



Sex Chromosomes and Sex Determination in Dioecious Agricultural Plants

Olga V. Razumova ^{1,2,*}, Oleg S. Alexandrov ¹, Karina D. Bone ¹, Gennady I. Karlov ¹, and Mikhail G. Divashuk ¹

- ¹ All-Russia Research Institute of Agricultural Biotechnology, 127550 Moscow, Russia
- ² Tsitsin Main Botanical Garden RAS, Botanicheskaya St., 4, 127276 Moscow, Russia

* Correspondence: razumovao@gmail.com

Abstract: Unlike in animals, dioecy among flowering plants is a rare phenomenon. The vast majority of angiosperm species have a bisexual flower that combines male (androecium) and female (gynoecium) reproductive organs. However, about a quarter of species have dioecious flowers, which can be located within the same plant (monoecious) or on different plants (dioecious). The flower formation in dioecious plants is determined by various genetic mechanisms. They become more complex from the work of a single gene to the functioning of full-fledged heteromorphic sex chromosomes, which can directly affect sex differentiation or participate in the balance determination of sex (where the formation of male or female flower organs depends on the ratio of X chromosomes to autosomes, for example). In recent years, the development of sequencing techniques, bioinformatics, and molecular biology has led to an increase in interest in the sex determination mechanisms among plants. It is noteworthy that a significant number of dioecious plants have economic value. At the same time, dioecious crops. In this review, we attempt to summarize the current information on sex chromosomes and the mechanisms of sex determination in dioecious plants, concentrating on species with agricultural importance.

Keywords: dioeciousness; flowering plants; monoecious plant; dioecious plant; sex chromosomes; dioecious crops; sex determination

1. Introduction

Dioecy is a relatively rare phenomenon in the world of angiosperms. Dioecious species make up about 5-7% of angiosperms [1–3]. The active development of bioinformatics methods, reductions in the cost of whole-genome sequencing methods, and the creation of chromosome assemblies in recent years have opened up wide-ranging opportunities for studying the mechanisms of sex determination in plants. Using new technologies, it was confirmed that dioecy arose repeatedly, in different ways, and in evolutionarily distant species [4]. The events associated with the emergence of dioecy have occurred hundreds or even thousands of times. Reverse events (a return to monoecy or hermaphroditism) have also occurred [5–7]. Consequently, it is necessary to study the sex determination mechanisms in each individual case of dioecy occurrence. Additionally, data obtained on one species cannot be interpolated to other, even phylogenetically close species. At the same time, species with dioecious flowers are quite often (about in 20% of cases) cultivated as agricultural crops [8]. The dioecy of agricultural crops must be considered in industrial cultivation. On the one hand, dioecy can be beneficial for growers, since all flowers of female plants produce fruit and the plants do not waste resources on "less profitable" male flowers. On the other hand, dioecy can create problems. For example, the sex of plants is often unknown at the juvenile stage of ontogenesis. However, growers need to know the sex of the plants so that the ratio of female to male plants is optimal for



Citation: Razumova, O.V.; Alexandrov, O.S.; Bone, K.D.; Karlov, G.I.; Divashuk, M.G. Sex Chromosomes and Sex Determination in Dioecious Agricultural Plants. *Agronomy* **2023**, *13*, 540. https://doi.org/10.3390/ agronomy13020540

Academic Editor: Agnieszka Golicz

Received: 30 December 2022 Revised: 27 January 2023 Accepted: 9 February 2023 Published: 14 February 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). efficient pollination when setting up a nursery. In other cases, the presence of male plants in a plantation is generally unacceptable, and they must be removed before the flowering phase. Sometimes, morphological or physiological differences between plants of different sexes must be considered in the production of products from dioecious species. Thus, it is essential to have tools for the early sex differentiation of seedlings.

It has been repeatedly reported that the frequency of dioecious species occurrence varies depending on geographical conditions. It is much greater in tropical and neotropical zones than in subpolar regions. Woody and climbing forms, as well as plants with succulent fruits distributed by animals, are also more likely to be dioecious than herbaceous plants with dry fruits [9]. What environmental factors lead to the appearance of dioecious species? What evolutionary mechanisms contribute to the fixation of dioecy in some species and its culling in others? Why are these dioecious species fixed in the course of evolution? What are the benefits of separating the sexes? These questions are still debatable. It was previously believed that a mutation in two genes is required for the occurrence of dioecy. This is the "two-gene model" [10,11]. However, recent research shows that this is not the only evolutionary path. The emergence of dioecious plants is also possible with a mutation in one locus ("single-gene model"). In nature, confirmations of both the first and second models have been found [12–17]. However, such models are created on the basis of the existence of natural selection in populations. They are incorrect for dioecious crops since these have been subjected to artificial selection for a long time. Despite these difficulties, modern methods of genomics and cytogenetics have helped scientists come to an understanding of the occurrence of dioecy, at least for some species.

The occurrence of dioecy does not always lead to the formation of heteromorphic sex chromosomes [18,19]. In addition, the degree of differentiation of sex chromosomes does not always correlate with the evolutionary age of dioecy in the species under study. Therefore, it is necessary to clearly differentiate the emergence and evolution of sex determination regions and heteromorphic sex chromosomes.

In recent years, the development of technologies for high-throughput genomic sequencing, the analysis of repeatomes and proteomes, GWAS (genome-wide association studies), and other omics technologies have raised the study of wildlife to a qualitatively new and deeper level. Having obtained data on simple model objects (Arabidopsis, Drosophila or C. elegans), researchers chose economically important plant and animal species as the next candidates for more detailed studies. At the same time, researchers immediately encountered a number of problems. In the case of plants, these problems usually include huge genome sizes (for example, *Triticum aestivum* genome 1C = 17.30 pg; *Allium cepa* 1C = 17.90 pg [20]) and a large amount of non-coding DNA that, in some species, makes up 50-85% of the genome size [21,22]. Dioecious crop species have additional challenges in accurate sequencing due to differences in genotypes between male and female plants (often more than 3% of the total genome size, for example, in *Humulus japonicus* or *Rumex* spp. [23,24]), the presence of a non-recombining region in the heterogametic sex, and a pseudoautosomal region (PAR) that greatly varies in size among species [25]. Despite these difficulties, a large amount of data about the genomes of various agricultural dioecious species has been accumulated to date (Table 1).

Species	Sex Chromosome System	Genome Sequenced *	Molecular Markers	Reference
Asparagus officinalis	XX/XY	Yes	Yes	[26]
Rumex acetosa	XX/XY1Y2	No	Yes	[27]
Spinacia oleracea	XX/XY	Yes	Yes	[28]
Cannabis sativa	XX/XY	Yes	Yes	[29]
Piper betle	unknown	No	Yes	[30]

Table 1. Genetic characteristics of the most agriculturally important dioecious plants species.

Species	Sex Chromosome System	Genome Sequenced *	Molecular Markers	Reference
Dioscorea cayenensis subsp. rotundata	ZZ/ZW	Yes	Yes	[31]
Carica papaya	XX/XY/XY	Yes	Yes	[32,33]
Vitis vinifera	monoecious	Yes	-	-
Diospyros kaki	XX/XY	Yes	Yes	[34]
Pistacia vera	ZW/WW	Yes	Yes	[35,36]
Hippophae rhamnoides	XX/XY	No	Yes	[37]
Humulus lupulus	XX/XY	Yes	Yes	[38]
Phoenix dactylifera	XX/XY	Yes	Yes	[39]
Myristica fragrans	XX/XY	No	Yes	[40]
Actinidia chinensis	XX/XY	Yes	Yes	[41]
Ilex paraguariensis	unknown	Yes	No	-
Morella rubra	ZW/WW	Yes	Yes	[42]
Amaranthus palmeri	XX\XY	Yes	Yes	[43]

Table 1. Cont.

 * All information about the availability of genome sequences is presented according to the NCBI website.

Here, we attempt to summarize the data available to date for a number of major cultivated dioecious crops, paying special attention to the influence of dioecious nature on the agronomic features of their cultivation (Table 2).

Table 2. Particular qualities of the most agriculturally important dioecious plants species.

Life-Forms	Species	Common Name	Part of a Plant Used by Humans	Direction of Use	What Type of Plants Are Used
	Asparagus officinalis	Asparagus	Stock	Vegetable	Male or supermale
	Rumex acetosa	Sorrel	Leaves	Green crops	Male and female
	Spinacia oleracea	Spinach	Leaves	Green crops	Male and female
Grass annual or perennial	Cannabis sativa	Hemp	Leaves, seeds, fibers,	Industrial, food or medicinal use	Female, male and monoecious
	Piper betle	Betel leaves	Leaves	Medicinal	Male and female
	Dioscorea spp.	Yam	Modified tubers	Food	Male and female
	Amaranthus palmeri	Green amaranth	-	Weeds	-
Woody perennial	Carica papaya	Papaya	Fruit	Fruit crops	Hermaphroditic
	Vitis vinifera	Grape	Fruit	Fruit crops	Only wild forms are dioecious; varieties of cultivated grapes are hermaphroditic
	Diospyros sp.	Persimmon	Fruit	Fruit crops	Female or monoecious; male needed for pollination
	Pistacia vera	Pistacia	Fruit	Nut-bearing crops	Female; male needed for pollination
	Hippophae rhamnoides	Sea buckthorn	Fruit	Berry crops, medicinal	Female; male needed for pollination

Life-Forms	Species	Common Name	Part of a Plant Used by Humans	Direction of Use	What Type of Plants Are Used
	Humulus lupulus	Нор	Fruit	Hop culture	Female; male plants are only needed for breeding to develop new cultivars
	Phoenix dactylifera	Date palm	Fruit	Fruit crops	Female; male needed for pollination
	Myristica fragrans	Nutmeg	Fruit	Cultural spices	Female; male needed for pollination
	Actinidia chinensis	Kiwifruit	Fruit	Fruit crops	Female; male needed for pollination
	Ilex paraguariensis	Yerba Mate	Leaves	Tea crops	Female and male
	Morella rubra	Chinese Bayberry	Fruit	Fruit crops	Female; male needed for pollination

Table 2. Cont.

2. Features of the Dioecious Crop Cultivation

2.1. Sex Chromosomes and Sex Determination in Herbaceous Crops

As a rule, vegetative parts are used in herbaceous dioecious crops, and their cultivation often does not need sexing, as in the cultivation of perennial fruit crops. However, many herbaceous dioecious plants are convenient model objects for laboratory studies. They demonstrate rapid growth and development, bloom in the first or third year, do not require large areas, and are easily introduced into in vitro cultures. Therefore, the sex chromosomes and mechanisms of sex determination in cultivated herbaceous plants have been most fully studied. Molecular sex markers for many of them have been created, although they is little demand for them in industry. At the same time, there is evidence that the male and female plants of some herbaceous crops may differ in productivity, regrowth time and technical characteristics [44].

2.1.1. Asparagus

The *Asparagus* genus includes about 200 species, among which there are hermaphroditic, dioecious, and monoecious representatives, but *Asparagus officinalis* L. has the greatest popularity in production [45]. Garden asparagus is an economically important perennial herbaceous vegetable crop that is widely grown in Asia, Europe and the Americas. The leader in production is China, and the sown area of this crop around the world is constantly increasing [46]. Juicy young shoots are used for food.

This is one of the first dioecious plants in which sex determination was studied [47]. Asparagus plants are diploid, 2n = 2x = 20, and their sex chromosomes are homomorphic and do not visually differ from each other [48,49]. The female plants are homogametic and have XX chromosomes (also known as genotype mm) in their karyotype, while the male plants are heterogametic (XY\Mm karyotype). At the same time, the homogametic karyotype YY (MM), the so-called supermale, is viable, has fertile pollen, and is androdiecious in phenotype—that is, it has male and hermaphrodite flowers. The viability of the YY homozygote indicates the early stages of the dioecy evolution path with the young Y chromosome [50]. The mechanism of sex regulation in asparagus is one of the most studied among dioecious plants. A small M locus (835 kb in size) located on homomorphic sex chromosomes is responsible for sex determination in this species [12]. The development of male and female reproductive organs requires the presence of two key genes located on the sex chromosomes. These are the gene that suppresses female function (SOFF) and the gene that promotes the development and functioning of the tapetum (aspTDF1). Transcriptome analysis revealed more than 500 differently expressed genes in male and female plants [51].

Asparagus DNA methylation was studied using whole-genome BS sequencing (bisulfite sequencing) [52]. A difference was found in the DNA methylation containe d in male and female flowers. However, no difference in methylation was found in the genes responsible for the formation of flowers.

Male and supermale plants are preferred in asparagus production [44,53]. Male plants have been shown to have higher shoots per plant, increased yield, early maturity, and increased longevity [54,55]. Therefore, attempts are being made to create hybrids only consisting of male plants. They are created in ways such as the cultivation of anthers in vitro followed by dihaploidization with colchicine [56,57] and the self-pollination of andromonoecious plants. The cultivation of anthers leads to the formation of supermale plants, which are extremely rare in natural populations. In the production of asparagus, rhizomes are planted, and the sex can only be determined after flowering, which occurs in the second or third year. In this regard, the development of molecular markers, which make it possible to identify male, female, and supermale plants at the early stages of germination, seemed relevant. To determine the sex of garden asparagus, different types of markers have been developed (RAPD [56], RFLP [58], AFLP [59,60], and SSR [61]). Currently, the most interesting and relevant marker is RM17, developed by Stone et al. using whole-genome sequencing data [62]. This marker is codominant—that is, it is able to detect not only male and female but also supermale plants.

