

Supplementary Materials

Isolation, Characterization and Biological Action of Type-1 Ribosome-Inactivating Proteins from Tissues of *Salsola soda* L.

Nicola Landi, Sara Ragucci, Lucía Citores, Angela Clemente, Hafiza Z. F. Hussain, Rosario Iglesias, José M. Ferreras and Antimo Di Maro

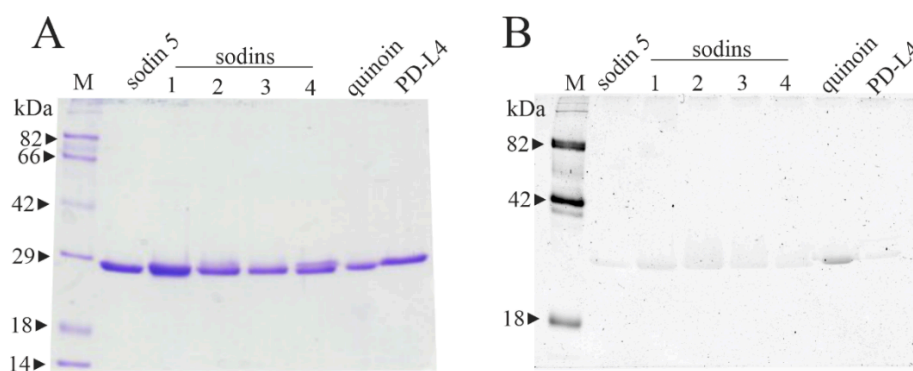


Figure S1. (A) SDS-PAGE analysis (B) and in gel staining for sugars of sodins, isolated from *S. soda* tissues. 3.0 μg (0.1 nmole) of sodin 5 (major form from *S. soda* seeds), sodins 1-4 (lanes 1, 2, 3 and 4), quinoioin (N-glycosylated type-1 RIP from *C. quinoa* seeds) and PD-L4 (non-glycosylated type-1 RIP from *P. dioica* leaves) were loaded, respectively. M, Coomassie blue (left panel) and CandyCane™ glycoproteins (right panel) molecular weight standards, respectively. SDS-PAGE was carried out on a 12% polyacrylamide separating gel in the presence of β -mercaptoethanol, followed by in-gel glycan detection using the Pro-Q Emerald 300 glycoprotein staining kit. Stained glycoproteins were visualised by UV trans-illumination.

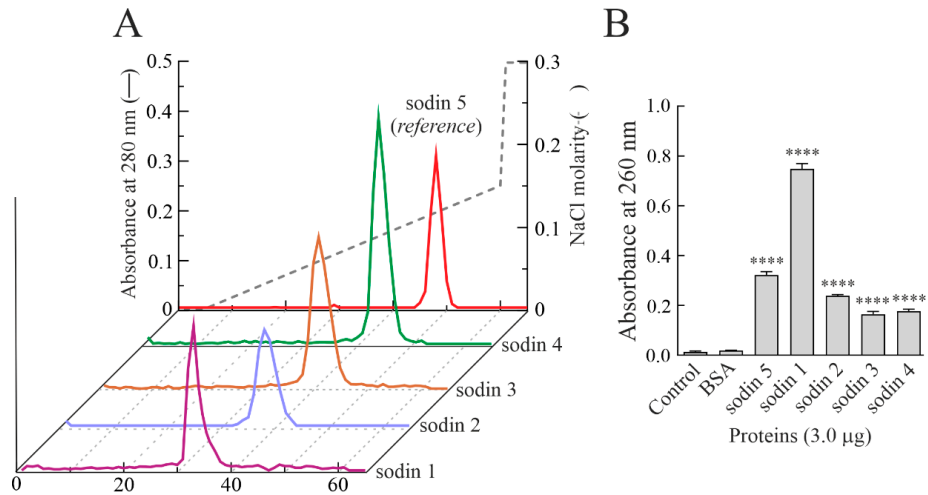


Figure S2. Protein purification and polynucleotide:adenosine glycosylase activity of sodins 1-4. **(A)** Elution profile after re-chromatography FPLC on an AKTA Purifier System from cation exchange chromatography using a Source 15S PE 4.6/100 column, showing peaks 1-4 (sodins 1-4) with the ability to release the β -fragment. The elution profile of sodin 5 was reported as reference. **(B)** Polynucleotide:adenosine glycosylase activity of BSA (negative control) or sodins 1-5 type-1 RIPs. Proteins (3.0 μ g) were assayed on salmon sperm DNA as described in Materials and methods. The mean results \pm SD of three experiments performed in triplicate are reported. Data were compared to control and analysed by One-way ANOVA with Dunnett's post hoc test (****, $p < 0.0001$).

