

Review

Plugging the Gaps in the Global PhenoCam Monitoring of Forests—The Need for a PhenoCam Network across Indian Forests

Karun Jose ¹, Rajiv Kumar Chaturvedi ^{1,*} , Chockalingam Jeganathan ² , Mukunda Dev Behera ³ 
and Chandra Prakash Singh ⁴ 

¹ Department of Humanities and Social Sciences, BITS Pilani K K Birla Goa Campus, Vasco da Gama 403726, India; p20200063@goa.bits-pilani.ac.in

² Department of Remote Sensing, Birla Institute of Technology (BIT), Mesra, Ranchi 835215, India

³ Centre for Oceans, Rivers, Atmosphere and Land Sciences (CORAL), Indian Institute of Technology (IIT) Kharagpur, Kharagpur 721302, India

⁴ Space Applications Centre, Ahmedabad 380015, India

* Correspondence: rajivc@goa.bits-pilani.ac.in

Abstract: Our understanding of the impact of climate change on forests is constrained by a lack of long-term phenological monitoring. It is generally carried out via (1) ground observations, (2) satellite-based remote sensing, and (3) near-surface remote sensing (e.g., PhenoCams, unmanned aerial vehicles, etc.). Ground-based observations are limited by space, time, funds, and human observer bias. Satellite-based phenological monitoring does not carry these limitations; however, it is generally associated with larger uncertainties due to atmospheric noise, land cover mixing, and the modifiable area unit problem. In this context, near-surface remote sensing technologies, e.g., PhenoCam, emerge as a promising alternative complementing ground and satellite-based observations. Ground-based phenological observations generally record the following key parameters: leaves (bud stage, mature, abscission), flowers (bud stage, anthesis, abscission), and fruit (bud stage, maturation, and abscission). This review suggests that most of these nine parameters can be recorded using PhenoCam with >90% accuracy. Currently, Phenocameras are situated in the US, Europe, and East Asia, with a stark paucity over Africa, South America, Central, South-East, and South Asia. There is a need to expand PhenoCam monitoring in underrepresented regions, especially in the tropics, to better understand global forest dynamics as well as the impact of global change on forest ecosystems. Here, we spotlight India and discuss the need for a new PhenoCam network covering the diversity of Indian forests and its possible applications in forest management at a local level.

Keywords: phenology; Indian PhenoCam; forest management; long term monitoring



Citation: Jose, K.; Chaturvedi, R.K.; Jeganathan, C.; Behera, M.D.; Singh, C.P. Plugging the Gaps in the Global PhenoCam Monitoring of Forests—The Need for a PhenoCam Network across Indian Forests. *Remote Sens.* **2023**, *15*, 5642. <https://doi.org/10.3390/rs15245642>

Academic Editors: Iñigo Molina, Jan Komarek and Marlena Kycko

Received: 22 August 2023

Revised: 11 November 2023

Accepted: 14 November 2023

Published: 06 December 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Phenology as an Indicator of the Impact of Climate Change on Forests

Phenology is the science that deals with the study of recurrent events in the biological lifecycle and how these lifecycle events are influenced by temporal changes in biotic and abiotic factors [1]. Plant phenology indicates significant seasonal events, such as sprouting, leaf development, flowering, fruiting, and leaf fall in a plant's life cycle, which influences ecosystem functions via controlling energy and carbon through interactions between the atmosphere and plant communities [2–4]. The climate is the major controlling factor (temperature, water availability, and day length), although non-climatic parameters, such as altitude, slope, shadings, soil properties, and nutrient availability, may also affect plant phenology [5,6]. The responsiveness of phenology to the annual variability in weather parameters serves as an essential indicator of climate change in terrestrial ecosystems [2,7]. Multiple studies across the globe have confirmed the impact of climate change on plant phenology, such as advanced spring greenup [8,9], delayed senescence [10,11], and a shift

in flowering and fruiting [12]. The shift in phenological timing has affected atmosphere–vegetation interactions, the land surface energy balance, vegetation productivity, the water cycle, and the global carbon cycle [2,13]. The increasing length of the growing season (LOS) has generally resulted in an increased uptake of CO₂, potentially increasing the annual net primary productivity (NPP) [14]. However, the NPP also shows a negative trend with an increasing LOS due to increasing temperature during the growing season, resulting in increased respiration [15]. A longer LOS and increasing air temperature may also affect plants' transpiration rates, which influences the global water cycle [16]. In addition, the shift in phenological parameters can also affect agriculture, human health, and forestry. For example, on a global scale, approximately 27% of cereal crop regions have experienced a longer LOS (2.3 days per year) since 1981, affecting food production in these areas [17]. Similarly, the increasing LOS has also affected processes related to the phenology of wood formation [18]. Thus, monitoring the phenological dynamics of vegetation is important for understanding the behavioral responses of the Earth's ecosystems to future climate change.

