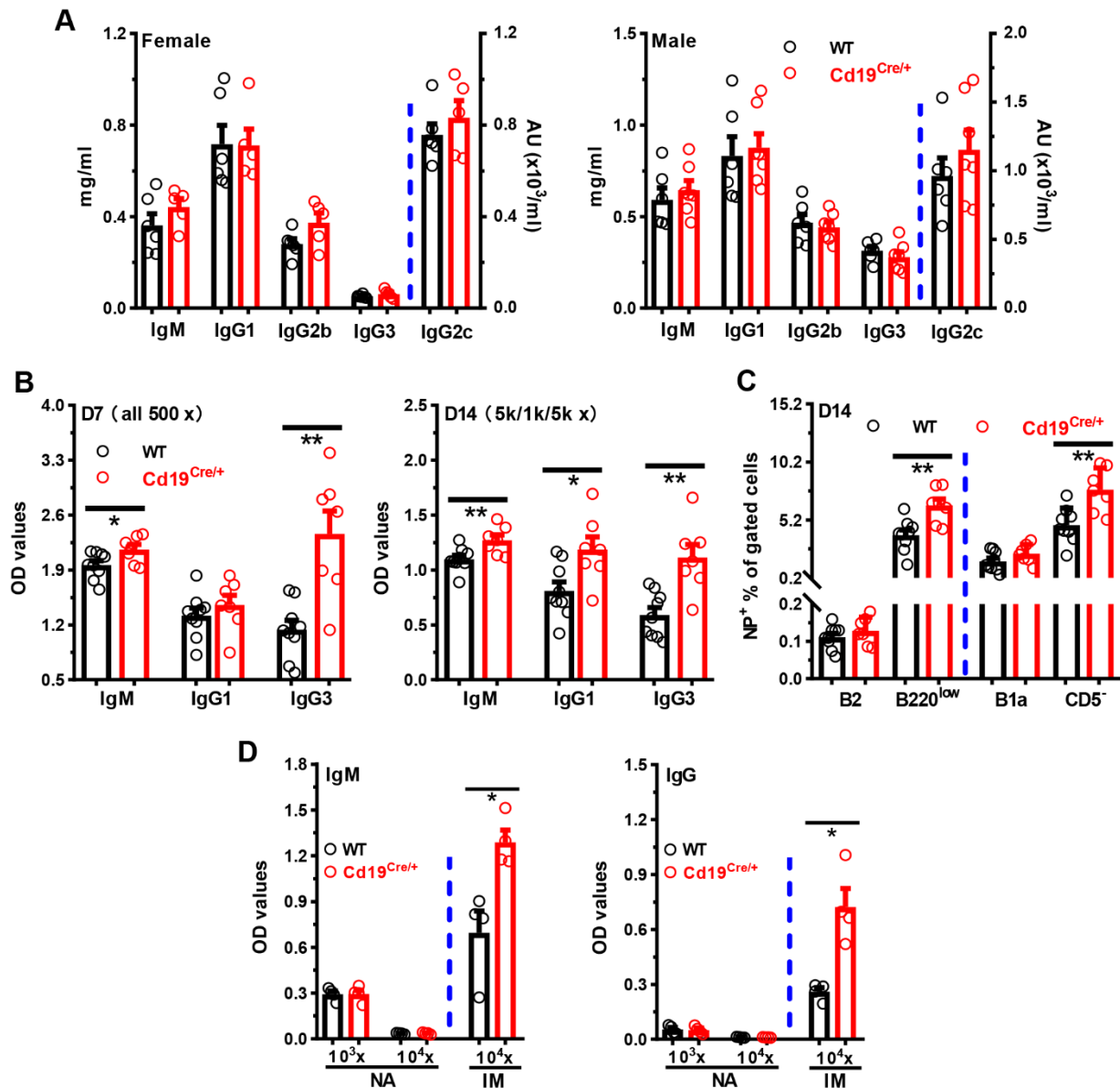
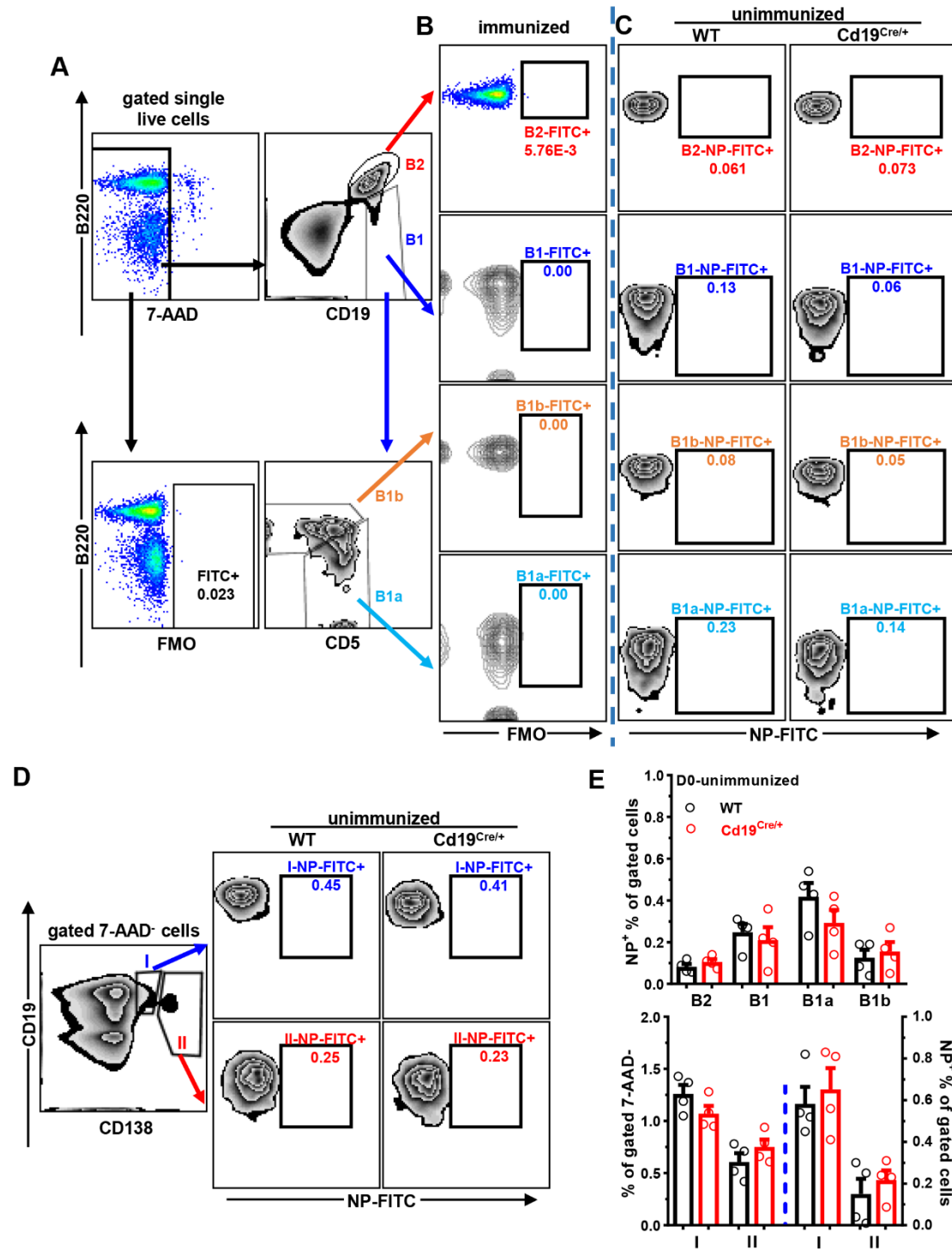


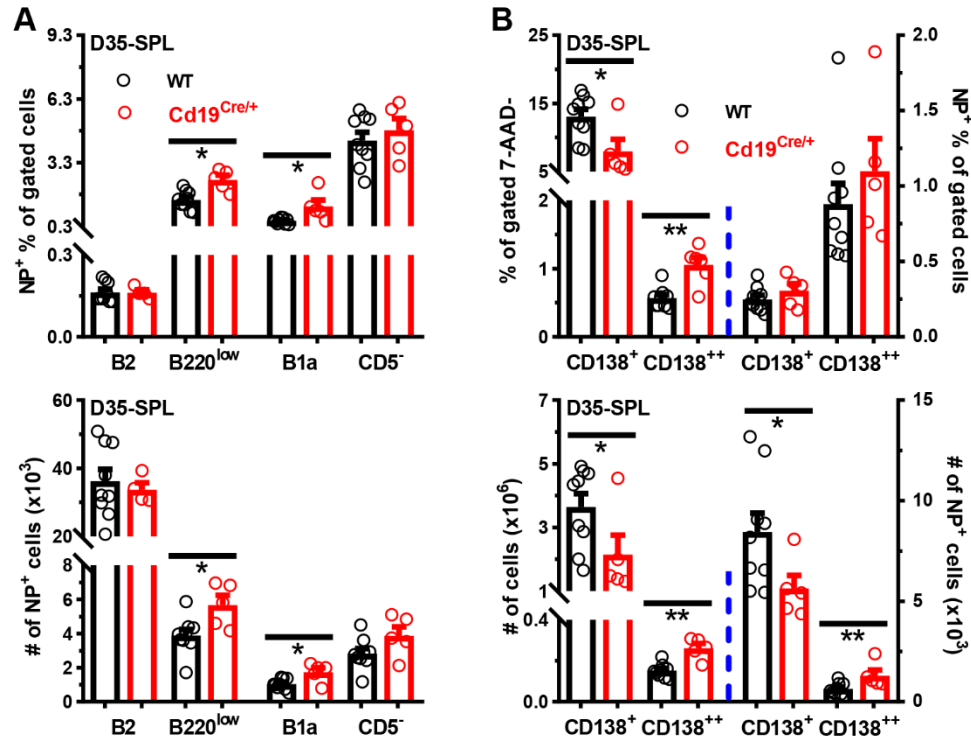
Supplementary Figure S1. Disturbed phenotypes of peripheral B cells in Cd19^{Cre/+} mice. Cells from the spleens of naive mice were stained with antibodies against CD45/B220/CD19/IgD & IgM (A&B) or CD45/CD19/CD23/AA4.1 & IgM (C&D) followed by analyses on FACS. A&C, representative FACS plots showing the gating strategies for B (CD19⁺), B2 (CD19⁺B220^{high})/B1 (CD19⁺B220^{low}) (left