2.1.2. Sorrel

The *Rumex* genus is one of the largest in the Polygonaceae family—it has about 200 species [63,64]. Sorrel is a perennial herbaceous crop. *Rumex* spp. differ according to their economic importance. Some species (primarily *R. acetosa*) are cultivated for food consumption, while others (*R. obtusifolius*, *R. crispus*, and *R. acetosa*) are a big problem for agriculture because they are weeds in some regions of Europe, North America and Asia [65–67]. In nature, *Rumex* spp. are widely distributed in Eurasia, North America, Africa, mainly in the Northern Hemisphere. In culture, sorrel is primarily known as a traditional plant in Europe and Asia. Being adaptable to various conditions, it is widely grown throughout Russia in household plots. According to the N. I. Vavilov All-Russian Institute of Plant Genetic Resources (VIR) classification, it is introduced into cultures, selection is carried out, and new varieties are created [68]. Some species of sorrel are used as fodder crops [69]. Additionally, sorrel species are rich in secondary metabolites, primarily flavonoids and antioxidants. Because of this, they are used in traditional medicine around the world. The use of sorrel substances to create medicines is widely studied [63,70,71].

The *Rumex* genus is one of the model groups for studying sex determination in plants [72]. Therefore, sex determination and sex chromosomes are widely studied in a number of *Rumex* species. Among these species, there are various systems for determining sex and sex chromosomes (completely hermaphroditic species, monoecious species with XY chromosomes and an active Y chromosome, and species with X:A balance sex determination, including multichromosomal species with the XY1Y2 system). Species such as R. acetosa, R. acetosella, R. hastatalus, and R. suffruticosus are dioecious and widely studied. The R. acetosa karyotype consists of 12 autosomes and sex chromosomes (XX in female plants and the XY1Y2 multichromosomal system in male plants) [73–76]. The sex chromosomes are the largest in the karyotype. Two Y chromosomes contain 26% of the total DNA. Sorrel Y chromosomes are heterochromatic and rich in repetitive DNA sequences [77]. In 1999, a group of tandem repeats specific to male sex chromosomes were isolated and called RAYSI (*Rumex acetosa* Y-chromosome-specific I) [78]. After that, a number of satellite DNA repeats specific for sorrel Y chromosomes were described. The repeat sequences of RAYSII and RAYSIII are AT-rich and show about 60% homology with RAYSI [73]. Repeat satDNA RAE180 is localized on two Y chromosomes and is also present on a pair of autosomes. [79]. In addition, it was shown that, in addition to satellite DNA, mobile elements played an important role in the evolution of sorrel Y chromosomes [80]. At the moment, we have not been able to find information about any differences in economically valuable traits in the male and female plants of *R. acetosa*. However, there is evidence of different morphologies in male and female plants in *R. lunaria* depending on temperature and humidity [81]. This

trait can potentially affect yield, but there are no data on the economic efficiency of growing male and female plants of this crop.

2.1.3. Spinacia

Spinach is a perennial herbaceous plant of the Chenopodiaceae family. It has been cultivated for over 2000 years. In industry, vegetable spinach (*Spinacia oleracea* L.) is primarily used [82]. Two related wild-growing species (*S. tetrandra* and *S. turkestanica*) with sex chromosomes are studied and used for the breeding improvement of this species [83]. Spinach leaves are eaten fresh or cooked. Spinach is cultivated in more than 50 countries around the world, and China is the leader in production [28,84]. The karyotype of *S. oleraceae* is 2n = 2x = 12. Male plants are heterogametic [85]. Like asparagus, the spinach sex chromosomes are evolutionarily young and homomorphic [86,87], and the YY karyotype is viable. It can be formed by self-pollination of andromonoecious XY plants, resulting in supermales. There are individual monoecious genotypes [88].

Currently, the spinach genome has been sequenced, including by PacBio and Hi-C scaffolding. The sex determination region (SDR) was determined; it is located on chromosome 1 or 4 (according to other sources) [28]. It has been shown that the size of the SDR is 24.1 Mb on the Y chromosome and 13 Mb on the X chromosome. At the same time, the SDR on the Y chromosome contains inversions (14.1 Mb) and duplications (10 Mb) that are specific for male plants [89].

Studies have shown differences in morphological traits between male, female, and monoecious spinach plants. Vitale et al. (1985) showed that male and female plants significantly differ in height and the biomass of roots and the vegetative part while monoecious (cosexual) plants differ in the studied traits between both male and female plants [90]. Another study revealed differences in the response of spinach plants of different sexes (for example, in yield) when fertilized with selenium. Female plants were more productive and contained fewer heavy metals than male plants when selenate was applied. In addition, the contents of vitamin C, chlorophyll and carotenes increased in female plants [91]. Similar data showing that male spinach plants are taller than females but that females have a higher biomass were obtained in a later study. The response to biotic stress was also found to differ depending on the sex of the spinach [92].

2.1.4. Hemp

Hemp is an extremely polymorphic species of the Cannabaceae family, the only species of the monotypic *Cannabis* genus. Normally, these are dioecious plants, with the XX/XY system of sex chromosomes and the 2n = 2x = 18 + XX/XY karyotype [93]. The *Cannabis sativa* genome has been completely sequenced (including by modern PacBio methods). The genome size is 818 Mb for female plants and 843 Mb for male plants [94–97]. Transcriptome analysis has revealed sex-related genes.

Hemp is one of the oldest cultivated crops. According to some data, its domestication is estimated to have occurred 7000–10,000 years ago [98,99]. The cannabis sex chromosome system is also very ancient [18]. It is a multifunctional crop grown for oil and fiber, which are used in the manufacturing of biocomposite materials, as well as in the food and paint industries. Hemp is also widely known as a plant for medicinal use. According to some reports, more than 25,000 products are obtained from hemp [100]. Currently, hemp is considered one of the "greenest" crops. It is considered to have a negative carbon footprint—that is, more CO_2 is absorbed during its cultivation than is emitted [101]. In the middle of the 20th century, the area covered by this crop was declining, but in recent years, hemp has become increasingly popular, and its production has increased [100]. Due to the presence of addictive alkaloids in plants (primarily tetrahydrocannabinol (THC) and cannabidiol), cannabis cultivation is strictly regulated in many countries.

Hemp is one of the few annual agricultural herbaceous plants that are dioecious in nature. In addition, male and female cannabis plants differ in habit. The dioecy of hemp seriously affects its agricultural practices. Male and female plants differ in ripening time.

Female plants are leafier, have more lignified cell walls due to the better (compared with male plants) accumulation of lignin, bloom later, and stay green for longer [102]. These features lead to different levels of fiber quality when growing hemp as a textile crop. Cultivation technology requires expensive and time-consuming manual weeding. When growing hemp for oil, more female plants and fewer male pollinating plants are required. When growing female cannabis plants for medicinal purposes, pollination should not be allowed, since unpollinated flowers are characterized by a higher THC content. In this case, the presence of male plants in the population is undesirable. In this regard, the cultivation of monoecious varieties is more cost-effective [103]. The nature of hemp's monoecy is not yet fully understood. A number of studies have shown that monoecious varieties are female plants [105]. Male plants have 18 autosomes and XY chromosomes in their karyotype. At the same time, sex is determined according to the balance mechanism.

To recognize the sex of plants at any stage of cultivation, molecular genetic markers have been developed in hemp [29,106,107]. However, due to the annual nature of the culture, it seems more promising to create monoecious varieties. However, the ratio of male to female flowers in monoecious plants, as well as the sex of cannabis plants in general, is a very unstable trait. There have been cases of the appearance of flowers of the opposite sex in dioecious plants, as well as sex changes under the influence of environmental factors [108–111]. In this regard, a detailed understanding of the mechanisms of the occurrence of dioecy in hemp, as well as the potential opportunity to influence this trait, is of undoubted agronomic importance.

2.1.5. Betel

Betel (*Piper betle* L.) is a perennial evergreen dioecious vine of the Piperaceae family. This crop is economically significant for Asian countries, primarily for India, where it is one of the most important export crops [112]. Betel is grown for its leaves, which are chewed after meals and widely used in traditional medicine in the countries of Central and Southeastern Asia [113–117]. This plant also has important religious and ceremonial significance. Traditionally, this culture is vegetatively propagated [118]. Betel rarely blooms, but there have been reports of economically significant differences in the varieties of male and female plants [30]. There is very little information about the chromosome numbers and the level of ploidy of this species. The main number of chromosomes of this species is x = 13; however, polyploid rows are reported in this culture. Apparently, plants with a high level of ploidy are mainly grown. It was shown that female plants are tetraploid and have 52 (2n = 4x = 52) chromosomes in their karyotype, while male plants are triploid and have 39 chromosomes (2n = 3x = 39) [119]. Sex markers have been created for betel plants, mainly to accompany the breeding process, which is currently not very developed due to the complexity in the biology of the culture [30,120,121].

2.1.6. Yams

Yams (*Dioscorea rotundata*) are one of the most important crops in a number of countries, primarily in the African continent, where Nigeria, Ghana, Côte d'Ivoire, and Benin are considered leaders in the production of yams [46]. The plant is grown for the production of tubers, which are then used for food. Normally, *D. rotundata* is a dioecious species; however, cases of monoecious occurrence have been observed. Despite the huge role of this crop in African countries, the selection of this species is difficult, largely due to its dioecious nature and problems with flowering (it is predominantly a vegetatively propagated crop) [122,123]. In addition, a change in the sex of plants under the influence of the environment is repeatedly observed during the growing season, which also complicates the breeding process of this crop [124]. The identification of sex chromosomes in yams is also difficult [125]. However, using modern sequencing techniques, the ZZ/ZW sex determination system has been demonstrated [125]. In 2020, the first yam genome obtained as long reads using the Oxford Nanopore Technologies technique was presented [126].

A number of studies have shown that the yam sex determination system is apparently multigenetic and probably still evolving [122,126–128].

2.2. Sex Chromosomes and Sex Determination of Perennial Tree Crops Grown for Fruits

Most dioecious species are perennial tree crops [129]. One of the differences between perennial tree crops and annual and perennial herbaceous crops is a long juvenile period—the reproductive stage often only occurs by the age of 5–8. This feature causes difficulties in growing such plants, since most perennial tree crops are cultivated for the fruits and seeds that appear from the ovary located on the female plants. Revealing the mechanisms of sex determination and the early diagnosis of the sex of such crops are critical tasks for production and breeding. The accumulation of repetitive DNA sequences in sex chromosomes may contribute to the development of sex-associated molecular markers, which is especially important for agricultural plants with a long juvenile period. In cultures such as papaya (*Carica papaya* L.), lychee (*Litchi chinensis* Sonn.), rambutan (*Nephelium lappaceum* L.), and longan (*Euphoria longan* Steud.), the sex determination system is complex. There are not only dioecious plants but also hermaphroditic, gynodioecious, and monoecious plants in the population.

Perennial tree crops grown for their vegetative parts are also found. The production of these crops generally does not require knowing the sex of the plant. However, some agronomically important indicators for these crops (the same as those described above for annual plants) may differ depending on the sex [130].

2.2.1. Papaya

Papaya (Carica papaya L.) is one of the most widely studied model plant species with sex chromosomes. The genome and SDR in papaya are the most widely studied among dioecious plants. Many molecular markers have been created to identify the sex of papaya plants, the genome and transcriptome of this species have been sequenced and assembled, sex chromosomes have been observed, and gene dose compensation and sex-related epigenetic modifications have been noted [131–137]. The papaya sex chromosomes were the second to be fully sequenced after the human sex chromosomes [137]. They are evolutionarily young and outwardly practically homomorphic [11,138]. One of the features of papaya is the presence of hermaphroditic plants—in fact, it is a trioecious species, not a dioecious species. There are two types of Y chromosomes in this species. The first type is represented by the Y chromosome, which is found in male plants. The second type is represented by the Yh chromosome, which causes the hermaphroditism of flowers. At the same time, the YY, YYh, and YhYh genotypes are lethal [33,139]. The non-recombining region on the Y chromosome of papaya was formed as a result of two inversions of retrotransposons and is 8.1 Mb, while the size of the analogous region of the X chromosome is 3.5 Mb. All genes in non-recombining regions have been annotated [140]. Wild papaya is generally dioecious but cultivated as a rule of the gynodioecy cultivar [131]. It has been shown that evolutionarily hermaphroditic plants have been fixed for about 4000 years during domestication [141]. In addition to differences in the ease of cultivation (when growing hermaphroditic plants, male pollinating plants that do not produce fruits are not needed) between female and hermaphroditic plants, there is dimorphism in the shape of the fruits. Fruits from hermaphrodite flowers are more elongated and commercial producers prefer them, rejecting the more rounded fruits of female plants [137]. However, plants can change the sex of their flowers during the growing process. Hermaphroditic flowers can stop the development of carpels, turning into functionally male flowers under the influence of environmental factors such as drought or high temperatures [142–144]. Such phenomena lead to economic losses. It is necessary to create varieties that are less susceptible to environmental factors [145].