2. Methods for Monitoring Phenology

Phenological monitoring methods can be classified into three types based on the spatial coverage of monitoring as follows: the species level, plot-regional level, and regional-global level (Figure 1). A human observer has traditionally monitored individual organisms to visually document changes in forest phenological parameters [2]. This method of collecting plant phenological data is known for its long temporal resolutions, enabling the availability of data from the 1900s in multiple locations worldwide [19]. However, it is worth noting that the spatial distribution of these data is currently limited to a few countries, primarily including the USA (National Phenology Network, Budburst, and National Ecological Observatory Network), Canada (Nature Watch), Sweden (Swedish National Phenology Network), Russia (Chronicles of Nature), Australia (Climate Watch), Pan European Phenology database (PEP725) and the UK (Nature's Calendar). Despite these efforts, ground-based phenological observations face certain limitations, such as being constrained to a limited number of species, being more prone to human errors, requiring significant labor, and being time-consuming [20,21].

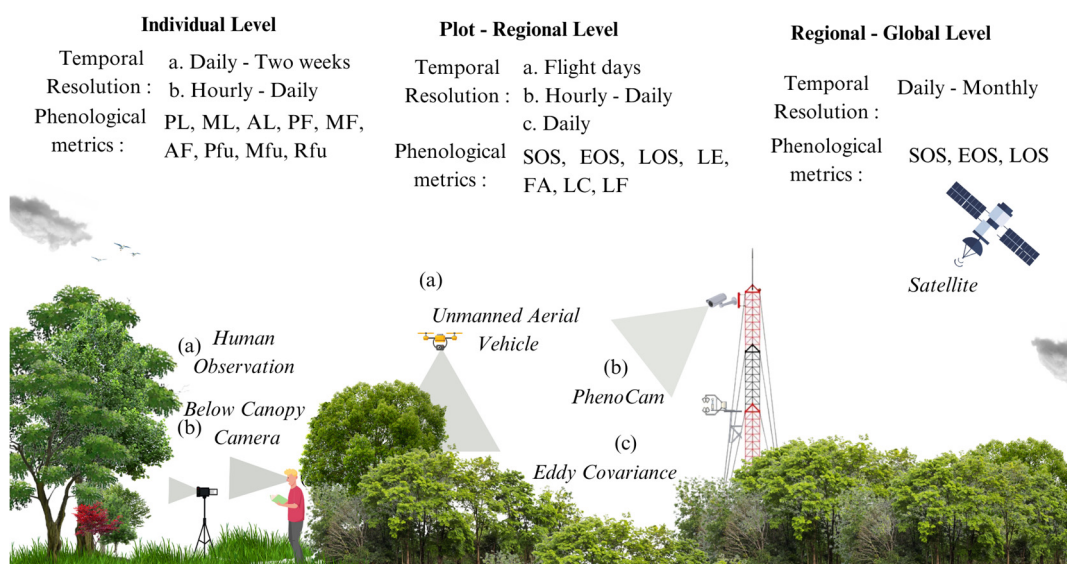


Figure 1. Overview of different methods of phenological monitoring. (Individual level: PL: production of young leaves, ML: maturation of leaves, AL: abscission of leaves, PF: production of young flowers, MF: maturation of flowers, AF: abscission of flowers, Pfu: production of young fruits, Mfu: maturation of fruits, Rfu: ripening of fruits; plot-regional level: SOS: start of season, EOS: end of season, LOS: length of season, LE: leaf expansion, FA: flower appearance, LC: leaf coloring, LF: leaf fall).

