

**Table S1.** *Aspergillus* strains used in this study.

Strains	Relevant genotype	References
FGSC4	<i>A. nidulans</i> wild type, <i>veA</i> <sup>+</sup>	FGSC <sup>a</sup>
RJMP1.59	<i>pyrG89; pyroA4; veA</i> <sup>+</sup>	[21]
TNJ36	<i>pyrG89; AfupyG</i> <sup>+</sup> ; <i>pyroA4; veA</i> <sup>+</sup>	[22]
$\Delta$ <i>vosA</i>	<i>pyrG89; pyroA4; \Delta vosA::AfupyG</i> <sup>+</sup> ; <i>veA</i> <sup>+</sup>	[13]
$\Delta$ <i>velB</i>	<i>pyrG89; pyroA4; \Delta velB::AfupyG</i> <sup>+</sup> ; <i>veA</i> <sup>+</sup>	[13]
$\Delta$ <i>vadH</i>	<i>pyrG89; pyroA4; \Delta vadH::AfupyG</i> <sup>+</sup> ; <i>veA</i> <sup>+</sup>	This study
C' <i>vadH</i>	<i>pyrG89; pyroA::vadH(p)::vadH::FLAG3x::pyroAb;</i> <i>\Delta vadH::AfupyrG</i> <sup>+</sup> ; <i>veA</i> <sup>+</sup>	This study
OE <i>vadH</i>	<i>pyrG89; AfupyrG</i> <sup>+</sup> ; <i>pyroA::nii(p)::vadH::FLAG::pyroA; veA</i> <sup>+</sup>	This study

<sup>a</sup> Fungal Genetic Stock Center

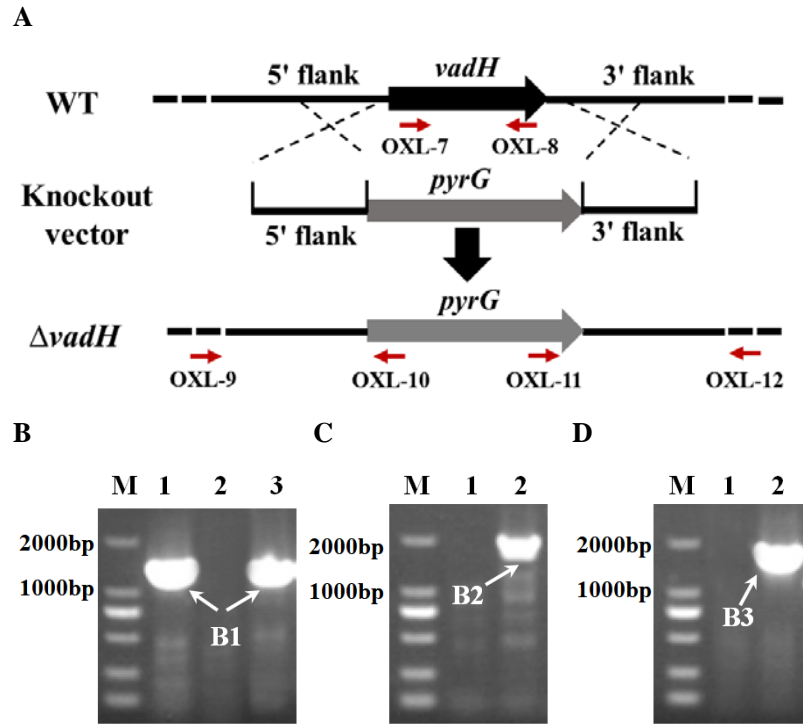
**Table S2.** Primers used in this study.

