

Supplementary Information

Synthesis and characterization of temperature-responsive

N-cyanomethylacrylamide-containing diblock copolymer assemblies
in water

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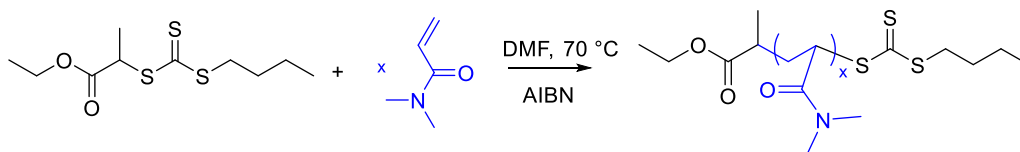
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1. Synthesis of the PDMAm macroRAFT agents



Scheme S1. Synthesis route for the PDMAm-TTC macroRAFT agents *via* RAFT-mediated solution polymerization in the presence of CTA-1.

Table S1. Experimental conditions and results for the RAFT-mediated polymerization of DMAm in DMF at 70 °C[#]

Entry	$\frac{[\text{DMAm}]_0}{[\text{CTA-1}]_0}$ ^a	$[\text{DMAm}]_0$ (mol L _{DMF} ⁻¹)	Time (min)	Conv. ^b (%)	$DP_{n,\text{th}}$ ^c	DP_n ^b	$M_{n,\text{th}}$ ^d (kg mol ⁻¹)	SEC DMF ^e	
								M_n ^e (kg mol ⁻¹)	\mathcal{D} ^e
M1	23	2.5	80	59	14	13	1.55	1.35	1.12
M2	29	2.4	93	64	19	23	2.55	2.22	1.09
M3	46	2.3	210	74	34	36	3.83	3.19	1.09

[#]Polymerizations were performed in the presence of a RAFT agent (CTA-1) using AIBN as a radical initiator at an initial molar ratio of CTA-1/AIBN: 1/0.05. ^a Initial monomer/CTA-1 molar ratio. ^b Determined by ¹H NMR (for the determination of the DP_n of M3 see Figure S1). ^c Theoretical number-average degree of polymerization, $DP_{n,\text{th}}$ calculated using the experimental conversion. ^d Theoretical number-average molar mass, $M_{n,\text{th}}$, calculated using $DP_{n,\text{th}}$. ^e Number-average molar mass, M_n and dispersity, \mathcal{D} , determined by SEC in DMF (+ LiBr 1 g L⁻¹) with a PMMA calibration.

