

# Commentary NLRP3 and Gut Microbiota Homeostasis: Progress in Research

Hongming Pan<sup>1,2,†</sup>, Yuting Jian<sup>1,†</sup>, Feijie Wang<sup>3</sup>, Shaokun Yu<sup>1</sup>, Jiannan Guo<sup>1</sup>, Juntao Kan<sup>3,\*</sup> and Wei Guo<sup>1,\*</sup>

- <sup>1</sup> Department of Pharmacology, School of Pharmacy, Fudan University, Shanghai 201203, China
- <sup>2</sup> Department of Gastrointestinal Surgery, Harbin Medical University Cancer Hospital,
- Harbin Medical University, Harbin 150081, China
- <sup>3</sup> Nutrilite Health Institute, Shanghai 201203, China
- \* Correspondence: junot.kan@amway.com (J.K.); guowei@fudan.edu.cn (W.G.); Tel.: +86-21-519-800-43 (W.G.)
- + These authors contributed equally to this work.

**Abstract:** The inflammasome is a platform for inflammatory signaling, and the NLRP3 inflammasome recognizes stimuli in vitro and in vivo, and releases inflammatory cytokines that trigger inflammation and pyroptosis. In the gut, the NLRP3 inflammasome is a key sensor for protecting the body from damage and exogenous pathogens. It plays a fundamental role in maintaining the stability of the gut's immune system. We focus on the role of NLRP3 as a key node in maintaining the homeostasis of gut microbiota which has not been fully highlighted in the past; gut microbiota and innate immunity, as well as the NLRP3 inflammasome, are discussed in this article.

Keywords: NLRP3; microbiota; intestinal mucosal immunity; inflammasome



Citation: Pan, H.; Jian, Y.; Wang, F.; Yu, S.; Guo, J.; Kan, J.; Guo, W. NLRP3 and Gut Microbiota Homeostasis: Progress in Research. *Cells* **2022**, *11*, 3758. https:// doi.org/10.3390/cells11233758

Academic Editors: Pujie Shi, Tiantian Lin, Lin Chen, Xin Yang, Caili Fu, Hyun-Gyun Yuk and Rong Fan

Received: 26 October 2022 Accepted: 16 November 2022 Published: 24 November 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

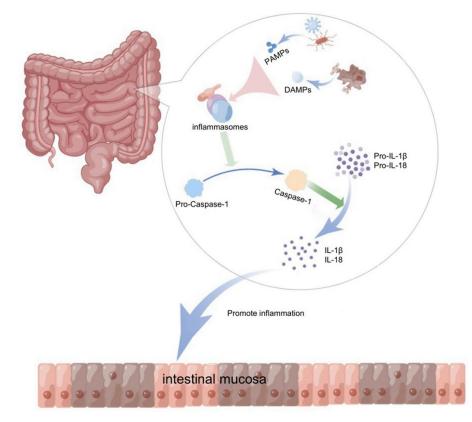
## 1. Introduction

The study of gut microbiota focuses on clarifying the composition of normal gut microbiota and maintaining a stable, normal intestinal flora structure in the host, which is crucial in maintaining the normal intestinal function of the host and resisting pathogenic factors. At present, the main work in the research field of gut microbiota is trying to understand the types and composition ratio of human gut microbiota, and even the distribution characteristics of flora, more accurately and comprehensively in different intestinal segments, find out the differences of gut microbiota among individuals in different health states and of different ethnicities, and explore the influence of interference factors such as antibiotics and dietary habits on intestinal flora composition [1-3]. Based on the current understanding of gut microbiota, people rebuild the host's disturbed gut microbiota through flora transplantation to restore the normal and stable state of the host's gut microbiota and maintain the normal intestinal state of the host [4]. However, the way of reconstruction after disturbance is still passive and relatively lagged. We still need to further clarify the key nodes in the gut microbiota change to find a breakthrough point with application value, so that we can take greater initiative in maintaining the stability of the gut microbiota. Therefore, we focus on the role of NLRP3 as a key node in the intestinal flora change to highlight the great potential of intervention against NLRP3 in the maintenance of the gut microbiota.

### 2. Inflammasome and Innate Immunity

The concept of inflammasomes was first proposed by Jurg Tschopp in 2002, and was composed of three main components: (1) pattern-recognition receptors (PRR) or NOD-like receptors (NLR) or AIM2-like receptors (ALR); (2) apoptosis-associated speck-like proteins containing caspase recruitment domains (ASCs); and (3) caspase proteases [5]. The polyprotein complex with those three parts is appproximately 700 kDa [6]. The innate immune system protects the body from pathogenic microorganisms and harmful substances and is involved in maintaining homeostasis in the gastrointestinal tract. As an important receptor and sensor in the innate immune system, the inflammasome plays

an important role in inflammatory signaling by recognizing pathogenic factors, such as bacteria, fungi, viruses, and products of cell damage, and by initiating inflammatory responses through pro-inflammatory factors [7]. The production of inflammatory factors and cell death mediated by inflammasome activation is closely related to the body's defense against pathogenic microbial infection and the maintenance of homeostasis, making this phenomenon an integral part of innate immunity. Something like pathogen-associated molecular patterns (PAMPs) from invading pathogens, such as lipopolysaccharide (LPS), peptidoglycan, muramyl dipeptide (MDP), bacterial RNA, and danger-associated molecular patterns (DAMPs) from damaged and dying cells in the host, triggered PRR [8]. PRR binds to the adapter ASC and effector pro-caspase-1 to complete NLRP3 inflammasome assembly, then promotes pro-caspase-1 self-cleavage to switch on the production of activated caspase-1, which further triggers IL-1-family cytokines, pro-IL-1 $\beta$  and pro-IL-18, into biologically activated inflammatory factors, IL-1 $\beta$  and IL-18, through proteolysis [9]. Therefore, the inflammasome is regarded as a key factor required for the secretion of IL-1 $\beta$  and IL-18, which play a vital role in innate and adaptive immunity and inflammatory responses in the body [10] (Figure 1).



