

# Immunomodulatory effects of endoscopic ultrasound-guided thermal ablation in patients with pancreatic ductal adenocarcinoma

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## Supplementary Material

### 1. Peripheral blood samples processing

PBMCs were isolated from fresh heparinized blood samples, previously diluted with buffered saline phosphate (PBS; ratio of 1 volume of PB for 2 volumes of PBS), by Ficoll-Hypaque density gradient (GE Healthcare Srl), after stratification on Ficoll-Hypaque (ratio of 3 volumes of PB for 1 volume of Ficoll-Hypaque) and centrifugation at 2500 x g for 20 minutes. The obtained PBMCs were subjected to repeated washing with buffered saline phosphate (PBS) and resuspended in a Roswell Park Memorial Institute (RPMI) 1640 medium integrated with 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES) 25 mmol/L, L-glutamine 2 mmol/L, Gentamicin 50 µg/mL and 8% human serum. PBMCs were then immediately titled through an automatic cell counter and frozen in liquid nitrogen in a medium consisting of 90% fetal bovine serum (FBS) and 10% dimethyl sulfoxide (DMSO).

Serum samples were obtained through centrifugation of PB samples at 3000 rpm for 20 minutes and immediately frozen in liquid nitrogen.

### 2. Cell samples processing for flow-cytometry analysis

The following five flow-cytometric panels were designed:

- Panel I: CX3CR1-FITC (clone: 2A9-1); CD115-PE (clone: 9-4D2-1E4); CCR2-PerCP-Cy5.5 (clone: K036C2); CD45-PE/Cy7 (clone: HI30); CD16-APC (clone: 3G8); CD14-APC/Cy7 (clone: M5E2); CD11b-Pacific Blue (clone: ICRF44); HLA-DR-Brilliant Violet 510 (clone: L243).
- Panel II: CXCR3-FITC (clone: G025H7); CCR6-PE (clone: 11A9); PD1-PerCP/Cy5.5 (clone: EH12.2H7); CXCR5-PE/Cy7 (clone: J252D4); CD45RO-APC (clone: UCHL1); ICOS-Super Bright 436 (clone: ISA-3); CD4-Brilliant Violet 510 (clone: A161A1).
- Panel III: CD8a-FITC (clone: HIT8a); CD25-PE (clone: RPA-T4); CD127-PerCP/Cy5.5 (clone: HIL-7R-M21); CD62L-PE/Cy7 (clone: DREG-56); SLAMF7 (CD319)-APC (clone: 162.1); CD27-APC/Cy7 (clone: O323); CD45RO-Pacific Blue (clone: UCHL1); CD4-Brilliant Violet 510 (clone: A161A1).
- Panel IV: CD14-Alexa Fluor 488 (clone: 63D3); CD138-PE dazzle (clone: MI15); CD45RB-PE (clone: MEM-55); IgM-Brilliant Violet 650 (clone: G20-127); CD21-Brilliant Ultra Violet 496 (clone: B-ly4); CD27-Brilliant Ultra Violet 737 (clone: L128); CD3-PerCP-Cy5.5 (clone: HIT3a); CD10-Alexa Fluor 700 (clone: HI10a); IgD-Brilliant Violet 421 (clone: IA6-2); CD38-Brilliant Violet 510 (clone: HB-7); CD19-Brilliant Violet 605 (clone: HIB19); CD73-Brilliant Ultra Violet 395 (clone: AD2); CXCR5-PE/Cy7 (clone: J252D4); CD11c-APC (clone: B-ly6).
- Panel V: CD14-Alexa Fluor 488 (clone: 63D3); CD3-PerCP-Cy5.5 (clone: HIT3a); CD24-APC/Cy7 (clone: ML5); CD38-Brilliant Violet 510 (clone: HB-7); CD19-Brilliant Violet 605 (clone: HIB19); CD27-Brilliant Ultra Violet 737 (clone: L128); TGFβ1-PE (clone: TW4-9E7); IL10-APC (clone: JES3-19F1).

After thawing, stored PBMCs suspensions were revitalized in a cell medium consisting of RPMI 1640-GlutaMax medium (Gibco), 10% heated-inactivated FBS (Euroclone SpA), 1% P/S (Gibco), 1% non-essential aminoacids (100x) (GibcoTM), 1 mM Sodium-pyruvate (100x) (Gibco), 50 uM 1-mercaptoethanol (GibcoTM) and 1% Penicillin-Streptomycin mixture (PenStrep, Gibco). Subsequently, revitalized PBMCs were counted adding Trypan blue (dilution 1:4) and resuspended at  $5 \times 10^6$  cells/ml.

After repeated washing with PBS, all PBMCs suspensions were live/dead single stained with Viakrome 808 fixable viability dye (Beckman Coulter, 2.5  $\mu$ l for  $5 \times 10^6$  cells) for identification of viable cells, and incubated for 20 minutes in dark room. Therefore, incubation with human FcR blocking reagent (Miltenyi Biotec) for 10 minutes at 4°C and staining with titled surface mAbs for 10 minutes in dark room were performed. After washing of fluorochrome-labeled PBMCs with a FACS buffer, consisting of Dulbecco's phosphate-buffered saline (DPBS) without  $\text{Ca}^{2+}$  e  $\text{Mg}^{2+}$  and 2% FBS, they were incubated with Fixation buffer (ThermoFisher Scientific, 200  $\mu$ l) for 20 minutes at 4°C.

PBMCs suspensions intended for immunophenotyping of B regulatory cells, before staining with the fixable viability dye, were first incubated for 5 hours at 37°C with CpG-ODN 2006 (Aurogene, 1  $\mu$ l for  $0.5 \times 10^6$  cells), Ionomycin (Sigma-Aldrich, 1  $\mu$ l), Phorbol 12-myristate 13-acetate (PMA, Sigma-Aldrich, 1  $\mu$ l) and GolgiStop (Beckton Dickinson, 1  $\mu$ l). For the intracellular staining, after performing surface staining, cells were incubated with the Cytotfix/Cytoperm fixation/permeabilization solution (Beckton Dickinson, 200  $\mu$ l) for 20 minutes at 4°C in dark room, washed with the Perm/Wash Buffer (Beckton Dickinson, 1 ml diluted 1:10 in  $\text{H}_2\text{O}$ ), incubated with the intracellular mAbs for 30 minutes in dark room, and still washed with the Perm/Wash Buffer.

All stained PBMCs were then resuspended in 200  $\mu$ l of FACS buffer and stored at 4°C in dark room until acquisition using the CytoFLEX LX Flow Cytometer.

For compensation adjustments the CompBeads Compensation Particles Set (Beckton Dickinson) was used. Before each flow cytometric acquisition, an instrument calibration was performed using the SpheroTM Rainbow Calibration Particles (Spherotech Inc.) for long-term performance tracking.

### 3. Serum samples processing for multiplex immunoassays

After thawing samples, 25  $\mu$ g of diluted serum (1:4 for the 37-BioPlex and 27-BioPlex assays, 1:16 for the 3-BioPlex assay) and an eight-point standard curve (plus blank) provided by the commercial assays were analysed for each of the different factors.

Using the 27-BioPlex and 37-BioPlex assays, diluted serum was incubated with the antibody-conjugated magnetic beads at room temperature for 1 hour on a plate shaker at  $850 \pm 50$  rpm. Using the 3-BioPlex assay, serum samples were first activated by adding 1 N-HCL (1:5) for 10 minutes at room temperature and neutralized by adding same volume of the 1.2 N NaOH/0.5 M HEPES buffer base. Diluted samples were then incubated with the antibody-conjugated magnetic beads on plate shaker at  $850 \pm 50$  rpm at room temperature for 2 hours.

Bead-complexes were then incubated with the detection antibody diluent (25  $\mu$ l) for 30 minutes (27-BioPlex and 37-BioPlex) or 1 hour (3-BioPlex) and with streptavidin-phycoerythrin (50  $\mu$ l) for 10 minutes (27-BioPlex and 37-BioPlex) or 30 minutes (3-BioPlex) on a plate shaker at  $850 \pm 50$  rpm at room temperature, after rinsing three times before each of these passages. After washing three times, beads were re-suspended in the assay buffer (125  $\mu$ l) for 30 seconds on a plate shaker at  $850 \pm 50$  rpm at room temperature and, thus, immediately read.

## Supplementary Tables

**Table S1.** Detailed immune phenotypes of cell subsets analysed by flow-cytometry.

	Immune cell subsets
<b>Panel I</b>	<p><i>CD45+CD115+CD11b+HLA-DR+</i> monocytes</p> <p><i>CD45+CD115+CD11b+HLA-DR-</i> immunosuppressive monocytes</p> <p><i>CD45+CD115+CD11b+HLA-DR+CD16-CD14+</i> classical monocytes</p> <p><i>CD45+CD115+CD11b+HLA-DR+CD16+CD14+</i> intermediate monocytes</p> <p><i>CD45+CD115+CD11b+HLA-DR+CD16+CD14<sup>low</sup></i> non-classical monocytes</p> <p><i>CD45+CD115+CD11b+HLA-DR+CD16+CX3CR1<sup>high</sup></i> resident monocytes</p> <p><i>CD45+CD115+CD11b+HLA-DR+CD16-CX3CR1<sup>low</sup>CD14+CCR2+</i> inflammatory monocytes</p>
<b>Panel II</b>	<p><i>CD4+</i> T cells</p> <p><i>CD4+CD45RO+</i> T central memory cells</p> <p><i>CD4+CD45RO+CD62L-CD27-</i> T effector memory cells</p> <p><i>CD4+CD45RO+CD62L-CD27-CD319(SLAMF7)+</i> T <i>CD4+</i> cytotoxic cells</p> <p><i>CD8+CD45RO+CD62L-CD27-CD319(SLAMF7)+</i> T <i>CD8+</i> cytotoxic cells</p> <p><i>CD4+CD127<sup>low</sup>CD25+</i> T regulatory cells</p>
<b>Panel III</b>	<p><i>CD4+CD45RO+CXCR3+CCR6-</i> T helper 1 cells</p> <p><i>CD4+CD45RO+CXCR3-CCR6-</i> T helper 2 cells</p> <p><i>CD4+CD45RO+CXCR3-CCR6+</i> T helper 17 cells</p> <p><i>CD4+CD45RO+CXCR3+CCR6+</i> T helper 1/17 cells</p> <p><i>CD4+CD45RO+CXCR5+</i> T follicular helper cells</p> <p><i>CD4+CD45RO+CXCR5+CXCR3+CCR6-</i> T follicular helper 1 cells</p> <p><i>CD4+CD45RO+CXCR5+CXCR3-CCR6-</i> T follicular helper 2 cells</p> <p><i>CD4+CD45RO+CXCR5+CXCR3-CCR6+</i> T follicular helper 17 cells</p> <p><i>CD4+CD45RO+CXCR5+CXCR3+CCR6+</i> T follicular helper 1/17 cells</p> <p><i>CD4+CD45RO+CXCR5+ICOS+PD-1+</i> activated T follicular helper cells</p>
<b>Panel IV</b>	<p><i>CD3-CD14-CD19+</i> B cells</p> <p><i>CD3-CD14-CD19+CD38+CD27+</i> antibody secreting B cells</p> <p><i>CD3-CD14-CD19+CD38+CD27+CD138-</i> antibody secreting plasmablasts</p> <p><i>CD3-CD14-CD19+CD38+CD27+CD138+</i> antibody secreting plasmacells</p> <p><i>CD3-CD14-CD19+CD38-CD27+</i> memory B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27+IgD+</i> unswitched memory B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27+IgD-</i> switched memory B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27+IgD-CD11c+CD21-</i> activated switched memory B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD-</i> double negative B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD-CD27-CXCR5+CD11c-</i> double negative 1 B cells</p>

	<p><i>CD3-CD14-CD19+CD38+/-CD27-IgD-CD27-CXCR5-CD11c+</i> double negative 2 B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD-CD27-CXCR5-CD11c-</i> double negative 3 B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD-CD27-CXCR5+CD11c+</i> double negative 4 B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD+</i> naïve B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD+CD10-CD45RB-CD21-CD11c+</i> activated naïve B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD+CD10+CD45RB-</i> transitional 1/2 B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD+CD10-CD45RB+</i> marginal zone peripheral B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD+CD27-CD10-CD45RB-CD21+/-CD11c-CXCR5+CD73+</i> follicular B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD+CD27-CD10-CD45RB-CD21+/-CD11c-CXCR5+CD73+CD21-IgM+</i> resting naïve B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD+CD27-CD10-CD45RB-CD21+/-CD11c-CXCR5+CD73+CD21-IgM-</i> anergic naïve B cells</p> <p><i>CD3-CD14-CD19+CD38+/-CD27-IgD+CD27-CD10-CD45RB-CD21+/-CD11c-CXCR5+CD73-</i> transitional CXCR5+ T3a B cells</p> <p><i>CD3-CD14-CD19+CD38+/-IgD+CD27-CD10-CD54RB-CD21+/-CD11c-CXCR5-CD73-</i> transitional CXCR5- T3a B cells</p>
<b>Panel V</b>	<p><i>CD3-CD14-CD19+CD24+CD38-</i> regulatory B cells</p> <p><i>CD3-CD14-CD19+CD24+CD38+</i> immature regulatory B cells</p> <p><i>CD3-CD14-CD19+CD24+CD27+</i> B10 regulatory B cells</p> <p><i>CD3-CD14-CD19+CD38+CD27+</i> antibody secreting regulatory B cells</p> <p><i>CD3-CD14-CD19+CD24+CD38-IL-10+</i> regulatory B cells expressing IL-10</p> <p><i>CD3-CD14-CD19+CD24+CD38+IL-10+</i> immature regulatory B cells expressing IL-10</p> <p><i>CD3-CD14-CD19+CD24+CD27+IL-10+</i> B10 regulatory B cells expressing IL-10</p> <p><i>CD3-CD14-CD19+CD38+CD27+IL-10+</i> antibody secreting regulatory B cells expressing IL-10</p> <p><i>CD3-CD14-CD19+CD24+CD38-TGF-β+</i> regulatory B cells expressing TGF-β</p> <p><i>CD3-CD14-CD19+CD24+CD38+TGF-β+</i> immature regulatory B cells expressing TGF-β</p> <p><i>CD3-CD14-CD19+CD24+CD27+TGF-β+</i> B10 regulatory B cells expressing TGF-β</p> <p><i>CD3-CD14-CD19+CD38+CD27+TGF-β+</i> antibody secreting regulatory B cells expressing TGF-β</p>

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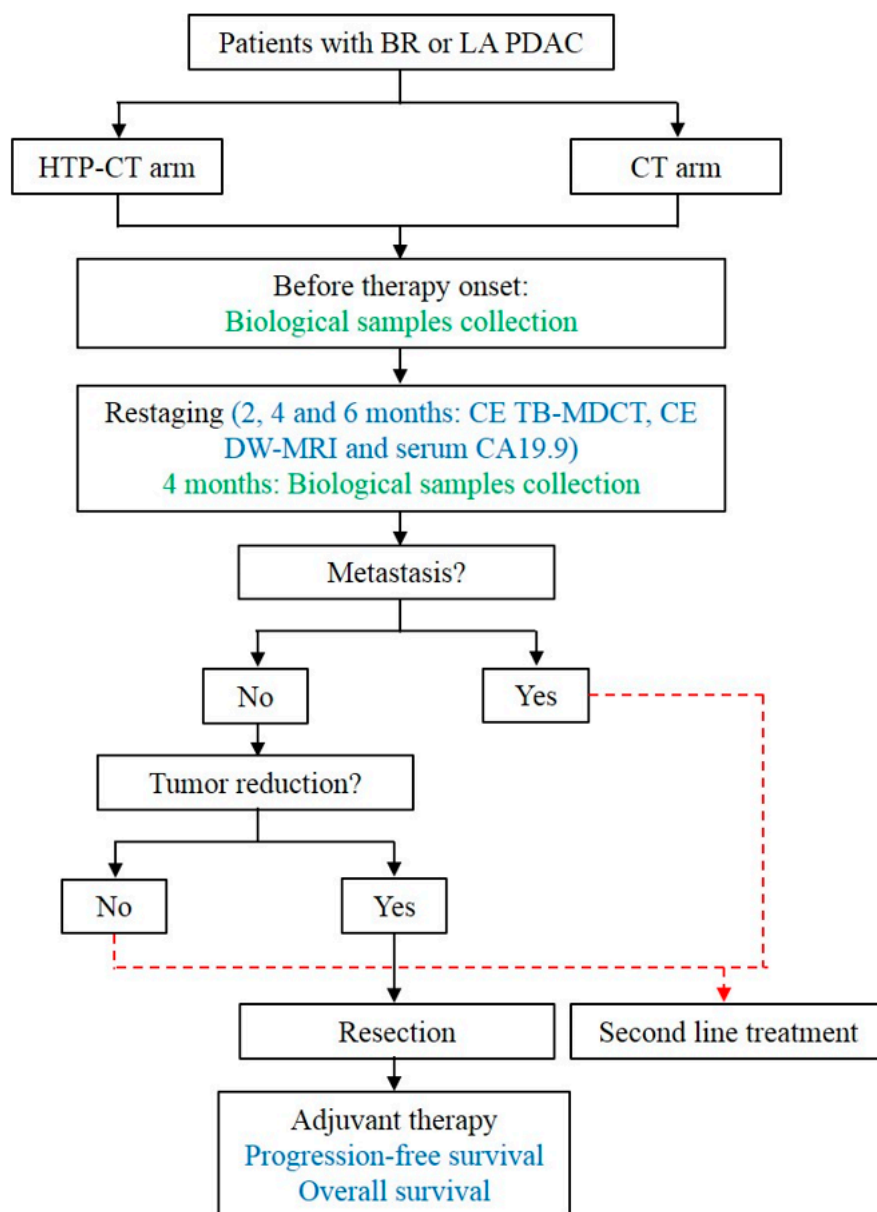


**Table S2.** Detail of soluble analytes quantified by multiplex Luminex assays.

	Soluble analytes		Soluble analytes
<b>Pro Human Inflammation Panel I Assay 37-BioPlex</b>	APRIL / TNFSF13	<b>Pro Human Cytokine Immunoassay 27-BioPlex</b>	FGF basic
	BAFF / TNFSF13B		Eotaxin
	sCD30 / TNFRSF8		G-CSF
	sCD163		GM-CSF
	Chitinase-3-like 1		IFN- $\gamma$
	gp130 / sIL-6R $\beta$		IL-1 $\beta$
	IFN- $\alpha$ 2		IL-1ra
	IFN- $\beta$		IL-2
	IFN- $\gamma$		IL-4
	IL-2		IL-5
	sIL-6R $\alpha$		IL-6
	IL-8		IL-7
	IL-10		IL-8
	IL-11		IL-9
	IL-12 (p40)		IL-10
	IL-12 (p70)		IL-12 (p70)
	IL-19		IL-13
	IL-20		IL-15
	IL-22		IL-17A
	IL-26		IP-10
	IL-27 (p28)	<b>Pro TGF-<math>\beta</math> Immunoassay 3-BioPlex</b>	MCP-1 (MCAF)
	IL-28A / IFN- $\lambda$ 2		MIP-1 $\alpha$
	IL-29 / IFN- $\lambda$ 1		MIP-1 $\beta$
	IL-32		PDGF-BB
	IL-34		RANTES
	IL-35		TNF- $\alpha$
	LIGHT / TNFSF14		VEGF
	MMP-1		TGF- $\beta$ 1
	MMP-2		TGF- $\beta$ 2
	MMP-3		TGF- $\beta$ 3
	Osteocalcin		
	Osteopontin		
	Pentraxin-3		
	sTNF-R1		
	sTNF-R2		
	TSLP		
	TWEAK / TNFSF12		

## Supplementary Figure legends

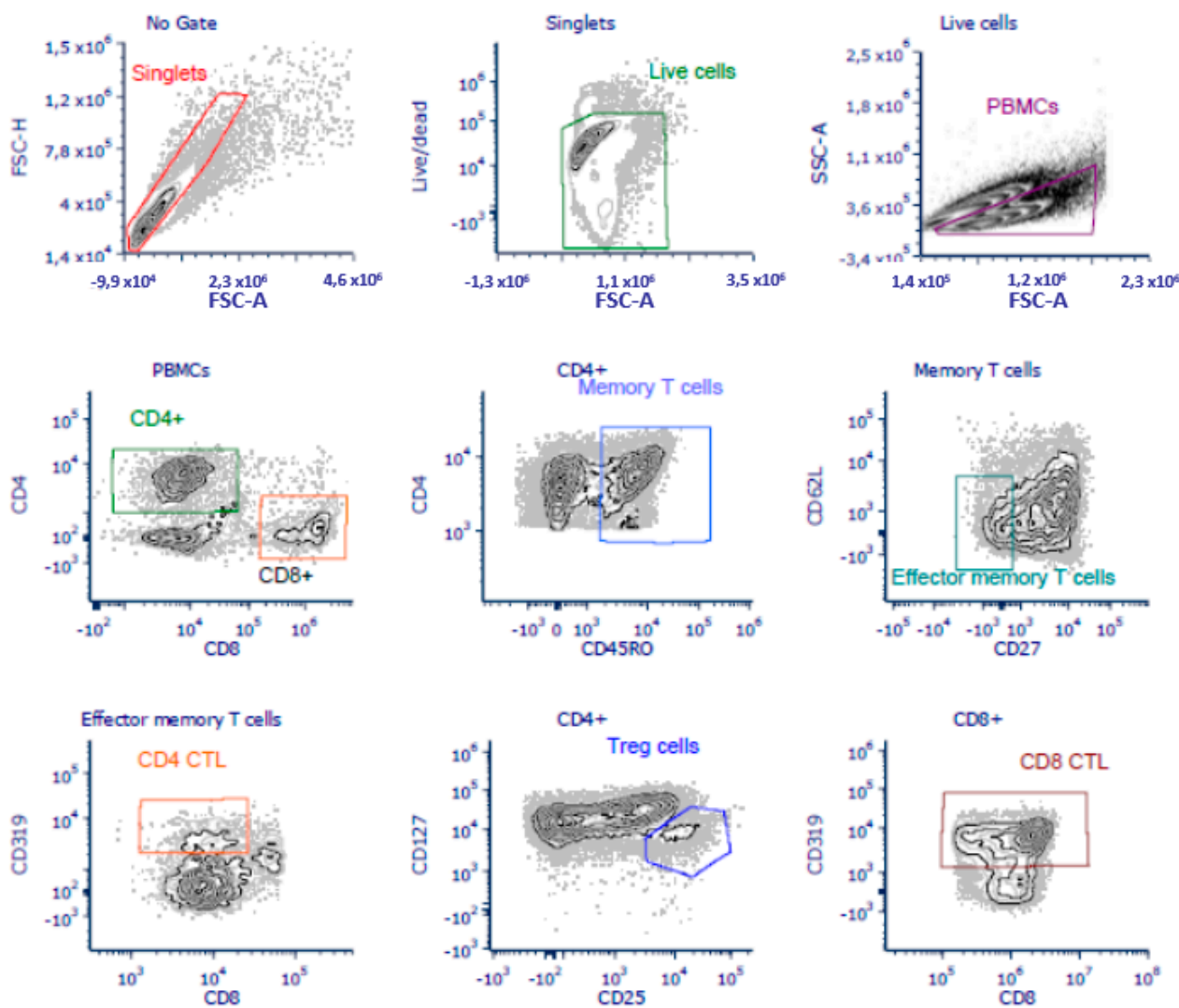
Figure S1. Clinical flow-chart of the randomized controlled study.



**Footnote:** BR: Borderline Resectable; LA: Locally Advanced; PDAC: Pancreatic Adenocarcinoma; HTP-CT: HybridTherm Probe ablation plus Chemotherapy; CT: Chemotherapy; CE: Contrast Enhanced; TB-MDCT: Total Body-MultiDetector Computed Tomography; DW-MRI: Double-Weighed Magnetic Resonance Imaging; CA19.9: Carbohydrate Antigen 19.9.

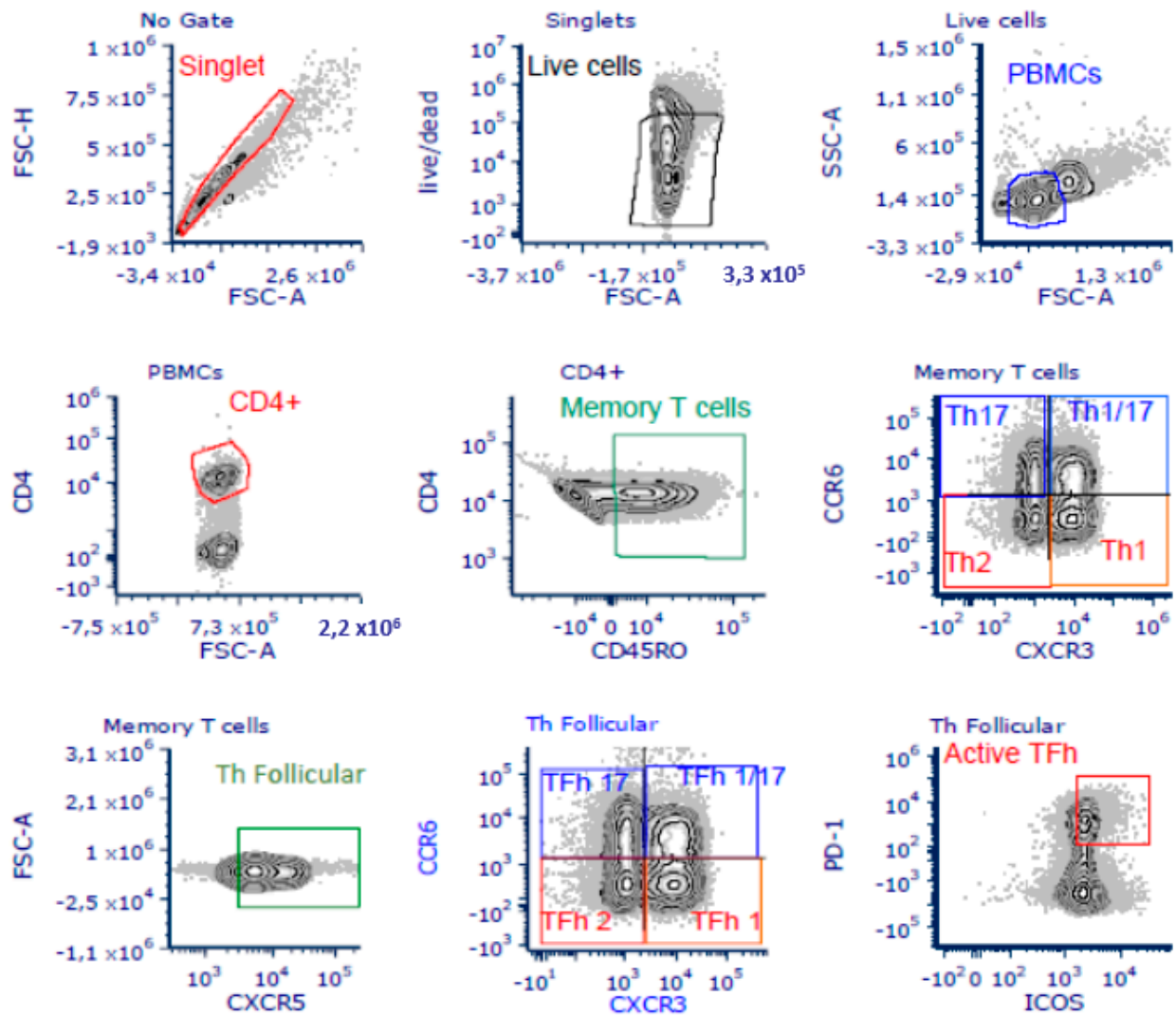
**Figure S2.** Flow-cytometry gating strategies for immunophenotyping of cytotoxic T cells and T regulatory cells (A), T helper and T follicular helper lymphocytes (B), B-cell subsets (C), B regulatory cell subsets (D) and monocyte subsets (E).

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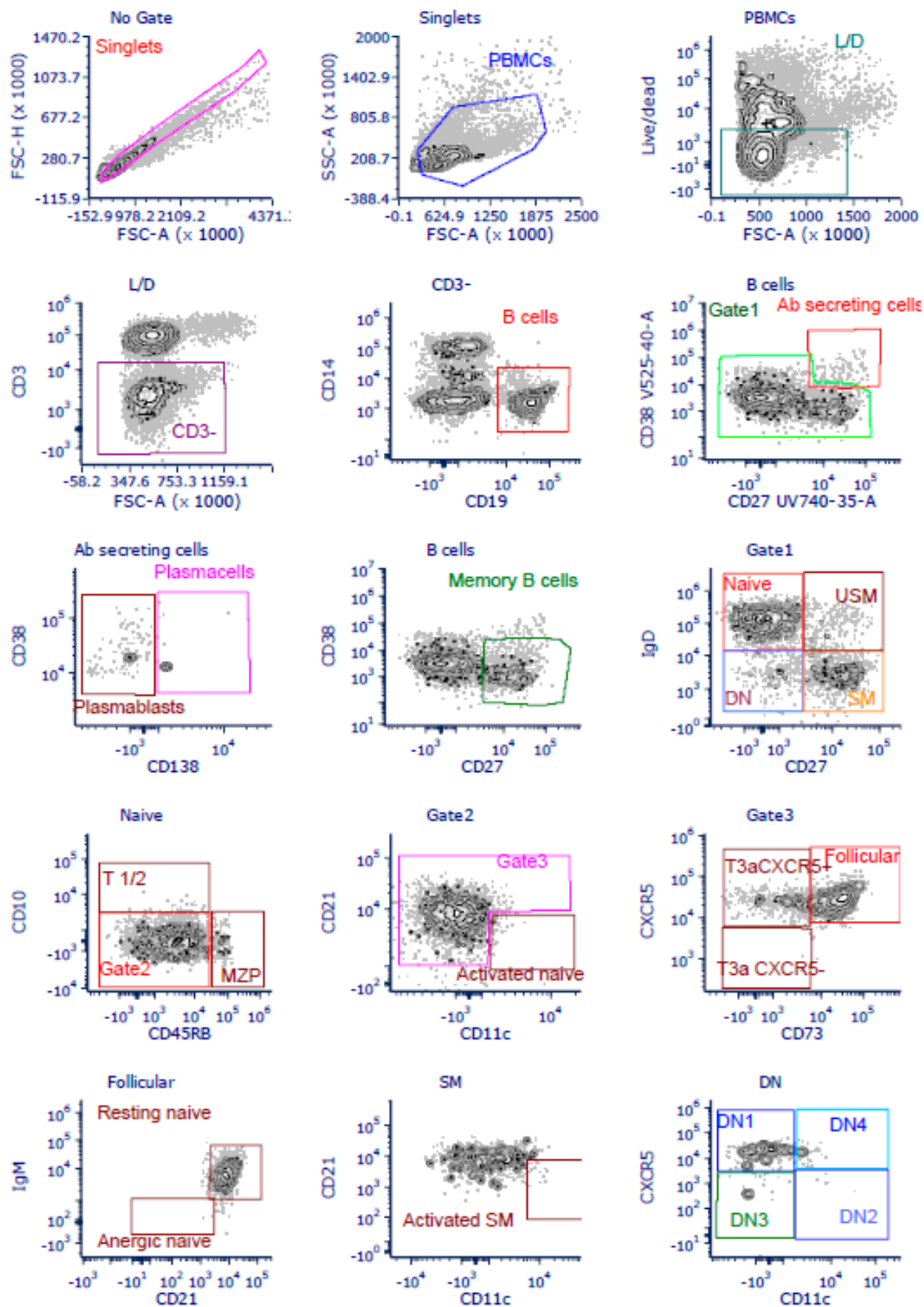
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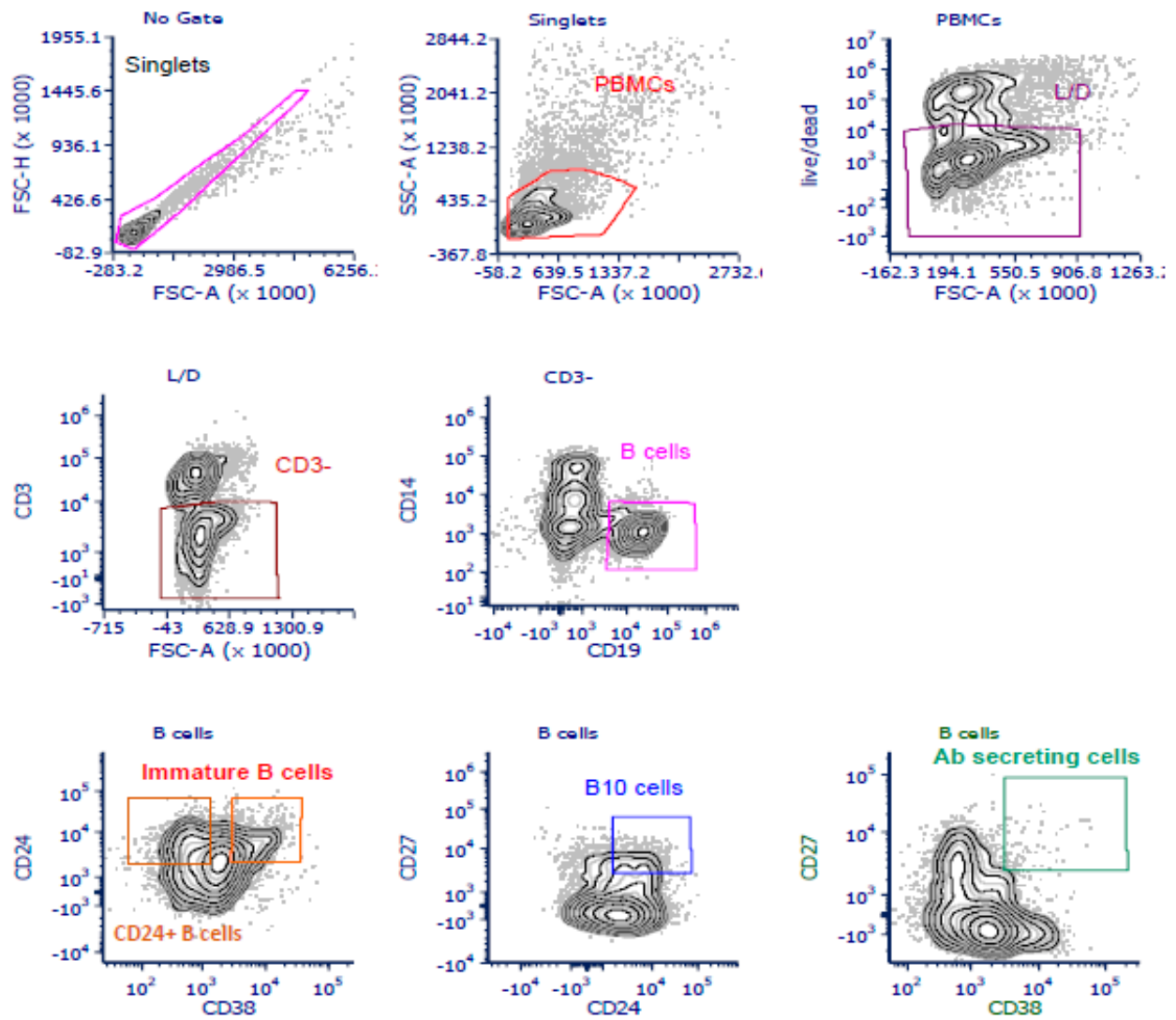
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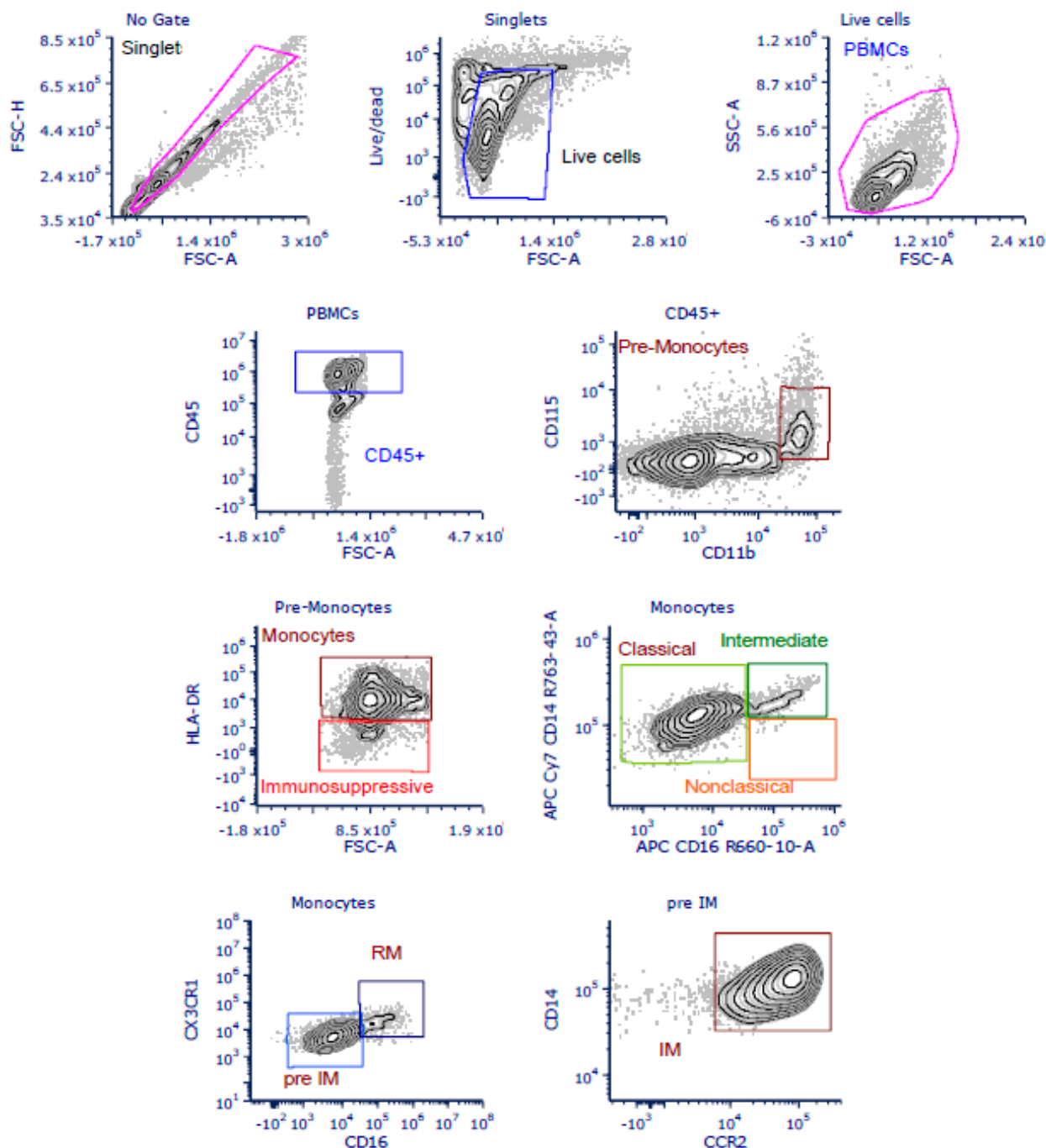
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**Footnote:** PBMCs: peripheral mononucleated cells; CTLs: cytotoxic T lymphocytes; Treg: T regulatory; Th: T helper; Tfh: T follicular helper; L/D: live/dead; Ab: antibody; USM: unswitched memory; SM: switched memory; DN: double negative; T 1/2: transitional 1/2; MZP: marginal zone peripheral; T3a: transitional 3a; RM: resident monocytes; IM: inflammatory monocytes.

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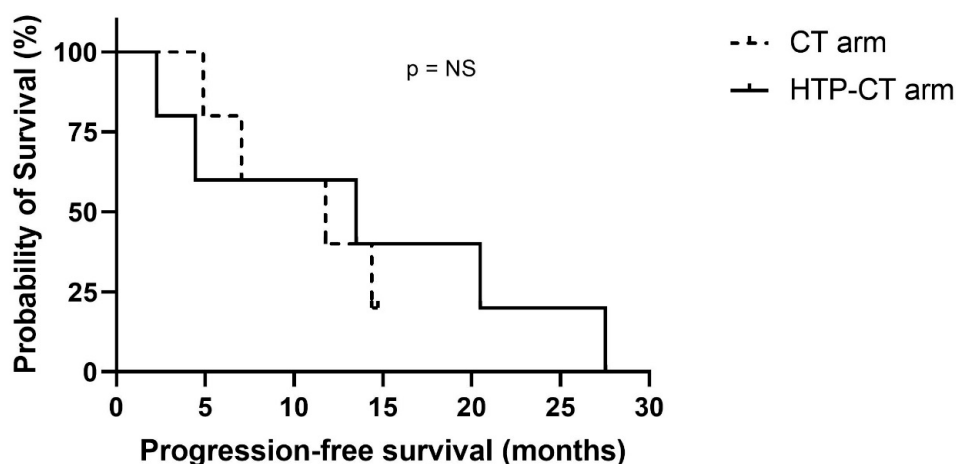
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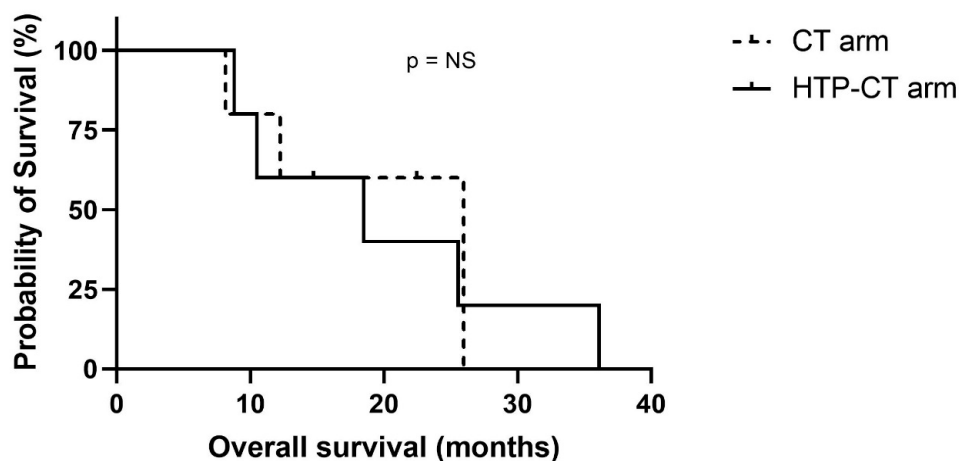
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**Figure S3.** Progression free and overall survival in patients treated with HybridTherm Probe ablation plus chemotherapy (HTP-CT arm) and those with chemotherapy alone (CT arm). Kaplan Meier curves showing the progression-free survival (A) and overall survival (B) of patients with borderline resectable and locally advanced pancreatic ductal adenocarcinoma treated with HybridTherm Probe ablation plus chemotherapy and those treated with chemotherapy only. Survival curves were compared using the Log-rank Mantel-Cox test: not significant (NS) =  $p > 0.05$ .

A

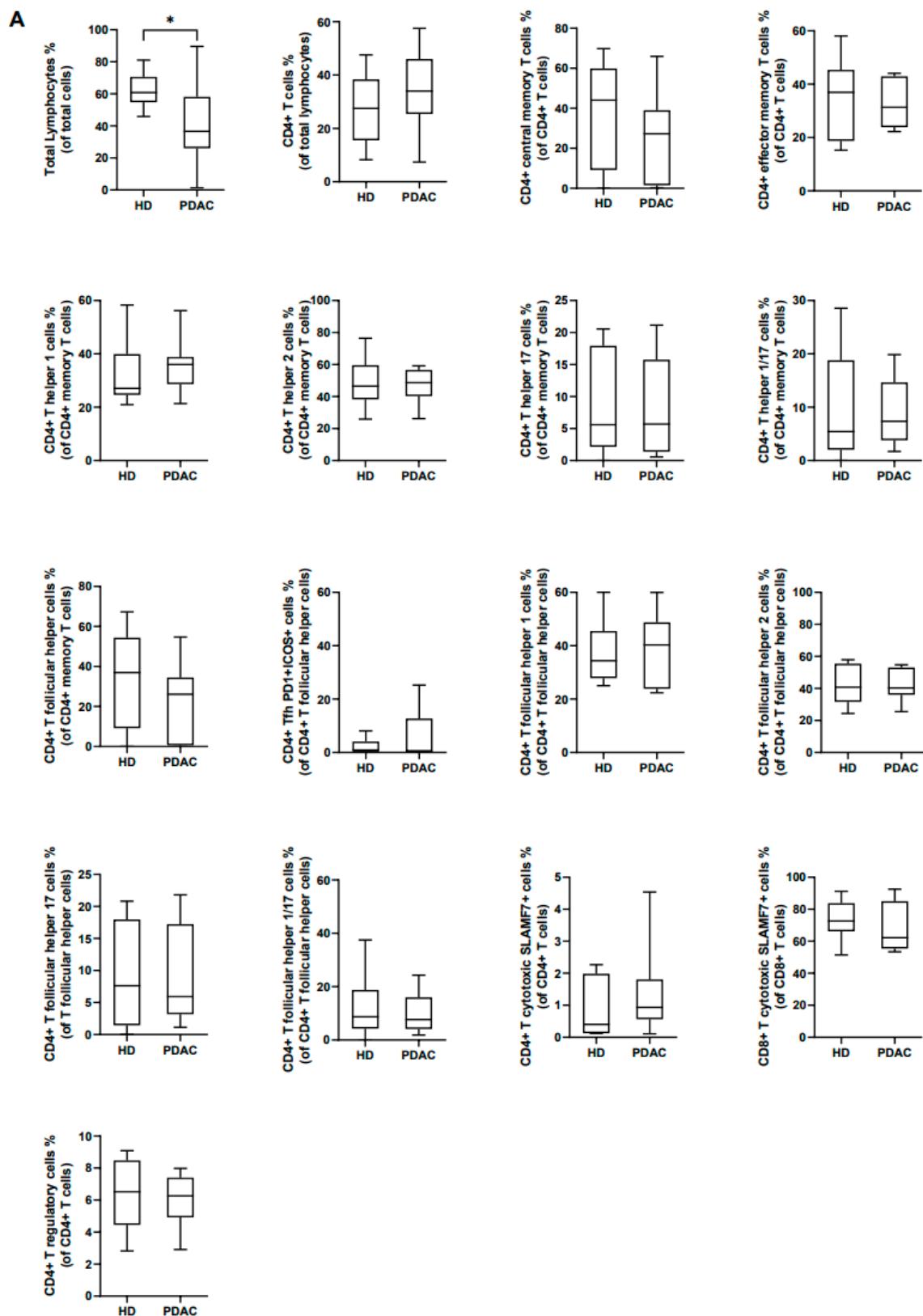


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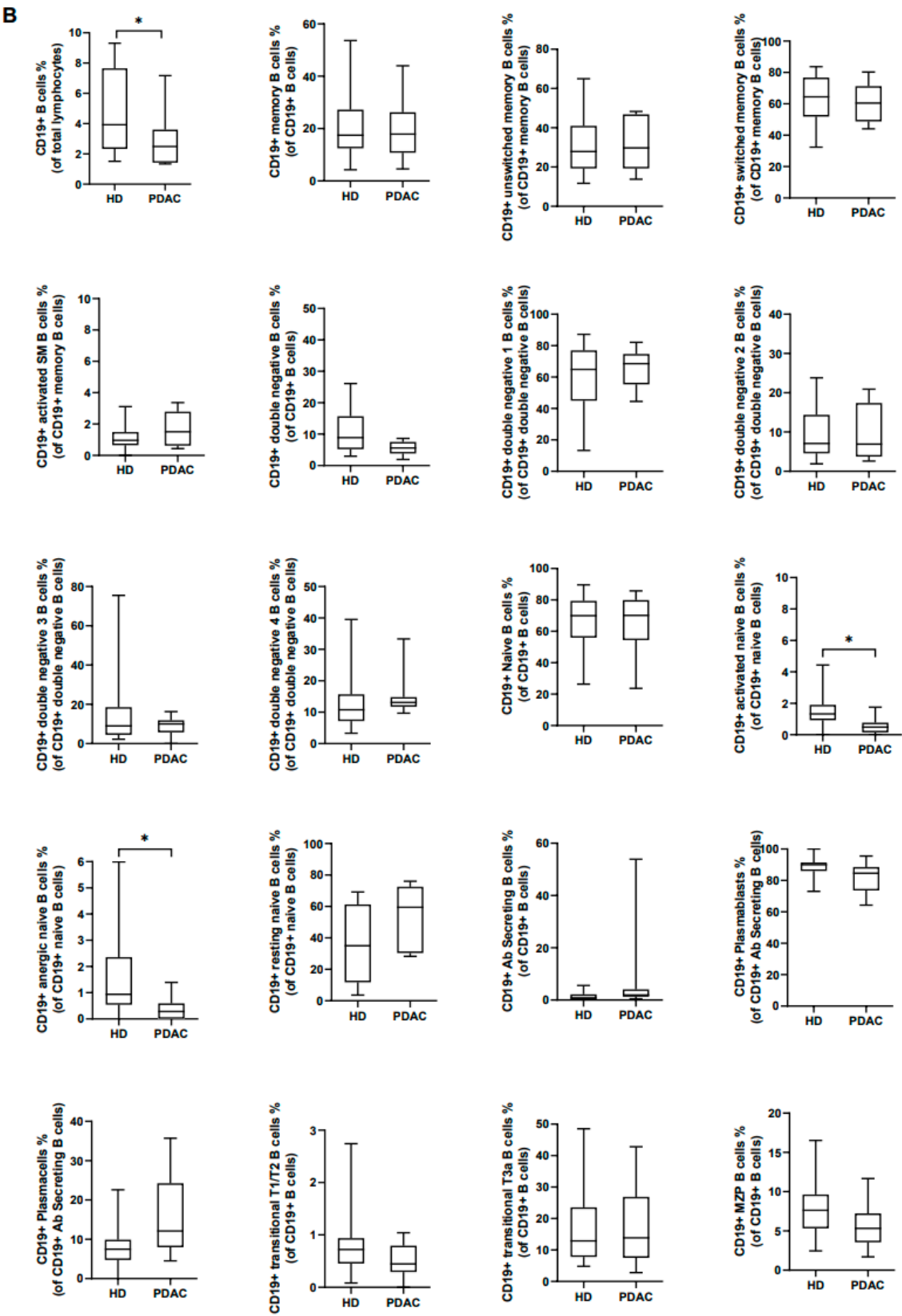


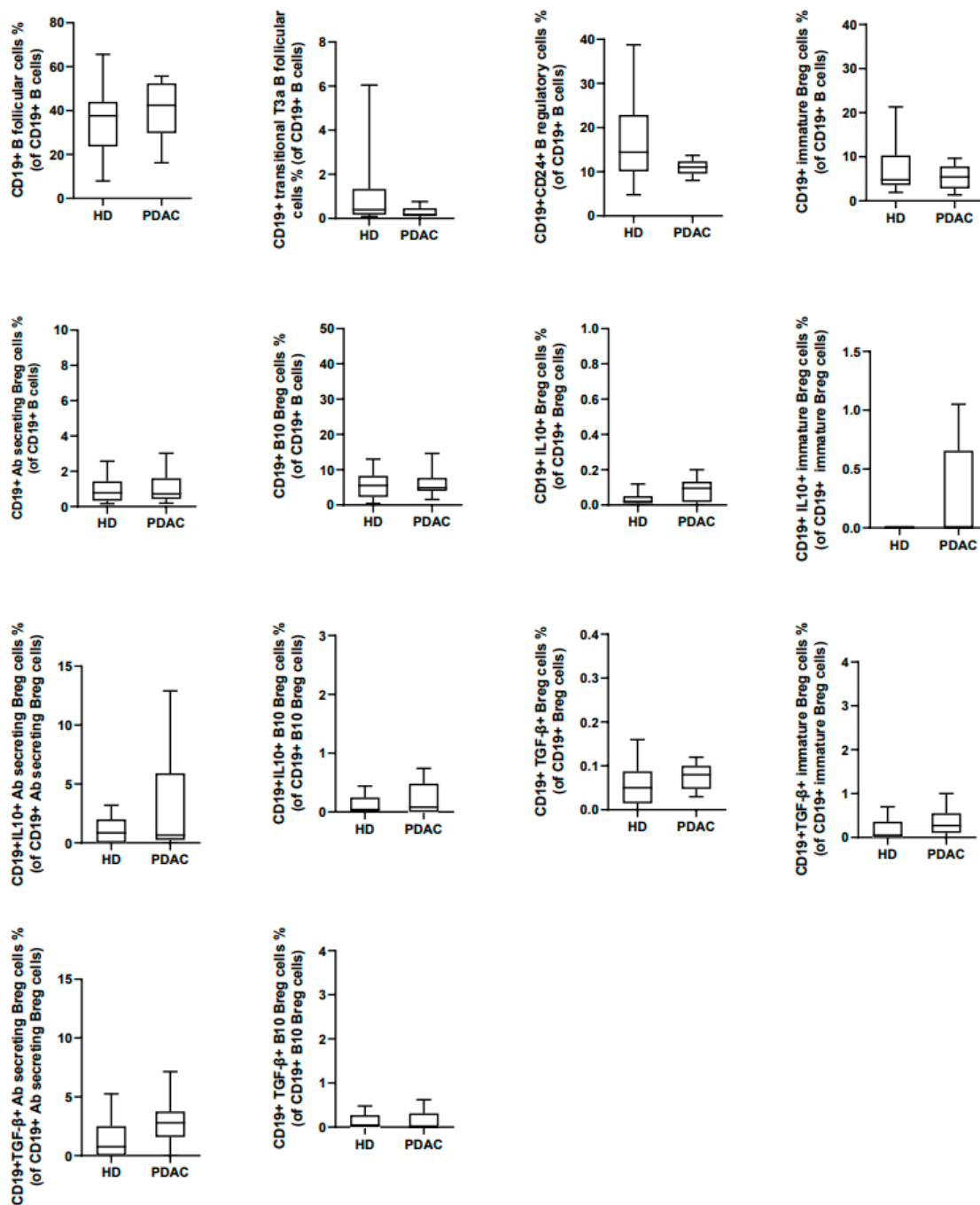


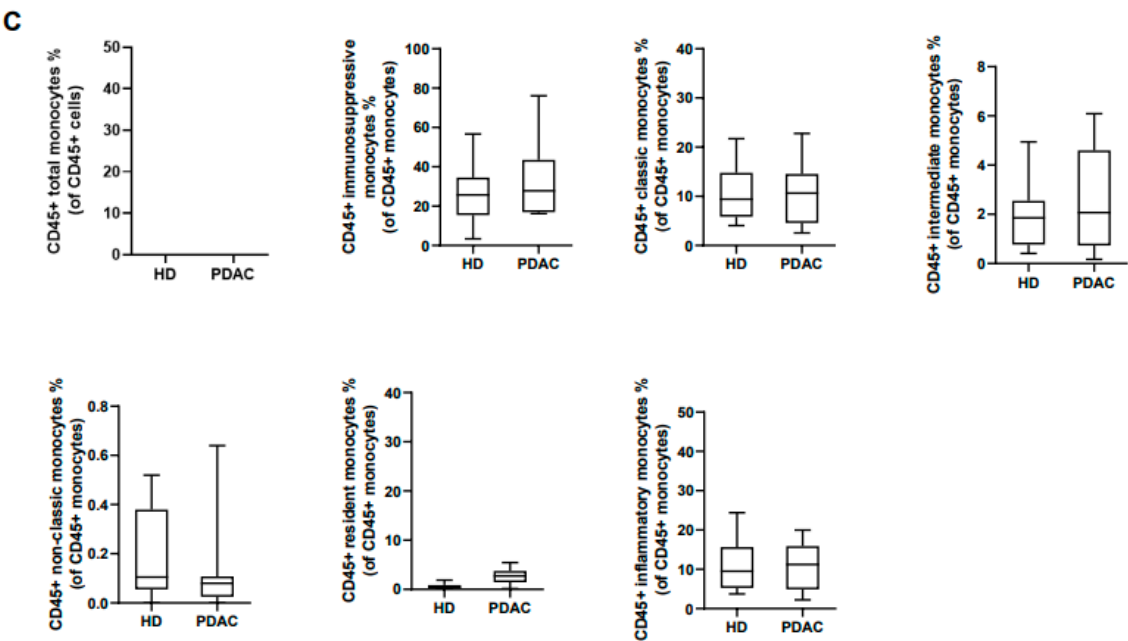
**Figure S4.** Differences of circulating immune cells (**A**: T-cell compartment; **B**: B-cell compartment; **C**: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (**D**: Pro Human Inflammation Panel I Assay; **E**: Pro TGF- $\beta$  Immunoassay; **F**: Pro Human Cytokine Immunoassay) between patients with pancreatic adenocarcinoma (PDAC) and sex- and age-matched healthy donors (HD) at baseline. Inter-group variables were compared using the Welch two sample t-test: \* =  $p < 0.05$ .