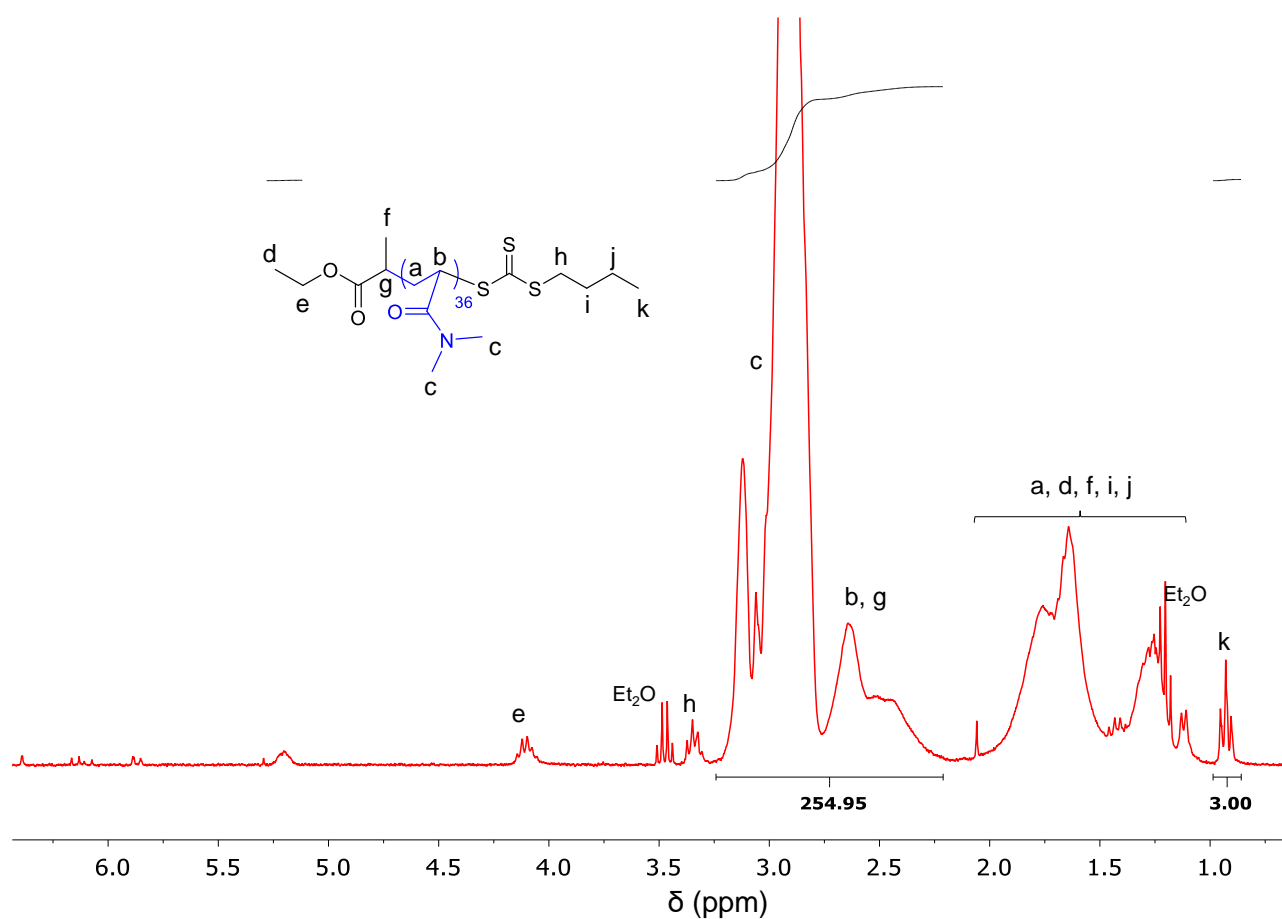


Figure S1. ^1H NMR spectrum of M3 (Table S1) in CDCl_3 .

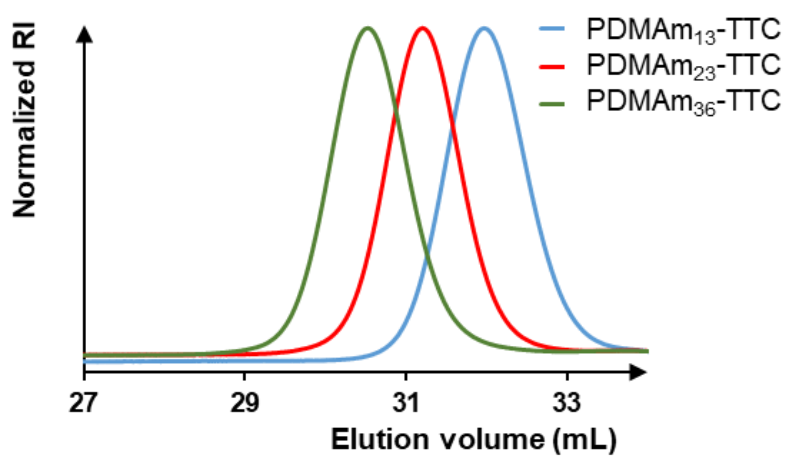


Figure S2. SEC chromatograms in DMF (+ LiBr) of the macroRAFT agents with different DP_{nS} .

