

## Supplementary Materials

# Lignin Extracted from Various Parts of Castor (*Ricinus communis* L.) Plant: Structural Characterization and Catalytic Depolymerization

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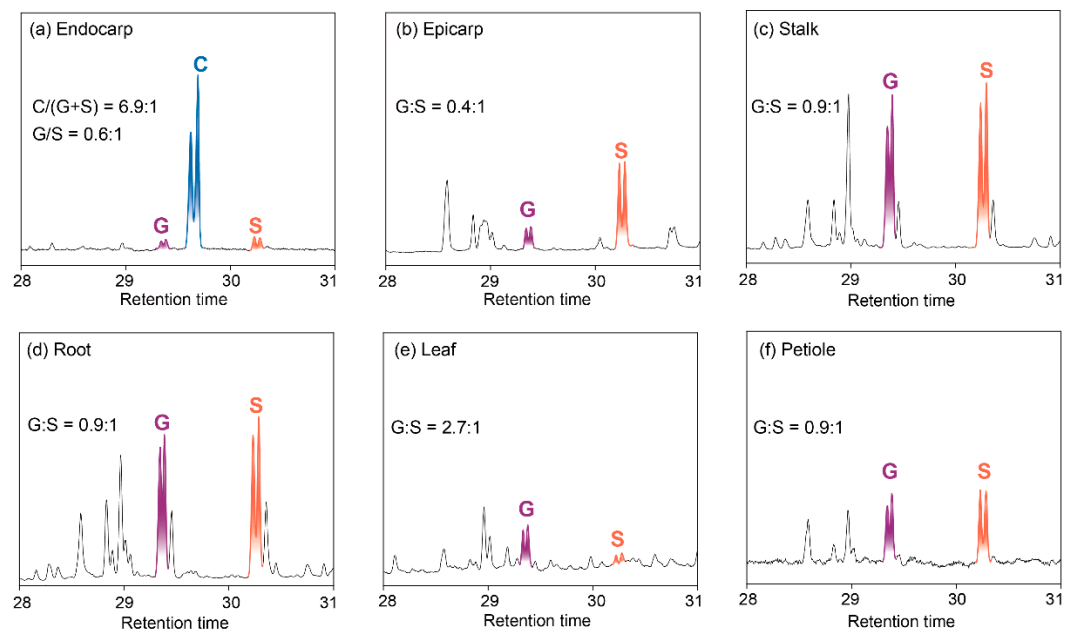
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**Table S1.** The contents of acid insoluble lignin (AIL), acid soluble lignin (ASL), total lignin content (TLC), cellulose, hemicellulose and extraction of the various raw materials.<sup>a</sup>

Samples	AIL	ASL	TLC	Cellulose	Hemicellulose	Extraction
Endocarp	59.5	1.1	13.3 <sup>b</sup>	19.3	10.6	9.1
Ericarp	14.4	2.4	16.8	42.1	27.2	2.5
Stalk	19.0	2.0	21.0	46.2	24.0	3.1
Root	21.8	3.9	25.7	44.7	19.5	3.8
Leaf	12.2	1.6	13.8	35.5	19.0	6.8
Petiole	8.9	2.2	11.1	37.7	17.7	4.5