Since the 1970s, satellite-based remote sensing has introduced new possibilities for worldwide phenological surveillance at the landscape scale with the advantage of continuously collecting data for large areas. Land surface phenology (LSP) is the term generally used to represent vegetation phenology derived using satellite-based remote sensing estimated from vegetation indices (VI) or biophysical variables [22,23]. Multiple studies have confirmed the shift in phenological timing at a regional and global scale due to a shift in climatic conditions, such as changes in precipitation, temperature patterns, and a shift in snowfall timings [4,7]. Using satellite data, the phenological study at a global scale has confirmed a significant advancement in the SOS by 0.38 days per year, a delay in the EOS by 0.45 days per year, and a global increase in the LOS by 0.8 days per year [24]. Similarly, several studies have also reported a shifting phenological period at the regional and continental levels [10,22]. However, the practical application of satellite-based remote sensing in phenological monitoring faces several constraints, including low temporal and spatial resolution, susceptibility to cloud cover, and the modifiable area unit problem (MAUP), where the pixel being monitored may not correspond precisely to the ground location at different times [25–27]. Additionally, while comparing ground observations and satellite-based monitoring, the major difference is the footprint mismatch. Ground observations record phenology at the individual species level, which makes it difficult to compare it with a satellite-based phenological product at the regional scale. This pronounced disparity in footprint size underscores the need for near-surface monitoring techniques that are capable of capturing phenological data at the plot level.

It is interesting to note that the first fully digital camera came into existence after a decade of the first operational earth observation satellite. In recent years, satellite-based monitoring techniques have been complemented by near-surface remote sensing methods, such as time-lapse digital cameras (PhenoCam) [28], in situ measurements taken with radiometric instruments [29], monitoring based on eddy covariance (EC) data [30], and images captured by unmanned aerial vehicles (UAV—temporal resolution depends on the frequency of the flight) [31]. These near-surface remote sensing techniques offer high temporal resolution and better spatial coverage compared to direct ground observations. They provide valuable data that can be used to develop, calibrate, and evaluate satellite-based phenological models and products. Among these four near-surface phenology monitoring techniques, PhenoCam networks have garnered significant attention in the global research community due to their simplicity in data processing and the automated continuous data acquisition capacity at high spatial and temporal (timings pre-programmed) resolutions.

3. PhenoCam as a Promising Technology for Long-Term and Continuous Phenological Monitoring

The PhenoCam system tracks vegetation phenology at a high spatial and temporal resolution using images from digital cameras mounted on a tall tower [32] or small towers situated at a vantage point. This innovative technique allows for the monitoring of vegetation phenology at both individual and local area levels with minimal atmospheric errors. Researchers can analyze the captured images to identify data patterns or anomalies, facilitating a deeper understanding of the vegetation dynamics [33] (Figure 2).

The PhenoCam system captures images in red, green, and blue (RGB) and, optionally, in infrared (IR) bands. Since these images are captured under open atmospheric conditions, fluctuations in brightness due to cloud cover and the time of day can influence the quality of data. However, these effects can be mitigated by calibrating the images using a reference panel kept in the field of view [34]. Data generated from PhenoCam imagery have been utilized for various purposes, such as to assess satellite phenological products [35–37]; to define and validate new phenological models [38]; to better understand the interactions between canopy phenology and the dynamics of ecosystems [39]; and to study the seasonal changes in leaf-level physiological processes that are connected to variations in leaf color [40,41].