2. Supplementary data

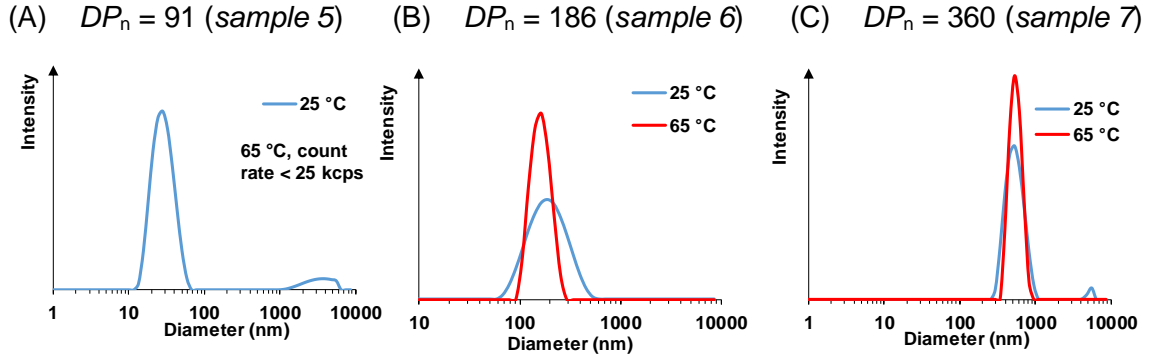


Figure S3. Temperature-dependent size distribution determined by DLS at 0.1 wt% in water for samples 5, 6 and 7 (PDMAM₃₆-*b*-PCMA_m).

