer cells to sites of inflammation or malignancy [114]. The aberrant methylation of CpG islands in the promoter region and the first exon of the CXCL14 gene is associated with its downregulation in gastric cancer [115]. Moreover, CXCL14 was found to be transcriptionally inactivated by promoter CpG hypermethylation in human prostate cancer [116]. CXCL12 and its receptor CXCR4 belong to same family of CXCL14 and are also associated with tumorigenesis. Interestingly, the demethylation of CXCR4 and the hypermethylation of CXCL12 and ESR1 are predictive marker of tumor stage, size, metastasis, and poor overall survival in breast cancer [117].

Multiple proinflammatory cytokines including interleukins are often stated to be epigenetically regulated in various forms of cancer, especially lung cancer. The expression of the IL-1B, IL-6, and IL-8 genes are regulated through promoter DNA methylation which have been reported to play crucial roles in lung cancer [118]. Interleukin-23, a member of the IL-6 superfamily, is stated to be epigenetically regulated in non-small-cell lung cancer (NSCLC) via both histone acetylation and DNA methylation [119]. The epigenetic silencing of IL12RB2, a subunit of the IL-12 receptor, is a recurrent event in human lung cancers [120]. Furthermore, IL12RB2 methylation has been found to be frequent in patients suffering

from both chronic obstructive pulmonary diseases (COPD) and non-small-cell lung cancer (NSCLC) [121].

3.5. Epigenetic Modifiers in Natural Killer Cells

NK cells are key mediators of the innate immune response, and they exert cytotoxic effects after the recognition of cancer cells and virus-infected cells [122]. Upon the recognition of tumor cells, NK-cell activation occurs through the interaction of NKG2D receptors on the surface of NK cells with ULBP ligands and the MHC class I chain-associated proteins MICA and MICB on the surface of tumor cells [123]. One study reported that VPA (a HDACi) upregulates NKG2D, the immunoreceptor that binds with MICA and MICB, thus leading to the enhancement of the NK-mediated lysis of cancer cells in AML [122,124]. EZH2, an HMT, was found to inhibit the differentiation and function of NK cells by downregulating NKG2D receptor expression. Moreover, EZH2-mediated H3K27me3 induces the silencing of the IL-15R, CD122, and NKG2D receptor proteins, hence suppressing NK-cell expansion and decreasing the cytotoxic targeting of tumor cells.

3.6. Epigenetic Modifiers in CAR-T Therapy

Cell-based therapies, such as chimeric antigen receptor (CAR) T-cell therapies, have led to enormous successes against several hematological malignancies. However, their success has been limited due to tumor antigen heterogeneity, tumor infiltration, and persistence. Nevertheless, the manipulation and modification of epigenetic and genetic cascades have been observed to trigger specific T-cell-signaling pathways, which can help to promote the expansion and persistence of CAR-T cells. In support of this idea, a hypomorphic mutation in the epigenetic modifier TET2 (a chromatin modifier that encodes the methyl cytosine dioxygenase enzyme that facilitates DNA demethylation to activate gene expression) has been shown to support the central memory phenotype in anti-CD19 CAR-T cells in chronic lymphocytic leukemia [125]. The inhibition of TET2 through S-2-hydroxyglutarate (S-2HG) was found to result in the formation of CD8+ central memory CAR T cells, which helps to overcome the issue of the persistence of CAR-T cells in patients with B-cell malignancies [126]. The pretreatment of lymphoma cells with decitabine, a DNMTi, was shown to lead to the increased expression of the surface antigen CD19 on lymphoma cells, making them more susceptible to CD19 CAR T cells; this was observed in two lymphoma patients who were treated with decitabine before CAR T-cell therapy and achieved complete remission [127]. The triple knockdown of the T-cell exhaustion signature genes PD-1, Tim-3, and Lag-3 dramatically increases the chromatin accessibility of the CD56 gene, leading to the increased expression of CD56 in CAR-T cells and making them more effective at infiltrating ovarian cancer [128]. Antigen heterogeneity and antigen loss are key obstacles for developing effective CAR T-cell therapy. EZH2 is associated with a low antigen presentation and poor immunogenicity, and targeting EZH2 with a selective inhibitor and CAR T-cell therapy was found to lead to the significant enhancement of the antitumor activity of CAR T cells [129].

4. Transcription Factor Circuitry in ICT Resistance

ICT resistance can be attributed to T-cell exhaustion and a lack of central and effector memory T-cell formation. Numerous transcription factors are associated with these T-cell fates, and a few of them have been reported to be epigenetically regulated including NR4A1, NR4A2, NR4A3, TBET, EOMES, TCF1 (TCF7), LEF1, BATE, NFAT and EGR2 [130,131]. The high expression of NR4A1, NR4A2, and NR4A3 has been related to the poor prognosis of many cancers, and there are reports that suggest that their binding to the target LEF1 promoters can be regulated through DNA methylation and histone acetylation levels [132]. T-bet has the ability to recruit a H3K27-demethylase-Jmjd3 and a H3K4-methyltransferase-Set7/9 complex to its target genes that empower T-bet to effectively modulate the epigenomic state of its target genes and T-cell fate [133]. EOMES has been reported to be a member of chromatin-modulating complexes containing BRG1, which has been observed in RUNX3 enhancers in

T-cell innate memory formations [134]. BATF epigenetically regulates activation-associated gene expression in tumor-infiltrated Treg cells [135]. Networks of TFs also determine the differentiation and phenotypes of the T-cell-like T-bet, which displaces Sin3A-histone deacetylase (HDAC1 and HDAC2) complexes to facilitate the differentiation of Th1 cells [136]. In response to IL-12 signals, the activation of STAT4 (required for the development of Th1 cells), facilitates chromatin remodeling at the enhancer regions of Th1 genes. EZH2 facilitates the correct expression of Tbx21 and GATA-3 for differentiating Th1 and Th2 cells through H3K27 trimethylation (H3K27me3) [137]. BATF regulates Th1 gene expression via the acetylation of T-bet and IFN- γ , considered an important checkpoint in T-cell differentiation [138].

5. Conclusions

Immunotherapy is a major breakthrough in cancer treatment, though it still has challenges. If we can better understand immunotherapy at the cellular and molecular levels, then we can deal with its underlying issues. This review is a step towards understanding the epigenetic mechanism involved in the significant components of immunotherapy. This understanding can enable us to develop new ideas and hypotheses in the study of novel combinatorial treatment/biomarkers for correct treatment plans, including epigenetic features that have the intrinsic advantage of “reversibility”. This will increase the chances of success in future cancer immunotherapies.