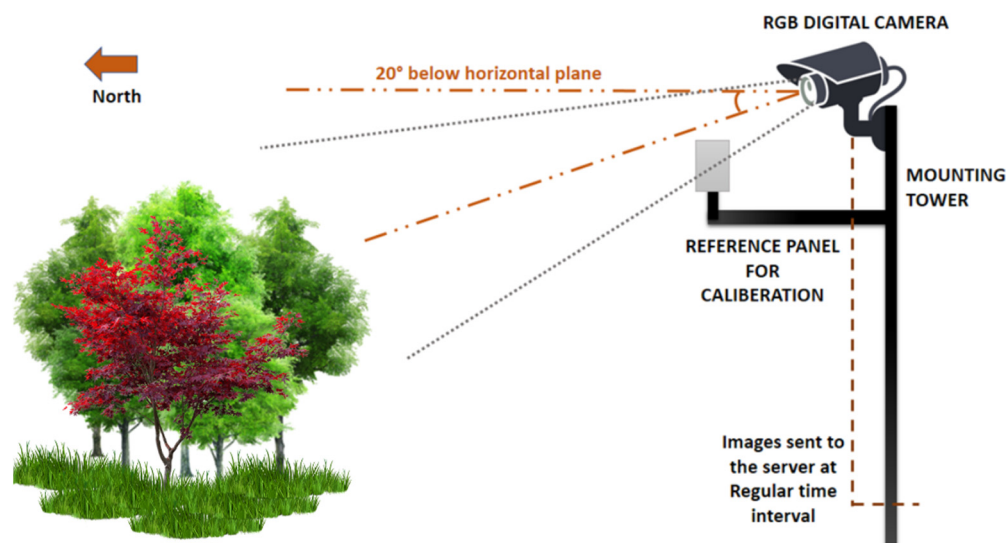


Figure 2. A representation of different components of the PhenoCam system.

In this review, we systematically compared observations from PhenoCam to satellite-based phenological products, the eddy covariance tower, and the ground observation of different ecosystems for which data are available in the literature. The comparison between PhenoCam and various other methods in terms of their performance, as reported in the existing literature, is depicted in Figure 3. The figure presents the correlation coefficients (r values) collected from relevant studies, providing a visual representation of the comparative performance of these methods for phenological monitoring. All the literature on global PhenoCam work referenced in this review is included in Table S1.

First, we compared PhenoCam-derived time series green chromatic coordinates (GCC) with MODIS and VIIRS satellite time series data (NDVI) (low-resolution satellites) (Figure 3a). In deciduous forests, the average correlation was found to be 0.87, while in grasslands, it was 0.85 ($n = 12$). The evergreen forest showed the lowest correlation with a value of 0.7 ($n = 7$). We also examined the correlation with moderate-resolution satellites such as Landsat and Sentinel. In grasslands, PhenoCam data demonstrated an average correlation of 0.91 ($n = 9$), while in deciduous forests, it was approximately 0.87 ($n = 4$). Furthermore (Figure 3b), we evaluated the correlation between extracting SOS and EOS from MODIS data and PhenoCam observations ($n = 12$). In the deciduous forest, the average correlation for SOS was 0.89, and for EOS, it was 0.81. In grasslands, the correlation between MODIS and PhenoCam for SOS was 0.94, while for EOS, it was 0.70. These studies show that Deciduous forests and Grasslands exhibit a stronger correlation with satellite data, primarily because they vividly display phenological stages. By contrast, Evergreen forests show a relatively lower correlation, which is mainly attributed to the absence of distinct phenological stages and the limitations in temporal resolution provided by satellite data.

Next, we compared PhenoCam time series GCC with gross primary productivity (GPP)-based phenological monitoring using the eddy covariance tower across different ecosystems (Figure 3c). Among the ecosystems studied, grasslands exhibited the highest correlation, with an average R-value of 0.91 ($n = 8$), followed by deciduous forests with an average R-value of 0.87 ($n = 11$), and evergreen forests with an R-value of 0.84 ($n = 5$). Lastly, we compared ground observation data with PhenoCam-derived GCC for various phenological metrics. In the deciduous forest, the correlation for budburst was found to be 0.96 ($n = 4$); for the leaf area index (LAI), it was 0.93 ($n = 2$); for the chlorophyll concentration, it was 0.95; and for leaf fall, it was 0.96 ($n = 2$) (Figure 3d).