Name	Sequence (5'-3') <sup>a</sup>	Purpose
OXL-1	AAACGGTGGATGCTCAGTGGTCG	5' flanking region of vadH
OXL-2	<i>GCITTTGGCCTGTATCATGACTTCAGATGATAAAG</i> GCGGACCCTGCGT	5' vadH with AfupyrG tail
OXL-3	<i>ATCGACCGAACCTAGGTAGGGTAGTTTCCCAGTT</i> CGTGCGTTGTCTG	3' vadH with AfupyrG tail
OXL-4	TCATTGTTACTACTCGGCCTCGG	3' flanking region of vadH
OXL-5	CCGCATCGAGGTACAGCAAAGGT	5' nested of vadH
OXL-6	TACATCTCACGAAGCACACTG	3' nested of vadH
OXL-7	GCCATGCCGTTGGTGCTGTTGCT	verification of vadH ORF
OXL-8	ACGCCCCCTTGATGCCCTTGTTAC	verification of vadH ORF
OXL-9	CTGGTGTCTCCCCTGAACTTTGT	verification of 5' flanking region
OXL-10	TTGCTGTTGCCAGGTGAGGGTAT	verification of 5' flanking region
OXL-11	<b>GATAAGCAAAGTGGACTGATAGC</b>	verification of 3' flanking region
OXL-12	<b>AGTCCTTTAGGCAGCAGTGATAG</b>	verification of 3' flanking region
OXL-15	<b>GGGGTACCCAGTCAAGCGACAACCTCTACTC</b>	vadH complement <i>Kpn</i> I-pHS13
OXL-16	<b>CCCAAGCTTCACTGAGTGACAAGCTCTCACAC</b>	vadH complement <i>Hind</i> III-pHS13
OXL-31	<b>CGGAATTCATGGAGACCGCAGAGCCTGTATC</b>	vadH overexpression <i>Eco</i> R I –pHS11
OXL-32	<b>CCCAAGCTTCTCGTCACCGCGCTTCTCAATGA</b>	vadH overexpression <i>Hind</i> III –pHS11
OHS-694	GCTGAAGTCATGATACAGGCCAAA	5' AfupyrG marker
OHS-695	ATCGTCGGGAGGTATTGTCTGTCAC	3' AfupyrG marker
VadH-QF	CAACATCACAATCACCATCACC	qRT-PCR for VadH
VadH-QR	TCATGAGCAGATACTCCGTTAC	qRT-PCR for VadH
Actin-QF	GGCAACATCGTTATGTCTGG	qRT-PCR for actin
Actin-QR	CCGATCCAGACGGAGTATTT	qRT-PCR for actin
AN9219-QF	GTCGCAATGTGTCTTCGTAAAC	qRT-PCR for AN9219
AN9219-QR	CACCGAGAACTTCATTTTGTGT	qRT-PCR for AN9219
AN7030-QF	GTCATCAAGGCTCTCTACGATT	qRT-PCR for AN7030
AN7030-QR	CTGATAACGTGGAAGAAATCGC	qRT-PCR for AN7030
AN1443-QF	CTACAAGTCGGCTTCACCTATG	qRT-PCR for AN1443
AN1443-QR	GGAACATCCTGTGTCTTAGGAG	qRT-PCR for AN1443
AN1951-QF	CCATCCCACCTCCAAACTT	qRT-PCR for AN1951
AN1951-QR	TCATCGCTTATCCCCCAAAG	qRT-PCR for AN1951
AN4190-QF	CATCGAGACCATCTACCAGAC	qRT-PCR for AN4190
AN4190-QR	GATAGTGACGGTCGAGTAAGTG	qRT-PCR for AN4190
AN9473-QF	AATGGTTCCTACGAATTTGCTG	qRT-PCR for AN9473
AN9473-QR	ATAACACTGAATGTAAAGCGGC	qRT-PCR for AN9473
AN6273-QF	TCTTTCCTACGACACCAAGTAC	qRT-PCR for AN6273
AN6273-QR	GAGGTGTAAGTGATCTGGTAGC	qRT-PCR for AN6273
AN0554-QF	GTCACCTCCGACATGAAGATTG	qRT-PCR for AN0554
AN0554-QR	CCTGGTACGAGATCATGTTGTA	qRT-PCR for AN0554
AN1941-QF	AACAACATGGGTATTTTGGCTC	qRT-PCR for AN1941
AN1941-QR	GCCATGTTAGATTGTGTTCTGG	qRT-PCR for AN1941