2.2.2. Vitis

Grapes are another perennial fruit species in which domestication appears to have perpetuated spontaneous monoecy. The wine grape (*Vitis vinifera* L.) is one of the most economically important fruit crops in the world. It is the third most important, after the tomato and potato, among all horticultural crops [146]. The *Vitis* genus includes about 70 species, all of which are dioecious, and only the cultivated grape *V. vinifera* ssp. vinifera is monoecious, although the wild subspecies *V. vinifera* ssp. sylvestris is also dioecious [147]. It has been shown that the emergence of hermaphroditic forms occurred about 6000–8000 years ago at the time of the domestication of wild grapes [5,148,149]. Like papaya, grapes have three sex-determining loci—Y (M) is male, X (F) is female, and Yh (H) is hermaphroditic. Unlike papaya, the HH, HM, or MM genotypes are viable in grapes. Three sex-linked grape genes (ViviPLATZ, VviFSEX, and APRT3) have been described, and the expression of each differs in hermaphroditic plants [149].

2.2.3. Diospyros

Sex reversion from dioecious to monoecious species in *Diospyros* spp. also occurred as a result of a polyploidization event. *Diospyros* is a very large genus of the Ebenaceae family, which includes several hundred species [150]. Some species of persimmons are edible, but *Diospyros kaki* L. is of the greatest economic importance. This species is widely grown in Asia (more than 80% of world production) and Europe (about 10%) for its fruits [151]. It is an autopolyploid species with 2n = 90 or 135 chromosomes (2n = 6x; 2n = 9x; x = 15) [152]. Despite the presence of at least one Y chromosome in the karyotype, this species is usually monoecious or has completely female plants, with rare male flowers [34]. Recent studies have shown that in the dioecious diploid species *D. lotius* L. (a close relative of *D. kaki*), the *OGI* gene located on the Y chromosome is responsible for sex formation. It encodes a small RNA that targets the autosomal *MeGI* gene, regulating anther fertility in a dose-dependent manner. In polyploid species (*D. kaki*, in particular), there is an insertion of a retroelement, named Kali, in the promoter region of the *OGI* gene that prevents the synthesis of small RNA and promotes the development of monoecy. The authors do not exclude that this mutation, which led to monoecy, could also be fixed as a result of artificial selection [19].

2.2.4. Pistachio

Pistachio (*Pistacia vera* L.) is an economically significant crop for Turkey, Iran, the USA, and a number of other countries [46]. It is a dioecious plant species, where females are needed in production but male plants are needed for pollination. There are also monoecious cultivars [153,154]. From the point of view of the evolution of sex and sex chromosomes, this crop is interesting regarding the ZZ/ZW (2n = 30) system of sex chromosomes [155]—that is, the heterogametic sex of the pistachio is female, like birds or some insect species. This system of sex chromosomes is rare in plants. A recent study on sequencing and assembling the genomes of the male and female plants showed that the pistachio W sex chromosome arose from three successive inversions [156].

2.2.5. Hippophae

Sea buckthorn (*Hippophae rhamnoides* L.) is a berry crop of the Elaeagnaceae family. It is a very commercial, extremely polymorphic species with many subspecies (*Hippophae* has about six to seven species in addition to sea buckthorn) [157,158]. All plants of this genus are strictly dioecious. Sea buckthorn berries contain a large number of vitamins and biologically active substances, and they are used in fresh and cooked food. The oil is obtained from the seeds of sea buckthorn, as well as from the pulp of the fruit. It contains linoleic and linolenic acids, carotenes, fatty acids (primarily omega-3, omega-6, omega-9, and palmitoleic acids), and stearic acids. This is a multipurpose plant, the fruits of which are classified as "superfoods" [159–161]. The ability to enter into symbiosis with nitrogenfixing bacteria and resistance to adverse environmental conditions (drought, high and low temperatures, etc.) makes sea buckthorn a promising crop for growing in poor and

polluted soils, as well as for land remediation. Sea buckthorn is most widespread in Asia, including China, India, and Russia. In Europe, it is popular in Germany, Sweden, and Denmark [162]. The sea buckthorn karyotype consists of 24 chromosomes (22 autosomes and a pair of X/Y sex chromosomes) [163]. In the genome (about 2.6 pg in size), researchers have observed a uniquely large number of satellites that may turn out to be good cy-

have observed a uniquely large number of satellites that may turn out to be good cytogenetic markers [163–165]. The development of molecular markers to determine the sex of sea buckthorn in the early stages of cultivation has been quite successful, but our experiments have shown the unstable operation of these markers, which may be due to sea buckthorn's polymorphism [166–168]. Morphologically, male and female plants can only be distinguished by their generative buds after flowering age has been reached. At the same time, mainly female plants are needed in agricultural production. Additionally, breeding is carried out independently for male pollinating plants and female plants [169]. These facts demonstrate the need for a deeper study of the genetic determination of the sex in sea buckthorn.

The closest relatives of sea buckthorn are dioecious plants from the *Shepherdia* Nutt. genus. *Shepherdia* berries are also used as food, and these plants are a promising crop due to their decorative qualities and high contents of biological substances [170,171]. The system of sex chromosomes and sex determination in *Shepherdia* has not been widely studied.

2.2.6. Humulus

Humulus lupulus L. is a dioecious woody vine of the Cannabaceae family that is of great importance in brewing. Hops are grown for the female buds, which contain substances that create the characteristic taste, aroma and bitterness of beer. Due to the popularity of this drink, hops are cultivated on every continent except Antarctica. The hop karyotype consists of 18 autosomes and a pair of sex chromosomes (XX/XY). The Y chromosome is the smallest in the karyotype, which may indicate its degeneration and evolutionary antiquity [172–174]. In commercial cultivation, the female varieties are vegetatively propagated. A very important task is to prevent the pollination of inflorescences, since setting seeds spoils the taste of beer [175]. Hop is a wind-pollinated crop with light pollen, so just one male plant in the area can cause a lot of economic damage [176]. To prevent pollination, all male plants in a nursery and nearby wild populations must be eradicated. As a part of this procedure, molecular genetic markers are used to identify sex in the early stages of ontogenesis [38,177,178]. In hop production, seedless triploid varieties are also cultivated. They are more productive and have a high growth rate [179,180]. However, they are prone to monoecy. A more detailed study of the mechanisms of sex determination in hops could open up opportunities for influencing this trait and contribute to the creation of high-yielding barren varieties.

2.2.7. Date Palm

The date palm (2n = 2x = 32) is one of the oldest dioecious crops. As with other crops grown for fruit, its production primarily requires female date palm plants. Males are required in smaller numbers for pollination (a ratio of 1:20 is used, i.e., one male plant pollinates 20 females). At the same time, the date palm blooms in the 5–6th year of cultivation—that is, it is not possible to morphologically detect the sex of a tree before this age. Many molecular markers have been developed to identify the sex of the date palm in the early stages of ontogenesis. The genetic mechanisms of sex determination in the date palm have been recently identified [181–185]. An association of two genes, presumably associated with sex, was found on chromosome 12 [186]. Moreover, several genome sequences of different date palm varieties have been published [187–190].

2.2.8. Myristica

Nutmeg (*Myristica fragrans* Houtt., 2n = 38 of 44, according to different communications [191]) is a tropical evergreen tree grown in Asia and Africa for its fruits. It is widely used around the world as a spice [192]. The world production of nutmeg is estimated at 139,000 tons [46]. In agricultural production, female plants are needed more. Attempts to create molecular markers have been made to identify sex in the early stages of vegetation [40,193,194]. However, sex chromosomes in *M. fragrans* have not been found.

2.2.9. Actinidia

Kiwifruit (Actinidia chinensis Planch.) is a plant of the Actinidiaceae family. The Actinidia genus includes 50–70 species [195]. Several species of kiwifruit are cultivated. The most popular are A. chinensis and A. deliciosa. A. arguta is grown in New Zealand, the USA, and some European countries, but the acreage occupied by this species is small [196]. A. kolomikta is popular in Russia in personal plots due to its winter hardiness. All species of the genus are dioecious. Morphologically, the flowers look bisexual, but female plants form flowers with sterile pollen while male plants have an underdeveloped and non-functioning pistil [197]. However, some species sometimes have bisexual flowers. Hermaphroditic flowers have been observed in A. arguta, A. chinensis, A. deliciosa, and A. eriantha Benth. [196]. Kiwifruit has only been cultivated for just over 100 years, but it has achieved great popularity around the world, and its production is about 4 million tons per year [46]. Polyploidy is widespread in species of the Actinidia genus [198]. Chromosomal numbers range from 2n = 58 to 2n = 174 (x = 29) [199]. Polyploidy does not appear to affect sex in kiwifruit. Male plants have at least one Y chromosome in the karyotype, and female plants have XX [200,201]. The sex chromosomes are small and homomorphic, with a small SDR [41,202]. The mechanism of sex determination has been widely studied. This mechanism is based on two genes (the Shy Girl gene (SyGI), the dominant suppressor of carpel development, and the Friendly Boy gene (FrBy), expressed in tapetum cells) [15]. Currently, the kiwifruit genome is actively being studied, and genetic maps and molecular markers are being created to aid the breeding of this crop [203–206]. However, hermaphroditism can be determined by additional genetic factors, which creates difficulties for sex determination based on molecular markers [206].

2.2.10. Ilex

Ilex L. is a large genus of dioecious plants in the Aquifoliaceae family with over 500 species [207]. *Ilex* plants are trees or shrubs, mostly found in nature in East Asia and South America. Some species are found in Europe, Africa, and Northern Australia [208]. Ilex species are grown as ornamental crops to decorate gardens or to make traditional Christmas wreaths [209,210]. The most economically significant plant of this genus is a Paraguayan holly (I. paraguariensis A.St.-Hil.; 2n = 40 [211]). It is widely used in a number of countries (mainly Latin America) for the manufacturing of the mate herbal tea drink. According to FAO data, the global production of mate leaves in 2020 exceeded 1,400,000 tons [46]. In this species, sexual dimorphism has been repeatedly shown in the intensity of photosynthesis, leaf area, and higher mortality of female plants. Thus, male plants have a larger leaf area than female plants, and the intensity of photosynthesis is higher in female plants, while the leaves of male plants make the taste of the drink less bitter [130,212–214]. Like Actinidia, I. paraguariensis is functionally dioecious, with morphologically hermaphroditic flowers, and one of the sexes is sterile or abortive [215]. From a genetic point of view, this species is poorly studied. Heteromorphic sex chromosomes have not been identified. Molecular markers for marker-associated selection and sex detection have been developed [216–218].

2.2.11. Chinese Bayberry

Chinese bayberry (*Morella rubra* Lour.) is widely cultivated in China for fruits. It is the only edible species of the Myricaceae family [219]. The chromosome number is 2n = 16, and the heterogametic sex is presumably female (ZW chromosome system) [220]. A comparison of the sequenced genomes of male and female plants revealed a small region (59 kb) on chromosome 8 that is specific to female plants [42]. The differential expression

of some genes, presumably associated with the development of sex in this species, was also shown [221].

2.3. Sex Chromosomes and Sex Determination of Weeds

In addition to dioecious crops, there are dioecious weed and parasitic species that have negative effects on the cultivation of agricultural plants. In addition to the species of sorrel mentioned above, examples of such weeds are *Amaranthus palmeri* S. Wats. and *A. tuberculatus* (Moq.) J.D. Sauer. These weeds cause enormous damage to US agriculture. They have huge seed productivity and a high resistance to herbicides [222]. The study of the sex determination mechanisms, reproduction features, and secondary sexual characteristics of these species may open up an opportunity for crop protection in the future [223].

3. Conclusions

Modern research on sex chromosomes and the mechanisms of sex determination in plants is focused on the identification of sex-determining genes and on the fundamental issues of the occurrence of dioecy, the evolution of sex and sex chromosomes in a separate family or a whole class of plants. As a rule, a rather limited list of model objects is studied, and the study itself (even if carried out using modern methods and omics technologies affecting the entire plant genome/transcriptome or proteome) focuses on differences in the generative sphere in male and female plants. At the same time, a significant proportion of dioecious plants are economically significant species that have been cultivated by humans for many centuries. It should be noted that their features are linked with their cultivation (Figure 1).



Figure 1. The use of dioecious plants depending on its sex.

According to Figure 1, dioeciousness has a greater or lesser effect on crop cultivation. It is most noticeable on fruit trees. Gardeners strive to grow as many female plants as possible due to the fact that females produce the target product. However, male plants are necessary for pollination and cannot be completely abandoned. However, the maximum effective sex ratio differs depending on the plant and the type of pollination (insect pollination or wind pollination). For example, sea buckthorn shows the best yield in a planting pattern where one mixed row of male and female plants falls between two rows of female plants (in a mixed row, male plants comprise one in five plants) [160]. The role of the pollinator in the formation of economically valuable traits cannot be directly assessed. In addition to dioecy, a long juvenile period also affects the breeding of tree crops. Half of the population during seed propagation comprises male plants. This fact leads to the loss of labor, time, and other resources that are spent on growing males before flowering. Therefore, sex

Additionally, the use of monoecious and parthenocarpic forms may be relevant in such cases. At the same time, male plants are only used in the breeding of some other plants, such as hops. The pollination of industrial plantings is unacceptable. The spatial isolation of hop plantings and the early complete culling of male plants is necessary. In plants in which the vegetative part is used, the role of dioecy is not so obvious and it seems that plants of any sex can be used. However, the different maturation times of male and female plants, as in hemp, or the different yields of male and female plants, as in asparagus, can shift the interest of producers to monoecious forms or plants of a specific sex. Thus, researchers should consider the final purpose of the cultivated crop, as well as its biological characteristics in general, not only focusing on the genes for the formation of pistils or stamens. Humans have a significant impact on the entire environment, and dioecious cultures are no exception.