**Figure 1.** Activated inflammasomes in the gut recognize stimuli and initiate inflammatory responses. Inflammasomes in the intestine recognize the pathogen-associated molecular patterns (PAMPS) of exogenous pathogens invading the host, or the damaged and dying cells release the dangerous-associated molecular patterns (DAMPs) in the body, which will stimulate ASC to assemble with pro-caspase-1 to form inflammasomes. The assembled inflammasomes promote pro-caspase-1 to self-cleavage and produce activated caspase-1, and the activated caspase-1 will convert pro-IL-1  $\beta$  and pro-IL-18 to IL-1  $\beta$  and IL-18, which in turn triggers an inflammatory response in the intestine (diagram by Figdraw).

#### 3. Gut Immunity and Gut Microbiota

The intestinal tract is not only the largest digestive organ but also an essential immune organ. Although the intestinal epithelium is directly challenged by microorganisms since it is directly exposed to stimuli from the external environment, the human intestine is at the

forefront of the immune system, with over 70% of all immune cells settled here. Human intestinal mucosa covers approximately 300 m<sup>2</sup> and is the main site of contact between the body and pathogens [11]. The human gut is colonized by a large number and species of gut microbiota, which form a dynamic and mutually beneficial symbiotic relationship with the human body and play a critical role in gut immunity [12]. Therefore, together with experimental epithelial cells (IEC), intraepithelial lymphocytes (IEL), lamina propria lymphocytes (LPL), and Peyer's patches (PP), the gut microbiota account for one of the primary components of the intestinal immune system [7,8]. Many microorganisms exist in the gut microbiota, including bacteria, archaea, fungi, protozoa, and viruses [13]. Among all, intestinal bacteria, with a number of approximately 10 [14], form a complex but relatively stable huge system. Based on microbial culturomics research, these bacteria are mainly divided into Firmicutes, Bacteroidetes, Actinobacteria, Proteobacteria, and Clostridium, of which Bacteroidetes and Firmicutes together account for approximately 90% of the total gut microbiota (25% and 65% respectively) [14]. However, intestinal environments, as well as gut microbiota, are intestinal segment and individual dependent. For example, gut microbiota is influenced by diet and genetics, which is most obvious in the composition of Bacteroidetes and Proteobacteria [15].

#### 3.1. Gut Microbiota Is Essential for Gut Immunity

Gut microbiota not only contributes to nutrient absorption and metabolism in the body but also competes with bacterial and fungal pathogens for nutrients and attachment sites and resists their colonization of the intestinal mucosa by producing antibacterial substances and absorbing foreign pathogenic factors [16]. Therefore, gut microbiota enables the functional gastrointestinal immune system; it is even possible to view the gut microbiome as an organ system that is important to the development of children and plays a key role in child development [17]. In the first five years of life, the intestinal flora will affect the development of the infant's immune system and preschool reasoning ability. During this period, the healthy development of the infant's intestinal flora will help the infant to build a perfect immune system and cognitive function [18,19]. In normal people, the gut microbiota plays an essential role in developing and functioning immune organs, such as the small intestine and intestinal lymph nodes in the ileum [17]. In recent research, the changes in the digestive tract of the body before and after gut microbiota transplantation were compared; the results confirmed that the recurrence of recurrent *Clostridioides difficile* infection (rCDI) was effectively curbed after fecal microbiota transplant (FMT) [20]. In the treatment of rCDI infection, FMT affects the life cycle of C. difficile by restoring the interaction between gut microbiota and metabolites, especially short-chain fatty acids (SCFA) and bile acids.

Up to now, there are sufficient clinical research data to support that FMT has become an effective treatment for *C. difficile* infection [21]. Ekekezie Chiazotam's survey shows that the overall efficacy rate of microbiota transplantation in the treatment and prevention of *C*. *difficile* infections is between 80% and 90%, with no serious adverse reactions during the treatment [22]. Gut microbiota is also indispensable for developing an optimal immune system. The gut microbiota colonized in the gastrointestinal tract can help the host establish a strong intestinal mucosal protective layer to improve the resistance of the host's digestive tract to colonization and foreign pathogen infection, so the development of intestinal microbiota has an important impact on the development of human immune system [23]. The transplantation experiment of intestinal microbiota in animal models has confirmed that intestinal microbiota can help the development of the host's innate and adaptive immune system. It also demonstrated that the underdeveloped host's intestinal immune system can be corrected by transplanting specific intestinal symbiotic bacteria. The rapid changes in intestinal microbiota will affect the development of innate immune cells and adaptive immune cells, thus affecting the homeostasis and immune system functions of the host's intestine [24,25].