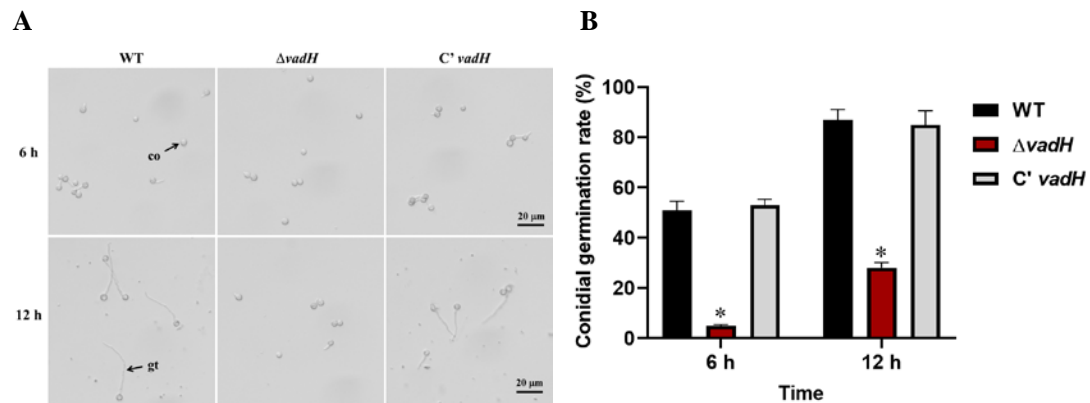
AN10218-QF	TTGCCTACGGTATCAATATGCA	qRT-PCR for AN10218
AN10218-QR	CTCGTTGATTTGCCATCTGTAC	qRT-PCR for AN10218
AN8043-QF	GATACTGTTATACCGCACCTCA	qRT-PCR for AN8043
AN8043-QR	TAATCTTGTGCACATTGCTGAC	qRT-PCR for AN8043
AN10694-QF	GGGGAAATGCAGATTATTGTCTG	qRT-PCR for AN10694
AN10694-QR	GCGAATTTTCAGACTCCCATTT	qRT-PCR for AN10694
AN0773-QF	TCACGCGAATAATCCTTGTTTC	qRT-PCR for AN0773
AN0773-QR	TTTCTTGATGTTTGCCCATAGC	qRT-PCR for AN0773
AN0493-QF	TGGACCATTACTTCCCTTAAGC	qRT-PCR for AN0493
AN0493-QR	TAGGTACAGTCGGTAGAGTCG	qRT-PCR for AN0493
AN3804-QF	TATCATTGACGGAAGGTCGTT	qRT-PCR for AN3804
AN3804-QR	GAAGTAGATGTGGACTCAGTGG	qRT-PCR for AN3804
AN8438-QF	CATTTGCGAGAGACGTTTACAC	qRT-PCR for AN8438
AN8438-QR	CGAGCAAATAAGAAGCCGTATC	qRT-PCR for AN8438
AN1460-QF	GTTTTTGAGCAGAGTGGAGATC	qRT-PCR for AN1460
AN1460-QR	ACTGCACAACTTTTTGATCCAG	qRT-PCR for AN1460
AN10326-QF	GTCTTGATGGCGATGTTCTTAC	qRT-PCR for AN10326
AN10326-QR	GGACAATTCATTGGCGATATCC	qRT-PCR for AN10326
AN2924-QF	GTTCGAGGCTGCTACTACTATC	qRT-PCR for AN2924
AN2924-QR	GAATCATATAGGATTGCGGCAC	qRT-PCR for AN2924
AN10003-QF	CGTACAGACAGCAAATCTGAAG	qRT-PCR for AN10003
AN10003-QR	GATCGAAAATTGTCTCCCCATG	qRT-PCR for AN10003
AN2002-QF	CAGTGCTCTCTTTAGCTATGGT	qRT-PCR for AN2002
AN2002-QR	GATAATCACCATTGCGCAGTAG	qRT-PCR for AN2002
AN2928-QF	GTTTCCACGAACATCATTAGCG	qRT-PCR for AN2928
AN2928-QR	GAAAGAGAGAAGACCGTGAAGA	qRT-PCR for AN2928
AN11378-QF	GATCTGTTGTTCGGGATACTGT	qRT-PCR for AN11378
AN11378-QR	CAACGCACTTATTTCCATAGCA	qRT-PCR for AN11378
AN3804-QF	TATCATTGACGGAAGGTCGTT	qRT-PCR for AN3804
AN3804-QR	GAAGTAGATGTGGACTCAGTGG	qRT-PCR for AN3804
AN9168-QF	TCAAGATGCCTACGGATTATAC	qRT-PCR for AN9168
AN9168-QR	GACCGACTCTCTACCATAAA	qRT-PCR for AN9168
AN11039-QF	GAAAAAGATCCCGGTTACAAGG	qRT-PCR for AN11039
AN11039-QR	GTTGAACTGGAGGTCAGACTTA	qRT-PCR for AN11039
AN9443-QF	CAAGCTTTCAAGATCAAAACGC	qRT-PCR for AN9443
AN9443-QR	TATCCTGAGACTGTGCACAAAT	qRT-PCR for AN9443
AN10385-QF	CTTGATACGTCGTTATTGTCCG	qRT-PCR for AN10385
AN10385-QR	CTTGTGAAAATGACGACCGTAG	qRT-PCR for AN10385
AN7836-QF	AAGACGGATAATTTTGCACAGG	qRT-PCR for AN7836
AN7836-QR	TTGGTCCTCATTAGTCTTCCAG	qRT-PCR for AN7836
AN2242-QF	CTACTGACTGCAAATTCGTGAG	qRT-PCR for AN2242
AN2242-QR	AATGGAGCAGATCGTGGTAATG	qRT-PCR for AN2242

