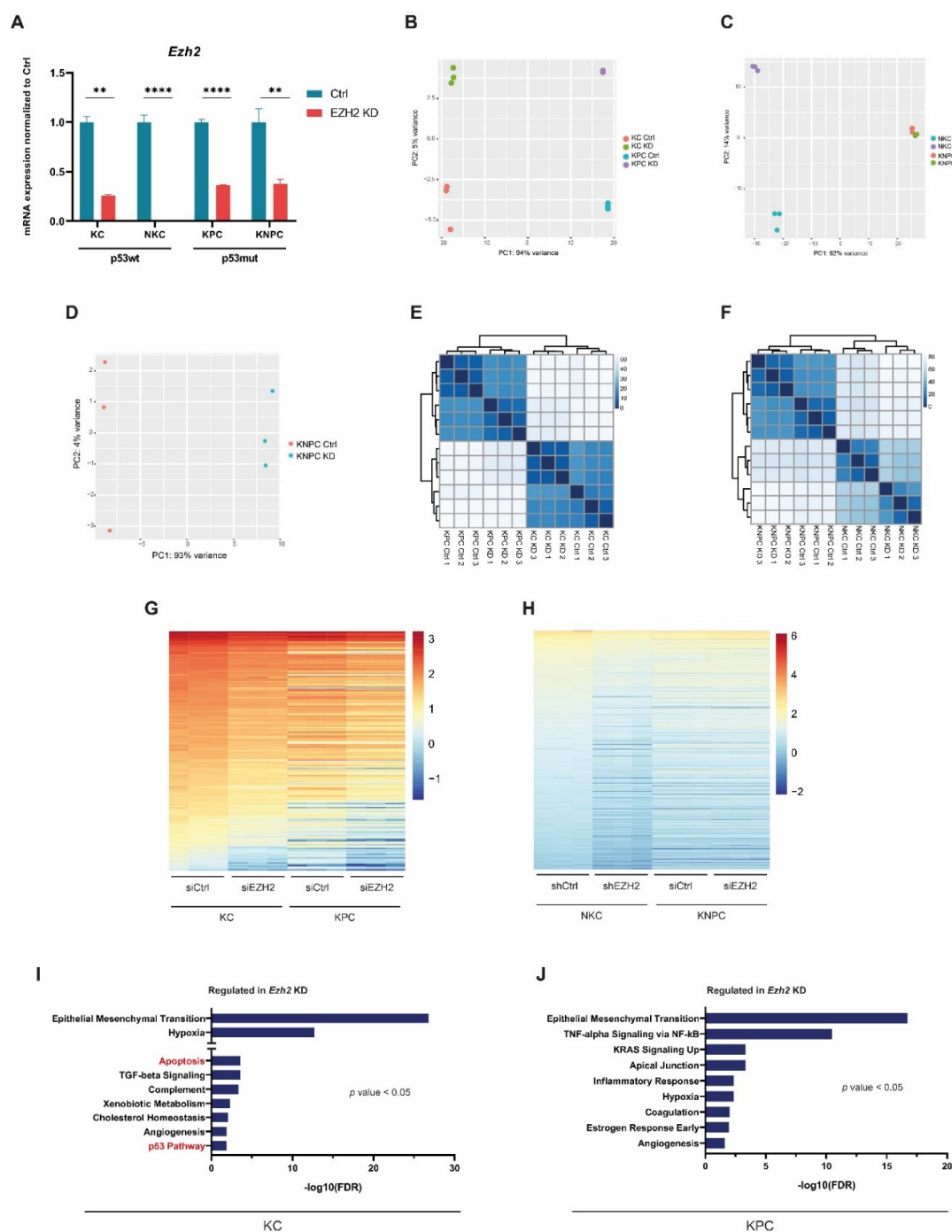
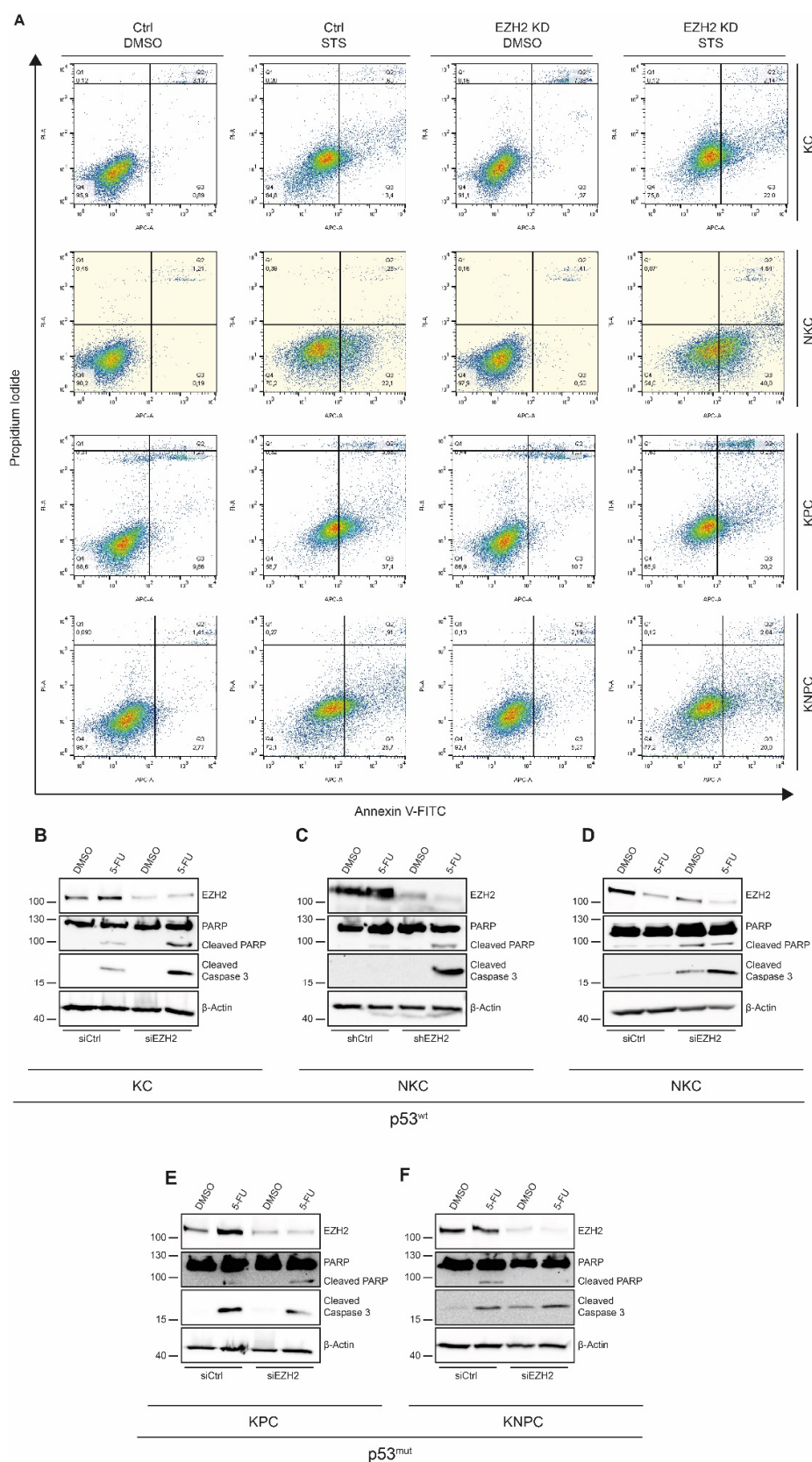