#### 3.2. The Gut Microbiota Disorder Is Associated with the Pathogenesis of Various Diseases

A large number of studies have shown that gut microbiota disorder is closely related to the occurrence of a variety of diseases, such as obesity, diabetes, cardiovascular and cerebrovascular diseases, inflammatory bowel disease (IBD), irritable bowel syndrome (IBS), autoimmune diseases, allergies, autistic disorders, depression, and senile dementia [26–28]. The results of fecal community analysis showed that the bacterial species diversity of fecal communities in patients with recurrent *C. difficile* infection was significantly reduced [29]. Disruption of the gut microbiota at the colonic sites and subsequent exposure to *C. difficile* are the first steps in disease pathogenesis. Changes in gut microbiota increase the content of certain free amino acids, especially proline, an essential growth factor of *C. difficile*, closely related to the pathogenesis of diseases [30,31]. Except for providing the energy to support the growth of *C. difficile*, increased proline also affects the production of Clostridium difficile toxin A and toxin B, which are the leading causes of hospital-acquired diarrhea in adults and can even lead to death in patients with severe symptoms [30,32,33]. Changes in the composition of the gut microbiota will affect the metabolic balance in the body and, therefore, involve the pathogenesis of the cardiovascular disease [34]. Metabolomics studies and germ-free mouse experiments have demonstrated that gut microbiota plays a key role in trimethylamine N-oxide (TMAO) production. Phosphatidylcholine from eggs, milk, red meat, and other food in the daily diet is metabolized in the gut to trimethylamine by gut microbiota, and then to TMAO by liver-derived flavin-containing monooxygenase. TMAO promotes the upregulation of multiple macrophage scavenger receptors associated with atherosclerosis. Therefore, increased TMAO intake can accelerate the development of atherosclerosis, enhanced macrophage cholesterol accumulation, and foam cell formation [35]. As mentioned above, gut microbiota can not only regulate the mucosal immune function of the intestinal tract but also affect the occurrence and development of cardiovascular diseases by affecting the body's nutrient metabolism.

#### 4. The Relationship between the NLRP3 Inflammasome and Intestinal Homeostasis

#### 4.1. NLRP3 Inflammasome Plays a Key Role in Intestinal Mucosal Immunity

As the natural cavity of the human body, the intestinal tract is frequently in contact with the external environment and foreign substances, thus serving as the primary way for many bacterial and viral pathogens to invade the human body. Stimulating molecules and pathogenic proteins contained in food and foreign microorganisms are the main inducers of the gut immune response; the intestinal defense system composed of intestinal microbiota, the intestinal epithelial barrier, and the local immune system can resist the stimulation of these stimulators or pathogens to the body, to maintain steady and intestinal health, despite the complex external environment [36]. As pathogen sensors, pattern-recognition receptors, such as the Toll-like receptor (TLR), NOD-like receptor (NLR), and RIG-I-like receptor (RLR), distributed in the intestinal epithelium can recognize the pathogen-associated molecular pattern (PAMP) ligand. Subsequently, downstream signaling pathways and molecular events are active. They induce the expression of anti-infective cytokines and other intestinal mucosal immune defense molecules to promote the occurrence of mucosal immune responses, which are crucial for maintaining gastrointestinal homeostasis [37]. Intestinal intraepithelial lymphocytes (IELs) and lamina propria T lymphocytes are the main immune cells of the gut, while dendritic cells are one of the professional antigen-presenting cells (APCs). As one of the most important members of the inflammasome family, the NLRP3 inflammasome is widely present in epithelial cells and immune cells. The molecular structure of NLRP3 contains a central nucleotide-binding and oligomerization (NACHT) domain which has ATPase activity, while the C-terminus of NLRP3 retains a leucine-rich repeat (LRR) domain to regulate the biological activity of NLRP3 and detects endogenous alarm proteins or microbial-derived ligands. When the NLRP3 inflammasome is activated by pathogens, such as bacterial toxins, ATP, reactive oxygen species (ROS), in the gut and danger signal molecules in the body, its downstream caspase-1 effector proteins activate inflammatory factors such as IL-1 $\beta$  and IL-18, thereby triggering an inflammatory response

5 of 9

in the gut. Hereby, NLRP3 plays a crucial role in the body's anti-infection immunity and the occurrence of inflammatory diseases such as IBD, IBS, and other immune-related intestinal diseases [38]. Accordingly, the role of NLRP3 inflammasome in intestinal mucosal immunity has received more and more attention and is expected to be a new target for drug discovery and development [39].