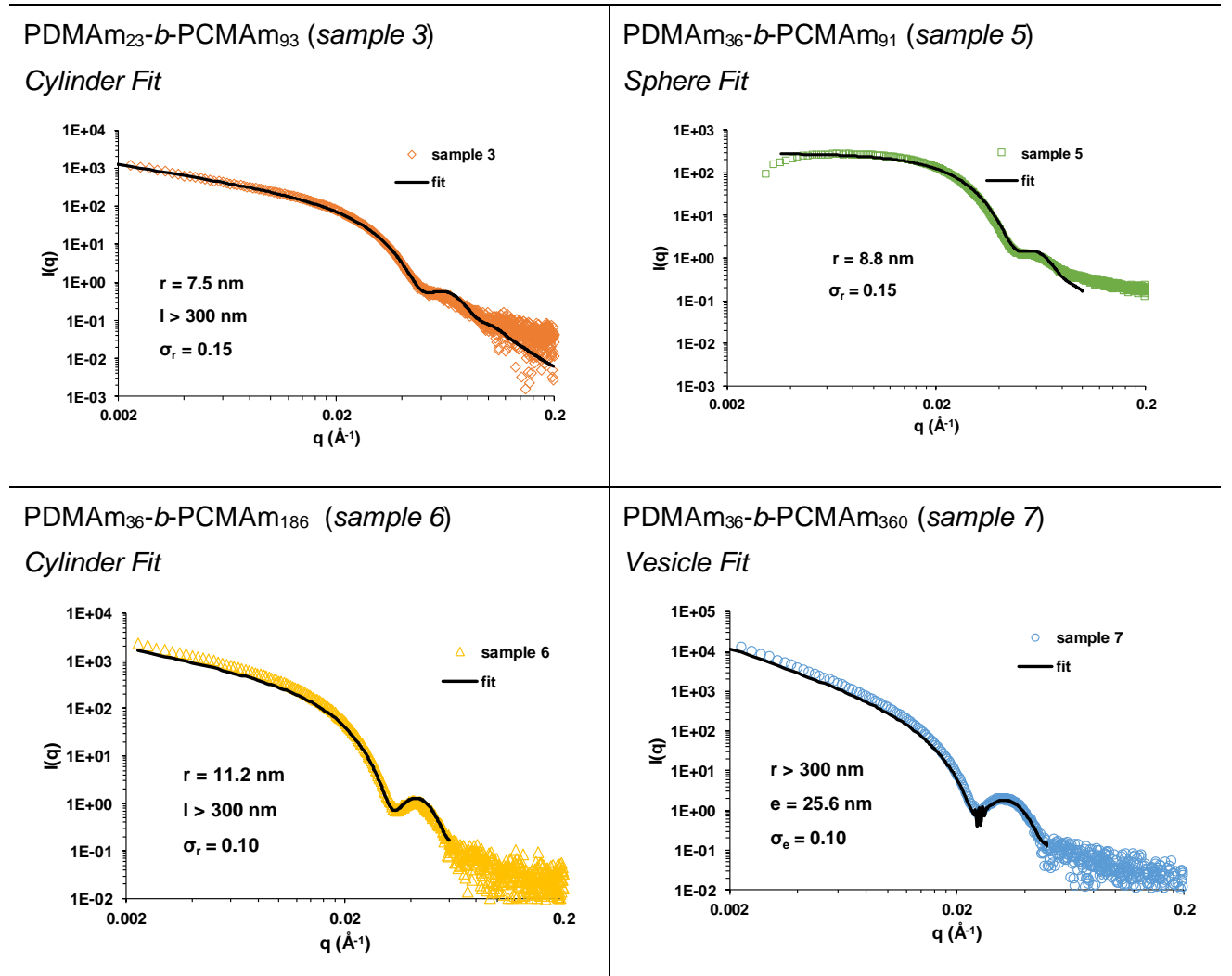


Figure S4. SAXS intensity (I) versus scattering vector (q) for selected diblock PDMAM-*b*-PCMA samples at 1 wt% in water at 25 °C (samples 3, 5, 6 and 7, Table 1).

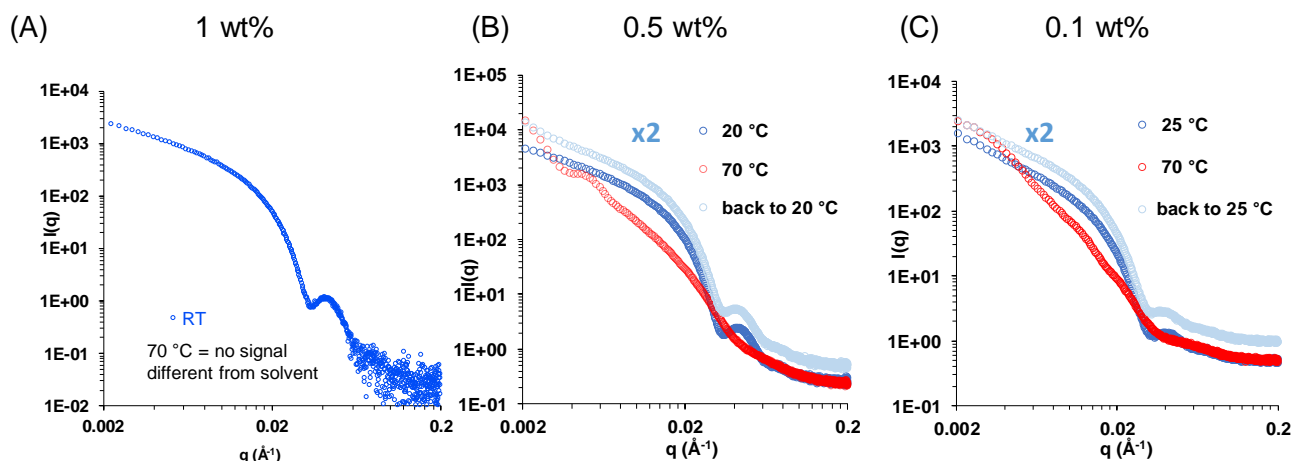


Figure S5. SAXS intensity (I) versus scattering vector (q) for PDMAm₃₆-*b*-PCMAm₁₈₆ (*sample 6*) in water at (A) 1 wt%; (B) 0.5 wt% and (C) 0.1 wt% in water at 25 °C (or 20 °C) and 70 °C. For the samples cooled back after heating, the intensity was multiplied by a factor of 2 in order to visualize the signal, since it is a perfect overlay with the initial sample.

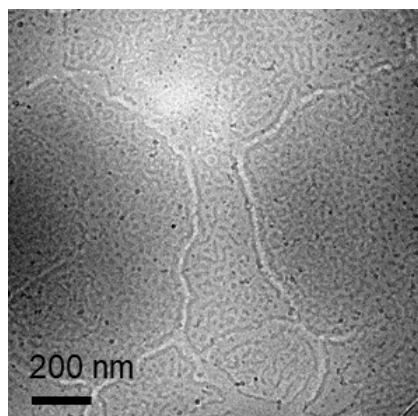


Figure S6. Selected cryo-TEM image of *sample 7* (PDMAm₃₆-*b*-PCMAm₃₆₀) prepared at 70 °C, showing the presence of additional morphologies different from standard vesicles. Sample concentration = 1 wt% in water.