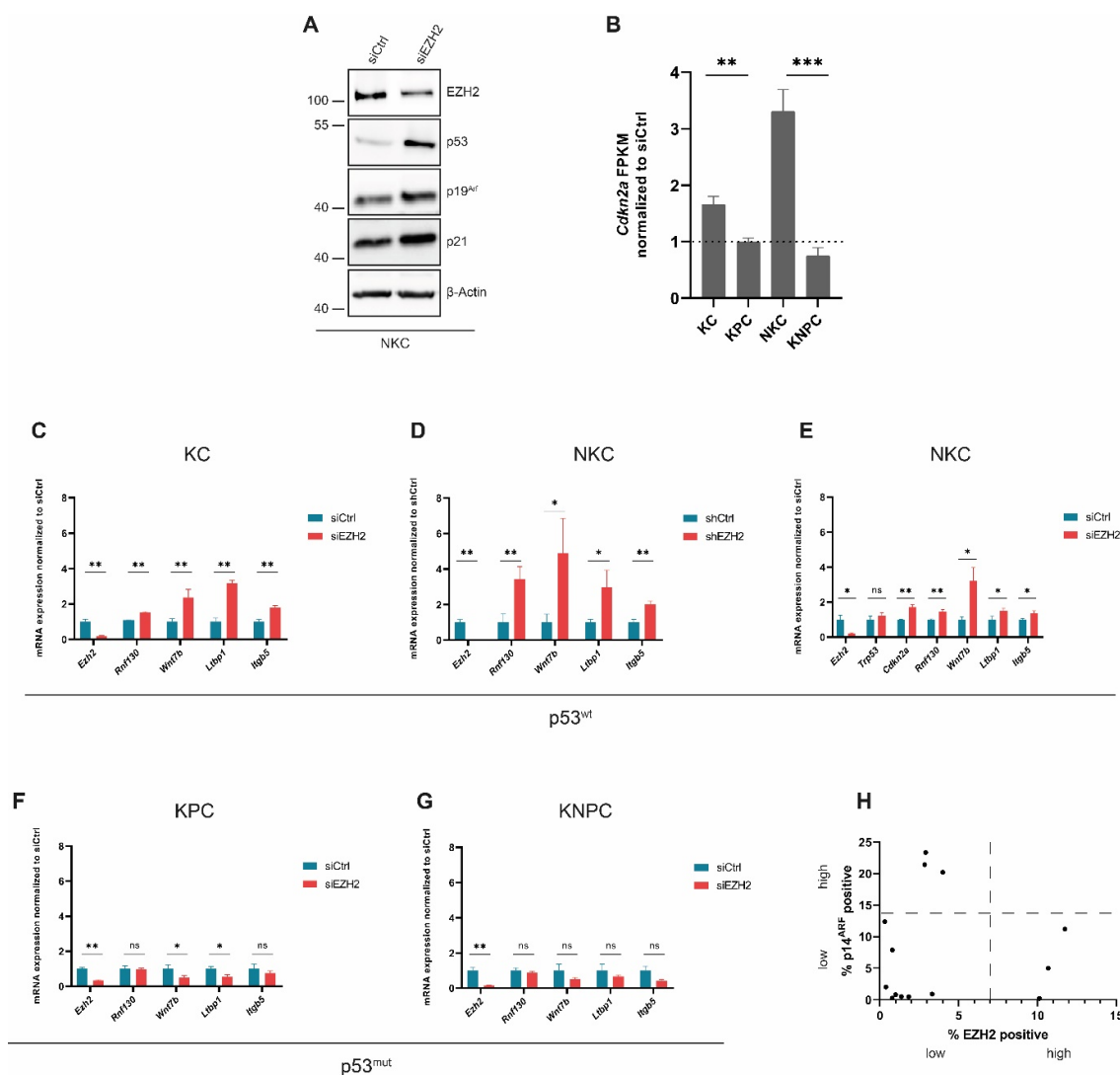
**Supplementary Figure S1.** Quantification of (immuno-) histological stainings depicted in Figure 1G. **A–C** Quantification of Ki-67 positive staining (**A**), Masson's Trichrome staining (**B**), and αSMA expression (**C**) of PDAC from orthotopically transplanted Panc1 cells (upper panel) and KrasG12D;Trp53R172H/+ (KPC) cells (lower panel). **D** Calculation of the activated stroma index by dividing the αSMA positive area by Masson's Trichrome blue area (collagen). For Masson's Trichrome and αSMA staining percentage of positively stained area of ten representative images of each tumor and for Ki-67 staining percentage of Ki-67 positive cells of ten representative images were measured using ImageJ Fiji. Each dot represents one mouse. Values represent mean ± SD. Significance was determined by two-tailed unpaired Student's t test; \*\*,  $p \leq 0.01$ ; ns, non-significant.