### 4.2. Downstream Effector IL-1

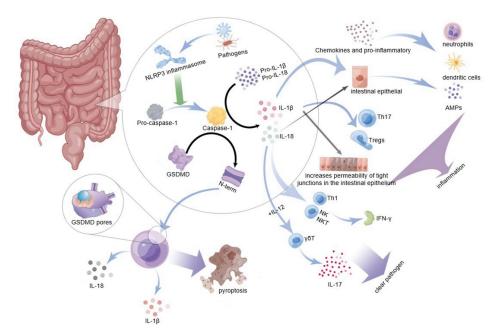
IL-1 is a multifunctional cytokine, which is activated by the initiation of the NLRP3 inflammasome. IL-1 is dominant in innate immunity, with strong pro-inflammatory activity, and is one of the most critical medium factors in inflammation and host anti-infection response. IL-1 can induce the expression of proinflammatory cytokines or chemokines such as IL-6, IL-8, and CCL2 in inflammatory sites. The changes in the expression of these inflammatory cytokines or chemokines recruit monocytes and neutrophils to migrate to infection, injury, and necrosis sites to engulf pathogens, inducing local inflammatory response, and even promoting the activation of dendritic cells and neutrophils; this series of chain reactions triggered by IL-1 will eventually help the host to resist the invasion of pathogens and maintain the homeostasis in the host's intestine [40–42]. In addition, Judy Lieberman's research shows that NLRP3 can affect lamina propria mononuclear cells to secrete more IL-1 $\beta$ . It can help the intestinal epithelium to produce antibacterial peptides and enhance the activation of Treg cells in the host body's immune response. The activated Tregs can boost the controllability and stability of intestinal inflammatory activities to keep the stability of intestinal links; IL-1 also co-mediates the differentiation of CD4 + T cells into Th17 cells with other cytokines [9,43]. The role of IL-1 in intestinal inflammation has been reported in several studies; as a typical pro-inflammatory cytokine IL-1 $\beta$  occupied a key position in inducing the increase in the epithelial tight junction permeability in the intestine, which plays an important role in amplifying the cascade of intestinal inflammation. A defective intestinal epithelial tight junction (TJ) barrier is recognized as an important contributor to intestinal inflammatory diseases; blocking IL-1 can improve the status of patients with Clostridium- and Salmonella-induced colitis. These findings suggest that IL-1 is a key pro-inflammatory cytokine in IBD-like diseases which plays a core role in promoting inflammatory damage [44,45].

#### 4.3. Downstream Effector IL-18

IL-18 not only activates monocytes but also stimulates the secretion of IFN- $\gamma$  by Th1 cells, natural killer (NK) cells, and natural killer T cells; it also promotes the production of  $\gamma\delta T$  cells which then secrete IL-17 when IL-18 is combined with IL-12, and this series of chain reactions activated by IL-18 helps the human body eliminate infectious microorganisms [46]. Moreover, IL-18 helps the body defend against bacterial and viral invasion and maintains intestinal microbiome homeostasis by activating intestinal epithelial cells to produce antimicrobial peptides. IL-18-activated  $\gamma\delta T$  cells play an important role in intestinal mucosal immunity by maintaining intestinal epithelial homeostasis. Evidence shows that the combination of IL-18 and IL-2 activates and induces the proliferation of human CD56 + CD11c + precursor NK cells and consequently triggers strong proliferation and activation of  $\gamma \delta T$  cells in the gastrointestinal inflamed mucosa of patients with ulcerative colitis [47]. By activating  $\gamma\delta T$  cells, IL-18 also drives the production of IL-17. It is crucial for resistance against pathogen invasion and maintenance of the steady state of the intestinal tract by recruiting neutrophils to the site of pathogen infection or inflammation [48,49]. In general, intestinal epithelial cells without intestinal infection and inflammation under physiological conditions are the main source of IL-18; IL-18 released by intestinal epithelial cells has the function of promoting intestinal epithelial repair, renewal, and maturation. Therefore, it helps maintain the normal function of the intestinal epithelial barrier against intestinal infection and inflammation. More IL-18 is produced downstream of the activated inflammasome to exert its biological functions when gastrointestinal infection and intestinal inflammation occur [50].

#### 4.4. NLRP3 Inflammasome and Gasdermin D

A recent study reported that caspase-1, activated by the NLRP3 inflammasome, can cleave and activate gasdermin D protein (GSDMD), which is a key mediator of innate immune defense. The study from Feng Shao's group confirmed that GSDMD is a direct-acting molecule of inflammation-induced pyroptosis, leading to inflammatory form cell death or pyroptosis [51]. This new cell death mode which is known as "inflammatory programmed cell death" has confirmed that inflammasomes play a role in promoting the host to resist the invasion of microbial pathogens and producing aseptic inflammation [52]. The molecular structure of gasdermins contains a cytotoxic N-terminal domain and a C-terminal repressor domain connected by a flexible linker. After being cleaved by caspase, the N-terminal domain fragment of GSDMD is released and oligomerized to form a bioactive p30 fragment with a molecular weight of 32 kD; p30 then binds to the cell membrane to form many honeycomb-like pores with an inner diameter of 10–14 nm, which further promotes the release of mature IL-1  $\beta$  and IL-18 [53,54]. IL-1  $\beta$  and IL-18 then activate an inflammatory response in the host's body. Furthermore, membrane pores formed by GsdmD p30 allow the inflow and outflow of various ions, proteins, and water, which cause cell osmotic pressure imbalance and cell swelling, eventually leading to inflammatory cell death or pyroptosis [55]. This phenomenon has been observed in a variety of physiological and pathophysiological conditions, from viral and bacterial infections to inflammation-related diseases, as shown in Figure 2.