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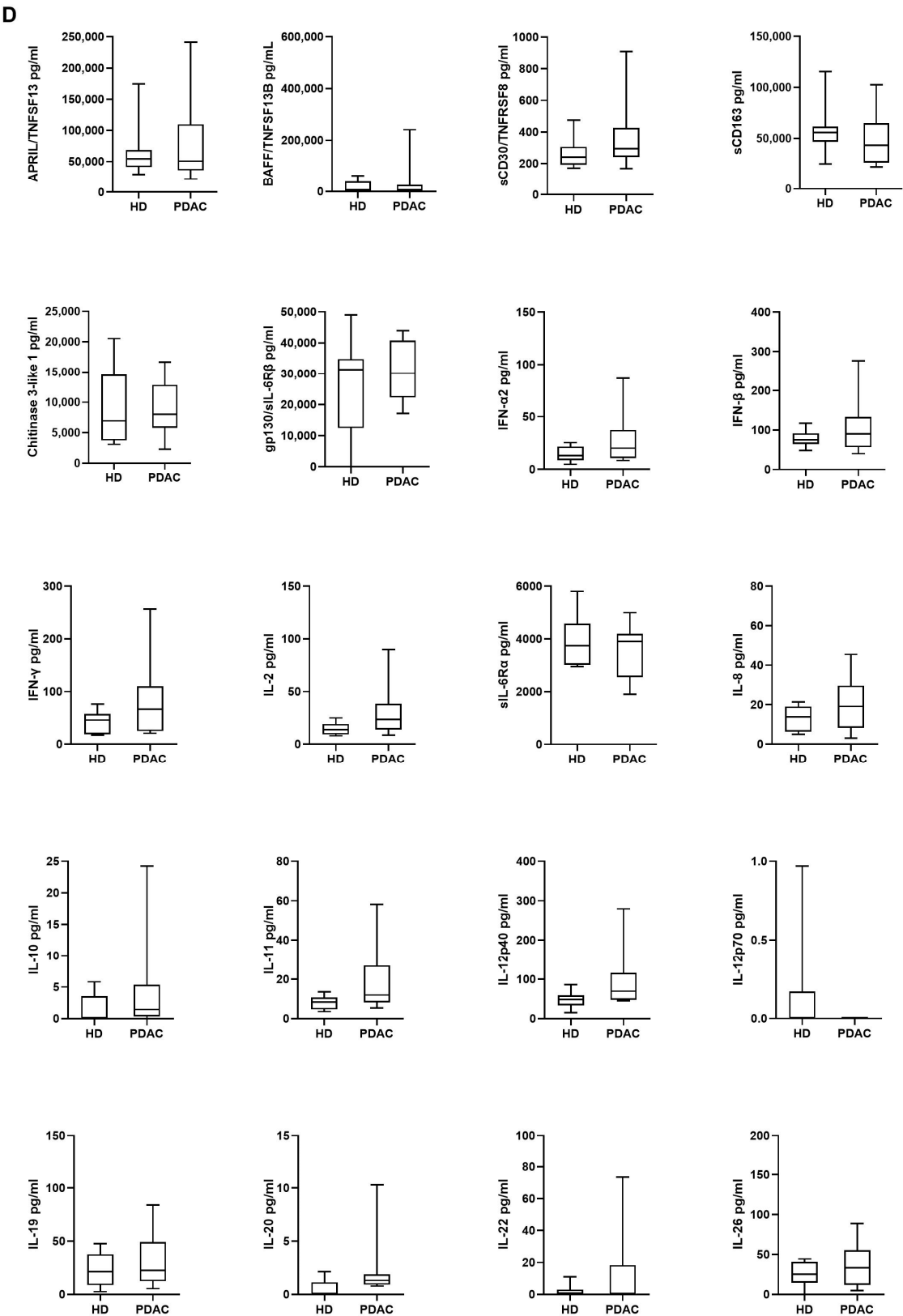


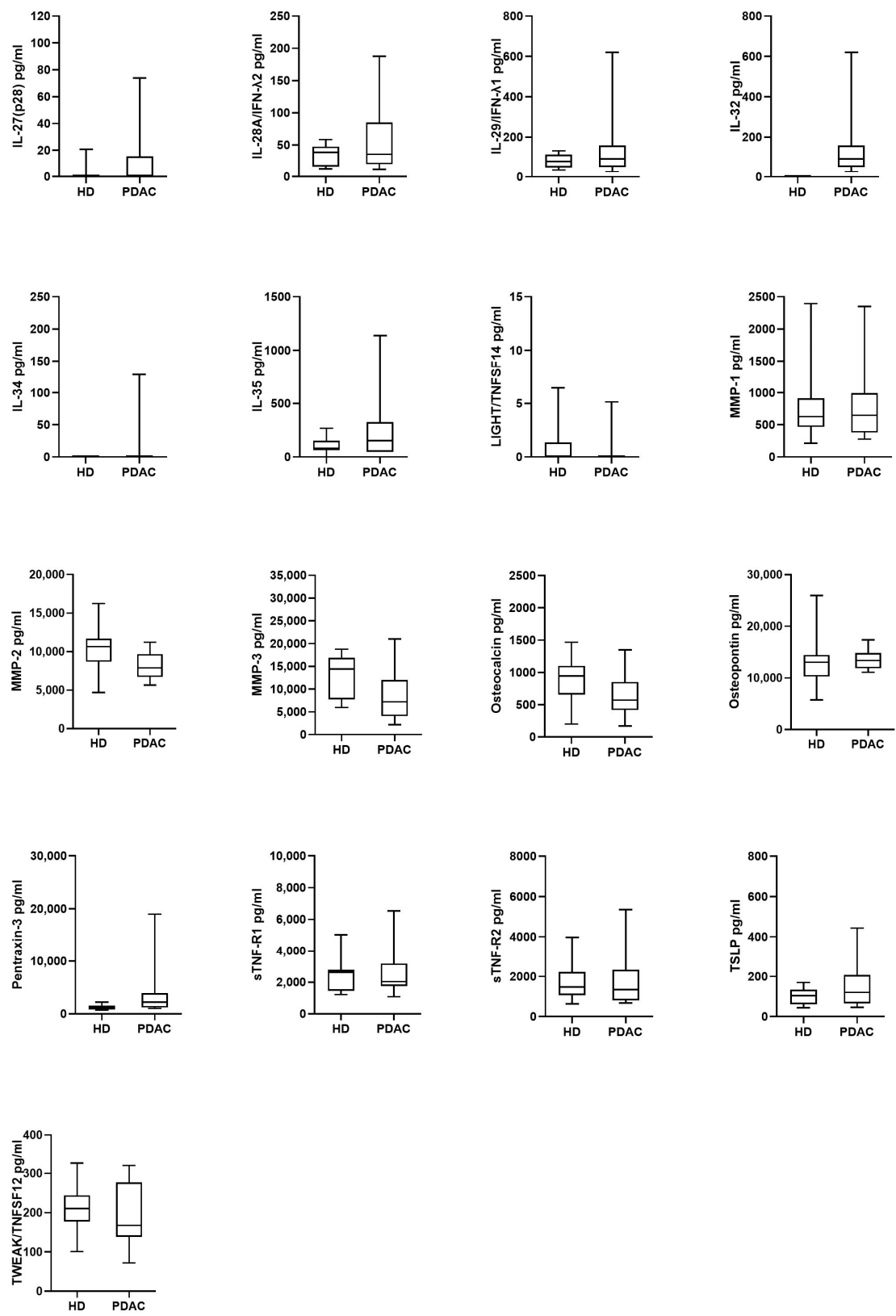


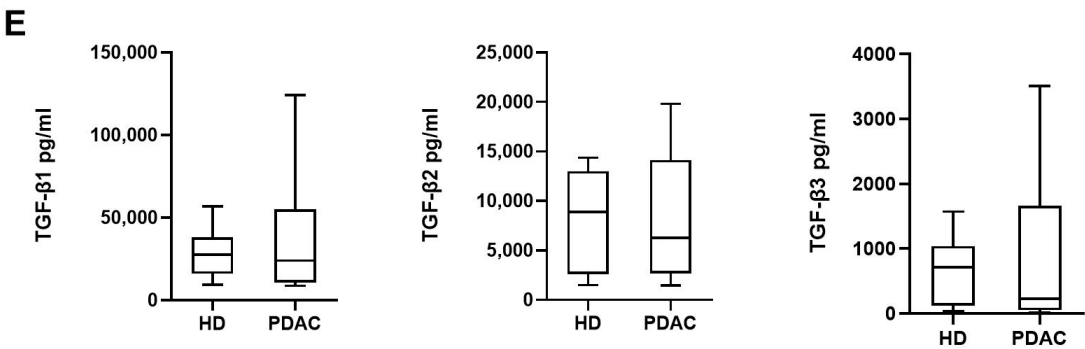


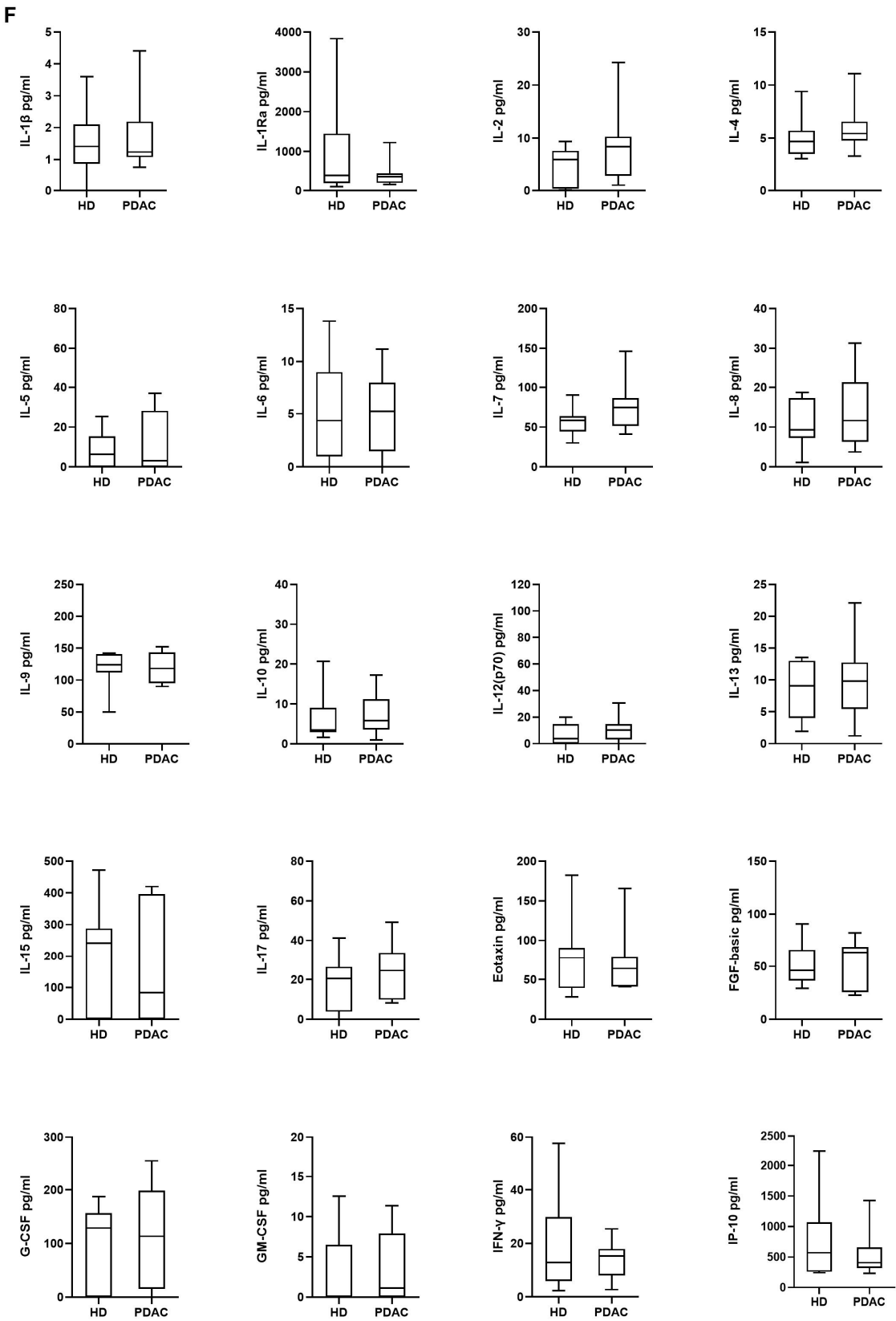
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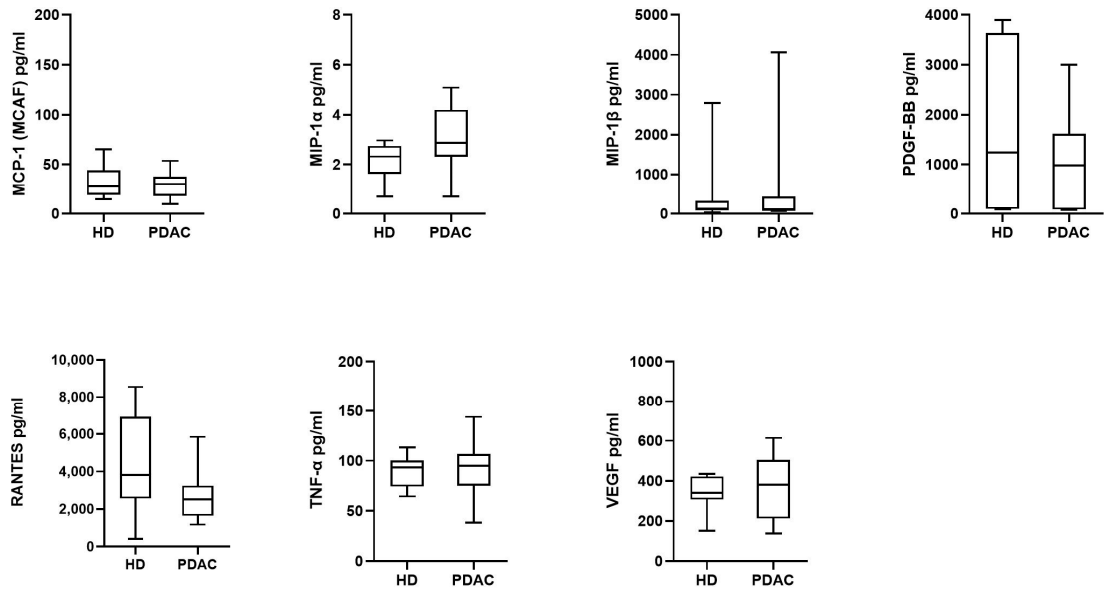


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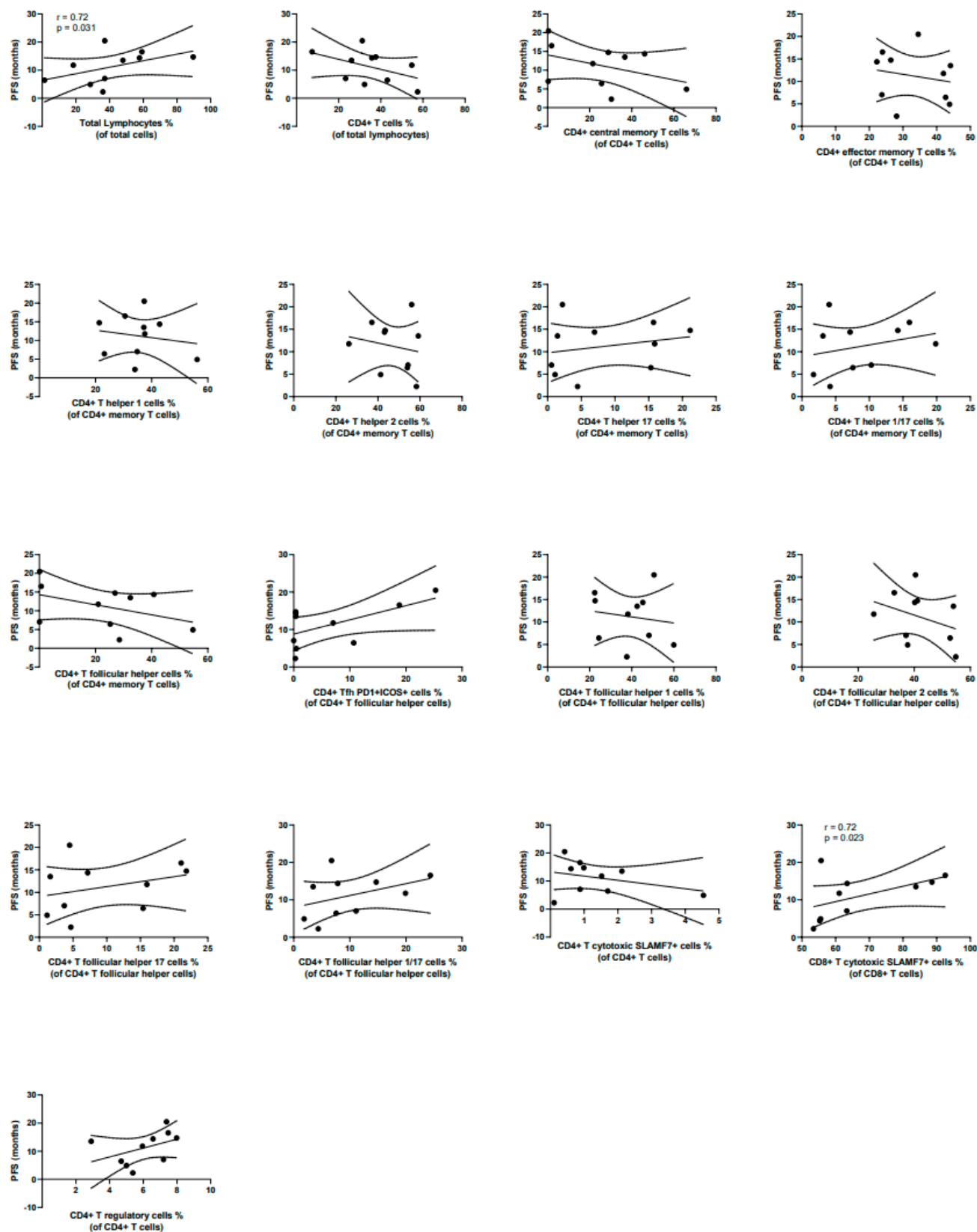
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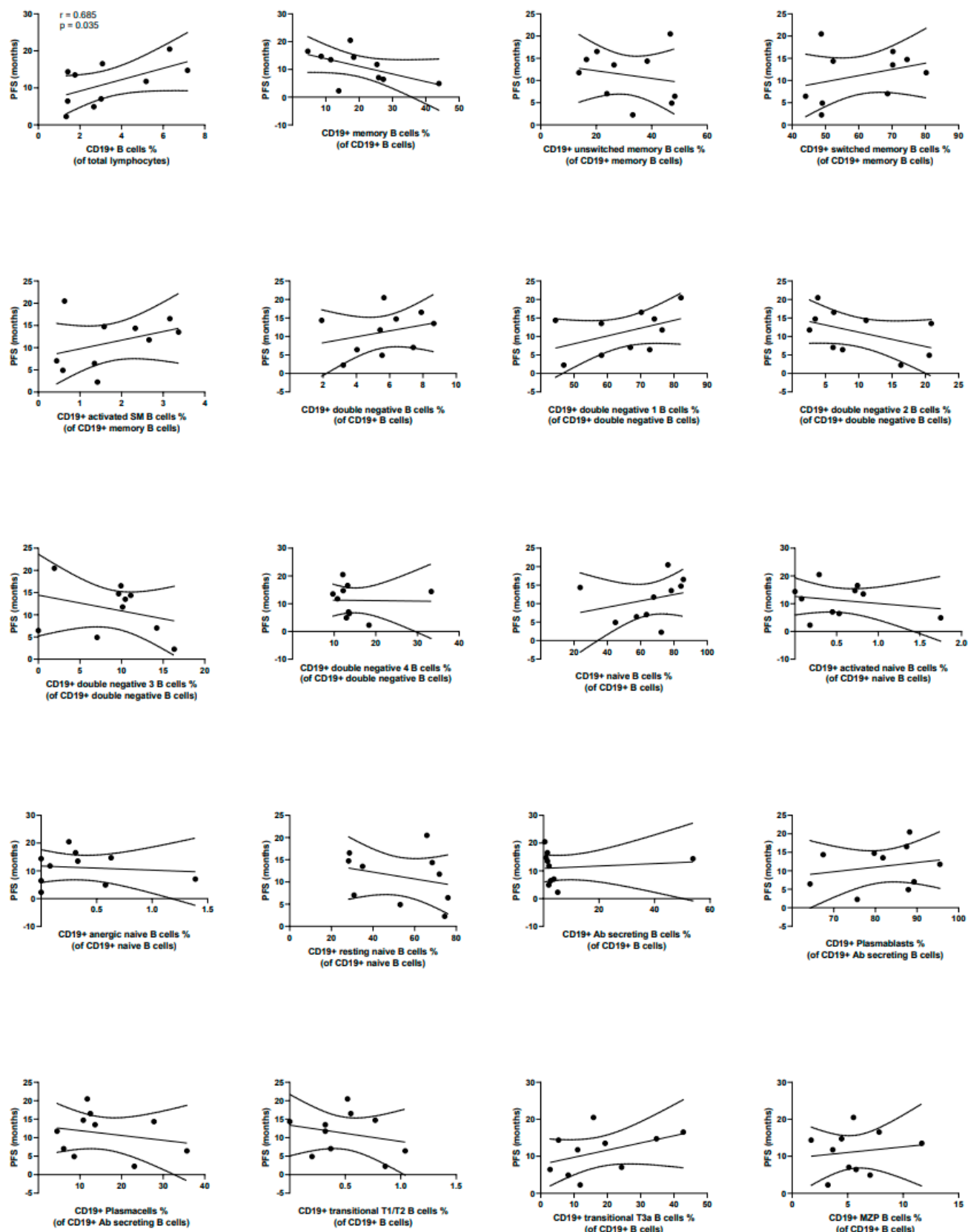


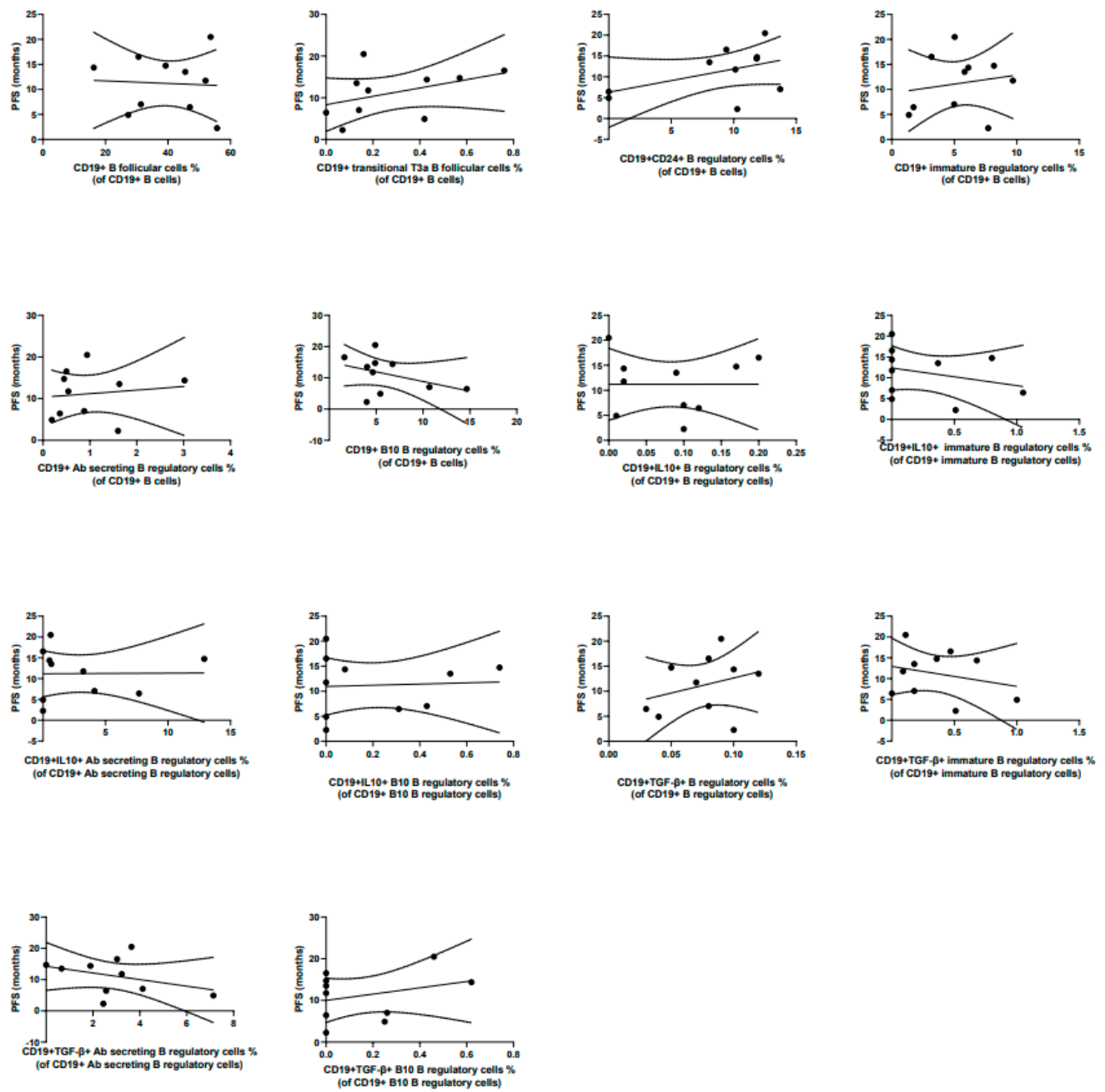


**Footnote:** Tfh: T follicular helper; SM: switched memory; Ab: antibody secreting; T1/2: type 1/2; T3a: type 3a; MZP: marginal zone peripheral; Breg: B regulatory.

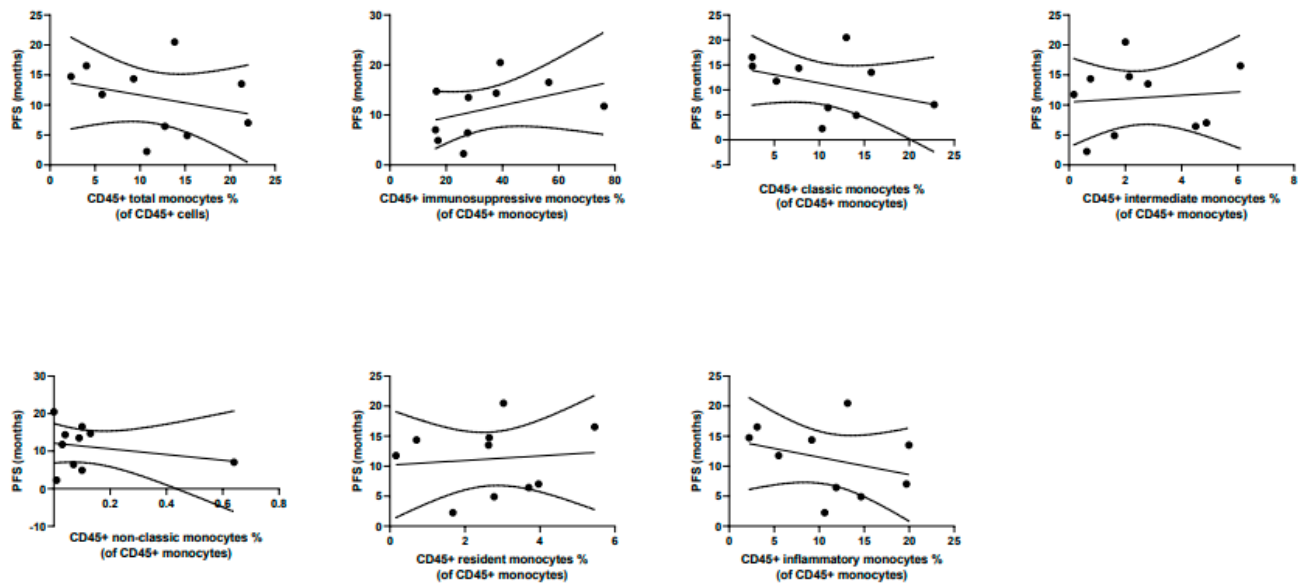
**Figure S5.** Correlation studies between circulating immune cells (A: T-cell compartment; B: B-cell compartment; C: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D: Pro Human Inflammation Panel I Assay; E: Pro TGF- $\beta$  Immunoassay; F: Pro Human Cytokine Immunoassay) at baseline and progression-free survival (PFS) after therapy onset in patients with pancreatic adenocarcinoma (PDAC). Correlation studies were performed using the Spearman's rho rank correlation test.

**A**

**B**



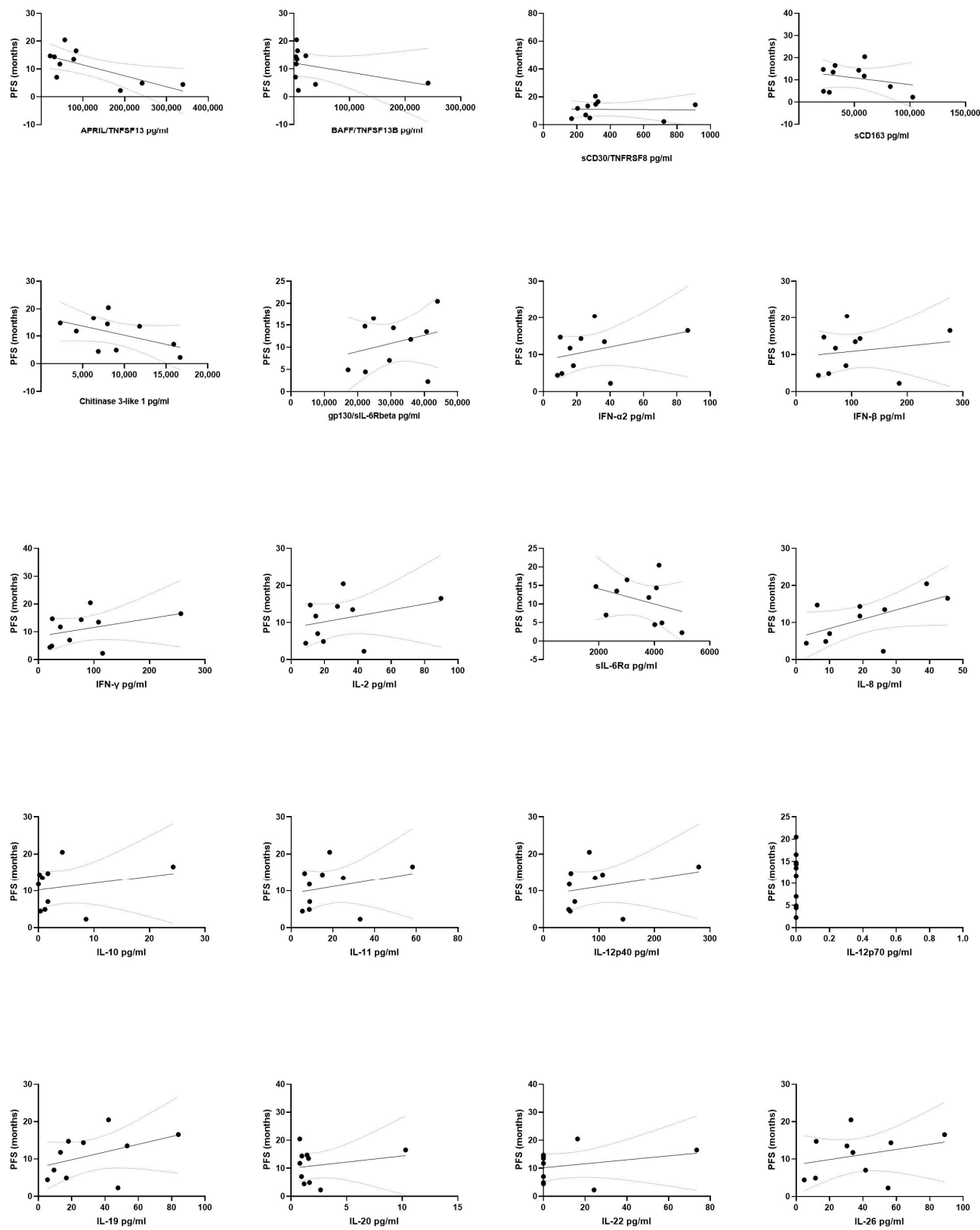
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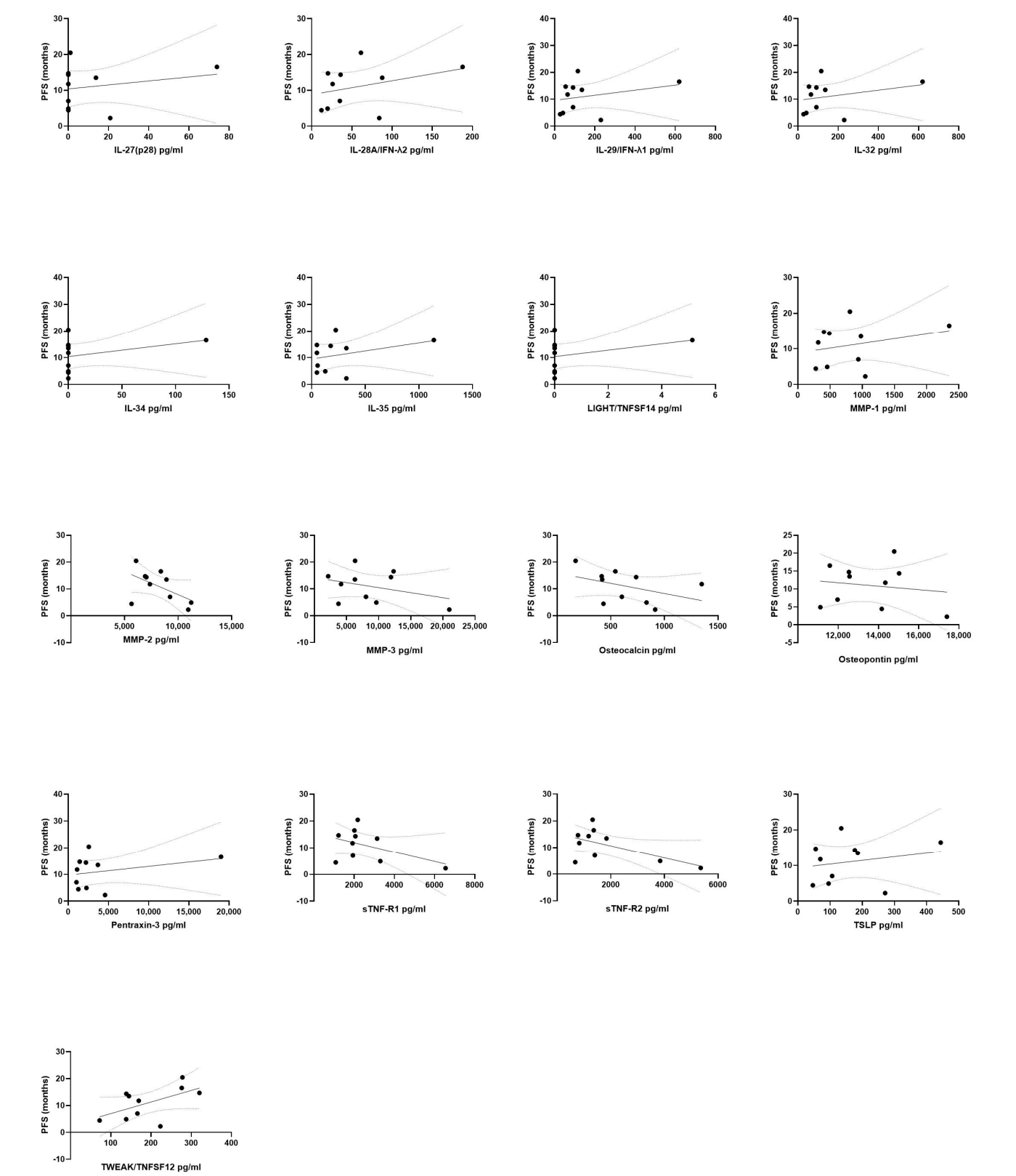


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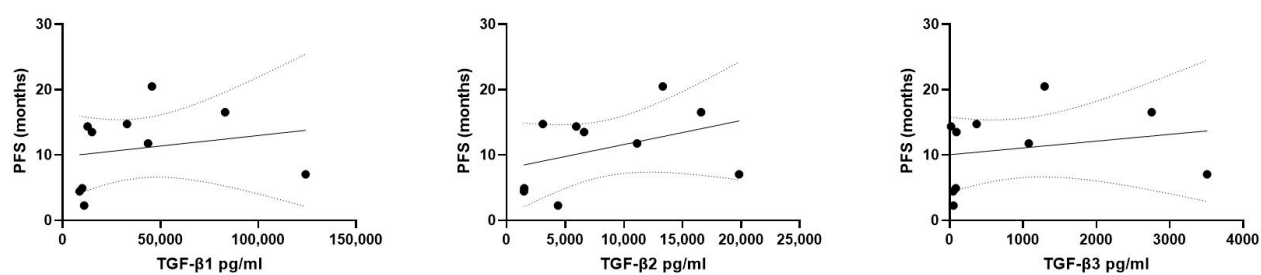
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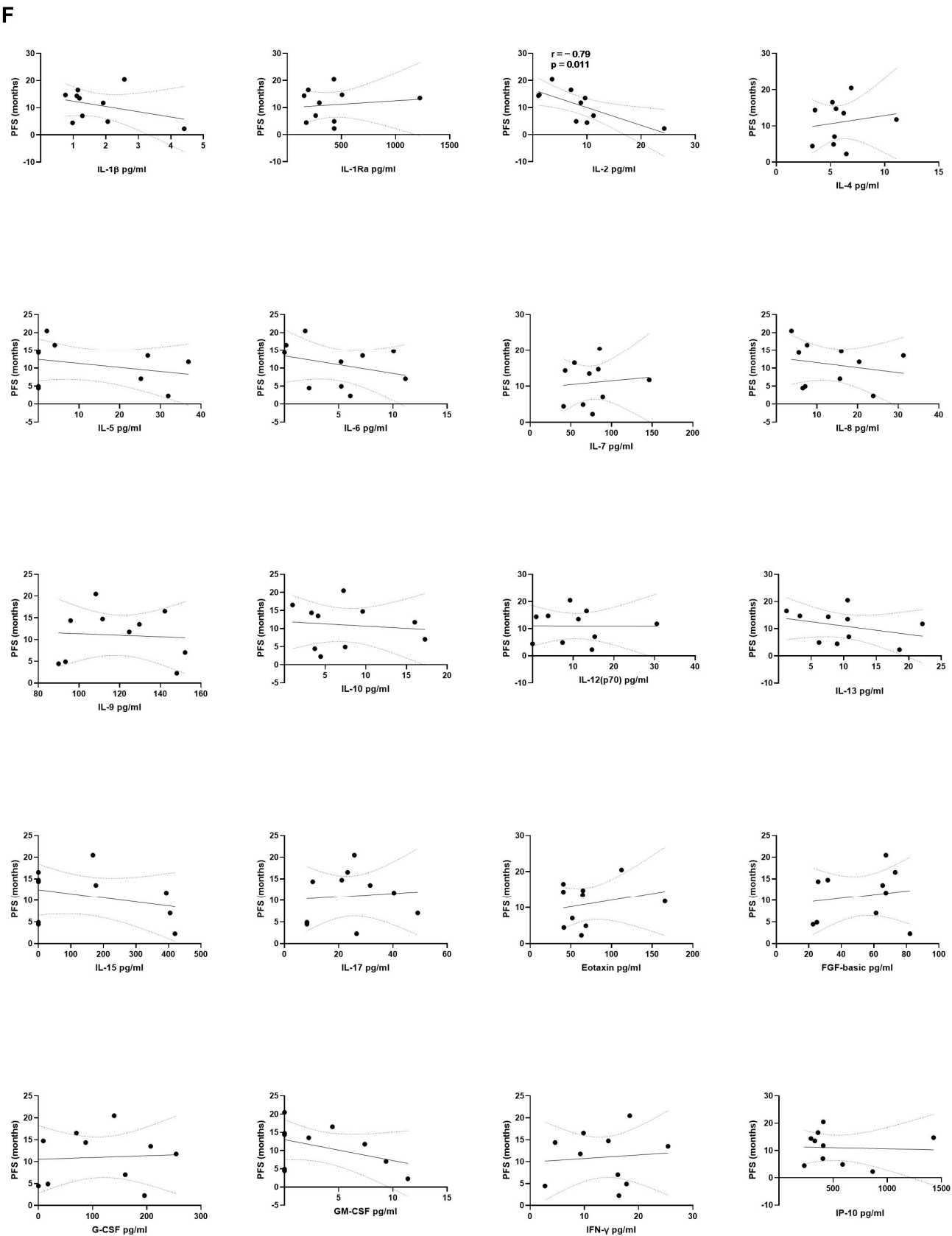
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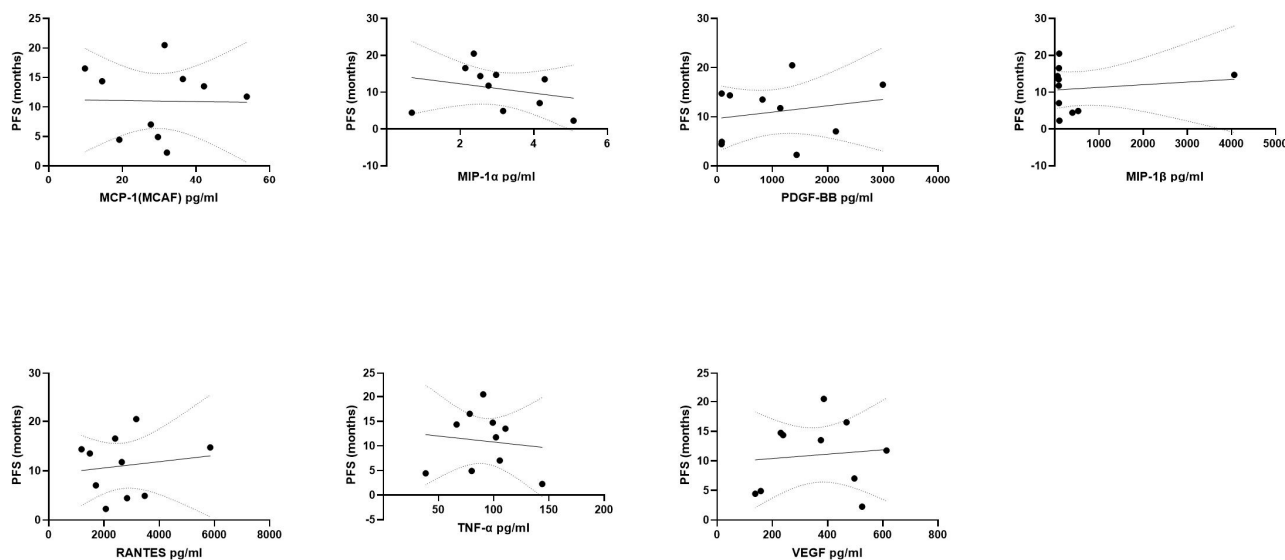






**E**

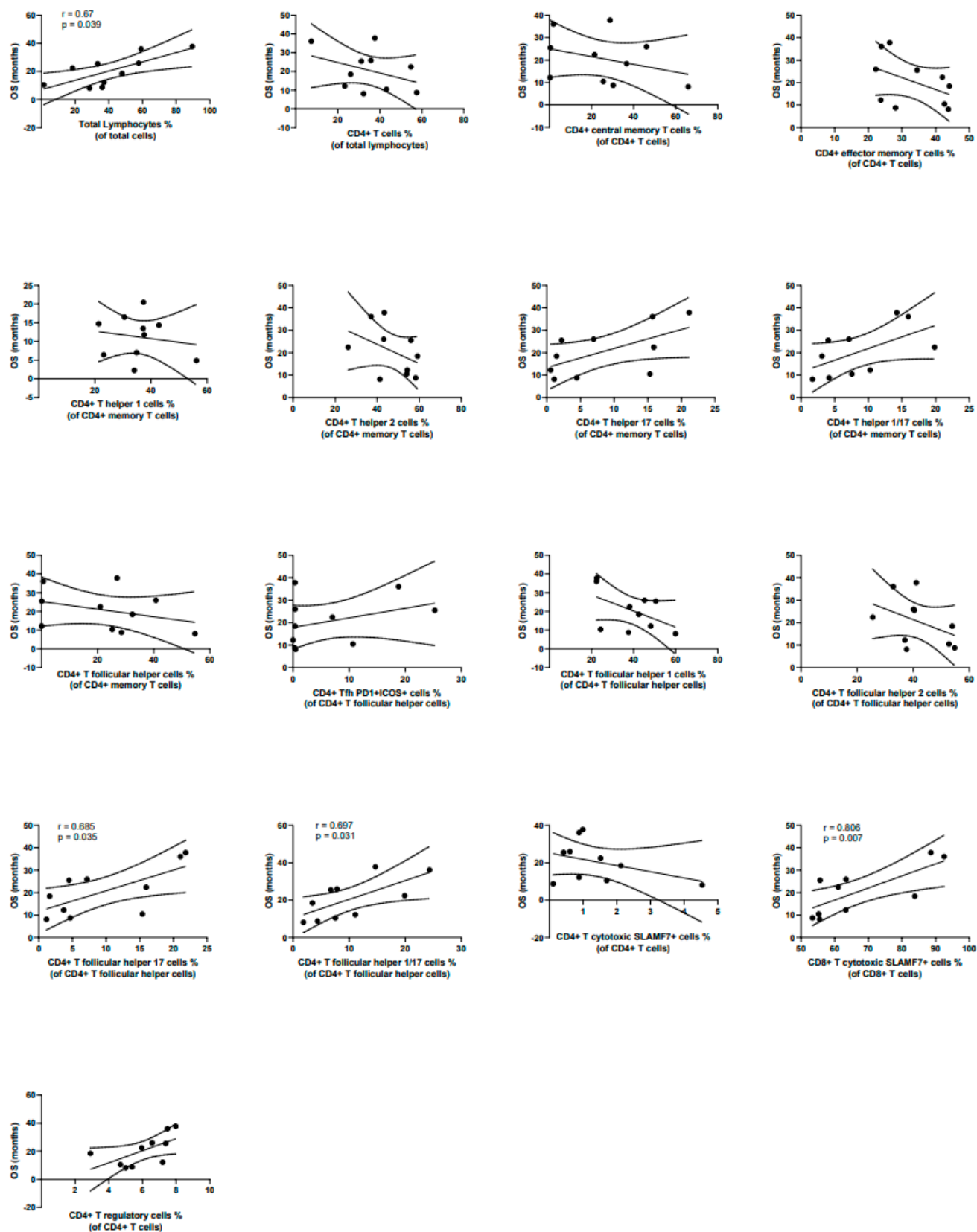


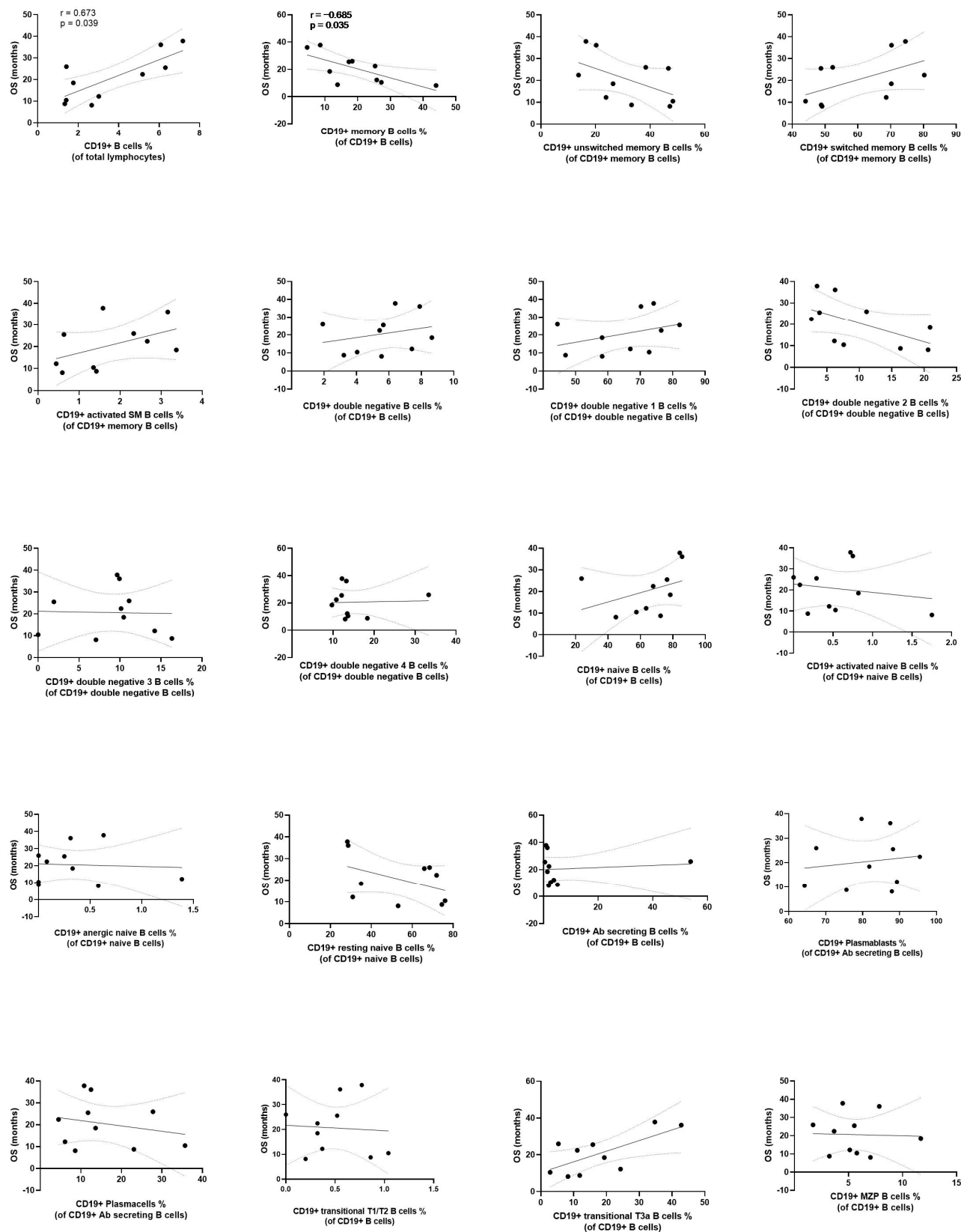


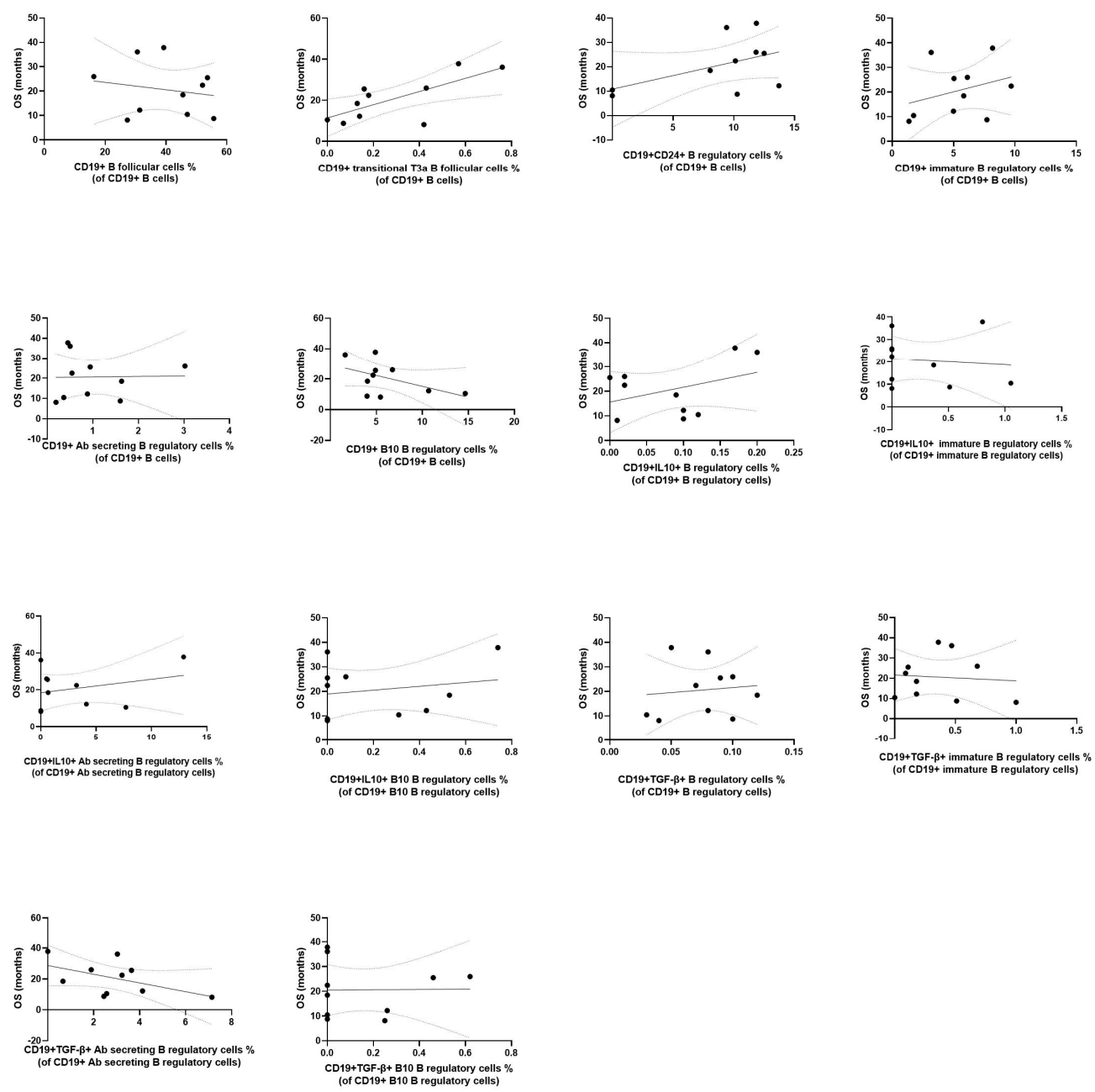
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**Figure S6.** Correlation studies between circulating immune cells (A: T-cell compartment; B: B-cell compartment; C: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D: Pro Human Inflammation Panel I Assay; E: Pro TGF- $\beta$  Immunoassay; F: Pro Human Cytokine Immunoassay) at baseline and overall survival (OS) after therapy onset in patients with pancreatic adenocarcinoma (PDAC). Correlation studies were performed using the Spearman's rho rank correlation test.