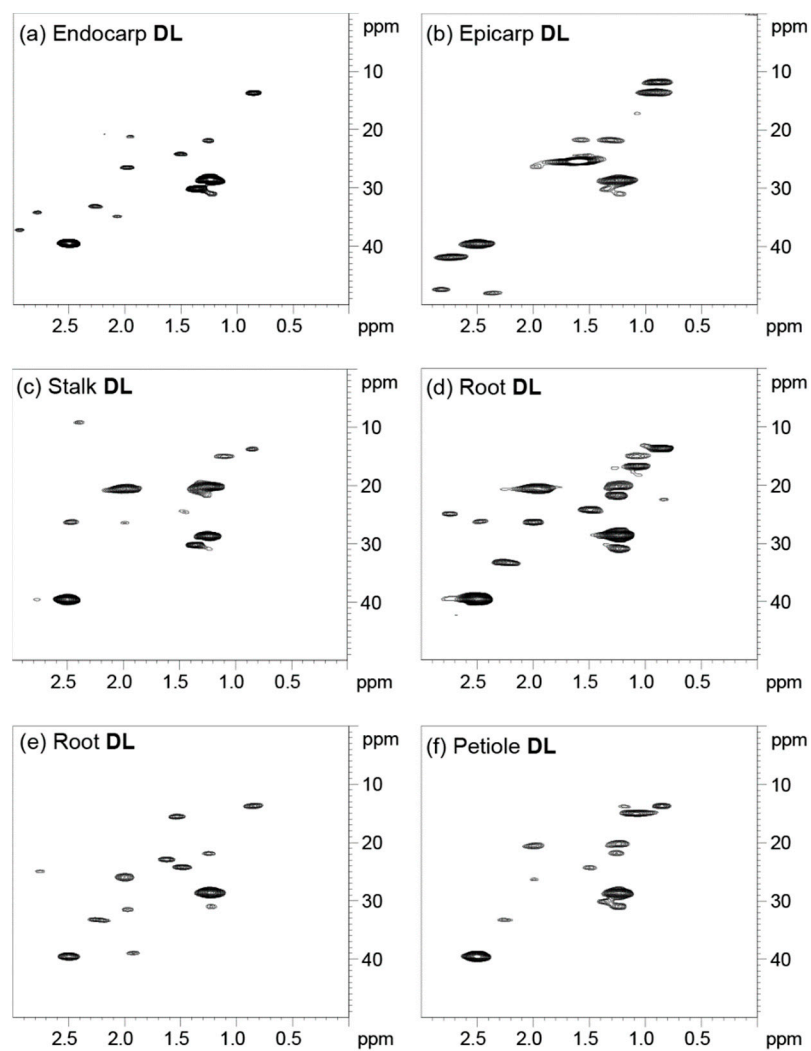
<sup>a</sup> The compositions of biomass were analyzed according to the procedures of the NREL method. <sup>b</sup> Based on the dilute HCl in 1,4-dioxane method.



**Figure S1.** GC spectra of thioacidolysis products from different parts of castor.

**Table S2.** The contents of acid insoluble lignin (AIL), acid soluble lignin (ASL), total lignin content (TLC), cellulose, and hemicellulose of the various DL samples.

Samples	AIL	ASL	TLC	Cellulose	Hemicellulose	Total
Endocarp	85.0	1.6	86.6	3.4	2.9	92.9
Epicarp	62.8	6.2	69.0	3.1	2.3	74.4
Stalk	75.3	1.4	76.7	8.2	1.6	86.5
Root	70.3	3.3	73.6	7.1	0.5	81.2
Leaf	52.0	6.4	58.4	5.9	3.5	67.8
Petiole	57.2	6.0	63.2	6.2	2.0	71.4



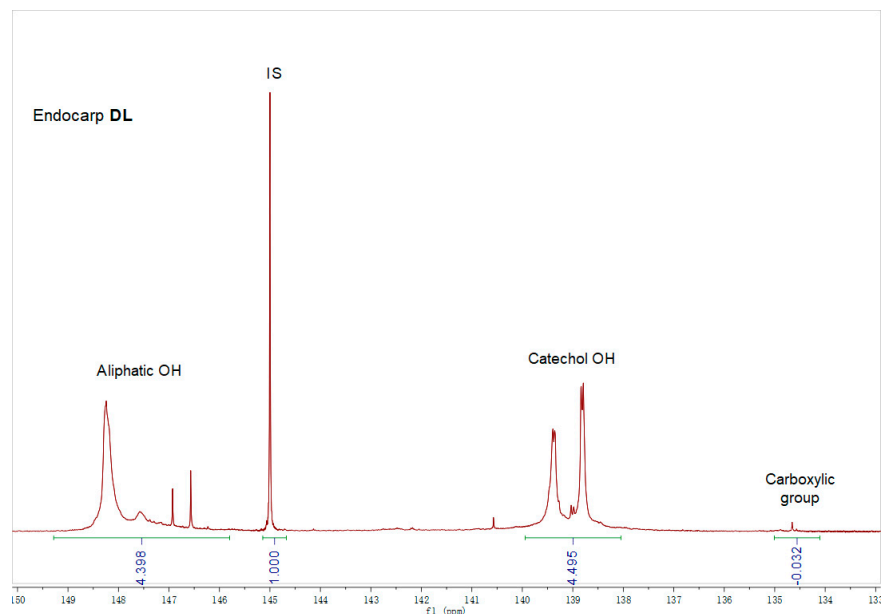
**Figure S2.** 2D NMR spectra of different DL samples.