Author Contributions: Conceptualization- A.K.S.; investigation, S.K., P.B., A.K.S.; writing- S.K., P.B., A.K.S.; review and editing- S.K., P.B., A.K.S.; graphics and visualization- P.B.; All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Buchbinder, E.I.; Desai, A. CTLA-4 and PD-1 Pathways: Similarities, Differences, and Implications of Their Inhibition. *Am. J. Clin. Oncol.* **2016**, *39*, 98–106. [[CrossRef](#)] [[PubMed](#)]
2. Freeman, G.J.; Long, A.J.; Iwai, Y.; Bourque, K.; Chernova, T.; Nishimura, H.; Fitz, L.J.; Malenkovich, N.; Okazaki, T.; Byrne, M.C.; et al. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *J. Exp. Med.* **2000**, *192*, 1027–1034. [[CrossRef](#)] [[PubMed](#)]
3. Seidel, J.A.; Otsuka, A.; Kabashima, K. Anti-PD-1 and Anti-CTLA-4 Therapies in Cancer: Mechanisms of Action, Efficacy, and Limitations. *Front. Oncol.* **2018**, *8*, 86. [[CrossRef](#)] [[PubMed](#)]
4. Anderson, A.C. Tim-3, a negative regulator of anti-tumor immunity. *Curr. Opin. Immunol.* **2012**, *24*, 213–216. [[CrossRef](#)] [[PubMed](#)]
5. Anderson, A.C. Tim-3: An emerging target in the cancer immunotherapy landscape. *Cancer Immunol. Res.* **2014**, *2*, 393–398. [[CrossRef](#)]
6. He, Y.; Cao, J.; Zhao, C.; Li, X.; Zhou, C.; Hirsch, F.R. TIM-3, a promising target for cancer immunotherapy. *Onco Targets Ther.* **2018**, *11*, 7005–7009. [[CrossRef](#)]
7. Huard, B.; Prigent, P.; Tournier, M.; Bruniquel, D.; Triebel, F. CD4/major histocompatibility complex class II interaction analyzed with CD4- and lymphocyte activation gene-3 (LAG-3)-Ig fusion proteins. *Eur. J. Immunol.* **1995**, *25*, 2718–2721. [[CrossRef](#)]
8. Triebel, F.; Jitsukawa, S.; Baixeras, E.; Roman-Roman, S.; Genevee, C.; Viegas-Pequignot, E.; Hercend, T. LAG-3, a novel lymphocyte activation gene closely related to CD4. *J. Exp. Med.* **1990**, *171*, 1393–1405. [[CrossRef](#)]
9. Yu, X.; Harden, K.; Gonzalez, L.C.; Francesco, M.; Chiang, E.; Irving, B.; Tom, I.; Ivelja, S.; Refino, C.J.; Clark, H.; et al. The surface protein TIGIT suppresses T cell activation by promoting the generation of mature immunoregulatory dendritic cells. *Nat. Immunol.* **2009**, *10*, 48–57. [[CrossRef](#)]
10. Johnston, R.J.; Comps-Agrar, L.; Hackney, J.; Yu, X.; Huseni, M.; Yang, Y.; Park, S.; Javinal, V.; Chiu, H.; Irving, B.; et al. The immunoreceptor TIGIT regulates antitumor and antiviral CD8(+) T cell effector function. *Cancer Cell* **2014**, *26*, 923–937. [[CrossRef](#)]
11. Abbas, H.A.; Hao, D.; Tomczak, K.; Barrodia, P.; Im, J.S.; Reville, P.K.; Alaniz, Z.; Wang, W.; Wang, R.; Wang, F.; et al. Single cell T cell landscape and T cell receptor repertoire profiling of AML in context of PD-1 blockade therapy. *Nat. Commun.* **2021**, *12*, 6071. [[CrossRef](#)]