A



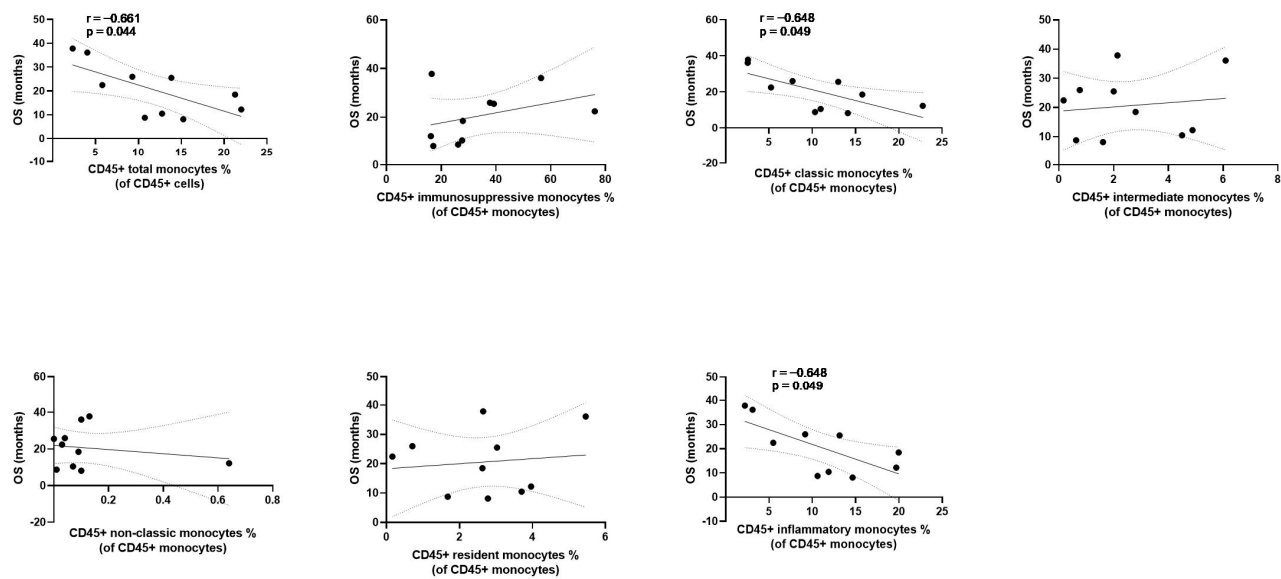
**B**



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C

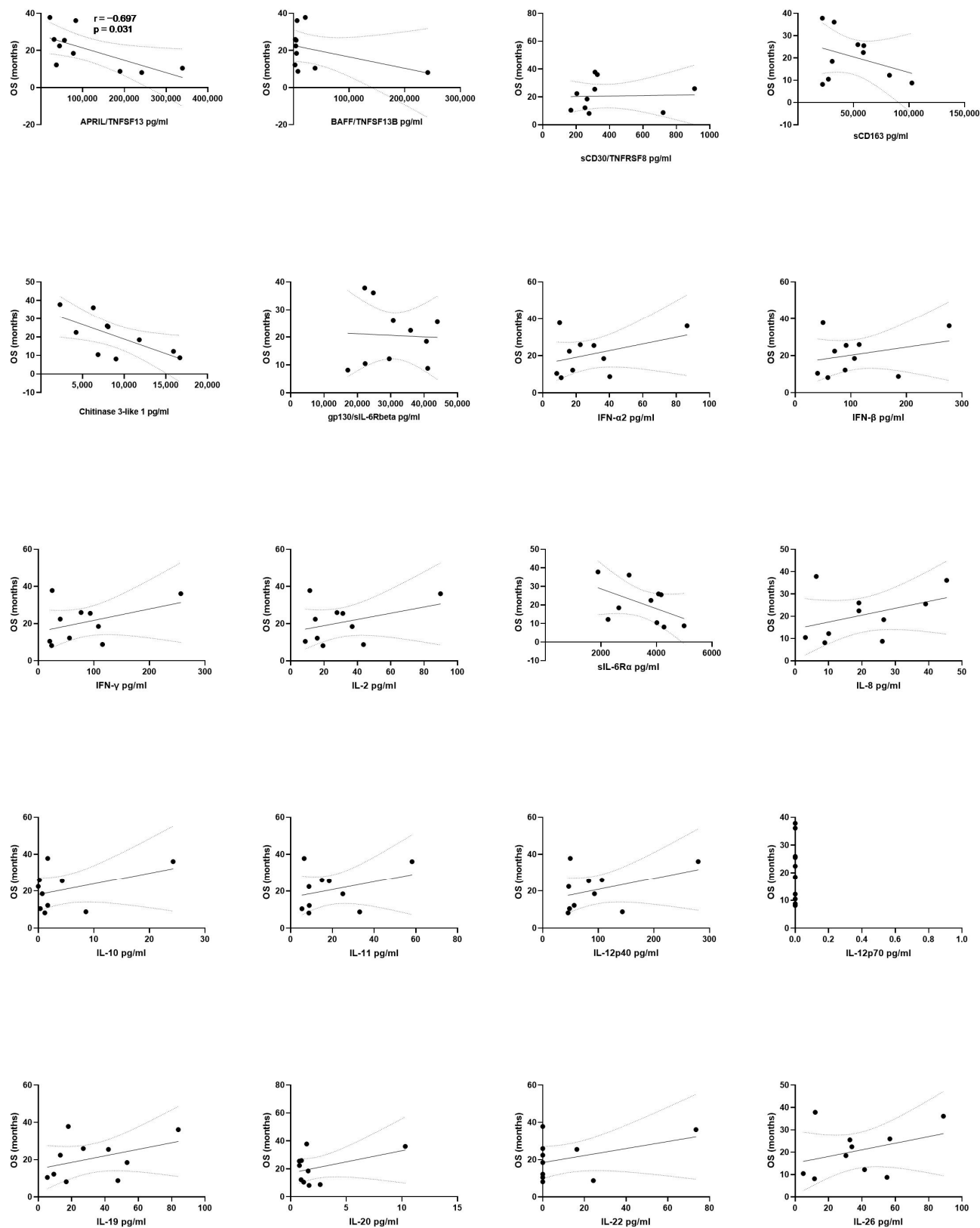


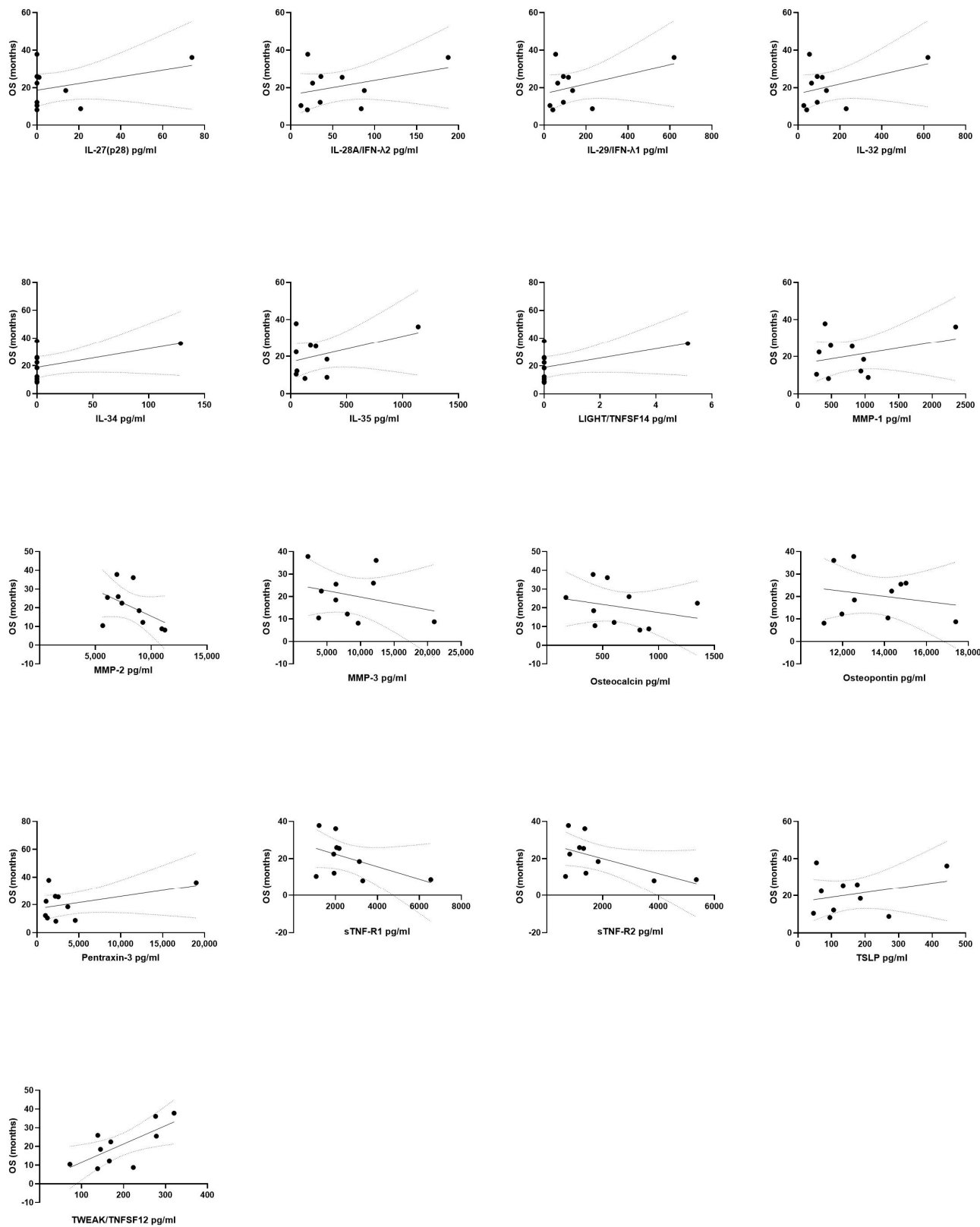
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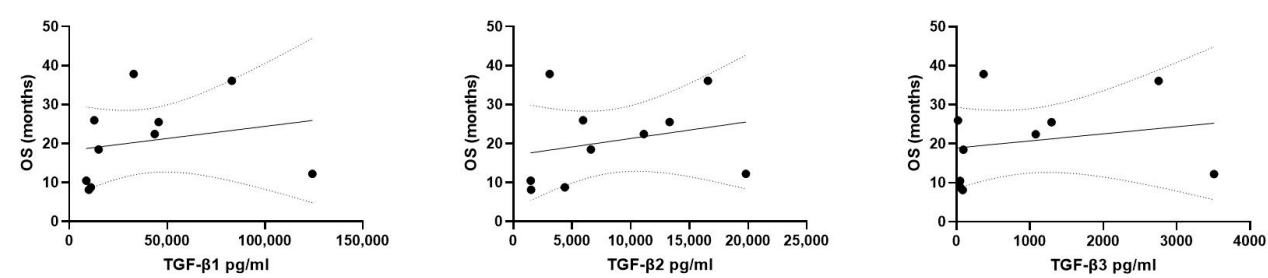


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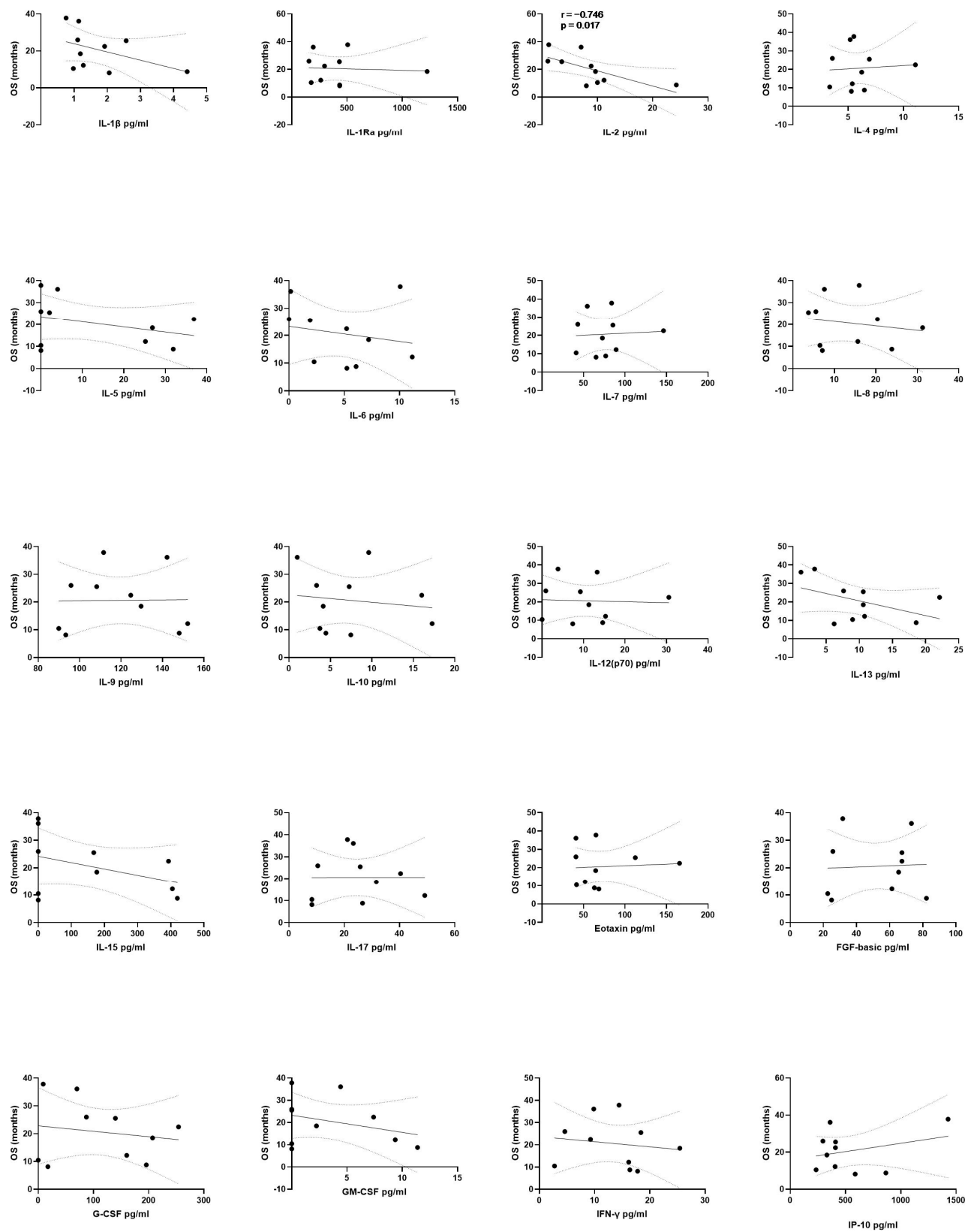




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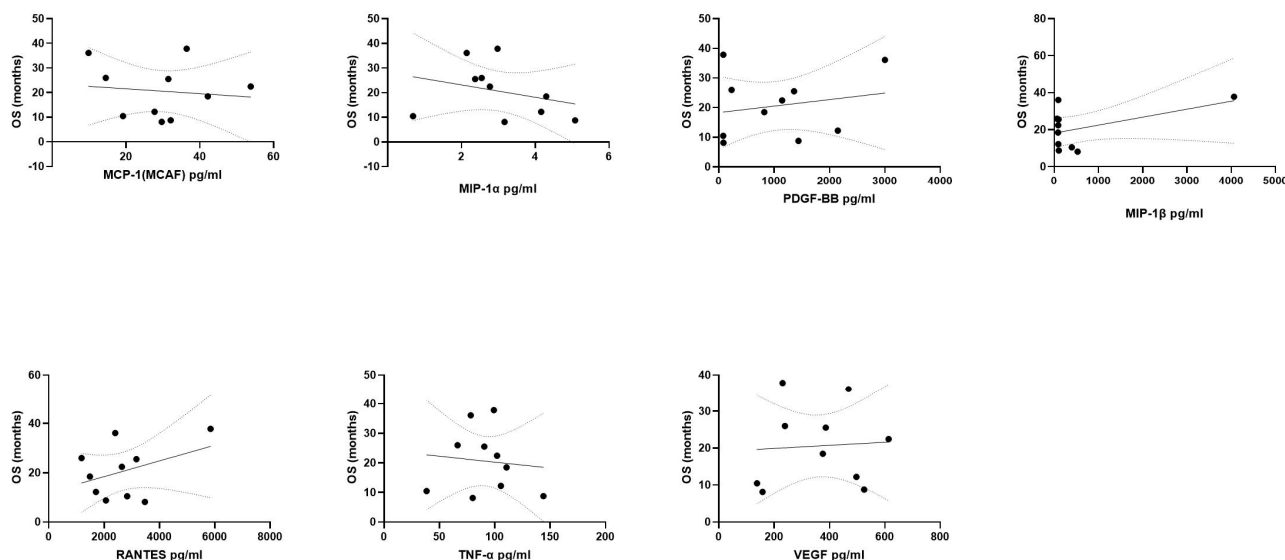


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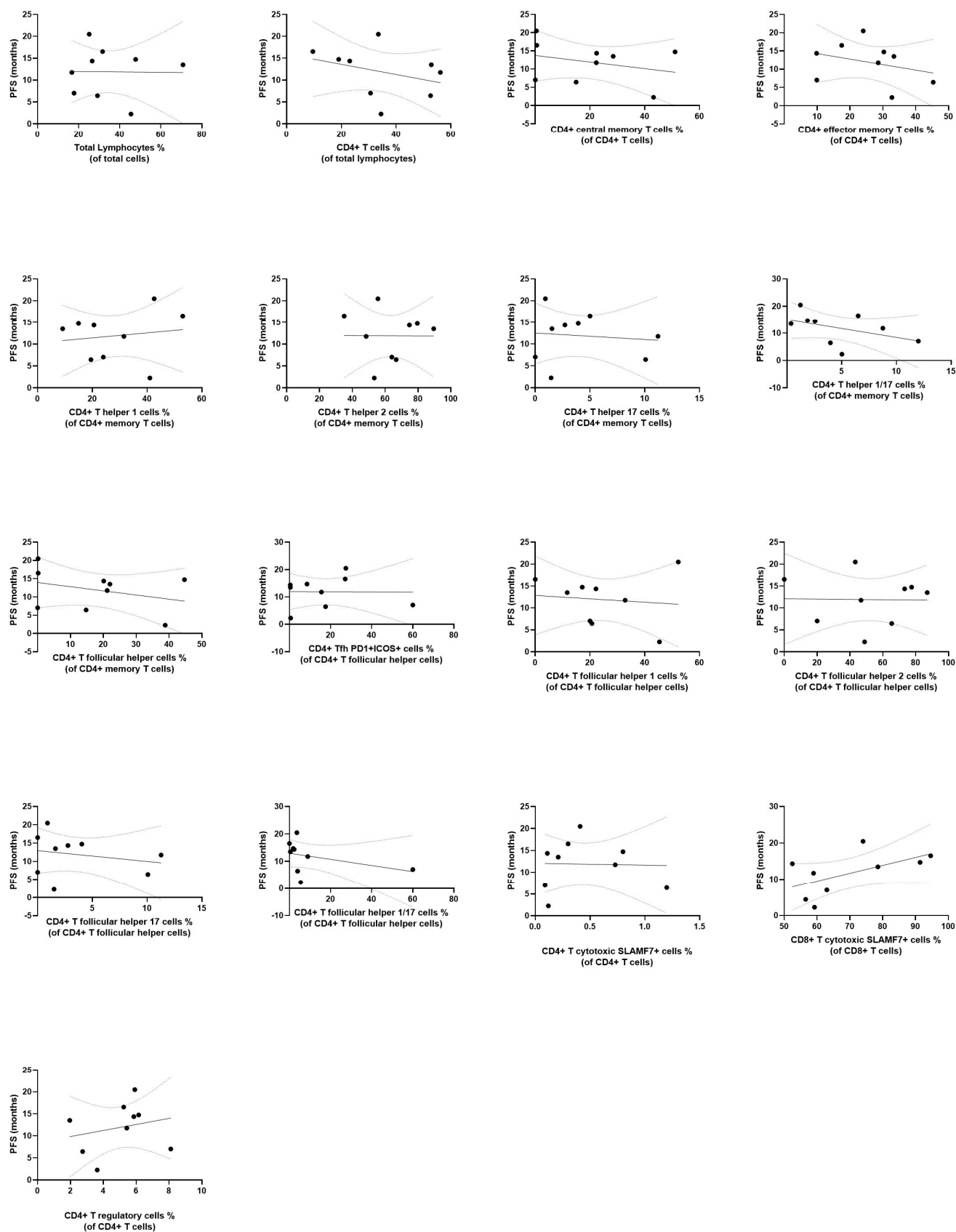
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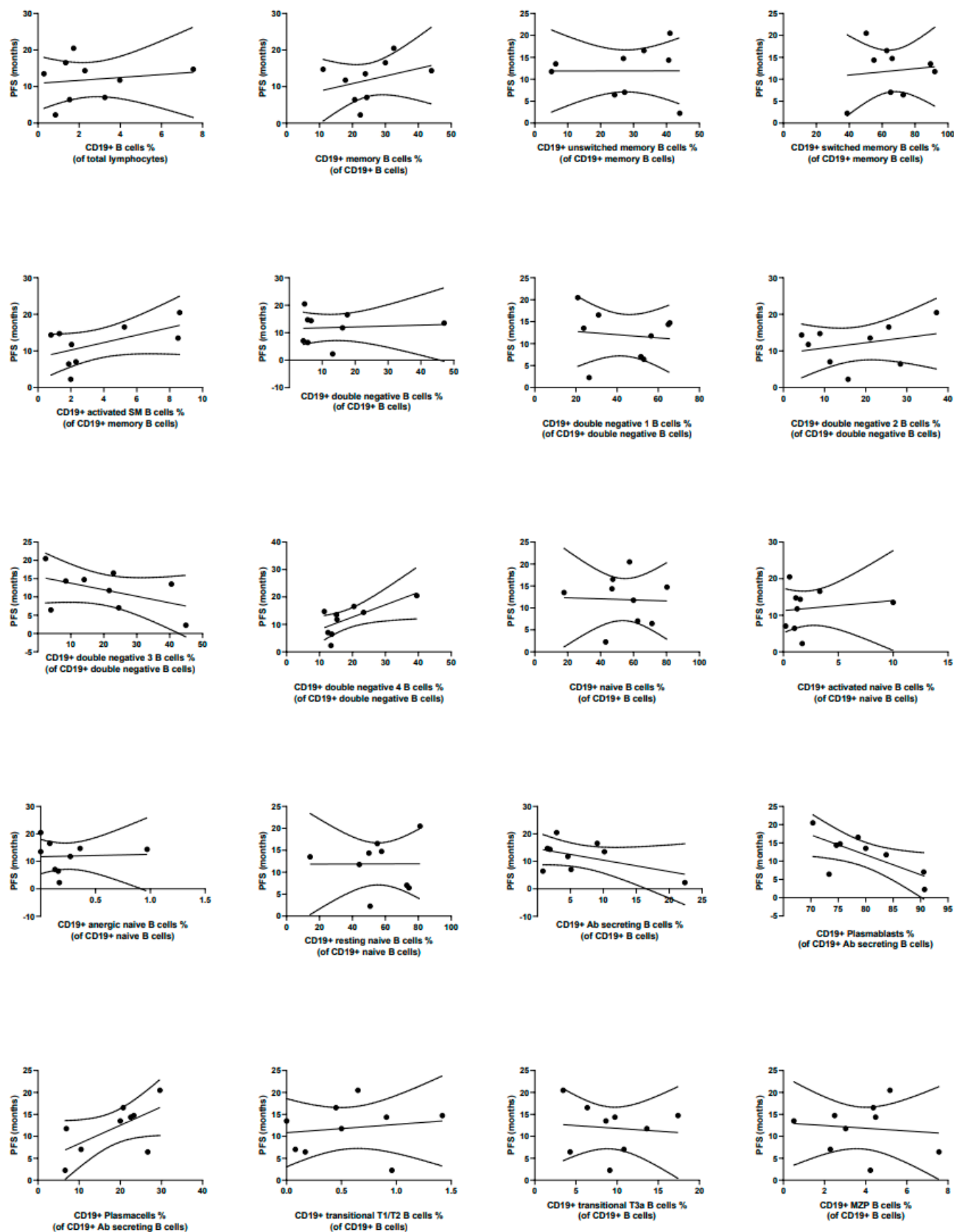
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**Footnote:** Te: T effector; Th1: T helper 1; Th2: T helper 2; Th17: T helper 17; Th1/17: T helper 1/17; Tfh: T follicular helper; Tfh1: T follicular helper 1; Tfh2: T follicular helper 2; Tfh17: T follicular helper 17; Tfh1/17: T follicular helper 1/17; Treg: T regulatory; UM: unswitched memory; SM: switched memory; ASM: activated switched memory; DN: double negative; DN1: double negative 1; DN2: double negative 2; DN3: double negative 3; DN4: double negative 4; AcN: activated naïve; AnN: anergic naïve; RN: resting naïve; Ab: antibody secreting; T1/2: transitional 1/2; T3a: transitional 3a; MZP: marginal zone peripheral; Breg: B regulatory; AbS: antibody secreting; Imm: immature; Mon: monocytes.

**Figure S7.** Correlation studies between circulating immune cells (A: T-cell compartment; B: B-cell compartment; C: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D: Pro Human Inflammation Panel I Assay; E: Pro TGF- $\beta$  Immunoassay; F: Pro Human Cytokine Immunoassay) at 4-month follow-up and progression-free survival (PFS) after therapy onset in patients with pancreatic adenocarcinoma (PDAC). Correlation studies were performed using the Spearman's rho rank correlation test.

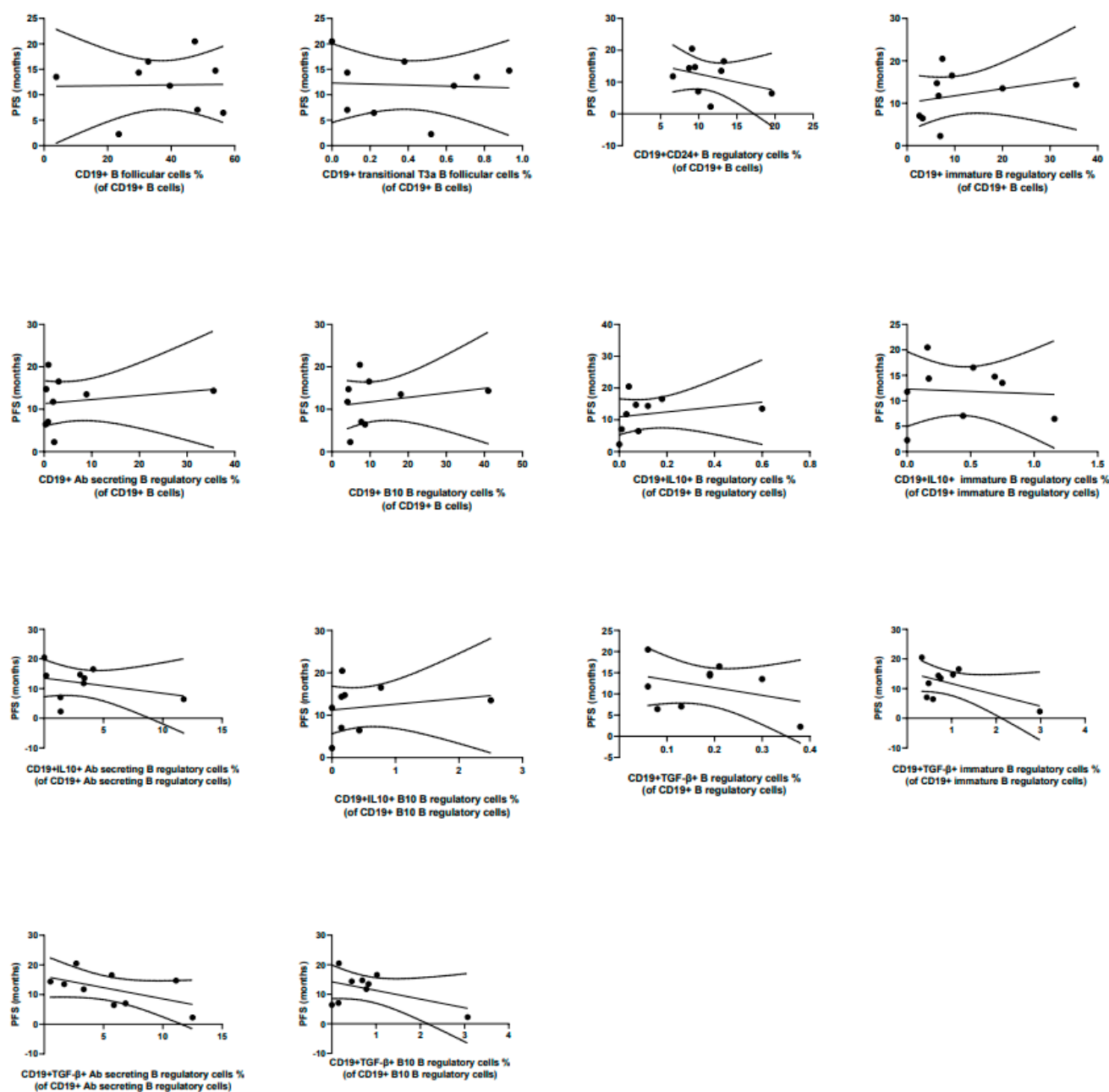
**A**

**B**

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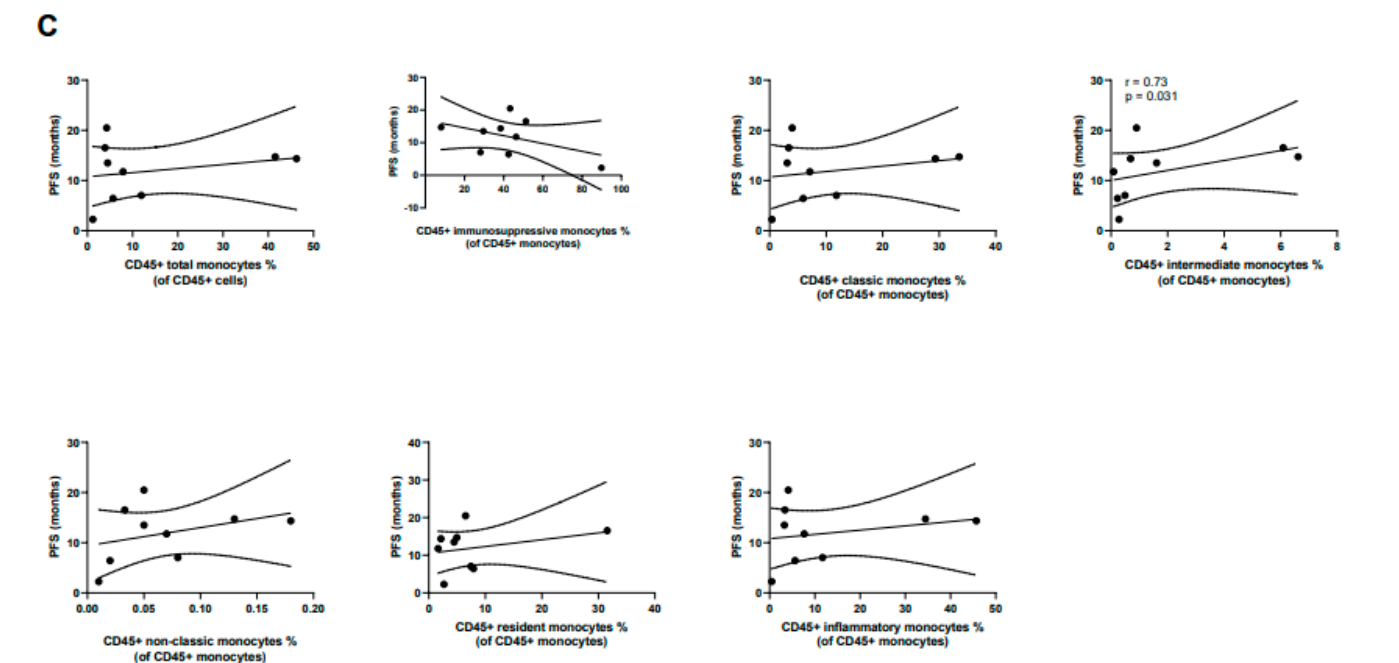
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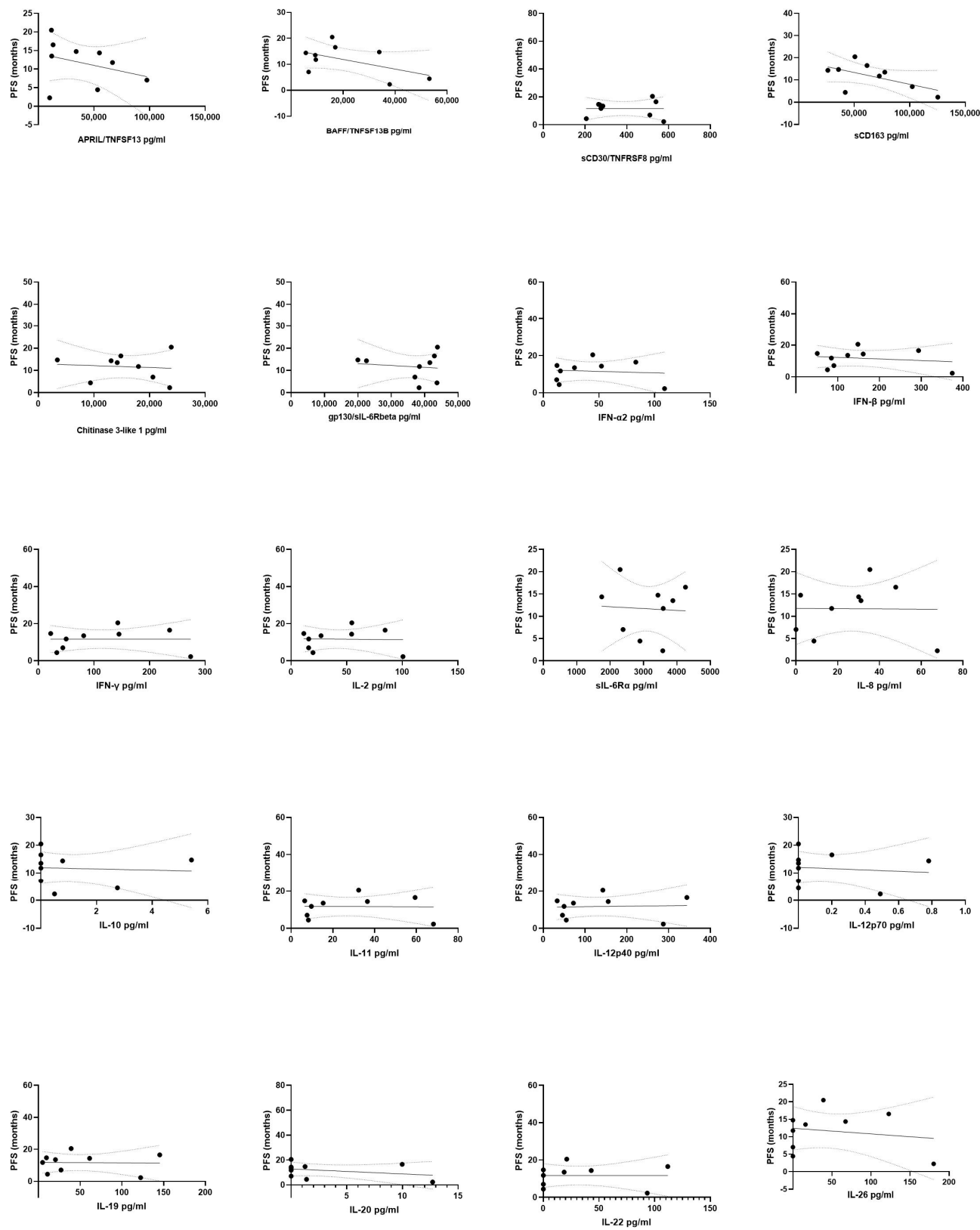
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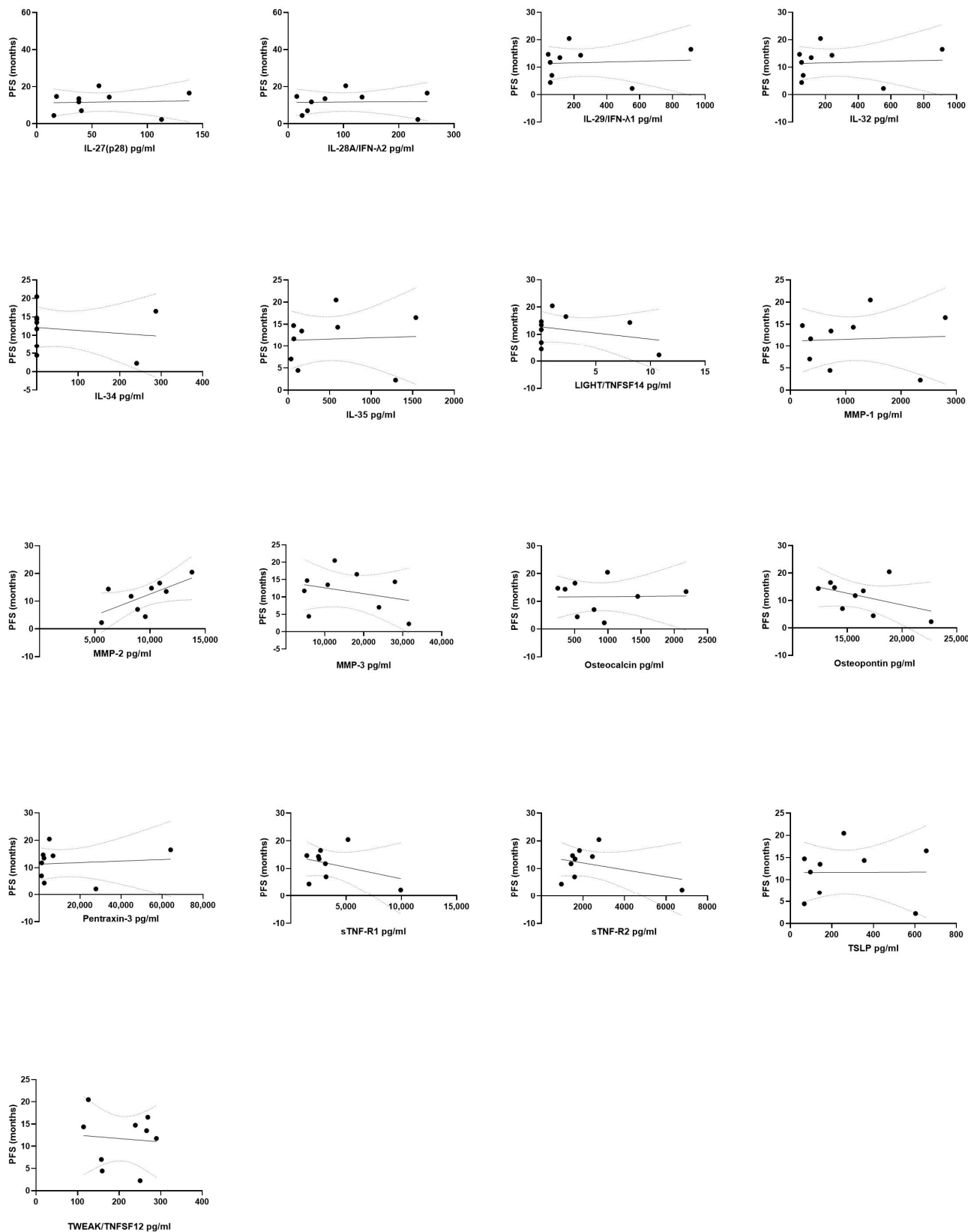


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D

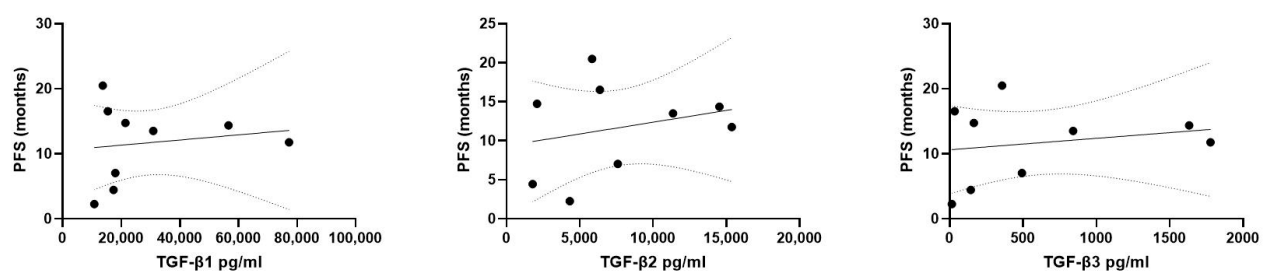


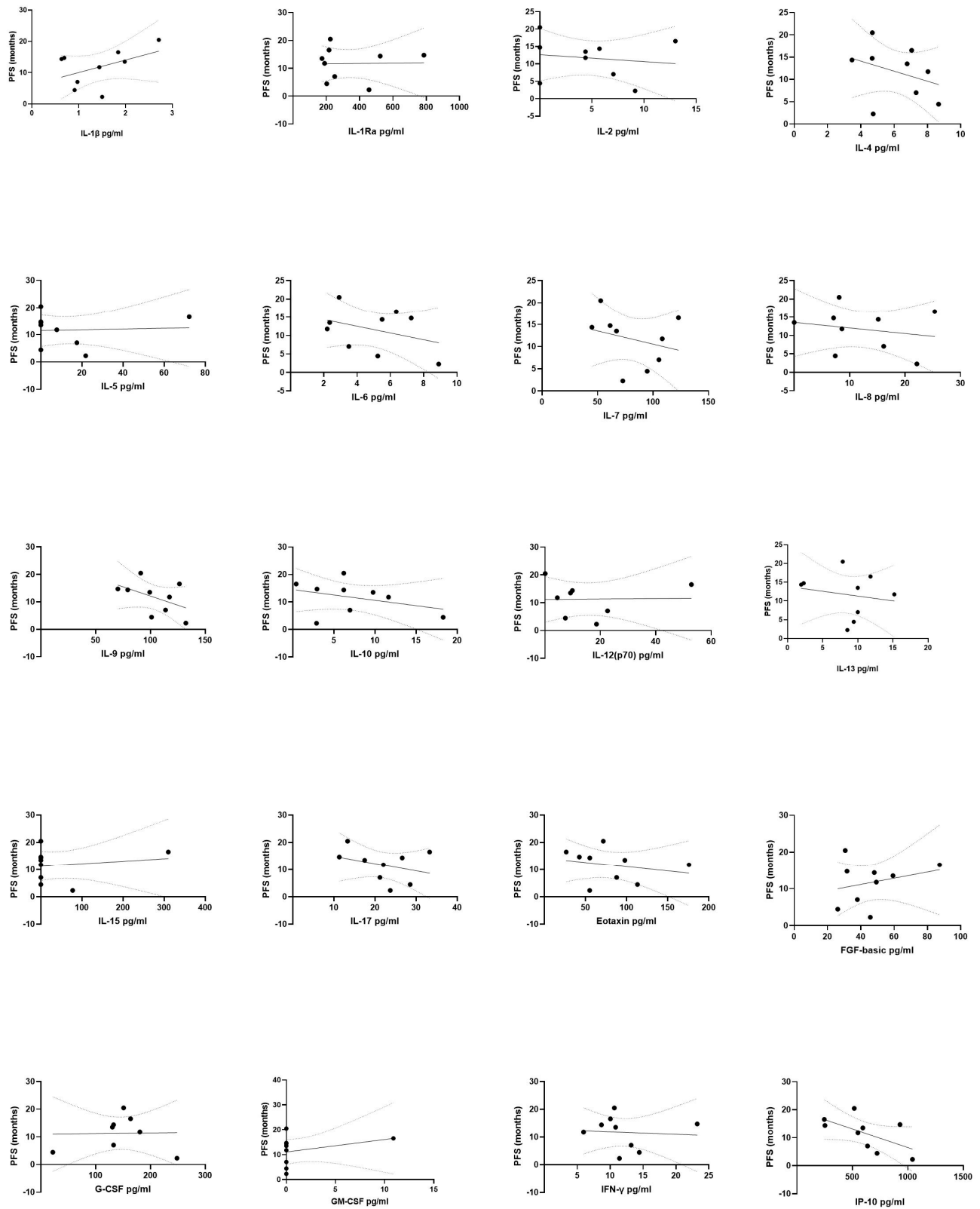


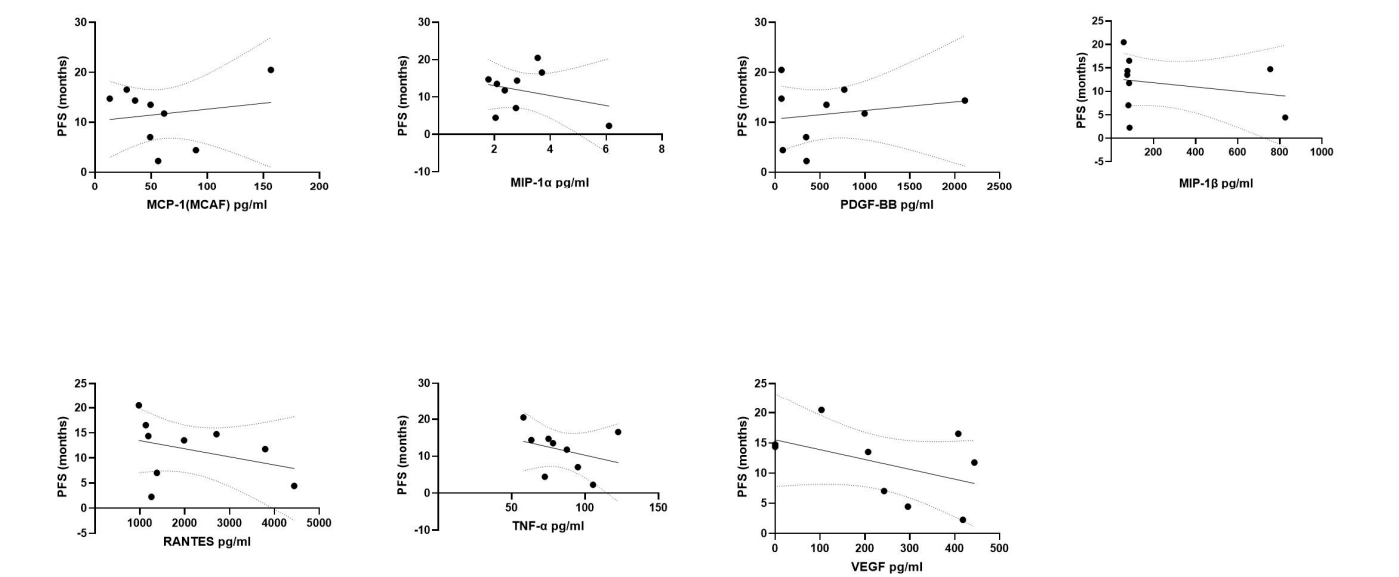
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**E**

**F**

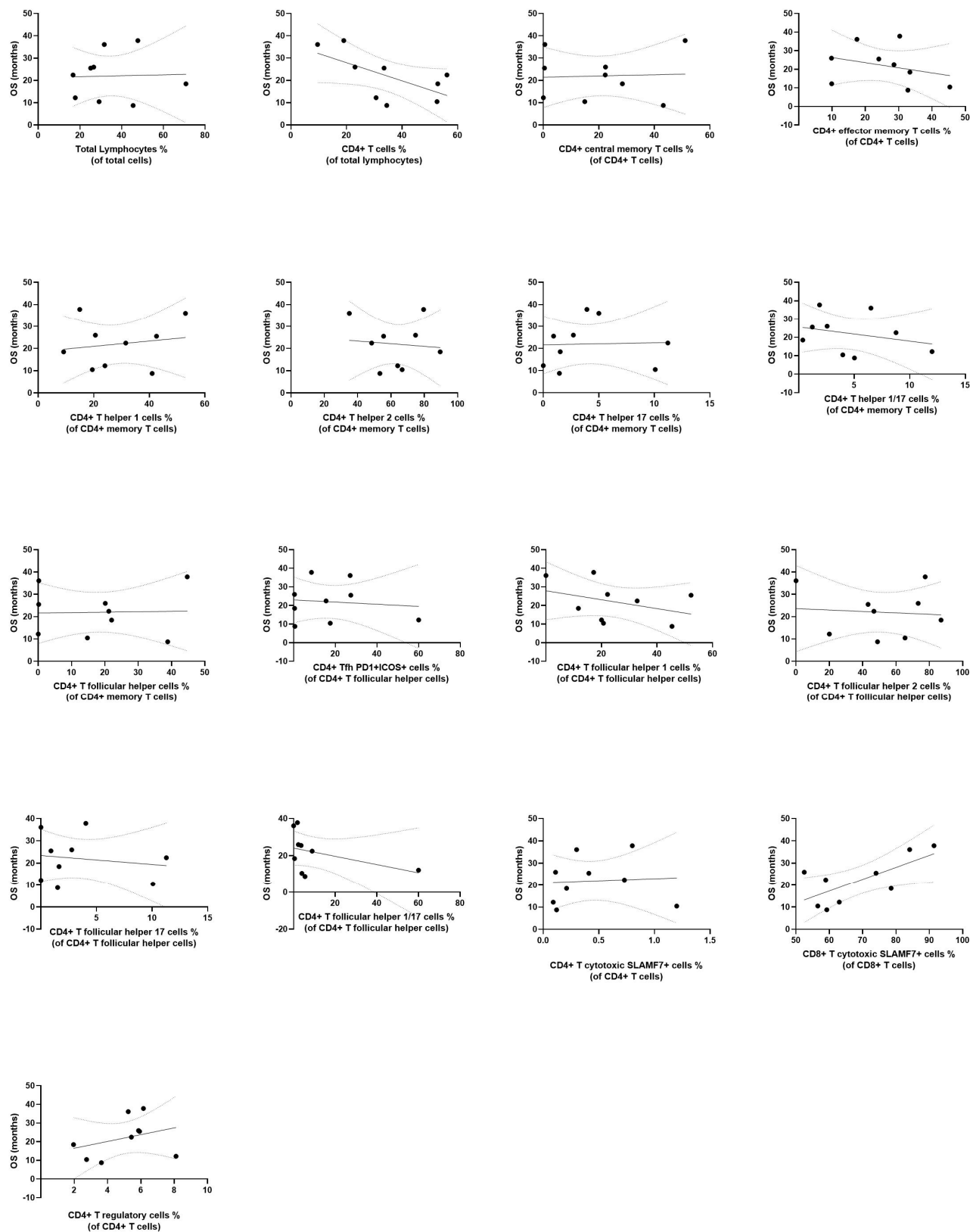


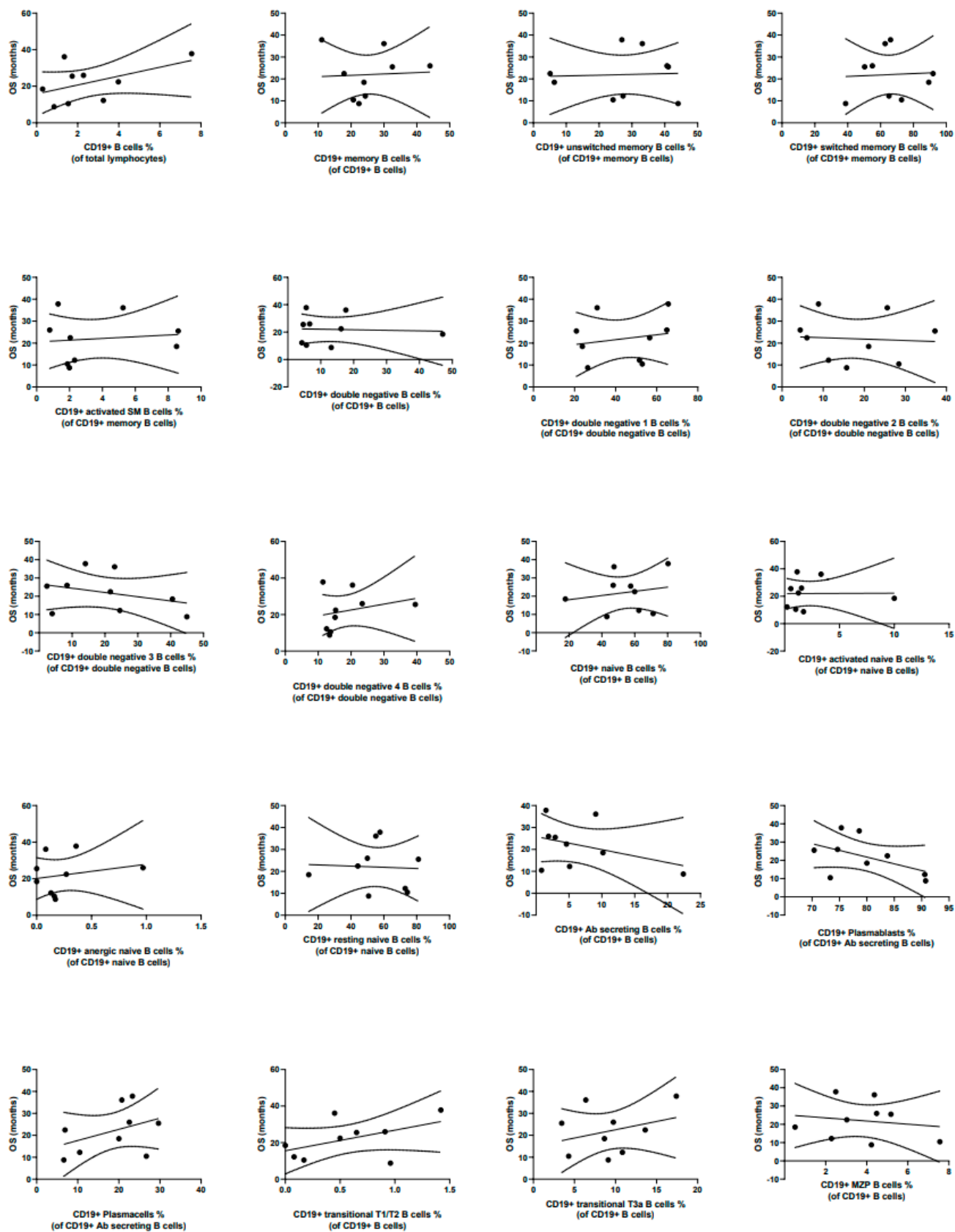
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**Figure S8.** Correlation studies between circulating immune cells (A: T-cell compartment; B: B-cell compartment; C: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D: Pro Human Inflammation Panel I Assay; E: Pro TGF- $\beta$  Immunoassay; F: Pro Human Cytokine Immunoassay) at 4-month follow-up and overall survival (OS) after therapy onset in patients with pancreatic adenocarcinoma (PDAC). Correlation studies were performed using the Spearman's rho rank correlation test.