On the one hand, dioecious plants have been subjected to the influence of artificial selection for a long time, and many of them have gone through a "bottleneck" in their evolution and have lost some of their genotypes. On the other hand, differences between males and females are often not limited to differences in flower structure. Unlike animals, secondary sexual characteristics in plants are usually not expressed; however, individual reports demonstrate their existence. Evolutionarily, this is explained by the different reproductive costs of male and female plants. Females generally require more carbon to produce fruits and seeds, while males (especially wind-pollinated plants) need to produce larger amounts of pollen. This determines differences in not only the type of flowers but also reactions to stress, biomass, productivity, and other traits [224–228]. The significance of a number of dioecious plants for humans indicates the relevance of their in-depth study, and the economic efficiency of growing plants of a certain sex should be evaluated in accordance with the goals of growing such a crop.

Theoretically, it is also interesting to evaluate the means of possible evolution in cultivated dioecious plants (to monoecy, in particular). It is possible that humans will continue to have significant impacts on the rate and direction of evolution for many species. A deeper understanding of the evolution and sex determination of cultivated dioecious plants, together with the characteristics of cultivation, can open up new opportunities for agricultural management.

Author Contributions: Conceptualization, O.V.R. and M.G.D.; validation, O.V.R., O.S.A. and M.G.D.; resources, O.V.R. and O.S.A.; data curation, O.V.R., K.D.B. and O.S.A.; writing—original draft preparation, O.V.R. and K.D.B.; writing—review and editing, all authors; supervision, G.I.K.; funding acquisition, O.V.R. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Russian Foundation for Basic Research, grant 20-316-70018 and 20-016-00145.

Data Availability Statement: No new data were created or analyzed in this study. Data sharing is not applicable for this article.

Acknowledgments: We would like to thank the Russian Foundation for Basic Research for research funding.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Renner, S.S. The relative and absolute frequencies of angiosperm sexual systems: Dioecy, monoecy, gynodioecy, and an updated online database. *Am. J. Bot.* **2014**, *101*, 1588–1596. [CrossRef] [PubMed]
- 2. Bachtrog, D.; Mank, J.E.; Peichel, C.L.; Kirkpatrick, M.; Otto, S.P.; Ashman, T.-L.; Hahn, M.W.; Kitano, J.; Mayrose, I.; Ming, R.; et al. Sex Determination: Why So Many Ways of Doing It? *PLoS Biol.* **2014**, *12*, e1001899. [CrossRef] [PubMed]
- 3. Vyskot, B.; Hobza, R. The genomics of plant sex chromosomes. *Plant Sci.* 2015, 236, 126–135. [CrossRef] [PubMed]
- Montalvão, A.P.L.; Kersten, B.; Fladung, M.; Müller, N.A. The Diversity and Dynamics of Sex Determination in Dioecious Plants. Front. Plant Sci. 2021, 11, 580488. [CrossRef] [PubMed]

- 5. Picq, S.; Santoni, S.; Lacombe, T.; Latreille, M.; Weber, A.; Ardisson, M.; Ivorra, S.; Maghradze, D.; Arroyo-Garcia, R.; Chatelet, P.; et al. A small XY chromosomal region explains sex determination in wild dioecious V. vinifera and the reversal to hermaphroditism in domesticated grapevines. *BMC Plant Biol.* **2014**, *14*, 229. [CrossRef]
- 6. Schaefer, H.; Renner, S.S. A three-genome phylogeny of *Momordica* (Cucurbitaceae) suggests seven returns from dioecy to monoecy and recent long-distance dispersal to Asia. *Mol. Phylogenetics Evol.* **2010**, *54*, 553–560. [CrossRef]
- Weiblen, G.D. Phylogenetic relationships of functionally dioecious *FICUS* (Moraceae) based on ribosomal DNA sequences and morphology. *Am. J. Bot.* 2000, *87*, 1342–1357. [CrossRef]
- 8. Testolin, R.; Pilkington, S.M.; Akagi, T. Editorial: Dioecy in Fruit Crops: The Gender Rise and Decline and Its Agronomic Impact. *Front. Plant Sci.* **2021**, *12*, 719588. [CrossRef]
- 9. Renner, S.S.; Ricklefs, R.E. Dioecy and its correlates in the flowering plants. Am. J. Bot. 1995, 82, 596–606. [CrossRef]
- 10. Charlesworth, B.; Charlesworth, D. A Model for the Evolution of Dioecy and Gynodioecy. Am. Nat. 1978, 112, 975–997. [CrossRef]
- 11. Charlesworth, D. Young sex chromosomes in plants and animals. *New Phytol.* **2019**, 224, 1095–1107. [CrossRef]
- 12. Harkess, A.; Zhou, J.; Xu, C.; Bowers, J.E.; Van der Hulst, R.; Ayyampalayam, S.; Mercati, F.; Riccardi, P.; McKain, M.R.; Kakrana, A.; et al. The asparagus genome sheds light on the origin and evolution of a young Y chromosome. *Nat. Commun.* **2017**, *8*, 1–10. [CrossRef] [PubMed]
- Harkess, A.; Huang, K.; van der Hulst, R.; Tissen, B.; Caplan, J.L.; Koppula, A.; Batish, M.; Meyers, B.C.; Leebens-Mack, J. Sex Determination by Two Y-Linked Genes in Garden Asparagus. *Plant Cell* 2020, *32*, 1790–1796. [CrossRef] [PubMed]
- 14. Henry, I.M.; Akagi, T.; Tao, R.; Comai, L. One Hundred Ways to Invent the Sexes: Theoretical and Observed Paths to Dioecy in Plants. *Annu. Rev. Plant Biol.* **2018**, *69*, 553–575. [CrossRef] [PubMed]
- 15. Akagi, T.; Henry, I.M.; Ohtani, H.; Morimoto, T.; Beppu, K.; Kataoka, I.; Tao, R. A Y-Encoded Suppressor of Feminization Arose via Lineage-Specific Duplication of a Cytokinin Response Regulator in Kiwifruit. *Plant Cell* **2018**, *30*, 780–795. [CrossRef] [PubMed]
- 16. Akagi, T.; Pilkington, S.M.; Varkonyi-Gasic, E.; Henry, I.M.; Sugano, S.S.; Sonoda, M.; Firl, A.; Mcneilage, M.A.; Douglas, M.J.; Wang, T.; et al. Two Y-chromosome-encoded genes determine sex in kiwifruit. *Nat. Plants* **2019**, *5*, 801–809. [CrossRef]
- 17. Li, H.; Wang, L.; Sun, P.; Suo, Y.; Han, W.; Mai, Y.; Diao, S.; Yuan, D.; Fu, J. Cytomorphological observation on development of pistil and stamen of male and hermaphrodite floral buds of diospyros kaki'Longyan Yeshi 1'. *Acta Hortic. Sin.* **2019**, *46*, 1897–1906.
- 18. Prentout, D.; Razumova, O.; Rhoné, B.; Badouin, H.; Henri, H.; Feng, C.; Käfer, J.; Karlov, G.; Marais, G.A. An efficient RNA-seq-based segregation analysis identifies the sex chromosomes of *Cannabis sativa*. *Genome Res.* **2020**, *30*, 164–172. [CrossRef]
- Akagi, T.; Henry, I.M.; Kawai, T.; Comai, L.; Tao, R. Epigenetic Regulation of the Sex Determination Gene *MeGI* in Polyploid Persimmon. *Plant Cell* 2016, 28, 2905–2915. [CrossRef]
- 20. Royal Botanic Gardens, Kew. Available online: https://cvalues.science.kew.org/search/angiosperm (accessed on 25 October 2022).
- Kubis, S.; Schmidt, T.; Heslop-Harrison, J.S. Repetitive DNA Elements as a Major Component of Plant Genomes. Ann. Bot. 1998, 82, 45–55. [CrossRef]
- Macas, J.; Novák, P.; Pellicer, J.; Čížková, J.; Koblížková, A.; Neumann, P.; Fuková, I.; Doležel, J.; Kelly, L.J.; Leitch, I.J. In Depth Characterization of Repetitive DNA in 23 Plant Genomes Reveals Sources of Genome Size Variation in the Legume Tribe *Fabeae*. *PLoS ONE* 2015, 10, e0143424. [CrossRef] [PubMed]
- Błocka-Wandas, M.; Sliwinska, E.; Grabowska-Joachimiak, A.; Musial, K.; Joachimiak, A.J. Male gametophyte development and two different DNA classes of pollen grains in *Rumex acetosa* L., a plant with an XX/XY1Y2 sex chromosome system and a female-biased sex ratio. *Sex. Plant Reprod.* 2007, 20, 171–180. [CrossRef]
- 24. Grabowska-Joachimiak, A.; Śliwińska, E.; Piguła, M.; Skomra, U.; Joachimiak, A.J. Genome size in Humulus lupulus L. and H. japonicus Siebold and Zucc. (*Cannabaceae*). Acta Soc. Bot. Pol. **2011**, 75, 207–214. [CrossRef]
- Carey, S.B.; Lovell, J.T.; Jenkins, J.; Leebens-Mack, J.; Schmutz, J.; Wilson, M.A.; Harkess, A. Representing sex chromosomes in genome assemblies. *Cell Genom.* 2022, 2, 100132. [CrossRef] [PubMed]
- Ahmad, N.; Tian, R.; Li, G.; Zhao, C.; Fan, S.; Sun, J.; Zhao, S.; Wang, X. Establishment of male-specific sequence-tagged site markers in *Asparagus officinalis*: An efficient tool for sex identification. *Plant Breed.* 2022, 141, 471–481. [CrossRef]
- 27. Korpelainen, H. A genetic method to resolve gender complements investigations on sex ratios in *Rumex acetosa*. *Mol. Ecol.* **2002**, *11*, 2151–2156. [CrossRef]
- She, H.; Xu, Z.; Zhang, H.; Li, G.; Wu, J.; Wang, X.; Li, Y.; Liu, Z.; Qian, W. Identification of a male-specific region (MSR) in *Spinacia* oleracea. Hortic. Plant J. 2021, 7, 341–346. [CrossRef]
- 29. Mandolino, G.; Carboni, A.; Forapani, S.; Faeti, V.; Ranalli, P. Identification of DNA markers linked to the male sex in dioecious hemp (*Cannabis sativa* L.). *Theor. Appl. Genet.* **1999**, *98*, 86–92. [CrossRef]
- 30. Samantaray, S.; Phurailatpam, A.; Bishoyi, A.K.; Geetha, K.A.; Maiti, S. Identification of sex-specific DNA markers in betel vine (*Piper betle L.*). *Genet. Resour. Crop. Evol.* **2011**, *59*, 645–653. [CrossRef]
- 31. Mondo, J.M.; Agre, P.A.; Asiedu, R.; Akoroda, M.O.; Asfaw, A. Genome-Wide Association Studies for Sex Determination and Cross-Compatibility in Water Yam (*Dioscorea alata* L.). *Plants* **2021**, *10*, 1412. [CrossRef]
- Deputy, J.; Ming, R.; Ma, H.; Liu, Z.; Fitch, M.; Wang, M.; Manshardt, R.; Stiles, J. Molecular markers for sex determination in papaya (*Carica papaya* L.). *Theor. Appl. Genet.* 2002, 106, 107–111. [CrossRef] [PubMed]
- 33. Ming, R.; Yu, Q.; Moore, P.H. Sex determination in papaya. Semin. Cell Dev. Biol. 2007, 18, 401–408. [CrossRef] [PubMed]