**Table S3.** NMR data for the signal assignments for DL samples from various parts of castor in DMSO-*d*<sub>6</sub>.

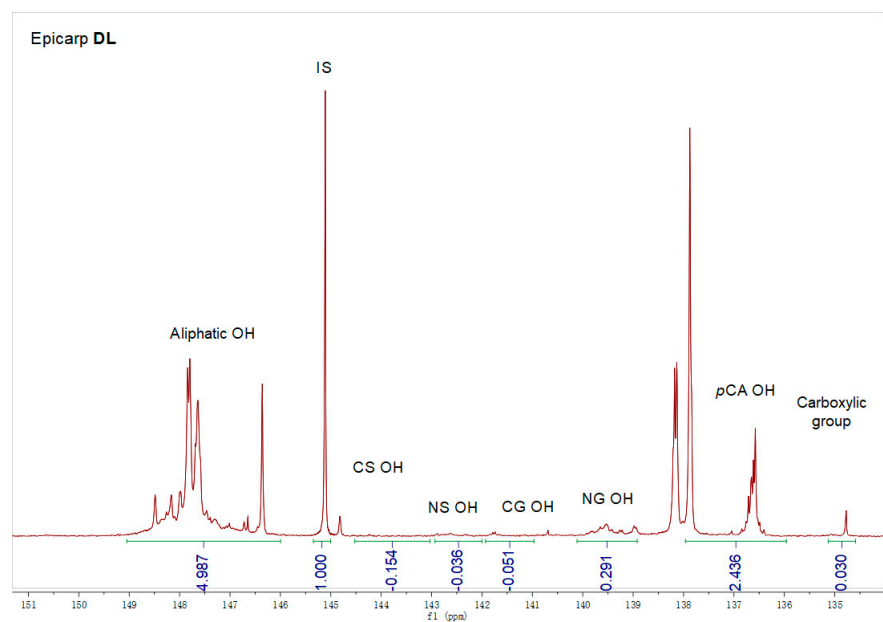
Label	$\delta_C/\delta_H$ (ppm)	Assignment
I <sub>α</sub>	75.6/4.93	C <sub>α</sub> -H <sub>α</sub> in benzodioxane substructures (I)
I <sub>β</sub>	78.2/4.13	C <sub>β</sub> -H <sub>β</sub> in benzodioxane substructures (I)
I <sub>γ</sub>	60.1/3.29-3.76	C <sub>γ</sub> -H <sub>γ</sub> in benzodioxane substructures (I)
II <sub>α</sub>	84.5/4.62	C <sub>α</sub> -H <sub>α</sub> in β-β resinol substructures (II)
II <sub>β</sub>	53.4/3.04	C <sub>β</sub> -H <sub>β</sub> in β-β resinol substructures (II)
II <sub>γ</sub>	70.7/3.74-4.09	C <sub>γ</sub> -H <sub>γ</sub> in β-β resinol substructures (II)
III <sub>α</sub>	71.4/4.89	C <sub>α</sub> -H <sub>α</sub> in β-O-4 substructures (III)
III <sub>β</sub>	86.4/4.15	C <sub>β</sub> -H <sub>β</sub> in β-O-4 substructures (III)
III <sub>γ</sub>	60.8/3.21-3.95	C <sub>γ</sub> -H <sub>γ</sub> in β-O-4 substructures (III)
IV <sub>α</sub>	86.5/5.48	C <sub>α</sub> -H <sub>α</sub> in phenylcoumaran (IV)
IV <sub>β</sub>	53.2/3.47	C <sub>β</sub> -H <sub>β</sub> in phenylcoumaran (IV)
IV <sub>γ</sub>	62.3/3.88	C <sub>γ</sub> -H <sub>γ</sub> in phenylcoumaran (IV)
X <sub>α</sub>	127.9/6.42	C <sub>α</sub> -H <sub>α</sub> in cinnamyl alcohol end-units (X)
X <sub>β</sub>	128.7/6.20	C <sub>β</sub> -H <sub>β</sub> in cinnamyl alcohol end-units (X)
X <sub>γ</sub>	61.4/4.10	C <sub>γ</sub> -H <sub>γ</sub> in cinnamyl alcohol end-units (X)
G <sub>2</sub>	110.7/6.92	C <sub>2</sub> -H <sub>2</sub> in guaiacyl units (G)
G <sub>5</sub>	114.5/6.68	C <sub>5</sub> -H <sub>5</sub> in guaiacyl units (G)
G <sub>6</sub>	119.0/6.77	C <sub>6</sub> -H <sub>6</sub> in guaiacyl units (G)
S <sub>2/6</sub>	103.5/6.68	C <sub>2/6</sub> -H <sub>2/6</sub> in syringyl units (S)
S' <sub>2/6</sub>	106.2/7.3	C <sub>2/6</sub> -H <sub>2/6</sub> in oxidized syringyl units (S')
<i>p</i> CA <sub>3/5</sub>	115.5/6.76	C <sub>3/5</sub> -H <sub>3/5</sub> in <i>p</i> -coumarate ( <i>p</i> CA)
<i>p</i> CA <sub>2/6</sub>	130.5/7.45	C <sub>2/6</sub> -H <sub>2/6</sub> in <i>p</i> -coumarate ( <i>p</i> CA)
<i>p</i> CA <sub>7</sub>	144.6/7.47	C <sub>7</sub> -H <sub>7</sub> in <i>p</i> -coumarate ( <i>p</i> CA)
<i>p</i> CA <sub>8</sub>	113.9/6.22	C <sub>8</sub> -H <sub>8</sub> in <i>p</i> -coumarate ( <i>p</i> CA)
I <sub>2</sub> , I <sub>5</sub> , I <sub>6</sub>	115.9/6.80, 117.6/6.94, 119.8/6.90	C <sub>2</sub> -H <sub>2</sub> , C <sub>5</sub> -H <sub>5</sub> , C <sub>6</sub> -H <sub>6</sub> , in catechol units (I)

