

Autologous iPSC-Derived Human Neuromuscular Junction to Model the Pathophysiology of Hereditary Spastic Paraplegia

SUPPLEMENTARY FIGURE TITLES AND LEGENDS

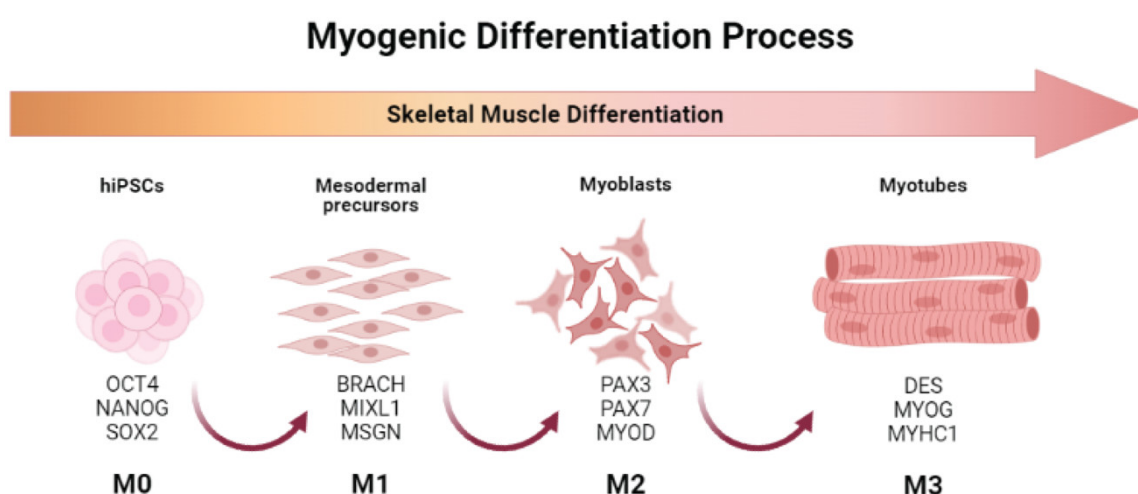


Figure S1. Myogenic differentiation process, related to figure 3. Schematic overview of the myogenic differentiation protocol for HSP and CTRL hiPSC lines towards myotubes. hiPSCs were kept pluripotent in M0 (E8 medium). Three different media were used to induce myogenic differentiation at three different steps: M1 to differentiate hiPSCs to the stage of mesodermal precursors, M2 to induce the myoblast stage, M3 to promote fusion of the myoblasts into myotubes. The qRT-PCR genes for checking the progression of the differentiation have been indicated. The protocol was adapted from Caron and colleagues (Caron et al., 2016).