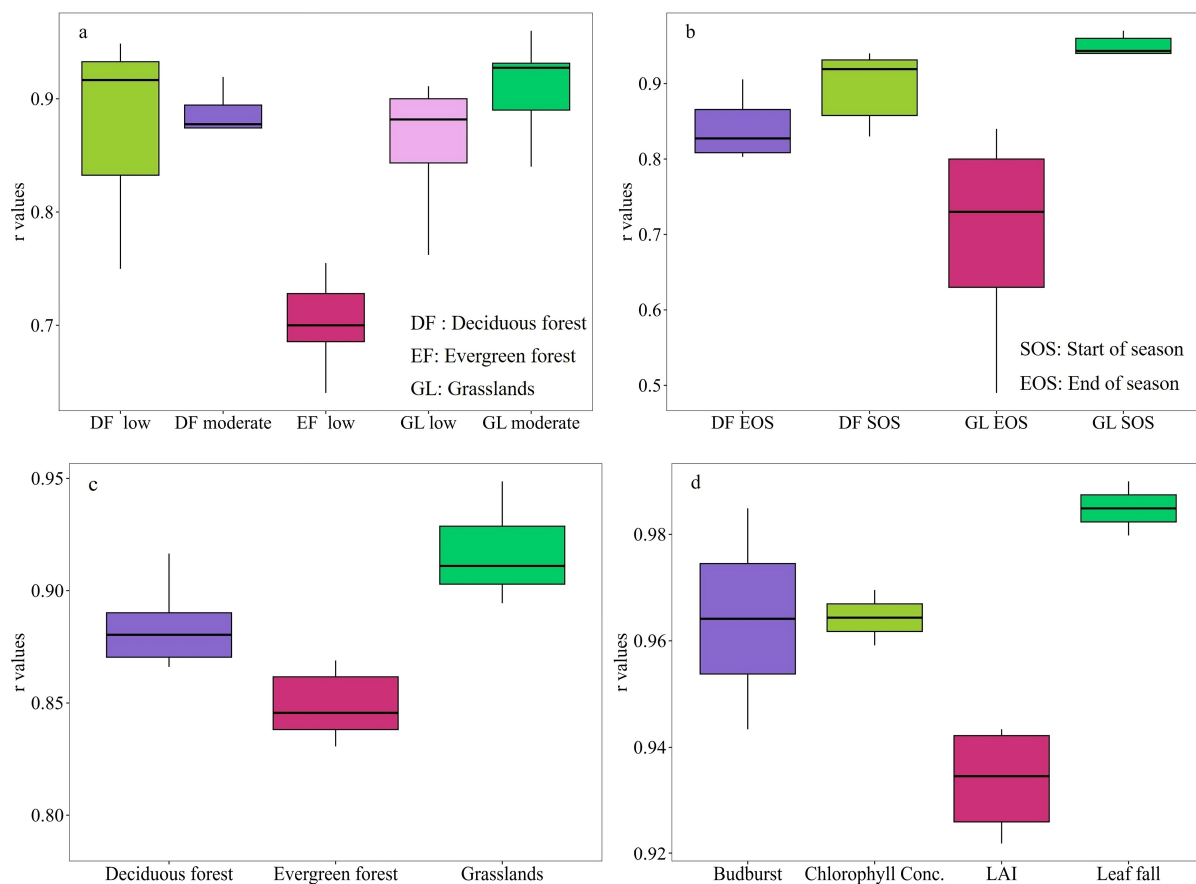


Figure 3. Correlation between PhenoCam data with other phenological monitoring methods, (a) PhenoCam to satellite-based NDVI, (b) PhenoCam-derived SOS and EOS to MODIS SOS and EOS, (c) PhenoCam GCC to eddy covariance gross primary productivity data and (d) PhenoCam to ground observations.

Traditionally, the observation of flowering and fruiting phenophases relied on labor-intensive direct ground observations. However, the introduction of the PhenoCam network has revolutionized this process, allowing for the more detailed monitoring of these phenophases. For instance, Chandra et al. (2022) utilized the PhenoCam network to monitor different stages of flowering (budding, initiation, and maturation) and fruiting (fruit development and maturation) phenology in *Rhododendron arboreum* Sm., located in the Himalayas. Their findings reveal a strong correlation between these phenophases and winter soil temperature [42]. Additionally, Mann et al. (2022) developed an automated method using time-lapse cameras to accurately extract flowering phenology (onset, peak, and end of flowering) for two Arctic species, achieving a high accuracy rate of over 90 percent compared to ground observation [43]. Also, Delpierre et al. (2020) reported a strong correlation ($r = 0.98$) between the within-population variability of budburst (WPVbb) calculated using PhenoCam data and ground observations. This supports the reliability of PhenoCam-derived metrics in capturing fine-scale phenological variations [44]. In addition to this, Kurc & Benton et al. (2010) employed linear regression analysis to demonstrate that greenness (GCC) is closely associated with soil moisture at depths ranging from 0.30 m to 1 m, which corresponds to the densest distribution of creosote bush roots [45]. This finding reinforces the link between PhenoCam-derived observations and ecological factors. Moreover, in the context of evergreen forests, where the correlation between PhenoCam-based observations compared to satellite phenological products tends to be low, Liu et al. (2020) discovered that red chromatic coordinates (RCC) offer a more precise prediction of GPP-based SOS and EOS [46]. This indicates the potential for an improved phenological assessment in evergreen forest ecosystems through the use of RCC data. These

studies highlight the potential of PhenoCam data, in combination with satellite data, to accurately monitor phenological changes and bridge the gap between satellite observations and real-world ecological dynamics.