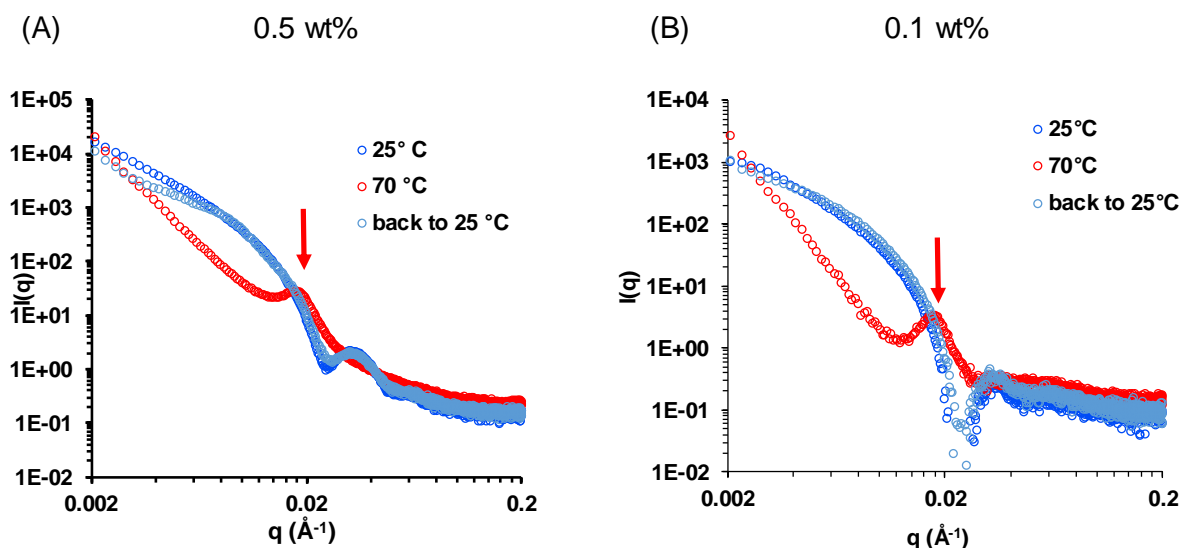


Figure S7. SAXS intensity (I) versus scattering vector (q) for PDMAm₃₆-*b*-PCMAm₃₆₀ (*sample 7*) in water at (A) 0.5 wt% and (B) 0.1 wt% at 25 °C (dark blue), 70 °C (red) and back to 25 °C (light blue). The arrows designate d , the reciprocal length corresponding to the inter-vesicle distance.

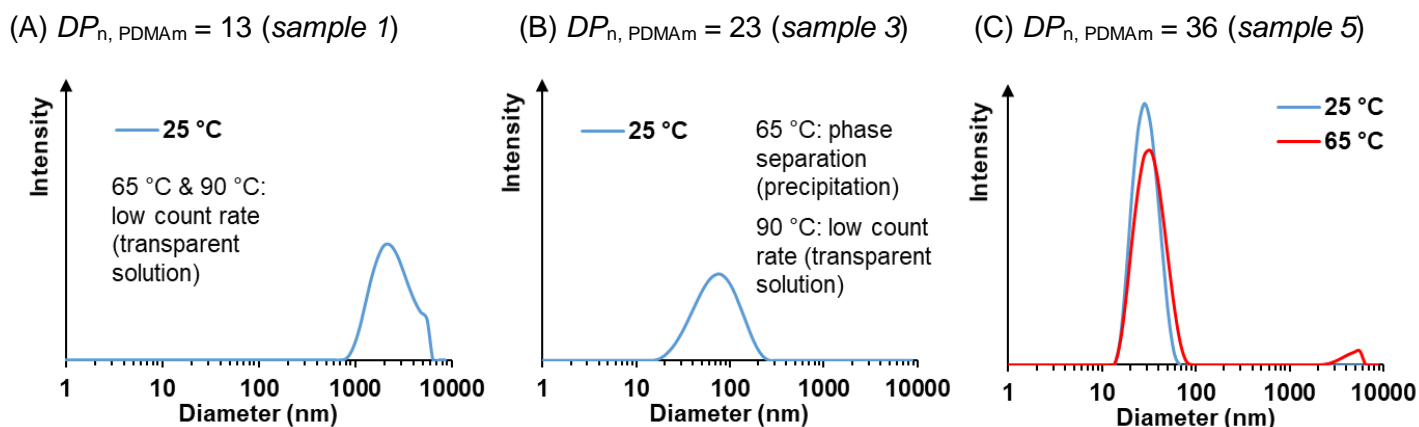


Figure S8. Size distributions determined by DLS of (A) PDMAm₁₃-*b*-PCMAm₉₀ (*sample 1*), (B) PDMAm₂₃-*b*-PCMAm₉₀ (*sample 3*) and (C) PDMAm₃₆-*b*-PCMAm₉₁ (*sample 5*) dispersions prepared at 1 wt% in water at various temperatures.