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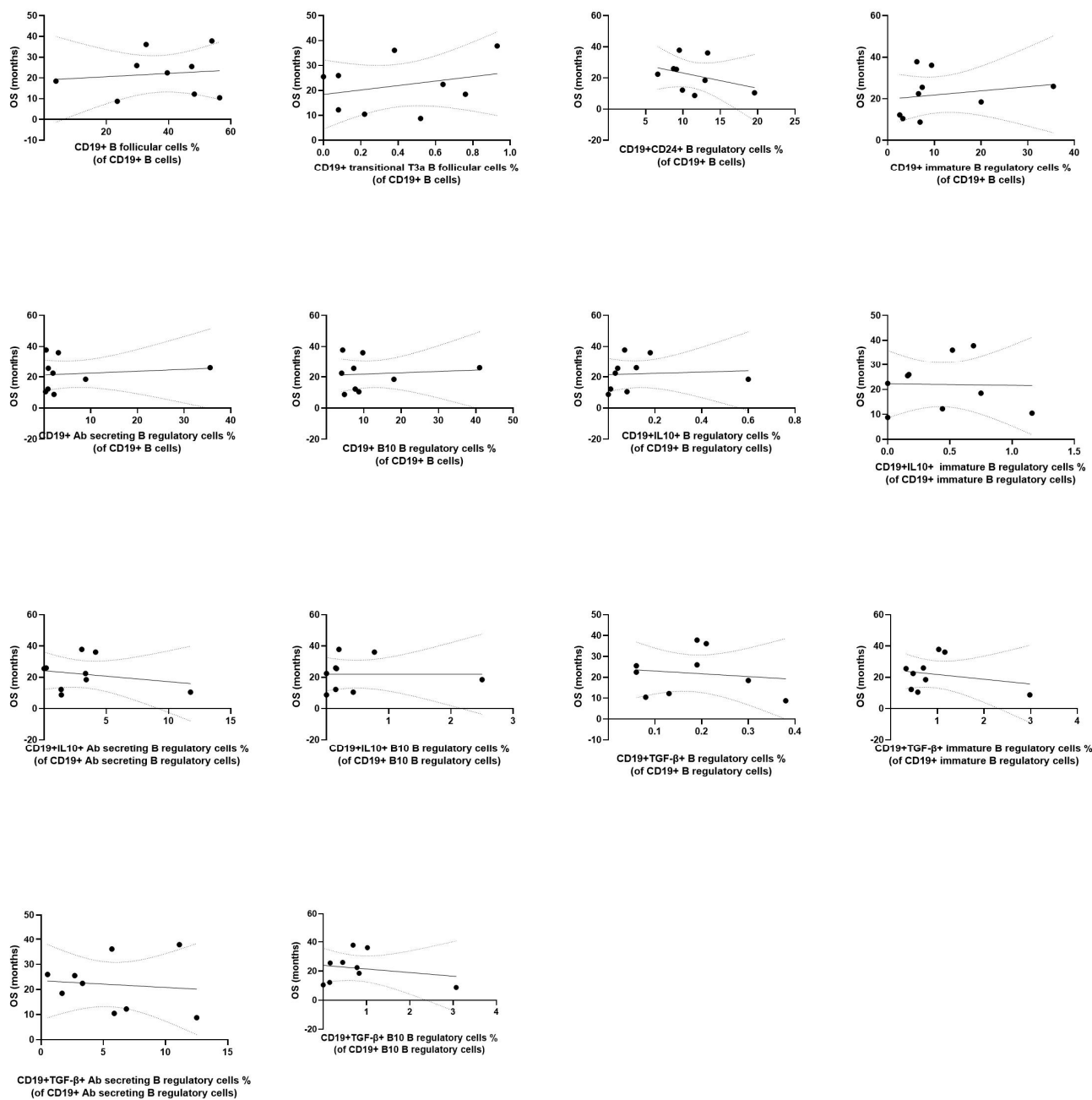


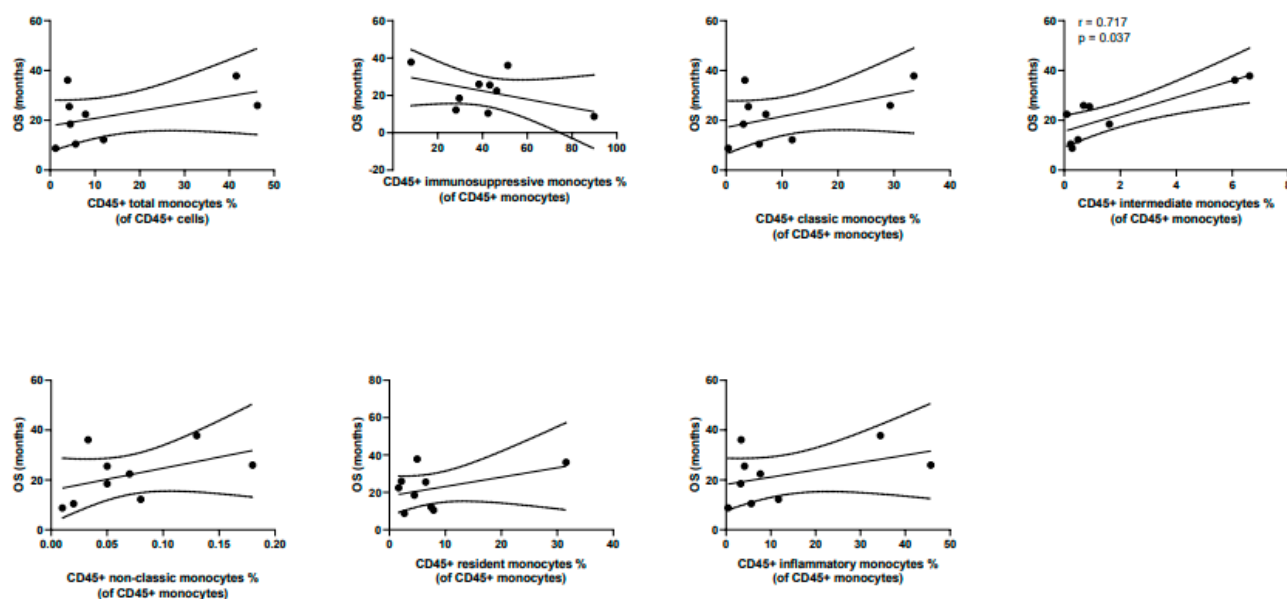
**A**

**B**

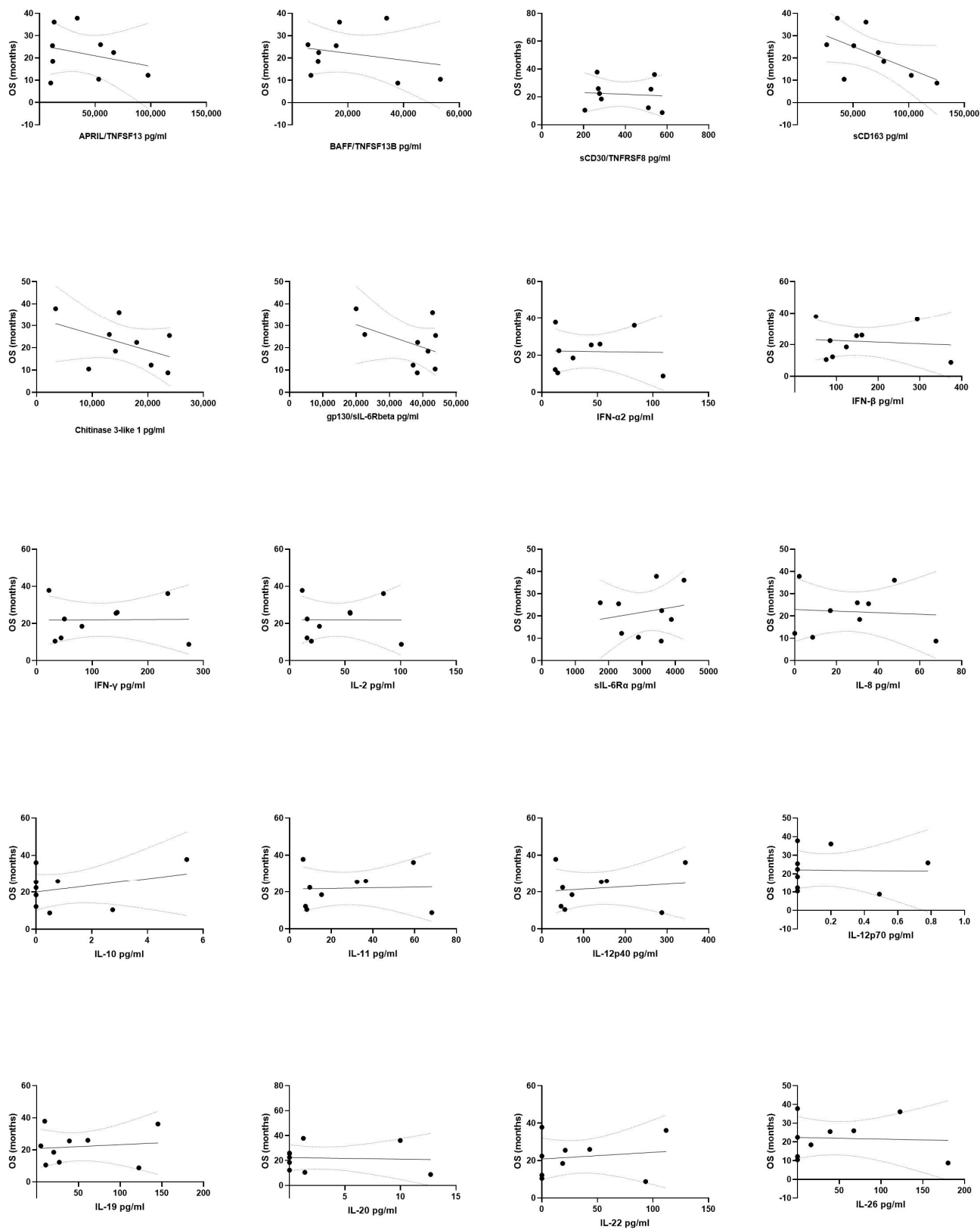
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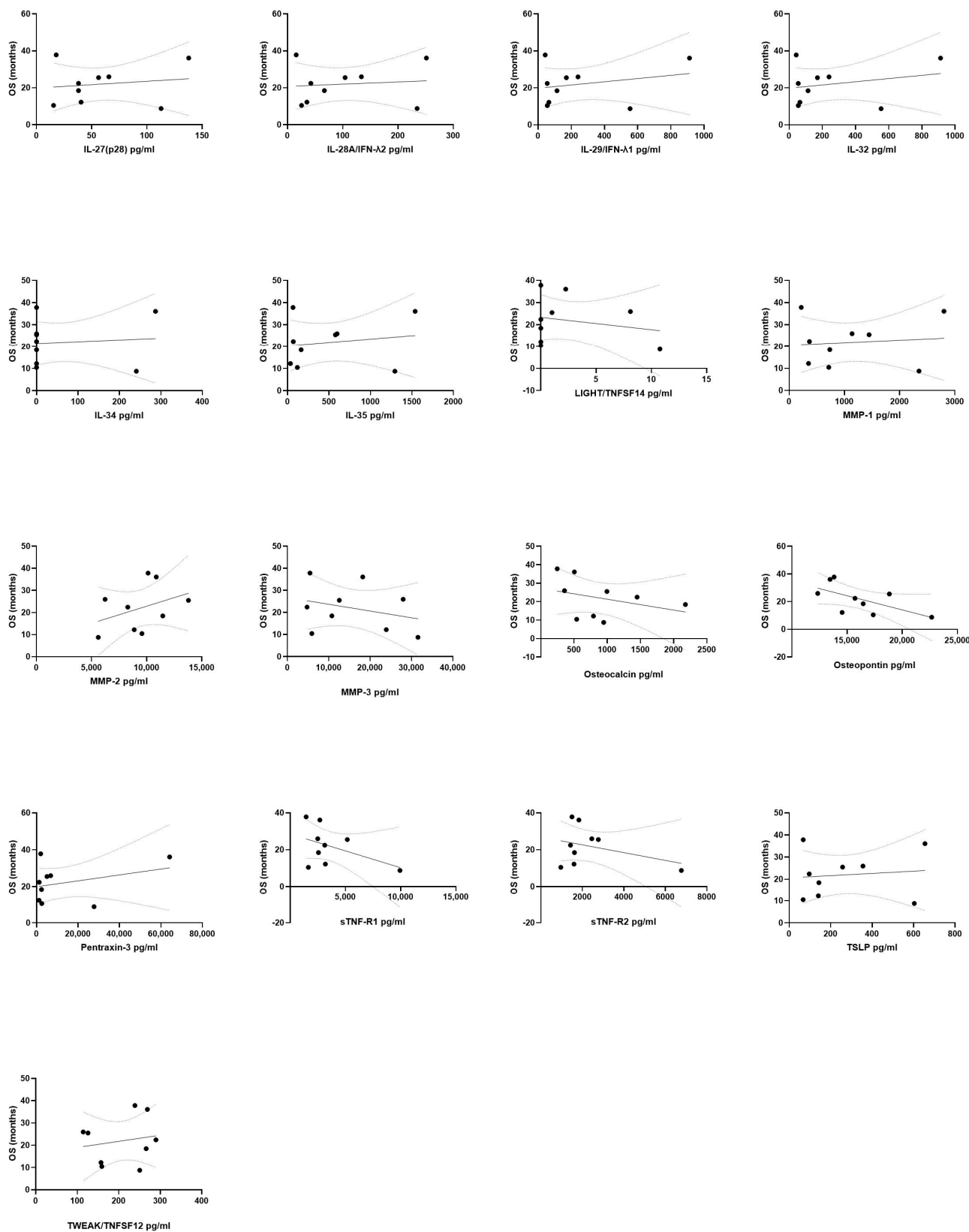
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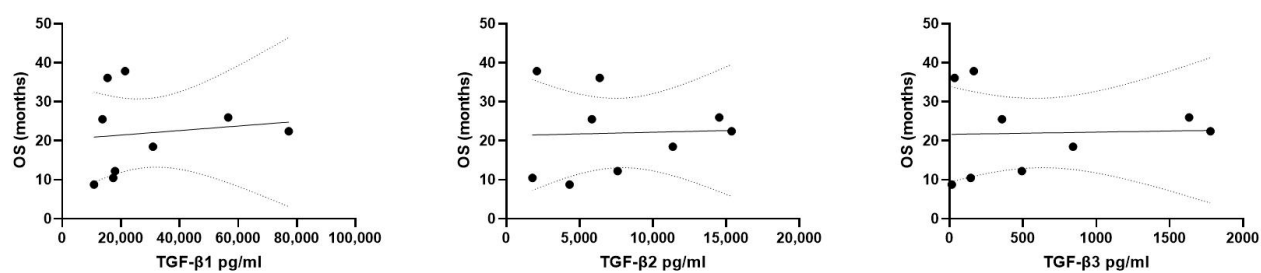


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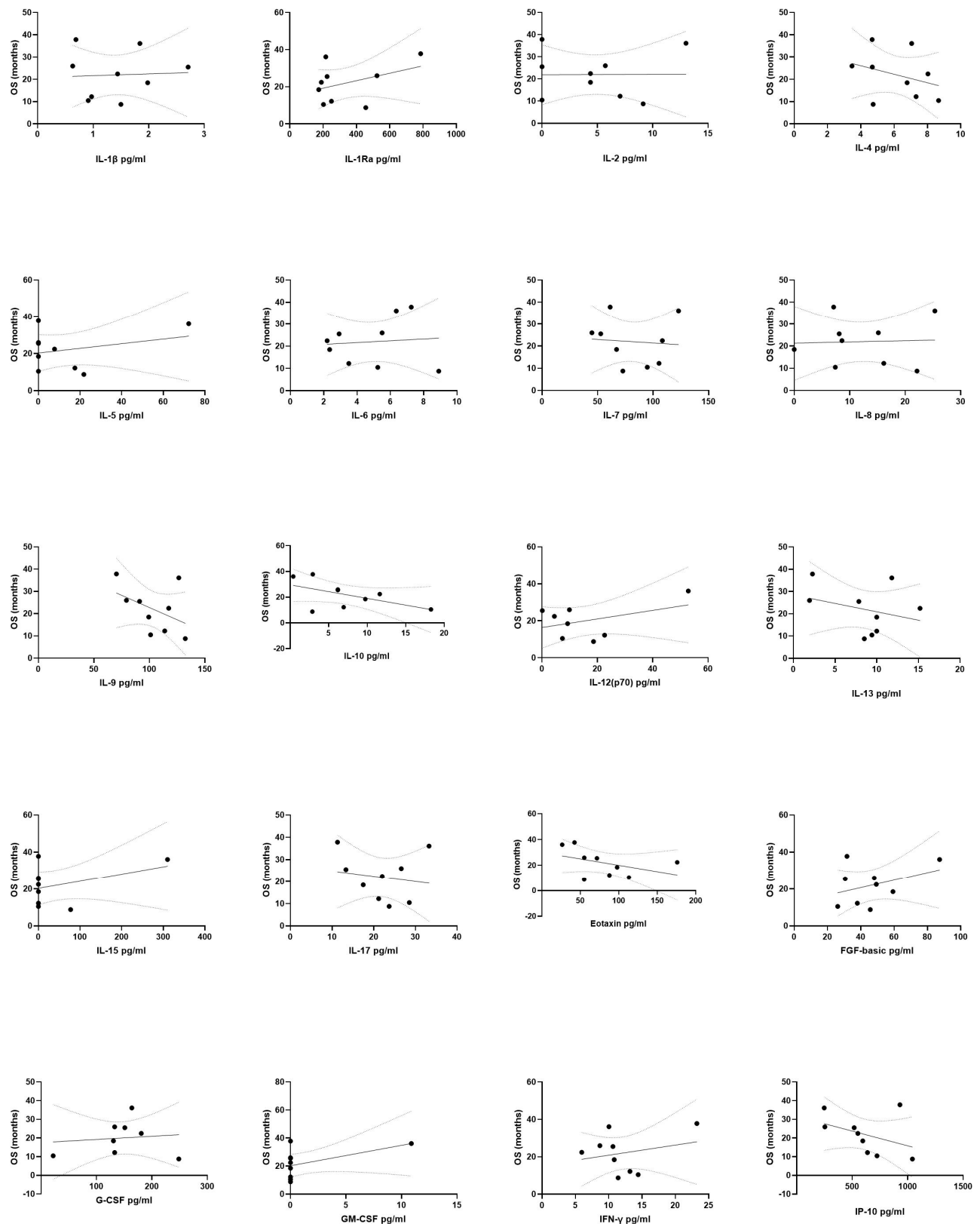
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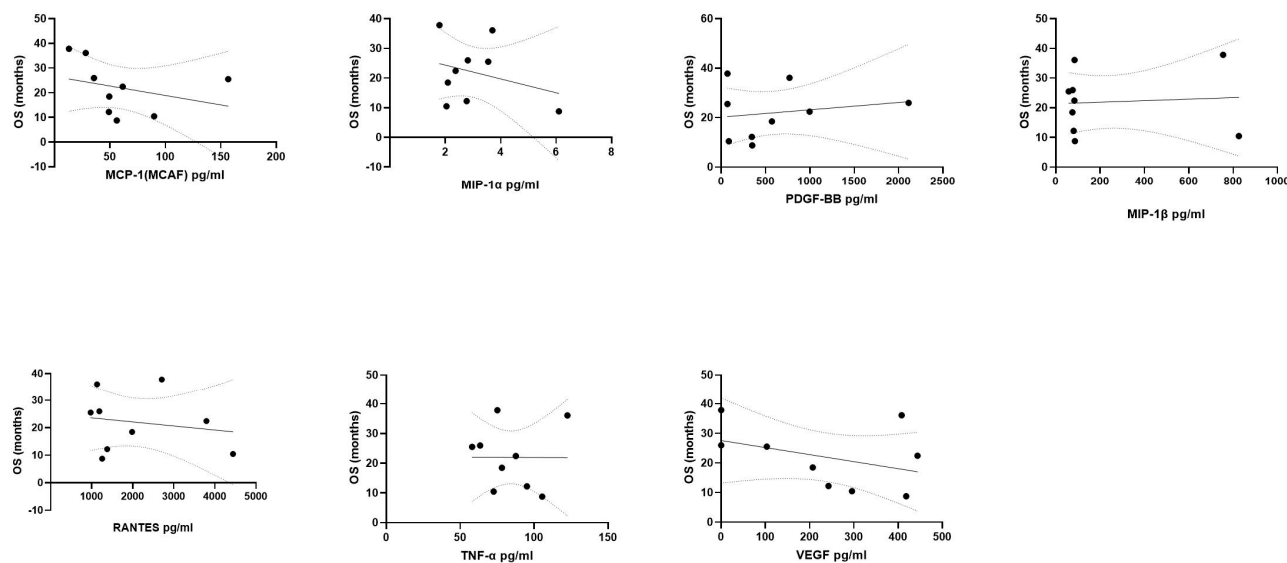


**E**

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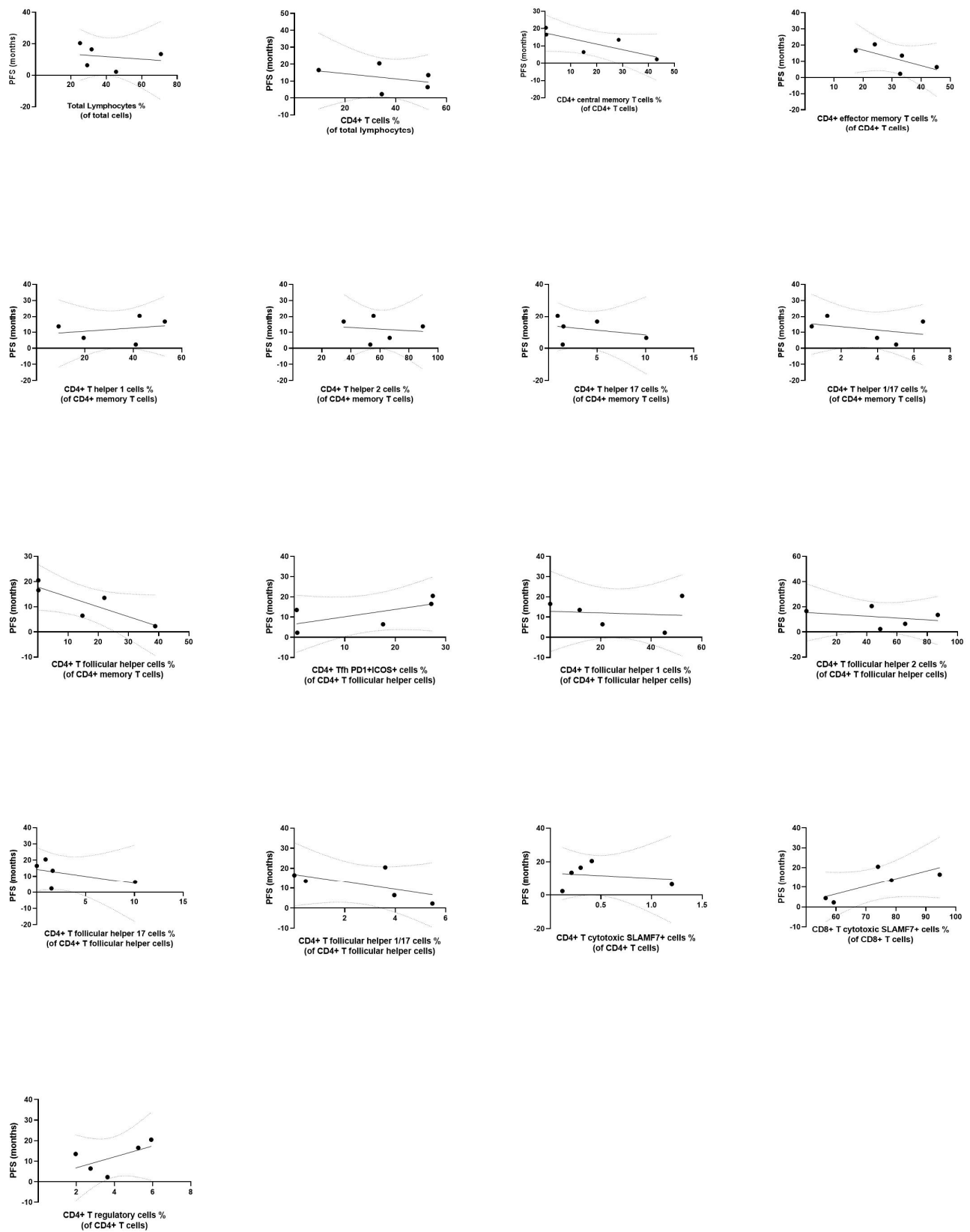


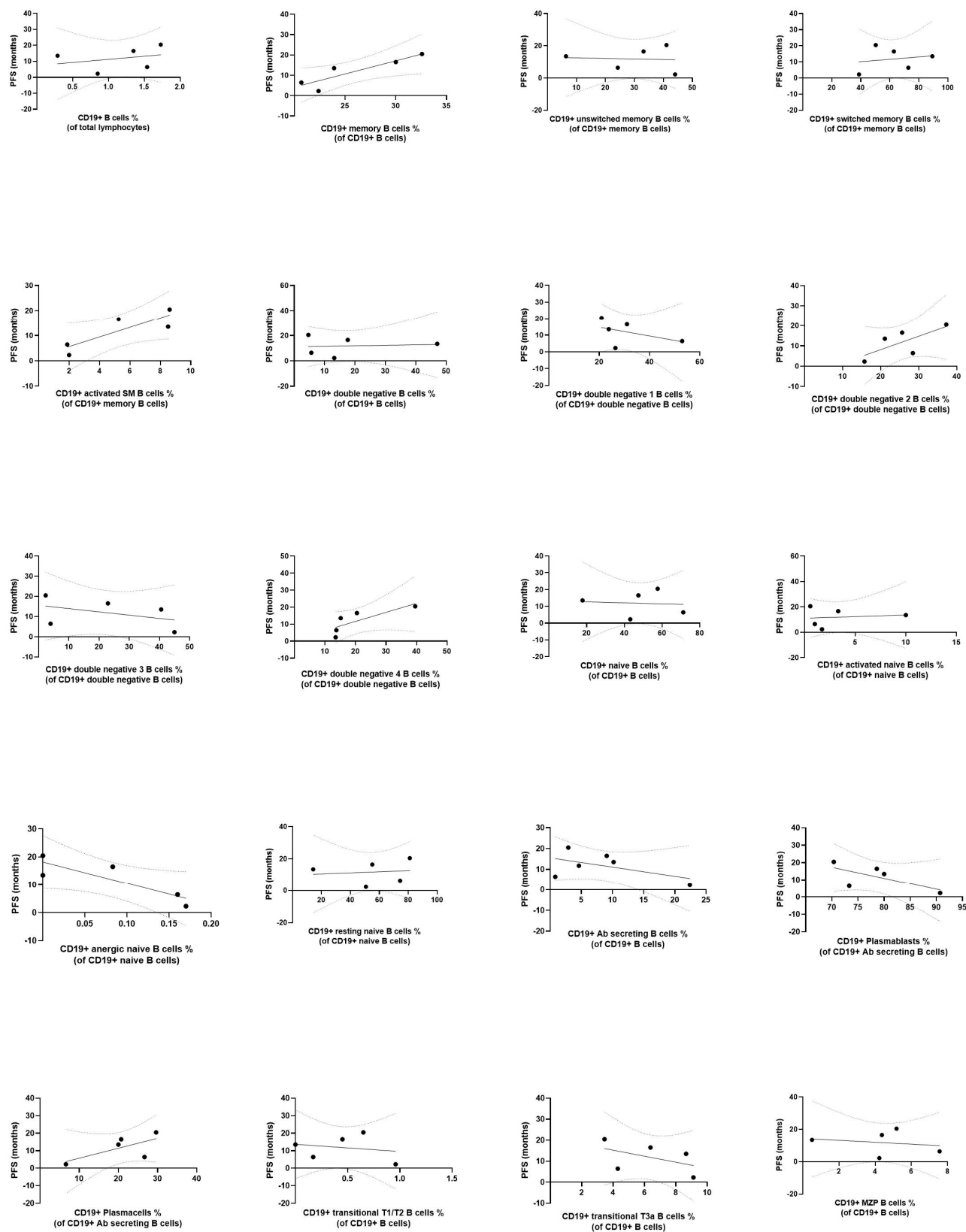


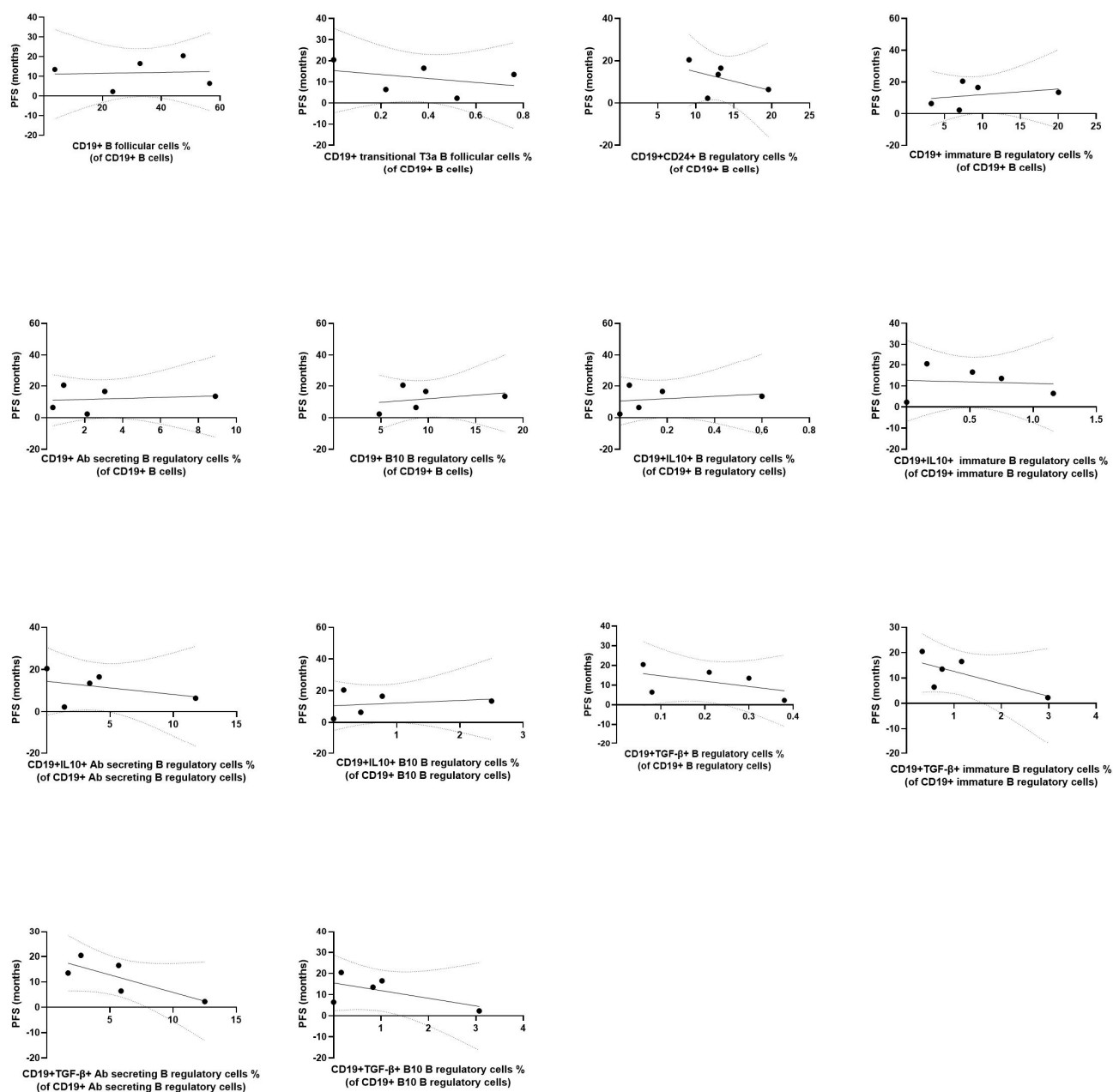


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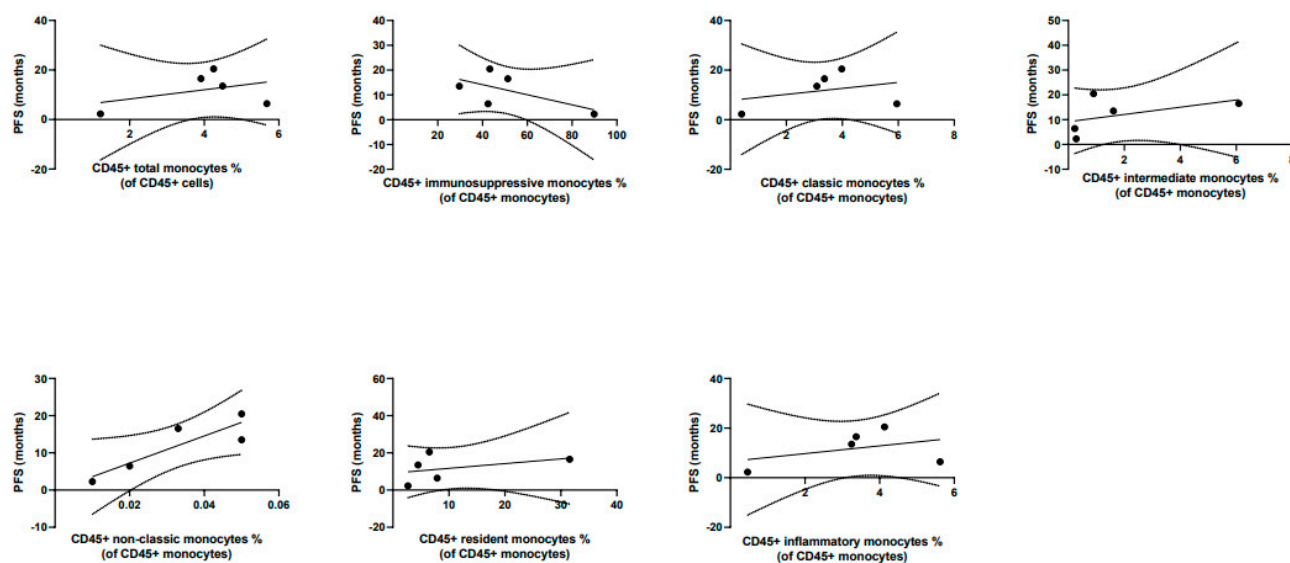
**Figure S9.** Correlation studies between circulating immune cells (A, G: T-cell compartment; B, H: B-cell compartment; C, I: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D, J: Pro Human Inflammation Panel I Assay; E, K: Pro TGF- $\beta$  Immunoassay; F, L: Pro Human Cytokine Immunoassay) at 4-month follow-up and progression-free survival (PFS) (A-F) and overall survival (OS) (G-L) after therapy onset in patients with pancreatic adenocarcinoma (PDAC) of the HybridTherm ablation plus chemotherapy (HTP-CT) arm. Correlation studies were performed using the Spearman's rho rank correlation test.

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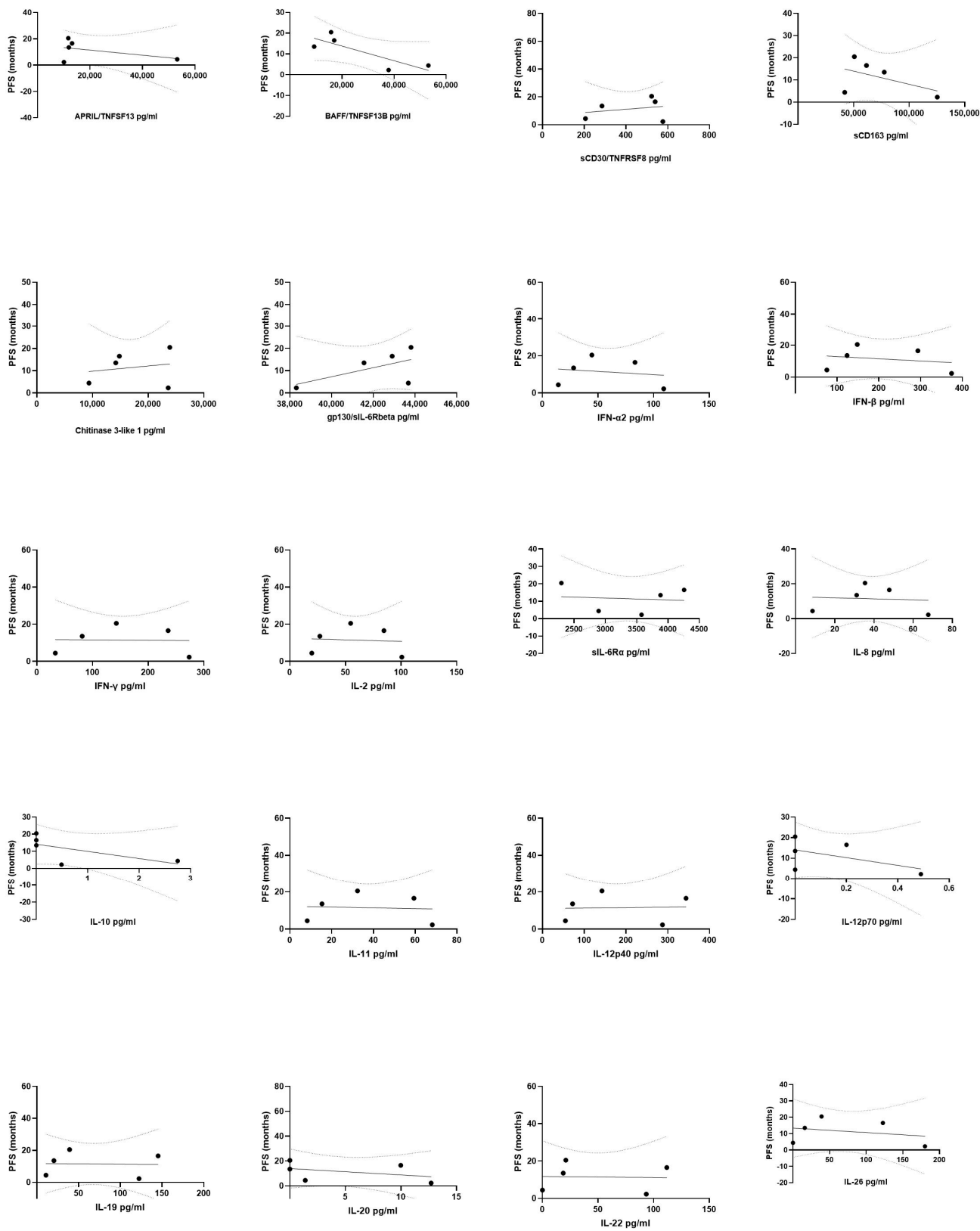
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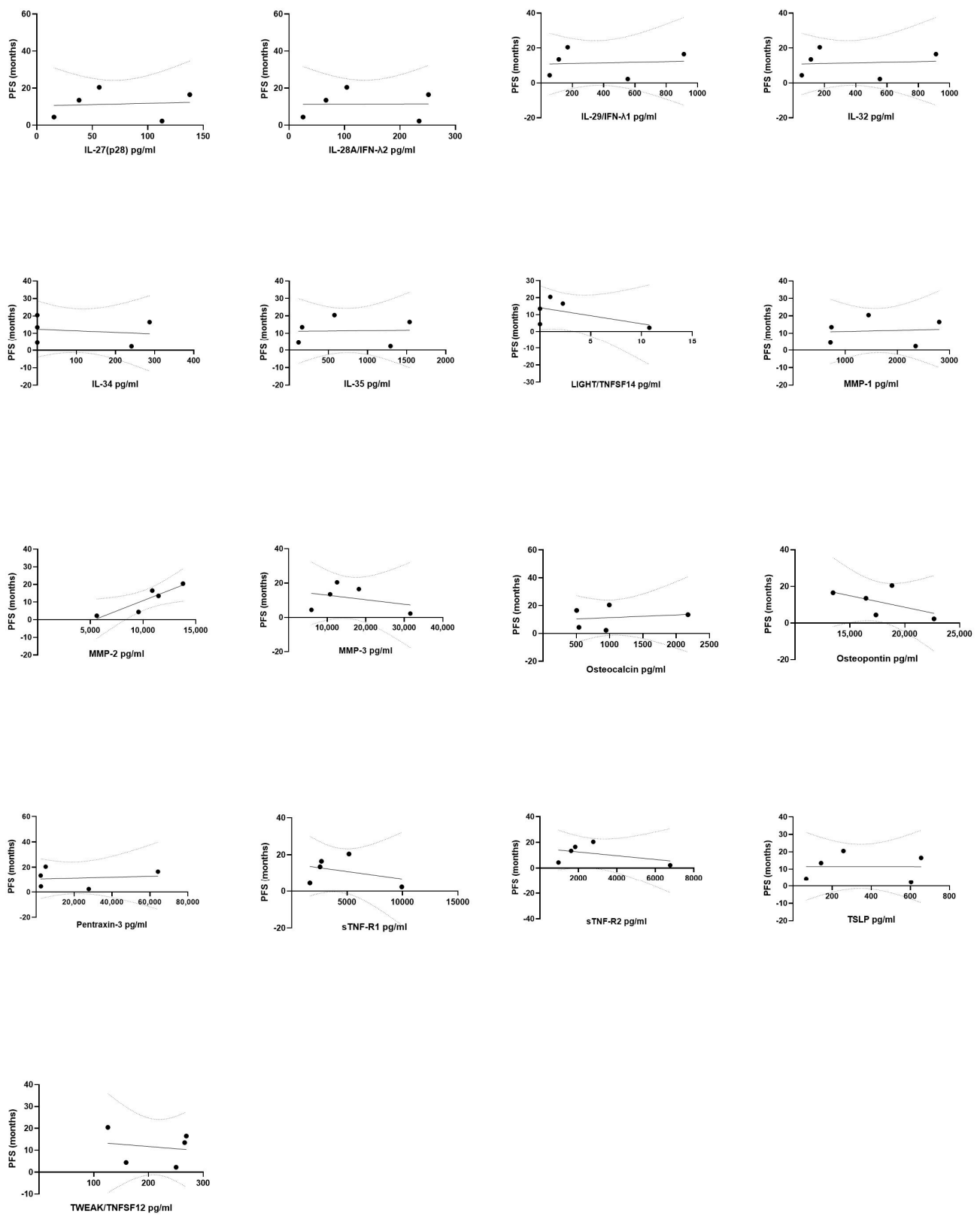


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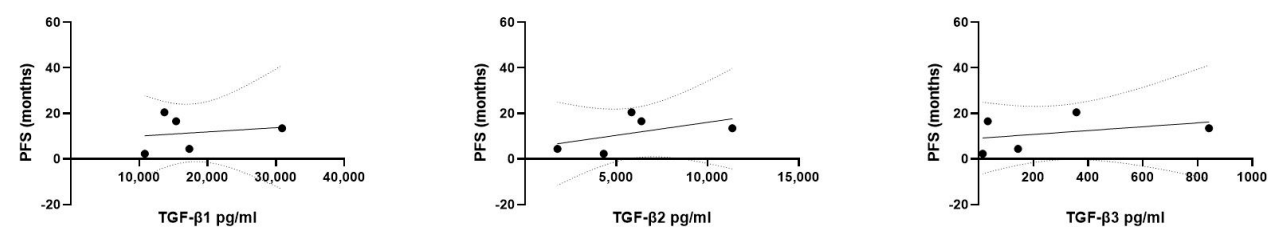
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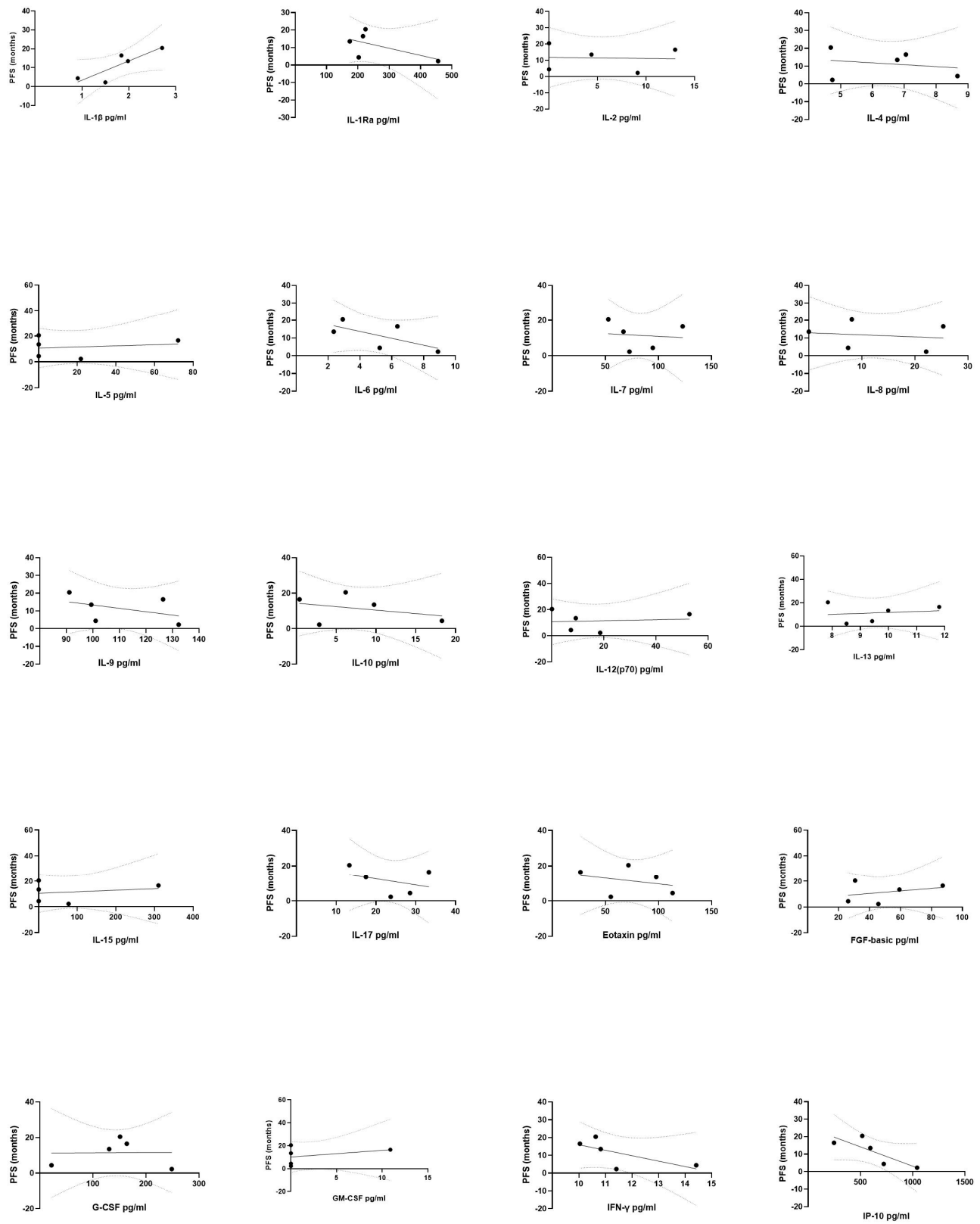