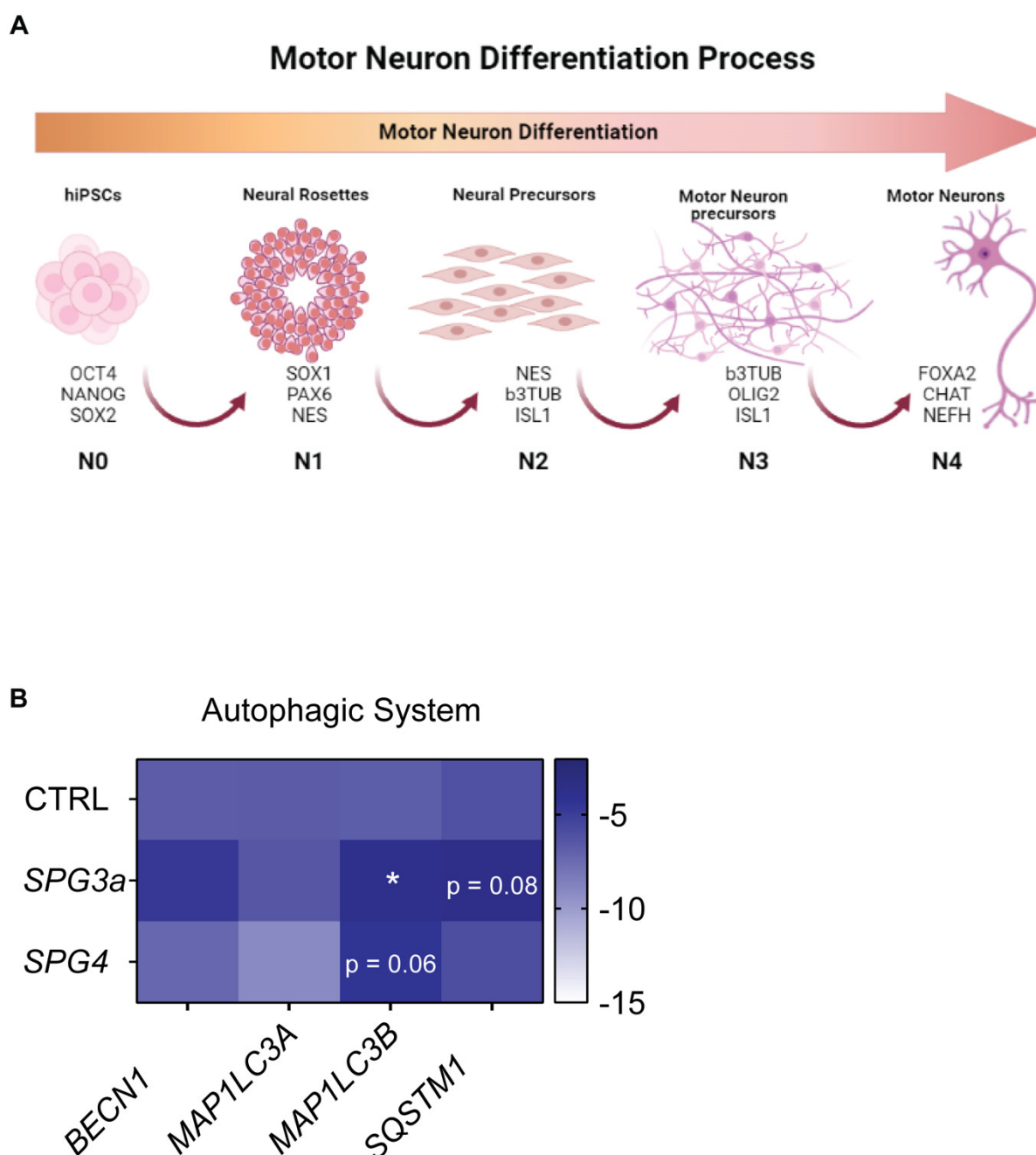


Figure S2. MN differentiation process, related to figure 4. **(A)** Schematic overview of the MN differentiation protocol for HSP and CTRL hiPSCs towards MNs. hiPSCs were kept pluripotent in N0 (E8 medium). Four distinct media were used to induce MN differentiation at four different steps: N1 to differentiate hiPSCs to the stage of neural rosettes, N2 to propagate the cells obtained, N3 to induce the MN precursors, N4 to obtain mature MNs. The qRT-PCR genes for checking the progression of the differentiation have been indicated. The protocol was adapted from Bianchi and colleagues (Bianchi et al., 2018). **(B)** Heat map for the genes regulating the autophagic proteolytic system (*BECN1*, *MAP1LC3A*, *MAP1LC3B* and *SQSTM1*) in CTRL and HSP-derived MNs. Each data point was represented as ΔCt , normalized for the housekeeping genes (*GAPDH* and *RPL13a*). Data were representative of independent experiments and values were expressed as mean \pm SEM. Significance of the differences for SPG-mutated vs. CTRL lines: * $p < 0.05$. For $p < 0.1$, the actual value was reported.