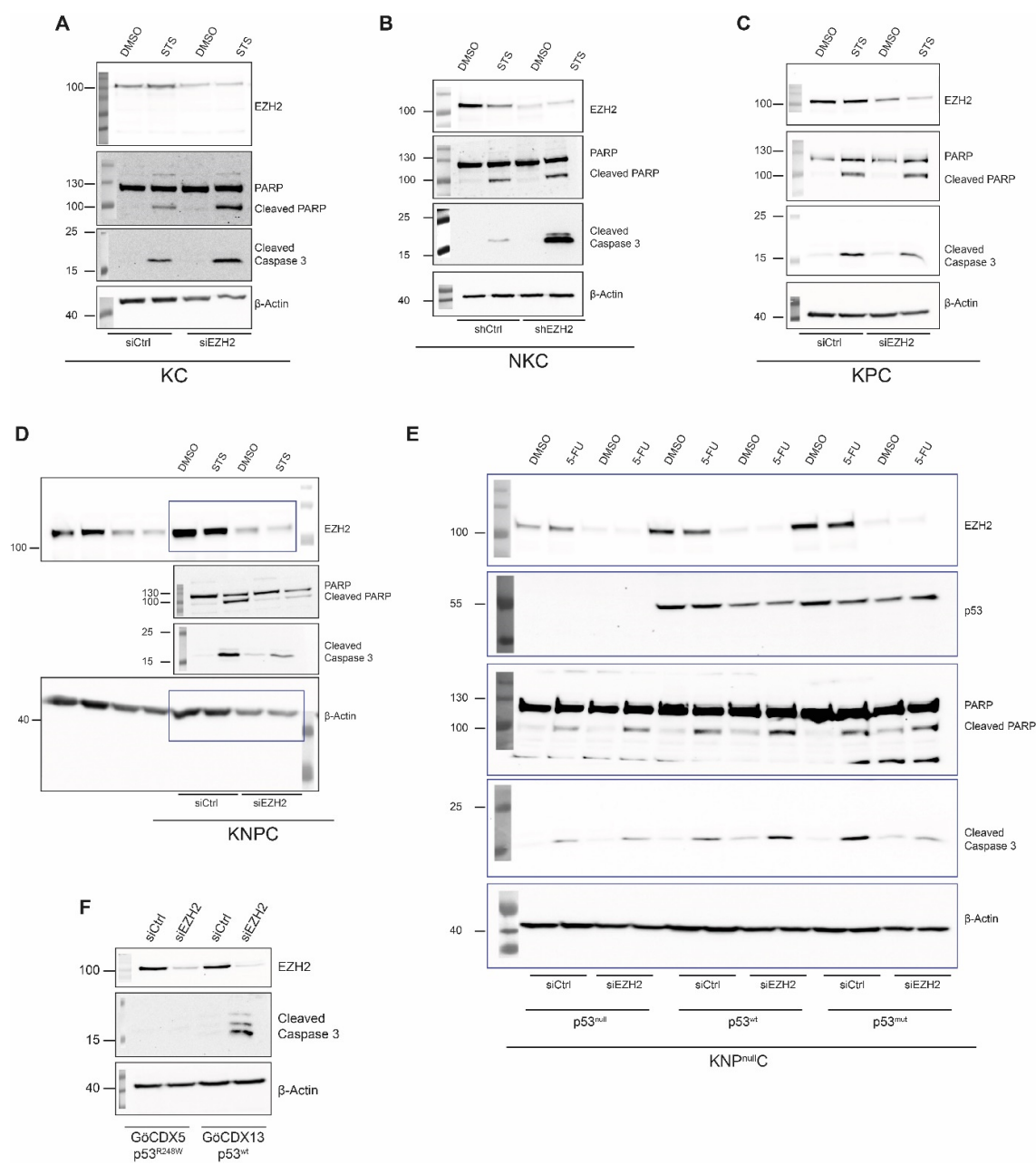
**Supplementary Figure S2.** *TP53*-status dependent EZH2 target gene regulation. **A** qRT-PCR analysis in the indicated p53wt and p53mut PDAC cells confirming knockdown of EZH2 ( $n = 3$ ). Values represent mean  $\pm$  SD. Significance was determined by Student's  $t$  test; \*\*,  $p \leq 0.01$ ; \*\*\*\*,  $p \leq 0.0001$ . **B–D** Principal component analysis (PCA) upon RNA-sequencing of KC and KPC cells (**B**) and NKC and KNPC cells (**C, D**) after EZH2 knockdown displaying distinct clusters of triplicates. **E, F** Sample-to-sample distances upon RNA-seq analysis of KC and KPC cells (**E**) and NKC and KNPC cells (**F**) upon EZH2 knockdown. **G, H** Heatmap illustrating genes being significantly downregulated (FPKM > 0.01,  $\log_2FC < -0.5$ ,  $q < 0.05$ ) upon knockdown of EZH2 in the indicated p53wt cells (KC: 188 genes, NKC: 964 genes) and its consequences on the expression of these genes in the respective p53mut cells. **I, J** Gene ontology (GO) analysis to reveal significantly upregulated pathways upon EZH2 depletion in the indicated PDAC cells ( $p < 0.05$ ).

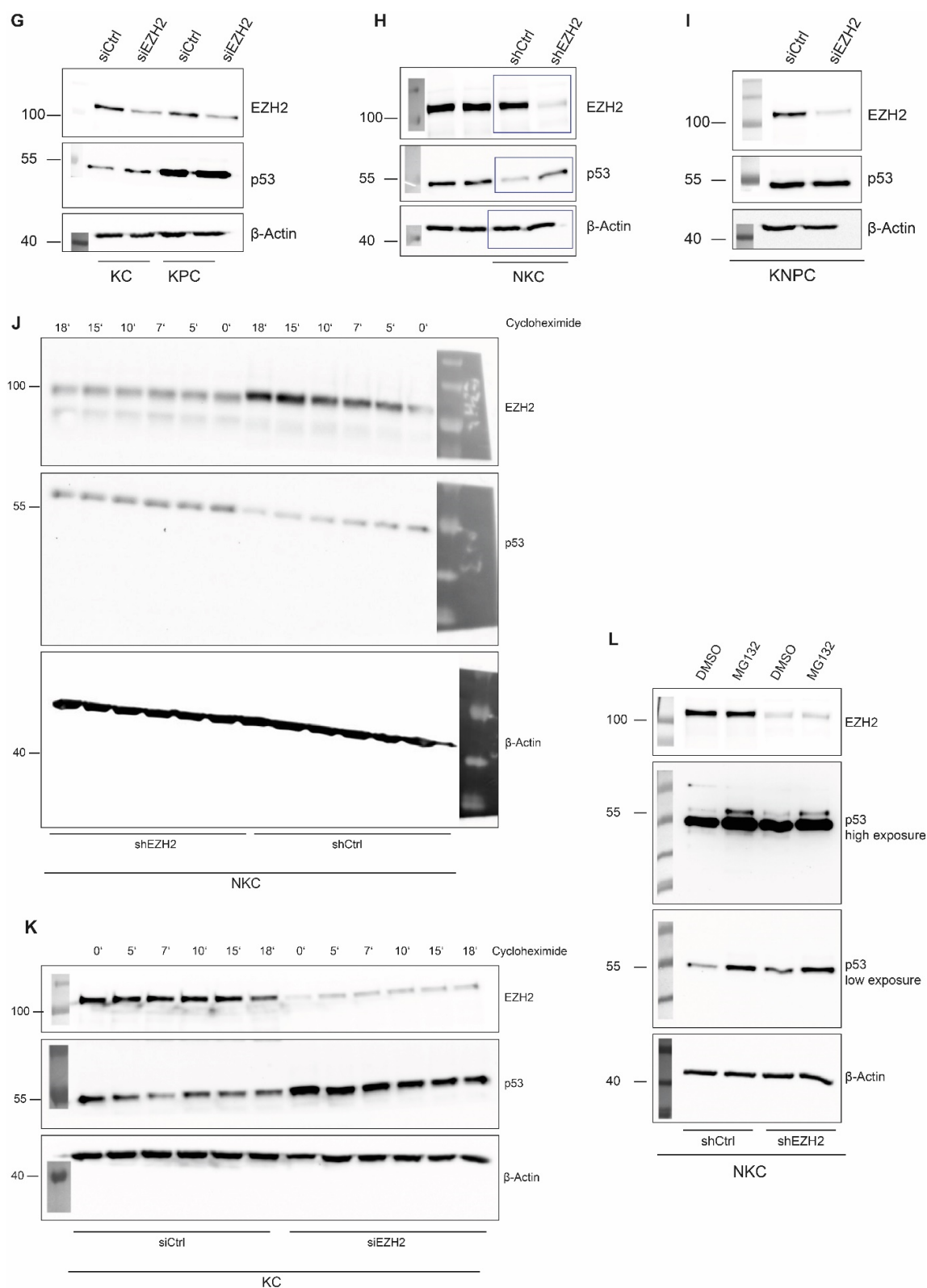


p53wt (KC, NKC) and p53mut (KPC, KNPC) expression and simultaneous treatment with staurosporine (STS). Representative flow cytometry results with the respective gating strategy upon Annexin-V/propidium iodide staining for each cell line referring to Figure 3. **B–F** Western blot analysis of apoptosis-related proteins upon knockdown of EZH2 and simultaneous treatment with 5-FU in the indicated p53wt and p53mut PDAC cells.