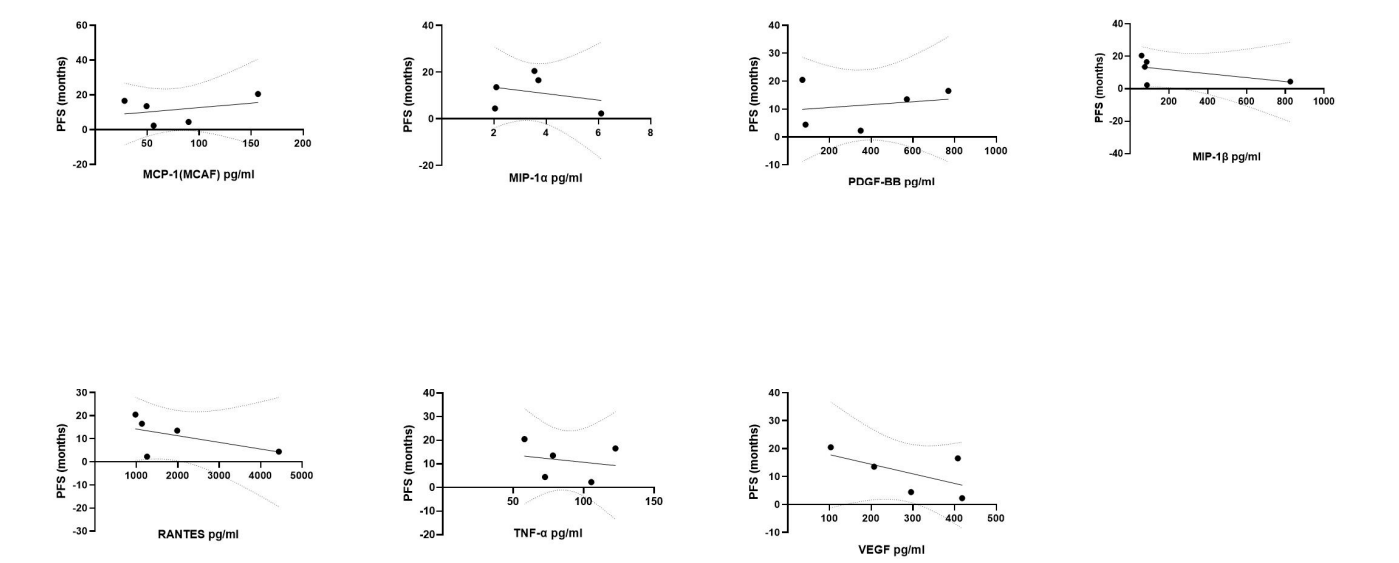


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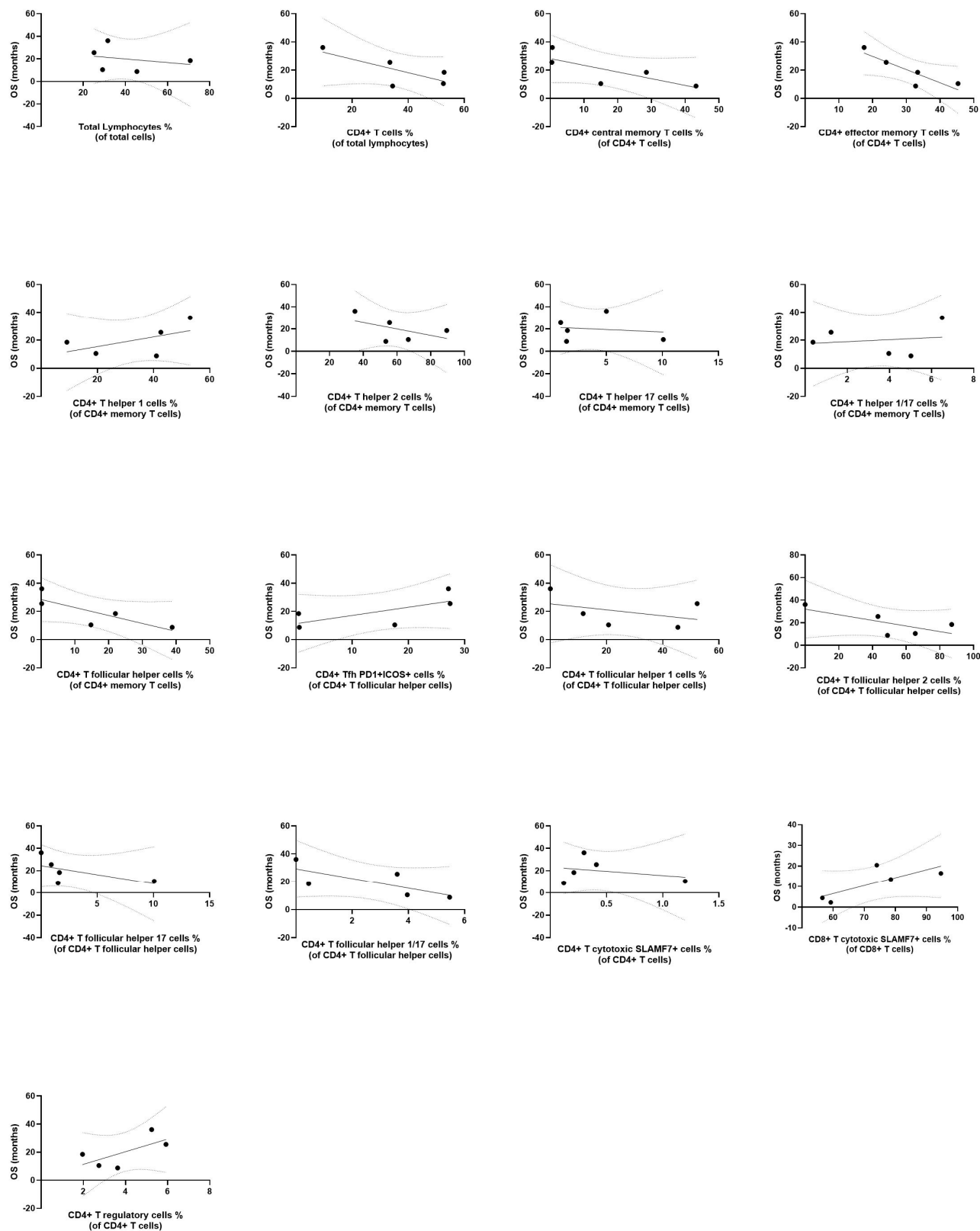




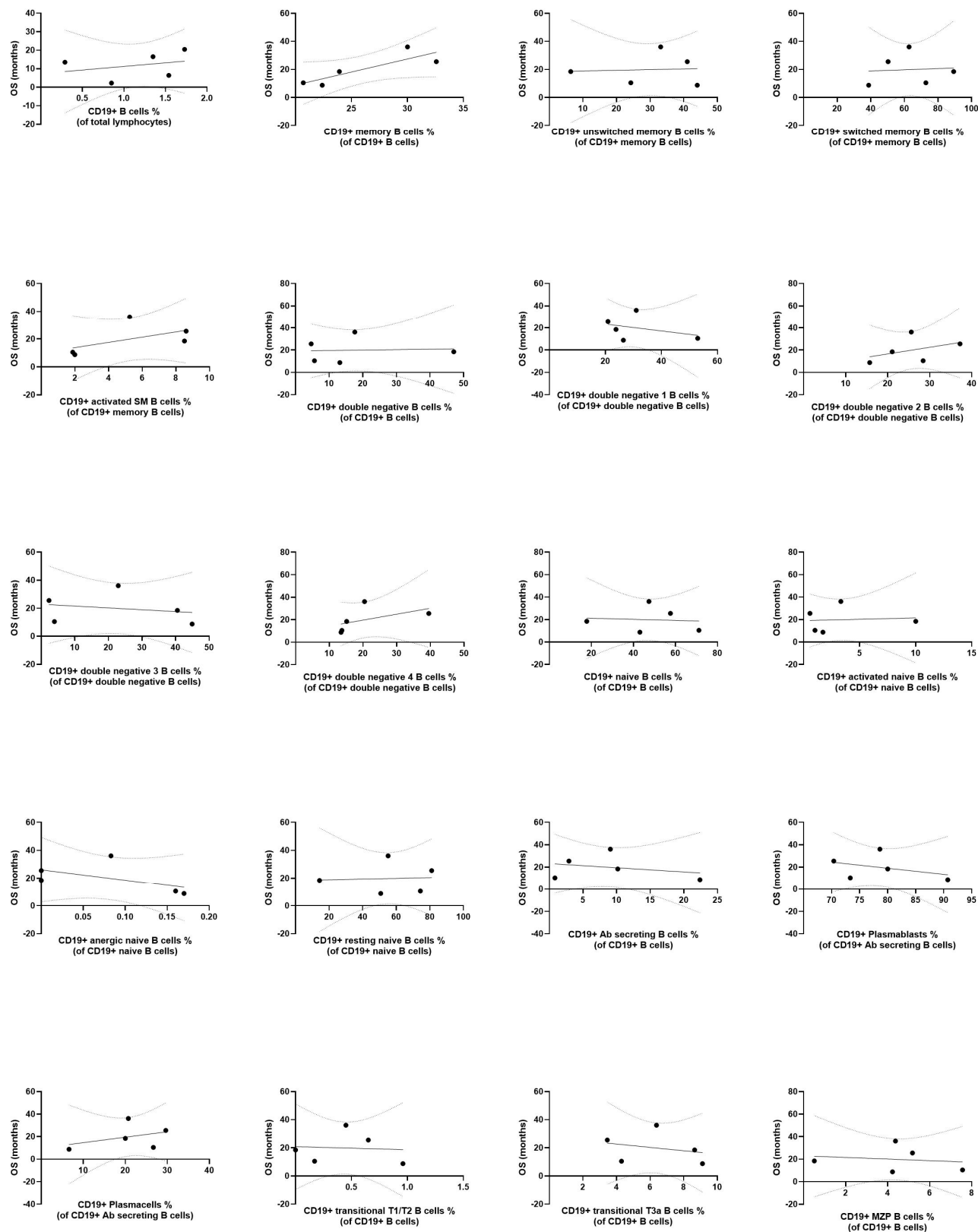
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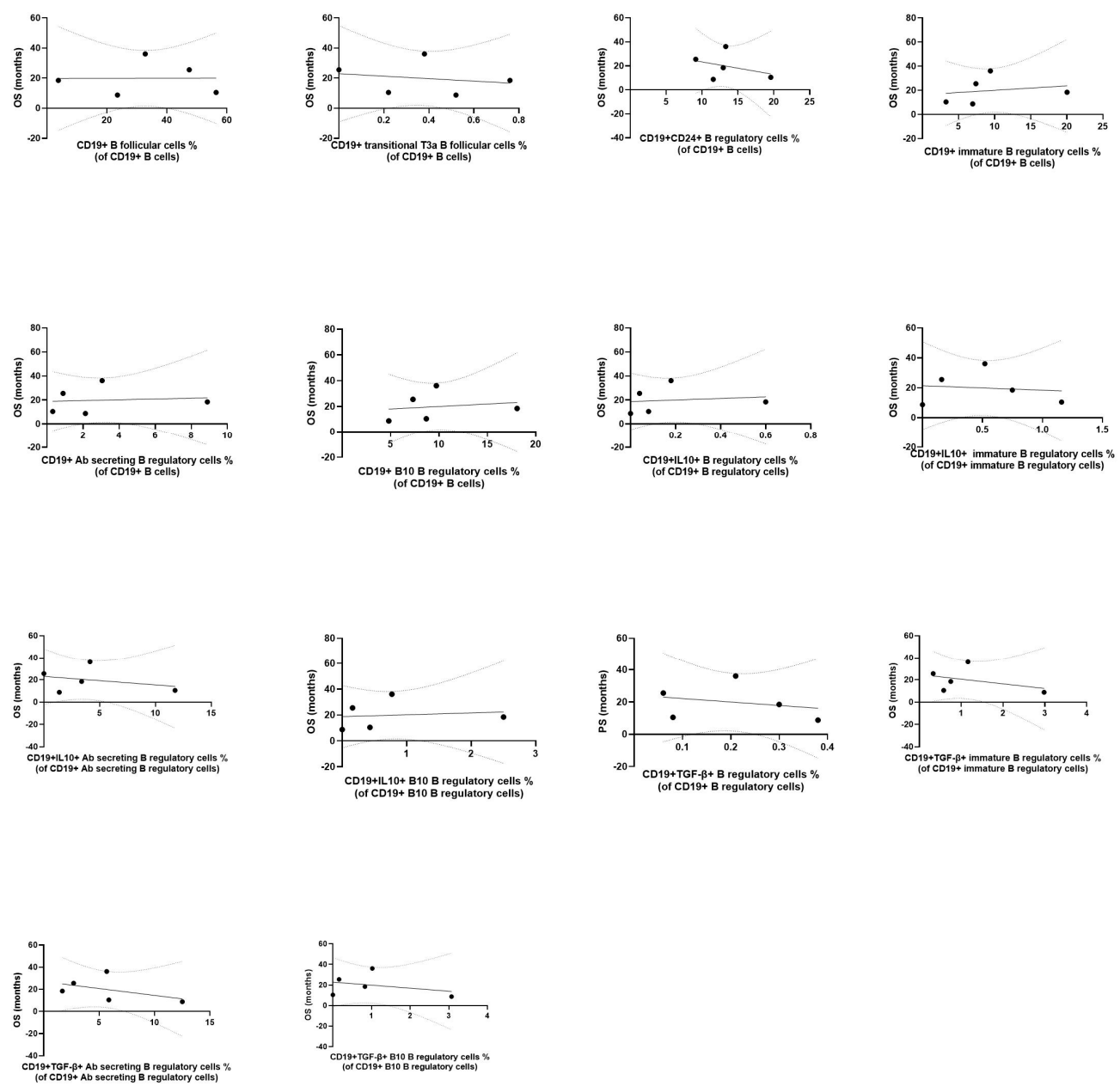
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G



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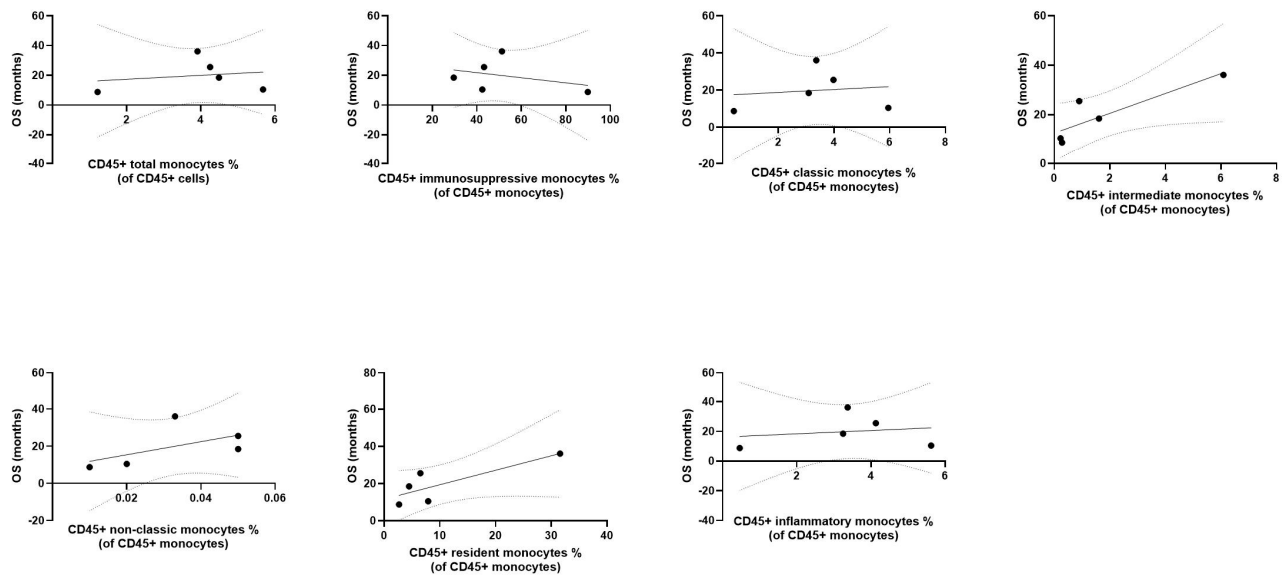


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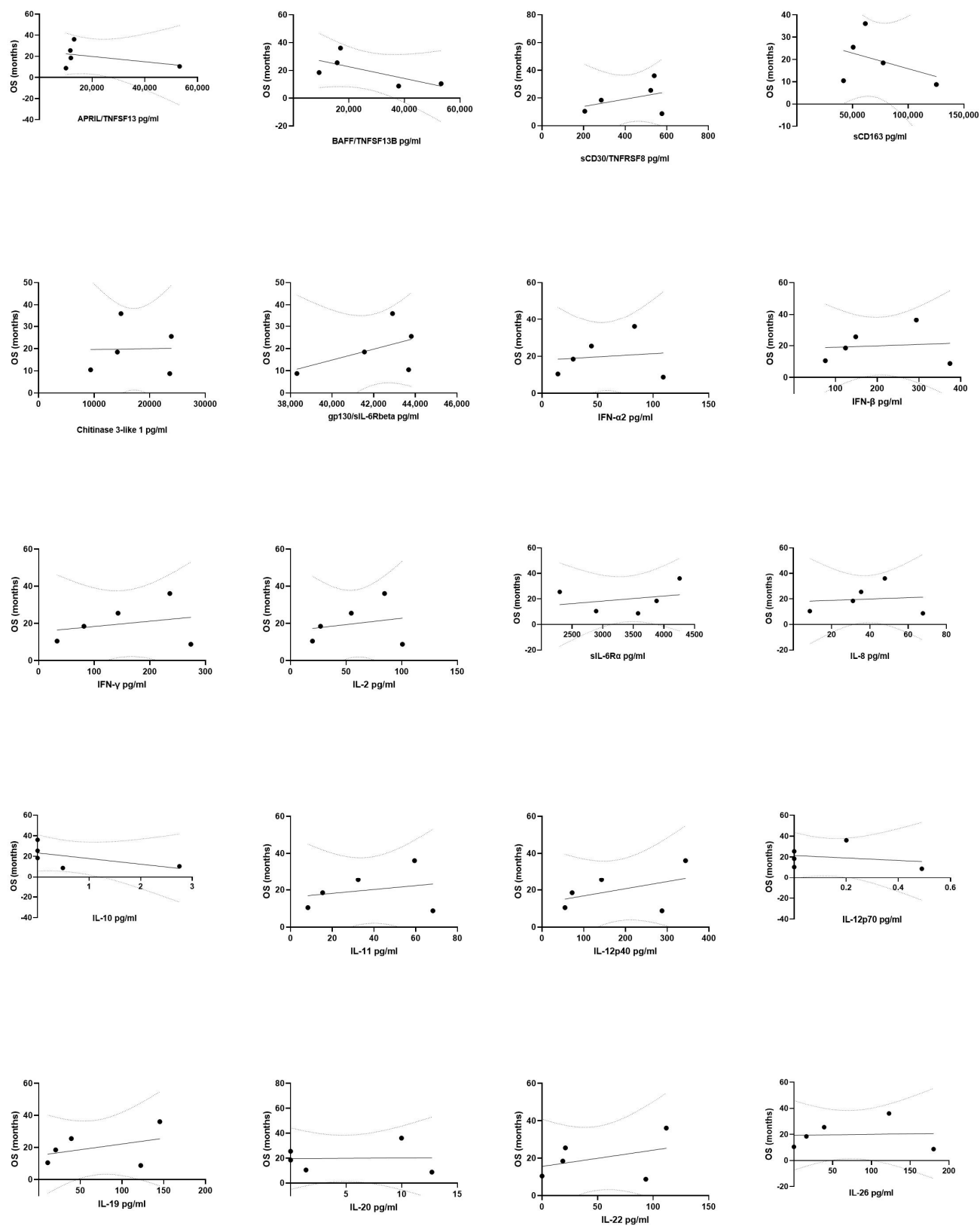
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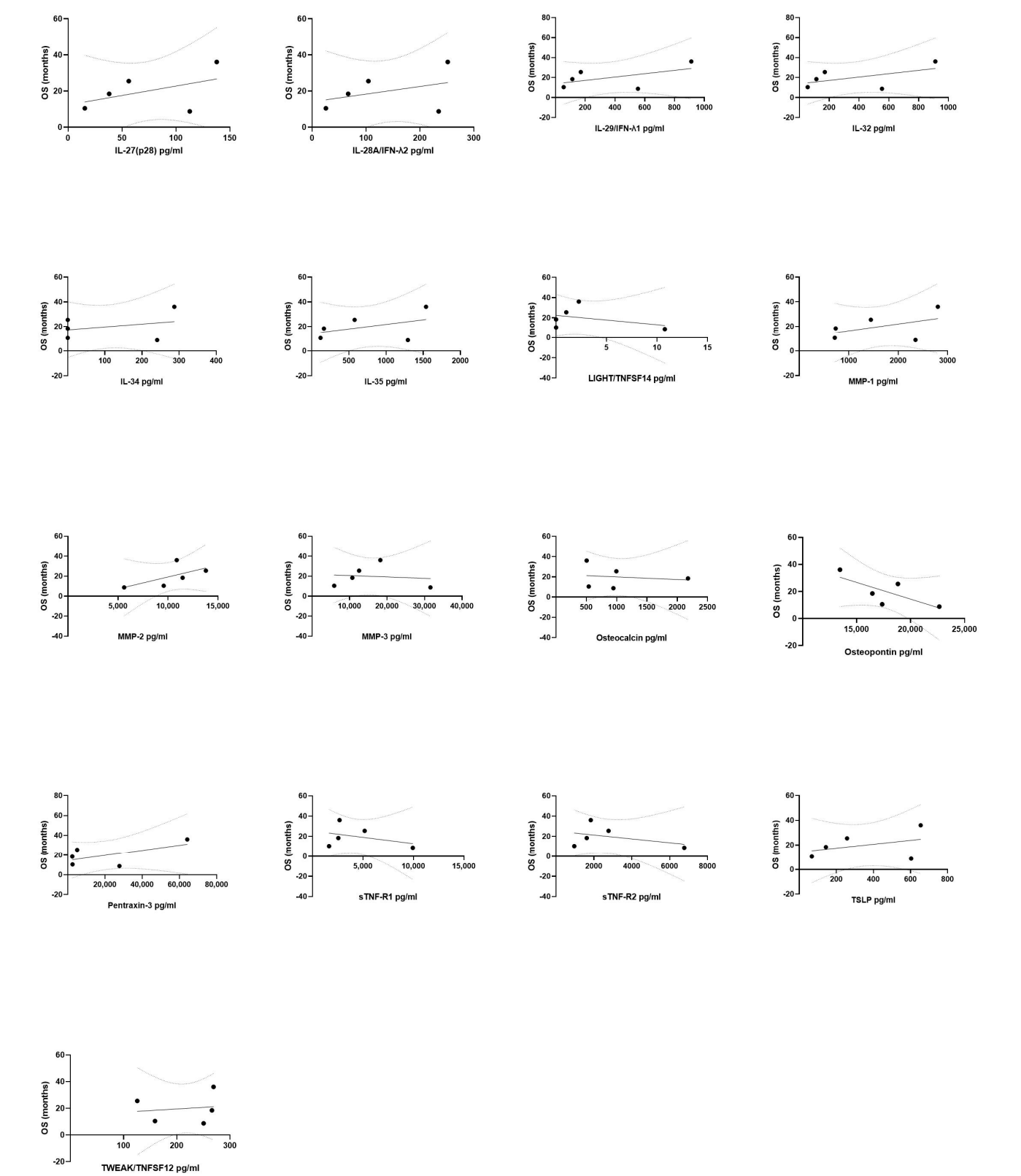
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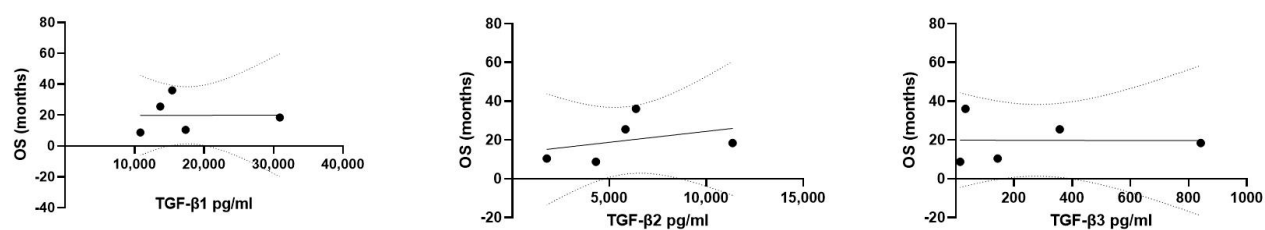




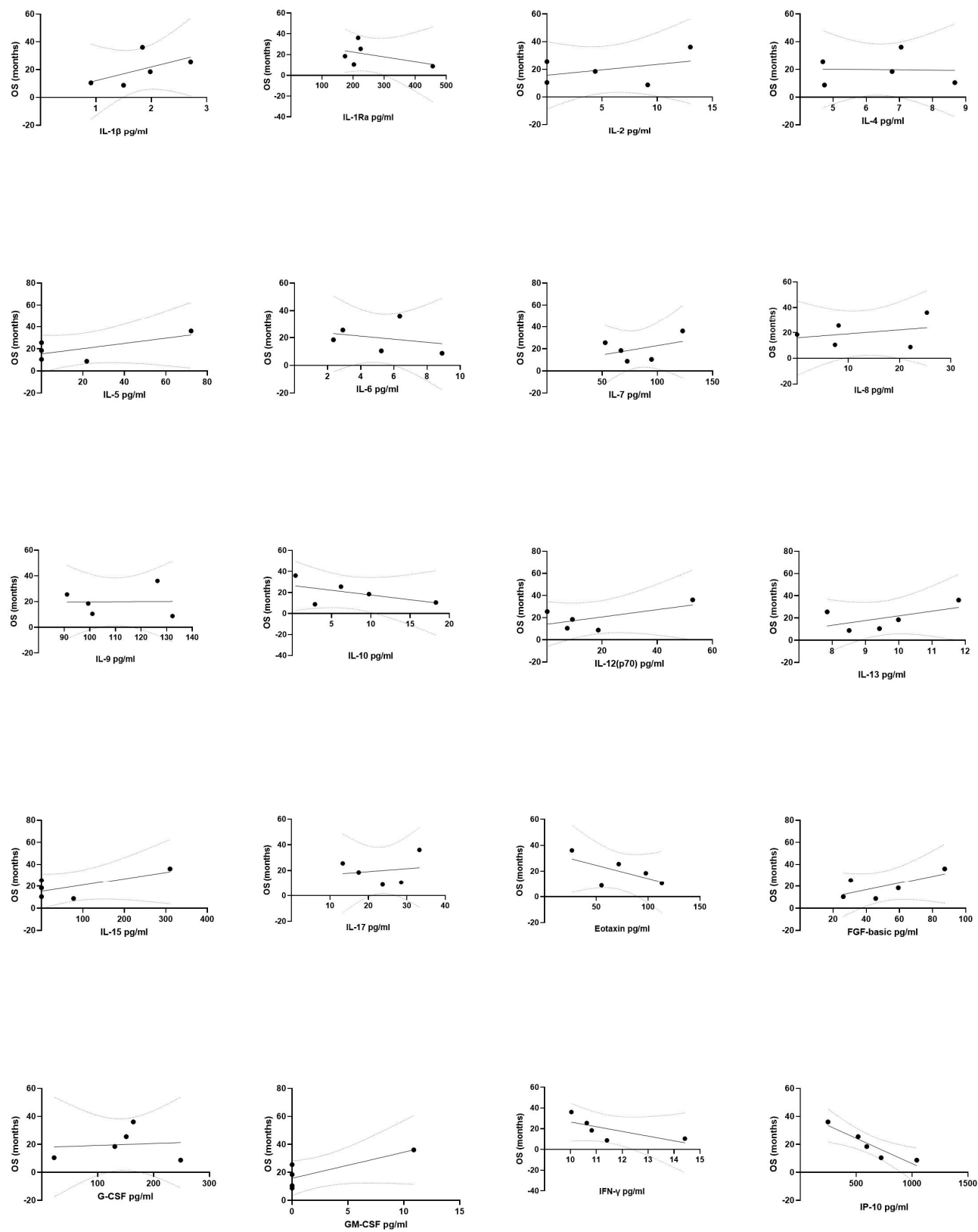
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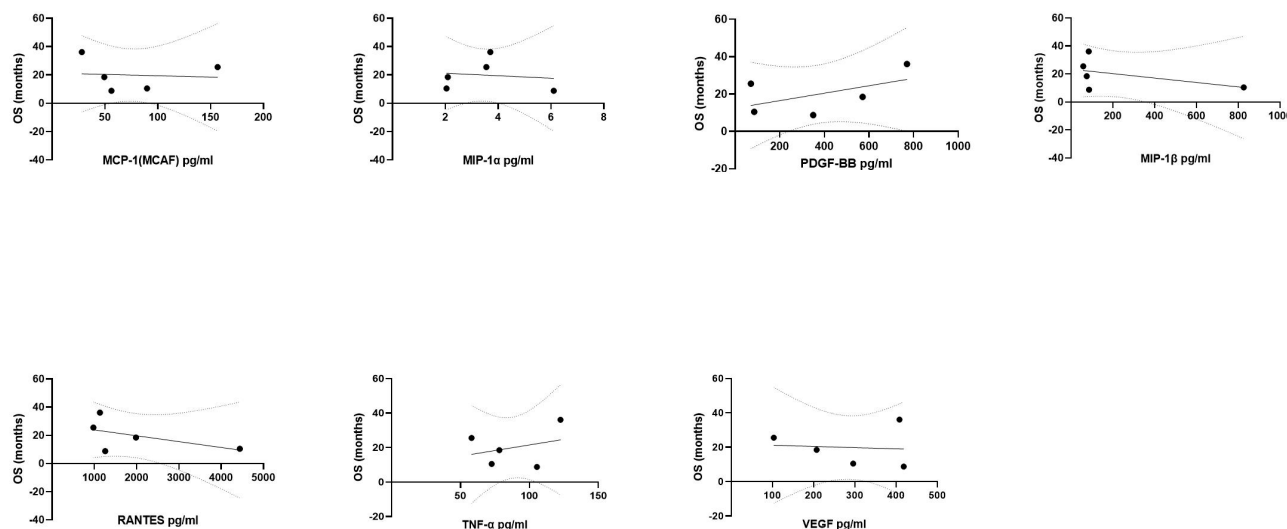
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**K**

L

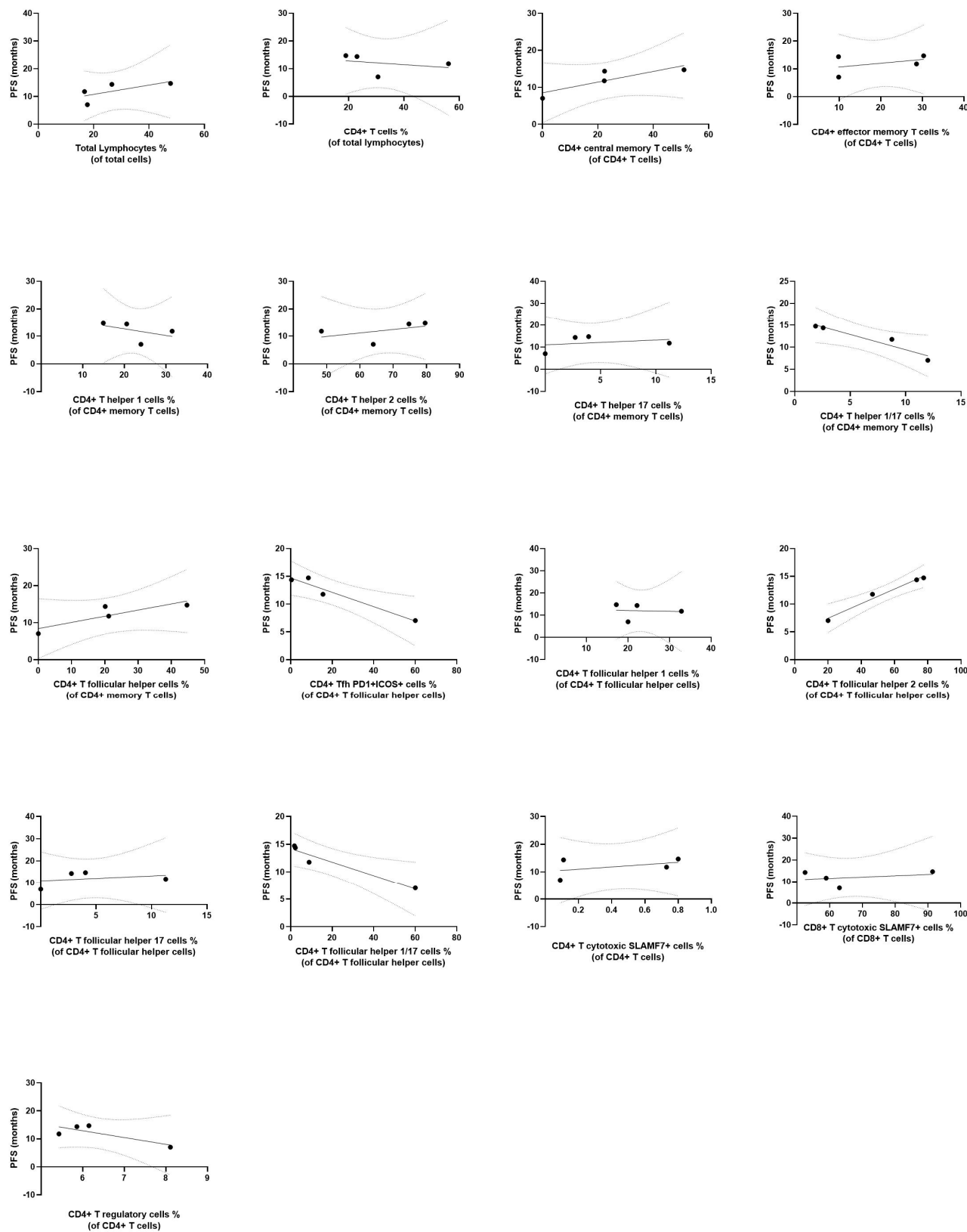




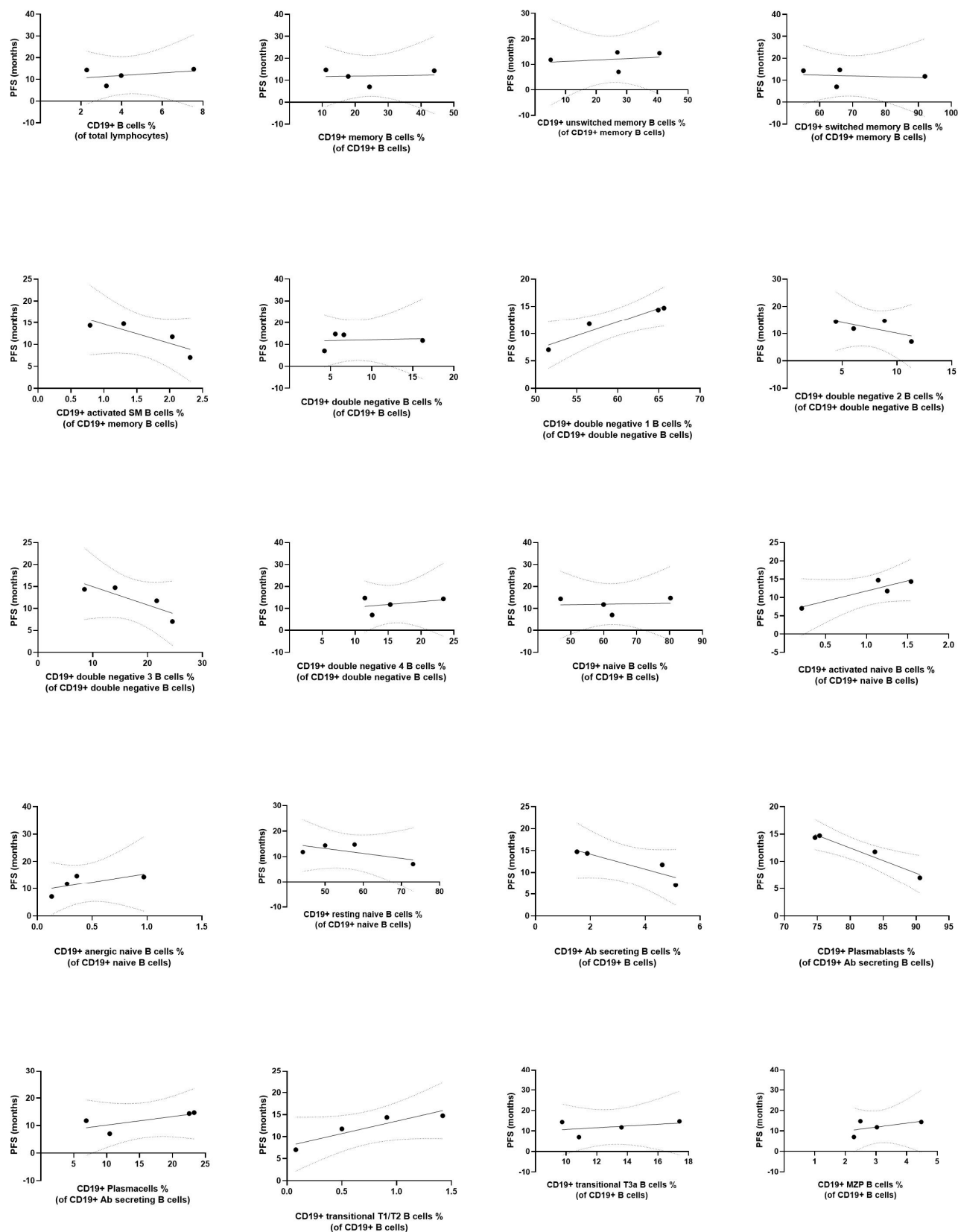
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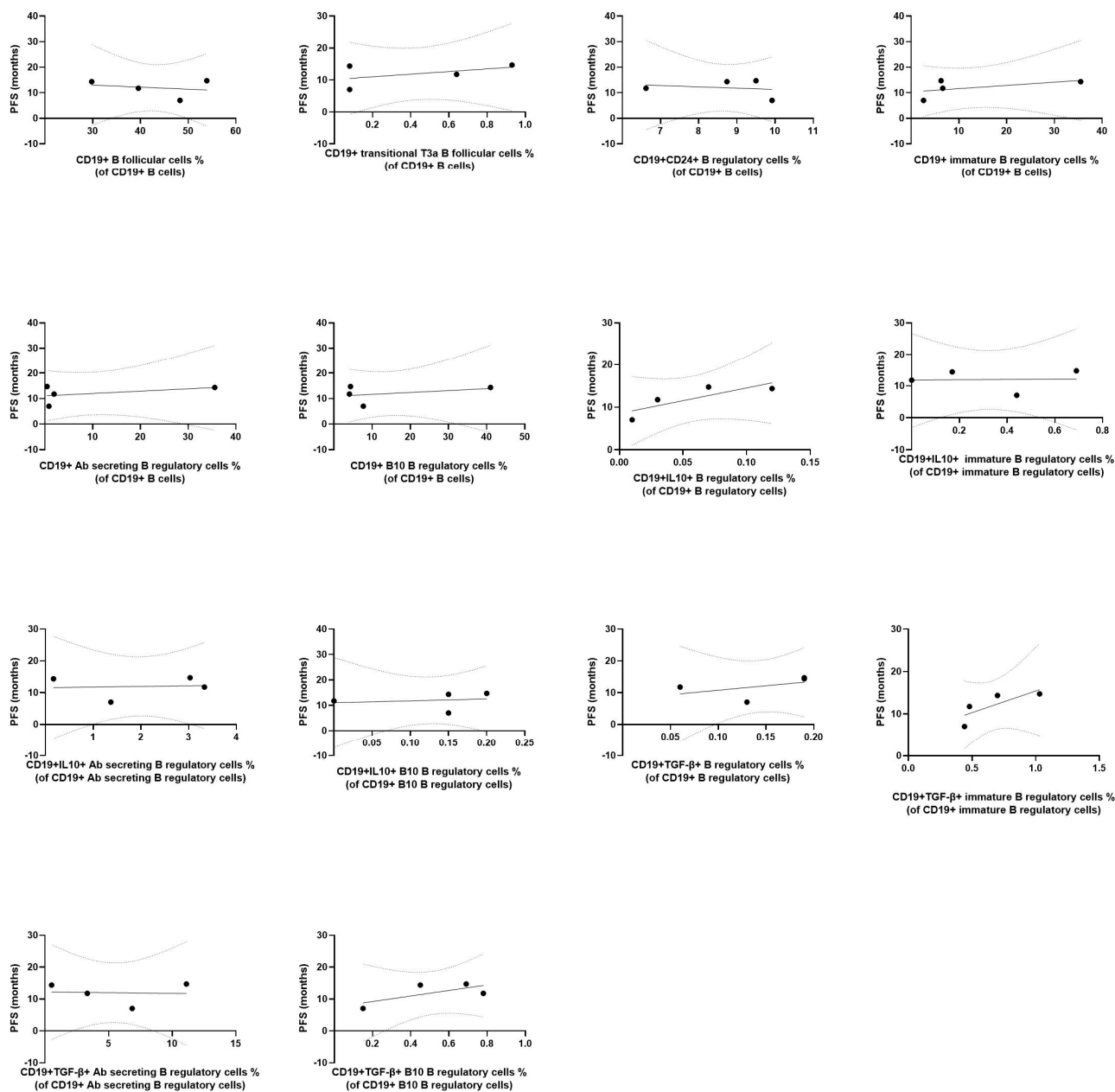
**Figure S10.** Correlation studies between circulating immune cells (A, G: T-cell compartment; B, H: B-cell compartment; C, I: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D, J: Pro Human Inflammation Panel I Assay; E, K: Pro TGF- $\beta$  Immunoassay; F, L: Pro Human Cytokine Immunoassay) at 4-month follow-up and progression-free survival (PFS) (A-F) and overall survival (OS) (G-L) after therapy onset in patients with pancreatic adenocarcinoma (PDAC) of the chemotherapy (CT) arm. Correlation studies were performed using the Spearman's rho rank correlation test.

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**A**

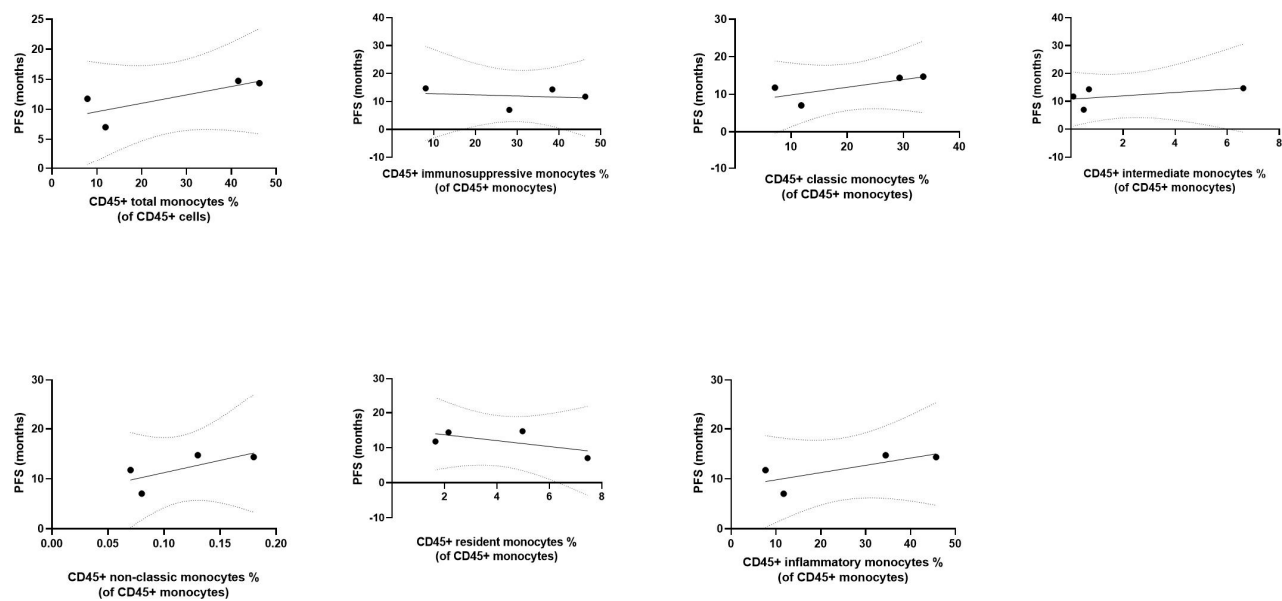
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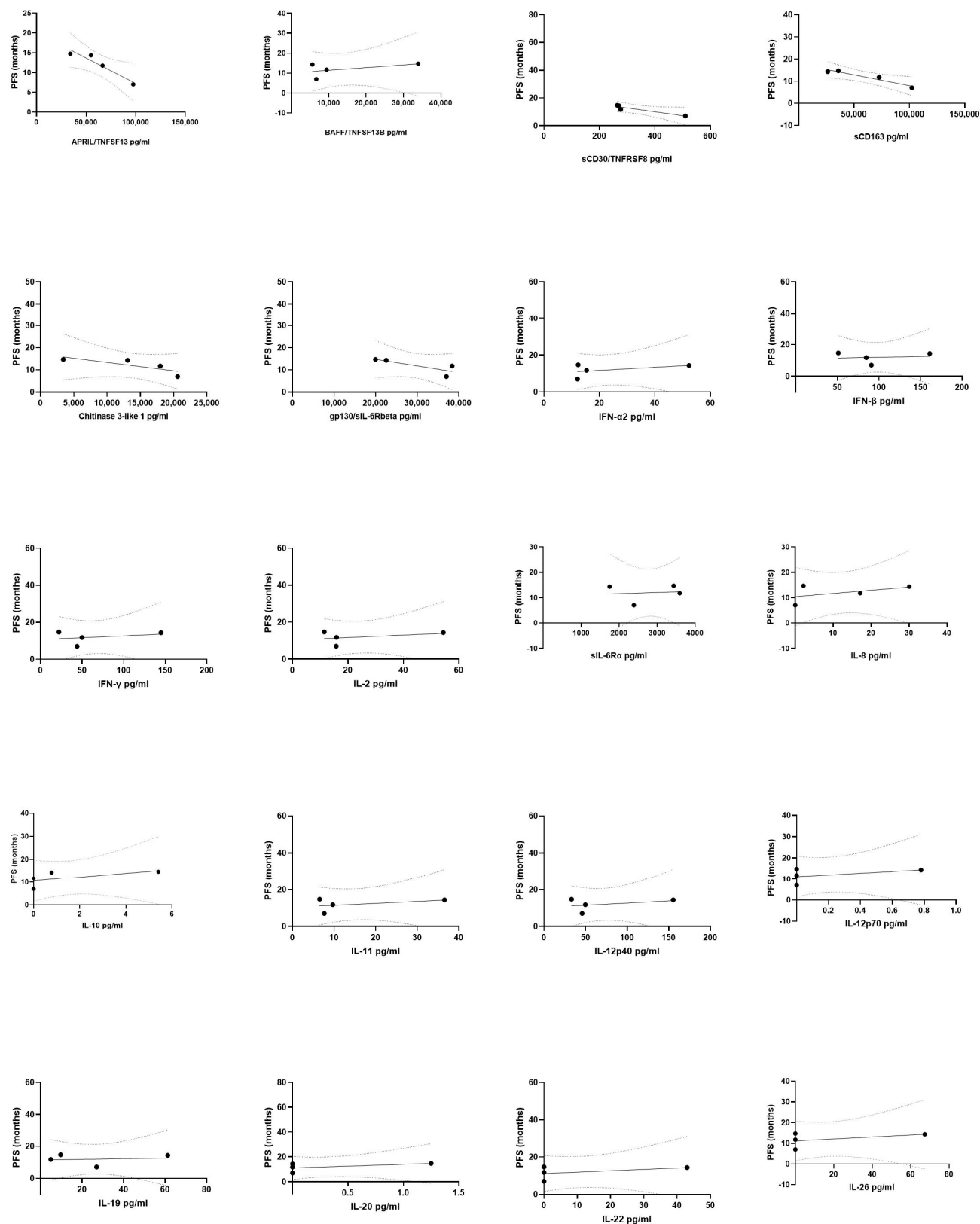


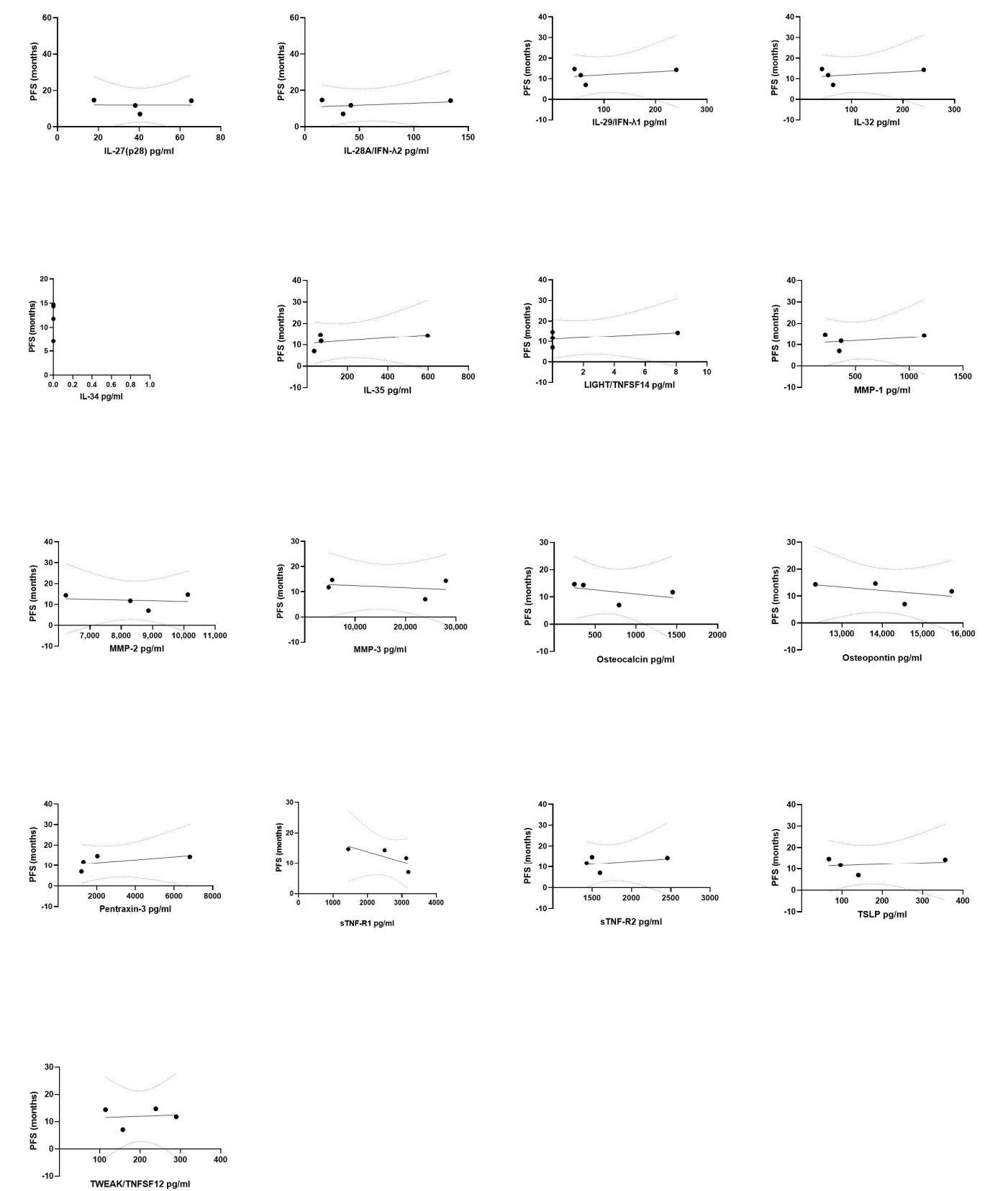


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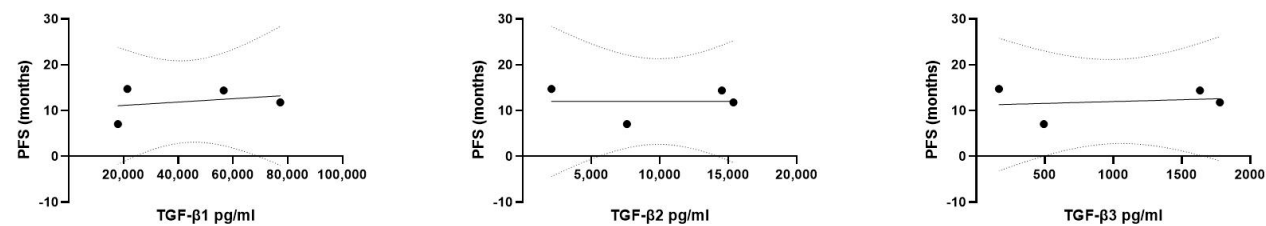