12. Wang, Y.; Zhang, H.; Liu, C.; Wang, Z.; Wu, W.; Zhang, N.; Zhang, L.; Hu, J.; Luo, P.; Zhang, J.; et al. Immune checkpoint modulators in cancer immunotherapy: Recent advances and emerging concepts. *J. Hematol Oncol* **2022**, *15*, 111. [[CrossRef](#)] [[PubMed](#)]
13. Rohaan, M.W.; Wilgenhof, S.; Haanen, J. Adoptive cellular therapies: The current landscape. *Virchows Arch.* **2019**, *474*, 449–461. [[CrossRef](#)]
14. Wang, Z.; Cao, Y.J. Adoptive Cell Therapy Targeting Neoantigens: A Frontier for Cancer Research. *Front. Immunol.* **2020**, *11*, 176. [[CrossRef](#)]
15. Rosenberg, S.A.; Restifo, N.P.; Yang, J.C.; Morgan, R.A.; Dudley, M.E. Adoptive cell transfer: A clinical path to effective cancer immunotherapy. *Nat. Rev. Cancer* **2008**, *8*, 299–308. [[CrossRef](#)]
16. Miller, J.S.; Soignier, Y.; Panoskaltsis-Mortari, A.; McNearney, S.A.; Yun, G.H.; Fautsch, S.K.; McKenna, D.; Le, C.; Defor, T.E.; Burns, L.J.; et al. Successful adoptive transfer and in vivo expansion of human haploidentical NK cells in patients with cancer. *Blood* **2005**, *105*, 3051–3057. [[CrossRef](#)]
17. Qu, C.; Zhang, H.; Cao, H.; Tang, L.; Mo, H.; Liu, F.; Zhang, L.; Yi, Z.; Long, L.; Yan, L.; et al. Tumor buster—Where will the CAR-T cell therapy ‘missile’ go? *Mol. Cancer* **2022**, *21*, 201. [[CrossRef](#)]
18. Donninger, H.; Li, C.; Eaton, J.W.; Yaddanapudi, K. Cancer Vaccines: Promising Therapeutics or an Unattainable Dream. *Vaccines* **2021**, *9*, 668. [[CrossRef](#)] [[PubMed](#)]
19. Kim, C.G.; Sang, Y.B.; Lee, J.H.; Chon, H.J. Combining Cancer Vaccines with Immunotherapy: Establishing a New Immunological Approach. *Int. J. Mol. Sci.* **2021**, *22*, 8035. [[CrossRef](#)]
20. Corral, L.G.; Haslett, P.A.; Muller, G.W.; Chen, R.; Wong, L.M.; Ocampo, C.J.; Patterson, R.T.; Stirling, D.I.; Kaplan, G. Differential cytokine modulation and T cell activation by two distinct classes of thalidomide analogues that are potent inhibitors of TNF-alpha. *J. Immunol.* **1999**, *163*, 380–386. [[CrossRef](#)] [[PubMed](#)]
21. Davies, F.E.; Raje, N.; Hideshima, T.; Lentzsch, S.; Young, G.; Tai, Y.T.; Lin, B.; Podar, K.; Gupta, D.; Chauhan, D.; et al. Thalidomide and immunomodulatory derivatives augment natural killer cell cytotoxicity in multiple myeloma. *Blood* **2001**, *98*, 210–216. [[CrossRef](#)]
22. Haslett, P.A.; Klausner, J.D.; Makonkawkeyoon, S.; Moreira, A.; Metatrapi, P.; Boyle, B.; Kunachiwa, W.; Maneekarn, N.; Vongchan, P.; Corral, L.G.; et al. Thalidomide stimulates T cell responses and interleukin 12 production in HIV-infected patients. *AIDS Res. Hum. Retroviruses* **1999**, *15*, 1169–1179. [[CrossRef](#)]
23. Singhal, S.; Mehta, J.; Desikan, R.; Ayers, D.; Roberson, P.; Eddlemon, P.; Munshi, N.; Anaissie, E.; Wilson, C.; Dhodapkar, M.; et al. Antitumor activity of thalidomide in refractory multiple myeloma. *N. Engl. J. Med.* **1999**, *341*, 1565–1571. [[CrossRef](#)]
24. Kennedy, L.B.; Salama, A.K.S. A review of cancer immunotherapy toxicity. *CA Cancer J. Clin.* **2020**, *70*, 86–104. [[CrossRef](#)] [[PubMed](#)]
25. Michot, J.M.; Bigenwald, C.; Champiat, S.; Collins, M.; Carbonnel, F.; Postel-Vinay, S.; Berdelou, A.; Varga, A.; Bahleda, R.; Hollebecque, A.; et al. Immune-related adverse events with immune checkpoint blockade: A comprehensive review. *Eur J. Cancer* **2016**, *54*, 139–148. [[CrossRef](#)]
26. Postow, M.A.; Sidlow, R.; Hellmann, M.D. Immune-Related Adverse Events Associated with Immune Checkpoint Blockade. *N. Engl. J. Med.* **2018**, *378*, 158–168. [[CrossRef](#)]
27. Ramos-Casals, M.; Brahmer, J.R.; Callahan, M.K.; Flores-Chavez, A.; Keegan, N.; Khamashta, M.A.; Lambotte, O.; Mariette, X.; Prat, A.; Suarez-Almazor, M.E. Immune-related adverse events of checkpoint inhibitors. *Nat. Rev. Dis. Primers* **2020**, *6*, 38. [[CrossRef](#)] [[PubMed](#)]
28. Beyes, S.; Bediaga, N.G.; Zippo, A. An Epigenetic Perspective on Intra-Tumour Heterogeneity: Novel Insights and New Challenges from Multiple Fields. *Cancers* **2021**, *13*, 4969. [[CrossRef](#)]
29. Easwaran, H.; Tsai, H.C.; Baylin, S.B. Cancer epigenetics: Tumor heterogeneity, plasticity of stem-like states, and drug resistance. *Mol. Cell* **2014**, *54*, 716–727. [[CrossRef](#)]
30. Darwiche, N. Epigenetic mechanisms and the hallmarks of cancer: An intimate affair. *Am. J. Cancer Res.* **2020**, *10*, 1954–1978. [[PubMed](#)]
31. Kugel, S.; Feldman, J.L.; Klein, M.A.; Silberman, D.M.; Sebastian, C.; Mermel, C.; Dobersch, S.; Clark, A.R.; Getz, G.; Denu, J.M.; et al. Identification of and Molecular Basis for SIRT6 Loss-of-Function Point Mutations in Cancer. *Cell Rep.* **2015**, *13*, 479–488. [[CrossRef](#)]
32. Peng, D.; Kryczek, I.; Nagarsheth, N.; Zhao, L.; Wei, S.; Wang, W.; Sun, Y.; Zhao, E.; Vatan, L.; Szeliga, W.; et al. Epigenetic silencing of TH1-type chemokines shapes tumour immunity and immunotherapy. *Nature* **2015**, *527*, 249–253. [[CrossRef](#)] [[PubMed](#)]
33. Topper, M.J.; Vaz, M.; Marrone, K.A.; Brahmer, J.R.; Baylin, S.B. The emerging role of epigenetic therapeutics in immuno-oncology. *Nat. Rev. Clin. Oncol.* **2020**, *17*, 75–90. [[CrossRef](#)] [[PubMed](#)]
34. Callahan, S.C.; Divenko, M.; Barrodia, P.; Singh, A.K.; Arslan, E.; Liu, Z.; Yang, J.; Anvar, N.; Amit, M.; Xie, T.; et al. KMT2D Loss Promotes Head and Neck Squamous Cell Carcinoma Through Enhancer Reprogramming and Modulation of Immune Microenvironment. *Biorxiv* **2021**. [[CrossRef](#)]
35. Kim, Y.D.; Park, S.M.; Ha, H.C.; Lee, A.R.; Won, H.; Cha, H.; Cho, S.; Cho, J.M. HDAC Inhibitor, CG-745, Enhances the Anti-Cancer Effect of Anti-PD-1 Immune Checkpoint Inhibitor by Modulation of the Immune Microenvironment. *J. Cancer* **2020**, *11*, 4059–4072. [[CrossRef](#)]

