

The POU-Domain Transcription Factor Oct-6/POU3F1 as a Regulator of Cellular Response to Genotoxic Stress

Cinzia Fionda, Danilo Di Bona, Andrea Kosta, Helena Stabile, Angela Santoni and Marco Cippitelli

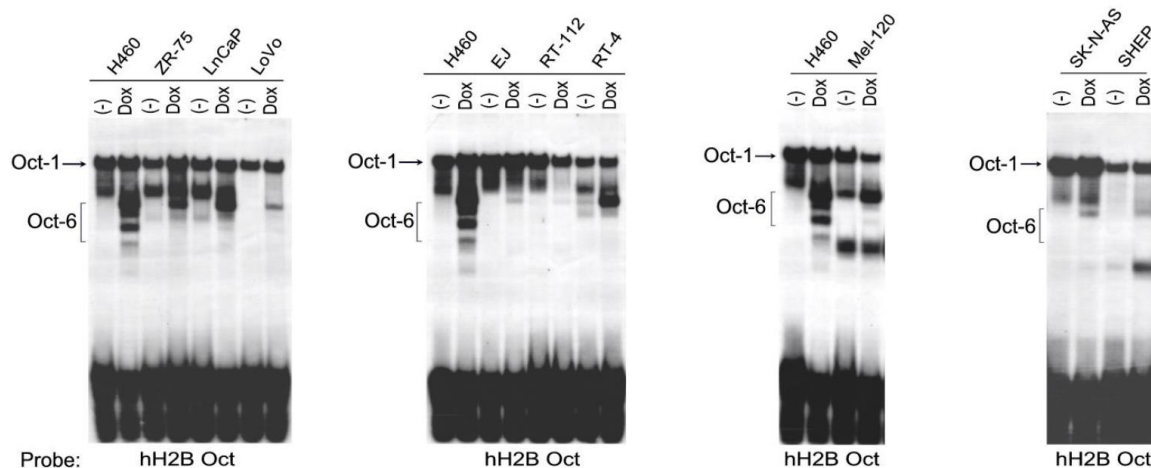


Figure S1. Doxorubicin induces Oct-6 DNA binding in different cancer cell lines. EMSA was performed using the hH2B-Oct probe in the presence of nuclear extracts (10 μ g) from H460 cells (positive control for each panel) or from the indicated cell line, unstimulated (-) or treated with Dox (0.5 μ M) for 24 h. DNA binding complexes with electrophoretic mobility corresponding to Oct-1 or Oct-6 are indicated in the figures.