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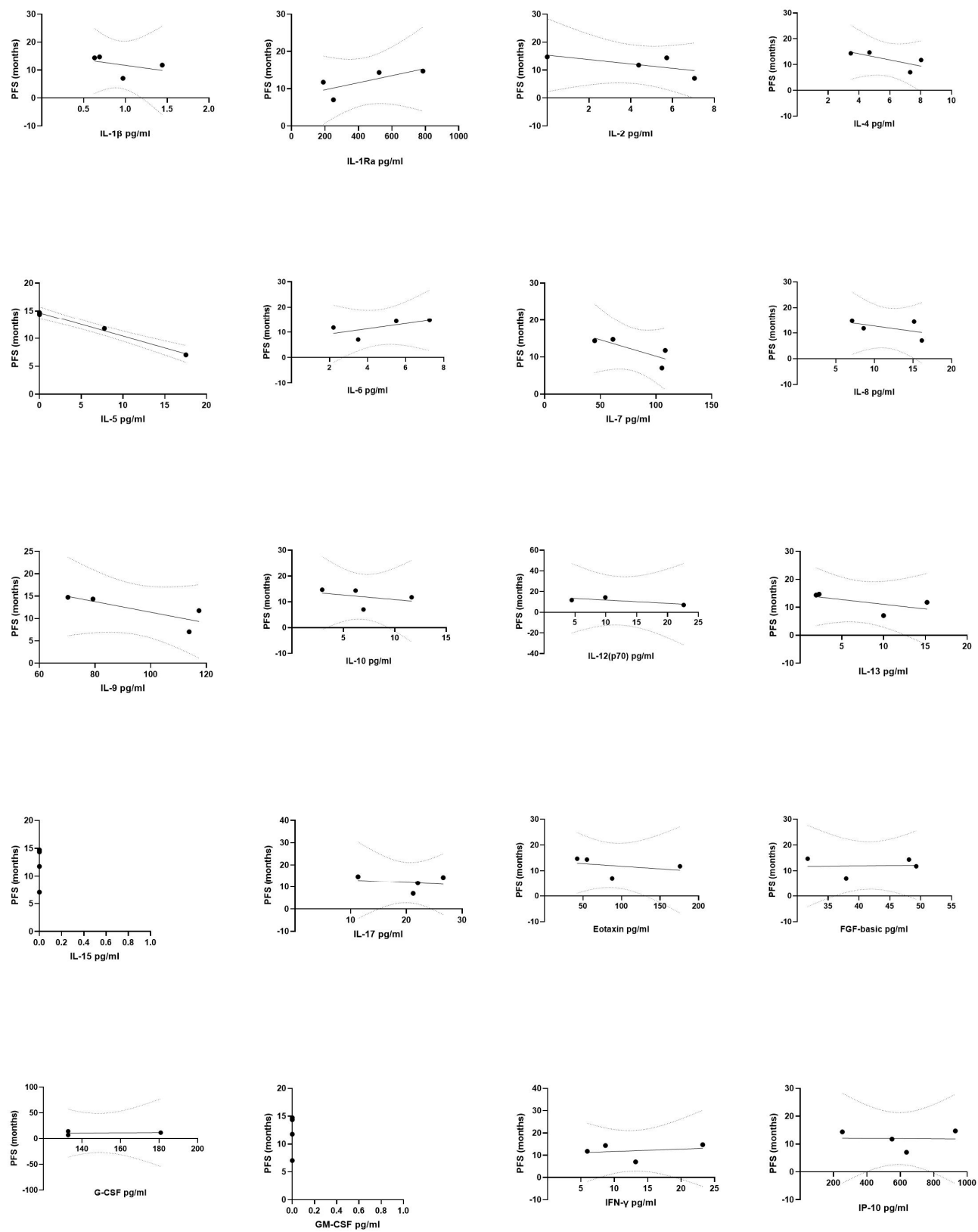


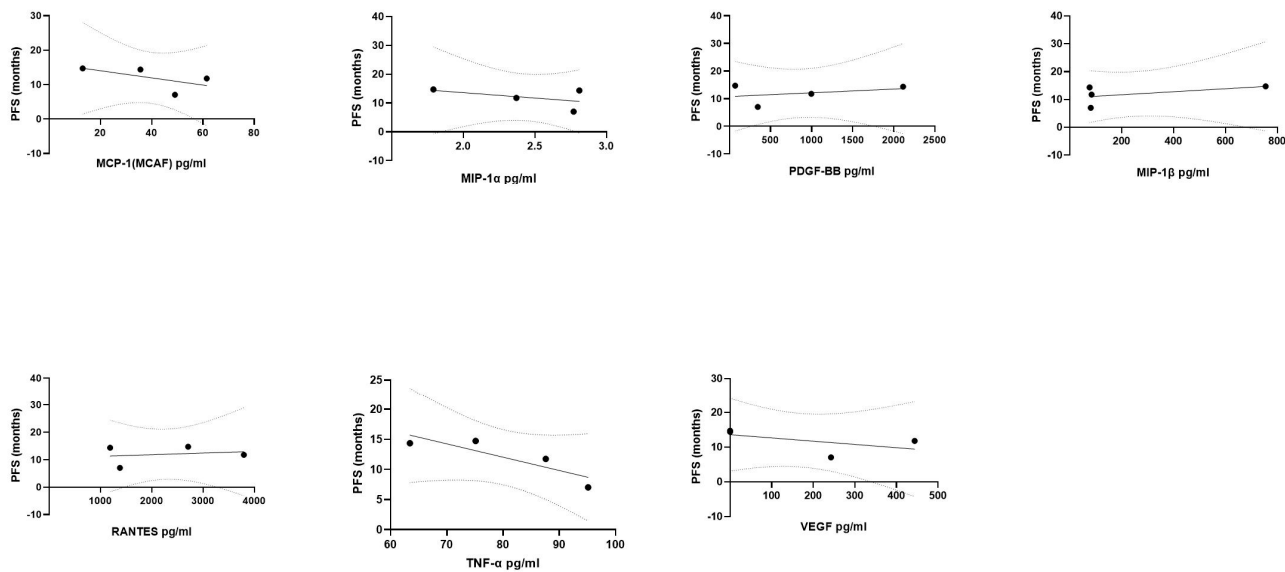


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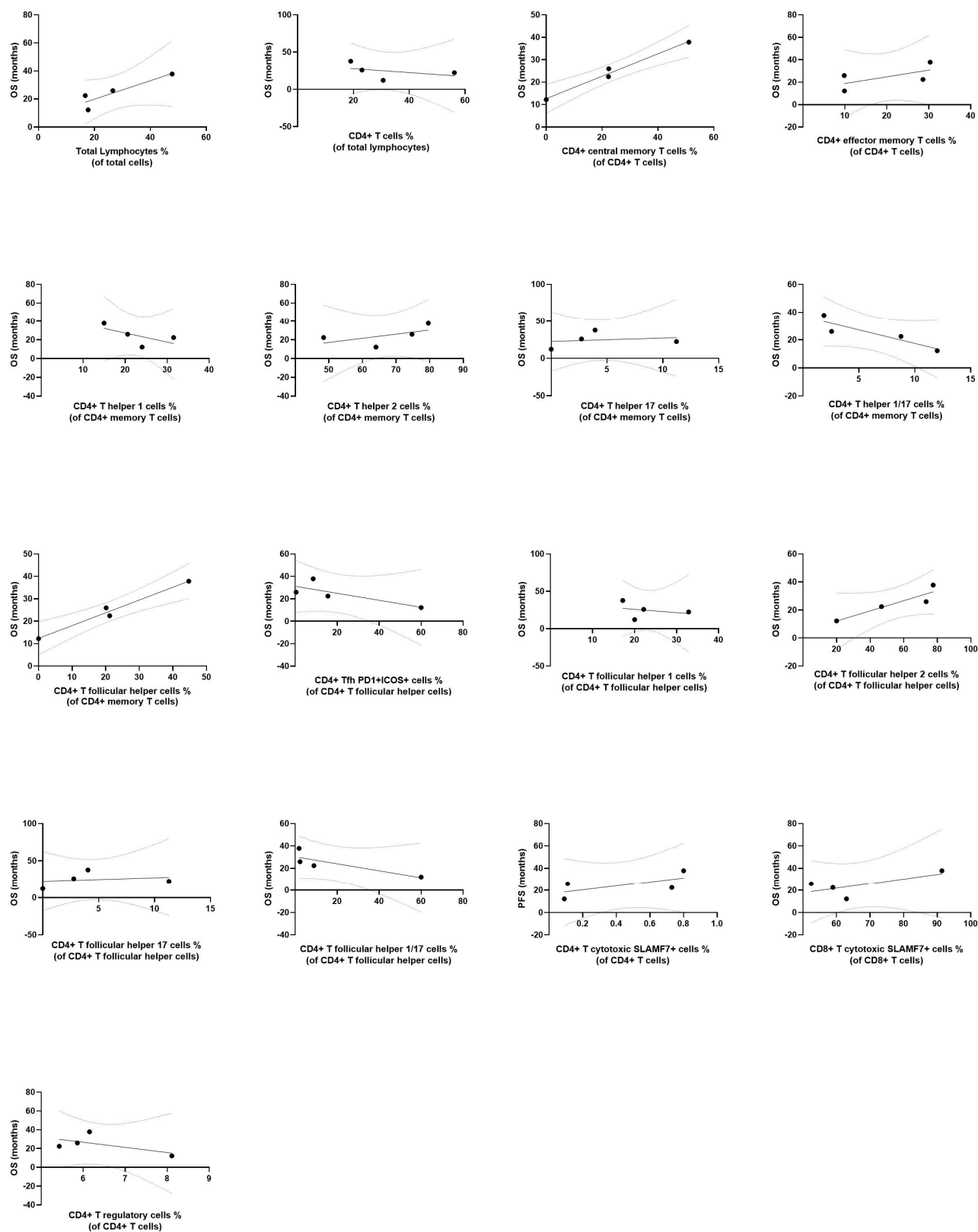


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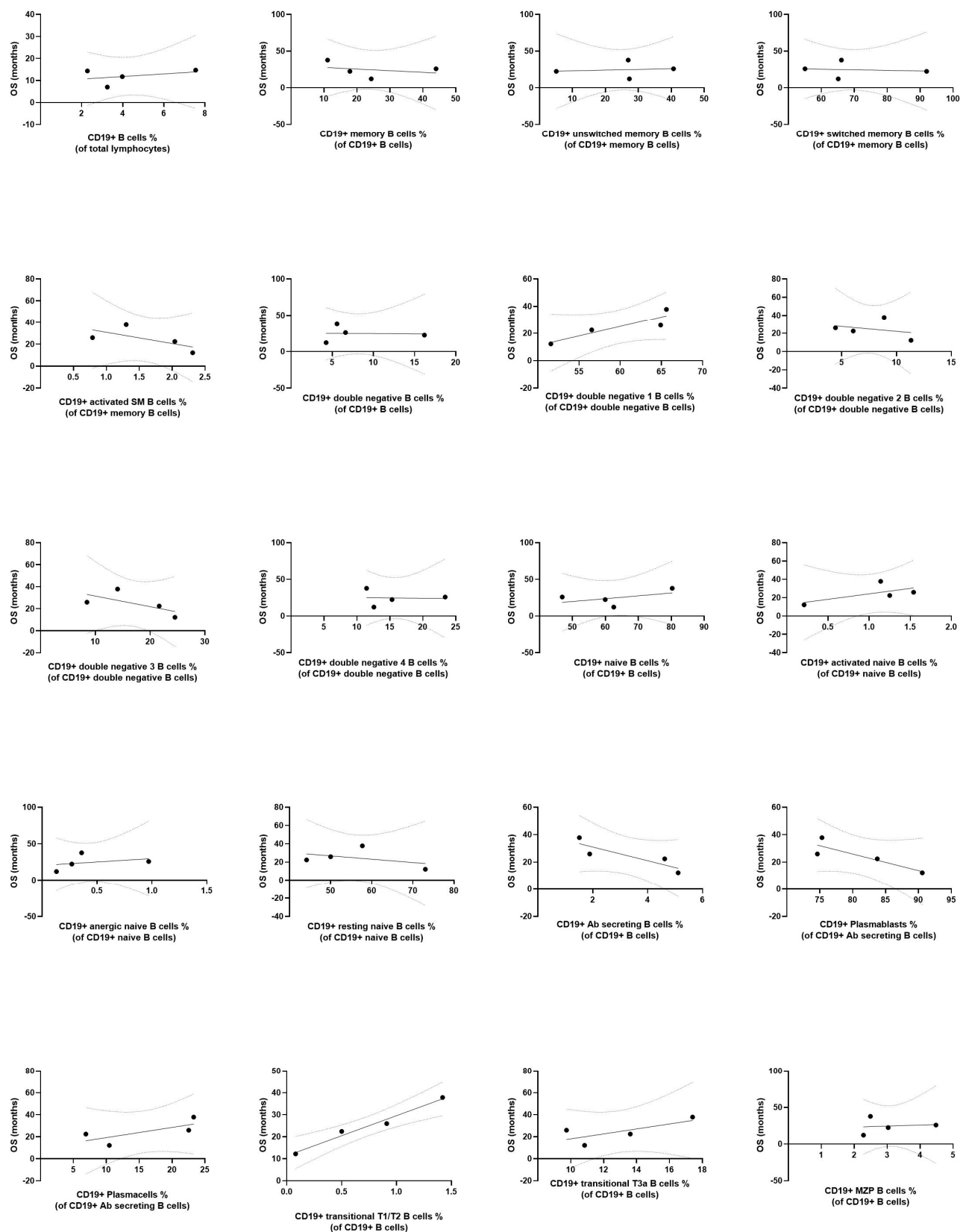




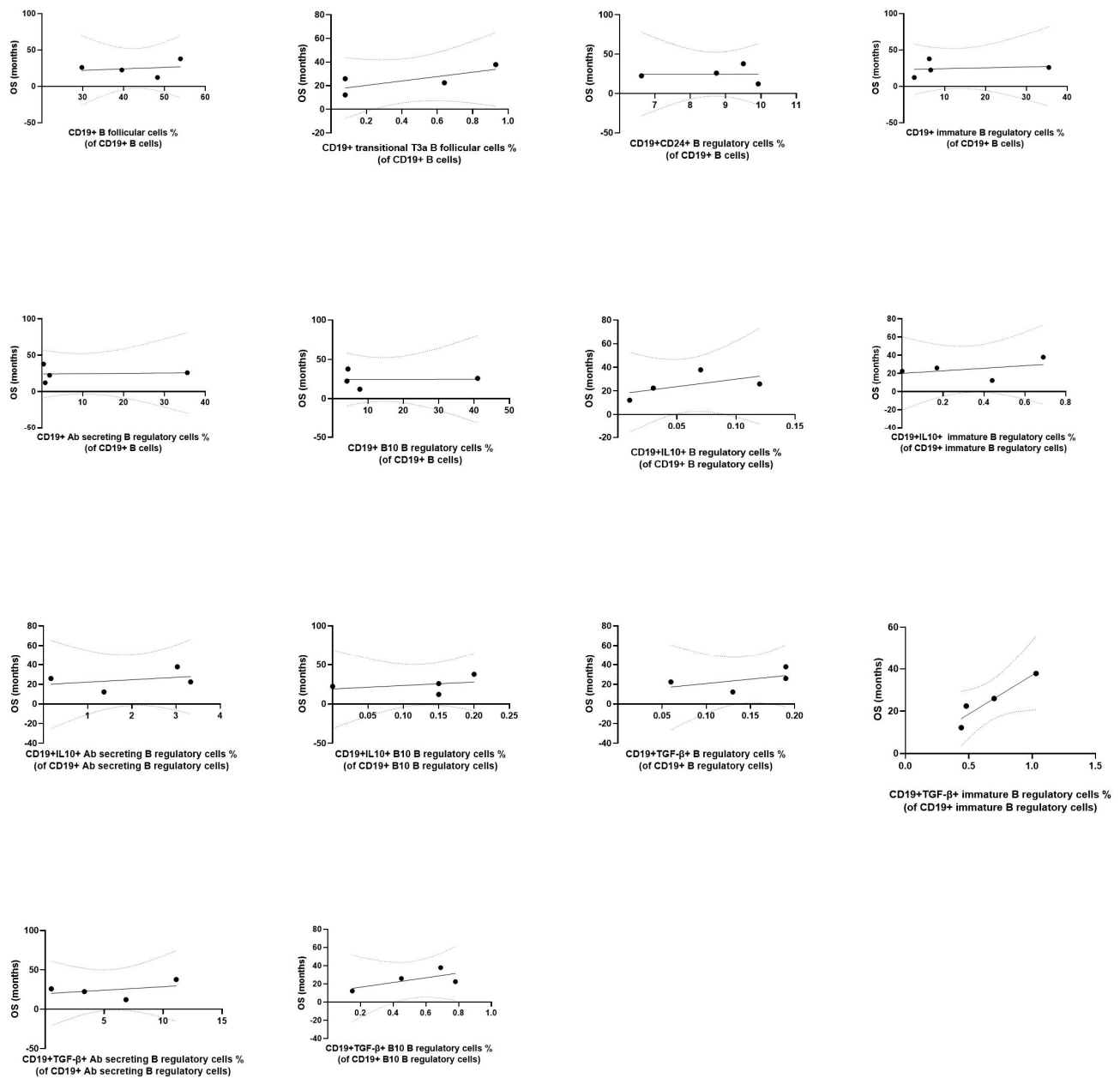
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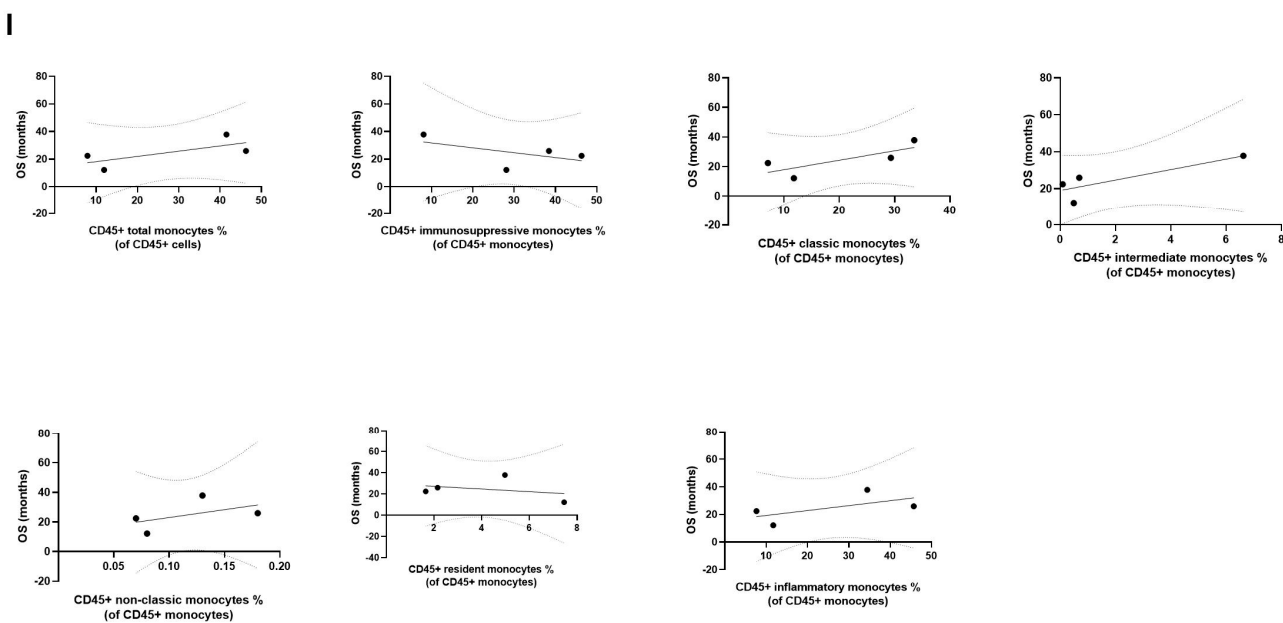


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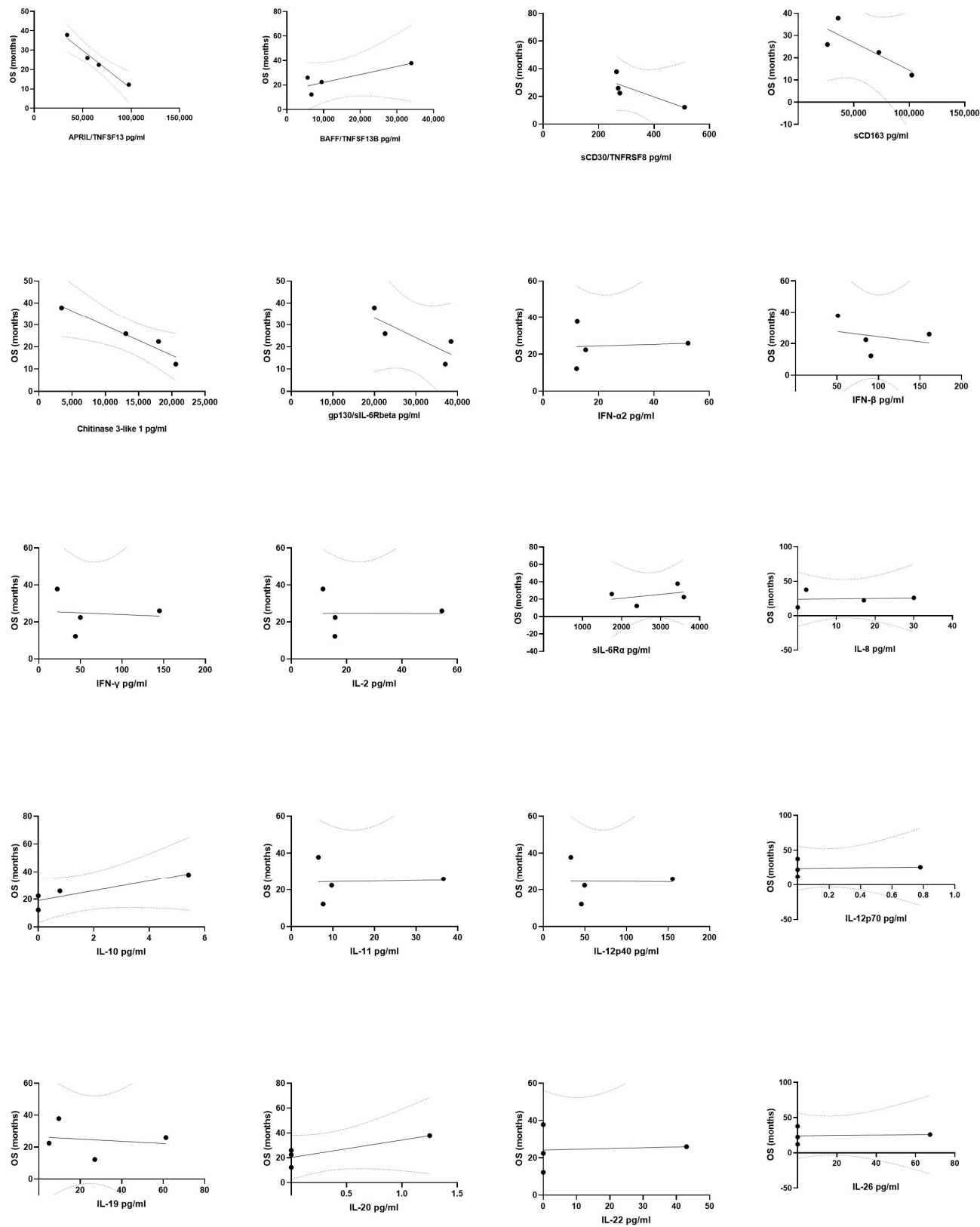


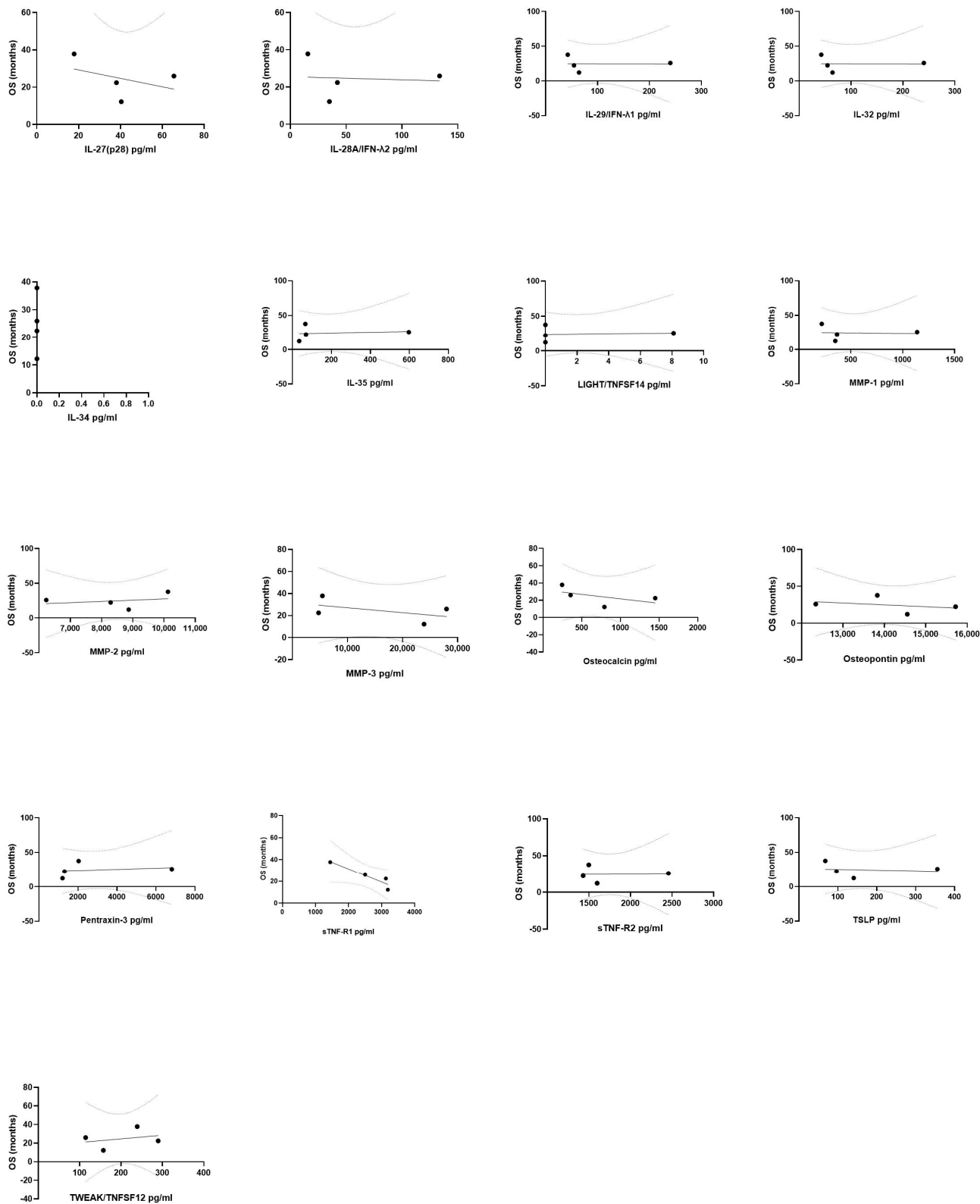






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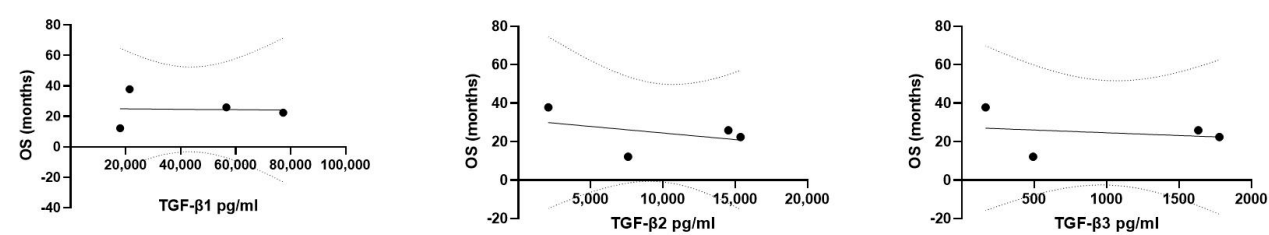
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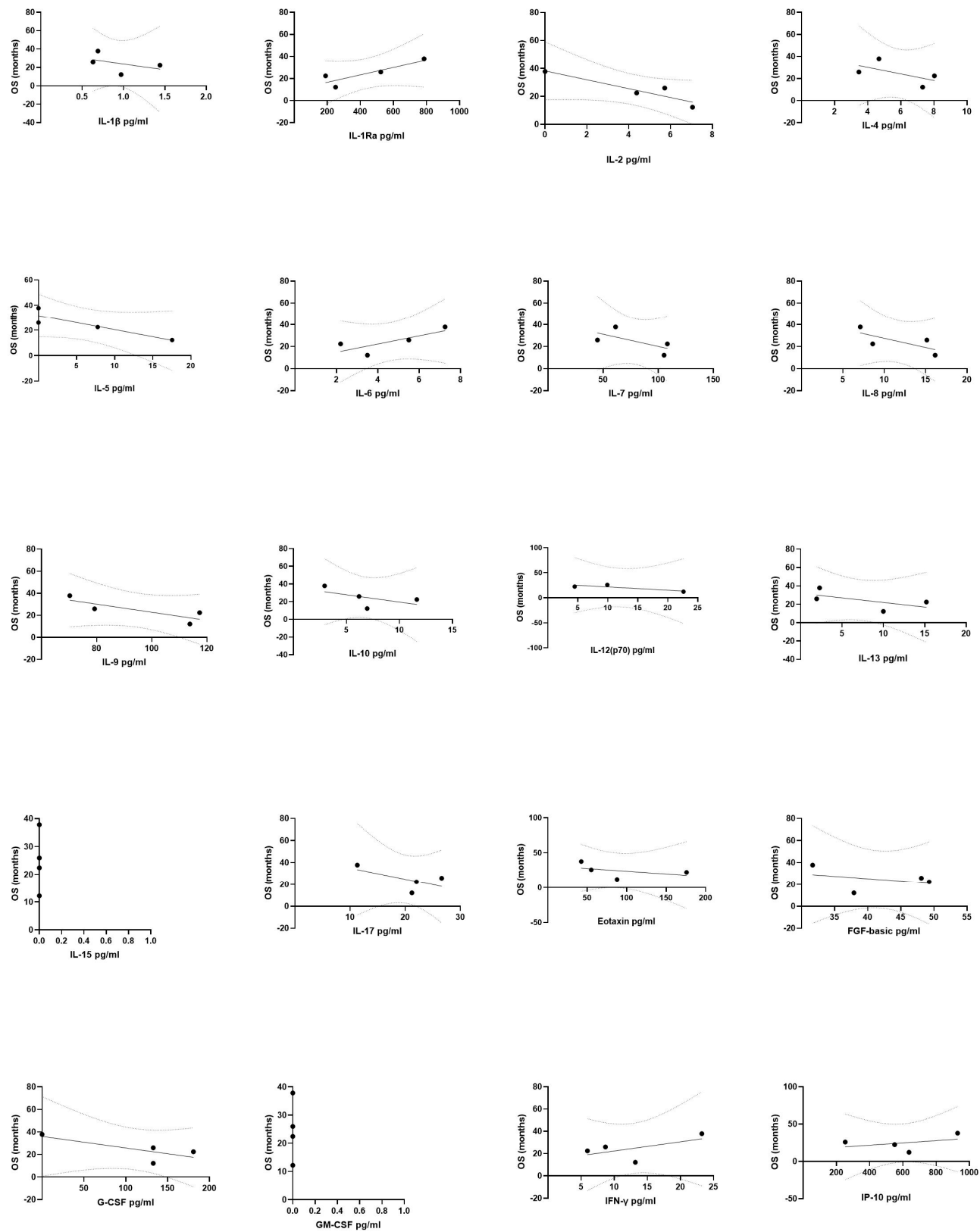
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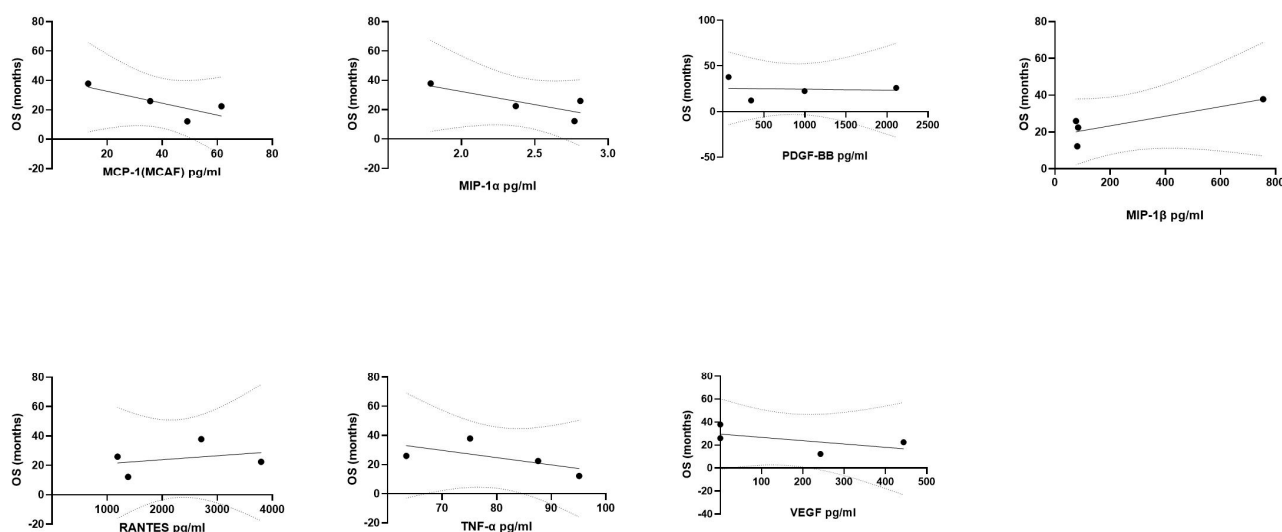
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K



L



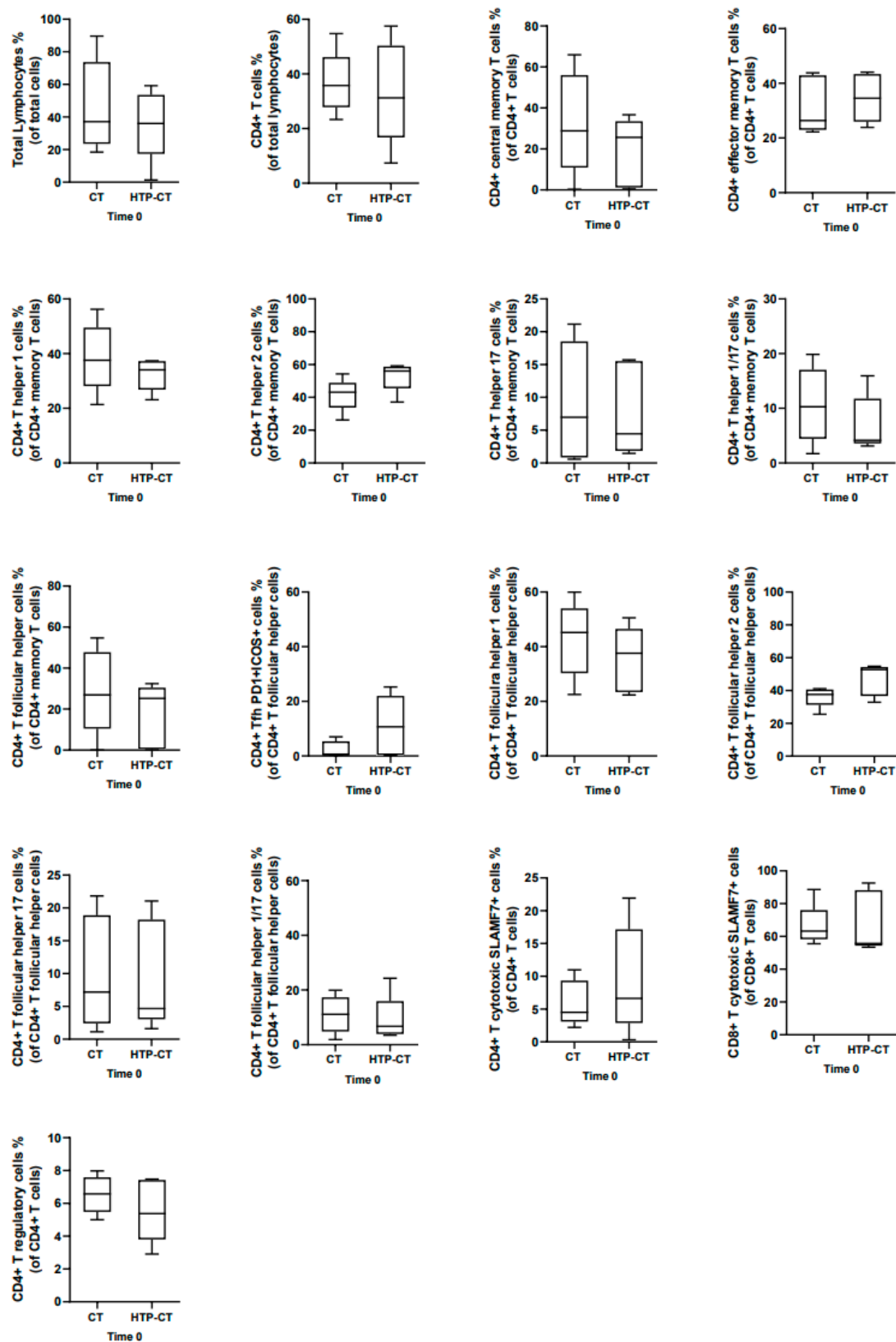


**Footnote:** Te: T effector; Th1: T helper 1; Th2: T helper 2; Th17: T helper 17; Th1/17: T helper 1/17; Tfh: T follicular helper; Tfh1: T follicular helper 1; Tfh2: T follicular helper 2; Tfh17: T follicular helper 17; Tfh1/17: T follicular helper 1/17; Treg: T regulatory; UM: unswitched memory; SM: switched memory; ASM: activated switched memory; DN: double negative; DN1: double negative 1; DN2: double negative 2; DN3: double negative 3; DN4: double negative 4; AcN: activated naïve; AnN: anergic naïve; RN: resting naïve; Ab: antibody secreting; T1/2: transitional 1/2; T3a: transitional 3a; MZP: marginal zone peripheral; Breg: B regulatory; AbS: antibody secreting; Imm: immature; Mon: monocytes.

**Figure S11.** Differences of circulating immune cells (A: T-cell compartment; B: B-cell compartment; C: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D: Pro Human Inflammation Panel I Assay; E: Pro TGF- $\beta$  Immunoassay; F: Pro Human Cytokine Immunoassay) between patients with pancreatic adenocarcinoma (PDAC) treated with HybridTherm Probe ablation plus chemotherapy (HTP-CT arm) and those treated with chemotherapy only (CT arm) at baseline. Inter-group variables were compared using the Welch two sample t-test: \* =  $p < 0.05$ .

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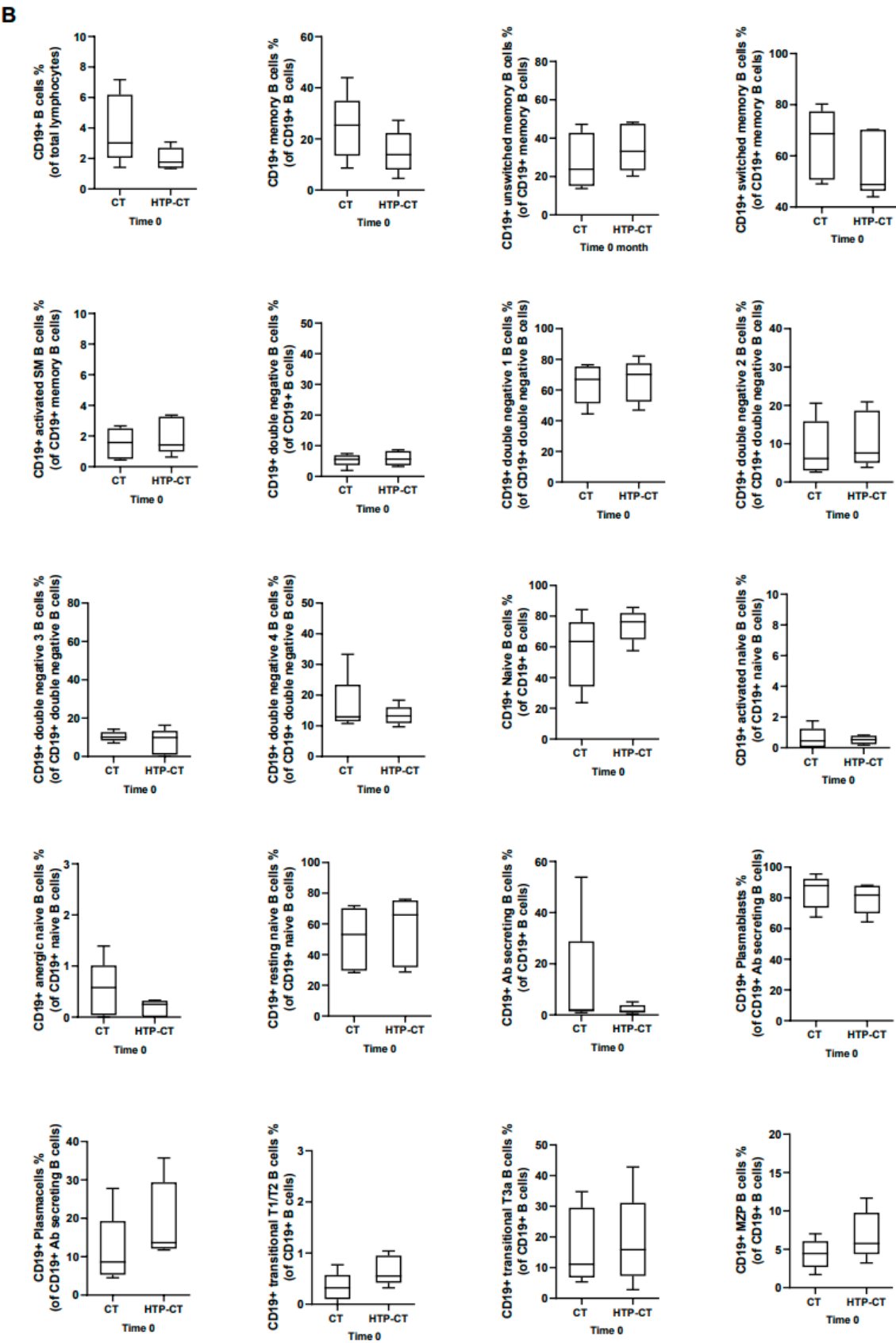
**A**

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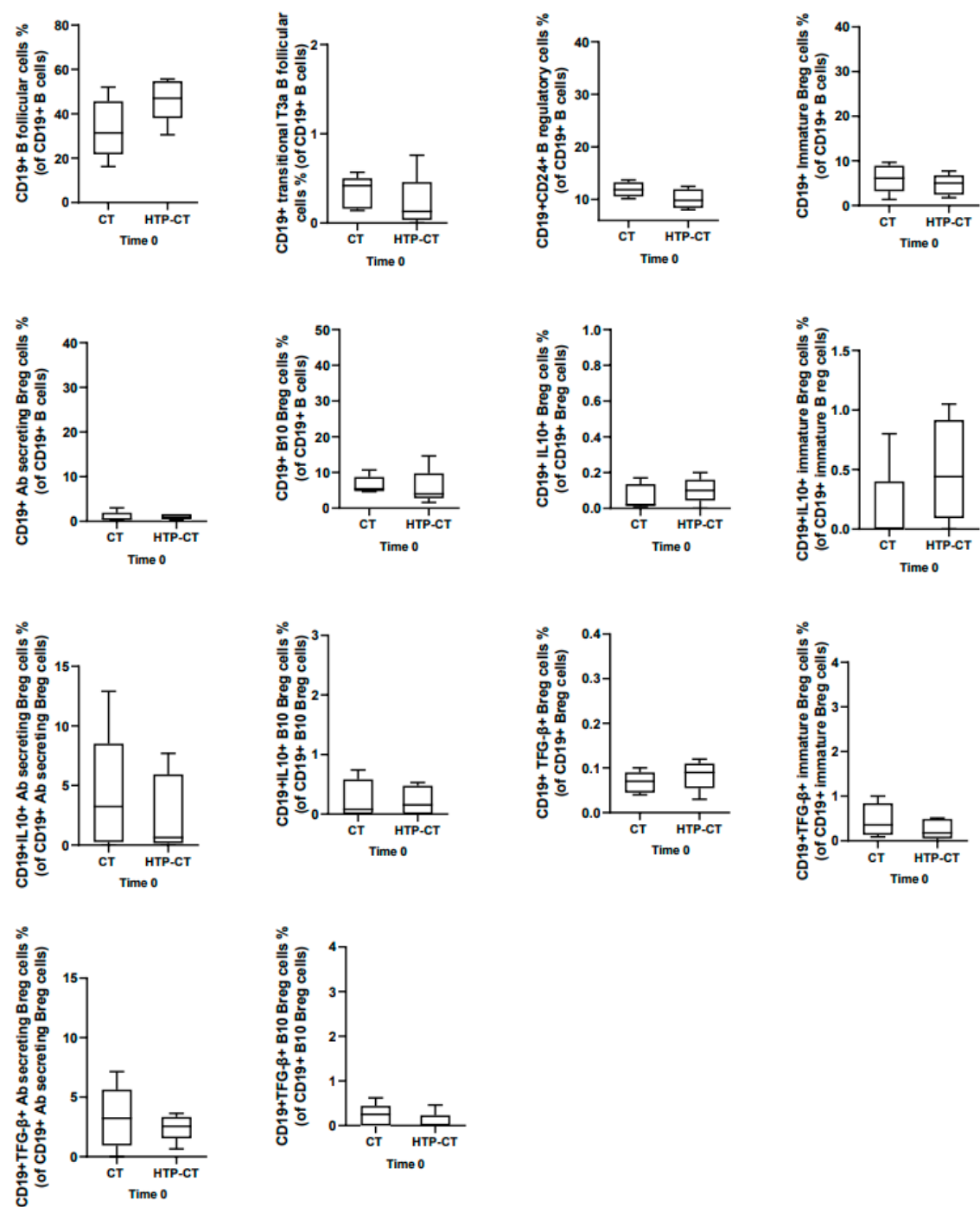
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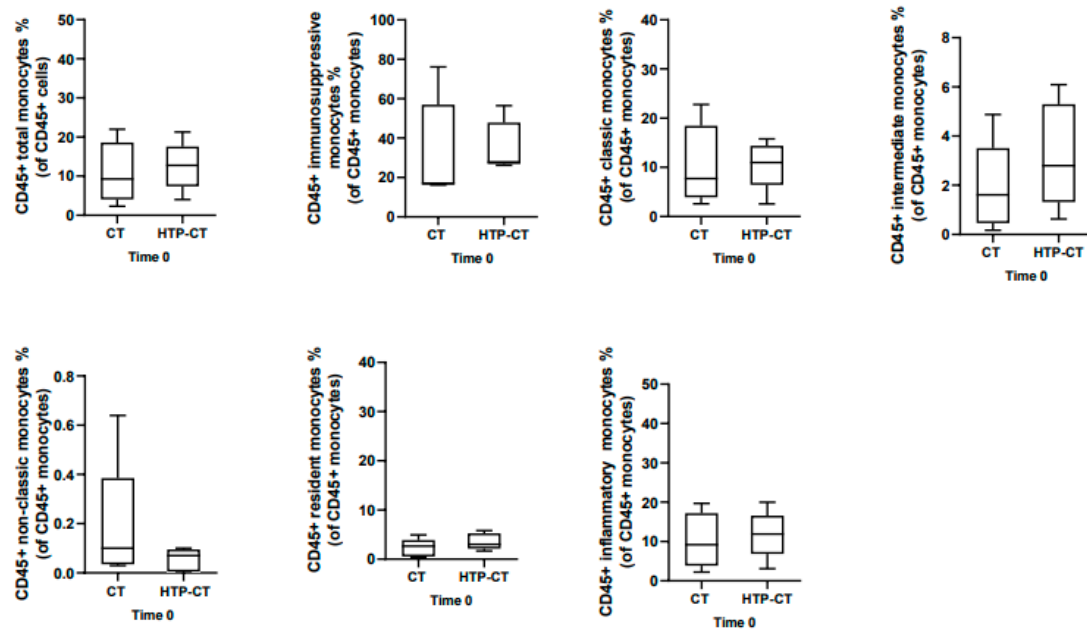
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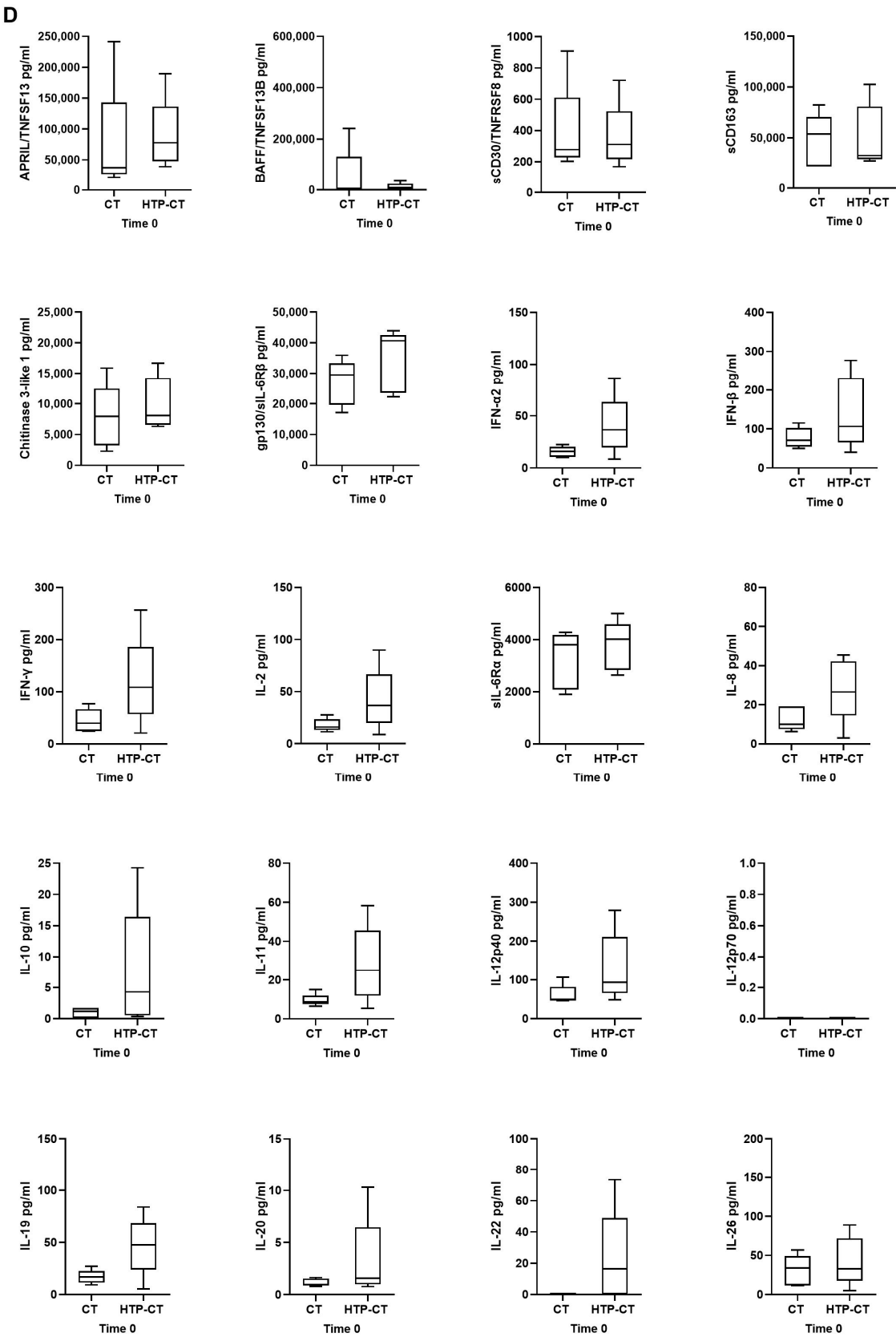
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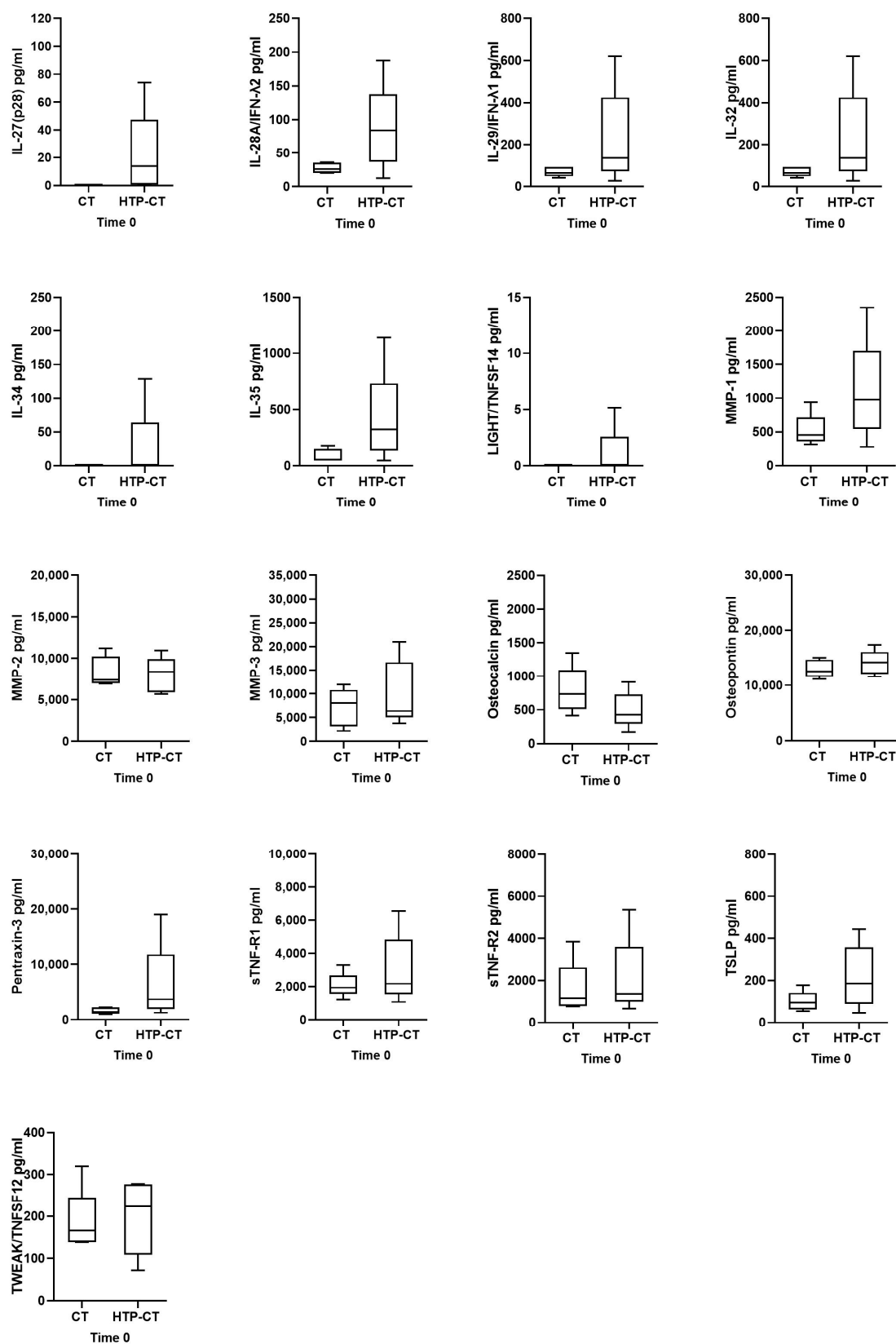
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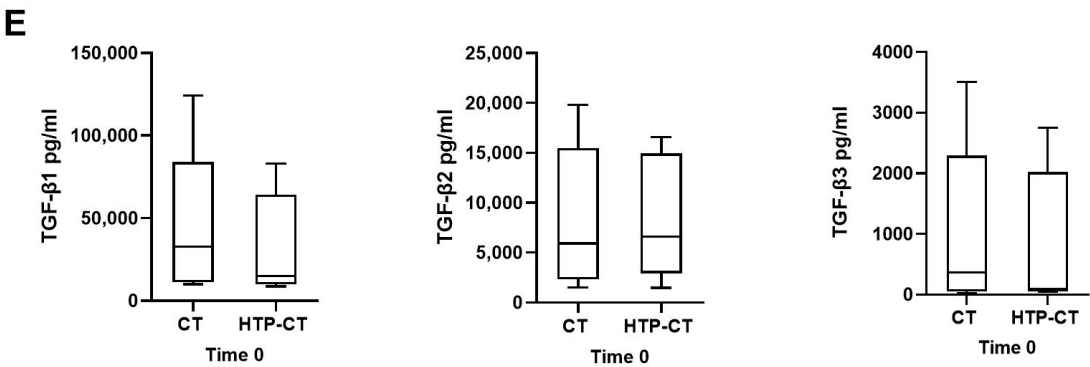
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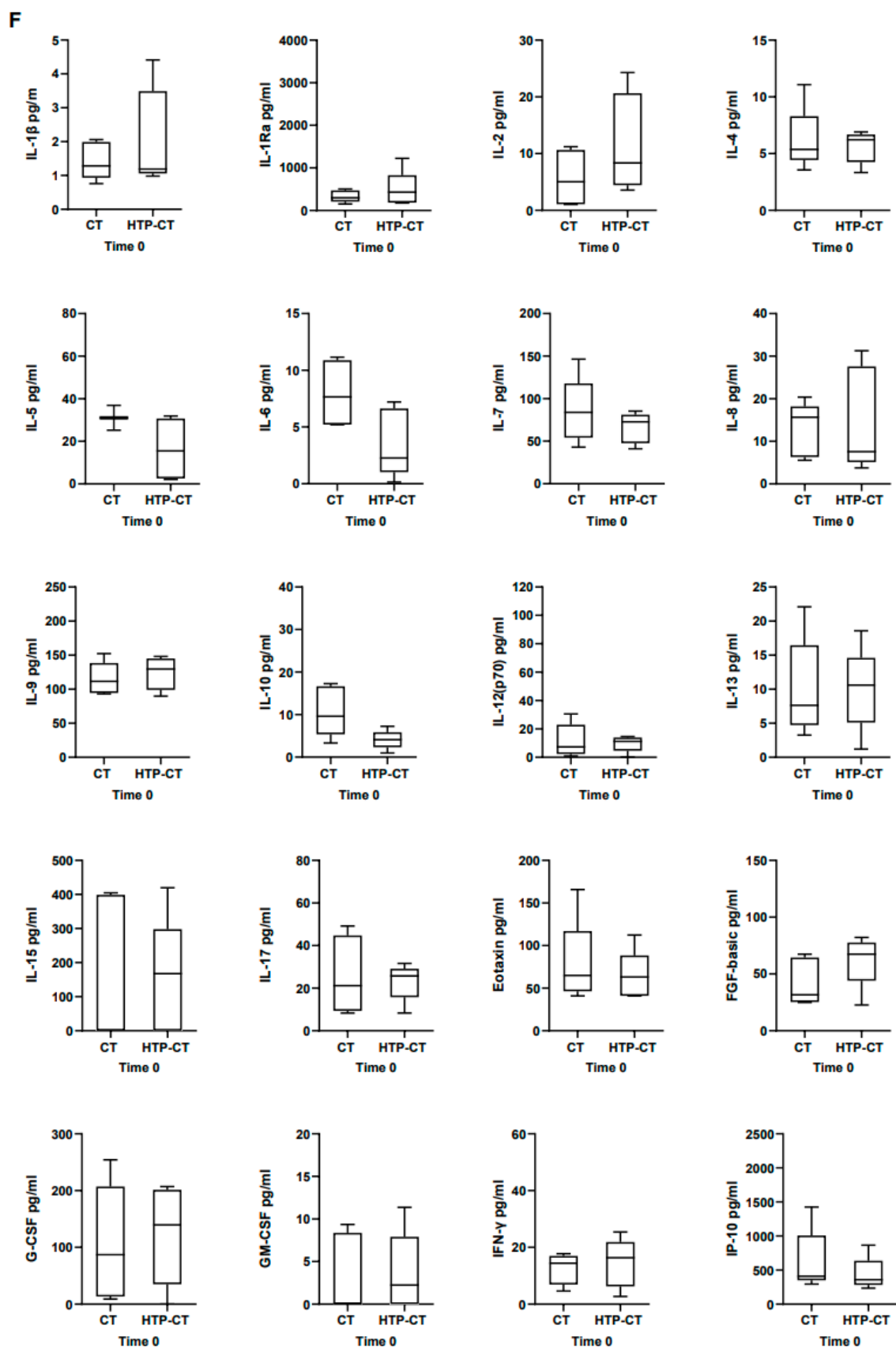
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**C**

