

Supporting Information for

Selective Synthesis of Furfuryl Alcohol from Biomass-Derived Furfural Using Immobilized Yeast Cells

Xue-Ying Zhang ¹, Zhong-Hua Xu ¹, Min-Hua Zong ¹, Chuan-Fu Wang ² and Ning Li ^{1,*}

¹ School of Food Science and Engineering, South China University of Technology, 381 Wushan Road, Guangzhou 510640, China; zxy1989318@126.com (Z.X.Y.); xzh199000@163.com (Z.H.X.); btmhzong@scut.edu.cn (M.H.Z.)

² National Institute of Clean-and-Low-Carbon Energy, Future Science Park, Beijing 102211, China; wangchuanfu@nicenergy.com.

* Correspondence: lining@scut.edu.cn (N.L.)

Materials

Poly(vinyl alcohol) (PVA 1788, 87-89% hydrolyzed) was purchased from Aladdin Industrial Co. (Shanghai, China). Gelatin was purchased from Kermel Chemical Reagent Co. Ltd. (Tianjing, China). Dibutyl phthalate (>98.5%), and carrageenan were bought from Macklin Biochemical Co. Ltd. (Shanghai, China); Agar was from Biofroxx Co. (Germany).

Preparation of gel beads

Agar beads were prepared according to a previous method [1], with some modifications. Agar of 2 g was dissolved in 100 mL deionized water under heating. Then warm agar solution (50°C) was dropped into the cooled dibutyl phthalate with gentle stirring for solidification. After solidification for 12 h at 4°C, agar beads were washed thoroughly with Tris-HCl buffer (100 mM, pH 8.0).

Preparation of gelatin beads was performed according to a previous method [2], with some modifications. Gelatin of 15 g was dissolved in 100 mL deionized water under heating. Then warm gelatin solution (50°C) was dropped into the cooled dibutyl phthalate with gentle stirring for solidification. After solidification for 1 h at 4°C, gelatin beads were transferred into 10% glutaraldehyde solution for crosslinking. After 1 h, the beads were washed thoroughly with Tris-HCl buffer (100 mM, pH 8.0).

Carrageenan beads were prepared according to He's method [3], with some modifications. Briefly, carrageenan of 3 g was dissolved in 100 mL 0.9% physiological saline under heating. Then warm solution (50°C) was dropped into the cooled dibutyl phthalate with gentle stirring for solidification. After solidification for 1 h at 4°C, gel beads were transferred into cooled 2% KCl solution (4°C), followed by incubation for 4 h. Then the beads were crosslinked by 5%

glutaraldehyde. After 1 h, the beads were washed thoroughly with 2% KCl solution. PVA beads were prepared according to a previous method [4], with some modifications. PVA of 5 g and sodium alginate of 0.25 g were dissolved in 50 mL deionized water under heating. Then warm solution (50°C) was dropped into a mixed solution of saturated boric acid and 100 mM calcium chloride with gentle stirring, followed by incubation of 2 h. The formed PVA beads were washed thoroughly with Tris-HCl buffer (100 mM, pH 8.0).

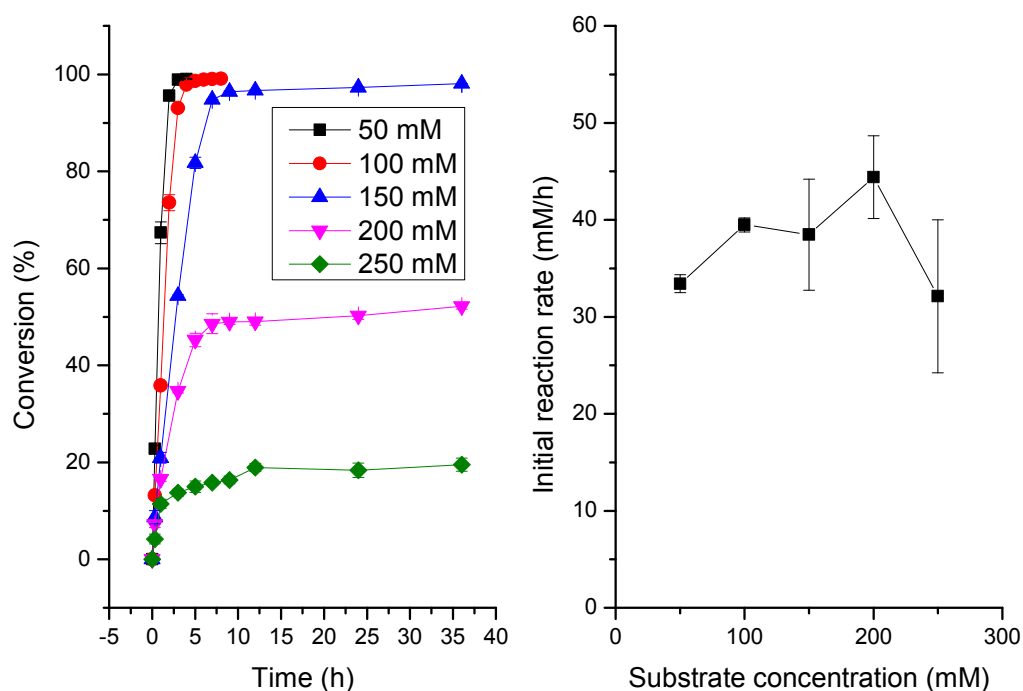


Figure S1 Time courses of substrate conversion in the reduction of furfural. The reaction conditions are the same as those in Figure 1d.

References

1. Vassileva, A.; Burhan, N.; Beschkov, V.; Spasova, D.; Radoevska, S.; Ivanova, V.; Tonkova, A. Cyclodextrin glucanotransferase production by free and agar gel immobilized cells of *bacillus circulans* atcc 21783. *Process Biochem.* **2003**, *38*, 1585-1591.
2. Aparecida de Assis, S.; Ferreira, B.S.; Fernandes, P.; Guaglianoni, D.G.; Cabral, J.M.S.; Oliveira, O.M.M.F. Gelatin-immobilized pectinmethylesterase for production of low methoxyl pectin. *Food Chem.* **2004**, *86*, 333-337.
3. He, Y.-C.; Xu, J.-H.; Su, J.-H.; Zhou, L. Bioproduction of glycolic acid from glycolonitrile with a new bacterial isolate of *alcaligenes* sp. Ecu0401. *Appl. Biochem. Biotechnol.* **2010**, *160*, 1428-1440.
4. Takei, T.; Ikeda, K.; Ijima, H.; Kawakami, K. Fabrication of poly(vinyl alcohol) hydrogel beads crosslinked using sodium sulfate for microorganism immobilization. *Process Biochem.* **2011**, *46*, 566-571.