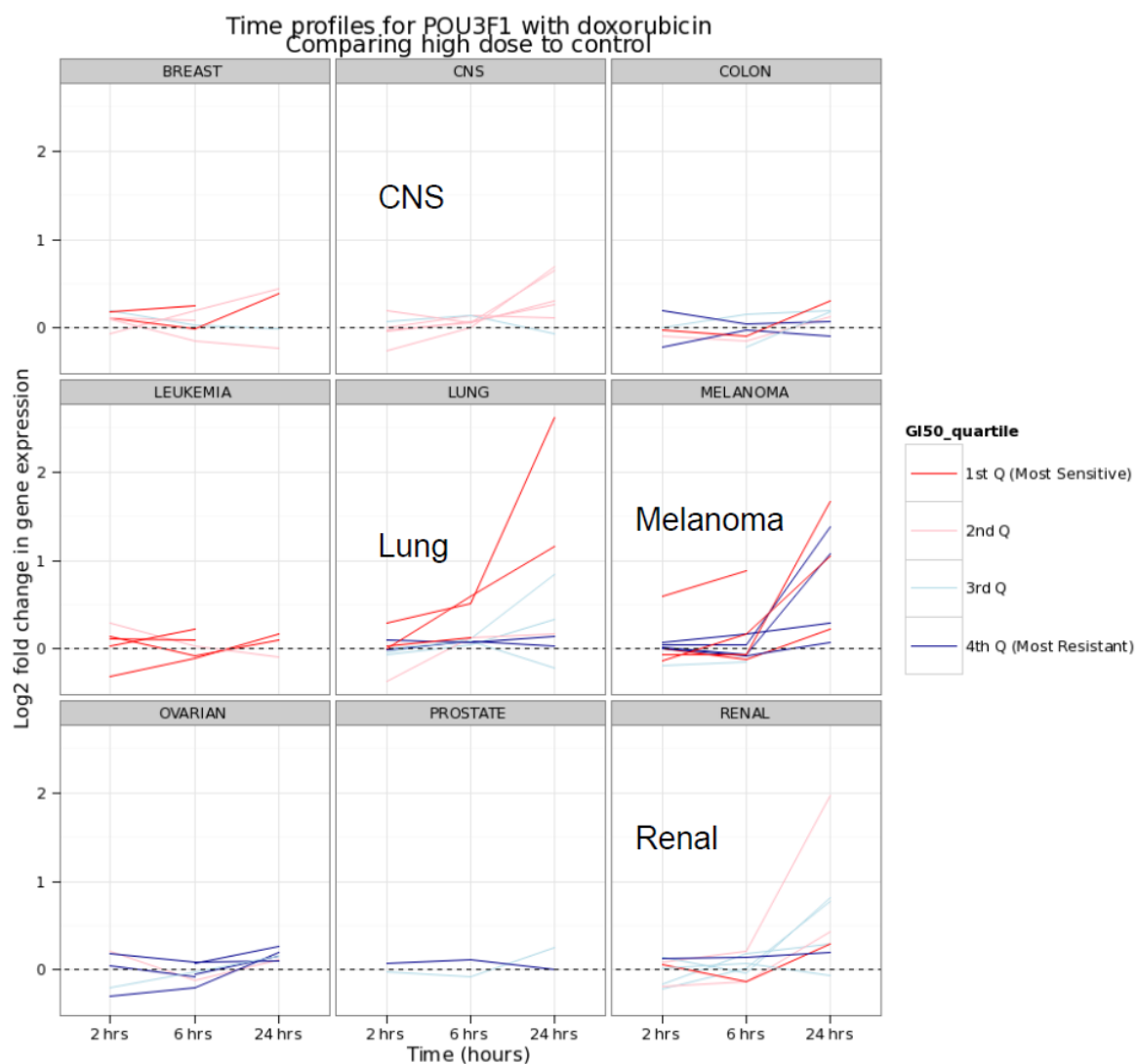


Figure S2. Time profiles for Oct-6 gene expression in different cell line lineages. Data shown in the picture were obtained using the interactive NCI Transcriptional Pharmacodynamics Workbench (NCI TPW) (<https://tpwb.nci.nih.gov/GeneExpressionNCI60/index.html>).