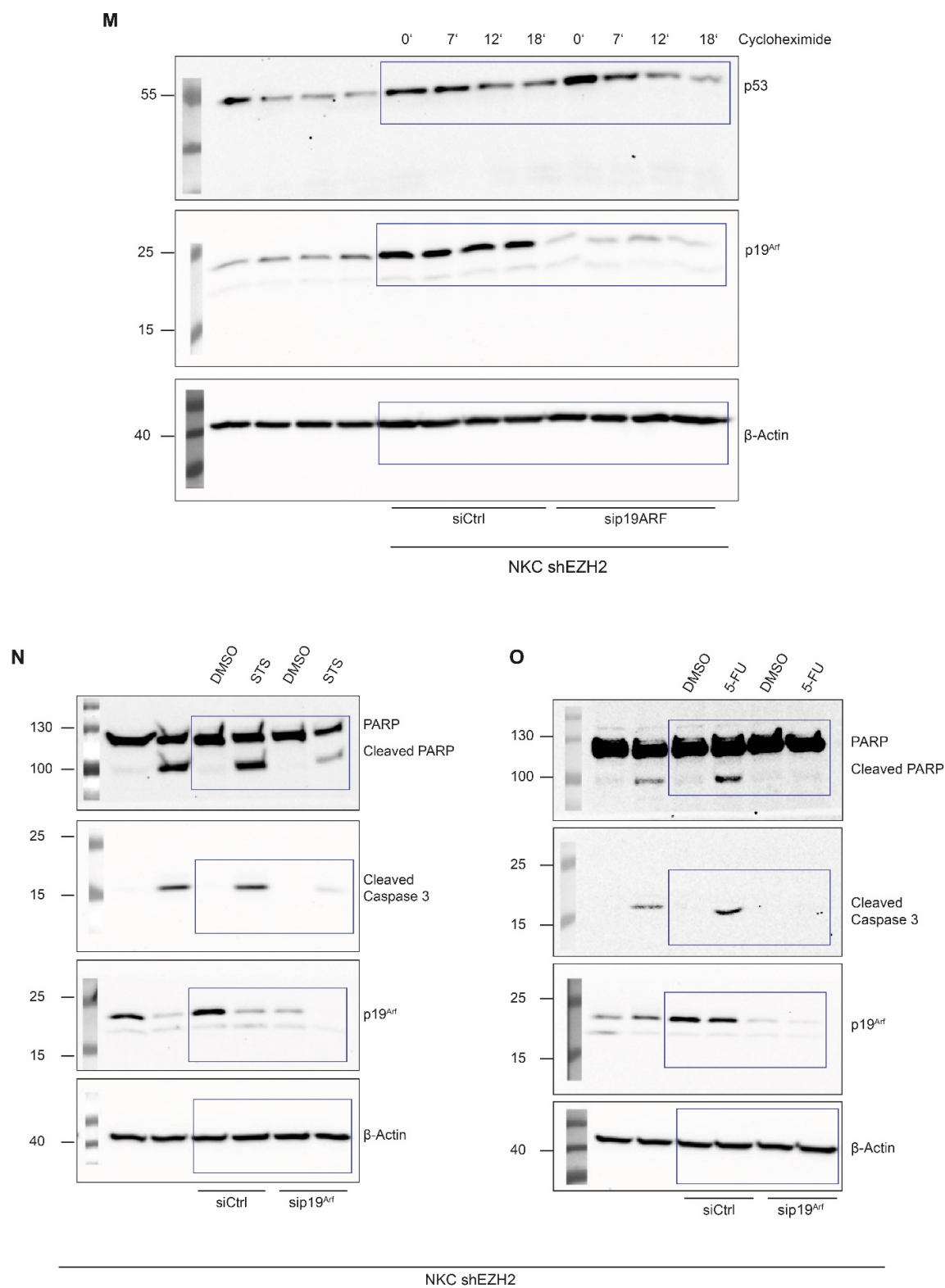


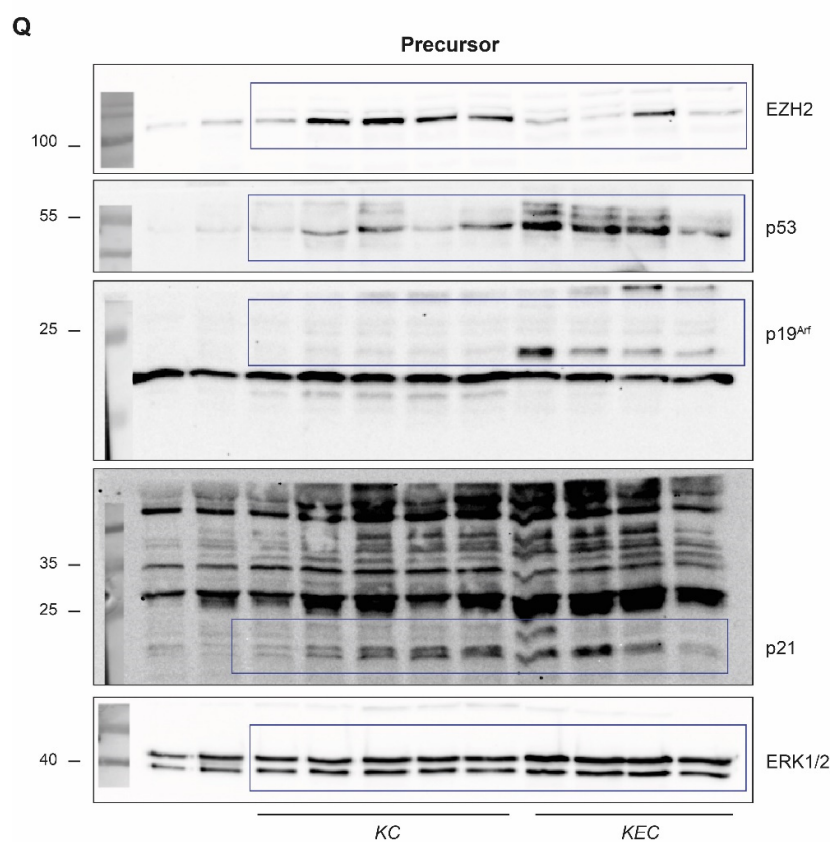
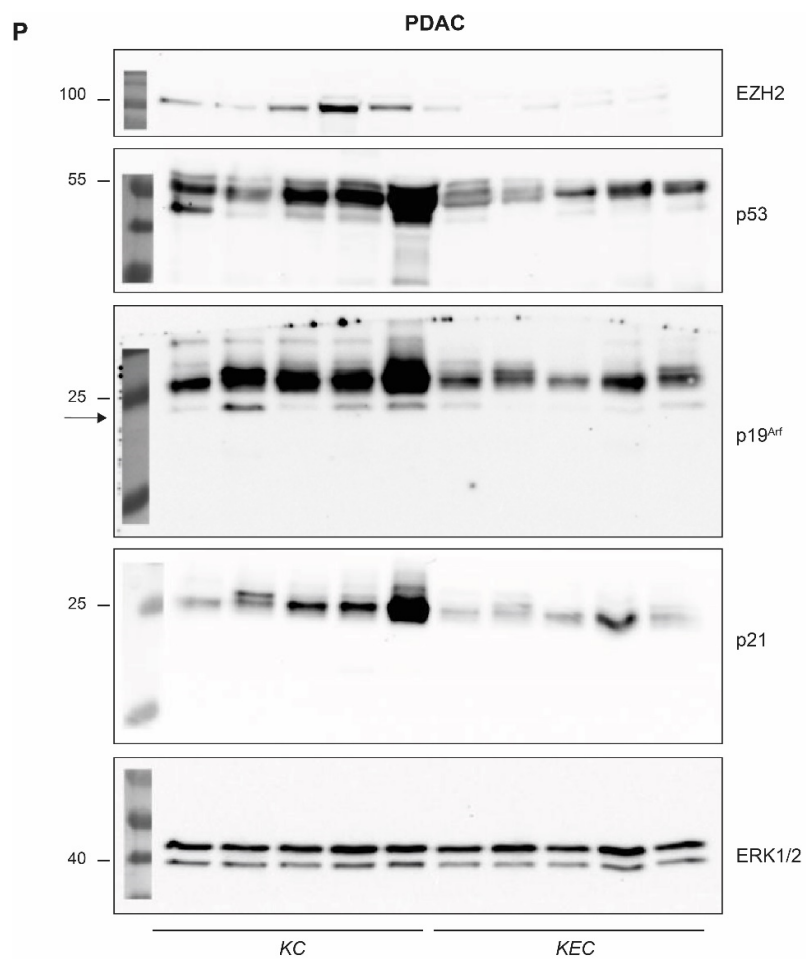
**Supplementary Figure S4.** EZH2-dependent target gene regulation differs in p53wt and p53mut PDAC. **A** Knockdown of EZH2 in p53wt NKC cells using siRNA following western blot analysis to investigate the consequences of EZH2 KD on the indicated proteins. **B** Normalized FPKM values of Cdkn2a expression after knockdown of EZH2 as revealed by RNA-seq in the indicated cell lines. Values represent mean  $\pm$  SD. Significance was determined by Student's t test; \*\*,  $p \leq 0.01$ ; \*\*\*,  $p \leq 0.001$ . **C–G** QRT-PCR analysis in the indicated PDAC cells upon knockdown of EZH2 validating the upregulation of these genes in p53wt cells ( $n = 3$ ). Values represent mean  $\pm$  SD. Significance was determined by Student's t test; \*,  $p \leq 0.05$ ; \*\*,  $p \leq 0.01$ ; ns, non-significant. **H** Table displaying number of patients in the indicated EZH2low/high and p14ARF low/high groups based on IHC staining.