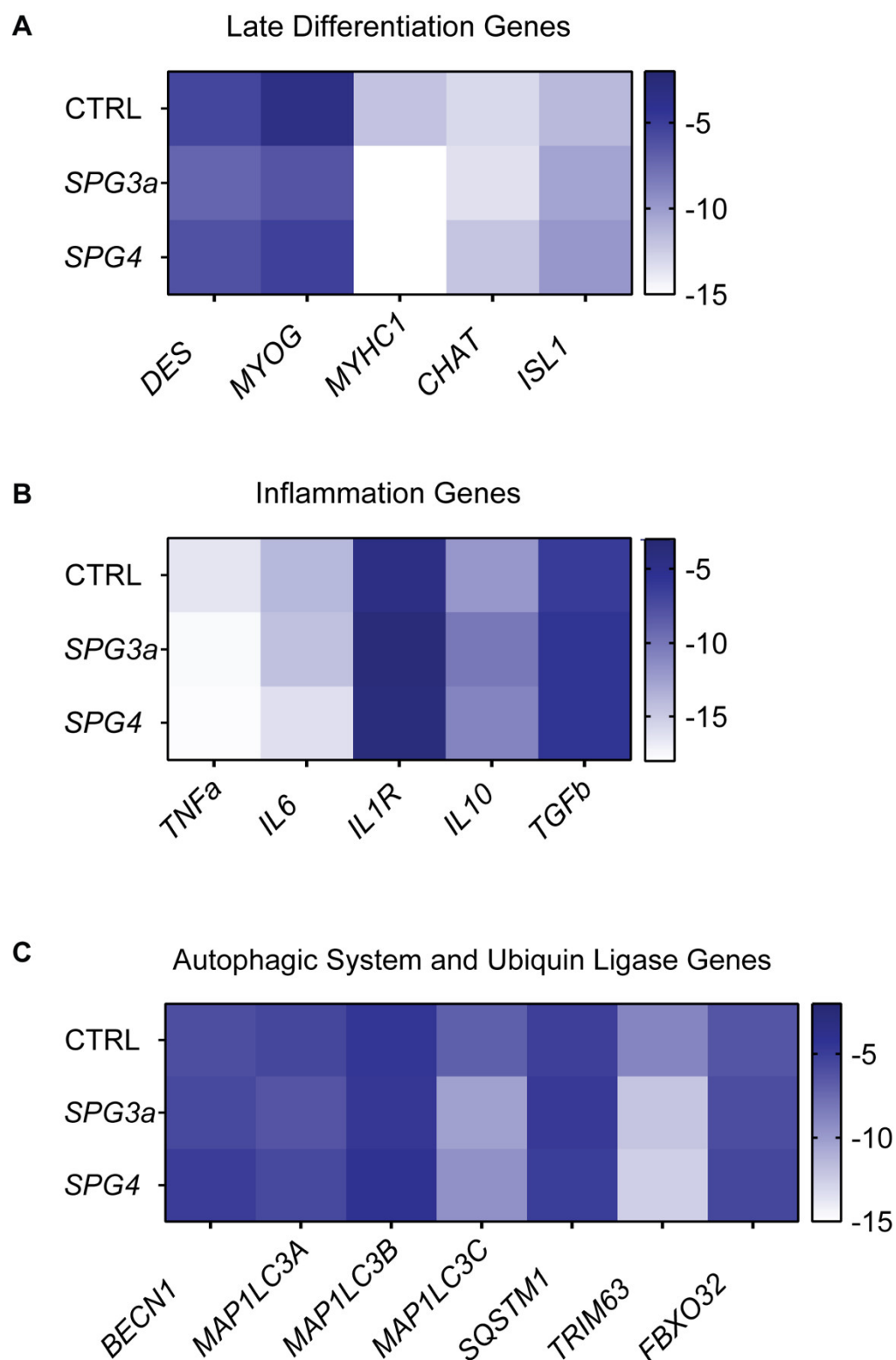


Figure S3. Gene expression profiles for NMJs, related to figure 6. Heat maps reporting genes regulating (A) the late-stage myogenic (*DES*, *MYOG* and *MYHC1*) and MN (*CHAT* and *ISL1*) differentiations, (B) inflammation (*TNFα*, *IL6*, *IL1R*, *IL10* and *TGFβ*) and (C) autophagic system (*BECN1*, *MAP1LC3A*, *MAP1LC3B*, *MAP1LC3C* and *SQSTM1*) or muscle specific E3 ubiquitin ligases (*TRIM63* and *FBXO32*). Each data point was represented as ΔCt , normalized for the housekeeping genes (*GAPDH* and *RPL13a*). Data were representative of independent experiments and values were expressed as mean \pm SEM.