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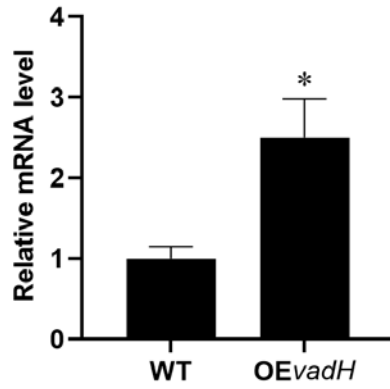
<sup>a</sup>Tail sequences are shown in italics. Restriction enzyme sites are in bold



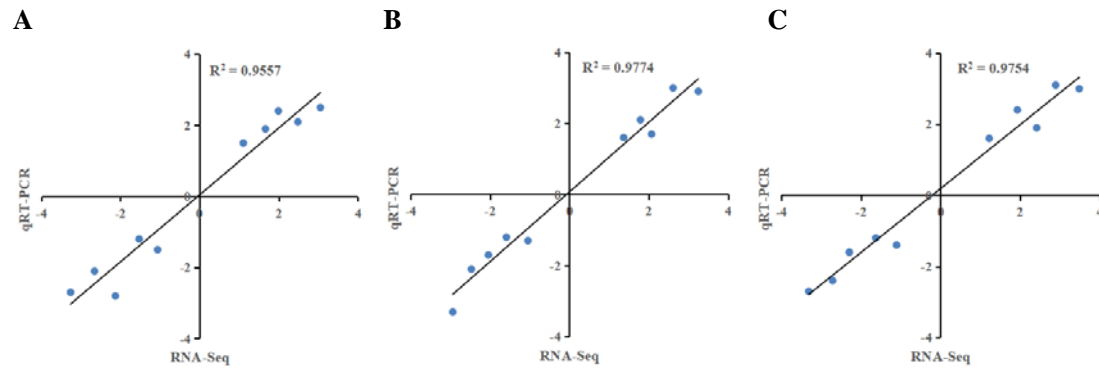
**Figure S1.** Verification of the gene-knockout mutant and complementary strain. (A) The verification strategy of *vadH*. (B) PCR verification results of OXL7 / OXL8. (C) PCR verification results of OXL9 / OXL10; (D) PCR verification results of OXL11 / OXL12. M: DL2000 DNA marker, 1: WT, 2:  $\Delta vadH$ , 3: C' *vadH*. B1: PCR amplification fragment using the primers OXL7 / OXL8; B2: PCR amplification fragment using the primers OXL9 / OXL10; B3: PCR amplification fragment using the primers OXL11 / OXL12.



**Figure S2.** Conidial germination of WT,  $\Delta vadH$  and the complemented strain. (A) Conidial germination at 6 and 12 h. co: conidium, gt: germ tube. (B) Statistical analyses of conidial germination rates. The asterisks represent significant level (\*  $p < 0.05$ ).



**Figure S3.** Verification of the mRNA level of *vadH* in OE*vadH*.



**Figure S4.** The qRT-PCR verification of RNA-Seq data in three stages. Five up-regulated and five downregulated genes were randomly selected and validated. **(A)** The qRT-PCR verification of RNA-Seq data in the vegetative growth stage. The selected DEGs for vegetative grow stage were as follows: AN9219 ( $\log_2$ Fold change in RNA-Seq: 3.05), AN7030 (2.48), AN1443 (1.99), AN1951 (1.66), AN4190 (1.10), AN9473 (-1.06), AN6273 (-1.53), AN0554 (-2.13), AN1941 (-2.66) and AN10218 (-3.26). **(B)** The qRT-PCR verification of RNA-Seq data in the asexual development stage. The selected DEGs for asexual development stage were as follows: AN8043 (3.25), AN10694 (2.61), AN0773 (2.07), AN0493 (1.79), AN3804 (1.37), AN8438 (-1.04), AN1460 (-1.59), AN10326 (-2.04), AN2924 (-2.47) and AN10003 (-2.94). **(C)** The qRT-PCR verification of RNA-Seq data in the sexual development stage. The selected DEGs for sexual development stage were as follows: AN2002 (3.48), AN2928 (2.89), AN11378 (2.42), AN3804 (1.93), AN9168 (1.22), AN11039 (-1.11), AN9443- (-1.63), AN10385 (-2.29), AN7836 (-2.71) and AN2242 (-3.31).