36. Zhu, H.; Bengsch, F.; Svoronos, N.; Rutkowski, M.R.; Bitler, B.G.; Allegranza, M.J.; Yokoyama, Y.; Kossenkov, A.V.; Bradner, J.E.; Conejo-Garcia, J.R.; et al. BET Bromodomain Inhibition Promotes Anti-tumor Immunity by Suppressing PD-L1 Expression. *Cell Rep.* **2016**, *16*, 2829–2837. [\[CrossRef\]](#)
37. Naik, R.R.; Singh, A.K.; Mali, A.M.; Khirade, M.F.; Bapat, S.A. A tumor deconstruction platform identifies definitive end points in the evaluation of drug responses. *Oncogene* **2016**, *35*, 727–737. [\[CrossRef\]](#)
38. Orouji, E.; Raman, A.T.; Singh, A.K.; Sorokin, A.; Arslan, E.; Ghosh, A.K.; Schulz, J.; Terranova, C.; Jiang, S.; Tang, M.; et al. Chromatin state dynamics confers specific therapeutic strategies in enhancer subtypes of colorectal cancer. *Gut* **2022**, *71*, 938–949. [\[CrossRef\]](#)
39. Singh, A.K.; Chandra, N.; Bapat, S.A. Evaluation of Epigenetic Drug Targeting of Heterogenous Tumor Cell Fractions Using Potential Biomarkers of Response in Ovarian Cancer. *Clin. Cancer Res.* **2015**, *21*, 5151–5163. [\[CrossRef\]](#)
40. Dong, Y.B., Jr.; Anand, K.S.; Bapat, S.; Clements, A.J. Transforming the future of treatment for ovarian cancer. *Clin. Exp. Pharmacol.* **2014**, *4*, 3.
41. O'Donnell, J.S.; Long, G.V.; Scolyer, R.A.; Teng, M.W.; Smyth, M.J. Resistance to PD1/PDL1 checkpoint inhibition. *Cancer Treat. Rev.* **2017**, *52*, 71–81. [\[CrossRef\]](#)
42. Pitt, J.M.; Vetizou, M.; Daillere, R.; Roberti, M.P.; Yamazaki, T.; Routy, B.; Lepage, P.; Boneca, I.G.; Chamaillard, M.; Kroemer, G.; et al. Resistance Mechanisms to Immune-Checkpoint Blockade in Cancer: Tumor-Intrinsic and -Extrinsic Factors. *Immunity* **2016**, *44*, 1255–1269. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Sharma, P.; Hu-Lieskovan, S.; Wargo, J.A.; Ribas, A. Primary, Adaptive, and Acquired Resistance to Cancer Immunotherapy. *Cell* **2017**, *168*, 707–723. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Pascual-Garcia, M.; Bonfill-Teixidor, E.; Planas-Rigol, E.; Rubio-Perez, C.; Iurlaro, R.; Arias, A.; Cuartas, I.; Sala-Hojman, A.; Escudero, L.; Martinez-Ricarte, F.; et al. LIF regulates CXCL9 in tumor-associated macrophages and prevents CD8(+) T cell tumor-infiltration impairing anti-PD1 therapy. *Nat. Commun.* **2019**, *10*, 2416. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Jones, P.A.; Takai, D. The role of DNA methylation in mammalian epigenetics. *Science* **2001**, *293*, 1068–1070. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Terranova, C.J.; Tang, M.; Maitituoheti, M.; Raman, A.T.; Ghosh, A.K.; Schulz, J.; Amin, S.B.; Orouji, E.; Tomczak, K.; Sarkar, S.; et al. Reprogramming of bivalent chromatin states in NRAS mutant melanoma suggests PRC2 inhibition as a therapeutic strategy. *Cell Rep.* **2021**, *36*, 109410. [\[CrossRef\]](#)
47. Kouzarides, T. Chromatin modifications and their function. *Cell* **2007**, *128*, 693–705. [\[CrossRef\]](#)
48. Audia, J.E.; Campbell, R.M. Histone Modifications and Cancer. *Cold Spring Harb. Perspect. Biol.* **2016**, *8*, a019521. [\[CrossRef\]](#)
49. Dawson, M.A.; Kouzarides, T. Cancer epigenetics: From mechanism to therapy. *Cell* **2012**, *150*, 12–27. [\[CrossRef\]](#)
50. McCaw, T.R.; Goel, N.; Brooke, D.J.; Katre, A.A.; Londono, A.I.; Smith, H.J.; Randall, T.D.; Arend, R.C. Class I histone deacetylase inhibition promotes CD8 T cell activation in ovarian cancer. *Cancer Med.* **2021**, *10*, 709–717. [\[CrossRef\]](#)
51. Preglej, T.; Hamminger, P.; Luu, M.; Bulat, T.; Andersen, L.; Goschl, L.; Stolz, V.; Rica, R.; Sandner, L.; Waltenberger, D.; et al. Histone deacetylases 1 and 2 restrain CD4+ cytotoxic T lymphocyte differentiation. *JCI Insight* **2020**, *5*, e133393. [\[CrossRef\]](#) [\[PubMed\]](#)
52. Hsu, F.C.; Belmonte, P.J.; Constans, M.M.; Chen, M.W.; McWilliams, D.C.; Hiebert, S.W.; Shapiro, V.S. Histone Deacetylase 3 Is Required for T Cell Maturation. *J. Immunol.* **2015**, *195*, 1578–1590. [\[CrossRef\]](#) [\[PubMed\]](#)
53. Nolz, J.C.; Harty, J.T. IL-15 regulates memory CD8+ T cell O-glycan synthesis and affects trafficking. *J. Clin. Investig.* **2014**, *124*, 1013–1026. [\[CrossRef\]](#) [\[PubMed\]](#)
54. Scheer, S.; Runting, J.; Bramhall, M.; Russ, B.; Zaini, A.; Ellemor, J.; Rodrigues, G.; Ng, J.; Zaph, C. The Methyltransferase DOT1L Controls Activation and Lineage Integrity in CD4(+) T Cells during Infection and Inflammation. *Cell Rep.* **2020**, *33*, 108505. [\[CrossRef\]](#)
55. Onodera, A.; Kiuchi, M.; Kokubo, K.; Kato, M.; Ogino, T.; Horiuchi, S.; Kanai, U.; Hirahara, K.; Nakayama, T. Menin Controls the Memory Th2 Cell Function by Maintaining the Epigenetic Integrity of Th2 Cells. *J. Immunol.* **2017**, *199*, 1153–1162. [\[CrossRef\]](#)
56. Cribbs, A.P.; Terlecki-Zaniewicz, S.; Philpott, M.; Baardman, J.; Ahern, D.; Lindow, M.; Obad, S.; Oerum, H.; Sampey, B.; Mander, P.K.; et al. Histone H3K27me3 demethylases regulate human Th17 cell development and effector functions by impacting on metabolism. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 6056–6066. [\[CrossRef\]](#) [\[PubMed\]](#)
57. O'Neill, R.E.; Cao, X. Co-stimulatory and co-inhibitory pathways in cancer immunotherapy. *Adv. Cancer Res.* **2019**, *143*, 145–194. [\[CrossRef\]](#)
58. Maitituoheti, M.A.S.; Tang, M.; Ho, L.-L.; Terranova, C.; Galani, K.; Keung, E.Z.; Creasy, C.A.; Wu, M.; Chen, J.; Chen, N.; et al. Enhancer Reprogramming in Melanoma Immune Checkpoint Therapy Resistance. *bioRxiv* **2022**.
59. Ni, L.; Dong, C. New B7 Family Checkpoints in Human Cancers. *Mol. Cancer Ther.* **2017**, *16*, 1203–1211. [\[CrossRef\]](#)
60. Wang, C.; Feng, H.; Cheng, X.; Liu, K.; Cai, D.; Zhao, R. Potential Therapeutic Targets of B7 Family in Colorectal Cancer. *Front. Immunol.* **2020**, *11*, 681. [\[CrossRef\]](#)
61. Saleh, R.; Toor, S.M.; Sasidharan Nair, V.; Elkord, E. Role of Epigenetic Modifications in Inhibitory Immune Checkpoints in Cancer Development and Progression. *Front. Immunol.* **2020**, *11*, 1469. [\[CrossRef\]](#) [\[PubMed\]](#)
62. Buttler, C.A.; Chuong, E.B. Emerging roles for endogenous retroviruses in immune epigenetic regulation. *Immunol. Rev.* **2022**, *305*, 165–178. [\[CrossRef\]](#) [\[PubMed\]](#)
63. Hu, C.; Liu, X.; Zeng, Y.; Liu, J.; Wu, F. DNA methyltransferase inhibitors combination therapy for the treatment of solid tumor: Mechanism and clinical application. *Clin. Epigenetics* **2021**, *13*, 166. [\[CrossRef\]](#) [\[PubMed\]](#)