However, RCC, GCC, and NDVI may not reveal the same timings of SOS or EOS as the physical processes associated with these indices are different. RCC and GCC reveal normalized energy from the red and green band from the PhenoCam RGB images, and they reveal that chlorophyll absorbed and reflected energy, hence revealing more about the chlorophyll pigment content in plants. On the other hand, NDVI uses red and NIR bands, and hence, it reveals more about the biomass as well as the health of the plant. During the senescence, the NIR value depends more on fiber content (intercellular arrangements of cells and pigment concentration, respectively), and the red reflection increases due to the absence of chlorophyll absorption and, hence, the magnitude of the NDVI value becomes low as the difference between NIR and RED is diminished [41,46]. Due to these reasons, RCC and GCC may show an early SOS, but NDVI may show a delayed SOS. Additionally, NDVI and other satellite-derived indices are influenced more by the soil background, atmospheric conditions, topography, etc., while GCC and RCC are relatively free (or negligible) from these influences. GCC and RCC could better demarcate leaf phenology, including SOS/EOS stages, while NDVI and other indices represent more plant growth and vigor with a better correlation of photosynthetic variables such as FAPAR/PAR [47,48]. This justifies the use of PhenoCam images for (leaf) phenological studies.

The comparison of ground-based observations with PhenoCam data consistently demonstrates a strong correlation (90%) across various ecosystems. The versatility of PhenoCam technology allows for the focused monitoring of specific species or multiple species, enabling the visual identification of various stages of plant life cycles, such as leafing (leaf buds, mature leaves, abscission of leaves), flowering (flower buds, anthesis of flowers, abscission of flowers), and fruiting (fruit buds, maturation of fruits, abscission of fruits). This review highlights the potential of PhenoCam as a promising alternative to labor-intensive ground observations. By utilizing PhenoCam cameras, researchers and environmentalists can streamline their data collection processes and contribute to a more comprehensive understanding of ecosystem dynamics. PhenoCam cameras, by replacing labor-intensive ground observation methods, represent a valuable tool for advancing ecological research and conservation efforts in poorer countries in the tropics.

4. Global Distribution of PhenoCams

PhenoCam is a viable alternative to the present method of ecosystem phenological monitoring that combines ground-based data with satellite observations [49]. Time-lapse and fixed digital cameras have demonstrated remarkable potential at capturing phenological metrics across a diverse range of biomes. These include high Arctic vegetation [50], North American grasslands [51], Amazon rainforests [52], grasslands in Japan [53], Australian grasslands [33], deciduous and evergreen forests of North America [54], tropical forests of Malaysia [55], and croplands [56]. In this review, we looked at only peer-reviewed articles that had the word “PhenoCam” in their titles or keywords. We carefully read each article and collected important information from them (see Table S1). From this list of 72 studies, we observed that a significant proportion of studies using PhenoCam were conducted in deciduous forests (31.5%), followed by grasslands (19.8%), evergreen forests (17.1%), and shrublands (8.1%) (Figure 4). This diverse distribution of research sites highlights the versatility and broad applicability of PhenoCam in different ecosystem types.

The recognition of the significance of PhenoCam systems in monitoring plant phenology has led to the establishment of several phenological networks worldwide. These networks include the PhenoCam Network—USA [32]—the Phenological Eyes Network—Japan [57]—the National Ecological Observatory Network—USA [58]—the EURO-Pheno Network—Europe [59]—the European Integrated Carbon Observation System (ICOS) Network—Europe [59]—and the Australian PhenoCam Network—Australia [60] (Table S2). When considering the spatial distribution of PhenoCam studies, it is notable that 53 percent

of the studies were from North America, followed by 28 percent from Europe and 11 percent from East Asia. This highlights a significant gap in PhenoCam-based monitoring in regions such as Africa, South America, South Asia, Central Asia, and Australia. There is a need to expand the PhenoCam network in these geographies to better understand their global forest dynamics and the impact of global change on forest ecosystems [61].