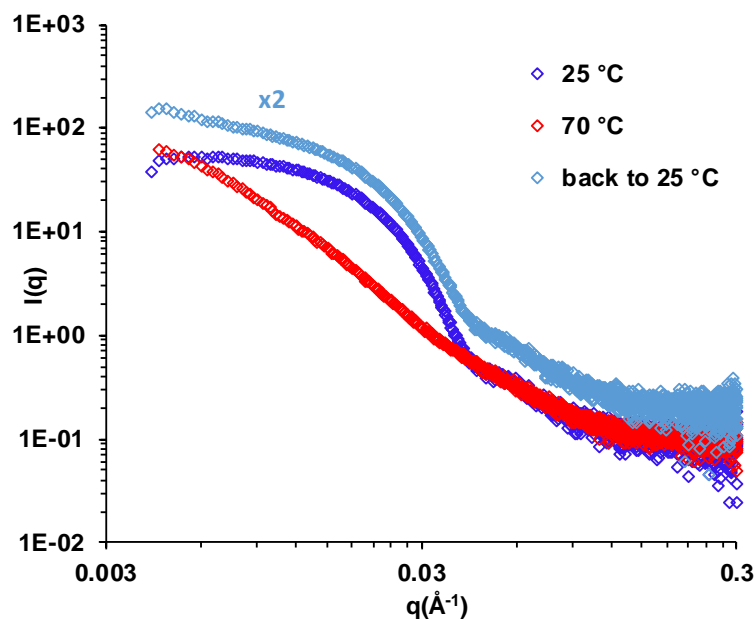


Figure S9. SAXS intensity (I) versus scattering vector (q) for PDMA_{m36}-*b*-PCMA_{m91} (*sample 5*) in water at 0.1 wt% at 25 °C (dark blue), 70 °C (red) and back to 25 °C (light blue, intensity multiplied by 2 in order to visualize the signal, since it is a perfect overlay with the initial sample before heating).

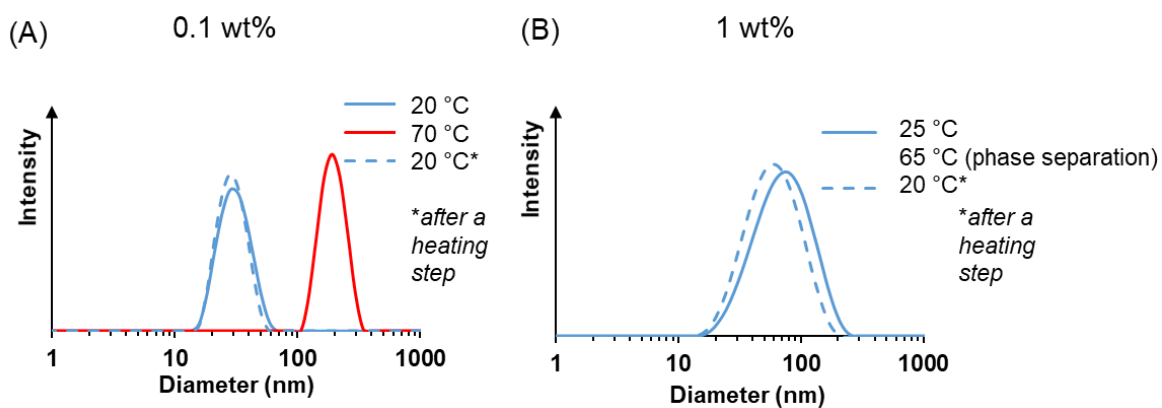


Figure S10. Size distribution of *sample 3* (PDMA_{m23}-*b*-PCMA_{m93}) at (A) 0.1 wt% and (B) 1 wt% in water determined by DLS at different temperatures. At 90 °C a very low scattering intensity was detected (kcps = 6 and 50, respectively for 0.1 wt% and 1 wt%).