64. Nijhof, I.S.; Casneuf, T.; van Velzen, J.; van Kessel, B.; Axel, A.E.; Syed, K.; Groen, R.W.; van Duin, M.; Sonneveld, P.; Minnema, M.C.; et al. CD38 expression and complement inhibitors affect response and resistance to daratumumab therapy in myeloma. *Blood* **2016**, *128*, 959–970. [[CrossRef](#)]
65. Woods, D.M.; Sodre, A.L.; Villagra, A.; Sarnaik, A.; Sotomayor, E.M.; Weber, J. HDAC Inhibition Upregulates PD-1 Ligands in Melanoma and Augments Immunotherapy with PD-1 Blockade. *Cancer Immunol. Res.* **2015**, *3*, 1375–1385. [[CrossRef](#)] [[PubMed](#)]
66. Braun, D.A.; Hou, Y.; Bakouny, Z.; Ficial, M.; Sant' Angelo, M.; Forman, J.; Ross-Macdonald, P.; Berger, A.C.; Jegede, O.A.; Elagina, L.; et al. Interplay of somatic alterations and immune infiltration modulates response to PD-1 blockade in advanced clear cell renal cell carcinoma. *Nat. Med.* **2020**, *26*, 909–918. [[CrossRef](#)] [[PubMed](#)]
67. Griffin, G.K.; Wu, J.; Iracheta-Vellve, A.; Patti, J.C.; Hsu, J.; Davis, T.; Dele-Oni, D.; Du, P.P.; Halawi, A.G.; Ishizuka, J.J.; et al. Epigenetic silencing by SETDB1 suppresses tumour intrinsic immunogenicity. *Nature* **2021**, *595*, 309–314. [[CrossRef](#)]
68. Harjes, U. SETDB1, a new target for immunotherapy. *Nat. Rev. Cancer* **2021**, *21*, 412. [[CrossRef](#)]
69. Dutta, R.; Khalil, R.; Mayilsamy, K.; Green, R.; Howell, M.; Bharadwaj, S.; Mohapatra, S.S.; Mohapatra, S. Combination Therapy of Mithramycin A and Immune Checkpoint Inhibitor for the Treatment of Colorectal Cancer in an Orthotopic Murine Model. *Front. Immunol.* **2021**, *12*, 706133. [[CrossRef](#)]
70. Andrieu, G.P.; Shafran, J.S.; Smith, C.L.; Belkina, A.C.; Casey, A.N.; Jafari, N.; Denis, G.V. BET protein targeting suppresses the PD-1/PD-L1 pathway in triple-negative breast cancer and elicits anti-tumor immune response. *Cancer Lett.* **2019**, *465*, 45–58. [[CrossRef](#)]
71. Adeegbe, D.O.; Liu, S.; Hattersley, M.M.; Bowden, M.; Zhou, C.W.; Li, S.; Vlahos, R.; Grondine, M.; Dolgalev, I.; Ivanova, E.V.; et al. BET Bromodomain Inhibition Cooperates with PD-1 Blockade to Facilitate Antitumor Response in Kras-Mutant Non-Small Cell Lung Cancer. *Cancer Immunol. Res.* **2018**, *6*, 1234–1245. [[CrossRef](#)]
72. Klumper, N.; Ralser, D.J.; Bawden, E.G.; Landsberg, J.; Zarbl, R.; Kristiansen, G.; Toma, M.; Ritter, M.; Holzel, M.; Ellinger, J.; et al. LAG3 (LAG-3, CD223) DNA methylation correlates with LAG3 expression by tumor and immune cells, immune cell infiltration, and overall survival in clear cell renal cell carcinoma. *J. Immunother. Cancer* **2020**, *8*, e000552. [[CrossRef](#)] [[PubMed](#)]
73. Wang, Y.; Wang, J.; Meng, J.; Jiang, H.; Zhao, J.; Qian, H.; Chen, T. Epigenetic Modification Mediates the Increase of LAG-3(+) T Cells in Chronic Osteomyelitis. *Inflammation* **2017**, *40*, 414–421. [[CrossRef](#)] [[PubMed](#)]
74. Li, X.; Hu, W.; Zheng, X.; Zhang, C.; Du, P.; Zheng, Z.; Yang, Y.; Wu, J.; Ji, M.; Jiang, J.; et al. Emerging immune checkpoints for cancer therapy. *Acta Oncol.* **2015**, *54*, 1706–1713. [[CrossRef](#)] [[PubMed](#)]
75. Moriyama, K.; Kukita, A.; Li, Y.J.; Uehara, N.; Zhang, J.Q.; Takahashi, I.; Kukita, T. Regulation of osteoclastogenesis through Tim-3: Possible involvement of the Tim-3/galectin-9 system in the modulation of inflammatory bone destruction. *Lab. Invest.* **2014**, *94*, 1200–1211. [[CrossRef](#)] [[PubMed](#)]
76. Zhang, H.; Song, Y.; Yang, H.; Liu, Z.; Gao, L.; Liang, X.; Ma, C. Tumor cell-intrinsic Tim-3 promotes liver cancer via NF-kappaB/IL-6/STAT3 axis. *Oncogene* **2018**, *37*, 2456–2468. [[CrossRef](#)]
77. Zhang, L.; Tian, S.; Pei, M.; Zhao, M.; Wang, L.; Jiang, Y.; Yang, T.; Zhao, J.; Song, L.; Yang, X. Crosstalk between histone modification and DNA methylation orchestrates the epigenetic regulation of the costimulatory factors, Tim3 and galectin9, in cervical cancer. *Oncol. Rep.* **2019**, *42*, 2655–2669. [[CrossRef](#)] [[PubMed](#)]
78. Zhang, L.; Tian, S.; Zhao, M.; Yang, T.; Quan, S.; Yang, Q.; Song, L.; Yang, X. SUV39H1-DNMT3A-mediated epigenetic regulation of Tim-3 and galectin-9 in the cervical cancer. *Cancer Cell Int.* **2020**, *20*, 325. [[CrossRef](#)]
79. Niebel, D.; Frohlich, A.; Zarbl, R.; Fietz, S.; de Vos, L.; Vogt, T.J.; Dietrich, J.; Sirokay, J.; Kuster, P.; Saavedra, G.; et al. DNA methylation regulates TIGIT expression within the melanoma microenvironment, is prognostic for overall survival, and predicts progression-free survival in patients treated with anti-PD-1 immunotherapy. *Clin. Epigenetics* **2022**, *14*, 50. [[CrossRef](#)]
80. Gibbs, Z.A.; Whitehurst, A.W. Emerging Contributions of Cancer/Testis Antigens to Neoplastic Behaviors. *Trends Cancer* **2018**, *4*, 701–712. [[CrossRef](#)]
81. Wang, C.; Gu, Y.; Zhang, K.; Xie, K.; Zhu, M.; Dai, N.; Jiang, Y.; Guo, X.; Liu, M.; Dai, J.; et al. Systematic identification of genes with a cancer-testis expression pattern in 19 cancer types. *Nat. Commun.* **2016**, *7*, 10499. [[CrossRef](#)]
82. Griffiths, E.A.; Srivastava, P.; Matsuzaki, J.; Brumberger, Z.; Wang, E.S.; Kocent, J.; Miller, A.; Roloff, G.W.; Wong, H.Y.; Paluch, B.E.; et al. NY-ESO-1 Vaccination in Combination with Decitabine Induces Antigen-Specific T-lymphocyte Responses in Patients with Myelodysplastic Syndrome. *Clin. Cancer Res.* **2018**, *24*, 1019–1029. [[CrossRef](#)]
83. Ishihara, M.; Kitano, S.; Kageyama, S.; Miyahara, Y.; Yamamoto, N.; Kato, H.; Mishima, H.; Hattori, H.; Funakoshi, T.; Kojima, T.; et al. NY-ESO-1-specific redirected T cells with endogenous TCR knockdown mediate tumor response and cytokine release syndrome. *J. Immunother. Cancer* **2022**, *10*, e003811. [[CrossRef](#)]
84. Grunewald, C.M.; Schulz, W.A.; Skowron, M.A.; Hoffmann, M.J.; Niegisch, G. Tumor immunotherapy—The potential of epigenetic drugs to overcome resistance. *Transl. Cancer Res.* **2018**, *7*, 1151–1160. [[CrossRef](#)]
85. Khan, A.N.; Gregorie, C.J.; Tomasi, T.B. Histone deacetylase inhibitors induce TAP, LMP, Tapasin genes and MHC class I antigen presentation by melanoma cells. *Cancer Immunol. Immunother.* **2008**, *57*, 647–654. [[CrossRef](#)] [[PubMed](#)]
86. Magner, W.J.; Kazim, A.L.; Stewart, C.; Romano, M.A.; Catalano, G.; Grande, C.; Keiser, N.; Santaniello, F.; Tomasi, T.B. Activation of MHC class I, II, and CD40 gene expression by histone deacetylase inhibitors. *J. Immunol.* **2000**, *165*, 7017–7024. [[CrossRef](#)] [[PubMed](#)]
87. Setiadi, A.F.; Omilusik, K.; David, M.D.; Seipp, R.P.; Hartikainen, J.; Gopaul, R.; Choi, K.B.; Jefferies, W.A. Epigenetic enhancement of antigen processing and presentation promotes immune recognition of tumors. *Cancer Res.* **2008**, *68*, 9601–9607. [[CrossRef](#)]