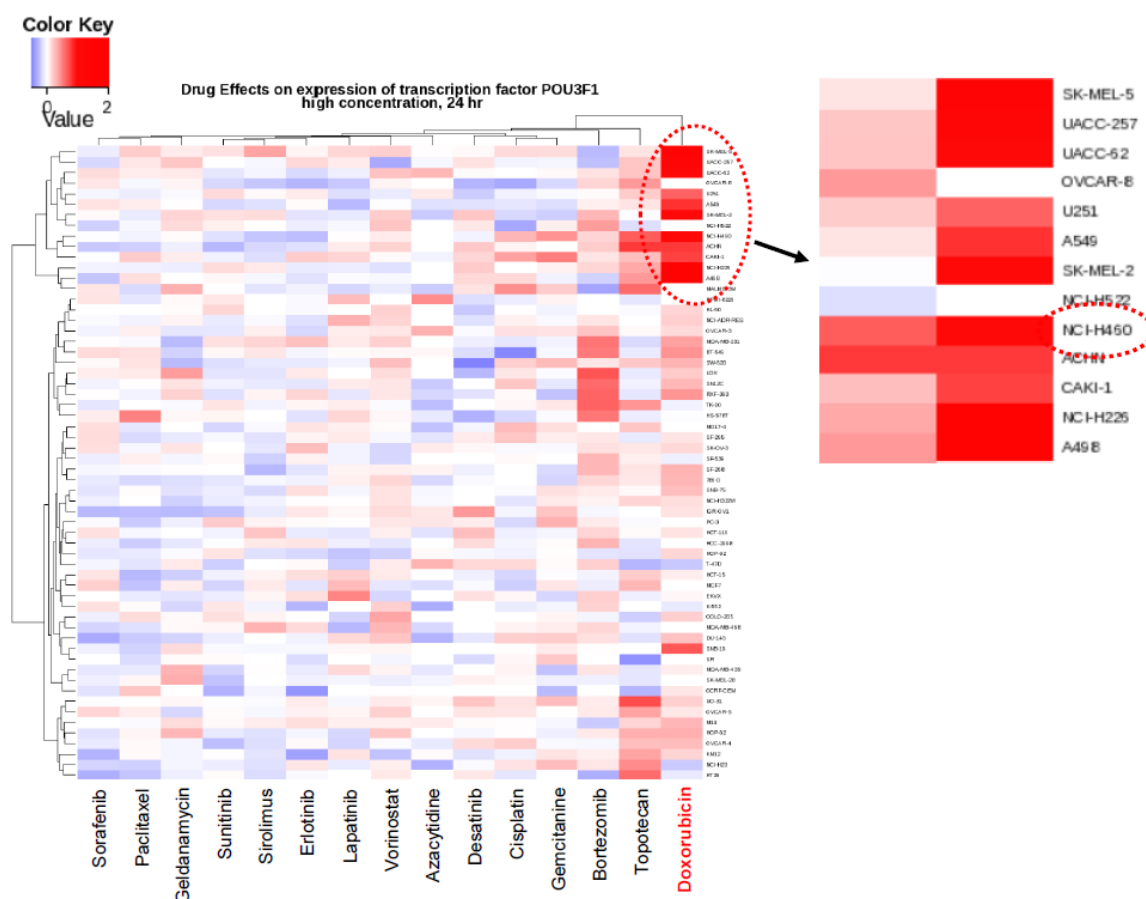


Figure S3. Heatmap of drug effects on expression of the transcription factor Oct-6: profiles in different cell line lineages treated with Dox. Expression of Oct-6 in the presence of 1 μ M Dox/24 h was evaluated in selected human cancer cell lines as indicated in the red circles. NCI Transcriptional Pharmacodynamics Workbench. In the red circle, heatmap of drug effects on expression of a specific transcription factor (POU3F1/Oct-6) on selected human cancer cell lines available at (<https://tpwb.nci.nih.gov/GeneExpressionNCI60/index.html>).