**Figure 2.** NLRP3 inflammasome in the intestine recognizes bacteria, viruses, or other pathogens and promotes the activation of pro-cassase-1 to caspase-1. On the one hand, caspase-1 will cleave GSDMD to produce an active segment N-terminal, which can bind to the cell membrane to form many 10–14 nm holes to cause pyroptosis and release IL-1  $\beta$  and IL-18 to promote inflammatory reaction. On the other hand, activated caspase-1 activates pro-IL-1  $\beta$  and pro-IL-18, activated IL-1  $\beta$ , further inducing the expression of inflammatory cytokines and chemokines in the inflammatory part; it can promote the recruitment and activation of neutrophils and dendritic cells in the inflammatory part, and promote the differentiation of Th17 and Tregs. IL-18 can activate intestinal epithelial cells to produce AMPs, stimulate the production of Th1, NK, and NKT to secrete IFN-  $\gamma$ , or combine with IL-12 to induce more  $\gamma\delta$  T cells which secrete IL-17. NLRP3 inflammasome promotes the local inflammatory response of the intestinal and helps remove the pathogenic factors invading the body to maintain intestinal homeostasis through the above way (diagram by Figdraw).

Pyroptosis induced by the activation of immune response after the recognition of risk factors by inflammasomes destroys the cells infected by pathogens; the replication of pathogens in the cells is inhibited in this way to avoid the further spread of pathogens in the intestine [56]. At the same time, IL-1-family cytokines, IL-1 $\beta$  and IL-18, that communicate with other cells will also be released to further activate the inflammatory activity and immune response against related pathogens, thus stimulating the immune system to eliminate pathogenic infection factors and helping maintain the stability of the gastrointestinal tract [57]. Although pyroptosis protects the body from pathogenic infection by antagonizing infection or endogenous dangers, hyperactivation of pyroptosis can lead to fatal sepsis [58]. Loss of GSDMD impairs the body's ability to clear the intestinal bacterial infection and enteroviral infection. However, the inflammasome has a complex mode of action in the intestine. Its detailed mechanism of action in intestinal immunity still needs more in-depth research.

#### 5. Conclusions

After a long co-evolution with the host, the gut microbiota formed a complex but effective intestinal immune system, helping the host resist the invasion of pathogens. Many studies have confirmed that the inflammasome plays a key role in intestinal immunity, but the detailed mechanism of the inflammasome in the intestinal immune system still needs more in-depth research. The discovery of the gasdermin family provides new directions and clues for the study of inflammasomes in intestinal immunity. However, its application as a potential therapeutic target for practical clinical therapeutic applications still needs further exploration [59]. The research on intestinal mucosal immunity has attracted increasing attention; as an important part of the body's immune defense line, intestinal mucosal immunity is indispensable for the maintenance of the body's homeostasis. Therefore, studying the mechanism and pathways of the intestinal immune system is of great significance for related diseases such as inflammatory bowel disease. The existing research shows that NLRP3 plays a key role in intestinal flora homeostasis and intestinal mucosal immunity. As a signaling molecule in the intestinal inflammatory response, NLRP3 provides new ideas to understand and improve the therapeutic effect of IBD. With the deepening of research work, NLRP3 may become a therapeutic target for intervening intestinal diseases such as inflammatory bowel disease and intestinal infection, but more clinical studies are needed before practical application.

**Author Contributions:** Writing—original draft preparation, H.P. and Y.J.; writing—review and editing, F.W., J.K. and W.G.; Gathered the references, S.Y., J.G. and J.K.; All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by Special Funds for People's Livelihood Research Medical and Health Project of Shanghai, grant number PKJ2019-Y22.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Acknowledgments: We thank Jun Du at Nutrilite Health Institute for providing technical support on this review.

**Conflicts of Interest:** The authors declare no conflict of interest.

#### References

- 1. Yuan, X.; Chang, C.; Chen, X.; Li, K. Emerging trends and focus of human gastrointestinal microbiome research from 2010–2021: A visualized study. *J. Transl. Med.* **2021**, *19*, 327. [CrossRef] [PubMed]
- Zhang, X.; Zhong, H.; Li, Y.; Shi, Z.; Ren, H.; Zhang, Z.; Zhou, X.; Tang, S.; Han, X.; Lin, Y.; et al. Sex- and age-related trajectories of the adult human gut microbiota shared across populations of different ethnicities. *Nat. Aging* 2021, 1, 87–100. [CrossRef]
- Zhang, G.; Wang, Q.; Tao, W.; Jiang, W.; Elinav, E.; Wang, Y.; Zhu, S. Glucosylated nanoparticles for the oral delivery of antibiotics to the proximal small intestine protect mice from gut dysbiosis. *Nat. Biomed. Eng.* 2022, *6*, 867–881. [CrossRef] [PubMed]