- Akagi, T.; Kajita, K.; Kibe, T.; Morimura, H.; Tsujimoto, T.; Nishiyama, S.; Kawai, T.; Yamane, H.; Tao, R. Development of molecular markers associated with sexuality in *Diospyros lotus* L. and their application in *D. kaki* Thunb. *J. Jpn. Soc. Hortic. Sci.* 2013, *83*, 214–221. [CrossRef]
- Khodaeiaminjan, M.; Kafkas, E.; Güney, M.; Kafkas, S. Development and linkage mapping of novel sex-linked markers for marker-assisted cultivar breeding in pistachio (*Pistacia vera* L.). *Mol. Breed.* 2017, 37, 1–9. [CrossRef]
- Kafkas, S.; Khodaeiaminjan, M.; Güney, M.; Kafkas, E. Identification of sex-linked SNP markers using RAD sequencing suggests ZW/ZZ sex determination in *Pistacia vera* L. *BMC Genom.* 2015, 16, 98. [CrossRef] [PubMed]
- Korekar, G.; Sharma, R.K.; Kumar, R.; Meenu; Bisht, N.C.; Srivastava, R.B.; Ahuja, P.S.; Stobdan, T. Identification and validation of sex-linked SCAR markers in dioecious *Hippophae rhamnoides* L. (Elaeagnaceae). *Biotechnol. Lett.* 2012, 34, 973–978. [CrossRef]
- 38. Danilova, T.V.; Karlov, G.I. Application of inter simple sequence repeat (ISSR) polymorphism for detection of sex-specific molecular markers in hop (*Humulus lupulus* L.). *Euphytica* **2006**, *151*, 15–21. [CrossRef]
- Amjad, Z.; Khan, Z.; Shan, M.; Khanum, P.; Saeed, A.; Akhtar, G. Validation of PCR-based Markers Associated with Sex Determination in Date Palm (*Phoenix dactylifera* L.). *Pak. J. Biochem. Biotechnol.* 2022, *3*, 26–33. [CrossRef]
- 40. Shaanker, U. Identification of sex-specific DNA markers in the dioecious tree, nutmeg (*Myristica fragrans* Houtt.). Noticiario de Recursos Fitogenéticos 2000.
- Fraser, L.G.; Tsang, G.K.; Datson, P.M.; De Silva, H.N.; Harvey, C.F.; Gill, G.P.; Crowhurst, R.N.; Mcneilage, M.A. A gene-rich linkage map in the dioecious species *Actinidia chinensis* (kiwifruit) reveals putative X/Y sex-determining chromosomes. *BMC Genom.* 2009, 10, 102. [CrossRef]
- Wang, Y.; Jia, H.-M.; Shen, Y.-T.; Zhao, H.-B.; Yang, Q.-S.; Zhu, C.-Q.; Sun, D.-L.; Wang, G.-Y.; Zhou, C.-C.; Jiao, Y.; et al. Construction of an anchoring SSR marker genetic linkage map and detection of a sex-linked region in two dioecious populations of red bayberry. *Hortic. Res.* 2020, 7, 1–9. [CrossRef] [PubMed]
- Montgomery, J.S.; Sadeque, A.; Giacomini, D.A.; Brown, P.J.; Tranel, P.J. Sex-specific markers for waterhemp (*Amaranthus tuberculatus*) and Palmer amaranth (*Amaranthus palmeri*). Weed Sci. 2019, 67, 412–418. [CrossRef]
- Uragami, A.; Ueno, R.; Yamasaki, A.; Matsuo, K.; Yamaguchi, T.; Tokiwa, H.; Takizawa, T.; Sakai, H.; Ikeuchi, T.; Watanabe, S.-I.; et al. Productive Differences between Male and Female Plants in White Asparagus Production Using the Rootstock-planting Forcing Culture Technique. *Hortic. J.* 2016, *85*, 322–330. [CrossRef]
- Norup, M.F.; Petersen, G.; Burrows, S.; Bouchenak-Khelladi, Y.; Leebens-Mack, J.; Pires, J.C.; Linder, H.P.; Seberg, O. Evolution of *Asparagus* L. (Asparagaceae): Out-of-South-Africa and multiple origins of sexual dimorphism. *Mol. Phylogenetics Evol.* 2015, 92, 25–44. [CrossRef] [PubMed]
- 46. Food and Agriculture Organization of the United Nations. Available online: https://www.fao.org/faostat/en/#data/QCL/visualize (accessed on 25 October 2022).
- 47. Flory, W.S. Genetic and cytological investigations on Asparagus officinalis L. Genetics 1932, 17, 432–467. [CrossRef]
- 48. Deng, C.-L.; Qin, R.-Y.; Wang, N.-N.; Cao, Y.; Gao, J.; Gao, W.-J.; Lu, L.-D. Karyotype of asparagus by physical mapping of 45S and 5S rDNA by FISH. *J. Genet.* 2012, *91*, 209–212. [CrossRef]
- Löptien, H. Giemsa-Banden auf Mitosechromosomen des Spargels (*Asparagus officinalis* L.) und des Spinats (*Spinacia oleracea* L.). Z Pflanz. 1976, 76, 225–230.
- 50. Ming, R.; Bendahmane, A.; Renner, S.S. Sex Chromosomes in Land Plants. Annu. Rev. Plant Biol. 2011, 62, 485–514. [CrossRef]
- Harkess, A.; Mercati, F.; Shan, H.; Sunseri, F.; Falavigna, A.; Leebens-Mack, J. Sex-biased gene expression in dioecious garden asparagus (*Asparagus officinalis*). New Phytol. 2015, 207, 883–892. [CrossRef]
- 52. Li, S.-F.; Lv, C.-C.; Lan, L.-N.; Jiang, K.-L.; Zhang, Y.-L.; Li, N.; Deng, C.-L.; Gao, W.-J. DNA methylation is involved in sexual differentiation and sex chromosome evolution in the dioecious plant garden asparagus. *Hortic. Res.* **2021**, *8*, 198. [CrossRef]
- 53. Sneep, J. The significance of andromonoecism for the breeding of *Asparagus officinalis* L. II. *Euphytica* 1953, *2*, 224–228. [CrossRef]
 54. Franken, A.A. Sex characteristics and inheritance of sex in asparagus (*Asparagus officinalis* L.). *Euphytica* 1970, *19*, 277–287.
- [CrossRef]
 55. Falloon, P.; Nikoloff, A. Asparagus: Value of individual plant yield and fern characteristics as selection criteria. *New Zealand J. Exp. Agric.* 1986, 14, 417–420. [CrossRef]
- 56. Moreno-Pinel, R.; Castro-López, P.; Die-Ramón, J.V.; Gil-Ligero, J. Asparagus (*Asparagus officinalis* L.) Breeding. In Advances in Plant Breeding Strategies: Vegetable Crops; Springer: Berlin/Heidelberg, Germany, 2021; pp. 425–469. [CrossRef]
- 57. Jain, S.M.; Al-Khayri, J.M.; Johnson, D.V. Advances in Plant Breeding Strategies: Vegetable Crops: Volume 10: Leaves, Flowerheads, Green Pods, Mushrooms and Truffles; Springer: Berlin/Heidelberg, Germany, 2021.
- 58. Ii, Y.; Uragami, A.; Uno, Y.; Kanechi, M.; Inagaki, N. RAPD-based analysis of differences between male and female genotypes of *Asparagus officinalis*. *Hortic. Sci.* **2012**, *39*, 33–37. [CrossRef]
- 59. Bracale, M.; Caporali, E.; Galli, M.; Longo, C.; Marziani-Longo, G.; Rossi, G.; Spada, A.; Soave, C.; Falavigna, A.; Raffaldi, F.; et al. Sex determination and differentiation in *Asparagus officinalis* L. *Plant Sci.* **1991**, *80*, 67–77. [CrossRef]
- 60. Reamon-Büttner, S.M.; Jung, C. AFLP-derived STS markers for the identification of sex in *Asparagus officinalis* L. *Theor. Appl. Genet.* **2000**, 100, 432–438. [CrossRef]
- 61. Mercati, F.; Riccardi, P.; Leebens-Mack, J.; Abenavoli, M.R.; Falavigna, A.; Sunseri, F. Single nucleotide polymorphism isolated from a novel EST dataset in garden asparagus (*Asparagus officinalis* L.). *Plant Sci.* **2013**, 203–204, 115–123. [CrossRef]

- 62. Stone, N.K.; Thomas, Z.M.; Roose, M.L. A new robust codominant sex-linked STS marker for asparagus. *Acta Hortic.* 2018, 1223, 51–58. [CrossRef]
- Vasas, A.; Orbán-Gyapai, O.; Hohmann, J. The Genus Rumex: Review of traditional uses, phytochemistry and pharmacology. J. Ethnopharmacol. 2015, 175, 198–228. [CrossRef]
- 64. Schuster, T.M.; Reveal, J.L.; Bayly, M.J.; Kron, K.A. An updated molecular phylogeny of *Polygonoideae* (Polygonaceae): Relationships of *Oxygonum, Pteroxygonum*, and *Rumex*, and a new circumscription of *Koenigia*. *Taxon* **2015**, *64*, 1188–1208. [CrossRef]
- Yazdi, S.A.F.; Rezvani, M.; Mohassel, M.H.R.; Ghanizadeh, H. Factors affecting seed germination and seedling emergence of sheep sorrel (*Rumex acetosella*). *Rom Agric Res* 2013, *30*, 373–380.
- 66. Kennedy, K.J.; Boyd, N.S.; Nams, V.O. Hexazinone and Fertilizer Impacts on Sheep Sorrel (*Rumex acetosella*) in Wild Blueberry. *Weed Sci.* 2010, *58*, 317–322. [CrossRef]
- 67. Kennedy, K.J.; Boyd, N.S.; Nams, V.O.; Olson, A.R. The Impacts of Fertilizer and Hexazinone on Sheep Sorrel (*Rumex acetosella*) Growth Patterns in Lowbush Blueberry Fields. *Weed Sci.* 2011, *59*, 335–340. [CrossRef]
- Talovina, G.V.; Smekalova, T.N. Distribution of wild relatives of cultivated sorrel (*Rumex acetosa* L.) over russia and aspects of their in situ conservation. *Vavilovia* 2018, 1, 33–39. [CrossRef]
- 69. Kukusheva, A.; Stepanov, A. Effect of mowing term on biometrics, yield and nutritional properties of hybrid (*Rumex patientia* × *Rumex tianschanicus*). *Bulg. J. Agric. Sci.* **2016**, 22, 948–954.
- Feduraev, P.; Skrypnik, L.; Nebreeva, S.; Dzhobadze, G.; Vatagina, A.; Kalinina, E.; Pungin, A.; Maslennikov, P.; Riabova, A.; Krol, O.; et al. Variability of Phenolic Compound Accumulation and Antioxidant Activity in Wild Plants of Some *Rumex* Species (*Polygonaceae*). *Antioxidants* 2022, 11, 311. [CrossRef]
- 71. Yıldırım, A.; Mavi, A.; Kara, A.A. Determination of Antioxidant and Antimicrobial Activities of *Rumex crispus* L. Extracts. *J. Agric. Food Chem.* **2001**, *49*, 4083–4089. [CrossRef]
- Cuñado, N.; Navajas-Pérez, R.; de la Herrán, R.; Rejón, C.R.; Rejón, M.R.; Santos, J.L.; Garrido-Ramos, M.A. The evolution of sex chromosomes in the genus *Rumex* (Polygonaceae): Identification of a new species with heteromorphic sex chromosomes. *Chromosom. Res.* 2007, 15, 825–833. [CrossRef]
- 73. Mariotti, B.; Manzano, S.; Kejnovský, E.; Vyskot, B.; Jamilena, M. Accumulation of Y-specific satellite DNAs during the evolution of *Rumex acetosa* sex chromosomes. *Mol. Genet. Genom.* **2008**, *281*, 249–259. [CrossRef]
- 74. Rejón, C.R.; Jamilena, M.; Ramos, M.G.; Parker, J.S. Cytogenetic and molecular analysis of the multiple sex chromosome system of Rumex acetosa. *Heredity* **1994**, *72*, 209–215. [CrossRef]
- Takenaka, Y. On the Special Autosomes with Reference to the Sex-determination of *Rumex acetosa* L. *Cytologia* 1937, 2, 995–1002.
 [CrossRef]
- 76. Kihara, H.; Ono, T. The sex-chromosomes of rumex acetosa. Mol. Genet. Genom. 1925, 39, 1–7. [CrossRef]
- 77. Wilby, A.S.; Parker, J.S. Recurrent patterns of chromosome variation in a species group. Heredity 1988, 61, 55–62. [CrossRef]
- 78. Shibata, F.; Hizume, M.; Kuroki, Y. Chromosome painting of Y chromosomes and isolation of a Y chromosome-specific repetitive sequence in the dioecious plant *Rumex acetosa*. *Chromosoma* **1999**, *108*, 266–270. [CrossRef]
- Shibata, F.; Hizume, M.; Kuroki, Y. Molecular cytogenetic analysis of supernumerary heterochromatic segments in *Rumex acetosa*. *Genome* 2000, 43, 391–397. [CrossRef]
- Steflova, P.; Tokan, V.; Vogel, I.; Lexa, M.; Macas, J.; Novak, P.; Hobza, R.; Vyskot, B.; Kejnovsky, E. Contrasting Patterns of Transposable Element and Satellite Distribution on Sex Chromosomes (XY1Y2) in the Dioecious Plant *Rumex acetosa*. *Genome Biol. Evol.* 2013, 5, 769–782. [CrossRef]
- 81. Bürli, S.; Pannell, J.R.; Tonnabel, J. Environmental variation in sex ratios and sexual dimorphism in three wind-pollinated dioecious plant species. *Oikos* 2022, 2022, e08651. [CrossRef]
- 82. Liu, Z.; She, H.; Xu, Z.; Zhang, H.; Li, G.; Zhang, S.; Qian, W. Quantitative trait loci (QTL) analysis of leaf related traits in spinach (Spinacia oleracea L.). *BMC Plant Biol.* **2021**, *21*, 1–14. [CrossRef]
- Hammer, K. Chenopodiaceae, Spinacia oleracea L. In Mansfeld's Encyclopedia of Agricultural and Horticultural Crops (Except Ornamentals), 1st English ed.; Hanelt, P., Institute of Plant Genetics and Crop Plant Research Berlin, Eds.; Springer: Berlin/Heidelberg, Germany; New York, NY, USA, 2001; pp. 249–251.
- 84. Stevenson, E.C. A genetic study of the heterogametic nature of the staminate plant in spinach (*Spinacia oleracea* L.). *Proc. Am. Soc. Hortic. Sci.* **1954**, *63*, 444–446.
- 85. Janick, J.; Stevenson, E.C. The effects of polyploidy on sex expression in Spinach. J. Hered. 1955, 46, 151–156. [CrossRef]
- 86. Janick, J.; Mahoney, D.L.; Pfahler, P.L. The trisomics of Spinacia oleracea. J. Hered. 1959, 50, 47-50. [CrossRef]
- 87. Khattak, J.Z.K.; Torp, A.M.; Andersen, S.B. A genetic linkage map of *Spinacia oleracea* and localization of a sex determination locus. *Euphytica* **2006**, *148*, 311–318. [CrossRef]
- Wang, H.; Liu, Z.; Wang, X.; Wu, J.; Zhang, H.; Xia, Z.; Qian, W. Genome-wide association studies for monoecism in spinach. *Acta Hortic. Sin.* 2019, 46, 1495–1502. [CrossRef]
- 89. She, H.; Liu, Z.; Xu, Z.; Zhang, H.; Cheng, F.; Wang, X.; Qian, W. The female (XX) and male (YY) genomes provide insights into the sex determination mechanism in spinach. *bioRxiv* 2020. [CrossRef]
- Vitale, J.J.; Freeman, D.C. Secondary sex characteristics in *Spinacia oleracea* L.: Quantitative evidence for the existence of at least three sexual morphs. *Am. J. Bot.* 1985, 72, 1061–1066. [CrossRef]