GSE116441

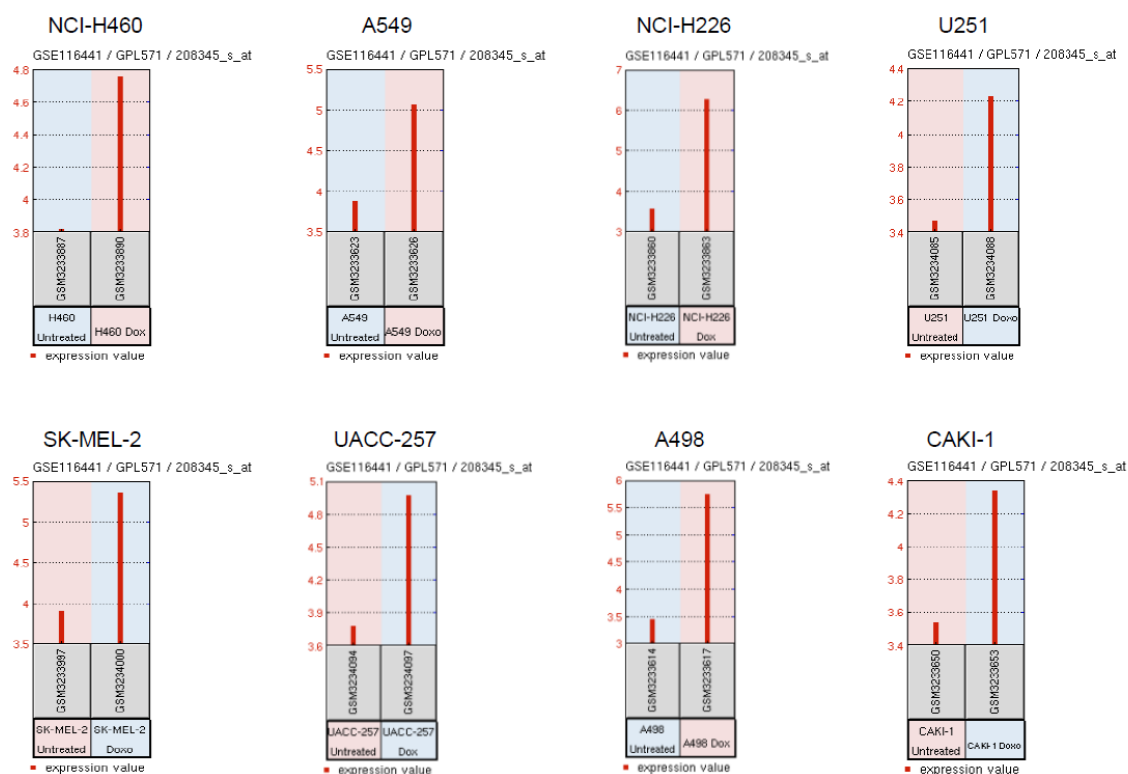


Figure S4. GEO2R analysis of the microarray public dataset GSE116441 available at (<http://www.ncbi.nlm.nih.gov/geo/>) for Oct-6 expression in selected cancer cell lines [(lung H460, A549, H226)–(melanoma SK-MEL-2, UACC-257)–(renal A498, CAKI-1)–(glioblastoma U251)] after treatment with Dox 1 μ M for 24 h.

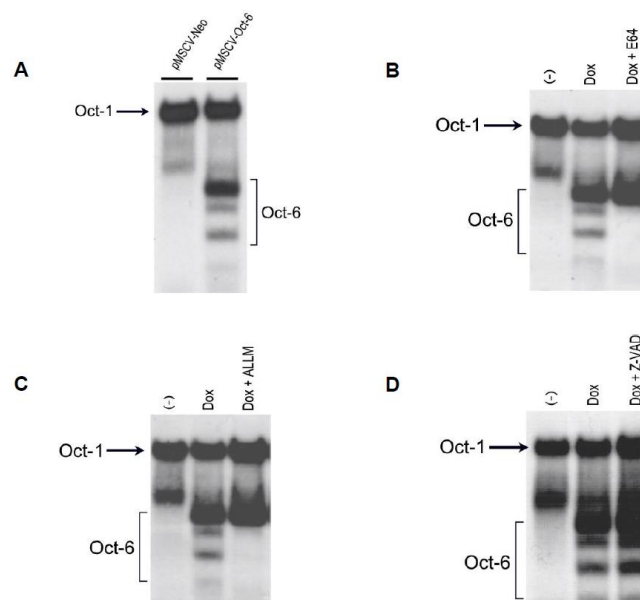


Figure S5. Characterization of Dox induced Oct-6 binding complexes. (A) EMSA showing the constitutive expression of Oct-6/Tst-1 in H460 cells stably transduced with the retroviral expression vector pMscv-Oct-6 or the control vector pMscv-Neo. (B–D) EMSA was performed using the 32 P-labeled hH2B-Oct probe in the presence of nuclear extracts (10 μ g) from unstimulated (–) or Dox treated H460 cells (24 h) in the absence or in the presence of E64 (100 μ M), ALLM (25 μ M) or Z-VAD (10 μ M). DNA binding complexes containing Oct-1 or Oct-6 are indicated in the figure. Data are representative of two independent experiments.

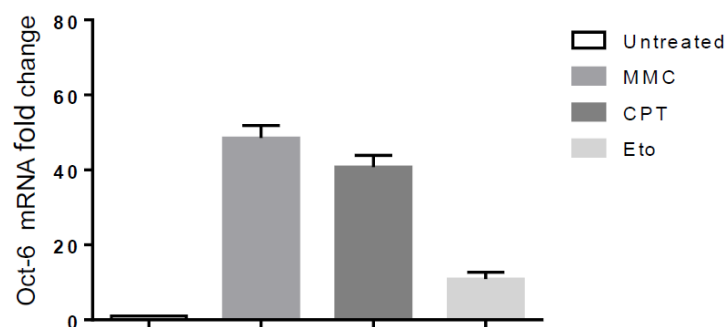


Figure S6. Different genotoxic drugs induce Oct-6 mRNA expression in H460 cells. Real Time PCR analysis of total mRNA obtained from H460 cells, untreated or treated with Mitomycin C 10 μ M (MMC), Cisplatin 5 μ g/ml (CPT) or Etoposide 10 μ M (Eto) for 24 h. Data, expressed as fold change units, were normalized with GAPDH. A representative experiment of two is shown.