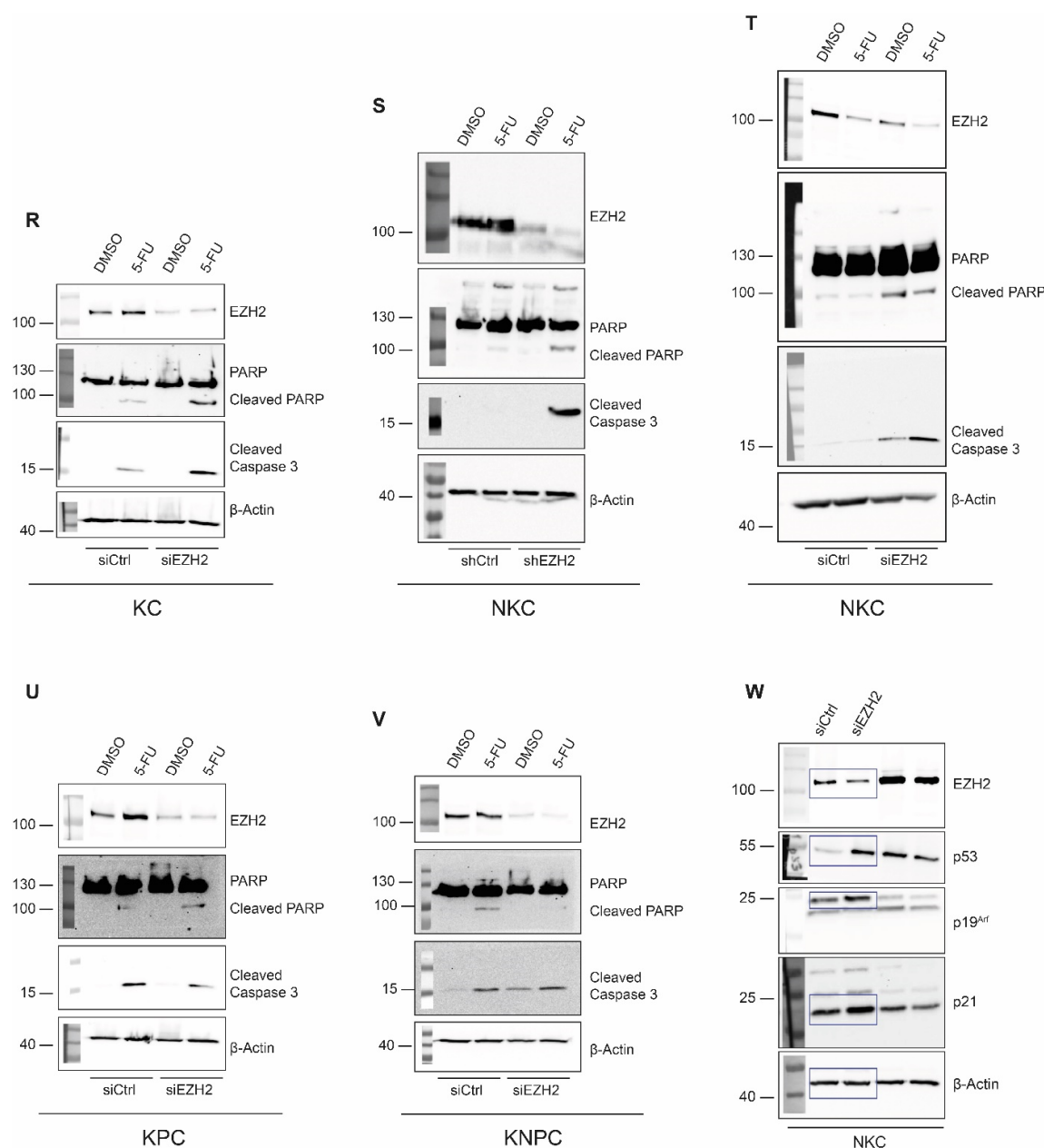












**Supplementary Figure S5.** Un-cut original Western Blot images. Original western blot images from those western blots shown throughout the manuscript. We used the PageRule Prestained Protein Ladder, 10 bis 180 kDa from Thermo Scientific (26616) having bands at 180, 130, 100, 70, 55, 40, 35, 25, 15, and 10 kDa. We did not label all bands of the molecular weight marker but only the most relevant. However, the other molecular weight sizes can be identified accordingly. For protein visualization, we used the Western Lightning Plus (NEL103E001EA) or Ultra (NEL111001EA) chemiluminescent substrate from PerkinElmer. However, the marker cannot be visualized with the chemiluminescent solution and was imaged separately. Subsequently, we added the marker from the raw colorimetric image onto the enhanced chemiluminescent image to compare the molecular weight of the protein of interest with the standard. Please note, that we pre-cut the membranes before antibody incubation around the expected molecular weight of the protein of interest in order to stain different proteins of distinct molecular weight on one membrane. Blue boxes indicate excerpt of protein lanes depicted in the respective Figures. **A** Un-cut western blot images referring to Fig. 3C. **B** Un-cut western blot images referring to Fig. 3E. **C** Un-cut western blot images referring to Fig. 3G. **D** Un-cut western blot images referring to Fig. 3I. **E** Un-cut western blot images referring to Fig. 3K. **F** Un-cut western blot images referring to Fig. 3L. **G** Un-cut western blot images referring to Fig. 4A