panel in A), or IgM^{high}IgD^{low} (I) & IgM^{low}IgD^{high} (II) cells within gated CD19⁺ B cells (right panel in A), or transitional T1 (CD19⁺AA4.1⁺IgM^{high}CD23^{low}), T2 (CD19⁺AA4.1⁺IgM^{high}CD23^{high}) & T3 (CD19⁺AA4.1⁺IgM^{low}CD23^{high}) B cells (C) in WT vs. Cd19^{Cre/+} mice. Numbers in brackets indicated the percentages of gated subsets within B cells. B&D, bar graphs showing the percent of IgM^{high}IgD^{low} & IgM^{low}IgD^{high} (B) or AA4.1⁺, T1, T2 & T3 (D) cells within gated B cells in spleens of mice. E&F, numbers of indicated B-cell subsets in the spleens (SPL, E) or peritoneal cavities (PCY, F) of mice (female, 8-10 weeks; n=5/4 for WT/Cd19^{Cre/+} group, respectively). G, bar graphs showing the percent of indicated B-cell subsets in the SPL or PCY of aged WT vs. Cd19^{Cre/+} mice (female, 16 weeks old, n=4 for each genotype). B-cell subsets were gated as illustrated in Figure 2A&C. Each symbol represented one single mouse, and results were expressed as mean \pm SEM (B, D-G). * $P < 0.05$.