88. Maeda, T.; Towatari, M.; Kosugi, H.; Saito, H. Up-regulation of costimulatory/adhesion molecules by histone deacetylase inhibitors in acute myeloid leukemia cells. *Blood* **2000**, *96*, 3847–3856. [\[CrossRef\]](#)
89. Wang, L.X.; Mei, Z.Y.; Zhou, J.H.; Yao, Y.S.; Li, Y.H.; Xu, Y.H.; Li, J.X.; Gao, X.N.; Zhou, M.H.; Jiang, M.M.; et al. Low dose decitabine treatment induces CD80 expression in cancer cells and stimulates tumor specific cytotoxic T lymphocyte responses. *PLoS ONE* **2013**, *8*, e62924. [\[CrossRef\]](#) [\[PubMed\]](#)
90. Federico, A.; Steinfass, T.; Larribere, L.; Novak, D.; Moris, F.; Nunez, L.E.; Umansky, V.; Utikal, J. Mithramycin A and Mithralog EC-8042 Inhibit SETDB1 Expression and Its Oncogenic Activity in Malignant Melanoma. *Mol. Ther. Oncolytics* **2020**, *18*, 83–99. [\[CrossRef\]](#)
91. Lin, J.; Guo, D.; Liu, H.; Zhou, W.; Wang, C.; Muller, I.; Kossenkov, A.V.; Drapkin, R.; Bitler, B.G.; Helin, K.; et al. The SETDB1-TRIM28 Complex Suppresses Antitumor Immunity. *Cancer Immunol. Res.* **2021**, *9*, 1413–1424. [\[CrossRef\]](#)
92. Truong, A.S.; Zhou, M.; Krishnan, B.; Utsumi, T.; Manocha, U.; Stewart, K.G.; Beck, W.; Rose, T.L.; Milowsky, M.I.; He, X.; et al. Entinostat induces antitumor immune responses through immune editing of tumor neoantigens. *J. Clin. Invest.* **2021**, *131*, e138560. [\[CrossRef\]](#)
93. Binnewies, M.; Roberts, E.W.; Kersten, K.; Chan, V.; Fearon, D.F.; Merad, M.; Coussens, L.M.; Gabrilovich, D.I.; Ostrand-Rosenberg, S.; Hedrick, C.C.; et al. Understanding the tumor immune microenvironment (TIME) for effective therapy. *Nat. Med.* **2018**, *24*, 541–550. [\[CrossRef\]](#)
94. Xie, Z.; Ago, Y.; Okada, N.; Tachibana, M. Valproic acid attenuates immunosuppressive function of myeloid-derived suppressor cells. *J. Pharmacol. Sci.* **2018**, *137*, 359–365. [\[CrossRef\]](#) [\[PubMed\]](#)
95. Xie, Z.; Ikegami, T.; Ago, Y.; Okada, N.; Tachibana, M. Valproic acid attenuates CCR2-dependent tumor infiltration of monocytic myeloid-derived suppressor cells, limiting tumor progression. *Oncoimmunology* **2020**, *9*, 1734268. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Wang, H.F.; Ning, F.; Liu, Z.C.; Wu, L.; Li, Z.Q.; Qi, Y.F.; Zhang, G.; Wang, H.S.; Cai, S.H.; Du, J. Histone deacetylase inhibitors deplete myeloid-derived suppressor cells induced by 4T1 mammary tumors in vivo and in vitro. *Cancer Immunol. Immunother.* **2017**, *66*, 355–366. [\[CrossRef\]](#) [\[PubMed\]](#)
97. Fontenot, J.D.; Rasmussen, J.P.; Williams, L.M.; Dooley, J.L.; Farr, A.G.; Rudensky, A.Y. Regulatory T cell lineage specification by the forkhead transcription factor foxp3. *Immunity* **2005**, *22*, 329–341. [\[CrossRef\]](#) [\[PubMed\]](#)
98. Marie, J.C.; Letterio, J.J.; Gavin, M.; Rudensky, A.Y. TGF-beta1 maintains suppressor function and Foxp3 expression in CD4+CD25+ regulatory T cells. *J. Exp. Med.* **2005**, *201*, 1061–1067. [\[CrossRef\]](#)
99. Burchill, M.A.; Yang, J.; Vogtenhuber, C.; Blazar, B.R.; Farrar, M.A. IL-2 receptor beta-dependent STAT5 activation is required for the development of Foxp3+ regulatory T cells. *J. Immunol.* **2007**, *178*, 280–290. [\[CrossRef\]](#)
100. Kim, H.P.; Leonard, W.J. CREB/ATF-dependent T cell receptor-induced FoxP3 gene expression: A role for DNA methylation. *J. Exp. Med.* **2007**, *204*, 1543–1551. [\[CrossRef\]](#)
101. Lal, G.; Zhang, N.; van der Touw, W.; Ding, Y.; Ju, W.; Bottinger, E.P.; Reid, S.P.; Levy, D.E.; Bromberg, J.S. Epigenetic regulation of Foxp3 expression in regulatory T cells by DNA methylation. *J. Immunol.* **2009**, *182*, 259–273. [\[CrossRef\]](#)
102. Chen, W.; Jin, W.; Hardegen, N.; Lei, K.J.; Li, L.; Marinos, N.; McGrady, G.; Wahl, S.M. Conversion of peripheral CD4+CD25- naive T cells to CD4+CD25+ regulatory T cells by TGF-beta induction of transcription factor Foxp3. *J. Exp. Med.* **2003**, *198*, 1875–1886. [\[CrossRef\]](#) [\[PubMed\]](#)
103. Tao, R.; de Zoeten, E.F.; Ozkaynak, E.; Chen, C.; Wang, L.; Porrett, P.M.; Li, B.; Turka, L.A.; Olson, E.N.; Greene, M.I.; et al. Deacetylase inhibition promotes the generation and function of regulatory T cells. *Nat. Med.* **2007**, *13*, 1299–1307. [\[CrossRef\]](#) [\[PubMed\]](#)
104. Zhang, H.; Xiao, Y.; Zhu, Z.; Li, B.; Greene, M.I. Immune regulation by histone deacetylases: A focus on the alteration of FOXP3 activity. *Immunol. Cell Biol.* **2012**, *90*, 95–100. [\[CrossRef\]](#)
105. Ohkura, N.; Hamaguchi, M.; Morikawa, H.; Sugimura, K.; Tanaka, A.; Ito, Y.; Osaki, M.; Tanaka, Y.; Yamashita, R.; Nakano, N.; et al. T cell receptor stimulation-induced epigenetic changes and Foxp3 expression are independent and complementary events required for Treg cell development. *Immunity* **2012**, *37*, 785–799. [\[CrossRef\]](#)
106. Wang, D.; Quiros, J.; Mahuron, K.; Pai, C.C.; Ranzani, V.; Young, A.; Silveria, S.; Harwin, T.; Abnousian, A.; Pagani, M.; et al. Targeting EZH2 Reprograms Intratumoral Regulatory T Cells to Enhance Cancer Immunity. *Cell Rep.* **2018**, *23*, 3262–3274. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Rosenzweig, J.M.; Glenn, J.D.; Calabresi, P.A.; Whartenby, K.A. KLF4 modulates expression of IL-6 in dendritic cells via both promoter activation and epigenetic modification. *J. Biol. Chem.* **2013**, *288*, 23868–23874. [\[CrossRef\]](#) [\[PubMed\]](#)
108. Shen, X.; He, Z.; Li, H.; Yao, C.; Zhang, Y.; He, L.; Li, S.; Huang, J.; Guo, Z. Distinct functional patterns of gene promoter hypomethylation and hypermethylation in cancer genomes. *PLoS ONE* **2012**, *7*, e44822. [\[CrossRef\]](#)
109. Ma, D.; Jiang, C.; Hu, X.; Liu, H.; Li, Q.; Li, T.; Yang, Y.; Li, O. Methylation patterns of the IFN-gamma gene in cervical cancer tissues. *Sci. Rep.* **2014**, *4*, 6331. [\[CrossRef\]](#)
110. Wang, F.; Xu, J.; Zhu, Q.; Qin, X.; Cao, Y.; Lou, J.; Xu, Y.; Ke, X.; Li, Q.; Xie, E.; et al. Downregulation of IFNG in CD4(+) T cells in lung cancer through hypermethylation: A possible mechanism of tumor-induced immunosuppression. *PLoS ONE* **2013**, *8*, e79064. [\[CrossRef\]](#)
111. Rincon-Orozco, B.; Halec, G.; Rosenberger, S.; Muschik, D.; Nindl, I.; Bachmann, A.; Ritter, T.M.; Dondog, B.; Ly, R.; Bosch, F.X.; et al. Epigenetic silencing of interferon-kappa in human papillomavirus type 16-positive cells. *Cancer Res.* **2009**, *69*, 8718–8725. [\[CrossRef\]](#)