and 4C. **H** Un-cut western blot images referring to Fig. 4B. **I** Un-cut western blot images referring to Fig. 4D. **J** Un-cut western blot images referring to Fig. 4F. **K** Un-cut western blot images referring to Fig. 4G. **L** Un-cut western blot images referring to Fig. 4H. **M** Un-cut western blot images referring to Fig. 5E. **N** Un-cut western blot images referring to Fig. 5F. **O** Un-cut western blot images referring to Fig. 5G. **P** Un-cut western blot images referring to Fig. 6B. **Q** Un-cut western blot images referring to Fig. 6H. **R** Un-cut western blot images referring to Supplementary Fig. 3B. **S** Un-cut western blot images referring to Supplementary Fig. 3C. **T** Un-cut western blot images referring to Supplementary Fig. 3D. **U** Un-cut western blot images referring to Supplementary Fig. 3E. **V** Un-cut western blot images referring to Supplementary Fig. 3F. **W** Un-cut western blot images referring to Supplementary Fig. 4A.

### Supplementary tables

**Supplementary Table S1.** sgRNA sequences and validation primers for CRISPR/Cas9-mediated *EZH2* knockout.

	<b>Murine <i>Ezh2</i></b>	<b>Human <i>EZH2</i></b>
<b>sgRNA sequence</b>	GTGGTGGATGCAACCCGAAA	GTGGTGGATGCAACCCGCAA
<b>Forward primers for knockout validation</b>	CCTGTGTAAGTGGGTGTGCT	TGCCTATTCGTGATGTTTGAAG
<b>Reverse primers for knockout validation</b>	GTTTGCTGTCACTTGGCTGG	TGTCAACAGCAGGGTGAGAAA

**Supplementary Table S2.** siRNA sequences.

	<b>Sense</b>	<b>Antisense</b>
<b>siEZH2</b>	GGAUACAGCCUGUGCACAUT	AUGUGCACAGGCUGUAUCCTC
<b>siP19<sup>Arf</sup></b>	GGCUAGAGAGGAUCUUGAGTT	CUCAAGAUCUCUCUAGCCTC

**Supplementary Table S3.** Primer for qRT-PCR.

<b>Target</b>	<b>Direction</b>	<b>Sequence</b>
<i>Cdkn2a</i>	forward	CGCAGGTTCTTGGTCACTGT
<i>Cdkn2a</i>	reverse	TGTTACGAAAGCCAGAGCG
<i>Ezh2</i>	forward	CAACCCGAAAGGGCAACAAA
<i>Ezh2</i>	reverse	ACC AGT CTG GAT AGC CCT CT
<i>Itgb5</i>	forward	GAAGTGCCACCTCGTGTGAA
<i>Itgb5</i>	reverse	GGACCGTGGATTGCCAAAGT
<i>Ltp1</i>	forward	GGTCGCATCAAGGTGGTCTTT
<i>Ltp1</i>	reverse	GTGGTGGTATTCCCCTTCTGG
<i>Rnf130</i>	forward	CTGCCCATCCACGGAGTTG
<i>Rnf130</i>	reverse	CAAGGCGATCCACTGTTTGA
<i>Rplp0</i>	forward	TGGGCAAGAACACCATGATG
<i>Rplp0</i>	reverse	AGTTTCTCCAGAGCTGGGTTGT
<i>Trp53</i>	forward	AGGTGTGCGTAGCACC
<i>Trp53</i>	reverse	CCCCACAACACCAAGT
<i>Wnt7b</i>	forward	CTTCACCTATGCCATCACGG
<i>Wnt7b</i>	reverse	TGGTTGTAGTAGCCTTGCTCT

**Supplementary Table S4.** Antibodies used for western blotting.

<b>Antibody</b>	<b>Company</b>	<b>Number</b>	<b>Dilution</b>
Actin-HRP	Sigma	A3854	1:40000
Cleaved caspase 3	Cell Signaling	9661	1:500
Erk (1/2)	Cell Signaling	9102	1:1000
EZH2	Cell Signaling	5246	1:1000
p19 <sup>Arf</sup>	Abcam	Ab80	1:1000
p19 <sup>Arf</sup>	Santa Cruz	32748	1:250
p53	Cell Signaling	2524	1:1000
PARP	Cell Signaling	9542	1:1000
Anti-mouse (IgG) HRP	Cell Signaling	7076	1:6500
Anti-rabbit (IgG) HRP	Cell Signaling	7074	1:6500
Anti-rat (IgG) HRP	Santa Cruz	2006	1:5000

**Supplementary Table S5.** Antibodies used for immunohistochemistry and immunofluorescence.

Antibody	Company	Number	Dilution
EZH2 (mouse)	Cell Signaling	5246	1:100
EZH2 (human)	Leica Biosystems	NCL-L-EZH2	1:50
Ki-67	Thermo Fisher	RM9106	1:600
αSMA	Agilent Dako	M0851	1:100
p14	Cell Signaling	2407	1:400
p19 <sup>Arf</sup>	Santa Cruz	32748	1:100
Alexa Fluor 488 donkey anti-rat	Invitrogen	A21208	1:500

**Supplementary Table S6.** Antibodies used for ChIP experiments.

Antibody	Company	Number	Amount
EZH2	Diagenode	C15410039-classic	2 µg
H3K4me3	Cell Signaling	9751	2 µg
Rabbit IgG	Diagenode	C15410206	2 µg

**Supplementary Table S7.** Primer for qRT-PCR following ChIP experiments.

Target	Direction	Sequence
<i>Cdkn2a</i> TSS	forward	GACCGTGAAGTTCAGC
<i>Cdkn2a</i> TSS	reverse	GGGGTCGCTTCTTCGG