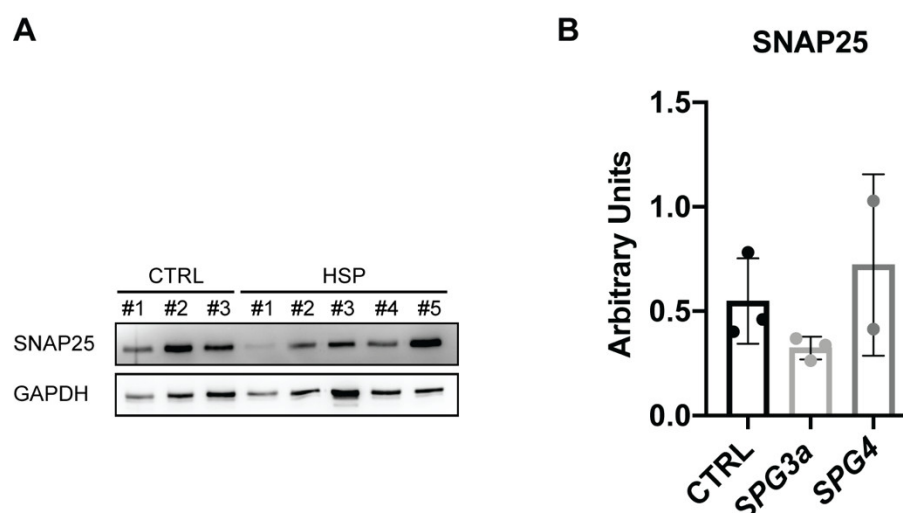


Figure S4. SNAP25 protein expression in NMJs, related to figure 6. WB analysis (**A**) and quantification (**B**) for the protein levels of SNAP25 in CTRL (three different hiPSC lines) and HSP-derived (#1, #2, #3 for *SPG3a*- and #4, #5 for *SPG4*-mutated genotypes) lines. Data were analyzed by one-way ANOVA followed by a Dunnett post hoc test. Data were representative of independent experiments and values were expressed as mean \pm SD.

SUPPLEMENTARY TABLES

Table S1. Genetic features of HSP patients, related to Experimental procedures (Study design and ethics statement).

Patient	Gender	Inheritance	Gene	Variant DNA	Protein	Variant Protein	Zygosity	Disorder
HSP #1	F	AD	<i>SPG3a</i>	c.1483C > T (exon 12)	ATLASTIN1	p.Arg495Trp	Heterozygous	<i>SPG3a</i> -HSP
HSP #2	M	AD	<i>SPG3a</i>	c.757G > A (exon 8)	ATLASTIN1	p.Val253Ile	Heterozygous	<i>SPG3a</i> -HSP
HSP #3	M	AD	<i>SPG3a</i>	c.757G > A (exon 8)	ATLASTIN1	p.Val253Ile	Heterozygous	<i>SPG3a</i> -HSP
HSP #4	M	AD	<i>SPG4</i>	c.1066G > A (exon 7)	SPASTIN	p.Glu356Lys	Heterozygous	<i>SPG4</i> -HSP
HSP #5	M	AD	<i>SPG4</i>	c.1496G > A (exon 13)	SPASTIN	p.Arg499His	Heterozygous	<i>SPG4</i> -HSP

AD: autosomal dominant.

Table S2. Clinical parameters of HSP patients, related to Experimental procedures (Study design and ethics statement).

Patient	HSP #1	HSP #2	HSP #3	HSP #4	HSP #5
Age at onset (years)	2	1	3	1	1,5
Age at diagnosis (years)	4,25	7	9	5	3
Gait	Pathologic	Pathologic	Pathologic	Pathologic	Pathologic
Lower limbs					
Muscle weakness	yes	yes	yes	yes	yes
Spasticity	yes	yes	yes	yes	yes
Hyperreflexia	yes	yes	yes	yes	yes

Treatment	Muscle contractures	yes	yes	yes	yes	yes
	Ankle clonus	yes	no	no	no	yes
	Babiski sign	yes	yes	yes	yes	yes
	GMFCS	II	I	I	I	III
	Botulinum toxin	4x (last in 2018)	3x (2003, 2005, 2012)	no	1x (2014)	1x (2012)
	Surgery	no	yes (2012)	no	no	yes (2012, 2014)
	Others	ADHD, dysarthria	no	no	Autism spectrum disorder	Intellectual disability

ADHD: attention deficit hyperactivity disorder.

Table S3. List of primers for qRT-PCR, related to Experimental procedures (Quantitative Real-Time PCR (qRT-PCR) analysis).

