

1 *Supplementary Material*

2 **Composites of cross-linked aggregates of Eversa®**
3 **Transform and magnetic nanoparticles. Performance**
4 **in the ethanolysis of soybean oil**

5 **Letícia Passos Miranda¹, José Renato Guimarães¹, Roberto Campos Giordano¹, Roberto**
6 **Fernandez-Lafuente^{2*}, Paulo Waldir Tardioli^{1*}**

7 ¹ Postgraduate Program in Chemical Engineering (PPGEQ), Department of Chemical Engineering, Federal
8 University of São Carlos (DEQ/UFSCar), Rod. Washington Luís, km 235, 13565-905, São Carlos, SP, Brazil;
9 lettypassos@gmail.com (L.P.M.); renatoge74@gmail.com (J.R.G.); roberto@ufscar.br (R.C.G.);
10 pwtardioli@ufscar.br (P.W.T.)

11 ² Departamento de Biocatálisis, ICP-CSIC, Campus UAM-CSIC, 28049, Madrid, Spain; rfl@icp.csic.es (R.F.-
12 L.)

13 * Correspondence: pwtardioli@ufscar.br (P.W.T.); rfl@icp.csic.es (R.F.-L.); Tel.: +55-16-3351-9362 (P.W.T.);
14 +34-91594804 (R.F.-L.)

15

16 Supplementary Material

17 1. Adsorption profile of Eversa Transform 2.0 on magnetic nanoparticles functionalized with 18 octyl and amine groups (SMNPs)

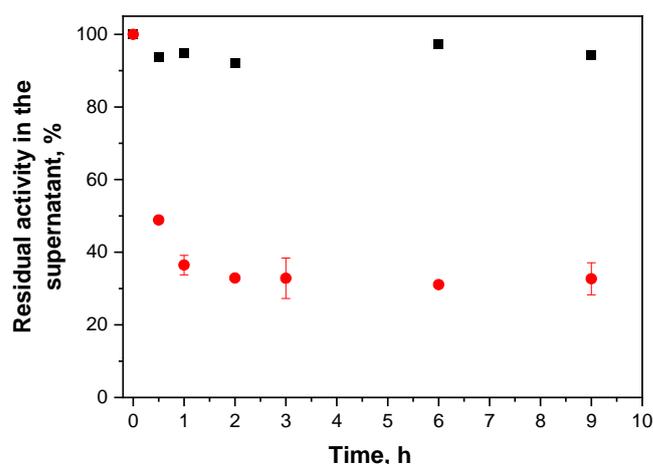
19 A solution was prepared containing PEI (section 3.4.3) and enzyme with a protein:PEI mass ratio
20 of 1:1 in 5mM sodium phosphate buffer. This mixture was incubated at 25 °C and stirred in a 3D
21 roller agitator (Kasvi, São José dos Pinhais, PR, Brazil) at 150 rpm. After 60 min, an aliquot of the
22 enzyme solution was added to a SMNPs suspension to a SMNPs/enzyme mass ratio of 3:1, and 5 mM
23 sodium phosphate buffer pH 7.0 was added to a final volume of 1 mL. Then, 3 mL of 5 mM sodium
24 phosphate buffer pH 7.0 were added instead of ethanol (precipitating agent in the CLEA
25 preparation). This mixture was incubated at 25 °C and stirred in a 3D roller agitator (Kasvi, São José
26 dos Pinhais, PR, Brazil) at 150 rpm. A solution control was prepared under the same conditions,
27 except for no addition of SMNPs. Hydrolytic activities (using tributyrin as substrate, section 3.2) were
28 measured in the supernatant and enzyme control throughout the reaction. The residual activity in
29 the supernatant was calculated according to the Eq. S1:

$$\text{Residual activity in the supernatant} = \frac{A_s}{A_{ci}} \times 100, \quad (\text{S1})$$

30 where A_s is the activity of the supernatant and A_{ci} is the initial activity of the solution control.