**Supplementary Table S8.** Favorable prognosis genes depicted in Figure 2C/D.

<i>CAMTA2</i>	<i>ABHD8</i>	<i>HCFC1</i>	<i>VPS54</i>	<i>SNAI3</i>	<i>GSE1</i>	<i>TBPL1</i>
<i>TSPYL2</i>	<i>SEMA6C</i>	<i>PPP1R10</i>	<i>MCM3AP</i>	<i>DOHH</i>	<i>LRRC29</i>	<i>ARIH2</i>
<i>ANAPC2</i>	<i>TMEM74B</i>	<i>SAFB</i>	<i>PMM1</i>	<i>RFX6</i>	<i>UAP1L1</i>	<i>RFX2</i>
<i>DEF8</i>	<i>GALT</i>	<i>KLHL36</i>	<i>ATP6V0A1</i>	<i>HERC1</i>	<i>KIF5A</i>	<i>SLC22A5</i>
<i>USP20</i>	<i>COG8</i>	<i>TMEM175</i>	<i>COQ10A</i>	<i>PRPSAP2</i>	<i>SLC27A3</i>	<i>DOPEY1</i>
<i>MICAL1</i>	<i>RAB11FIP3</i>	<i>ABCA5</i>	<i>RAB4B</i>	<i>UNC13B</i>	<i>DYNLL2</i>	<i>BAHCC1</i>
<i>SOCS2</i>	<i>EPOR</i>	<i>SSBP4</i>	<i>NEIL1</i>	<i>GNG7</i>	<i>PNISR</i>	<i>ASPHD1</i>
<i>SLC43A2</i>	<i>DENND4B</i>	<i>BRSK1</i>	<i>RBM4B</i>	<i>SNTA1</i>	<i>NCALD</i>	<i>MPDZ</i>
<i>ARNT2</i>	<i>NECAB3</i>	<i>BBS5</i>	<i>QTRT1</i>	<i>TACO1</i>	<i>INPP5B</i>	<i>MLLT6</i>
<i>WDR37</i>	<i>NAT9</i>	<i>TYK2</i>	<i>SPATC1L</i>	<i>NCDN</i>	<i>RAD51C</i>	<i>SMPD1</i>
<i>INPP5K</i>	<i>NFASC</i>	<i>CBX7</i>	<i>ZBTB46</i>	<i>HNRNPA0</i>	<i>TRMT10B</i>	<i>ULK3</i>
<i>PITPNA</i>	<i>PKD1</i>	<i>RNF166</i>	<i>CXXC4</i>	<i>ALKBH5</i>	<i>TAPT1</i>	<i>XAB2</i>
<i>PPP1R3F</i>	<i>ARMCX2</i>	<i>RBM14</i>	<i>IPO13</i>	<i>PSPH</i>	<i>TSPYL1</i>	<i>PPP6R2</i>
<i>MAMDC4</i>	<i>PNPLA6</i>	<i>ATP1B2</i>	<i>BTBD6</i>	<i>MAVS</i>	<i>POLR3H</i>	<i>MAP2K6</i>
<i>ARMC5</i>	<i>TMEM91</i>	<i>APH1B</i>	<i>WHRN</i>	<i>DVL2</i>	<i>NAP1L2</i>	<i>ANKRD13B</i>
<i>RNF167</i>	<i>B4GAT1</i>	<i>PLD6</i>	<i>PIP5K1C</i>	<i>CD81</i>	<i>INTS3</i>	<i>RBM5</i>
<i>CCDC106</i>	<i>IRGQ</i>	<i>PTPRS</i>	<i>IRF2BP1</i>	<i>GOLGA3</i>	<i>WDR83</i>	<i>RMND5B</i>
<i>MUM1</i>	<i>INCA1</i>	<i>ZFP3</i>	<i>ST3GAL2</i>	<i>FRS3</i>	<i>PFDN5</i>	<i>GAS8</i>
<i>SFXN5</i>	<i>RNASEH2C</i>	<i>GDF11</i>	<i>EFHC1</i>	<i>DIDO1</i>	<i>ANGEL1</i>	<i>PHKA2</i>
<i>TLE2</i>	<i>ACSF3</i>	<i>NCOA5</i>	<i>SDR39U1</i>	<i>IQCD</i>	<i>SYT5</i>	<i>GPX4</i>
<i>CBX6</i>	<i>GADD45G</i>	<i>NICN1</i>	<i>FOXA2</i>	<i>IGIP</i>	<i>NELL2</i>	<i>FKBP8</i>
<i>KCND1</i>	<i>CBX8</i>	<i>HDAC4</i>	<i>CCDC130</i>	<i>ADD1</i>	<i>REC8</i>	<i>RNPC3</i>
<i>RBM10</i>	<i>POMT1</i>	<i>KLHDC4</i>	<i>CERK</i>	<i>MAGEE1</i>	<i>HEXDC</i>	<i>ATP6AP1</i>
<i>TBC1D13</i>	<i>ARGLU1</i>	<i>ADGRL1</i>	<i>FN3K</i>	<i>HDAC3</i>	<i>SCG2</i>	<i>MC1R</i>
<i>RAD51D</i>	<i>PELP1</i>	<i>LIPE</i>	<i>SYNGR1</i>	<i>HIRA</i>	<i>STK40</i>	<i>MVB12B</i>
<i>FXR2</i>	<i>OAZ1</i>	<i>CCM2</i>	<i>FAM219A</i>	<i>FASTK</i>	<i>KCNH2</i>	<i>PDE3B</i>
<i>VAMP2</i>	<i>PPP1R12C</i>	<i>MBLAC1</i>	<i>NUDT18</i>	<i>TBC1D17</i>	<i>CALM1</i>	<i>SPECC1L</i>
<i>KLHL22</i>	<i>ZBTB40</i>	<i>DNMT3A</i>	<i>DCLK2</i>	<i>PRKAR2B</i>	<i>NEK9</i>	<i>MZF1</i>
<i>RAB6B</i>	<i>SPATA7</i>	<i>GRK6</i>	<i>RCAN2</i>	<i>SOWAHA</i>	<i>B4GALT6</i>	<i>DCAF15</i>
<i>EVL</i>	<i>MED9</i>	<i>SALL2</i>	<i>NPR1</i>	<i>ENPP5</i>	<i>TEX264</i>	<i>MIS12</i>
<i>TMEM203</i>	<i>KCTD17</i>	<i>RGS11</i>	<i>SLC25A44</i>	<i>NXF1</i>	<i>TMEM150C</i>	<i>TSC1</i>
<i>PRAF2</i>	<i>GPR162</i>	<i>MKS1</i>	<i>ID4</i>	<i>RAB17</i>	<i>CLEC3B</i>	<i>PHKG2</i>
<i>CYB561D1</i>	<i>MCF2L</i>	<i>UNC119B</i>	<i>TRIM39</i>	<i>RANBP3</i>	<i>JMJD6</i>	<i>AKAP8L</i>
<i>ZBTB48</i>	<i>SNAPC4</i>	<i>LUC7L</i>	<i>PIGT</i>	<i>RFXAP</i>	<i>TUBGCP6</i>	<i>PPP1R21</i>
<i>ELMOD3</i>	<i>SNRNP70</i>	<i>RAPGEF4</i>	<i>THEM4</i>	<i>MRPL38</i>	<i>ITGAE</i>	<i>CERS4</i>
<i>PHLDB3</i>	<i>TTC13</i>	<i>TBL1X</i>	<i>CHPF2</i>	<i>DEXI</i>	<i>ATP6V0E2</i>	<i>CUL9</i>
<i>SLC25A11</i>	<i>CCDC92</i>	<i>SPPL2B</i>	<i>TMEM240</i>	<i>ACCS</i>	<i>SFI1</i>	<i>FBXO46</i>
<i>VASH1</i>	<i>MTHFR</i>	<i>RGS9</i>	<i>CCND2</i>	<i>HAP1</i>	<i>SMDT1</i>	<i>TECPR1</i>

