

Transfer of a Rational Crystal Contact Engineering Strategy between Diverse Alcohol Dehydrogenases

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LkADH    1  TDRLKGKVAIVTGGTLGIGLAIADKFVEEGAKVVITGRHADVGEKAASIGGTDVIRFVQ
LbADH    1  SNRLDGKVAIITGGTLGIGLAIATKFVEEGAKVMITGRHSDVGEKAASVGTDPQIQFFQ
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LkADH    61 HDASDEAGWTKLFDTTEEAFGPVTTVVNNAGIAVSKSVEDTTTEEWRKLLSVNLDGVFFG
LbADH    61 HDSSDEAGWTKLFDATEKAFGPVSTLVNNAGIAVNKSVEETTAEWRKLLAVNLDGVFFG
          **  ***  *****  **  *****  *  *****  *****  *****  *****

LkADH    121 TRLGIRMKNGKLGASIIINMSSIEGFVGDP TLGAYNASKGAVRIMSKSAALDCALKDYDV
LbADH    121 TRLGIRMKNGKLGASIIINMSSIEGFVGDP SLGAYNASKGAVRIMSKSAALDCALKDYDV
          *****  *****  *****  *****  *****  *****  *****

LkADH    181 RVNTVHPGYIKTEPLVDDLEGAEEMMSORTKTPMGHIGEPNDIAWICVYLASDESKFATGA
LbADH    181 RVNTVHPGYIKTEPLVDDLPGAEEAMSORTKTPMGHIGEPNDIAYICVYLASNESKFATGS
          *****  *****  *****  *****  *****  *****  *****

LkADH    241 EFVVDGGYTAQ
LbADH    241 EFVVDGGYTAQ
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Figure S1. Sequence alignment of *LkADH* and *LbADH* wild type with an amino acid sequence homology of 88.5 %. The optimal global alignment was generated with the Needleman-Wunsch algorithm (EMBOSS, output format: pair, matrix: BLOSUM62) [1]. Identical amino acids are marked with an asterisk. Crystal contacts (distance between amino acids < 4 Å) are marked for *LkADH* (cyan) and *LbADH* (green). The positions selected for mutations, T102 and Q126, are underlined. RMSD of the crystal contacts are listed in Supplementary Table S2.

Table S1. Data collection and refinement statistics of X-ray diffraction experiments of *LkADH* wild type and mutant Q126K crystals (values in parentheses are for highest resolution shell).

<i>LkADH</i> Variant	Wild Type	Q126K
Data Collection		
PDB ID	7P36	7P7Y
Beamline	SLS Beamline X06DA	SLS Beamline X06DA
Wavelength, Å	1.000	1.000
Space group	I 2 2 2	I 2 2 2
Cell dimensions, Å	55.87, 79.97, 114.59	55.84, 80.04, 114.93
No. of molecules per asymmetric unit	1	1
Resolution, Å	50–1.14 (1.21–1.14)	50–1.25 (1.32–1.25)
<i>I</i> /σ(<i>I</i>)	71.4 (26.9)	49.61 (14.32)
CC (1/2)	100.0 (99.8)	100 (99.3)
Completeness, %	100.0 (99.8)	99.2 (95.3)
Redundancy	12.8 (11.9)	11.62 (6.43)

R _{meas} , %	2.3 (8.3)	3.3 (10.9)
Refinement		
Resolution, Å	1.14	1.25
No. unique reflections	93486 (6458)	70930 (4734)
R _{work} / R _{free}	0.093 / 0.107	0.088 / 0.132
No. atoms		
Protein	1865	1865
Water	387	344
Other	23	16
B-factors		
Overall	7.37	9.1
Protein main chain	6.39	7.95
Protein side chain	8.24	10.27
R.m.s. deviations		
Bond lengths, Å	0.012	0.012
Bond angles, °	1.82	1.68
Ramachandran plot		
Most favored, %	98	98
Additional allowed, %	2	2

Table S2. RMSD_{side chain} of crystal contacts present in both *LkADH* and *LbADH* wild type and calculated after side chain and backbone alignment of one tetramer. Crystal contacts were defined by a distance < 4 Å between amino acid residues of neighboring tetramers. The contact RMSD_{side chain} was then calculated by alignment of those amino acid residues forming a contact with PyMOL (v.2.3.; Schroedinger). Therefore, a smaller contact RMSD value indicates, that the corresponding crystal contacts in *LbADH* and *LkADH* wild type crystals are locally more similar.

Amino acid residue	Contacts	Contact RMSD _{side chain} , Å
H39	D74, E77, R127	1.26
V42	D41, T70	0.95
K45	E66	1.26
D54	T52, D54	4.89
K71	D197	1.58
T102	K48	1.25
Q126	A40/S40, D41, T70, L123, R127	1.13
E203	R208	2.59

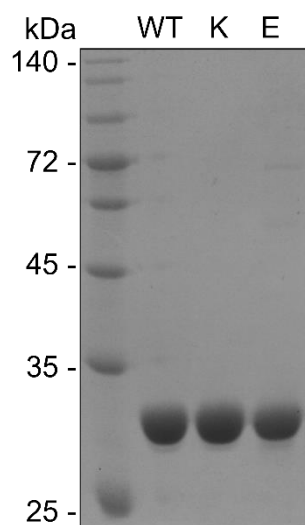


Figure S2. Illustration of protein purity analysis using SDS-PAGE of *LkADH* wild type (WT) and mutants Q126K (K) and T102E (E). The proteins purified by IMAC were loaded onto a 12.5% SDS gel (2.5 µg protein) under reducing conditions. Bands between 25–35 kDa correspond to *LkADH* monomers.