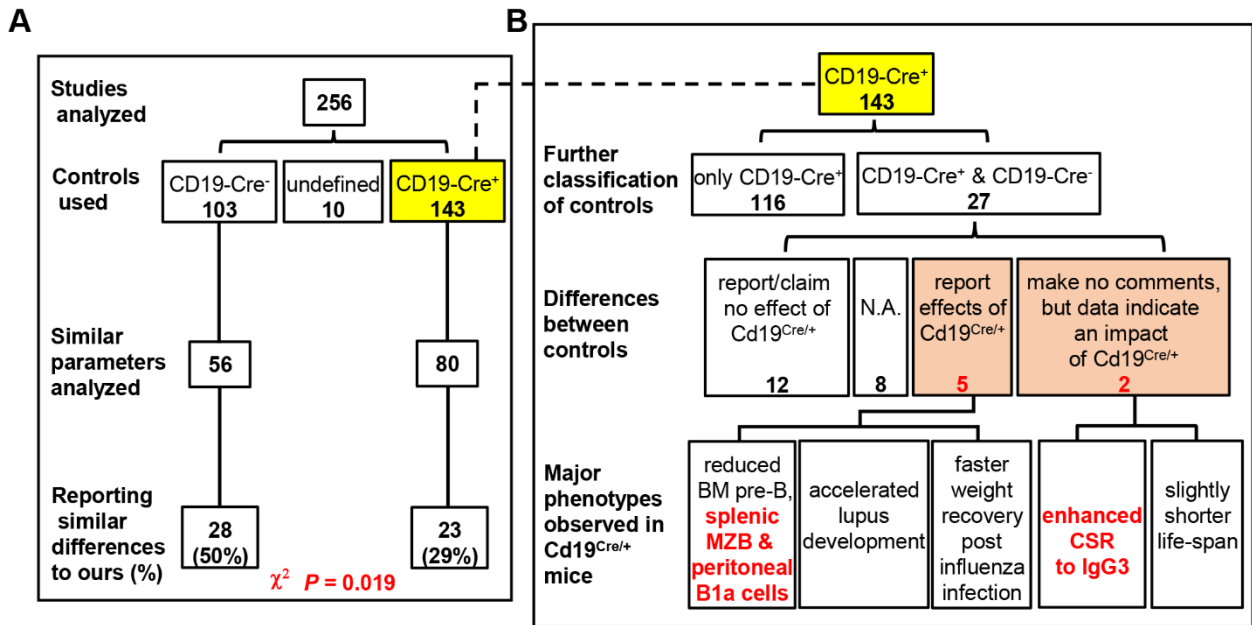
Supplementary Figure S2. Normal baseline sera antibody levels but enhanced humoral immune responses after immunization in Cd19^{Cre/+} mice. **A**, bar graphs showing levels of total IgM, IgG1, IgG2b, IgG3 and IgG2c in sera of female (~ 8 weeks old, n=6/5 for WT/Cd19^{Cre/+}, respectively) or male (8-10 weeks old, n=6/7 for WT/Cd19^{Cre/+}, respectively) mice. **B**, bar graphs showing levels of NP-specific IgM, IgG1 & IgG3 in sera of male WT or Cd19^{Cre/+} mice on D7 & D14 post immunization. **C**, a bar graph showing the percent of NP⁺ cells within gated blood B2 (CD19⁺B220^{high}), B220^{low} (CD19⁺B220^{low}), B1a (CD19⁺B220^{low}CD5⁺) or CD5⁻(CD19⁺B220^{low}CD5⁻) B cells in male mice on D14 post immunization. Mice (male, 11-13 weeks old, n=9/7 for WT/Cd19^{Cre/+}, respectively) were immunized with NP-Ficoll (10 μ g/100 μ l/mouse in PBS) intraperitoneally on D0 (**B&C**). **D**, bar graphs showing levels of NP-specific IgM & IgG in sera of naive (NA) or NP-Ficoll immunized (IM) WT or Cd19^{Cre/+} mice (male, ~12 weeks old, n=4 for each genotype). Numbers in the upper left corner (**B**) or below bars (**D**) indicated the dilutions of sera for detection of each antibody subtype. Each symbol represented one single mouse of the indicated genotype, and results were expressed as mean \pm SEM. * $P < 0.05$; ** $P < 0.01$.