- Golubkina, N.A.; Kosheleva, O.V.; Krivenkov, L.V.; Dobrutskaya, H.G.; Nadezhkin, S.; Caruso, G. Intersexual differences in plant growth, yield, mineral composition and antioxidants of spinach (*Spinacia oleracea* L.) as affected by selenium form. *Sci. Hortic.* 2017, 225, 350–358. [CrossRef]
- 92. Pérez-Llorca, M.; Vilas, J.S. Sexual dimorphism in response to herbivory and competition in the dioecious herb *Spinacia oleracea*. *Plant Ecol.* **2019**, 220, 57–68. [CrossRef]
- 93. Divashuk, M.G.; Alexandrov, O.S.; Razumova, O.V.; Kirov, I.V.; Karlov, G.I. Molecular Cytogenetic Characterization of the Dioecious Cannabis sativa with an XY Chromosome Sex Determination System. *PLoS ONE* **2014**, *9*, e85118. [CrossRef]
- Van Bakel, H.; Stout, J.M.; Cote, A.G.; Tallon, C.M.; Sharpe, A.G.; Hughes, T.R.; Page, J.E. The draft genome and transcriptome of *Cannabis sativa. Genome Biol.* 2011, 12, R102. [CrossRef]
- Grassa, C.J.; Weiblen, G.D.; Wenger, J.P.; Dabney, C.; Poplawski, S.G.; Motley, S.T.; Michael, T.P.; Schwartz, C.J. A new *Cannabis* genome assembly associates elevated cannabidiol (CBD) with hemp introgressed into marijuana. *New Phytol.* 2021, 230, 1665–1679. [CrossRef]
- Laverty, K.U.; Stout, J.M.; Sullivan, M.J.; Shah, H.; Gill, N.; Holbrook, L.; Deikus, G.; Sebra, R.; Hughes, T.R.; Page, J.E.; et al. A physical and genetic map of *Cannabis sativa* identifies extensive rearrangements at the *THC/CBD acid synthase* loci. *Genome Res.* 2018, 29, 146–156. [CrossRef] [PubMed]
- 97. Gao, S.; Wang, B.; Xie, S.; Xu, X.; Zhang, J.; Pei, L.; Yu, Y.; Yang, W.; Zhang, Y. A high-quality reference genome of wild *Cannabis* sativa. Hortic. Res. **2020**, *7*, 1–11. [CrossRef] [PubMed]
- 98. Schultes, R.E.; Klein, W.M.; Plowman, T.; Lockwood, T.E. Cannabis: An example of taxonomic neglect. In *Cannabis and Culture*; De Gruyter Mouton: New York, NY, USA, 1975; pp. 21–38. [CrossRef]
- 99. Merlin, M.D. Archaeological evidence for the tradition of psychoactive plant use in the old world. *Econ. Bot.* **2003**, *57*, 295–323. [CrossRef]
- 100. Crini, G.; Lichtfouse, E.; Chanet, G.; Morin-Crini, N. Applications of hemp in textiles, paper industry, insulation and building materials, horticulture, animal nutrition, food and beverages, nutraceuticals, cosmetics and hygiene, medicine, agrochemistry, energy production and environment: A review. *Environ. Chem. Lett.* **2020**, *18*, 1451–1476. [CrossRef]
- 101. Sorrentino, G. Introduction to emerging industrial applications of cannabis (*Cannabis sativa* L.). *Rendiconti Lince Sci. Fis. Nat.* **2021**, 32, 233–243. [CrossRef]
- 102. Faux, A.-M.; Draye, X.; Lambert, R.; D'Andrimont, R.; Raulier, P.; Bertin, P. The relationship of stem and seed yields to flowering phenology and sex expression in monoecious hemp (*Cannabis sativa* L.). *Eur. J. Agron.* **2013**, 47, 11–22. [CrossRef]
- 103. Baldini, M.; Ferfuia, C.; Piani, B.; Sepulcri, A.; Dorigo, G.; Zuliani, F.; Danuso, F.; Cattivello, C. The Performance and Potentiality of Monoecious Hemp (*Cannabis sativa* L.) Cultivars as a Multipurpose Crop. *Agronomy* **2018**, *8*, 162. [CrossRef]
- Razumova, O.V.; Alexandrov, O.S.; Divashuk, M.G.; Sukhorada, T.I.; Karlov, G.I. Molecular cytogenetic analysis of monoecious hemp (*Cannabis sativa* L.) cultivars reveals its karyotype variations and sex chromosomes constitution. *Protoplasma* 2015, 253, 895–901. [CrossRef]
- 105. Faux, A.-M.; Berhin, A.; Dauguet, N.; Bertin, P. Sex chromosomes and quantitative sex expression in monoecious hemp (*Cannabis sativa* L.). *Euphytica* **2013**, *196*, 183–197. [CrossRef]
- Törjék, O.; Bucherna, N.; Kiss, E.; Homoki, H.; Finta-Korpelová, Z.; Bócsa, I.; Nagy, I.; Heszky, L.E. Novel male-specific molecular markers (MADC5, MADC6) in hemp. *Euphytica* 2002, 127, 209–218. [CrossRef]
- 107. Mandolino, G.; Ranalli, P. The Applications of Molecular Markers in Genetics and Breeding of Hemp. J. Ind. Hemp 2002, 7, 7–23. [CrossRef]
- Moliterni, V.M.; Cattivelli, L.; Ranalli, P.; Mandolino, G. The sexual differentiation of *Cannabis sativa* L.: A morphological and molecular study. *Euphytica* 2004, 140, 95–106. [CrossRef]
- Faux, A.-M.; Draye, X.; Flamand, M.-C.; Occre, A.; Bertin, P. Identification of QTLs for sex expression in dioecious and monoecious hemp (*Cannabis sativa* L.). *Euphytica* 2016, 209, 357–376. [CrossRef]
- Heslop-Harrison, J.; Heslop-Harrison, Y. Studies on Flowering-Plant Growth and Organogenesis: III. Leaf Shape Changes Associated with Flowering and Sex Differentiation in Cannabis Sativa. In *Proceedings of the Royal Irish Academy. Section B: Biological, Geological, and Chemical Science*; Royal Irish Academy: Dublin, Ireland, 1957; Volume 59, pp. 257–283.
- Adal, A.M.; Doshi, K.; Holbrook, L.; Mahmoud, S.S. Comparative RNA-Seq analysis reveals genes associated with masculinization in female Cannabis sativa. *Planta* 2021, 253, 1–17. [CrossRef] [PubMed]
- 112. Guha, P. Betel Leaf: The Neglected Green Gold of India. J. Hum. Ecol. 2006, 19, 87–93. [CrossRef]
- 113. Biswas, P.; Anand, U.; Saha, S.C.; Kant, N.; Mishra, T.; Masih, H.; Bar, A.; Pandey, D.K.; Jha, N.K.; Majumder, M.; et al. Betelvine (*Piper betle* L.): A comprehensive insight into its ethnopharmacology, phytochemistry, and pharmacological, biomedical and therapeutic attributes. *J. Cell. Mol. Med.* **2022**, *26*, 3083–3119. [CrossRef] [PubMed]
- 114. Taukoorah, U.; Lall, N.; Mahomoodally, F. *Piper betle* L. (betel quid) shows bacteriostatic, additive, and synergistic antimicrobial action when combined with conventional antibiotics. *S. Afr. J. Bot.* **2016**, *105*, 133–140. [CrossRef]
- 115. Gundala, S.R.; Yang, C.; Mukkavilli, R.; Paranjpe, R.; Brahmbhatt, M.; Pannu, V.; Cheng, A.; Reid, M.D.; Aneja, R. Hydroxychavicol, a betel leaf component, inhibits prostate cancer through ROS-driven DNA damage and apoptosis. *Toxicol. Appl. Pharmacol.* 2014, 280, 86–96. [CrossRef]
- 116. Bari, S.; Khandokar, L.; Haque, E.; Romano, B.; Capasso, R.; Seidel, V.; Haque, A.; Rashid, M.A. Ethnomedicinal uses, phytochemistry, and biological activities of plants of the genus *Gynura*. *J. Ethnopharmacol.* **2021**, 271, 113834. [CrossRef]

- 117. Madhumita, M.; Guha, P.; Nag, A. Processing and Potential Health Benefits of Betel Leaf (*Piper betle* L.). In *Herbal Medicine in India*; Springer: Singapore, 2020; pp. 237–246. [CrossRef]
- 118. Verma, A.; Kumar, N.; Ranade, S.A. Genetic diversity amongst landraces of a dioecious vegetatively propagated plant, betelvine (*Piper betle* L.). *J. Biosci.* **2004**, *29*, 319–328. [CrossRef]
- 119. Phurailatpam, A.K.; Geetha, K.A.; Maiti, S. Ploidy distinction in male and female plants of betelvine (*Piper betle* L.): A study by flow cytometry. *Genet. Resour. Crop. Evol.* **2018**, *65*, 1565–1570. [CrossRef]
- 120. Das, S.; Parida, R.; Sandeep, I.S.; Nayak, S.; Mohanty, S. Biotechnological intervention in betelvine (*Piper betle* L.): A review on recent advances and future prospects. *Asian Pac. J. Trop. Med.* **2016**, *9*, 938–946. [CrossRef]
- 121. Sheeja, T.E.; Bindu, K.H.; Anto, P.; Dhanya, K.; Siju, S.; Kumar, T.V. A SCAR marker based method for sex determination in dioecious betel vine (*Piper betle*). *Ind. J. Agric. Sci.* 2013, *83*, 1409–1410.
- 122. Denadi, N.; Gandonou, C.; Missihoun, A.A.; Zoundjihékpon, J.; Quinet, M. Plant Sex Prediction Using Genetic Markers in Cultivated Yams (*Dioscorea rotundata* Poir.) in Benin. *Agronomy* **2020**, *10*, 1521. [CrossRef]
- Terauchi, R.; Kahl, G. Sex determination in *Dioscorea tokoro*, a wild yam species. In *Sex Determination in Plants*; Ainsworth, C.C., Ed.; BIOS Scientific Publishers: Oxford, UK, 1999; pp. 163–171.
- 124. Martin, F.W. Sex Ratio and Sex Determination in Dioscorea. J. Hered. 1966, 57, 95–99. [CrossRef]
- 125. Tamiru, M.; Natsume, S.; Takagi, H.; White, B.; Yaegashi, H.; Shimizu, M.; Yoshida, K.; Uemura, A.; Oikawa, K.; Abe, A.; et al. Genome sequencing of the staple food crop white Guinea yam enables the development of a molecular marker for sex determination. *BMC Biol.* 2017, *15*, 1–20. [CrossRef] [PubMed]
- 126. Sugihara, Y.; Darkwa, K.; Yaegashi, H.; Natsume, S.; Shimizu, M.; Abe, A.; Hirabuchi, A.; Ito, K.; Oikawa, K.; Tamiru-Oli, M.; et al. Genome analyses reveal the hybrid origin of the staple crop white Guinea yam (*Dioscorea rotundata*). Proc. Natl. Acad. Sci. USA 2020, 117, 31987–31992. [CrossRef]
- 127. Agre, P.; Nwachukwu, C.; Olasanmi, B.; Obidiegwu, J.; Nwachukwu, E.; Adebola, P.; Dekoeyer, D.; Asrat, A. Sample Preservation and Plant Sex Prediction in White Guinea yam (*Dioscorea rotundata* Poir.). J. Appl. Biotechnol. Rep. 2020, 7, 145–151. [CrossRef]
- 128. Sugihara, Y.; Kudoh, A.; Oli, M.T.; Takagi, H.; Natsume, S.; Shimizu, M.; Abe, A.; Asiedu, R.; Asfaw, A.; Adebola, P.; et al. Population Genomics of Yams: Evolution and Domestication of *Dioscorea* Species. In *Population Genomics*; Springer: Cham, Switzerland, 2021; pp. 1–28. [CrossRef]
- 129. Renner, S.S.; Feil, J.P. Pollinators of tropical dioecious angiosperms. Am. J. Bot. 1993, 80, 1100–1107. [CrossRef]
- 130. Rakocevic, M.; Medrado, M.; Martim, S.; Assad, E. Sexual dimorphism and seasonal changes of leaf gas exchange in the dioecious tree *llex paraguariensis* grown in two contrasted cultivation types. *Ann. Appl. Biol.* **2009**, *154*, 291–301. [CrossRef]
- 131. Zhou, P.; Zhang, X.; Ma, X.; Yue, J.; Liao, Z.; Ming, R. Methylation related genes affect sex differentiation in dioecious and gynodioecious papaya. *Hortic. Res.* 2022, *9*, uhab065. [CrossRef] [PubMed]
- 132. Ming, R.; Hou, S.; Feng, Y.; Yu, Q.; Dionne-Laporte, A.; Saw, J.H.; Senin, P.; Wang, W.; Ly, B.V.; Lewis, K.L.T.; et al. The draft genome of the transgenic tropical fruit tree papaya (*Carica papaya* Linnaeus). *Nature* **2008**, 452, 991–996. [CrossRef] [PubMed]
- Liu, J.; Han, J.; Sharma, A.; Wai, C.M.; Ming, R.; Yu, Q. Transcriptional regulation of dosage compensation in *Carica papaya*. Sci. Rep. 2021, 11, 5854. [CrossRef] [PubMed]
- 134. Lemos, E.G.M.; Silva, C.L.S.P.; Zaidan, H.A. Identification of sex in *Carica papaya* L. using RAPD markers. *Euphytica* 2002, 127, 179–184. [CrossRef]
- 135. Hsu, T.-H.; Gwo, J.-C.; Lin, K.-H. Rapid sex identification of papaya (*Carica papaya*) using multiplex loop-mediated isothermal amplification (mLAMP). *Planta* **2012**, *236*, 1239–1246. [CrossRef]
- 136. Chaves-Bedoya, G.; Nuñez, V. A SCAR marker for the sex types determination in Colombian genotypes of *Carica papaya*. *Euphytica* **2007**, *153*, 215–220. [CrossRef]
- 137. Liao, Z.; Yu, Q.; Ming, R. Development of male-specific markers and identification of sex reversal mutants in papaya. *Euphytica* **2017**, *213*, 1–12. [CrossRef]
- Yu, Q.; Hou, S.; Hobza, R.; Feltus, F.A.; Wang, X.; Jin, W.; Ming, R. Chromosomal location and gene paucity of the male specific region on papaya Y chromosome. *Mol. Genet. Genom.* 2007, 278, 177–185. [CrossRef]
- 139. Urasaki, N.; Tarora, K.; Shudo, A.; Ueno, H.; Tamaki, M.; Miyagi, N.; Adaniya, S.; Matsumura, H. Digital Transcriptome Analysis of Putative Sex-Determination Genes in Papaya (*Carica papaya*). *PLoS ONE* **2012**, *7*, e40904. [CrossRef]
- 140. Wang, J.; Na, J.-K.; Yu, Q.; Gschwend, A.R.; Han, J.; Zeng, F.; Aryal, R.; VanBuren, R.; Murray, J.E.; Zhang, W.; et al. Sequencing papaya X and Y^h chromosomes reveals molecular basis of incipient sex chromosome evolution. *Proc. Natl. Acad. Sci. USA* 2012, 109, 13710–13715. [CrossRef]
- 141. VanBuren, R.; Zeng, F.; Chen, C.; Zhang, J.; Wai, C.M.; Han, J.; Aryal, R.; Gschwend, A.R.; Wang, J.; Na, J.-K.; et al. Origin and domestication of papaya Y^h chromosome. *Genome Res.* 2015, *25*, 524–533. [CrossRef]
- 142. Lin, H.; Liao, Z.; Zhang, L.; Yu, Q. Transcriptome analysis of the male-to-hermaphrodite sex reversal induced by low temperature in papaya. *Tree Genet. Genomes* **2016**, *12*, 1–14. [CrossRef]
- 143. Ramos, H.C.C.; Pereira, M.G.; Da Silva, F.F.; Viana, A.P.; Ferreguetti, G.A. Seasonal and genetic influences on sex expression in a backcrossed segregating papaya population. *Crop. Breed. Appl. Biotechnol.* **2011**, *11*, 97–105. [CrossRef]
- 144. Martelleto, L.A.P.; Ribeiro, R.D.L.D.; Sudo-Martelleto, M.; Vasconcellos, M.A.D.S.; Pereira, M.B. Expressão da esterilidade feminina e da carpeloidia em mamoeiro sob diferentes ambientes de cultivo protegido. *Rev. Bras. Frutic.* 2011, 33, 1185–1193. [CrossRef]