- El-Salhy, M.; Winkel, R.; Casen, C.; Hausken, T.; Gilja, O.H.; Hatlebakk, J.G. Efficacy of Fecal Microbiota Transplantation for Patients with Irritable Bowel Syndrome at 3 Years After Transplantation. *Gastroenterology* 2022, 163, 982–994. [CrossRef]
- Martinon, F.; Burns, K.; Tschopp, J. The inflammasome: A molecular platform triggering activation of inflammatory caspases and processing of proIL-beta. *Mol. Cell* 2002, 10, 417–426. [CrossRef]
- 6. Atianand, M.K.; Rathinam, V.A.; Fitzgerald, K.A. SnapShot: Inflammasomes. Cell 2013, 153, 272. [CrossRef]
- Zahid, A.; Li, B.; Kombe, A.J.K.; Jin, T.; Tao, J. Pharmacological Inhibitors of the NLRP3 Inflammasome. *Front. Immunol.* 2019, 10, 2538. [CrossRef]
- 8. Broz, P.; Dixit, V.M. Inflammasomes: Mechanism of assembly, regulation and signalling. *Nat. Rev. Immunol.* **2016**, *16*, 407–420. [CrossRef]
- 9. Van de Veerdonk, F.L.; Netea, M.G.; Dinarello, C.A.; Joosten, L.A.B. Inflammasome activation and IL-1β and IL-18 processing during infection. *Trends Immunol.* **2011**, *32*, 110–116. [CrossRef]
- 10. Fields, J.K.; Günther, S.; Sundberg, E.J. Structural Basis of IL-1 Family Cytokine Signaling. *Front. Immunol.* **2019**, *10*, 1412. [CrossRef]
- 11. Hooper, L.V. Epithelial cell contributions to intestinal immunity. Adv. Immunol. 2015, 126, 129–172.
- 12. Koboziev, I.; Reinoso Webb, C.; Furr, K.L.; Grisham, M.B. Role of the enteric microbiota in intestinal homeostasis and inflammation. *Free Radic. Biol. Med.* **2013**, *68*, 122–133. [CrossRef]
- 13. Gérard, P. Gut microbiota and obesity. Cell. Mol. Life Sci. 2016, 73, 147–162. [CrossRef]
- 14. Lagier, J.-C.; Khelaifia, S.; Alou, M.T.; Ndongo, S.; Dione, N.; Hugon, P.; Caputo, A.; Cadoret, F.; Traore, S.I.; Seck, E.H.; et al. Culture of previously uncultured members of the human gut microbiota by culturomics. *Nat. Microbiol.* **2016**, *1*, 179. [CrossRef]
- 15. Costea, P.I.; Hildebrand, F.; Arumugam, M.; Bäckhed, F.; Blaser, M.J.; Bushman, F.D.; de Vos, W.M.; Ehrlich, S.D.; Fraser, C.M.; Hattori, M.; et al. Enterotypes in the landscape of gut microbial community composition. *Nat. Microbiol.* **2018**, *3*, 8–16. [CrossRef]
- 16. Cheng, H.-Y.; Ning, M.-X.; Chen, D.-K.; Ma, W.-T. Interactions Between the Gut Microbiota and the Host Innate Immune Response Against Pathogens. *Front. Immunol.* **2019**, *10*, 607. [CrossRef]
- Ronan, V.; Yeasin, R.; Claud, E.C. Childhood Development and the Microbiome-The Intestinal Microbiota in Maintenance of Health and Development of Disease During Childhood Development. *Gastroenterology* 2020, 160, 495–506. [CrossRef] [PubMed]
- Kaczmarczyk, M.; Löber, U.; Adamek, K.; Węgrzyn, D.; Skonieczna Żydecka, K.; Malinowski, D.; Łoniewski, I.; Markó, L.; Ulas, T.; Forslund, S.K.; et al. The gut microbiota is associated with the small intestinal paracellular permeability and the development of the immune system in healthy children during the first two years of life. *J. Transl. Med.* 2021, 19, 177. [CrossRef] [PubMed]
- Guzzardi, M.A.; Ederveen, T.H.A.; Rizzo, F.; Weisz, A.; Collado, M.C.; Muratori, F.; Gross, G.; Alkema, W.; Iozzo, P. Maternal pre-pregnancy overweight and neonatal gut bacterial colonization are associated with cognitive development and gut microbiota composition in pre-school-age offspring. *Brain Behav. Immun.* 2021, 100, 311–320. [CrossRef] [PubMed]
- Martinez-Gili, L.; McDonald, J.A.K.; Liu, Z.; Kao, D.; Allegretti, J.R.; Monaghan, T.M.; Barker, G.F.; Miguéns Blanco, J.; Williams, H.R.T.; Holmes, E.; et al. Understanding the mechanisms of efficacy of fecal microbiota transplant in treating recurrent Clostridioides difficile infection and beyond: The contribution of gut microbial-derived metabolites. *Gut Microbes* 2020, *12*, 1810531. [CrossRef]
- Ekekezie, C.; Perler, B.K.; Wexler, A.; Duff, C.; Lillis, C.J.; Kelly, C.R. Understanding the Scope of Do-It-Yourself Fecal Microbiota Transplant. Am. J. Gastroenterol. 2020, 115, 603–607. [CrossRef] [PubMed]
- Allegretti, J.R.; Mullish, B.H.; Kelly, C.; Fischer, M. The evolution of the use of faecal microbiota transplantation and emerging therapeutic indications. *Lancet* 2019, 394, 420–431. [CrossRef] [PubMed]
- Wang, X.; Zhang, P.; Zhang, X. Probiotics Regulate Gut Microbiota: An Effective Method to Improve Immunity. *Molecules* 2021, 26, 6076. [CrossRef]
- Xiang, Q.; Wu, X.; Pan, Y.; Wang, L.; Cui, C.; Guo, Y.; Zhu, L.; Peng, J.; Wei, H. Early-Life Intervention Using Fecal Microbiota Combined with Probiotics Promotes Gut Microbiota Maturation, Regulates Immune System Development, and Alleviates Weaning Stress in Piglets. *Int. J. Mol. Sci.* 2020, 21, 503. [CrossRef]
- 25. Xue, J.; Ajuwo, K.M.; Fang, R. Mechanistic insight into the gut microbiome and its interaction with host immunity and inflammation. *Anim. Nutr.* **2020**, *6*, 421–428.
- 26. Shen, J.; Obin, M.S.; Zhao, L. The gut microbiota, obesity and insulin resistance. Mol. Asp. Med. 2012, 34, 39–58. [CrossRef]
- Radhakrishnan, S.T.; Alexander, J.L.; Mullish, B.H.; Gallagher, K.I.; Powell, N.; Hicks, L.C.; Hart, A.L.; Li, J.V.; Marchesi, J.R.; Williams, H.R.T. Systematic review: The association between the gut microbiota and medical therapies in inflammatory bowel disease. *Aliment. Pharm. Ther.* 2021, 55, 26–48. [CrossRef]
- 28. Li, Y.; Luo, Z.-Y.; Hu, Y.-Y.; Bi, Y.-W.; Yang, J.-M.; Zou, W.-J.; Song, Y.-L.; Li, S.; Shen, T.; Li, S.-J.; et al. The gut microbiota regulates autism-like behavior by mediating vitamin B6 homeostasis in EphB6-deficient mice. *Microbiome* **2020**, *8*, 120. [CrossRef]
- Chang, J.Y.; Antonopoulos, D.A.; Kalra, A.; Tonelli, A.; Khalife, W.T.; Schmidt, T.M.; Young, V.B. Decreased diversity of the fecal Microbiome in recurrent Clostridium difficile-associated diarrhea. *J. Infect. Dis.* 2008, 197, 435–438. [CrossRef] [PubMed]
- 30. Poutanen, S.M.; Simor, A.E. Clostridium difficile-associated diarrhea in adults. CMAJ 2004, 171, 51–58. [CrossRef] [PubMed]
- Rodríguez Carrio, J.; López, P.; Sánchez, B.; González, S.; Gueimonde, M.; Margolles, A.; de Los Reyes-Gavilán, C.G.; Suárez, A. Intestinal Dysbiosis Is Associated with Altered Short-Chain Fatty Acids and Serum-Free Fatty Acids in Systemic Lupus Erythematosus. *Front. Immunol.* 2017, *8*, 23. [CrossRef]