112. Schoenborn, J.R.; Dorschner, M.O.; Sekimata, M.; Santer, D.M.; Shnyreva, M.; Fitzpatrick, D.R.; Stamatoyannopoulos, J.A.; Wilson, C.B. Comprehensive epigenetic profiling identifies multiple distal regulatory elements directing transcription of the gene encoding interferon-gamma. *Nat. Immunol.* **2007**, *8*, 732–742. [[CrossRef](#)]
113. Santangelo, S.; Cousins, D.J.; Winkelmann, N.; Triantaphyllopoulos, K.; Staynov, D.Z. Chromatin structure and DNA methylation of the IL-4 gene in human T(H)2 cells. *Chromosome Res.* **2009**, *17*, 485–496. [[CrossRef](#)] [[PubMed](#)]
114. Starnes, T.; Rasila, K.K.; Robertson, M.J.; Brahmi, Z.; Dahl, R.; Christopherson, K.; Hromas, R. The chemokine CXCL14 (BRAF) stimulates activated NK cell migration: Implications for the downregulation of CXCL14 in malignancy. *Exp. Hematol.* **2006**, *34*, 1101–1105. [[CrossRef](#)] [[PubMed](#)]
115. Hu, C.; Lin, F.; Zhu, G.; Xue, X.; Ding, Y.; Zhao, Z.; Zhang, L.; Shen, X. Abnormal hypermethylation of promoter region downregulates chemokine CXC ligand 14 expression in gastric cancer. *Int. J. Oncol.* **2013**, *43*, 1487–1494. [[CrossRef](#)] [[PubMed](#)]
116. Song, E.Y.; Shurin, M.R.; Tourkova, I.L.; Gutkin, D.W.; Shurin, G.V. Epigenetic mechanisms of promigratory chemokine CXCL14 regulation in human prostate cancer cells. *Cancer Res.* **2010**, *70*, 4394–4401. [[CrossRef](#)] [[PubMed](#)]
117. Ramos, E.A.; Camargo, A.A.; Braun, K.; Slowik, R.; Cavalli, I.J.; Ribeiro, E.M.; Pedrosa Fde, O.; de Souza, E.M.; Costa, F.F.; Klassen, G. Simultaneous CXCL12 and ESR1 CpG island hypermethylation correlates with poor prognosis in sporadic breast cancer. *BMC Cancer* **2010**, *10*, 23. [[CrossRef](#)]
118. Tekpli, X.; Landvik, N.E.; Anmarkud, K.H.; Skaug, V.; Haugen, A.; Zienolddiny, S. DNA methylation at promoter regions of interleukin 1B, interleukin 6, and interleukin 8 in non-small cell lung cancer. *Cancer Immunol. Immunother.* **2013**, *62*, 337–345. [[CrossRef](#)]
119. Baird, A.M.; Leonard, J.; Naicker, K.M.; Kilmartin, L.; O’Byrne, K.J.; Gray, S.G. IL-23 is pro-proliferative, epigenetically regulated and modulated by chemotherapy in non-small cell lung cancer. *Lung Cancer* **2013**, *79*, 83–90. [[CrossRef](#)]
120. Suzuki, M.; Iizasa, T.; Nakajima, T.; Kubo, R.; Iyoda, A.; Hiroshima, K.; Nakatani, Y.; Fujisawa, T. Aberrant methylation of IL-12Rbeta2 gene in lung adenocarcinoma cells is associated with unfavorable prognosis. *Ann. Surg. Oncol.* **2007**, *14*, 2636–2642. [[CrossRef](#)]
121. Suzuki, M.; Wada, H.; Yoshino, M.; Tian, L.; Shigematsu, H.; Suzuki, H.; Alaa, M.; Tamura, H.; Fujiwara, T.; Nagato, K.; et al. Molecular characterization of chronic obstructive pulmonary disease-related non-small cell lung cancer through aberrant methylation and alterations of EGFR signaling. *Ann. Surg. Oncol.* **2010**, *17*, 878–888. [[CrossRef](#)]
122. Conte, M.; De Palma, R.; Altucci, L. HDAC inhibitors as epigenetic regulators for cancer immunotherapy. *Int. J. Biochem. Cell Biol.* **2018**, *98*, 65–74. [[CrossRef](#)] [[PubMed](#)]
123. Maio, M.; Covre, A.; Fratta, E.; Di Giacomo, A.M.; Taverna, P.; Natali, P.G.; Coral, S.; Sigalotti, L. Molecular Pathways: At the Crossroads of Cancer Epigenetics and Immunotherapy. *Clin. Cancer Res.* **2015**, *21*, 4040–4047. [[CrossRef](#)] [[PubMed](#)]
124. Park, J.; Thomas, S.; Munster, P.N. Epigenetic modulation with histone deacetylase inhibitors in combination with immunotherapy. *Epigenomics* **2015**, *7*, 641–652. [[CrossRef](#)]
125. Fraietta, J.A.; Nobles, C.L.; Sammons, M.A.; Lundh, S.; Carty, S.A.; Reich, T.J.; Cogdill, A.P.; Morrisette, J.J.D.; DeNizio, J.E.; Reddy, S.; et al. Disruption of TET2 promotes the therapeutic efficacy of CD19-targeted T cells. *Nature* **2018**, *558*, 307–312. [[CrossRef](#)] [[PubMed](#)]
126. Foskolou, I.P.; Barbieri, L.; Vernet, A.; Bargiela, D.; Cunha, P.P.; Velica, P.; Suh, E.; Pietsch, S.; Matuleviciute, R.; Rundqvist, H.; et al. The S enantiomer of 2-hydroxyglutarate increases central memory CD8 populations and improves CAR-T therapy outcome. *Blood Adv.* **2020**, *4*, 4483–4493. [[CrossRef](#)]
127. Li, S.; Xue, L.; Wang, M.; Qiang, P.; Xu, H.; Zhang, X.; Kang, W.; You, F.; Xu, H.; Wang, Y.; et al. Decitabine enhances cytotoxic effect of T cells with an anti-CD19 chimeric antigen receptor in treatment of lymphoma. *Onco Targets Ther.* **2019**, *12*, 5627–5638. [[CrossRef](#)]
128. Zou, F.; Lu, L.; Liu, J.; Xia, B.; Zhang, W.; Hu, Q.; Liu, W.; Zhang, Y.; Lin, Y.; Jing, S.; et al. Engineered triple inhibitory receptor resistance improves anti-tumor CAR-T cell performance via CD56. *Nat. Commun.* **2019**, *10*, 4109. [[CrossRef](#)] [[PubMed](#)]
129. Kailayangiri, S.; Altvater, B.; Lesch, S.; Balbach, S.; Gottlich, C.; Kuhnemundt, J.; Mikesch, J.H.; Schelhaas, S.; Jamitzky, S.; Meltzer, J.; et al. EZH2 Inhibition in Ewing Sarcoma Upregulates G(D2) Expression for Targeting with Gene-Modified T Cells. *Mol. Ther.* **2019**, *27*, 933–946. [[CrossRef](#)]
130. Seo, W.; Jerin, C.; Nishikawa, H. Transcriptional regulatory network for the establishment of CD8(+) T cell exhaustion. *Exp. Mol. Med.* **2021**, *53*, 202–209. [[CrossRef](#)]
131. Yang, R.; Cheng, S.; Luo, N.; Gao, R.; Yu, K.; Kang, B.; Wang, L.; Zhang, Q.; Fang, Q.; Zhang, L.; et al. Distinct epigenetic features of tumor-reactive CD8+ T cells in colorectal cancer patients revealed by genome-wide DNA methylation analysis. *Genome Biol.* **2019**, *21*, 2. [[CrossRef](#)] [[PubMed](#)]
132. Jiang, C.; He, J.; Xu, S.; Wang, Q.; Cheng, J. NR4A1 promotes LEF1 expression in the pathogenesis of papillary thyroid cancer. *Cell Death Discov.* **2022**, *8*, 46. [[CrossRef](#)] [[PubMed](#)]
133. Miller, S.A.; Weinmann, A.S. Molecular mechanisms by which T-bet regulates T-helper cell commitment. *Immunol. Rev.* **2010**, *238*, 233–246. [[CrossRef](#)] [[PubMed](#)]
134. Istaces, N.; Splittgerber, M.; Lima Silva, V.; Nguyen, M.; Thomas, S.; Le, A.; Achouri, Y.; Calonne, E.; Defrance, M.; Fuks, F.; et al. EOMES interacts with RUNX3 and BRG1 to promote innate memory cell formation through epigenetic reprogramming. *Nat. Commun.* **2019**, *10*, 3306. [[CrossRef](#)]