Supplementary Figure S3. Comparable background stainings of NP-Ficoll in unimmunized WT vs. *Cd19^{Cre/+}* mice. Splenic or blood cells were stained with antibodies against CD45/B220/CD19/CD5 (**A-C**) or CD45/CD19/CD138 (**D**) plus 7-AAD and NP-FITC. **A**, representative FACS plots showing the gating strategies for B2, B1, B1a or B1b cells. **B**, representative FACS plots illustrating the signal in FITC channel in cells from an immunized WT mice (depicted in Figure 4) stained with all antibodies but without NP-FITC (FMO, fluorescence minus one/FITC). **C&D**, representative FACS plots showing percent of NP-FITC⁺ cells within gated B2 (B2-NP-FITC⁺), B1 (B1-NP-FITC⁺), B1a (B1a-NP-FITC⁺) & B1b (B1b-NP-FITC⁺) (**C**), or gated CD19⁺CD138^{low} (I-NP-FITC⁺)/CD19^{low}CD138^{high} (II-NP-FITC⁺) (**D**) in unimmunized WT vs. *Cd19^{Cre/+}* mice. **E**, bar graphs showing the percent of NP⁺ cells within gated B-cell subsets or frequencies of CD19⁺CD138^{low} (I)/CD19^{low}CD138^{high} (II) in the two groups of mice (female, 12 weeks old, n=4 for each genotype). Each symbol represented one single mouse of the indicated genotype, and results were expressed as mean \pm SEM (**E**).



Supplementary Figure S4. Increased numbers of NP⁺ B cells in spleens of Cd19^{Cre/+} mice immunized with NP-Ficoll. Mice (female, ~ 10 weeks of age) were immunized with NP-Ficoll (10 µg/100µl/mouse in PBS) intraperitoneally on D0. Cells in spleens (SPL) on D35 post immunization were stained with 7-AAD, NP-FITC plus antibodies against B220/CD19/CD5 & CD138. **A**, bar graphs showing the percent (*upper panel*) and numbers (*low panel*) of NP⁺ cells within gated CD19⁺B220^{high} B2, CD19⁺B220^{low} (B220^{low}), CD19⁺B220^{low}CD5⁺ B1a or CD19⁺B220^{low}CD5⁻ (CD5⁻) B cells in WT vs. Cd19^{Cre/+} mice. **B**, bar graphs showing the percent (*upper panel*) and numbers (*low panel*) of CD19⁺CD138^{low} (CD138⁺)/CD19⁺CD138^{high} (CD138⁺⁺) or NP⁺ cells within gated CD138⁺/CD138⁺⁺ B cells in the two groups of mice. Each symbol represented one single mouse of the indicated genotype (n=9/5 for WT/Cd19^{Cre/+} group, respectively), and results were expressed as mean ± SEM. * *P* < 0.05; ** *P* < 0.01.



Supplementary Figure S5. Survey of studies. In total 256 articles (Supplementary File 1) where the *Cd19^{Cre/+}* transgene was used to delete loxp-flanked sequence in B cells were analyzed, among which 10 studies contained no information on control mice or used pooled mice with different genotypes as the control group (undefined in A). CD19-Cre⁻ control mice, including WT (*gene^{+/+}*), floxed hemizygotes (*gene^{fl/+}*) or homozygotes (*gene^{fl/fl}*), were used in 103 studies, whereas mice with one *Cd19^{Cre/+}* knockin allele (CD19-Cre⁺) in combination with WT or floxed heterozygous (*gene^{fl/+}*) alleles were included as controls in the remaining 143 papers. Within these 246 articles, 136 (56 out of 103 using CD19-Cre⁻ and 80 out of 143 using CD19-Cre⁺ mice as controls, respectively) studies contained the necessary information allowing for comparisons between the control and *Cd19^{Cre/+}gene^{fl/+}* or *Cd19^{Cre/+}gene^{fl/fl}* mice in at least one of the following parameters: percent/numbers of MZB & B1a cells, or antibody levels *in vitro/vivo*. At least one of the following differences was present for the results reported to be considered similar (to the phenotype of Cd19^{Cre/+} mice described in this manuscript): reduced MZB or B1a cells, and increased antibody levels in sera or culture supernatants. χ^2 tests were employed to compare the results from published articles using CD19-Cre⁻ (56) or CD19-Cre⁺ (80) mice as controls (A). B, major impacts of the single *Cd19^{Cre/+}* knockin allele reported in the literatures. Among 27 studies using both CD19-Cre⁺ & CD19-Cre⁻ mice as controls, 5 studies reported significant differences between the two types of controls used, while data in two other publications indicated an augmented CSR to IgG3 of B cells or a reduced long-term survival rate in mice with a *Cd19^{Cre/+}* knockin allele. N.A.: no assessment on the impact of *Cd19^{Cre/+}* transgene could be made as *Cd19^{Cre/+}gene^{fl/+}* heterozygotes and CD19-Cre⁻ mice were used as controls.