Gene	Primer Direction	Primer Sequence (5' > 3')
<i>ACTA</i>	forward	GAAGATTTCGTCGTCCTGAGAA
	reverse	ACCACATGTGGATCACCAAG
<i>ADAM22</i>	forward	GAAGACGAAAGTCGGCACGA
	reverse	TGAATGACGTTCCAAAGGCAT
<i>ANK2</i>	forward	ACCTGCAATCAGAATGGACTCA
	reverse	TGCAATGTGAAGAGCGGTATT
<i>b3TUB</i>	forward	CCTCCGTGTAGTGACCCTT
	reverse	GGCCTTTGGACATCTCTTCAG
<i>BRACH</i>	forward	ACCCAGTTCATAGCGGTGAC
	reverse	AAGCTTTTGCAAATGGATTG
<i>BECN1</i>	forward	AACAGCGTTTGTAGTTCTGACA
	reverse	GACTACGACTTGTGTAGCGTC
<i>CASK</i>	forward	TTGAAATCGTAAAGCGAGCTGA
	reverse	CAGTAGCGTAGAGCTTCCAGTA
<i>CDX2</i>	forward	GACGTGAGCATGTACCCTAGC
	reverse	GCGTAGCCATTCCAGTCCT
<i>CHAT</i>	forward	CCCTGATGCCTTCATCCA
	reverse	GTAGGTGGGCACCAGTCTTC
<i>CHRNA1</i>	forward	GATGAAGTAAATCAGATCGTGACAACC
	reverse	TTCACACCGCCATAGTCATCTGGATTCCAT
<i>CHRNA1</i>	forward	AGAATGGCCAGTGGGAGAATAT
	reverse	CTATTGGGTTTGGGGAAGAGAA
<i>CHRNA1</i>	forward	GTCCCAGACTTACAGCACCAATG
	reverse	GATGAGGATGGCGACAGAGGAG
<i>CHRNA1</i>	forward	AACGGGGAGTGGGAGATAGTCCA
	reverse	CTTCTTGACCCCTCAGACAGCA
<i>CHRNA1</i>	forward	CTCTCAGACGTACAATGCCGAA
	reverse	GTGGCGCAGCCGCGGGGACATGG
<i>c-MYC</i>	forward	TCCTCGGATTCTCTGCTCTCCT
	reverse	AGAAGGTGATCCAGACTCTGACCT
<i>CLTA</i>	forward	CGATTGCAGTCAGAGCCTGAAAG

	reverse	TAGCTGCTCGTCCTGTCTTGCA
CLTB	forward	CAACGGTCCTGCTGATGGCTAC
	reverse	GCCATTCCTGTTCCGTGACCTT
DES	forward	TCGGCTCTAAGGGCTCCTC
	reverse	CGTGGTCAGAACTCCTGGTT
DLG1	forward	TCGAACTAGCCAGAAGCGATCC
	reverse	TGCCTGGCTTGCCACCATTCAT
DLG2	forward	GCCGGTGATTATCCTGGGG
	reverse	CGCTTTGGCCTCGTAGTATGA
DLG3	forward	TGGAGGTCAACCTGCTCAAAGG
	reverse	TAGGCGTCCATCCTTCTGAGCA
DLG4	forward	ACCGAGGCGAATTGTGATCC
	reverse	CCGTCTGACCCGCATTCTT
DNM1	forward	ATCGCCACTTGGCTGACCGTAT
	reverse	CCTCCTTCTCAATGGACAGTAGC
DNM2	forward	CGAGTCACTGTCCTGGTACAAG
	reverse	TGCTCCGTGTTGAAGATGGCGA
DNM3	forward	GCTCACCATCAGCAACATTGGC
	reverse	CCGAACTTTCAGGTTGTCCAAGG
DOC2B	forward	CACCAAGACCTTCAACATCTGCC
	reverse	GCCTTGCTTCTGTGAGCTGTAC
EN1	forward	CGCAGCAGCCTCTCGTATG
	reverse	CCTGGAACCTCCGCCTTGAG
EOMES	forward	CTGCCCCACTACAATGTGTTCCG
	reverse	GCGCCTTTGTTATTGGTGAGTTT
FOXA2	forward	GGAGCAGCTACTATGCAGAGC
	reverse	CGTGTTTCATGCCGTTTCATCC
FOXO32	forward	GCCTTTGTGCCTACAACCTGAA
	reverse	CTGCCCTTTGTCTGACAGAAT
GAPDH	forward	TCAAGAAGGTGGTGAAGCAGG
	reverse	ACCAGGAAATGAGCTTGACAAA
GATA4	forward	CGACACCCCAATCTCGATATG
	reverse	GTTGCACAGATAGTGACCCGT
GDF-3	forward	ACACCTGTGCCAGACTAAGATGCT
	reverse	TGACGGTGGCAGAGGTTCTTACAA
HAND2	forward	ATGAGTCTGGTAGGTGGTTTTCC
	reverse	CATACTCGGGGCTGTAGGACA
HB9	forward	CTTCTGTTTCTCCGCTTCCT
	reverse	CACCTCGCTCATGCTCAG
HOXC4	forward	ACGAGAAAGAGAGTGGGAGAGA