**Table S4.** Typical chemical shifts and integration regions for DL samples from various parts of castor in the <sup>31</sup>P NMR spectrum.

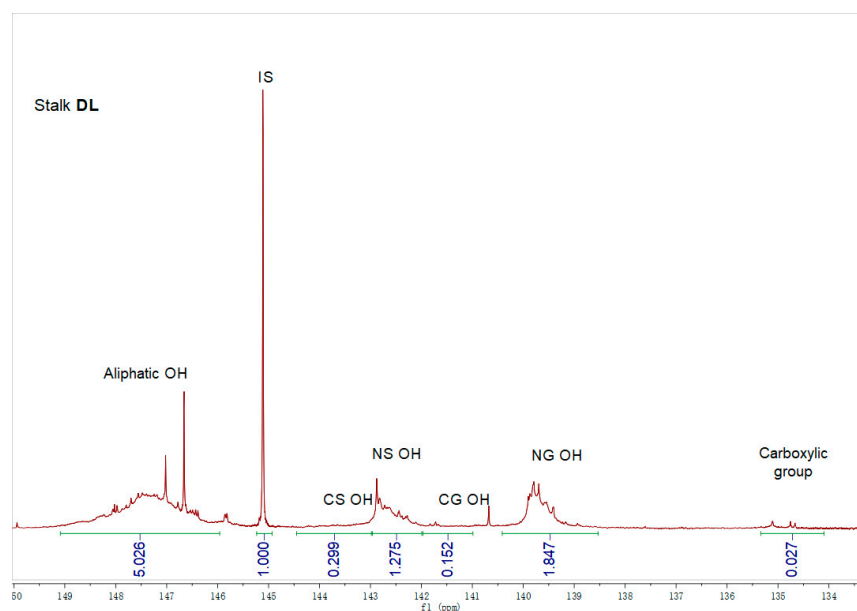
Structure	$\delta$ (ppm)
Aliphatic OH	145.4–150
Condensed Guaiacyl OH	141.42–142.17
Noncondensed Guaiacyl OH	138.79–140.17
Condensed Syringyl OH	143.2–144.5
Noncondensed Syringyl OH	142.17–143.2
<i>p</i> -coumarate ( <i>p</i> CA) OH	136.7–137.8
Catechol OH	138.0–140.0
Carboxylic group	134.2–135.5