- 145. Moreira, N.F.; Pereira, T.N.S.; Catarina, R.S.; Cortes, D.F.M.; Vettorazzi, J.C.F.; Ramos, H.C.C.; Viana, A.P.; Pereira, M.G. Quantification of floral abnormalities in a population generated from sexual polymorphism aiming at recurrent selection in papaya. *Bragantia* 2019, 78, 158–165. [CrossRef]
- 146. Alston, J.M.; Sambucci, O. Grapes in the World Economy. In *The Grape Genome. Compendium of Plant Genomes*; Cantu, D., Walker, M., Eds.; Springer: Cham, Switzerland, 2019. [CrossRef]
- 147. Moore, M.O. Classification and systematics of eastern North American Vitis L. (Vitaceae) north of Mexico. Sida 1991, 14, 345.
- 148. This, P.; Lacombe, T.; Thomas, M.R. Historical origins and genetic diversity of wine grapes. *Trends Genet.* **2006**, 22, 511–519. [CrossRef]
- 149. Badouin, H.; Velt, A.; Gindraud, F.; Flutre, T.; Dumas, V.; Vautrin, S.; Marande, W.; Corbi, J.; Sallet, E.; Ganofsky, J.; et al. The wild grape genome sequence provides insights into the transition from dioecy to hermaphroditism during grape domestication. *Genome Biol.* **2020**, *21*, 1–24. [CrossRef] [PubMed]
- 150. Duangjai, S.; Samuel, M.R.; Munzinger, J.; Forest, F.; Wallnöfer, B.; Barfuss, M.H.; Fischer, G.; Chase, M.W. A multi-locus plastid phylogenetic analysis of the pantropical genus Diospyros (Ebenaceae), with an emphasis on the radiation and biogeographic origins of the New Caledonian endemic species. *Mol. Phylogenetics Evol.* **2009**, *52*, 602–620. [CrossRef] [PubMed]
- 151. Giordani, E. History and Current Status of Worldwide Production. In *The Persimmon Genome*; Springer: Cham, Switzerland, 2022; pp. 1–10. [CrossRef]
- 152. Tamura, M.; Tao, R.; Yonemori, K.; Utsunomiya, N.; Sugiura, A. Ploidy Level and Genome Size of Several *Diospyros* Species. J. Jpn. Soc. Hortic. Sci. **1998**, 67, 306–312. [CrossRef]
- 153. Kafkas, S.; Açar, İ.; Gözel, H. A project on developing monoecious pistachio (*Pistacia vera* L.) populations and determination of sex mechanism in Pistacia. *Options Méditérr* **2003**, *63*, 57–60.
- 154. Sheikhi, A.; Arab, M.M.; Brown, P.J.; Ferguson, L.; Akbari, M. Pistachio (*Pistacia* spp.) Breeding. In *Advances in Plant Breeding* Strategies: Nut and Beverage Crops; Springer: Cham, Switzerland, 2019; pp. 353–400. [CrossRef]
- 155. Sola-Campoy, P.J.; Robles, F.; Schwarzacher, T.; Rejón, C.R.; De La Herrán, R.; Navajas-Pérez, R. The Molecular Cytogenetic Characterization of Pistachio (*Pistacia vera* L.) Suggests the Arrest of Recombination in the Largest Heteropycnotic Pair HC1. *PLoS ONE* **2015**, *10*, e0143861. [CrossRef] [PubMed]
- 156. Kafkas, S.; Ma, X.; Zhang, X.; Topçu, H.; Navajas-Pérez, R.; Wai, C.M.; Tang, H.; Xu, X.; Khodaeiaminjan, M.; Güney, M.; et al. Pistachio genomes provide insights into nut tree domestication and ZW sex chromosome evolution. *Plant Commun.* 2022, *in press.* [CrossRef]
- 157. Singh, R.; Dwivedi, S.K.; Bala, M. Biosystematics and Botanical Descriptions of Seabuckthorn (*Hippophae* Sp.) in India. In *The Seabuckthorn Genome*; Springer: Cham, Switzerland, 2022; pp. 1–21. [CrossRef]
- 158. Mir, N.A.; Geelani, S.M. Seabuckthorn (*Hippophae* sp.): A Unique high altitude multipurpose plant species growing in cold regions. *Int. J. Adv. Res. Sci. Eng.* 2018, 7, 1–12.
- 159. Zeb, A. Important therapeutic uses of sea buckthorn (Hippophae): A review. J. Biol. Sci. 2004, 4, 687–693.
- 160. Li, T.S.; Schroeder, W. Sea Buckthorn (Hippophae rhamnoides L.): A Multipurpose Plant. Horttechnology 1996, 6, 370–380. [CrossRef]
- Krejcarová, J.; Straková, E.; Suchý, P.; Herzig, I.; Karásková, K. Sea buckthorn (*Hippophae rhamnoides* L.) as a potential source of nutraceutics and its therapeutic possibilities—A review. *Acta Veter Brno* 2015, *84*, 257–268. [CrossRef]
- 162. Singh, V. Global Distribution of Seabuckthorn (*Hippophae* Sp.) Resources and Their Utilization. In *The Seabuckthorn Genome*; Springer: Cham, Switzerland, 2022; pp. 345–368. [CrossRef]
- 163. Puterova, J.; Razumova, O.; Martinek, T.; Alexandrov, O.; Divashuk, M.; Kubat, Z.; Hobza, R.; Karlov, G.; Kejnovsky, E. Satellite DNA and Transposable Elements in Seabuckthorn (*Hippophae rhamnoides*), a Dioecious Plant with Small Y and Large X Chromosomes. *Genome Biol. Evol.* 2017, *9*, 197–212. [CrossRef]
- 164. Yu, L.; Diao, S.; Zhang, G.; Yu, J.; Zhang, T.; Luo, H.; Duan, A.; Wang, J.; He, C.; Zhang, J. Genome sequence and population genomics provide insights into chromosomal evolution and phytochemical innovation of *Hippophae rhamnoides*. *Plant Biotechnol. J.* 2022, 20, 1257–1273. [CrossRef]
- Luo, X.; Liu, J.; He, Z. Oligo-FISH Can Identify Chromosomes and Distinguish *Hippophaë rhamnoides* L. Taxa. *Genes* 2022, 13, 195.
 [CrossRef] [PubMed]
- Sharma, A.; Zinta, G.; Rana, S.; Shirko, P. Molecular identification of sex in Hippophae rhamnoides L. using isozyme and RAPD markers. For. Stud. China 2010, 12, 62–66. [CrossRef]
- 167. Persson, H.A.; Nybom, H. Genetic Sex Determination and RAPD Marker Segregation in the Dioecious Species Sea Buckthorn (*Hippophae Rhamnoides* L.). *Hereditas* **2004**, 129, 45–51. [CrossRef]
- 168. Zhou, W.; Wang, Y.; Zhang, G.; Luan, G.; Chen, S.; Meng, J.; Wang, H.; Hu, N.; Suo, Y. Molecular Sex Identification in Dioecious Hippophae rhamnoides L. via RAPD and SCAR Markers. *Molecules* **2018**, *23*, 1048. [CrossRef] [PubMed]
- 169. Stobdan, T.; Mishra, G.P.; Yadav, A.; Chaurasia, O.P. Methods in Seabuckthorn Breeding. In *The Seabuckthorn Genome*; Springer: Cham, Switzerland, 2022; pp. 331–344. [CrossRef]
- 170. Chen, J.-J.; Kratsch, H.; Norton, J.; Sun, Y.; Rupp, L. Nodulation and Plant Growth of Shepherdia × utahensis 'Torrey' Topdressed with Controlled-release Fertilizer. *Hortscience* **2020**, *55*, 1956–1962. [CrossRef]
- 171. Riedl, K.M.; Choksi, K.; Wyzgoski, F.J.; Scheerens, J.C.; Schwartz, S.J.; Reese, R.N. Variation in Lycopene and Lycopenoates, Antioxidant Capacity, and Fruit Quality of Buffaloberry (*Shepherdia argentea* [Pursh] Nutt.). *J. Food Sci.* 2013, 78, C1673–C1679. [CrossRef]