LRSAM1	LPCAT1	MRPL53	SAFB2	POMGNT1	DXO	LENG1
CBFA2T2	PKIG	SCN1B	ZBTB4	NAA60	GPANK1	SRR
FAM53B	USF2	RWDD2A	CLDN15	CACNA1A	DNAJC18	OTUD5
BRF1	RNF146	TOP3A	EID2B	RECQL5	IGHMBP2	FZD8
NUAK2	MAP1LC3A	GCH1	MAGEH1	IKZF4	GNAO1	EFNB3
TBKBP1	STRADA	RBM6	CXXC1	SIN3B	CEP164	WRAP53
DLG4	TRIM3	CCDC159	ENGASE	SH2B1	GFOD2	MRPL57
ATOH8	CCDC57	DTX3	ELP5	CRY2	FXYD6	CHD3
SGSM2	MCOLN1	PCGF3	FBXO44	EMC10	NEU3	CNPY2
NCAM1	PRR3	RABL6	PBXIP1	RANGRF	ZDHHC1	FIZ1
CEP250	DCUN1D2	TERF2IP	MAP1S	UCKL1	GBA2	KATNB1
B3GNT8	SCAMP5	NINL	CAMKK1	SIRT3	SLC30A4	STARD3NL
SLC2A8	MADD	SLC16A13	NT5C3B	FLCN	EPC1	SHC2
TMEM86B	PTCH2	APBA3	ASPCR1	SLC29A4	PLEKHB1	SDSL
NAIF1	VPS16	MAP3K12	CIRBP	TMOD2	MEGF8	KDM2B
SEC61A2	PCED1A	PEBP1	CDK10	ALDH9A1	LUC7L3	PAF1
CCDC28B	LLGL2	EXOC3	FAM120B	AP2A2	ZZEF1	R3HCC1
PHF10	SAT2	ASB1	MIF4GD	ARFGAP1	ELMO2	VPS18
CLSTN3	MOCS1	ATP8B2	WDR19	ATP5D	RGL2	TMEM59L
ATP6V0D1	ANKRD39	ING5	METTL3	ELMO1	SLC25A27	TPGS1
BCAS3	ATP1A3	MYBBP1A	KRBA1	ACBD4	ANKRD54	IRX2
PI4KB	SENP3	STK11IP	RAI2	TATDN2	GPR137	TSPAN33
KCTD2	RNF216	STAT5B	MINK1	FBXW4	SUOX	FKRP
CHST12	MTG2	LIN37	MAGED2	DCXR	LRRC56	NPM2
GIGYF1	CTNS	IZUMO4	SLC25A45	DCAF8	GPATCH3	SREBF1
MXD4	CTC1	TSPAN7	FAAP20	PLCG1	RABEP1	KCTD13
SLC26A11	IFFO1	DPY19L3	SLC25A29	ASCL2	PLD3	IQSEC1
SLC16A11	TNRC6C	PGPEP1	SLC38A10	TFEB	MED22	HELQ
SOBP	UNK	GTPBP6	PACS2	TAOK2	PRKACA	KLHDC3
VPS53	PPP3CB	TNS2	CENPT	FYN	POLR3F	TRAPPC12
ZMAT1	GKAP1	ARHGEF11	TANGO2	DIRAS1	CLPP	PTOV1
ABHD17A	UPF3A	TMEM220	SMARCA1	MAN1C1	DPH7	HDAC10
MBD3	SLC23A2	SLC22A17	METTL16	FAM193A	PAPSS2	SPHK2
GPRASP1	NMNAT3	KDM8	SGSH	EGLN2	AXIN2	USP27X
DPH1	ARMCX1	ITGA7	PPP1R9B	RFNG	AKT1S1	TDRP
CRTC1	PGS1	TAF1C	TBCB	FAM222A	POU6F1	ZFAND2B
UBOX5	FBXL16	ZBTB49	SEPHS1	INTS1	KBTBD7	FAM83F
APBB1	LRRC75A	ZSWIM1	TFPT	GGA1	SSTR2	RASIP1
FLYWCH2	GPS2	ZSWIM7	TMEM8B	NCKAP5L	PKNOX1	LYRM9
EXOG	AHI1	CFD	NPDC1	MRPL34	IMMP1L	GTF3C1
ARID3A	RAB11B	SLC25A14	ATP6V0B	CCDC96	KLC1	ATOX1
TMEM104	CYB5D2	LRRC14	PIN1	SORBS3	PGBD1	AGFG2
ARAP1	CAPN10	CTDNEP1	KCTD7	LSM10	APBB3	SGK3
SAP30L	PMPCA	FAM117A	ERCC2	NAAA	WBP1	CISH
ENPP2	RUFY3	INPP5E	MON1A	WNT4	IP6K1	NDUFA3
VPS37D	UNC119	FBXO9	WDR81	P3H3	TRMU	PABPN1
CDIP1	IQCC	CPT1C	ABCB8	SPG7	ACADVL	EFCC1
TRAF1	DPP7	DUSP28	TRIM25	RFX1	TMEM198	ROGDI
MARK1	TSPYL4	UQCR11	CSAD	PDZRN3	NAGLU	TIMM13
PNPLA7	ORAI2	CYB561D2	ACADSB	DISP1	PPP1R26	WIPI2
GPR173	U2AF1L4	YIF1B	MAP6	SMIM4	ZDHHC14	STXBP1
NEURL4	TXLNA	NISCH	RGAG4	STIM2	UBXN6	PALM
TRAF3	NRIP2	RNASEK	AGER	RSAD1	MAST3	ATG4D
EBF4	LRWD1	TOM1L2	SLC25A4	CPLX1	RXRA	CHGB
CASKIN2	FBXO31	TRO	TIMM22	TSC2	PIAS4	CHMP6
GABBR1	DMPK	ZCWPW1	MTERF4	MTG1	ATG16L2	SMPD2
TSEN54	FAM69B	ACACB	LMBR1L	MTMR4	SCAP	
CD99L2	MTA1	KLHDC1	ZSCAN2	IFT88	MST1	
HSD17B14	GABARAP	ZFP57	CALY	CLCN5	TBXA2R	
PDZD4	CSNK1D	ANKRD16	KAT2A	RPAIN	GRIK5	

**Supplementary Table S9.** Genes being significantly upregulated in p53wt but not in p53mut cells upon EZH2 knockdown.

<i>Anxa6</i>	<i>Dtna</i>	<i>Igf2bp3</i>	<i>Maged2</i>	<i>Pbbp</i>	<i>Slfn2</i>	<i>Tpm2</i>
<i>Camk2b</i>	<i>Efemp2</i>	<i>Igf2r</i>	<i>Map1lc3a</i>	<i>Rdh10</i>	<i>Spon2</i>	<i>Trp53inp2</i>
<i>Cdkn2a</i>	<i>Fhdc1</i>	<i>Itgb5</i>	<i>Masp1</i>	<i>Rnf130</i>	<i>Sprr1a</i>	<i>Tuft1</i>
<i>Ddah2</i>	<i>Foxg1</i>	<i>Ltbp1</i>	<i>Neat1</i>	<i>Serpib6b</i>	<i>Tbc1d16</i>	<i>Wnt7b</i>
<i>Dnaaf9</i>	<i>Fzd6</i>	<i>Ltbp3</i>	<i>Palld</i>	<i>Slc44a2</i>	<i>Tcf24</i>	<i>Zfp62</i>