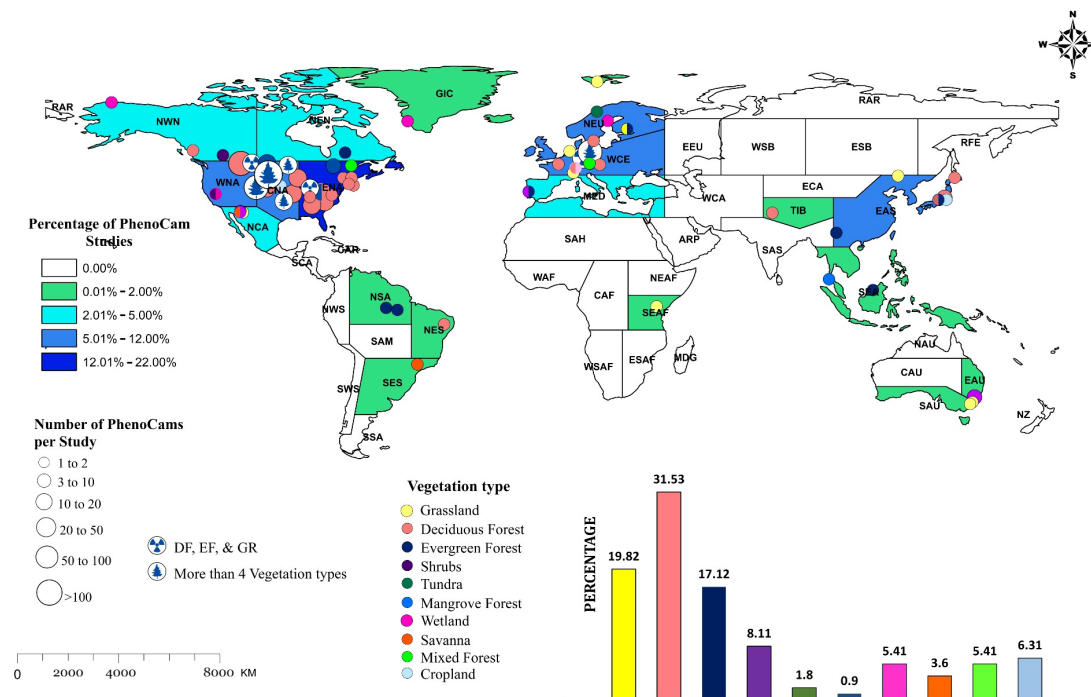


Figure 4. Distribution of published PhenoCam-based studies as per IPCC AR6 regions.

Here, we spotlight India as a case study and discuss the need for a PhenoCam network in India and its potential role in improving forest management in this country. The experiences from the Indian PhenoCam network could be useful in the expansion of PhenoCam networks in geographies with similar socio-economic circumstances, such as the other countries of South Asia, Africa, etc. By expanding the reach of PhenoCam networks to currently underrepresented regions, we can enhance the comprehensive monitoring of vegetation phenology worldwide. This could enable a more holistic perspective on the impact of climate change and contribute to a deeper understanding of ecological processes on a global scale.

5. The State of Phenological Monitoring in India

India is a country that has enormous diversity and endemism of plant species, with a total forest cover of 713,789 sq. km, which is about 21.71% of its total geographical area [62]. According to Champion and Seth, Indian forests have been classified into 16 different forest types and 255 sub-forest types [63]. Each of these forest and sub-forest types exhibits a distinct phenological behavior, necessitating enhanced phenological monitoring in Indian forests. Currently, forest phenological monitoring in India is mainly carried out using traditional ground observations and satellite remote-sensing techniques [20,64,65]. Ground-based phenological studies mainly focused on deciduous forests (tropical moist deciduous forests and tropical dry deciduous forests) and evergreen forests (tropical wet evergreen forests) [66–69] and a few on tropical and subtropical moist forests in the north-east of India [70–72] (Figure 5). The first-ever formal (from the available literature) phenological study in India was conducted from 15 April 1975 to 20 December 1976 and from 14 February 1978 to 15 September 1979 in the dry deciduous forests of Bandipur [64].