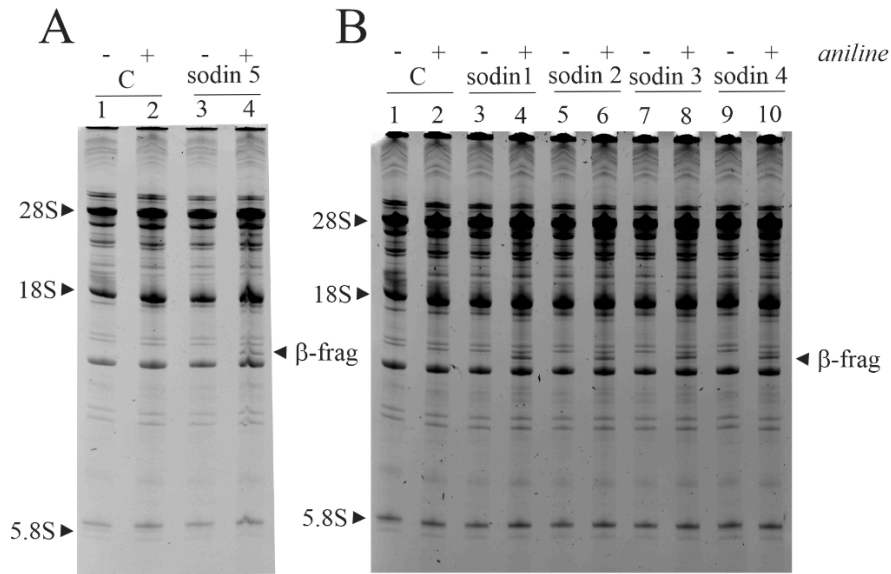


Figure S3. rRNA N-glycosylase activity of sodins from *S. soda* seeds assayed on rabbit ribosomes. Ribosomes were incubated with (A) 3.0 μ g of type-1 RIP sodin 5, (lanes 3 and 4) as positive control and (B) 3.0 μ g of sodins 1-4 (lanes 3-10, with or without aniline treatment). Following incubation, rRNA was extracted, treated with acid aniline and separated as reported in Materials and methods section. (+) and (-) indicate with and without aniline treatment. ' β -frag' indicates the position of Endo's fragment released by aniline treatment of rRNA from rabbit ribosomes.

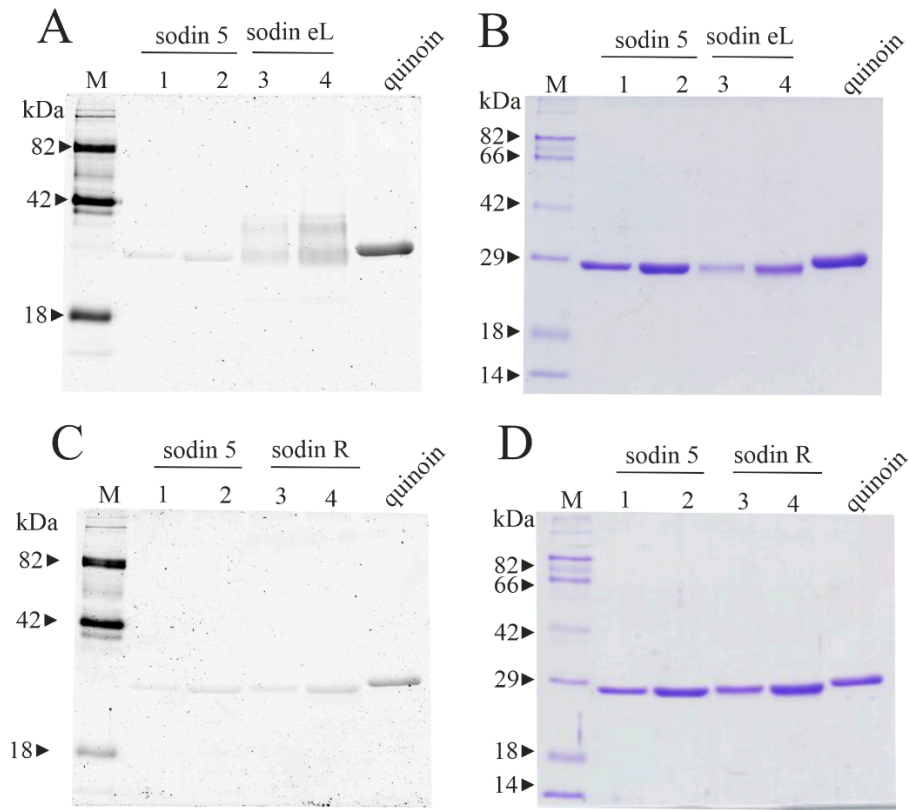


Figure S4. Staining for sugars of sodins isolated from *S. soda* tissues after SDS-PAGE. (A) and (B) 1.5 and 3.0 μg (0.05 and 0.1 nmole, respectively) of sodin 5 (lanes 1 and 2, respectively) and sodin eL (type-1 RIP from *S. soda* edible leaves; lanes 3 and 4, respectively) as well as 3.0 μg (0.1 nmole) of quinoin were loaded, respectively. (C) and (D) 1.5 and 3.0 μg (0.05 and 0.1 nmole, respectively) of sodin 5 (lanes 1 and 2, respectively) and sodin R (type-1 RIP from *S. soda* roots; lanes 3 and 4, respectively) as well as 3.0 μg (0.1 nmole) of quinoin were loaded, respectively. M, CandyCane™ glycoproteins (left panels) and Coomassie blue (right panels) molecular weight standards, respectively. SDS-PAGE was carried out on a 12% polyacrylamide separating gel in the presence of β -mercaptoethanol, followed by in-gel glycan detection using the Pro-Q Emerald 300 glycoprotein staining kit. Stained glycoproteins were visualised by UV trans-illumination.