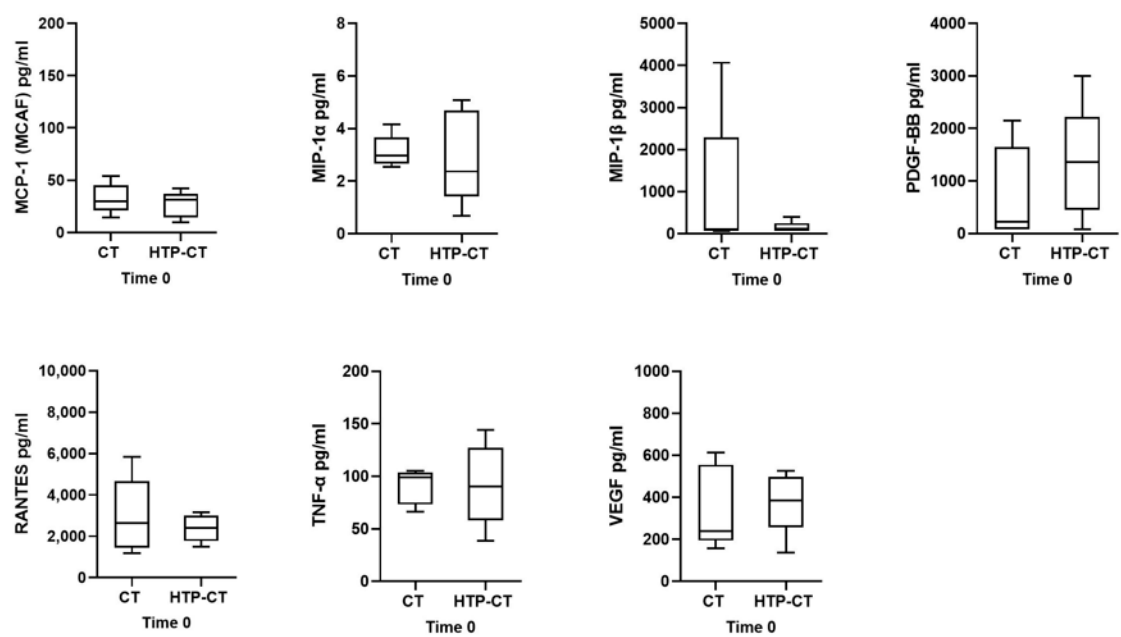






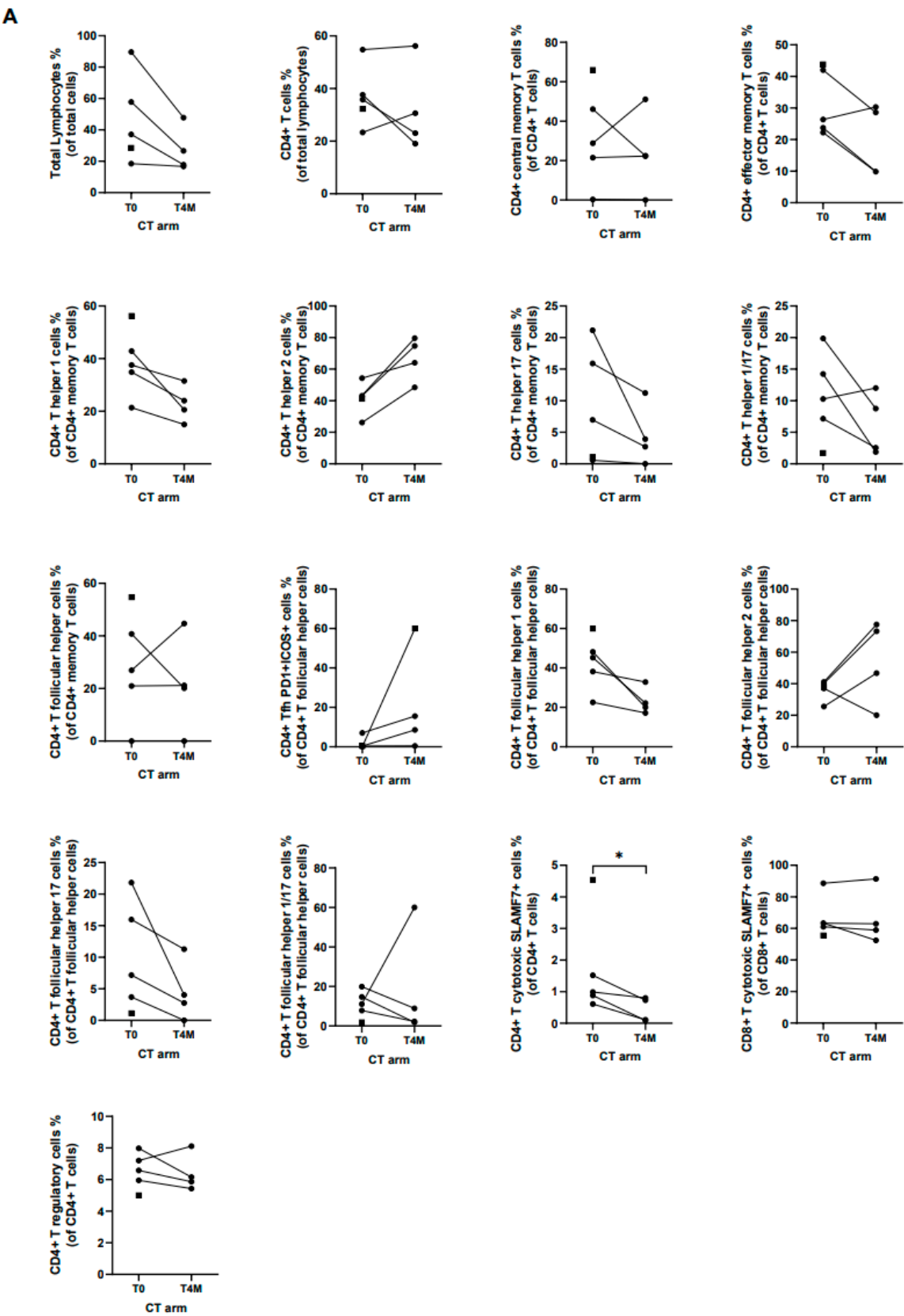






**Footnote:** Tfh: T follicular helper; SM: switched memory; Ab: antibody secreting; T1/2: type 1/2; T3a: type 3a; MZP: marginal zone peripheral; Breg: B regulatory.

**Figure S12.** Changes of circulating immune cells (A: T-cell compartment; B: B-cell compartment; C: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D: Pro Human Inflammation Panel I Assay; E: Pro TGF- $\beta$  Immunoassay; F: Pro Human Cytokine Immunoassay) between baseline (T0) and 4 months (T4M) after therapy onset in patients with pancreatic adenocarcinoma (PDAC) treated with chemotherapy only (CT arm). Square dot represents the patient in the CT arm who did not reach 4-month follow-up. Intra-group variables were compared using the paired t-test: \* =  $p < 0.05$ .

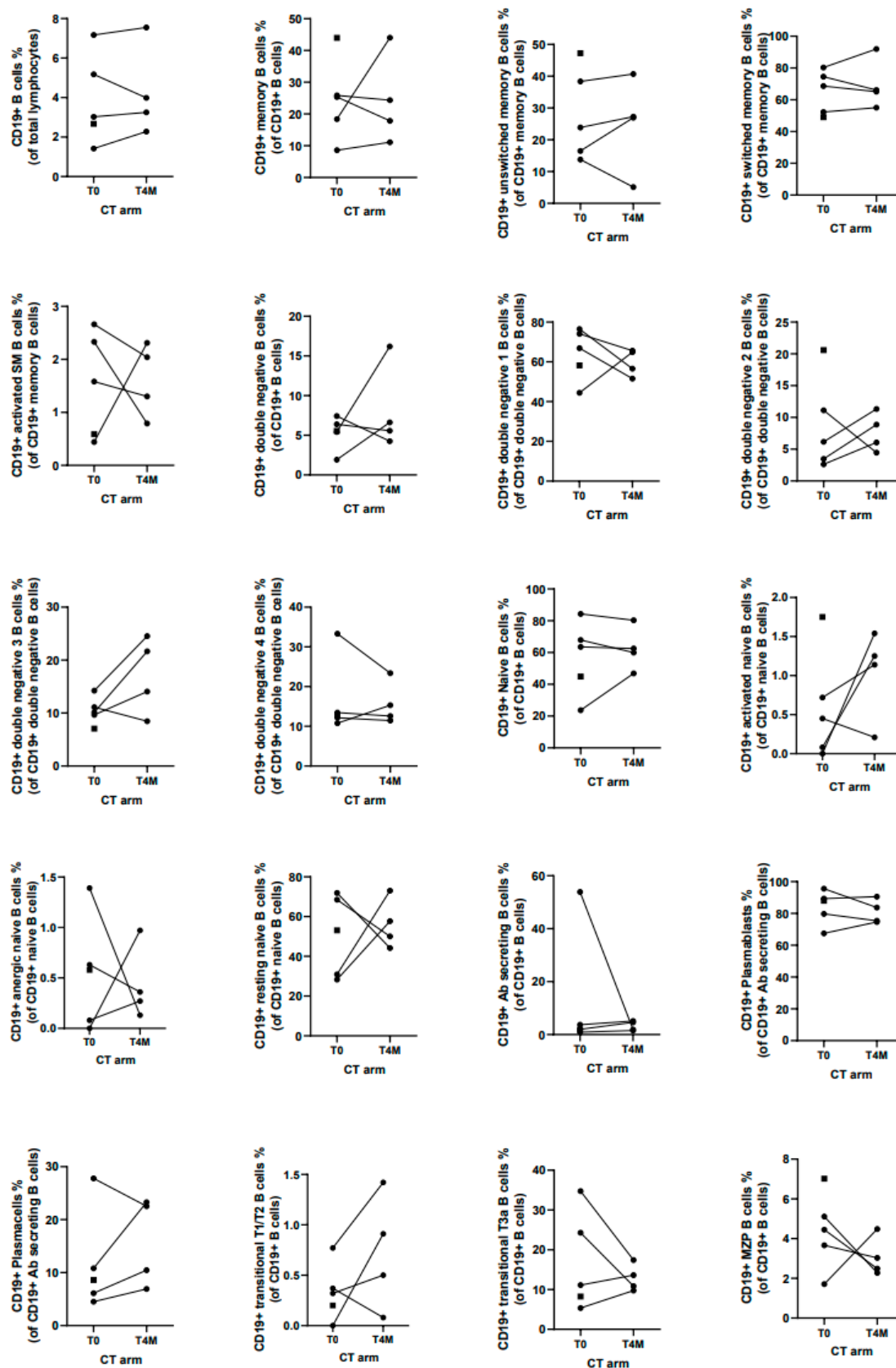


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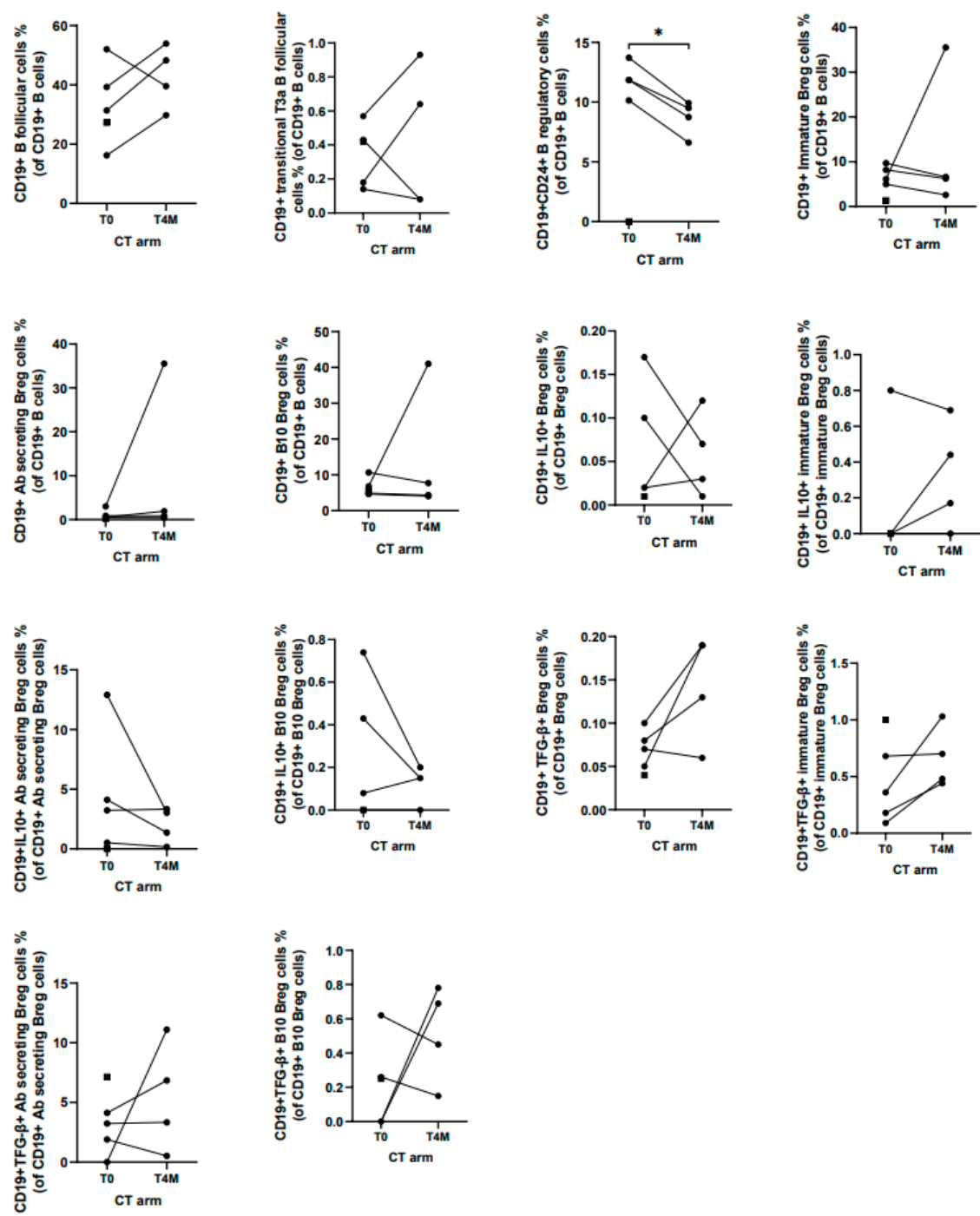
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**B**



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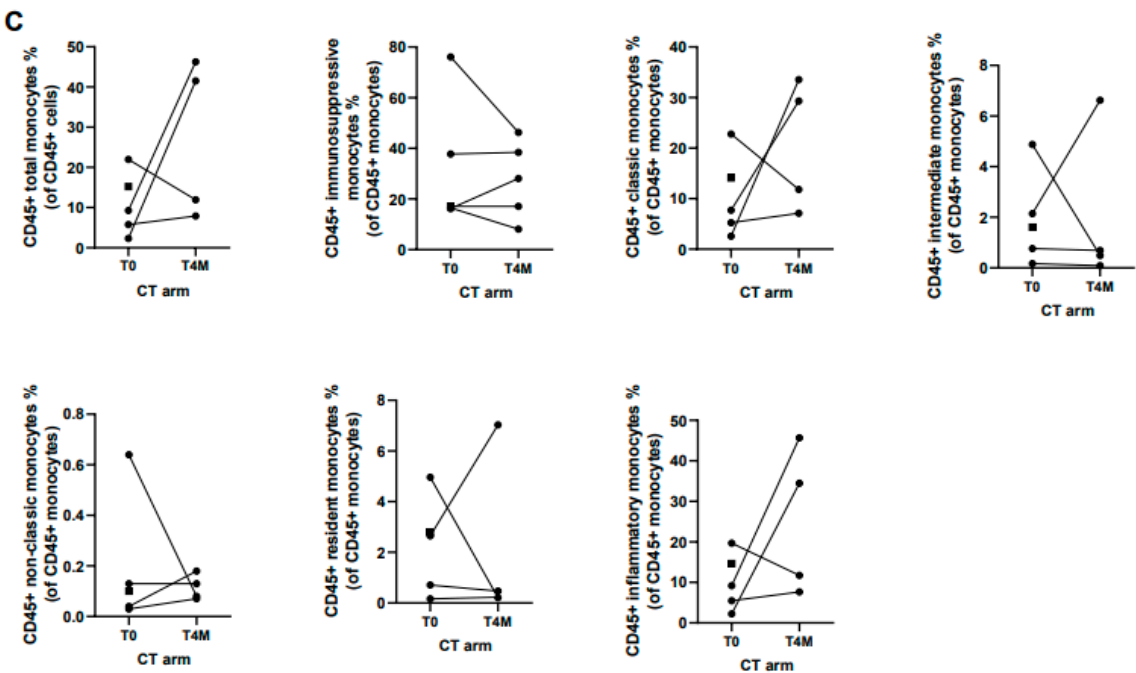
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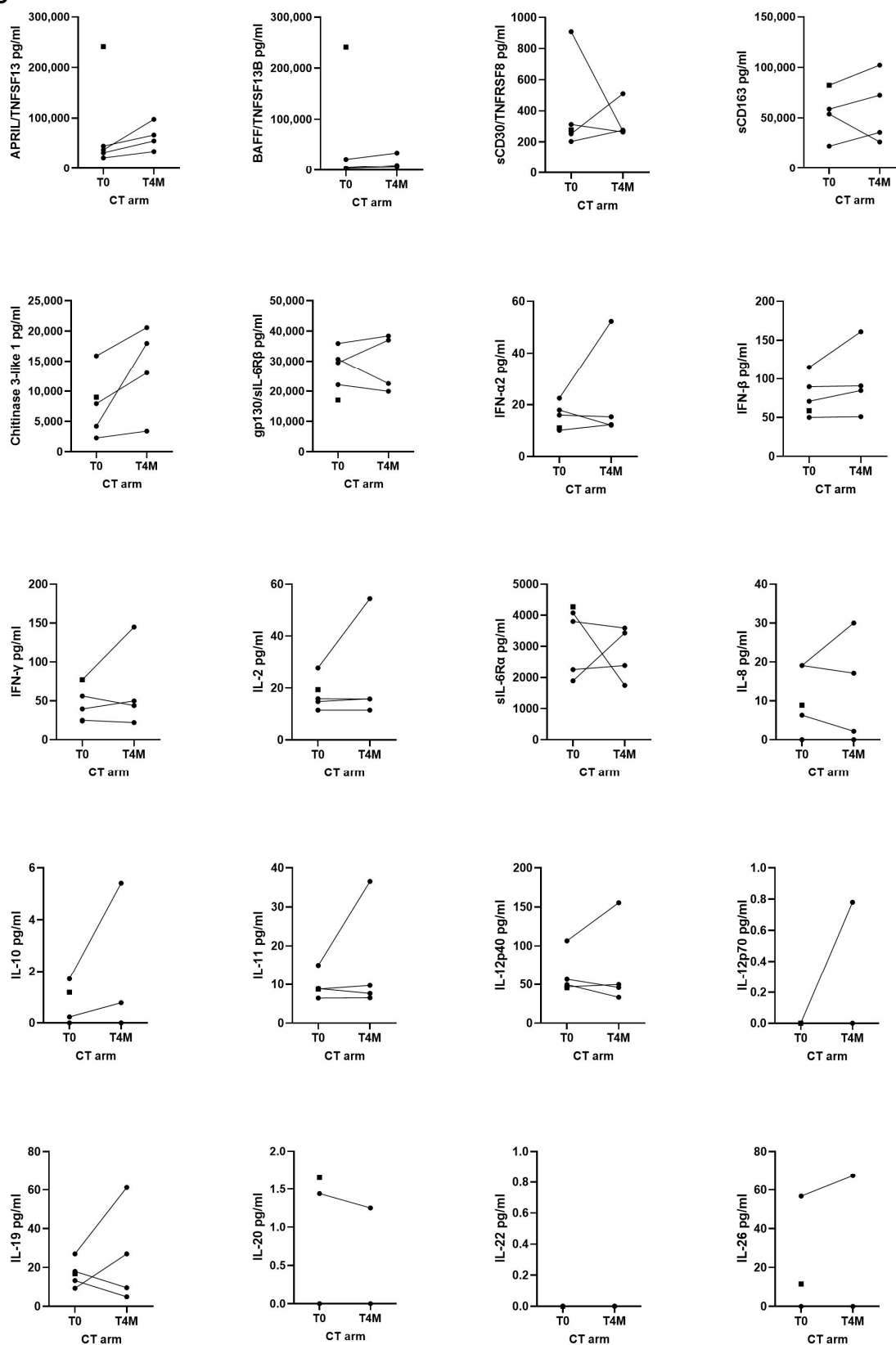


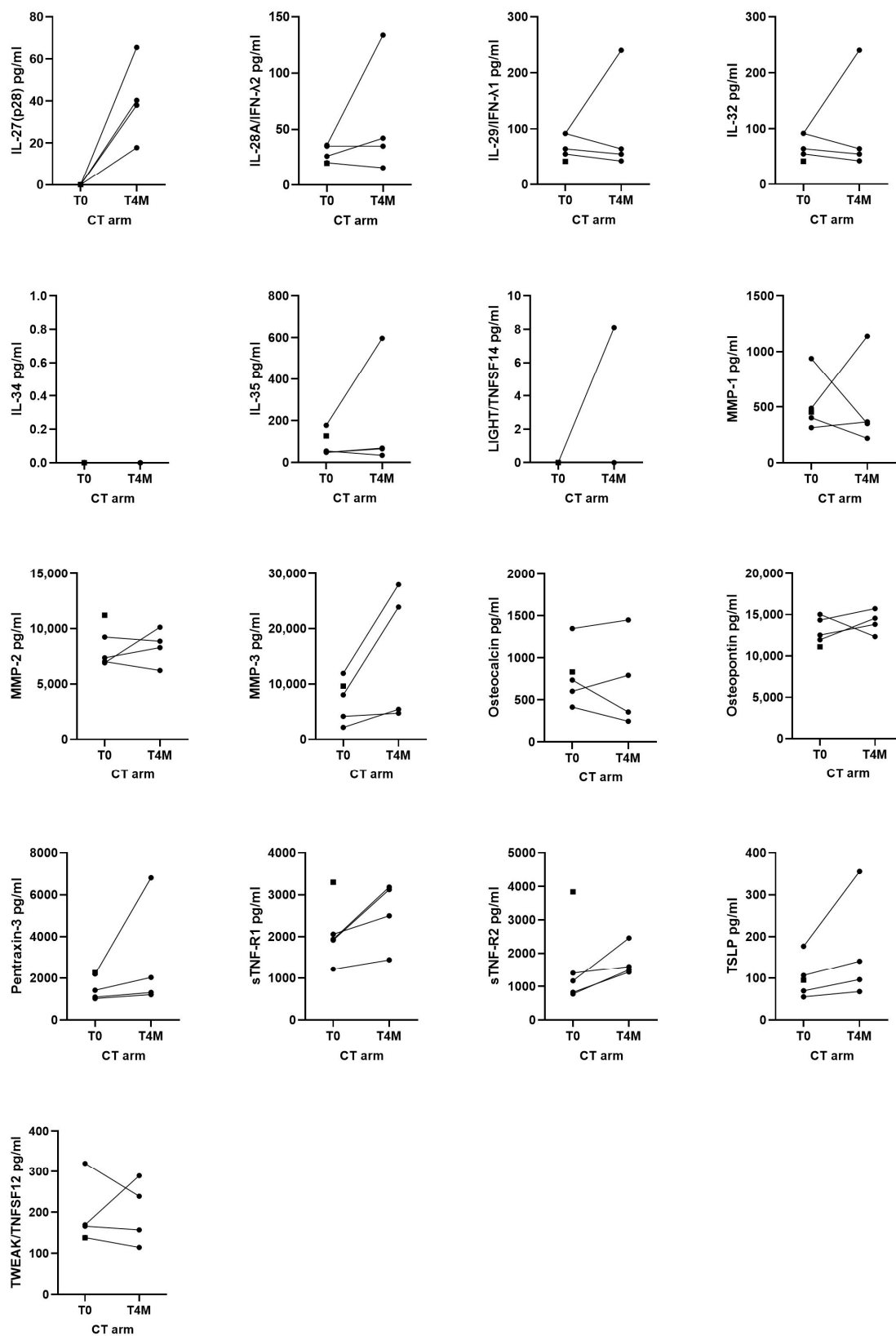
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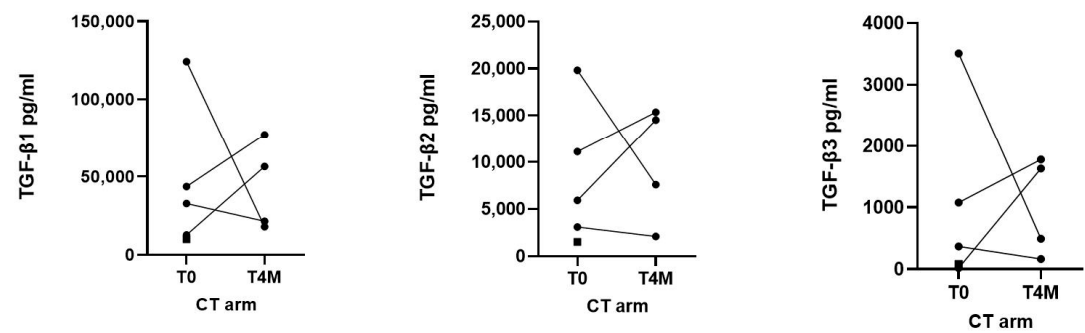


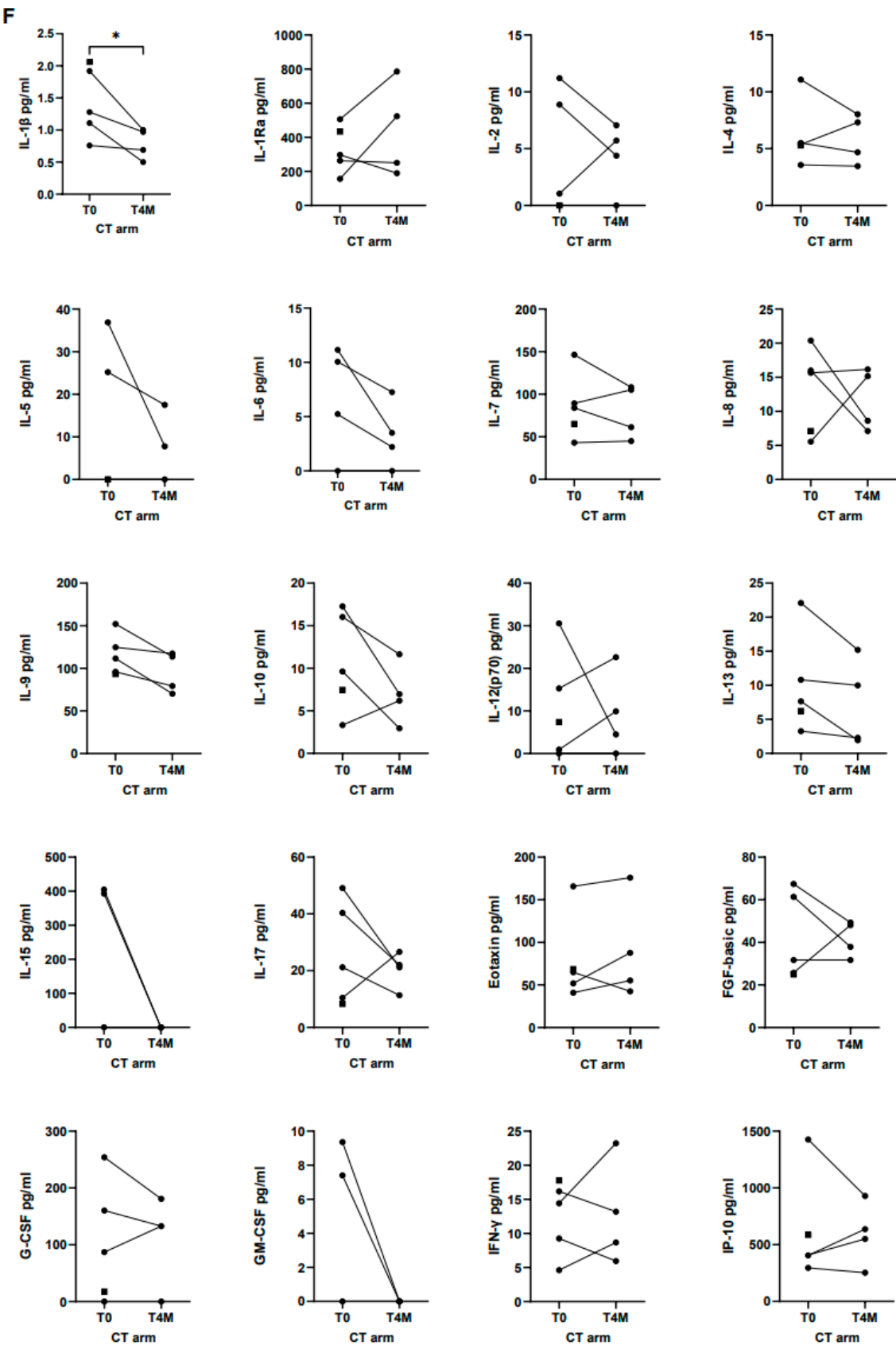
**D**

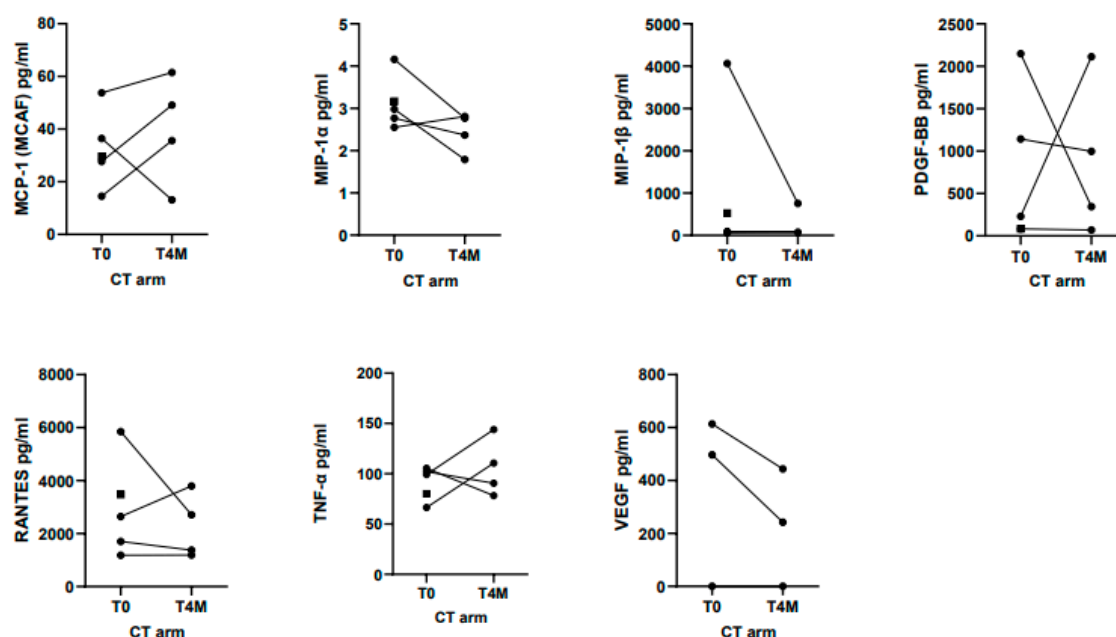




E



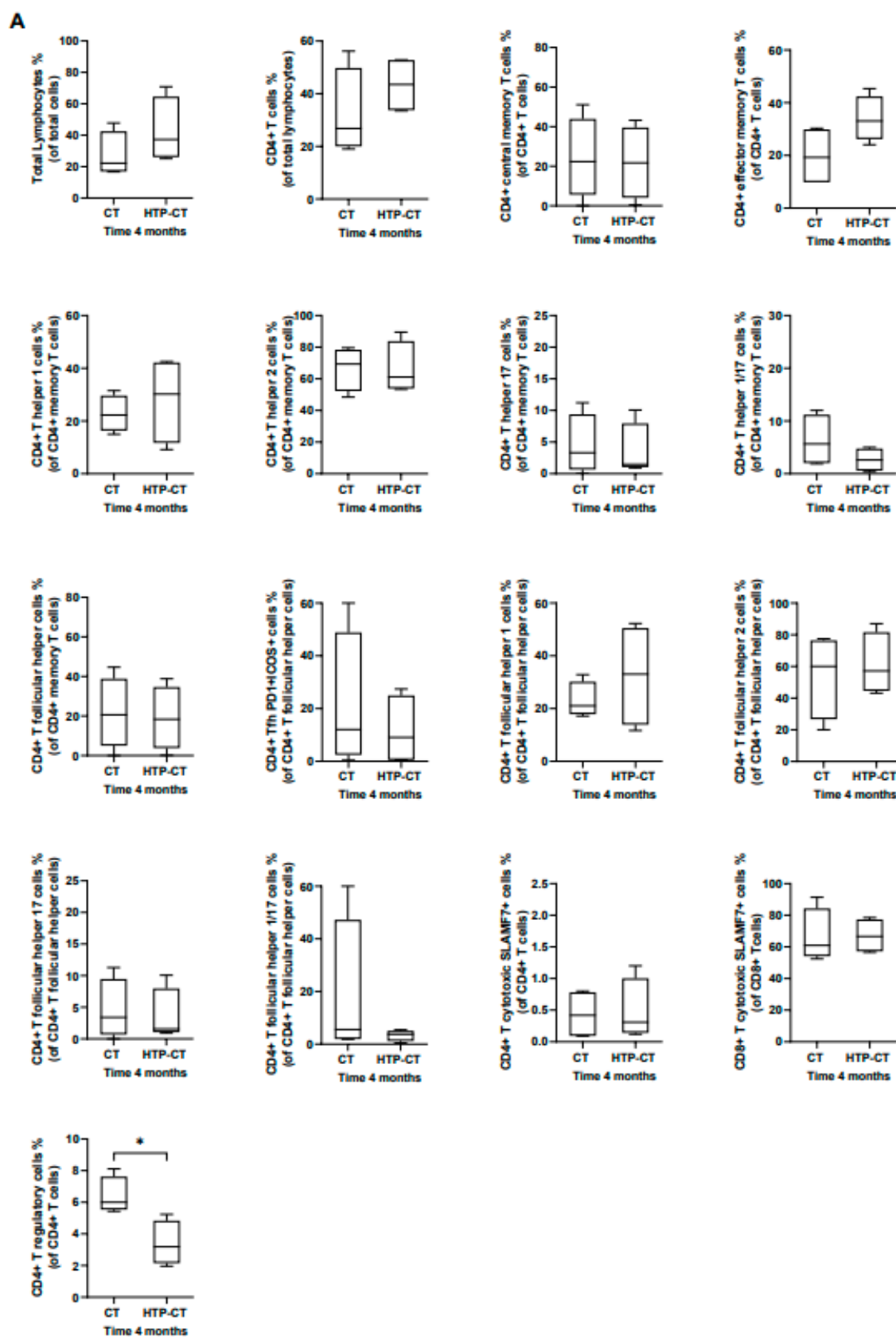




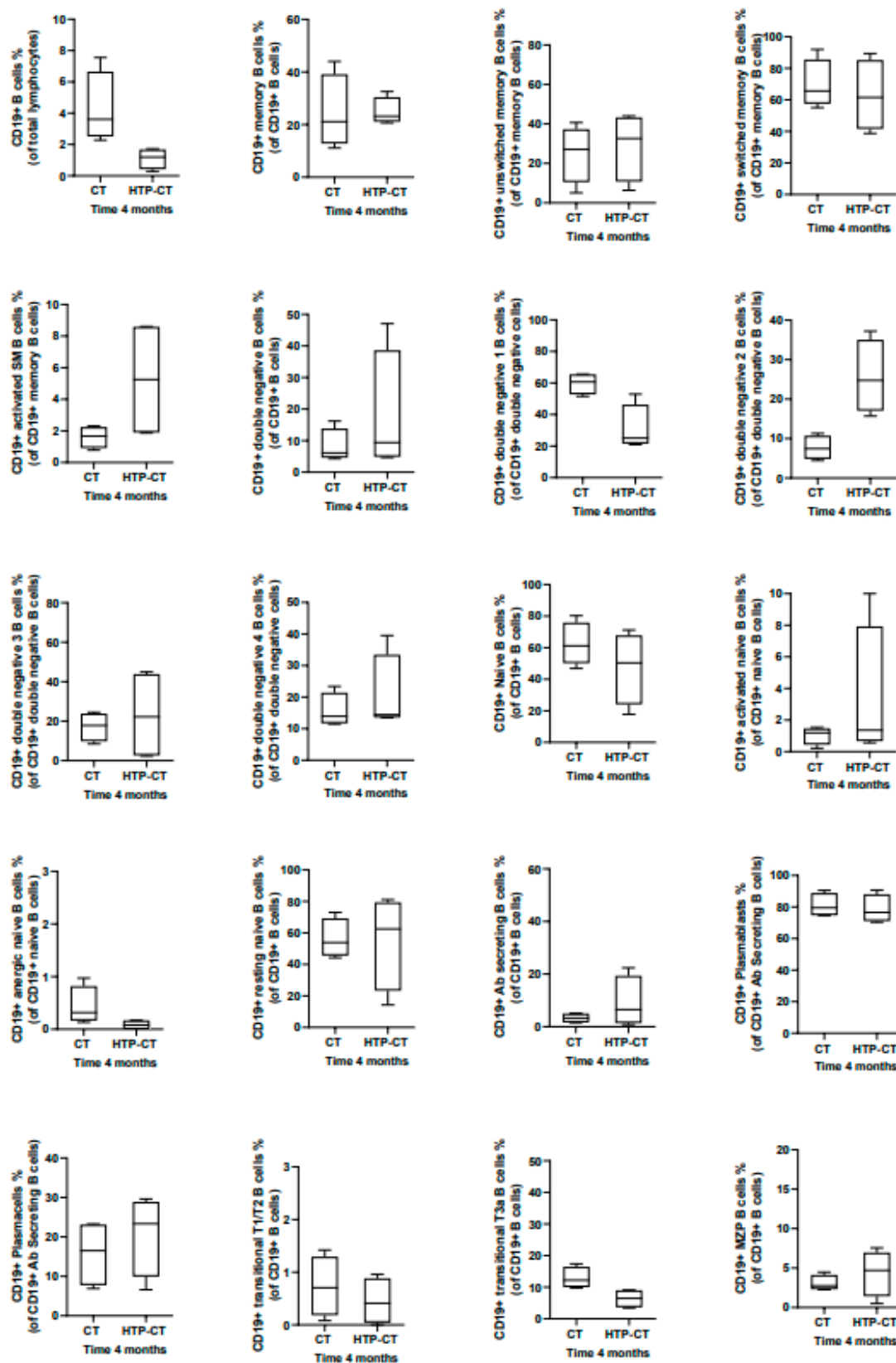
**Footnote:** Te: T effector; Th1: T helper 1; Th2: T helper 2; Th17: T helper 17; Th1/17: T helper 1/17; Tfh: T follicular helper; Tfh1: T follicular helper 1; Tfh2: T follicular helper 2; Tfh17: T follicular helper 17; Tfh1/17: T follicular helper 1/17; Treg: T regulatory; UM: unswitched memory; SM: switched memory; ASM: activated switched memory; DN: double negative; DN1: double negative 1; DN2: double negative 2; DN3: double negative 3; DN4: double negative 4; AcN: activated naïve; AnN: anergic naïve; RN: resting naïve; Ab: antibody secreting; T1/2: transitional 1/2; T3a: transitional 3a; MZP: marginal zone peripheral; Breg: B regulatory; AbS: antibody secreting; Imm: immature; Mon: monocytes.

**Figure S13.** Differences of circulating immune cells (A: T-cell compartment; B: B-cell compartment; C: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D: Pro Human Inflammation Panel I Assay; E: Pro TGF- $\beta$  Immunoassay; F: Pro Human Cytokine Immunoassay) between patients with pancreatic adenocarcinoma (PDAC) treated with HybridTherm Probe ablation plus chemotherapy (HTP-CT arm) and those treated with chemotherapy only (CT arm) at 4-months follow-up after therapy onset. Inter-group variables were compared using the Welch two sample t-test: \* =  $p < 0.05$ .