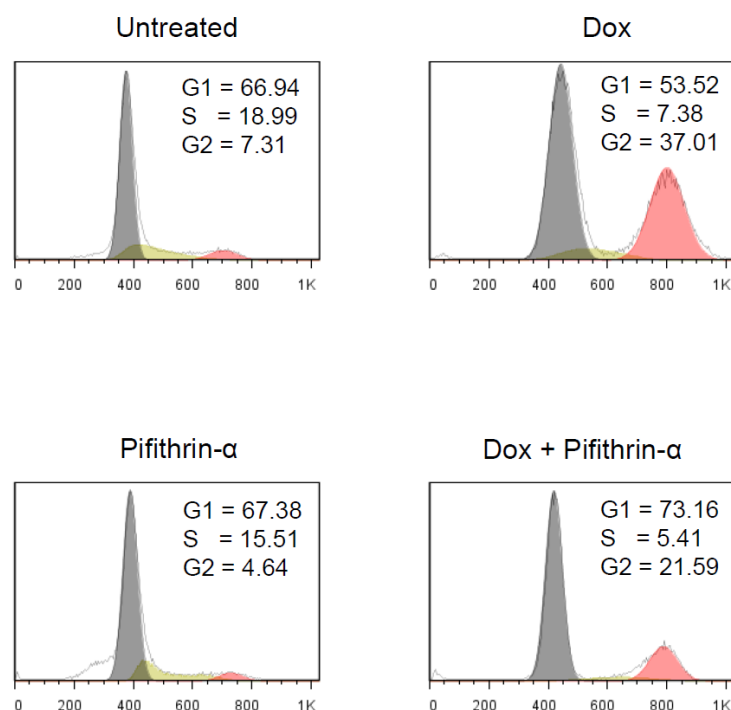


Figure S7. Cell-cycle analysis of H460 cells treated with Dox in the presence of Pifithrin- α . H460 cells, left untreated (-) or treated with Dox 0.5 μ M in the presence or in the absence of Pifithrin- α (30 μ M) for 24 h, were fixed and stained with PI to analyze cell distribution among the different cell cycle phases. A representative experiment is shown.

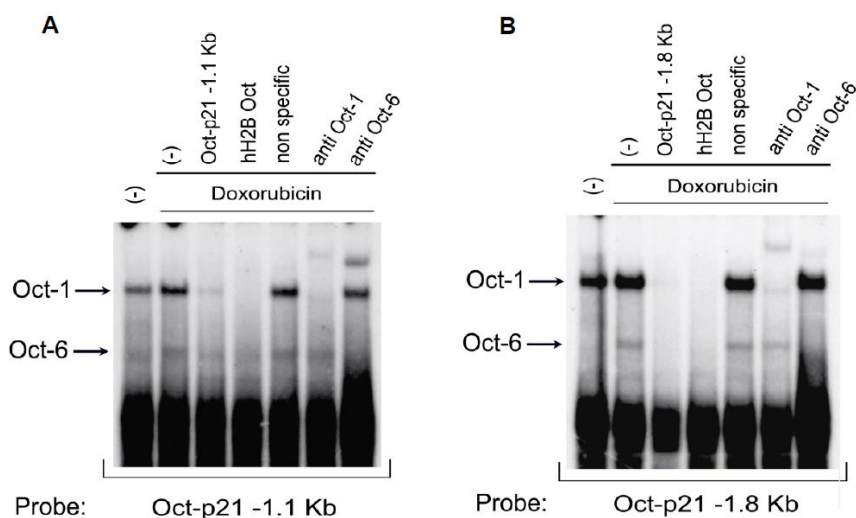


Figure S8. Oct-6 regulates p21 expression: direct binding to p21 promoter. EMSA was performed using two different consensus Octamer sequences from p21 promoter [1.1 Kb (left panel) (A) and -1.8 kb (right panel) (B)] as probe, in the presence of nuclear extracts (10 µg) from unstimulated (-) or Dox treated H460 cells for 24 h. Where indicated, 100 ng of specific consensus Oct or non-specific cold competitor was added to the reaction mixture to confirm specificity. Where indicated, anti-Oct-1 or anti-Oct-6 antibody was added to the reaction mixture. DNA binding complexes containing Oct-1 or Oct-6 are indicated in the figures. EMSA shown in the figure are representative of 3 independent experiments, all displaying similar results.



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