- 172. Divashuk, M.; Alexandrov, O.; Kroupin, P.; Karlov, G. Molecular Cytogenetic Mapping of *Humulus lupulus* Sex Chromosomes. *Cytogenet. Genome Res.* **2011**, 134, 213–219. [CrossRef]
- 173. Karlov, G.; Danilova, T.; Horlemann, C.; Weber, G. Molecular cytogenetics in hop (*Humulus lupulus* L.) and identification of sex chromosomes by DAPI-banding. *Euphytica* 2003, *132*, 185–190. [CrossRef]
- 174. Neve, R.A. Sex Chromosomes in the Hop Humulus lupulus. Nature 1958, 181, 1084–1085. [CrossRef]
- 175. Neve R., A. Hops; Chapman and Hall: London, UK, 1991.
- 176. Čerenak, A.; Kolenc, Z.; Sehur, P.; Whittock, S.P.; Koutoulis, A.; Beatson, R.; Buck, E.; Javornik, B.; Škof, S.; Jakše, J. New Male Specific Markers for Hop and Application in Breeding Program. *Sci. Rep.* 2019, 9, 14223. [CrossRef]
- 177. Polley, A.; Ganal, M.W.; Seigner, E. Identification of sex in hop (*Humulus lupulus*) using molecular markers. *Genome* **1997**, 40, 357–361. [CrossRef]
- Jakše, J.; Štajner, N.; Kozjak, P.; Čerenak, A.; Javornik, B. Trinucleotide microsatellite repeat is tightly linked to male sex in hop (*Humulus lupulus L.*). Mol. Breed. 2007, 21, 139–148. [CrossRef]
- Haunold, A. Cytology, Sex Expression, and Growth of a Tetraploid X Diploid Cross in Hop (*Humulus lupulus* L.) 1. Crop. Sci. 1971, 11, 868–871. [CrossRef]
- 180. Beatson, R.A.; Ferguson, A.R.; Weir, I.E.; Graham, L.T.; Ansell, K.A.; Ding, H. Flow cytometric identification of sexually derived polyploids in hop (*Humulus lupulus* L.) and their use in hop breeding. *Euphytica* 2003, *134*, 189–194. [CrossRef]
- Younis, R.A.; Ismail, O.M.; Soliman, S.S. Identification of sex-specific DNA markers for date palm (*Phoenix dactylifera* L.) using RAPD and ISSR techniques. *Res. J. Agric. Biol. Sci.* 2008, 4, 278–284.
- Jaskani, M.J.; Awan, F.S.; Ahmad, S.; Khan, I.A. Maryam Development of molecular method for sex identification in date palm (*Phoenix dactylifera* L.) plantlets using novel sex-linked microsatellite markers. 3 Biotech 2016, 6, 1–7. [CrossRef]
- Intha, N.; Chaiprasart, P. Sex determination in date palm (*Phoenix dactylifera* L.) by PCR based marker analysis. *Sci. Hortic.* 2018, 236, 251–255. [CrossRef]
- Elmeer, K.; Mattat, I. Marker-assisted sex differentiation in date palm using simple sequence repeats. 3 Biotech 2012, 2, 241–247. [CrossRef]
- 185. Dhawan, C.; Kharb, P.; Sharma, R.; Uppal, S.; Aggarwal, R.K. Development of male-specific SCAR marker in date palm (*Phoenix dactylifera* L.). *Tree Genet. Genomes* **2013**, *9*, 1143–1150. [CrossRef]
- 186. Torres, M.F.; Mathew, L.S.; Ahmed, I.; Al-Azwani, I.K.; Krueger, R.; Rivera-Nuñez, D.; Mohamoud, Y.A.; Clark, A.G.; Suhre, K.; Malek, J.A. Genus-wide sequencing supports a two-locus model for sex-determination in *Phoenix. Nat. Commun.* 2018, 9, 1–9. [CrossRef]
- 187. Zhang, X.; Tan, J.; Yang, M.; Yin, Y.; Al-Mssallem, I.S.; Yu, J. Date Palm Genome Project at the Kingdom of Saudi Arabia. In *Date Palm Biotechnology*; Springer: Dordrecht, The Netherlands, 2011; pp. 427–448. [CrossRef]
- Al-Mssallem, I.S.; Hu, S.; Zhang, X.; Lin, Q.; Liu, W.; Tan, J.; Yu, X.; Liu, J.; Pan, L.; Zhang, T.; et al. Genome sequence of the date palm *Phoenix dactylifera* L. *Nat. Commun.* 2013, 4, 2274. [CrossRef]
- 189. Al-Dous, E.K.; George, B.; E Al-Mahmoud, M.; Al-Jaber, M.Y.; Wang, H.; Salameh, Y.M.; Al-Azwani, E.K.; Chaluvadi, S.; Pontaroli, A.C.; DeBarry, J.; et al. De novo genome sequencing and comparative genomics of date palm (*Phoenix dactylifera*). *Nat. Biotechnol.* 2011, 29, 521–527. [CrossRef]
- Mathew, L.S.; Spannagl, M.; Al-Malki, A.; George, B.; Torres, M.F.; Al-Dous, E.K.; Al-Azwani, E.K.; Hussein, E.; Mathew, S.; Mayer, K.F.; et al. A first genetic map of date palm (*Phoenix dactylifera*) reveals long-range genome structure conservation in the palms. *BMC Genom.* 2014, 15, 285. [CrossRef] [PubMed]
- 191. Nair, R.R. Chromosome number analysis in different sex types and open-pollinated seedlings of nutmeg (*Myristica fragrans* Houtt). *J. Plant. Crops* **2019**, 47, 197–201. [CrossRef]
- 192. Olajide, O.A.; Ajayi, F.F.; Ekhelar, A.I.; Awe, S.O.; Makinde, J.M.; Alada, A.A. Biological effects of *Myristica fragrans* (nutmeg) extract. *Phytother. Res. Int. J. Devoted Pharmacol. Toxicol. Eval. Nat. Prod. Deriv.* **1999**, *13*, 344–345. [CrossRef]
- 193. Mintah, F.D. Sex Determination in Nutmeg Seedlings Using Scar Primers. J. Hortic. Plant Res. 2018, 3, 40–47. [CrossRef]
- 194. Nikam, D.P.; Ingale, P.C.; Gokhale, N.B.; Lajurkar, V.G. Sex Determination in Nutmeg (*Myristica fragrance* Hott.) by using RAPD Markers. *Indian Hortic. J.* 2016, 6, 148–149.
- 195. Ferguson, A.R. Botanical description. In The Kiwifruit Genome; Springer: Cham, Switzerland, 2016; pp. 1–13. [CrossRef]
- 196. Ferguson, A.R. Kiwifruit: The wild and the cultivated plants. In *Advances in Food and Nutrition Research;* Academic Press: Cambridge, MA, USA, 2013; Volume 68, pp. 15–32.
- 197. Mcneilage, M.A. Gender variation in Actinidia deliciosa, the kiwifruit. Sex. Plant Reprod. 1991, 4, 267–273. [CrossRef]
- 198. Ferguson, A.R.; Huang, H. Genetic resources of kiwifruit: Domestication and breeding. Hortic. Rev. 2007, 33, 1–121.
- 199. Yan, G.; Yao, J.; Ferguson, A.R.; Mcneilage, M.A.; Seal, A.G.; Murray, B.G. New reports of chromosome numbers in *Actinidia* (Actinidiaceae). N. Z. J. Bot. **1997**, 35, 181–186. [CrossRef]
- Gill, G.P.; Harvey, C.F.; Gardner, R.C.; Fraser, L.G. Development of sex-linked PCR markers for gender identification in *Actinidia*. *Theor. Appl. Genet.* 1998, 97, 439–445. [CrossRef]
- Seal, A.G.; Ferguson, A.R.; De Silva, H.N.; Zhang, J.-L. The effect of 2n gametes on sex ratios in *Actinidia. Sex. Plant Reprod.* 2012, 25, 197–203. [CrossRef]
- He, Z.-C.; Li, J.Q.; Cai, Q.; Wang, Q. The cytology of *Actinidia, Saurauia* and *Clematoclethra* (Actinidiaceae). *Bot. J. Linn. Soc.* 2005, 147, 369–374. [CrossRef]

- 203. Zhang, Q.; Liu, C.; Liu, Y.; VanBuren, R.; Yao, X.; Zhong, C.; Huang, H. High-density interspecific genetic maps of kiwifruit and the identification of sex-specific markers. *DNA Res.* 2015, *22*, 367–375. [CrossRef] [PubMed]
- 204. Shirkot, P.; Sharma, D.R.; Mohapatra, T. Molecular identification of sex in Actinidia deliciosa var. deliciosa by RAPD markers. *Sci. Hortic.* **2002**, *94*, 33–39. [CrossRef]
- 205. Hale, I.; Melo, A.; Gustafson, H. Sex-linked molecular markers for two cold-hardy kiwifruit species, *Actinidia arguta* and *A. kolomikta. Eur. J. Hortic. Sci.* 2018, 83, 236–246. [CrossRef]
- 206. Chłosta, I.; Kwolek, D.; Sliwinska, E.; Góralski, G.; Popielarska-Konieczna, M. Sex-Linked Molecular Markers Identify Female Lines in Endosperm-Derived Kiwifruit Callus and in Regenerants. *Plants* 2021, 10, 526. [CrossRef] [PubMed]
- 207. Choi, Y.H.; Sertic, S.; Kim, H.K.; Wilson, E.G.; Michopoulos, F.; Lefeber, A.W.M.; Erkelens, C.; Kricun, S.D.P.; Verpoorte, R. Classification of *Ilex* Species Based on Metabolomic Fingerprinting Using Nuclear Magnetic Resonance and Multivariate Data Analysis. *J. Agric. Food Chem.* 2005, 53, 1237–1245. [CrossRef]
- Gottlieb, A.M.; Giberti, G.C.; Poggio, L. Molecular analyses of the genus *Ilex* (Aquifoliaceae) in southern South America, evidence from AFLP and ITS sequence data. *Am. J. Bot.* 2005, *92*, 352–369. [CrossRef] [PubMed]
- Evens, Z.N.; Stellpflug, S. Holiday Plants with Toxic Misconceptions. WestJEM 21.2 March Issue 2012, 13, 538–542. [CrossRef]
 [PubMed]
- 210. Yao, X.; Zhang, F.; Corlett, R.T. Utilization of the Hollies (Ilex L. spp.): A Review. Forests 2022, 13, 94. [CrossRef]
- 211. Barral, G.; Poggio, L. Chromosome numbers and DNA content from *Ilex argentina* (Aquifoliaceae). *Bol Soc Argent Bot* **1995**, 30, 243–248.
- 212. Gaiad, S.; Rakocevic, M.; Reissmann, C.B. N sources affect growth, nutrient content, and net photosynthesis in maté (*Ilex paraguariensis* St. Hil.). *Braz. Arch. Biol. Technol.* 2006, 49, 689–697. [CrossRef]
- 213. Rakocevic, M.; Costes, E.; Assad, E. Structural and physiological sexual dimorphism estimated from three-dimensional virtual trees of yerba-mate (*Ilex paraguariensis*) is modified by cultivation environment. *Ann. Appl. Biol.* **2011**, *159*, 178–191. [CrossRef]
- Rakocevic, M.; Medrado, M.J.S. Quality of yerba-mate leaves originating from male and female plants. *Pesqui. Florest. Bras.* 2007, 54, 71–83.
- Gottlieb, A.M.; Poggio, L. Genomic screening in dioecious "yerba mate" tree (*Ilex paraguariensis* A. St. Hill., Aquifoliaceae) through representational difference analysis. *Genetica* 2010, 138, 567–578. [CrossRef]
- 216. Golan-Goldhirsch, A.; Jones, R.; Rowland, L. AFLP markers for sex determination in an ilex species. *Acta Hortic.* 2001, 546, 591–595. [CrossRef]
- 217. Gauer, L.; Cavalli-Molina, S. Genetic variation in natural populations of maté (*Ilex paraguariensis* A. St.-Hil., Aquifoliaceae) using RAPD markers. *Heredity* 2000, *84*, 647–656 . [CrossRef]
- Torimaru, T.; Tani, N.; Tsumura, Y.; Hiraoka, K.; Tomaru, N. Development and polymorphism of simple sequence repeat DNA markers for the evergreen shrub *Ilex leucoclada* M. *Mol. Ecol. Notes* 2004, *4*, 531–533. [CrossRef]
- 219. Chen, K.; Xu, C. Red bayberry: Botany and horticulture. In *Horticultural Reviews*; John Wiley & Sons Inc.: Oxford, UK, 2010; pp. 83–114.
- 220. Stokes, J. Cytological Studies in the Myricaceae. Bot. Gaz. 1937, 99, 387–399. [CrossRef]
- 221. Jia, H.; Zhao, L.; Wang, Y.; Wu, H.; Zhao, H.; Zhu, Y.; Jiao, Y.; Wang, G.; Zhou, C.; Huang, C.; et al. Comparative Transcriptome Analysis Reveals Sex-Biased Expression of Hormone-Related Genes at an Early Stage of Sex Differentiation in Red Bayberry (*Morella rubra*). Horticulturae 2022, 8, 183. [CrossRef]
- 222. Ward, S.M.; Webster, T.M.; Steckel, L.E. Palmer Amaranth (*Amaranthus palmeri*): A Review. Weed Technol. 2013, 27, 12–27. [CrossRef]
- 223. Tranel, P.J.; Riggins, C.; Bell, M.S.; Hager, A.G. Herbicide Resistances in Amaranthus tuberculatus: A Call for New Options. J. Agric. Food Chem. 2011, 59, 5808–5812. [CrossRef] [PubMed]
- Mesgaran, M.B.; Matzrafi, M.; Ohadi, S. Sex dimorphism in dioecious Palmer amaranth (*Amaranthus palmeri*) in response to water stress. *Planta* 2021, 254, 1–10. [CrossRef] [PubMed]
- 225. Barrett, S.C.; Hough, J. Sexual dimorphism in flowering plants. J. Exp. Bot. 2012, 64, 67–82. [CrossRef] [PubMed]
- 226. Dawson, T.E.; Geber, M.A. Sexual Dimorphism in Physiology and Morphology. In *Gender and Sexual Dimorphism in Flowering Plants*; Springer: Berlin/Heidelberg, Germany, 1999; pp. 175–215. [CrossRef]
- Harris, M.S.; Pannell, J.R. Roots, shoots and reproduction: Sexual dimorphism in size and costs of reproductive allocation in an annual herb. *Proc. R. Soc. B Boil. Sci.* 2008, 275, 2595–2602. [CrossRef] [PubMed]
- 228. Obeso, J.R. The costs of reproduction in plants. New Phytol. 2002, 155, 321–348. [CrossRef] [PubMed]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.