- Reed, A.D.; Fletcher, J.R.; Huang, Y.Y.; Thanissery, R.; Rivera, A.J.; Parsons, R.J.; Stewart, A.K.; Kountz, D.J.; Shen, A.; Balskus, E.P.; et al. The Stickland Reaction Precursor trans-4-Hydroxy-l-Proline Differentially Impacts the Metabolism of Clostridioides difficile and Commensal Clostridia. *mSphere* 2022, 7, e0092621. [CrossRef] [PubMed]
- Bouillaut, L.; Self, W.T.; Sonenshein, A.L. Proline-dependent regulation of Clostridium difficile Stickland metabolism. *J. Bacteriol.* 2012, 195, 844–854. [CrossRef] [PubMed]
- 34. Xu, H.; Wang, X.; Feng, W.; Liu, Q.; Zhou, S.; Liu, Q.; Cai, L. The gut microbiota and its interactions with cardiovascular disease. *Microb. Biotechnol.* **2020**, *13*, 637–656. [CrossRef] [PubMed]
- 35. Wang, Z.; Klipfell, E.; Bennett, B.J.; Koeth, R.; Levison, B.S.; Dugar, B.; Feldstein, A.E.; Britt, E.B.; Fu, X.; Chung, Y.-M.; et al. Gut flora metabolism of phosphatidylcholine promotes cardiovascular disease. *Nature* **2011**, *472*, 57–63. [CrossRef]
- 36. Haq, S.; Grondin, J.; Banskota, S.; Khan, W.I. Autophagy: Roles in intestinal mucosal homeostasis and inflammation. *J. Biomed. Sci.* **2019**, *26*, 19. [CrossRef]
- Kurashima, Y.; Kiyono, H. Mucosal Ecological Network of Epithelium and Immune Cells for Gut Homeostasis and Tissue Healing. Annu. Rev Immunol. 2017, 35, 119–147. [CrossRef]
- 38. Zhao, C.; Zhao, W. NLRP3 Inflammasome—A Key Player in Antiviral Responses. Front. Immunol. 2020, 11, 211. [CrossRef]
- Song, Y.; Zhao, Y.; Ma, Y.; Wang, Z.; Rong, L.; Wang, B.; Zhang, N. Biological functions of NLRP3 inflammasome: A therapeutic target in inflammatory bowel disease. *Cytokine Growth Factor Rev.* 2021, 60, 61–75. [CrossRef]
- Zambetti, L.P.; Mortellaro, A. NLRPs, microbiota, and gut homeostasis: Unravelling the connection. J. Pathol. 2014, 233, 321–330. [CrossRef]
- 41. Guarda, G.; So, A. Regulation of inflammasome activity. Immunology 2010, 130, 329–336. [CrossRef] [PubMed]
- 42. Paget, C.; Doz-Deblauwe, E.; Winter, N.; Briard, B. Specific NLRP3 Inflammasome Assembling and Regulation in Neutrophils: Relevance in Inflammatory and Infectious Diseases. *Cells* **2022**, *11*, 1188. [CrossRef] [PubMed]
- 43. Liu, X.; Zhang, Z.; Ruan, J.; Pan, Y.; Magupalli, V.G.; Wu, H.; Lieberman, J. Inflammasome-activated gasdermin D causes pyroptosis by forming membrane pores. *Nature* **2016**, *535*, 153–158. [CrossRef] [PubMed]
- Rawat, M.; Nighot, M.; AlSadi, R.; Gupta, Y.; Viszwapriya, D.; Yochum, G.; Koltun, W.; Ma, T.Y. IL1B Increases Intestinal Tight Junction Permeability by Up-regulation of MIR200C-3p, Which Degrades Occludin mRNA. *Gastroenterology* 2020, 159, 1375–1389. [CrossRef]
- 45. Afonina, I.S.; Müller, C.; Martin, S.J.; Beyaert, R. Proteolytic Processing of Interleukin-1 Family Cytokines: Variations on a Common Theme. *Immunity* **2015**, *42*, 991–1004. [CrossRef]
- Papadakis, K.A.; Zhu, D.; Prehn, J.L.; Landers, C.; Avanesyan, A.; Lafkas, G.; Targan, S.R. Dominant role for TL1A/DR3 pathway in IL-12 plus IL-18-induced IFN-gamma production by peripheral blood and mucosal CCR9+ T lymphocytes. *J. Immunol.* 2005, 174, 4985–4990. [CrossRef] [PubMed]