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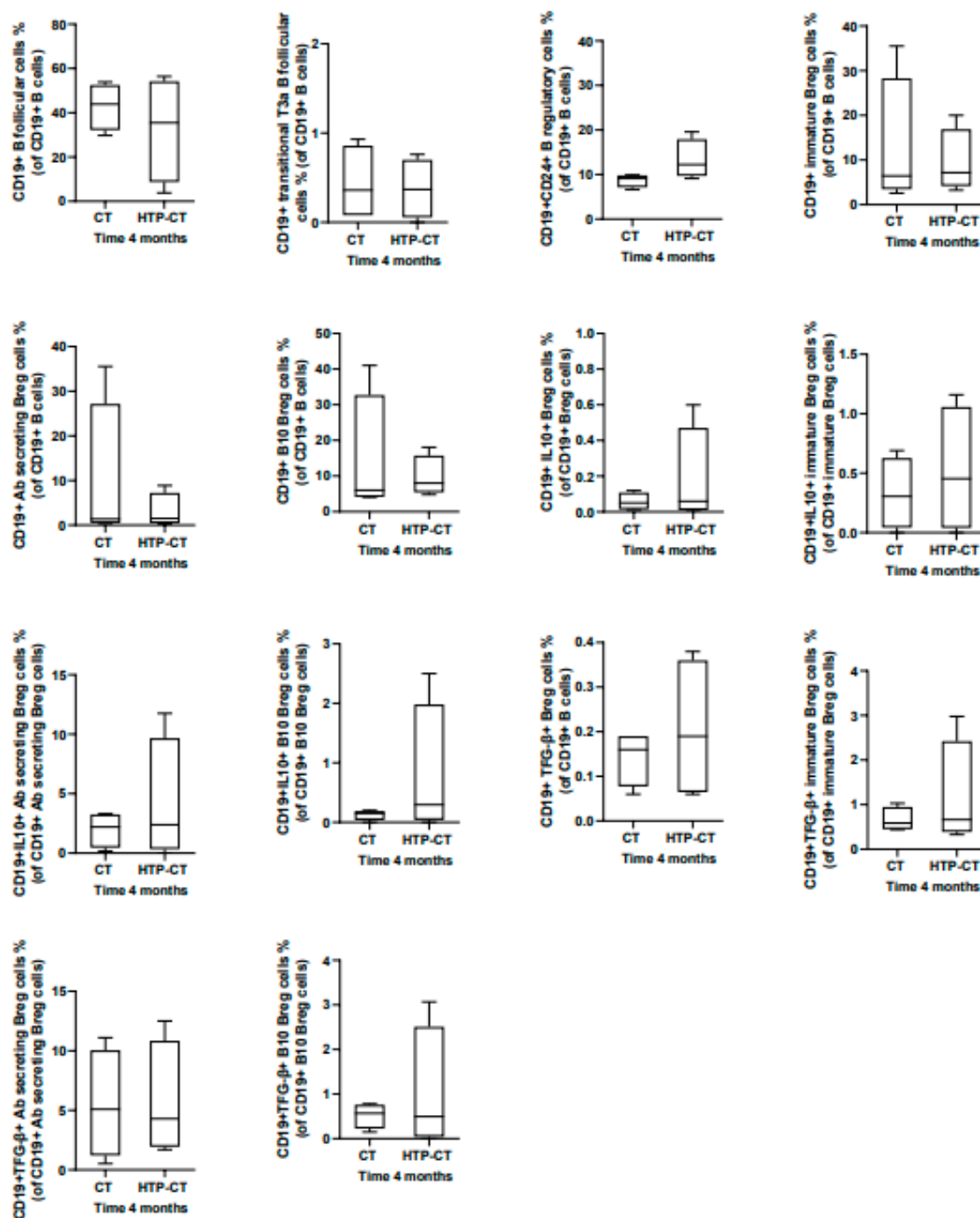
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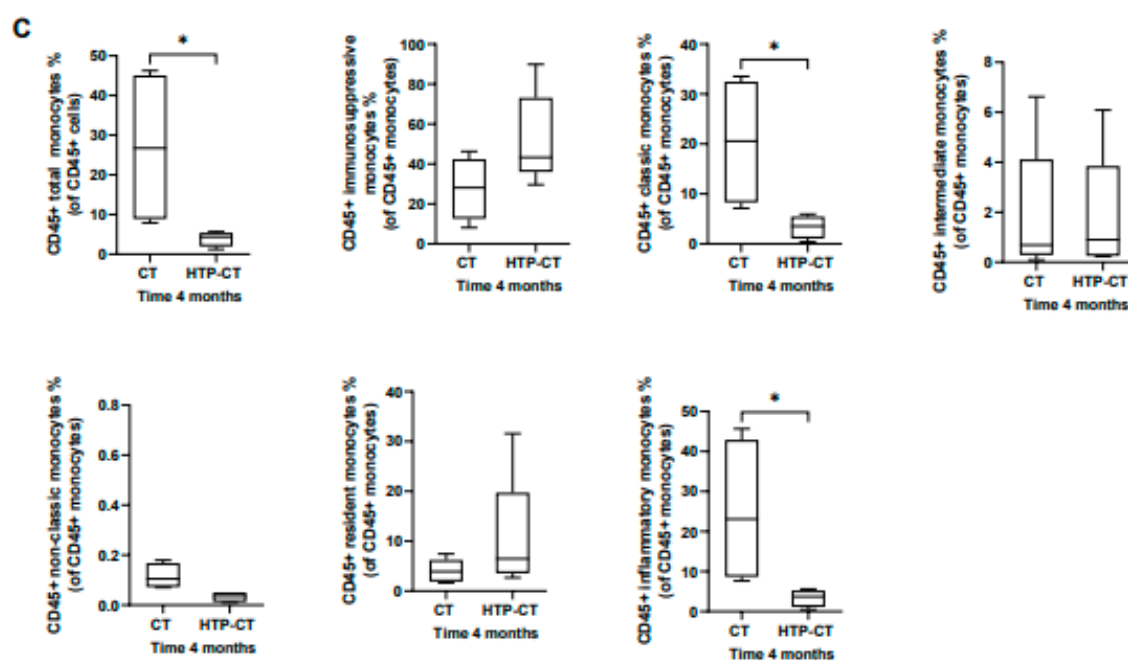


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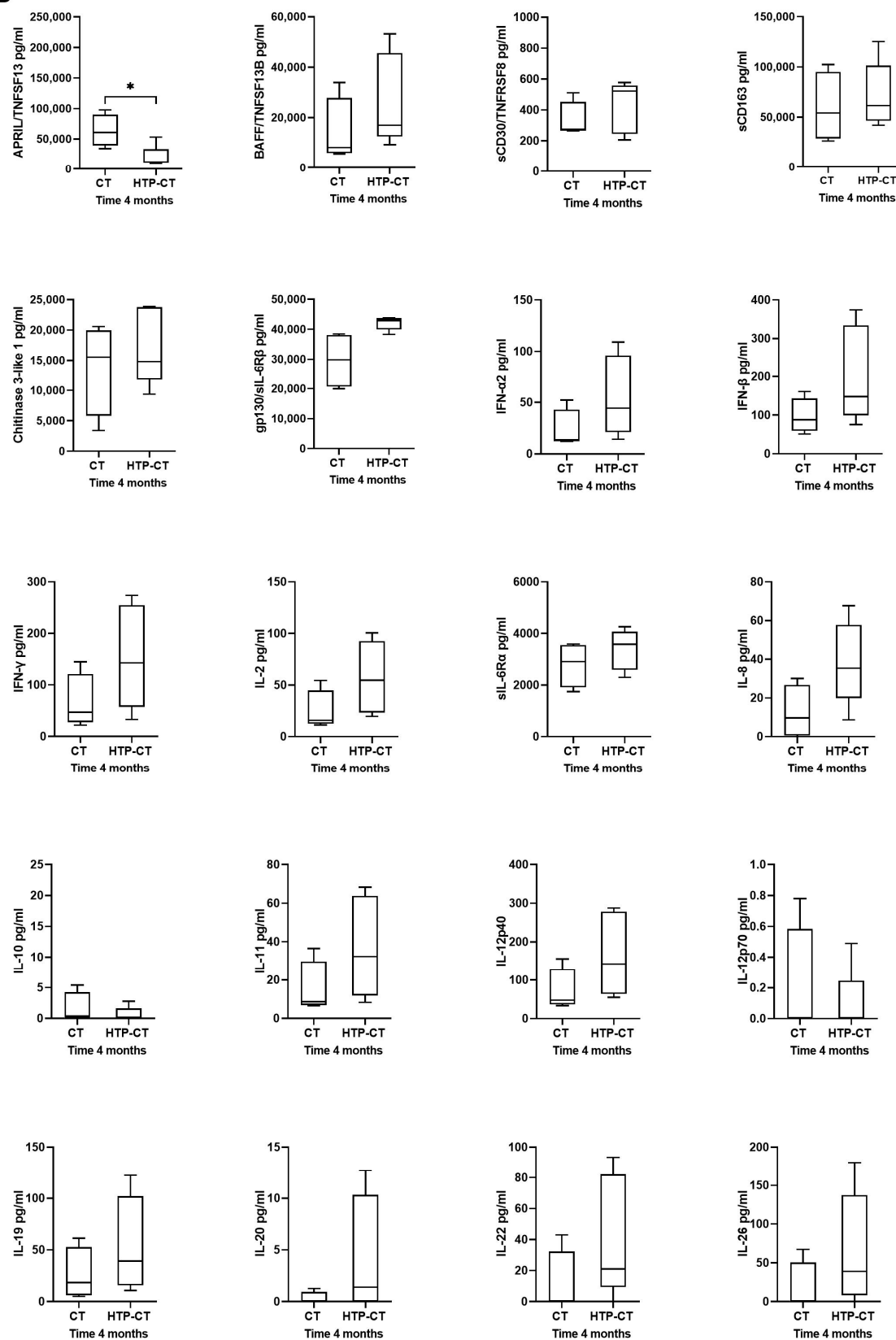
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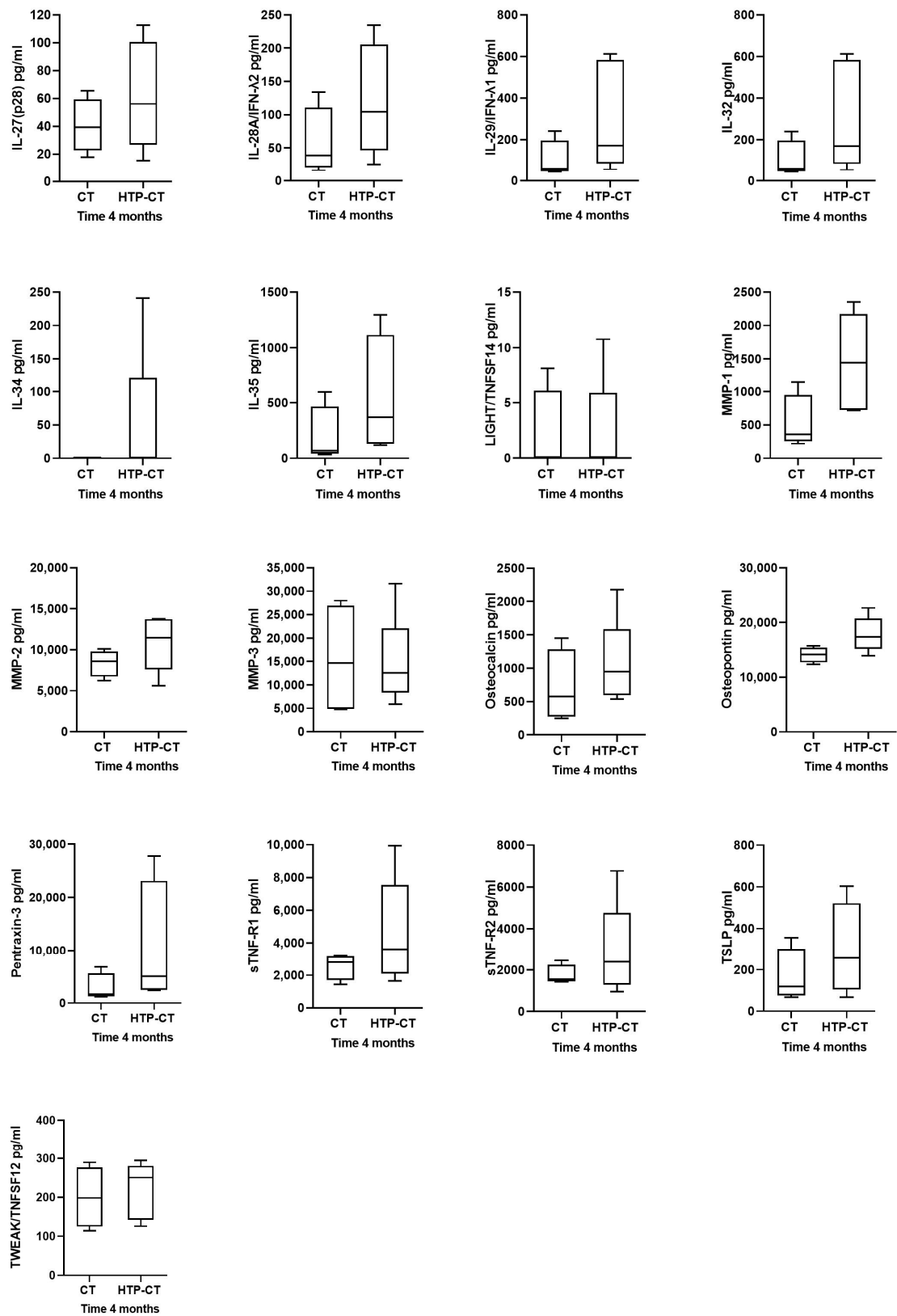
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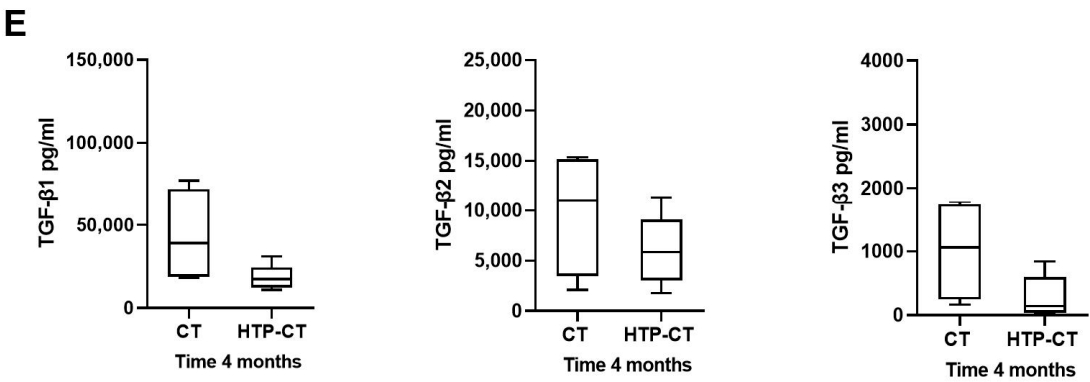


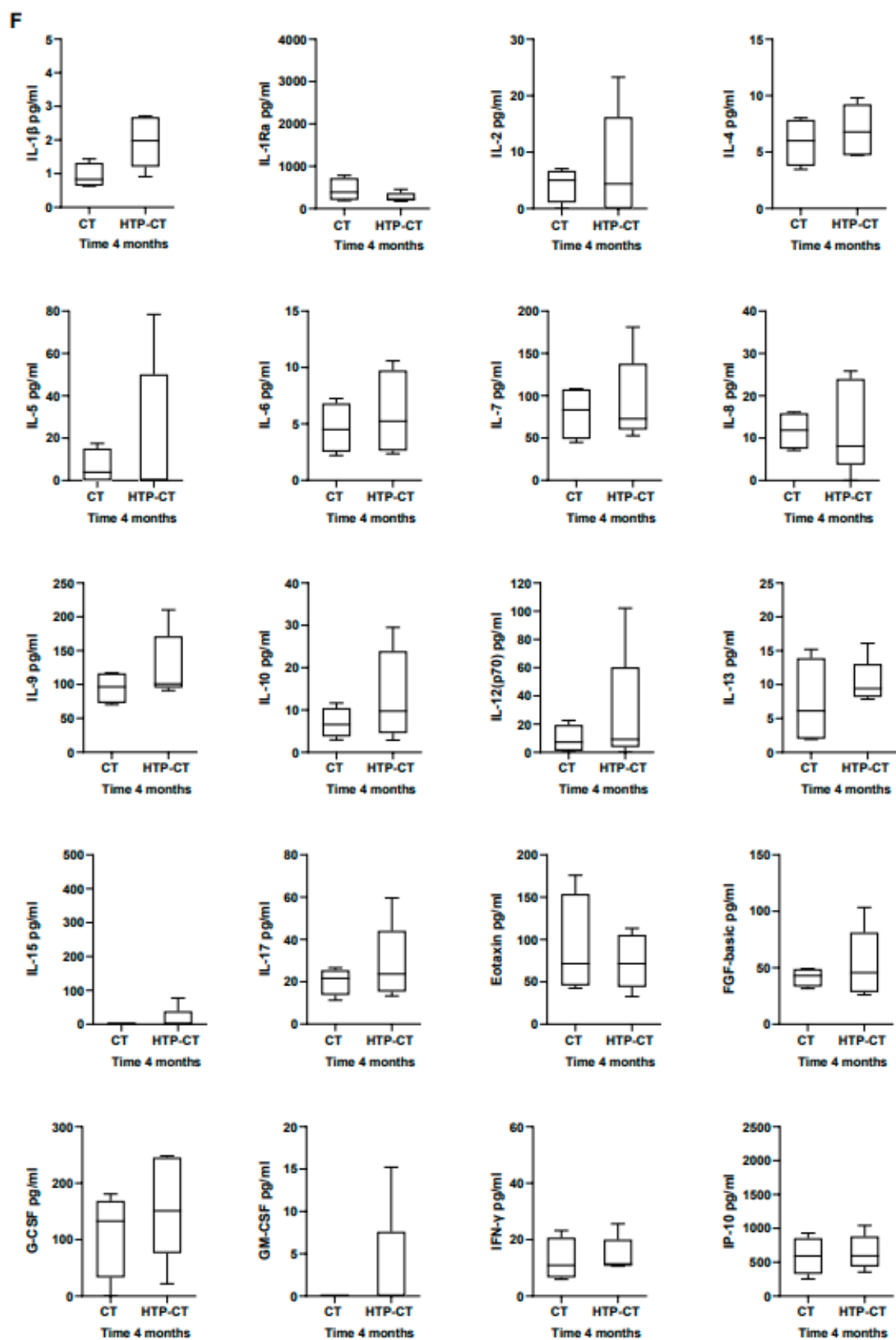


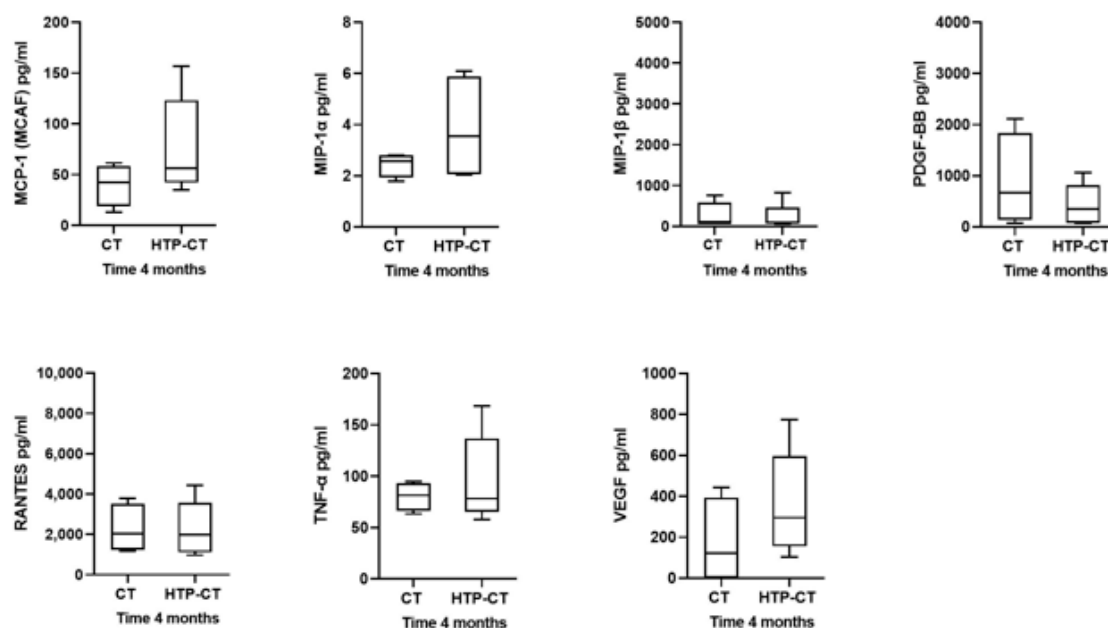


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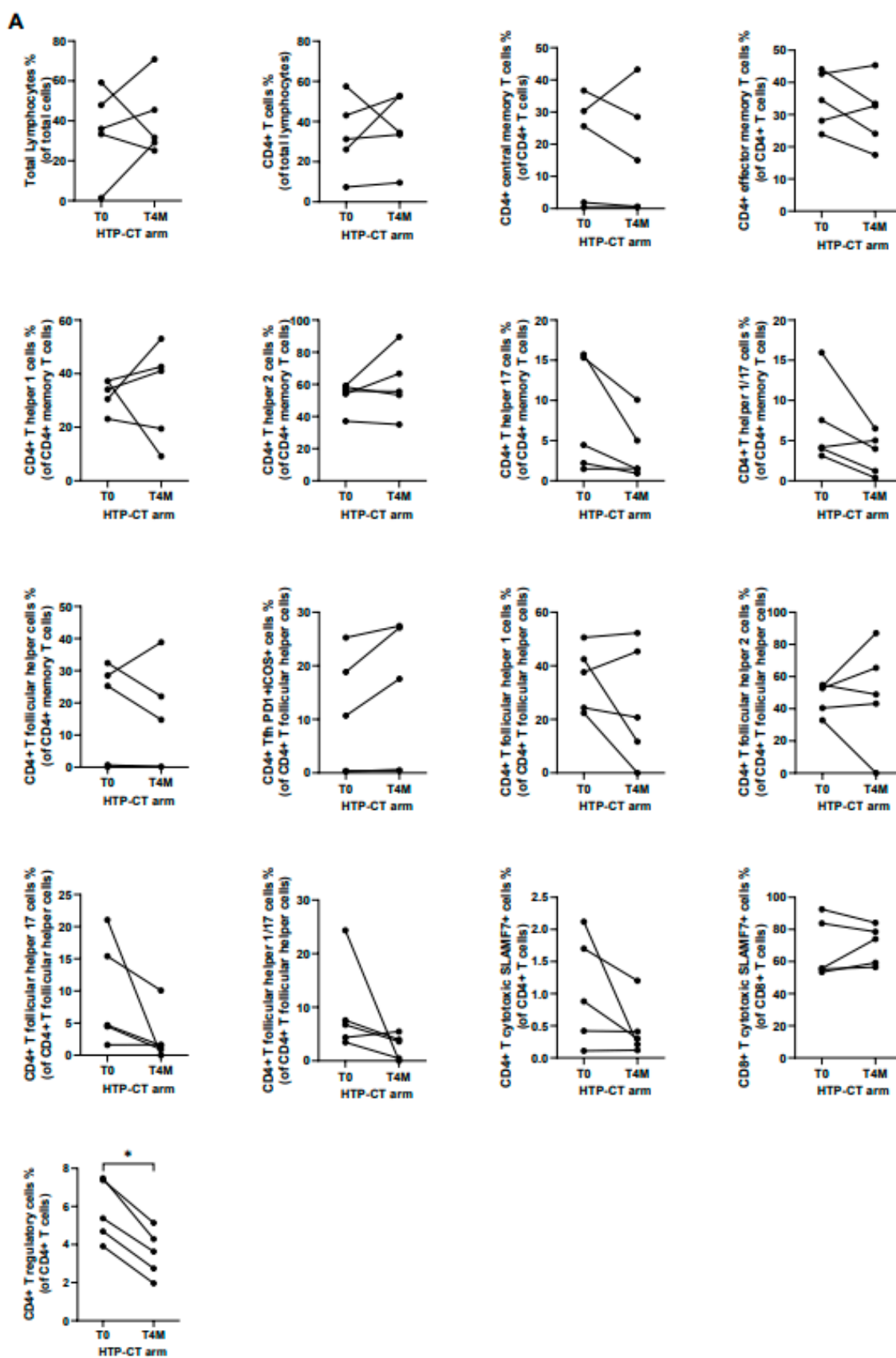


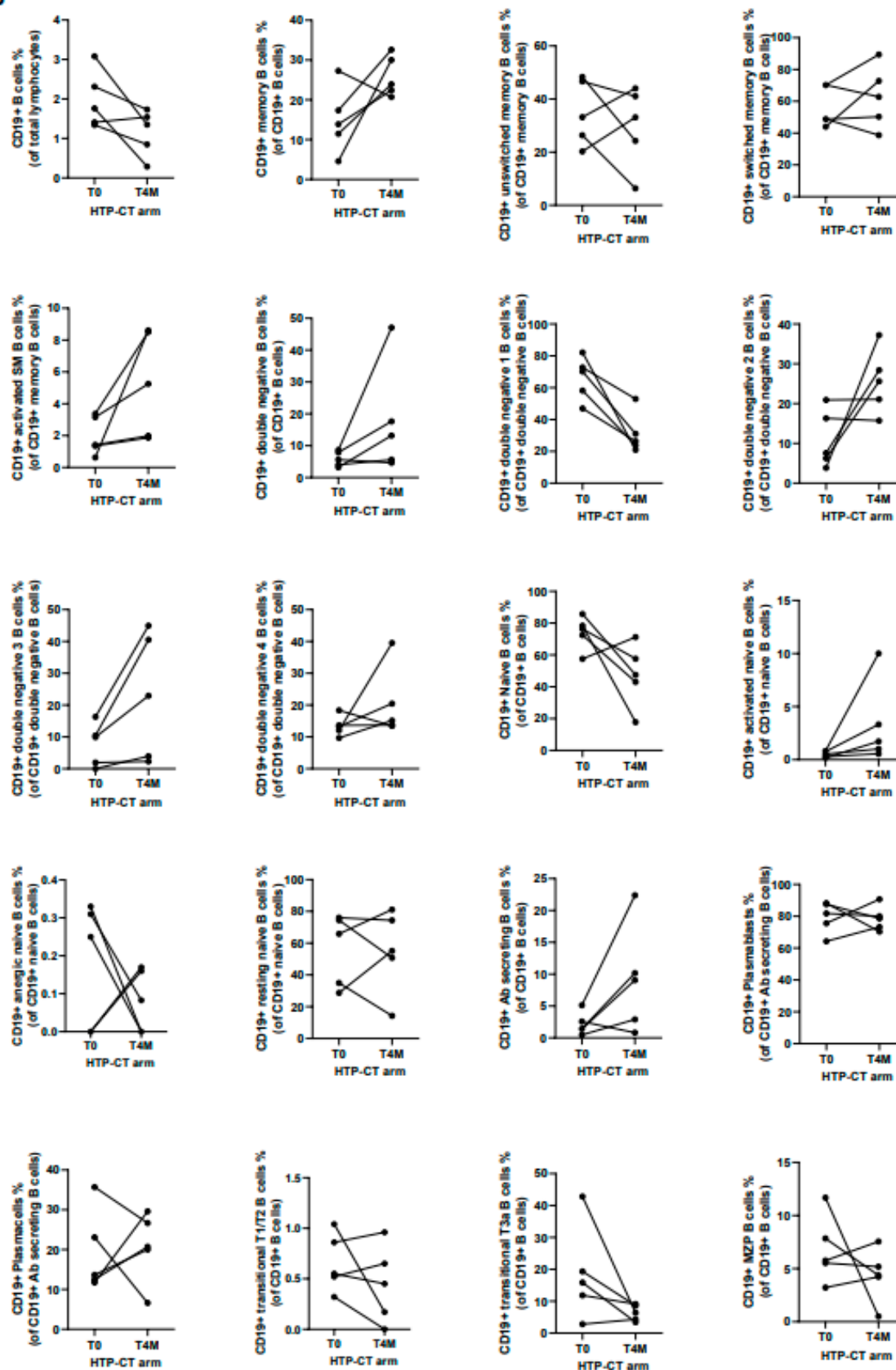




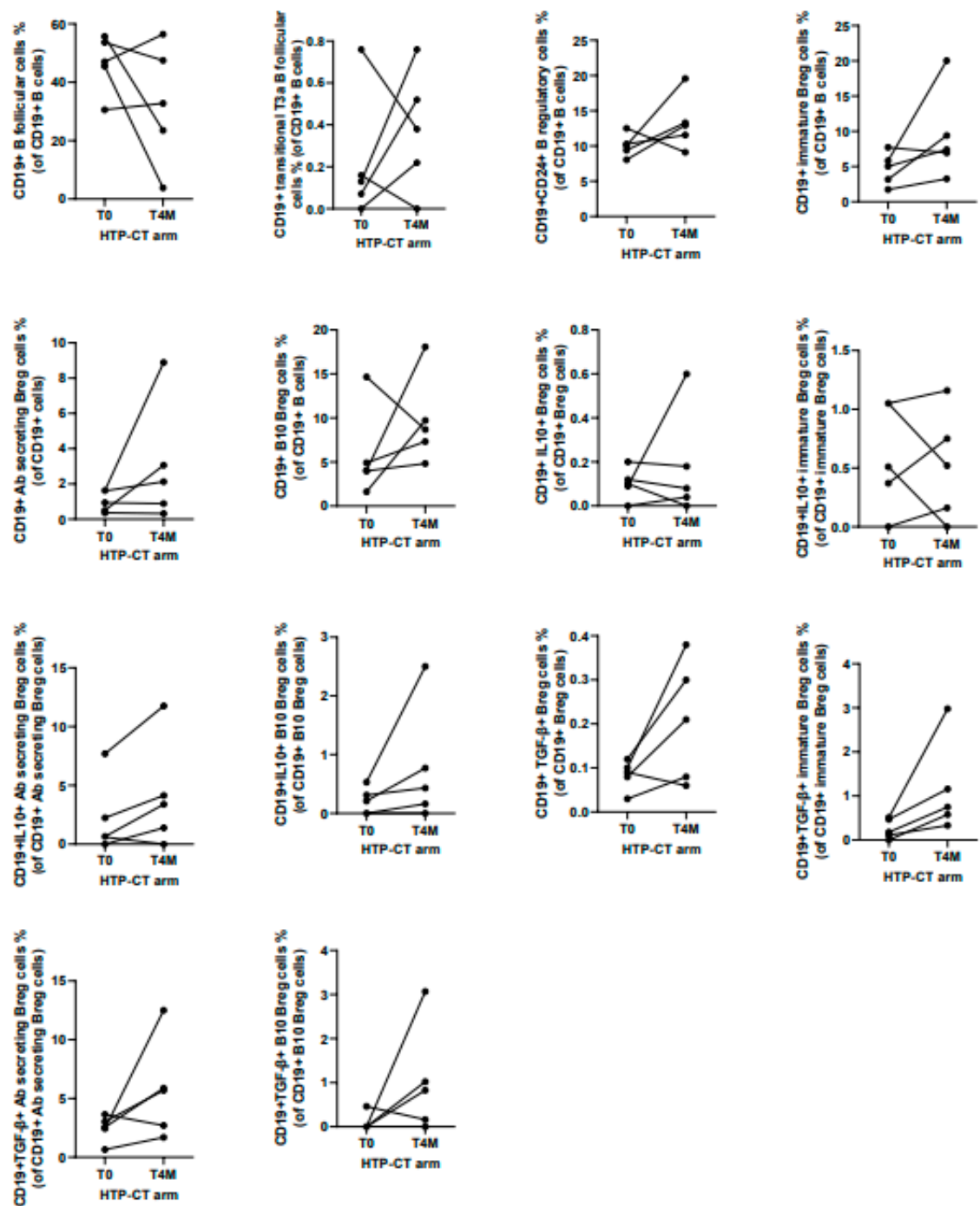
**Footnote:** Te: T effector; Th1: T helper 1; Th2: T helper 2; Th17: T helper 17; Th1/17: T helper 1/17; Tfh: T follicular helper; Tfh1: T follicular helper 1; Tfh2: T follicular helper 2; Tfh17: T follicular helper 17; Tfh1/17: T follicular helper 1/17; Treg: T regulatory; UM: unswitched memory; SM: switched memory; ASM: activated switched memory; DN: double negative; DN1: double negative 1; DN2: double negative 2; DN3: double negative 3; DN4: double negative 4; AcN: activated naïve; AnN: anergic naïve; RN: resting naïve; Ab: antibody secreting; T1/2: transitional 1/2; T3a: transitional 3a; MZP: marginal zone peripheral; Breg: B regulatory; AbS: antibody secreting; Imm: immature; Mon: monocytes.

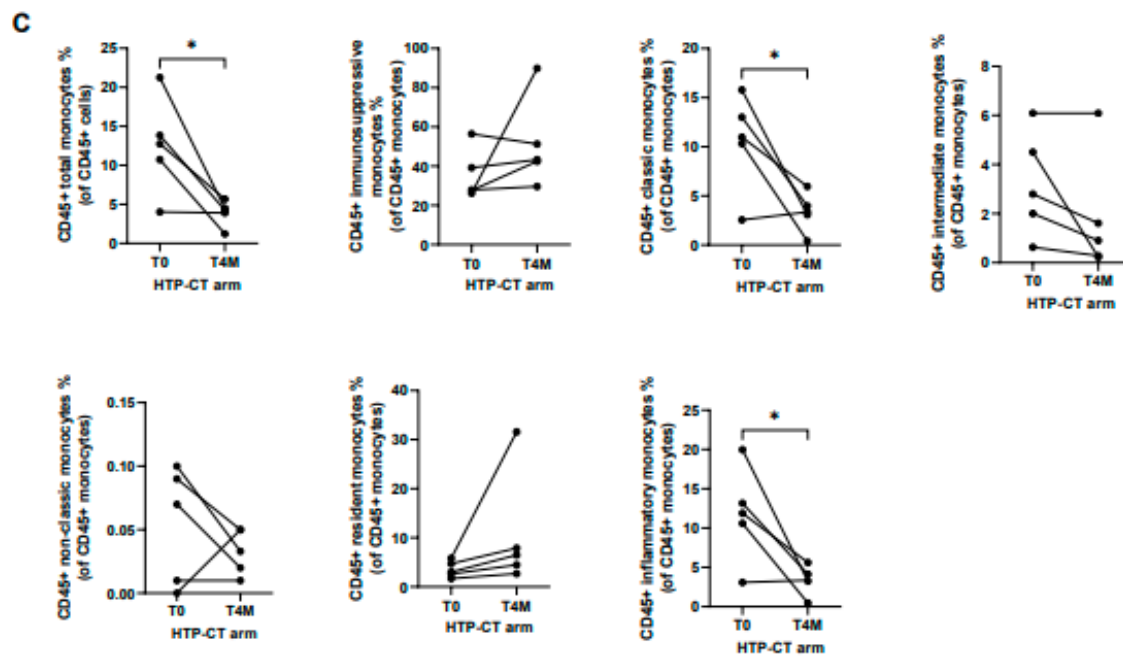
**Figure S14.** Changes of circulating immune cells (A: T-cell compartment; B: B-cell compartment; C: monocyte compartment) and concentrations of inflammatory and immune-related cytokines and chemokines (D: Pro Human Inflammation Panel I Assay; E: Pro TGF- $\beta$  Immunoassay; F: Pro Human Cytokine Immunoassay) between baseline (T0) and 4 months (T4M) after therapy onset in patients with pancreatic adenocarcinoma (PDAC) treated with HybridTherm Probe ablation plus chemotherapy (HTP-CT arm). Intra-group variables were compared using the paired t-test: \* =  $p < 0.05$ .



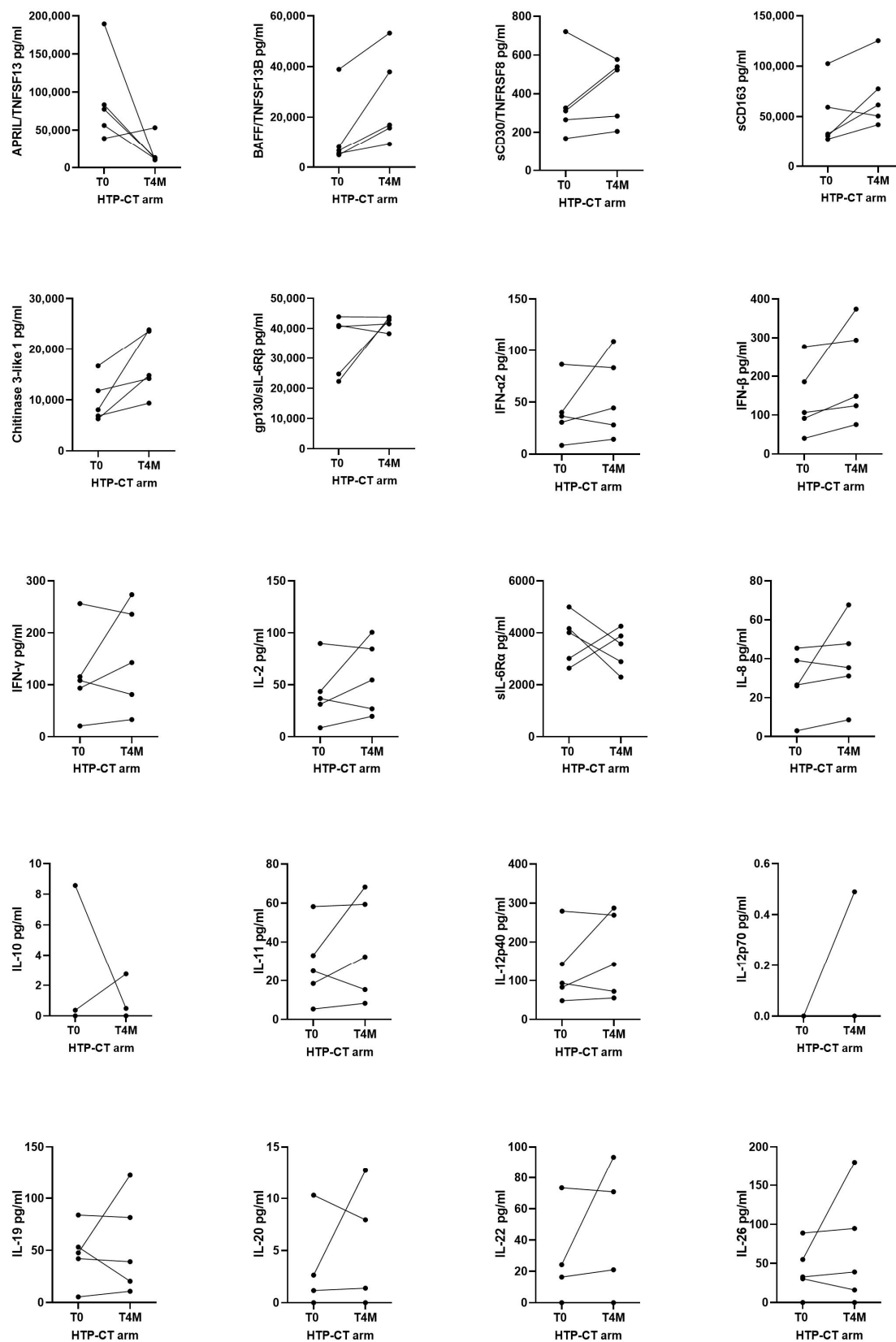
**B**

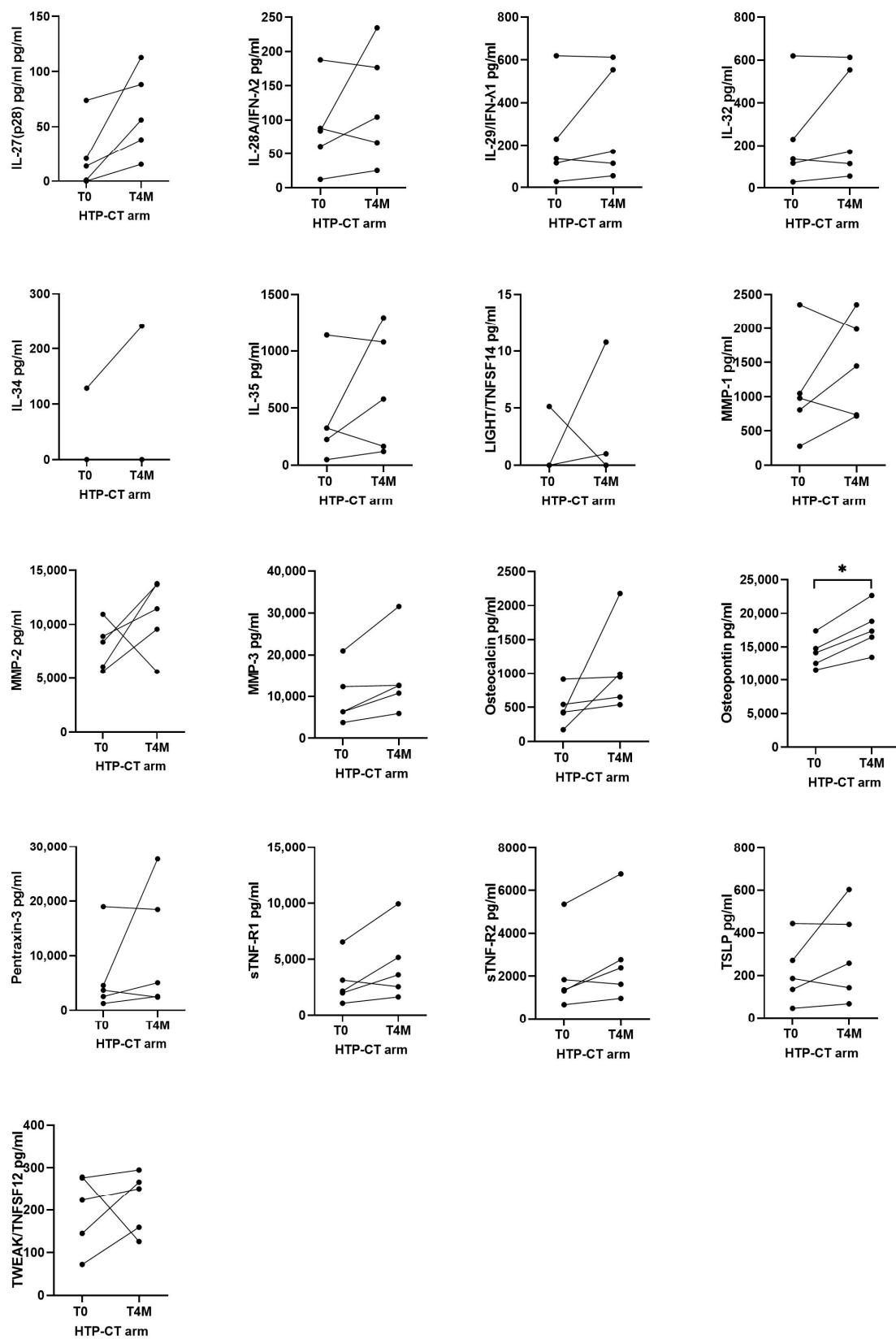






D



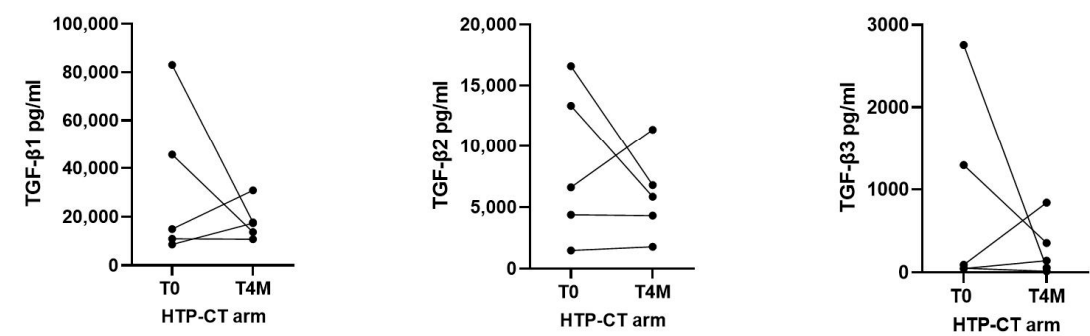


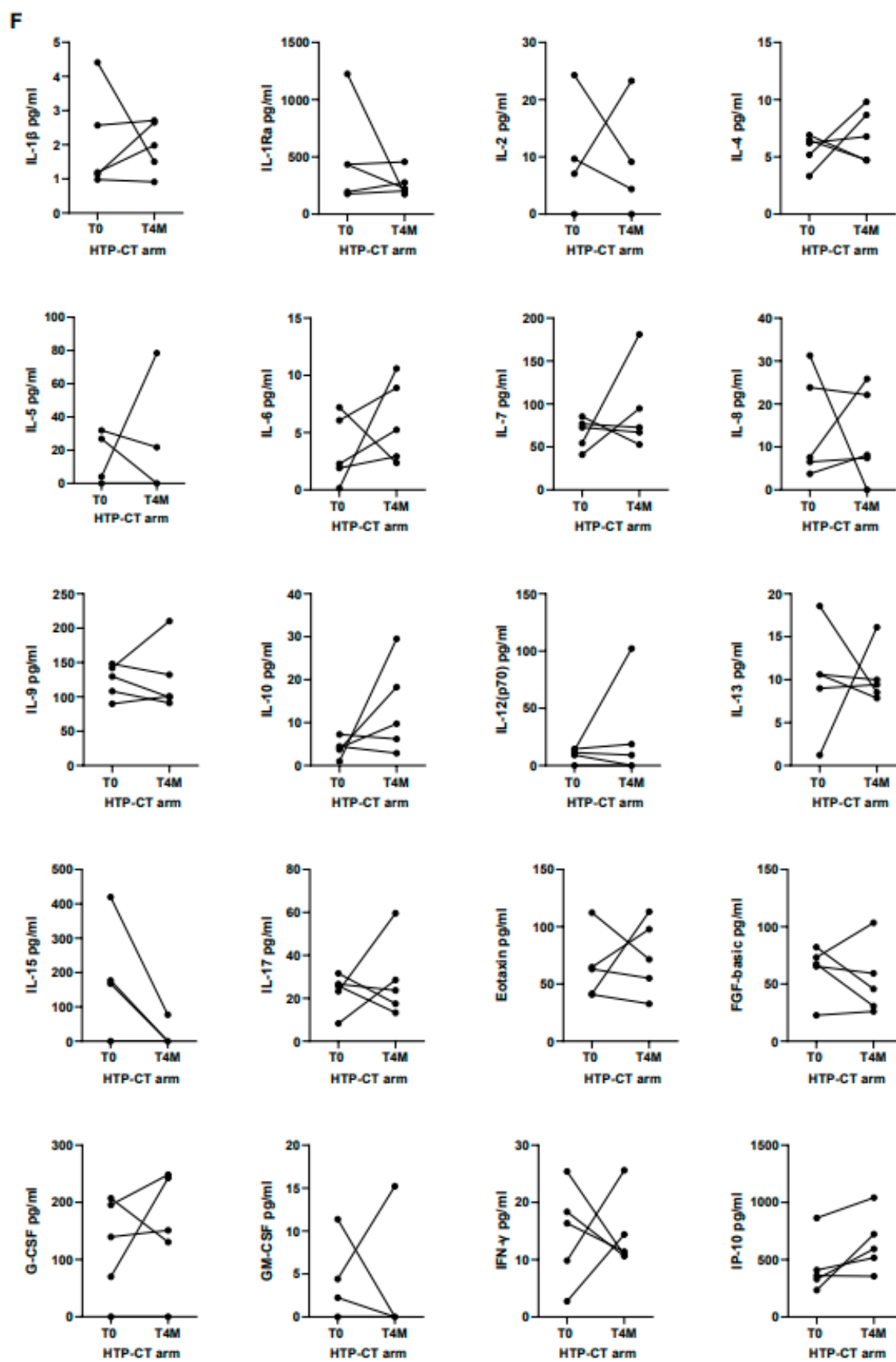
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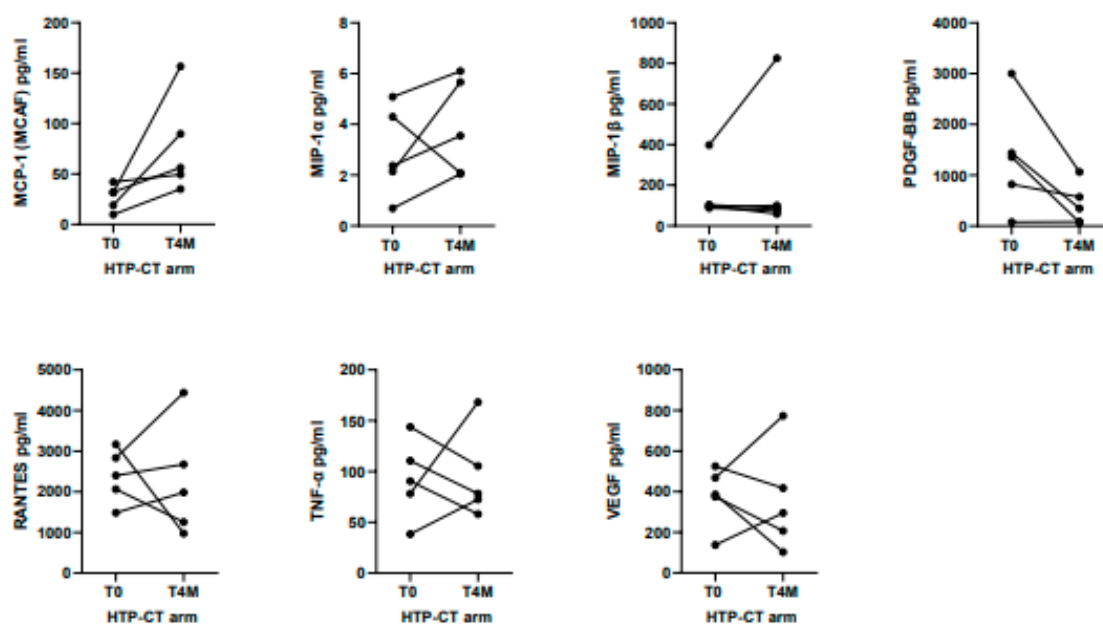
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E



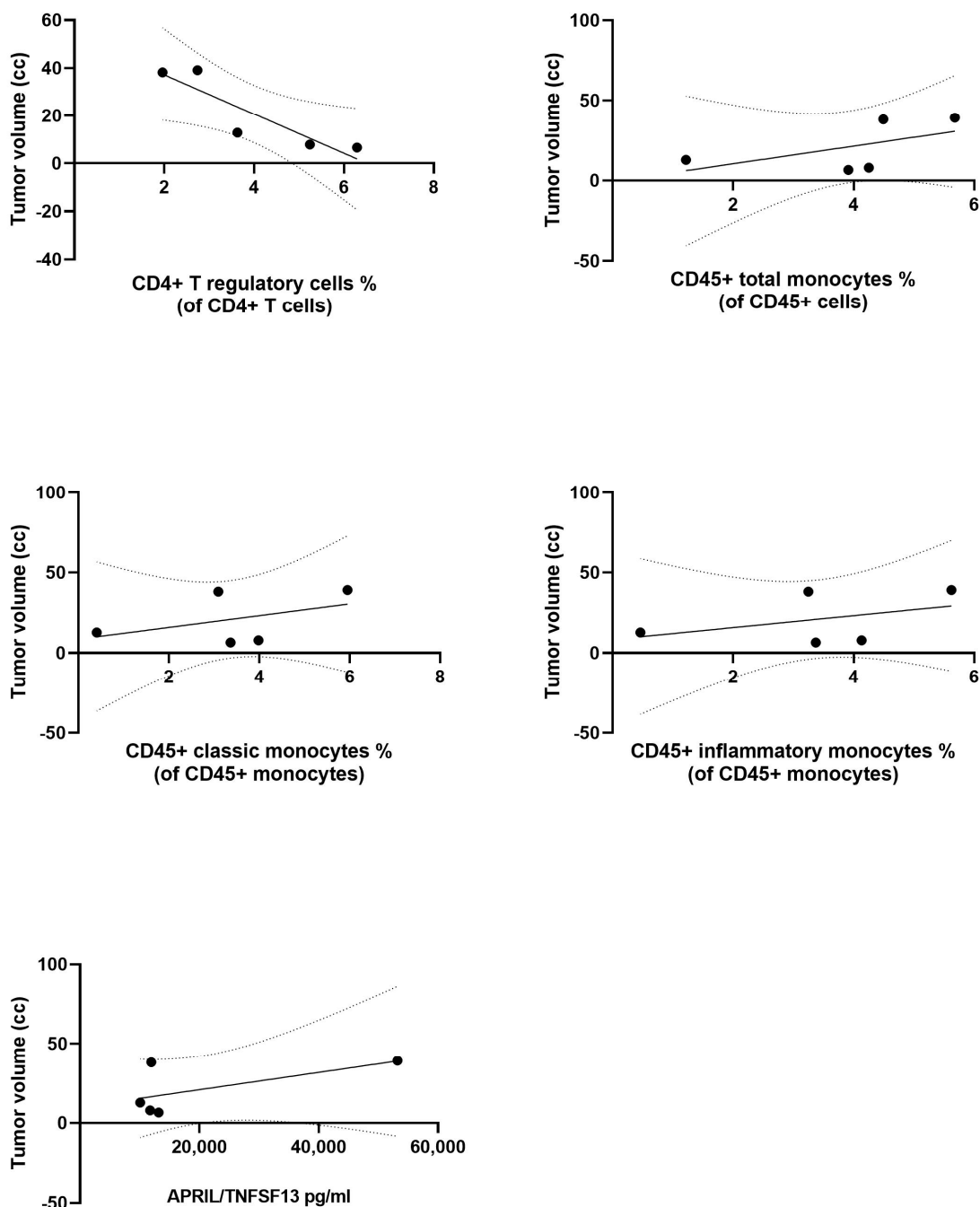




**Footnote:** Te: T effector; Th1: T helper 1; Th2: T helper 2; Th17: T helper 17; Th1/17: T helper 1/17; Tfh: T follicular helper; Tfh1: T follicular helper 1; Tfh2: T follicular helper 2; Tfh17: T follicular helper 17; Tfh1/17: T follicular helper 1/17; Treg: T regulatory; UM: unswitched memory; SM: switched memory; ASM: activated switched memory; DN: double negative; DN1: double negative 1; DN2: double negative 2; DN3: double negative 3; DN4: double negative 4; AcN: activated naïve; AnN: anergic naïve; RN: resting naïve; Ab: antibody secreting; T1/2: transitional 1/2; T3a: transitional 3a; MZP: marginal zone peripheral; Breg: B regulatory; AbS: antibody secreting; Imm: immature; Mon: monocytes.

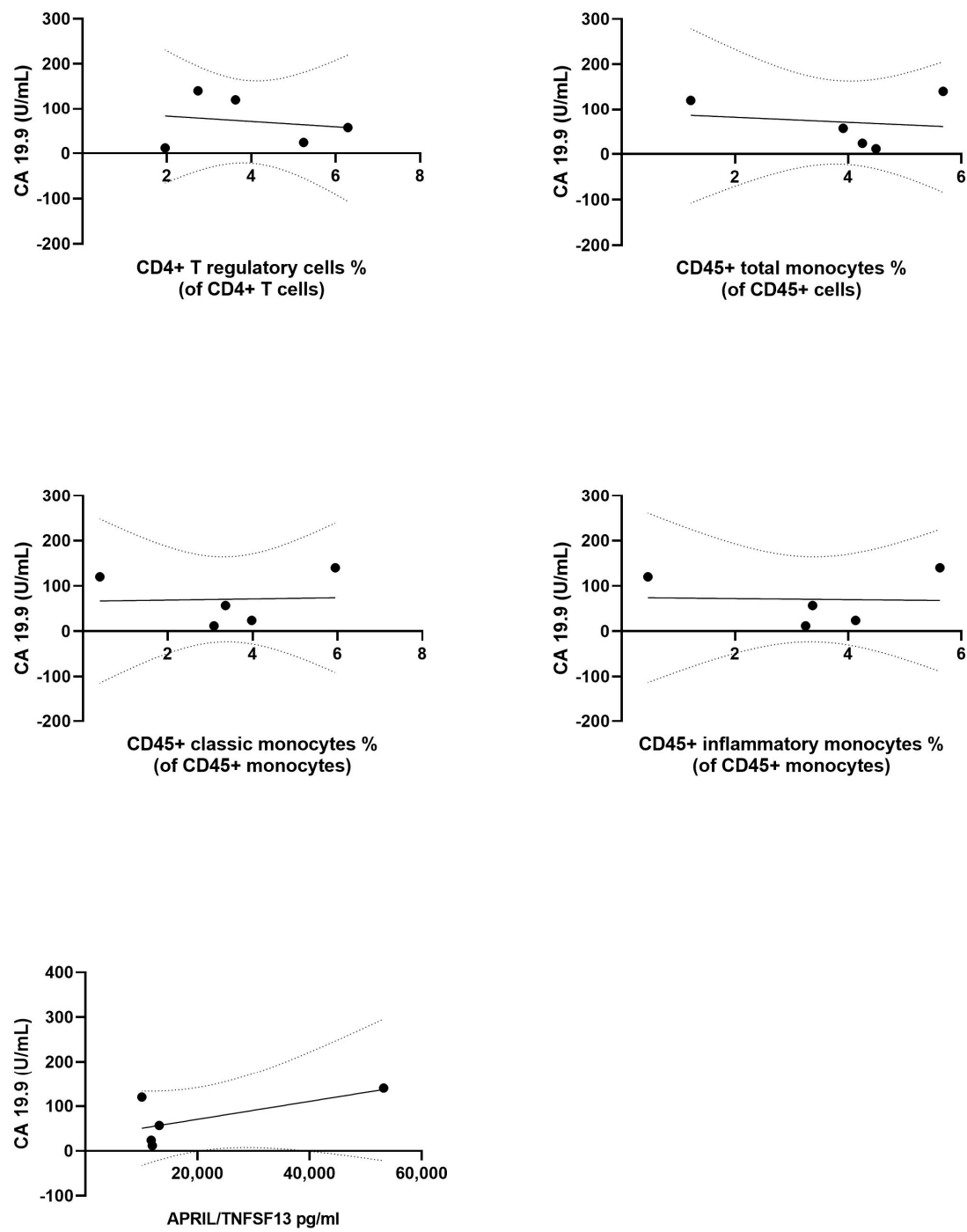
**Figure S15.** Correlation studies between circulating immune cells and cytokines significantly different in patients with pancreatic adenocarcinoma (PDAC) treated with HybridTherm Probe ablation plus chemotherapy (HTP-CT arm) from those treated with chemotherapy only (CT arm) at 4-month follow-up, and the tumor volume (A) and serum levels of tumor marker CA19-9 (B). Correlation studies were performed using the Spearman's rho rank correlation test.

**A**





**B**



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