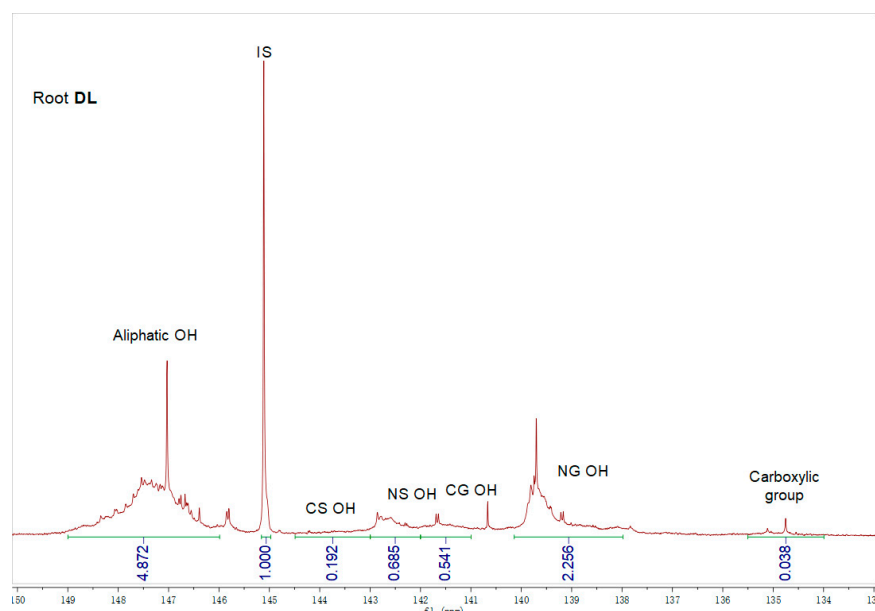
**Figure S3.** Quantitative  $^{31}\text{P}$  NMR spectrum of the Endocarp DL sample.



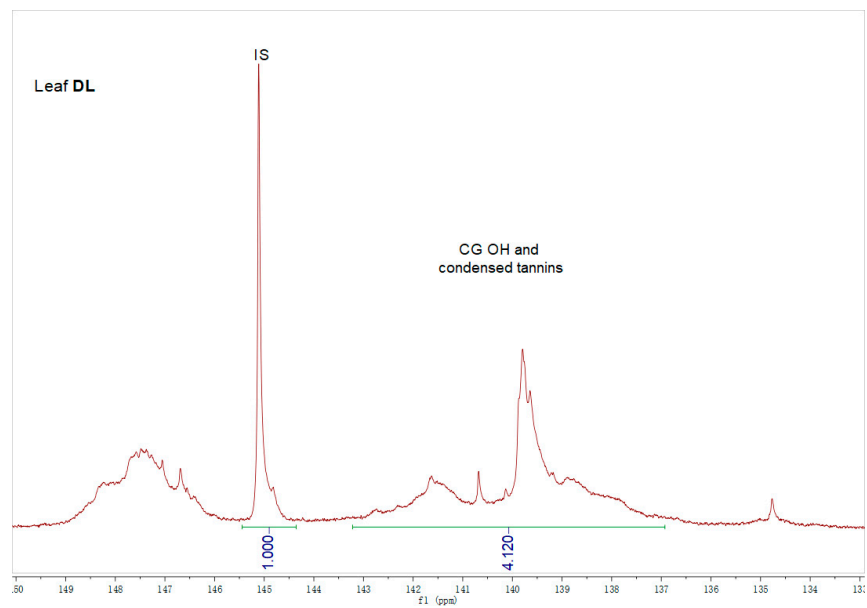
**Figure S4.** Quantitative  $^{31}\text{P}$  NMR spectrum of the Epicarp DL sample.



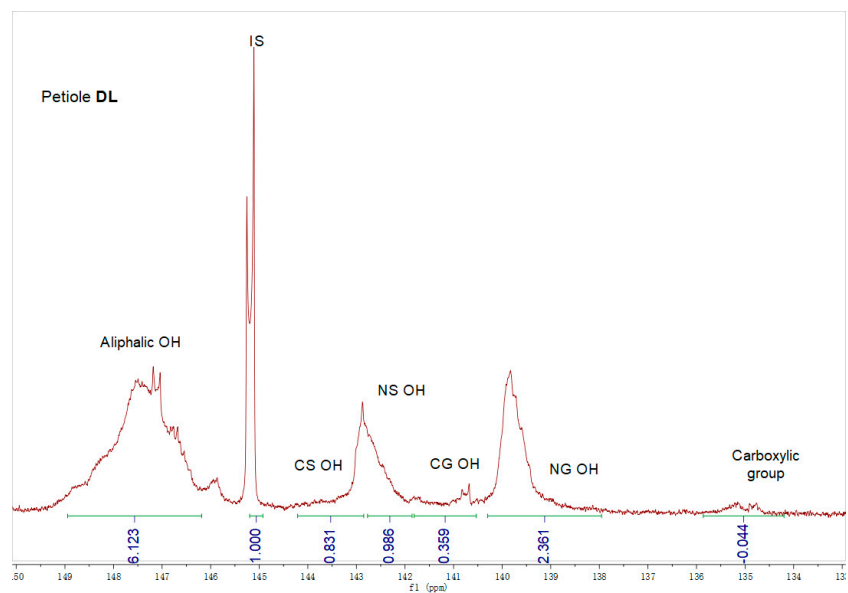
**Figure S5.** Quantitative  $^{31}\text{P}$  NMR spectrum of the Stalk DL sample.