135. Itahashi, K.; Irie, T.; Yuda, J.; Kumagai, S.; Tanegashima, T.; Lin, Y.T.; Watanabe, S.; Goto, Y.; Suzuki, J.; Aokage, K.; et al. BATF epigenetically and transcriptionally controls the activation program of regulatory T cells in human tumors. *Sci. Immunol.* **2022**, *7*, eabk0957. [[CrossRef](#)]
136. Chang, S.; Collins, P.L.; Aune, T.M. T-bet dependent removal of Sin3A-histone deacetylase complexes at the Ifng locus drives Th1 differentiation. *J. Immunol.* **2008**, *181*, 8372–8381. [[CrossRef](#)]
137. Tumes, D.J.; Onodera, A.; Suzuki, A.; Shinoda, K.; Endo, Y.; Iwamura, C.; Hosokawa, H.; Koseki, H.; Tokoyoda, K.; Suzuki, Y.; et al. The polycomb protein Ezh2 regulates differentiation and plasticity of CD4(+) T helper type 1 and type 2 cells. *Immunity* **2013**, *39*, 819–832. [[CrossRef](#)]
138. Kurachi, M.; Barnitz, R.A.; Yosef, N.; Odorizzi, P.M.; DiIorio, M.A.; Lemieux, M.E.; Yates, K.; Godec, J.; Klatt, M.G.; Regev, A.; et al. The transcription factor BATF operates as an essential differentiation checkpoint in early effector CD8+ T cells. *Nat. Immunol.* **2014**, *15*, 373–383. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.