	reverse	GGAGGTCTGGGGTTGAG
<i>HPRT</i>	forward	TGACACTGGCAAAACAATGCA
	reverse	GGTCCTTTTCACCAGCAAGCT
<i>hTERT</i>	forward	AAATGCGGCCCTGTTTCT
	reverse	CAGTGCGTCTTGAGGAGCA
<i>IL10</i>	forward	GACTTTAAGGGTTACCTGGGTTG
	reverse	TCACATGCGCCTTGATGTCTG
<i>IL1R</i>	forward	ATGAAATTGATGTTTCGTCCCTGT
	reverse	ACCACGCAATAGTAATGTCCTG
<i>IL6</i>	forward	ACTCACCTCTTCAGAACGAATTG
	reverse	CCATCTTTGGAAGGTTTCAGGTTG
<i>ISL1</i>	forward	GCGGAGTGTAATCAGTATTTGGA
	reverse	GCATTTGATCCCGTACAACCT
<i>KLF-4</i>	forward	CGGACATCAACGACGTGAG
	reverse	GACGCCTTCAGCACGAACT
<i>MAP1LC3A</i>	forward	AACATGAGCGAGTTGGTCAAG
	reverse	GCTCGTAGATGTCCGCGAT
<i>MAP1LC3B</i>	forward	AAGGCGCTTACAGCTCAATG
	reverse	CTGGGAGGCATAGACCATGT
<i>MIXL1</i>	forward	CTGAGGAGCCATGACTGACA
	reverse	TGGGAGTGTGGGCTTAAAC
<i>MSGN</i>	forward	CTGCACACCCTCCGGAATTA
	reverse	AGGAGGTCTGTGAGTTCCCC
<i>MYHC1</i>	forward	GACATTGACCACACCCAGTATAA
	reverse	CAGCTTCTCATCTCGCATCTC
<i>MYOD</i>	forward	CCGCCTGAGCAAAGTAAATG
	reverse	CGATATAGCGGATGGCGTT
<i>MYOG</i>	forward	CAGGCTCAAGAAGGTGAATGA
	reverse	CGATGTACTGGATGGCACTG
<i>NANOG</i>	forward	TGGCCGAAGAATAGCAATGGTGTG
	reverse	TTCCAGGTCTGGTTGCTCCACATT
<i>NEFH</i>	forward	GTGAAGAGTGTCCGATTGGCT
	reverse	ACACAGAGGGAATTTTGGGGA
<i>NEFM</i>	forward	ACAACCACGACCTCAGCAGCTA
	reverse	GTTGAGGAGGTCTCTGGTATTCTG
<i>NES</i>	forward	TCAGCTTTCAGGACCCCAAG
	reverse	TGGGAGCAAAGATCCAAGACG
<i>NEUROD1</i>	forward	CTGCTCAGGACCTACTAACAACAA
	reverse	GTCCAGCTTGGAGGACCTT
<i>NEUROG2</i>	forward	TCCTCTTCTCCTTCAACTCC
	reverse	GCCAAAGTCACAGCAACG
<i>NFL</i>	forward	CCAAGACCTCCTCAACGTGAAG
	reverse	ATGCTTCCCACGCTGGTGAAC
<i>NKX2.5</i>	forward	ACCTCAACAGCTCCCTGACTCT
	reverse	ATAATCGCCGCCACAACTCTCC
<i>OCT4</i>	forward	CGAGCAATTTGCCAAGCTCCTGAA
	reverse	GCCGCAGCTTACACATGTTCTTGA