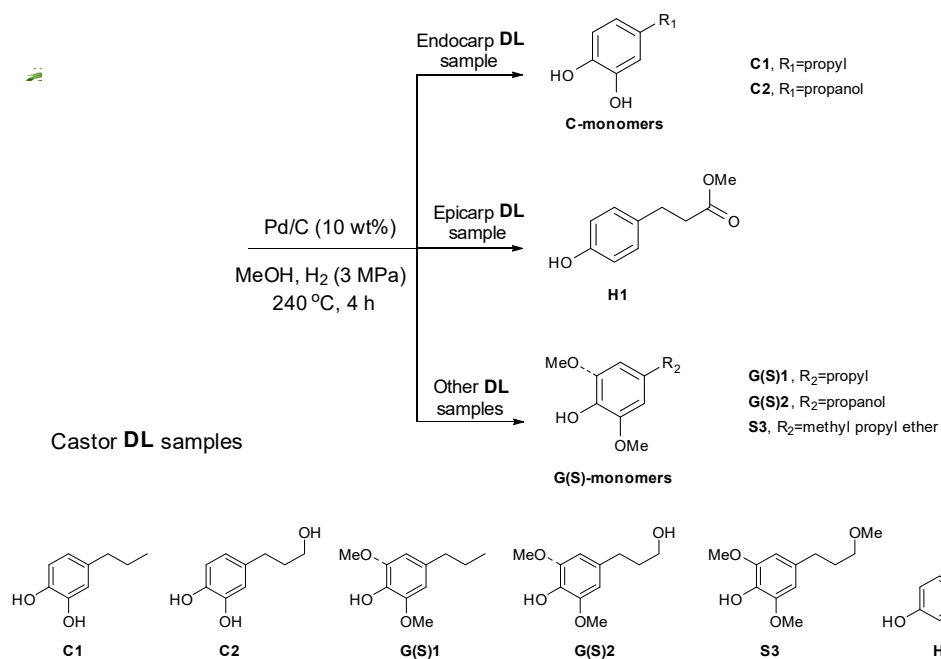
**Figure S6.** Quantitative  $^{31}\text{P}$  NMR spectrum of the Root DL sample.



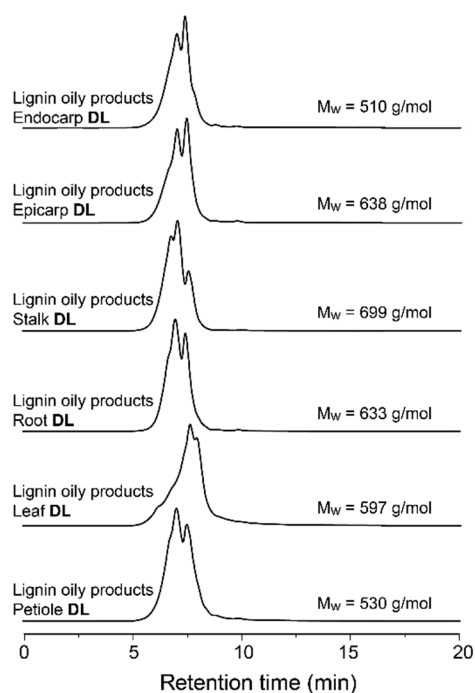
**Figure S7.** Quantitative  $^{31}\text{P}$  NMR spectrum of the Leaf DL sample.



**Figure S8.** Quantitative  $^{31}\text{P}$  NMR spectrum of the Petiole DL sample.



**Figure S9.** Products distribution of Pd/C-catalyzed depolymerization of various castor DL samples.



**Figure S10.** GPC spectra of lignin oily products from Pd/C-catalyzed hydrogenolysis of various DL samples of castor plants. Reaction conditions: sample (50 mg), Pd/C (10 mg), MeOH (10 mL), H<sub>2</sub> (3 MPa), 240 °C, 4 h.