- Sun, X.; Cai, Y.; Fleming, C.; Tong, Z.; Wang, Z.; Ding, C.; Qu, M.; Zhang, H.-G.; Suo, J.; Yan, J. Innate γδT17 cells play a protective role in DSS-induced colitis via recruitment of Gr-1+CD11b+ myeloid suppressor cells. *Oncoimmunology* 2017, *6*, e1313369. [CrossRef]
- Lalor, S.J.; Dungan, L.S.; Sutton, C.E.; Basdeo, S.A.; Fletcher, J.M.; Mills, K.H.G. Caspase-1-processed cytokines IL-1beta and IL-18 promote IL-17 production by gammadelta and CD4 T cells that mediate autoimmunity. *J. Immunol.* 2011, 186, 5738–5748. [CrossRef]
- Martin, B.; Hirota, K.; Cua, D.J.; Stockinger, B.; Veldhoen, M. Interleukin-17-producing gammadelta T cells selectively expand in response to pathogen products and environmental signals. *Immunity* 2009, *31*, 321–330. [CrossRef]
- 50. Chiang, H.-Y.; Lu, H.-H.; Sudhakar, J.N.; Chen, Y.-W.; Shih, N.-S.; Weng, Y.-T.; Shui, J.-W. IL-22 initiates an IL-18-dependent epithelial response circuit to enforce intestinal host defence. *Nat. Commun.* **2022**, *13*, 874. [CrossRef]
- Ding, J.; Wang, K.; Liu, W.; She, Y.; Sun, Q.; Shi, J.; Sun, H.; Wang, D.-C.; Shao, F. Pore-forming activity and structural autoinhibition of the gasdermin family. *Nature* 2016, 535, 111–116. [CrossRef] [PubMed]
- 52. Shi, J.; Zhao, Y.; Wang, K.; Shi, X.; Wang, Y.; Huang, H.; Zhuang, Y.; Cai, T.; Wang, F.; Shao, F. Cleavage of GSDMD by inflammatory caspases determines pyroptotic cell death. *Nature* 2015, 526, 660–665. [CrossRef] [PubMed]
- Aglietti, R.A.; Estevez, A.; Gupta, A.; Ramirez, M.G.; Liu, P.S.; Kayagaki, N.; Ciferri, C.; Dixit, V.M.; Dueber, E.C. GsdmD p30 elicited by caspase-11 during pyroptosis forms pores in membranes. *Proc. Natl. Acad. Sci. USA* 2016, 113, 7858–7863. [CrossRef] [PubMed]
- 54. Yao, X.; Zhang, C.; Xing, Y.; Xue, G.; Zhang, Q.; Pan, F.; Wu, G.; Hu, Y.; Guo, Q.; Lu, A.; et al. Remodeling of the gut microbiota by hyperactive NLRP3 induces regulatory T cells to maintain homeostasis. *Nat. Commun.* **2017**, *8*, 1896. [CrossRef] [PubMed]
- 55. Kolbrink, B.; Riebeling, T.; Kunzendorf, U.; Krautwald, S. Plasma Membrane Pores Drive Inflammatory Cell Death. *Front. Cell Dev. Biol.* 2020, *8*, 817. [CrossRef]
- 56. Xia, S. Biological mechanisms and therapeutic relevance of the gasdermin family. Mol. Aspects Med. 2020, 76, 100890. [CrossRef]
- 57. Lu, F.; Lan, Z.; Xin, Z.; He, C.; Guo, Z.; Xia, X.; Hu, T. Emerging insights into molecular mechanisms underlying pyroptosis and functions of inflammasomes in diseases. *J. Cell. Physiol.* **2019**, *235*, 3207–3221. [CrossRef]
- Chen, Y.; Luo, R.; Li, J.; Wang, S.; Ding, J.; Zhao, K.; Lu, B.; Zhou, W. Intrinsic Radical Species Scavenging Activities of Tea Polyphenols Nanoparticles Block Pyroptosis in Endotoxin-Induced Sepsis. ACS Nano 2022, 16, 2429–2441. [CrossRef] [PubMed]
- 59. Xia, X.; Wang, X.; Zheng, Y.; Jiang, J.; Hu, J. What role does pyroptosis play in microbial infection? *J. Cell. Physiol.* 2018, 234, 7885–7892. [CrossRef]