<i>OLIG2</i>	forward	CACAGAGCAGTGGGGAGTG
	reverse	GCACACAGCGGTACCTTTTC
<i>PAX3</i>	forward	ATTAAGCCACACATGCCGGT
	reverse	TACACAAGGAAGCCCCTGCT
<i>PAX6</i>	forward	AACGATAACATACCAAGCGTGT
	reverse	GGTCTGCCCCGTTCAACATC
<i>PAX7</i>	forward	GGGCCTCCTGCTTGTTTAT
	reverse	CCATCTGGCTGGACTTCAAT
<i>REX1</i>	forward	TGGAGGAATACCTGGCATTGACCT
	reverse	AGCGATTGCGCTCAGACTGTCATA
<i>RPL13a</i>	forward	CCTGGAGGAGAAGAGGAAAGAGA
	reverse	TTGAGGACCTCTGTGTATTTGTCAA
<i>SHANK2</i>	forward	CGTGGCAAGCCGGAATAAG
	reverse	AAGGACCAGGTGATTCCCTCC
<i>SLC17A6</i>	forward	GGGTTTTTCGGAGCTGCCATA
	reverse	CACACCCTCAACAAGTCCCTG
<i>SLC17A7</i>	forward	CAGAGTTTTTCGGCTTTGCTATTG
	reverse	GCGACTCCGTTCTAAGGGTG
<i>SNAP25</i>	forward	ACCAGTTGGCTGATGAGTCG
	reverse	CAAAGTCCTGATACCAGCATCTT
<i>SOX1</i>	forward	AAAGTCAAAACGAGGCGAGA
	reverse	AAGTGCTTGGACCTGCCTTA
<i>SOX2</i>	forward	TGGCGAACCATCTCTGTGGT
	reverse	CCAACGGTGTCAACCTGCAT
<i>SOX17</i>	forward	GTGGACCGCACGGAATTTG
	reverse	GGAGATTCACACCGGAGTCA
<i>SQSTM1</i>	forward	GACTACGACTTGTGTAGCGTC
	reverse	AGTGTCCGTGTTTCACCTTCC
<i>STON1</i>	forward	TATGAGAGTGCCTACCAGGCAG
	reverse	ACTGAACAGTAGCAAATGGATACC
<i>STON2</i>	forward	GTGTGGAGGATAAACCGACTGC
	reverse	CGTGATTGGCAAATCTGGAAGGC
<i>STX1A</i>	forward	TAAAGAGCATCGAGCAGTCCA
	reverse	GACATGACCTCCACAACTTTCT
<i>SVOP</i>	forward	TGGACTCTGTACTATGGCATCC
	reverse	TGGCCCAGAATACCTCAATCA
<i>SYN1</i>	forward	TGCTCAGCAGTACAACGTACC
	reverse	GACACTTGCGATGTCCTGGAA
<i>SYNDIG1</i>	forward	ACACCCTGTCCTACGATGTG
	reverse	TCTCTGTGTGCGTTGAGTAGTC
<i>SYP</i>	forward	CTCGGCTTTGTGAAGGTGCT
	reverse	CTGAGGTCACTCTCGGTCTTG
<i>SYT2</i>	forward	CGTGGACAACCTCCACTGAGAGT

	reverse	CAACCACAGCAATGGCGATCAG
SYT3	forward	AGCCCTCCTACTTGGACATGG
	reverse	GGGATGTTTGGCTCGGTTTGA
SYT6	forward	TACGCTACGATTACGAGACCG
	reverse	GCTTCCACAAAAGTCCTTGGC
SYT12	forward	GTTCCCAATTACGACTACAGG
	reverse	TCAATGCTGAGACTGCCTTTG
TBR1	forward	TCACTGGAGGTTTCAAGGAGGC
	reverse	TTTCTTGGCGCATCCAGTGAGC
TGFβ	forward	CAATTCCTGGCGATACCTCAG
	reverse	GCACAACCTCCGGTGACATCAA
TNFα	forward	CCTCTCTCTAATCAGCCCTCTG
	reverse	GAGGACCTGGGAGTAGATGAG
TRIM64	forward	CTTCCAGGCTGCAAATCCCTA
	reverse	ACACTCCGTGACGATCCATGA
VAMP1	forward	ACATGACCAGTAACAGACGACT
	reverse	ACGTTACACGTATGATGTCC

Table S4. List of antibodies for IF and WB, related to Experimental procedures (IF, Western blot).