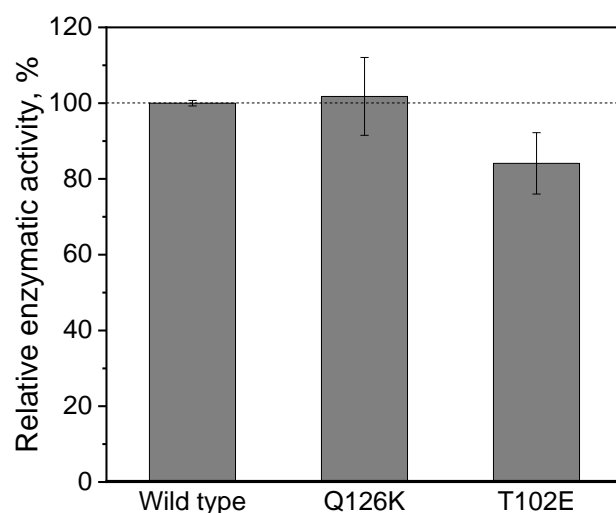


Figure S3. Maximum relative enzymatic activity of *LkADH* mutants Q126K and T102E compared to *LkADH* wild type. Purified protein solutions of *LkADH* variants were adjusted to 6 mg L⁻¹. Enzymatic activity was measured for 10 min at 340 nm (45 °C) with addition of 180 µL buffer (20 mM HEPES, pH 7.0; 1 mM MgCl₂) containing 0.5 mM NADPH and 10 mM acetophenone to a final volume of 200 µL (n = 3).

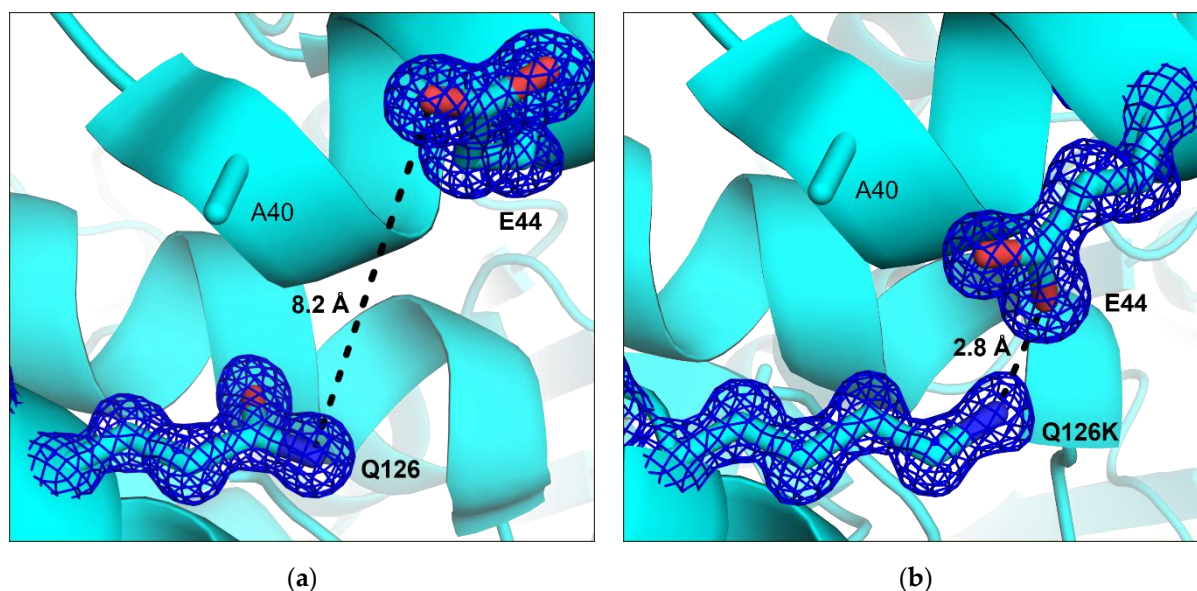


Figure S4. Electron density map (blue) of *LkADH* crystal contact at position (a) Q126 and (b) Q126K with well-defined electron density of the lysine and glutamic acid side chain. The map represents the structure factor amplitude difference $2F_o - F_c$ with a contour level of 1.0 σ (calculated with *REFMAC* [2]).

References

1. Madeira, F., Park, Y.M., Lee, J., Buso, N., Gur, T., Madhusoodanan, N., Lopez, R., The EMBL-EBI search and sequence analysis tools APIs in 2019. *Nucleic Acids Res.* **2019**, 47, W636–W641.
2. Murshudov, G.N., Vagin, A.A., and Dodson, E.J., Refinement of Macromolecular Structures by the Maximum-Likelihood Method. *Acta Crystallogr., Sect. D: Biol. Crystallogr.* **1997**. 53, 240–255.