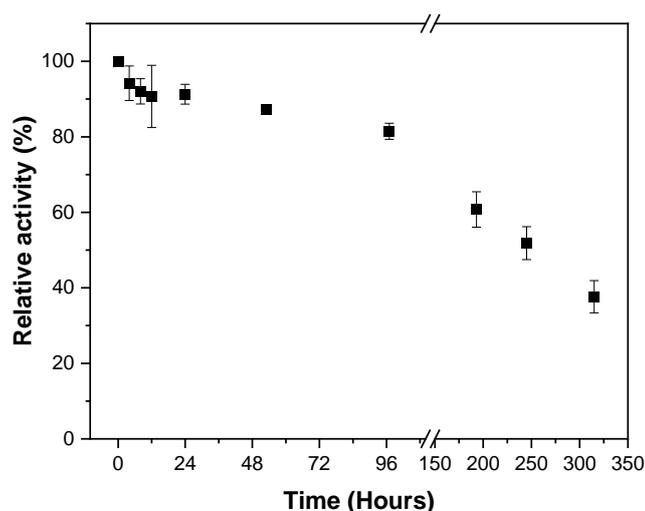
31 For an adsorption assay of the enzyme non-treated with PEI, 5 mg of protein was offered to 1 g
32 of SMPNs in 10 mL of 5 mM sodium phosphate buffer, pH 7.0. This mixture was incubated at 25 °C
33 and stirred in a 3D roller agitator (Kasvi, São José dos Pinhais, PR, Brazil) at 150 rpm. The solution
34 control was prepared under the same conditions, except for no addition of SMNPs. During the
35 reaction, hydrolytic activities (using tributyrin as substrate, section 3.2) were measured in the
36 supernatant and in the solution control throughout the reaction. The residual activity in the
37 supernatant was calculated according to the Eq. S1.

38 The enzyme non-treated with PEI was fully adsorbed on SMNPs after 30 min reaction, while the
39 enzyme treated with PEI was slowly adsorbed (Figure S1), but after 30 min reaction around 50% of
40 the enzyme was adsorbed on the SMNPs, reaching a adsorption plateau of around 70%.



41

42 **Figure S1.** Adsorption profile of Eversa Transform 2.0 treated with polyethyleneimine (Section 3.4.3)
43 on silica magnetic nanoparticles (SMNPs) functionalized with octyl and amine groups at 25 °C under
44 150 rpm stirring (3D roller agitator). Hydrolytic activities (using tributyrin as substrate, Section 3.2)
45 were measured in a solution control (■) and in the supernatant (●).

46 **2. Inactivation profile of liquid Eversa Transform 2.0**

47

48 **Figure S2.** Profile of inactivation of liquid Eversa® Transform 2.0 at 60 °C and pH 7.0 (100 mM sodium
 49 phosphate buffer) using an enzyme concentration of 5 mg protein/mL. The activity of the enzyme
 50 initial solution was taken as 100%. Hydrolytic activities were measured with tributyrin as substrate
 51 (Section 3.2).

52 **3. Water content in transesterification reactions of soybean oil with ethanol using liquid and**
 53 **magnetic CLEA of Eversa Transform 2.0 (Eversa-mCLEA)**

54 **Table S1.** Experimental conditions (enzyme load and water content) for transesterification reactions
 55 using liquid and immobilized Eversa.

Biocatalyst	Eversa-mCLEA				Liquid Eversa	
Enzyme load (U_{est}/g_{oil})	4.0	7.0	7.0	12.0	7.0	7.0
Water (% w/w _{oil})	6.1 ^a	6.9 ^a	1.6 ^b	2.8 ^b	2.8 ^b	6.5 ^a

56 ^a Amount of water present in the reaction (water from the biocatalyst plus water added to the reaction).

57 ^b Reactions without adding water. The water present in these reactions comes from the biocatalyst.

58 Reaction conditions: 15 g refined soybean oil, oil/ethanol molar ratio of 1:6, 40 °C, 1700-2000 rpm stirring in a
 59 vortex flow reactor. Biocatalyst activity (esterification activity, U_{est}/g): liquid Eversa = 106.36, Eversa-mCLEA =
 60 106.17. Water content determined by Karl Fischer method.

61



© 2020 by the authors. Submitted for possible open access publication under the terms
 and conditions of the Creative Commons Attribution (CC BY) license
 (<http://creativecommons.org/licenses/by/4.0/>).

62