Protein	Antibody Name (#Catalog Number)	Provider	IF	WB
ATL1	ATL1 Monoclonal Antibody (1F6B12) (Rabbit monoclonal) (#MA5-31641)	Thermo Fisher Scientific		1:1000
ACTN2	Anti-SARCOMERIC ALPHA ACTININ (EA-53) (Mouse monoclonal) (#ab9465)	Abcam	1:200	1:500
GAPDH	Anti-GAPDH Antibody (Rabbit polyclonal) (G9545)	Merck		1:1000
ISL1	Recombinant Anti-ISLET 1 antibody (EP4182) - Neural Stem Cell Marker (Rabbit monoclonal) (#ab109517)	Abcam	1:500	1:60
LIN28	LIN-28 (S-15) (Goat polyclonal) (#sc-54032)	Santa Cruz Biotechnology	1:50	
MF20	MYHC1 (MF20) (Mouse monoclonal)	Development Studies Hybridoma Bank (DSHB)	1:300	1:3
MYOD	MYOD1 (D8G3) XP Rabbit mAb (Rabbit monoclonal) (#13812)	Cell Signaling	1:300	1:1000
NANOG	NANOG (Rabbit polyclonal) (#PA1-097)	Thermo Fisher Scientific	1:200	
NEFH	Anti-NEUROFILAMENT HEAVY POLYPEPTIDE antibody (Rabbit polyclonal) (#ab8135)	Abcam	1:1000	1:100
NEST	Purified anti-NESTIN Antibody (10C2) (Mouse monoclonal) (#656801)	BioLegend		1:250
OCT4	Anti-OCT4 - Embryonic Stem Cell Marker (Goat polyclonal) (#ab27985)	Abcam	1:200	
OCT4	Anti-OCT4 antibody (Rabbit polyclonal) (#ab19857)	Abcam		1:150
PAX6	PAX6 (D3A9V) XP Rabbit mAb (Rabbit monoclonal) (#60433)	Cell Signaling	1:100	1:150
SOX2	SOX-2 (Y-17)	Santa Cruz Biotechnology	1:50	

	(Goat polyclonal) (#sc-17320)			
SPAST	Anti-SPASTIN antibody (Sp3G11/1) (Mouse monoclonal) (#ab31850)	Abcam	1:200	
SSEA4	SSEA-4 (MC813) (Mouse monoclonal) (#sc-59368)	Santa Cruz Biotechnology	1:50	
TBA1B	Acetyl- α -TUBULIN (Lys40) (D20G3) (Rabbit monoclonal) (#5335)	Cell Signaling	1:500	
TRA-1-60	TRA-1-60 (TRA-1-60) (Mouse monoclonal) (#sc-21705)	Santa Cruz Biotechnology	1:50	
TUBA4A	Anti- α -TUBULIN antibody (B-5-1-2) (Mouse monoclonal) (#T6074)	Merck	1:1000	
TUBB3	Anti-beta III TUBULIN antibody (2G10) - Neuronal Marker (Mouse monoclonal) (#ab78078)	Abcam	1:500	1:300

Supplementary Video S1. related to figure 6. Phase-contrast video-micrograph of spontaneously contracting hiPSC-derived CTRL myotubes co-cultured with hiPSC-derived CTRL MNs at day 9.

Supplementary Video S2. related to figure 6. Phase-contrast video-micrograph of spontaneously contracting hiPSC-derived *SPG3a*-mutated myotubes co-cultured with hiPSC-derived *SPG3a*-mutated MNs at day 9.

Supplementary Video S3. related to figure 6. Phase-contrast video-micrograph of spontaneously contracting hiPSC-derived *SPG4*-mutated myotubes co-cultured with hiPSC-derived *SPG4*-mutated MNs at day 9.