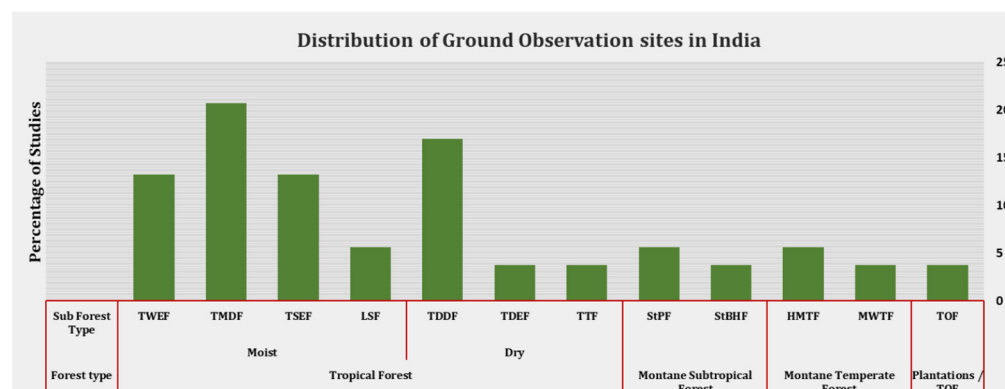


Figure 5. The distribution of ground phenology monitoring sites in India from published journals since 1975.

However, only a few studies have been published for phenological observations over a long period (more than five years), such as the study by Datta & Rane at Pakke Wildlife Sanctuary and Tiger Reserve, Arunachal Pradesh, which lasted six years from 1997 to 1999 and 2009 to 2011, and the study by Suresh & Sukumar at Mudumalai Dry Forests, Tamil Nadu, which lasted from 2000 to 2008 [73,74].

Even though India has 37 (Figure 6, Table S3) distinct ground-based phenology observation sites, it has been reported that there are only six long-term forest phenology monitoring sites in India (most of them are ongoing, and only a few are published): Mudumalai-Tamil Nadu (30 years), Pakke Tiger Reserve-Arunachal Pradesh (1997–2000 and 2009–ongoing), dry forest in Rishi Valley-Andhra Pradesh (10 years), Kalakkad Mundanthurai Tiger Reserve-Tamil Nadu (20+ years), Anamalai Hills-Tamil Nadu (4 years) and dry tropical forest in Nigdale-Maharashtra (7 years) [75]. In addition, India has a long-term, large-scale citizen science initiative for plant monitoring called SeasonWatch. Hitherto, SeasonWatch (<https://www.seasonwatch.in/>, accessed on 15 October 2023) has registered over 24,617 plants and collected over 6.6 lakh observations on the phenology of these trees.

In addition to this, several satellite-based phenological monitoring studies have been conducted since 1980 [76–78]. However, nationwide ground validation is not possible for satellite-based phenological products due to a lack of ground observations. The published literature provides information for ground validation in a few locations; however, it is difficult to identify the exact date of the year (DOY) from these linguistic distributions (such as early spring, the end of the dry period, and the post or pre-monsoon period). In addition, most of the ground-based phenological studies in India have monitored the major nine phenological stages in the cycle, such as leaves (bud stage, mature, abscission), flowers (bud stage, anthesis, abscission), and fruit (bud stage, maturation, and abscission). However, it is impossible to monitor all these nine phenological stages with a high level of accuracy using satellite data, which is one of the significant limitations of satellite remote sensing in phenology monitoring.

Scientists have employed near-surface digital cameras to circumvent these impediments for phenological monitoring. Sharma et al. (2021) installed a camera in the Western Himalayas to track *Betula*, utilize phenological temporal trends, and record snow cover patterns [79]. They effectively tracked the phases of greenup, leaf maturity, senescence, and dormancy and reported that greenup occurred four days earlier in 2018 than in 2017, whereas dormancy occurred one day later. Parihar et al. (2013) used digital cameras to monitor forest phenology in Madhav National Park in Madhya Pradesh and compared their findings to satellite data [80]. However, the use of digital cameras for phenology monitoring in India is limited, with studies often focusing on a restricted range of plant species and short monitoring periods.

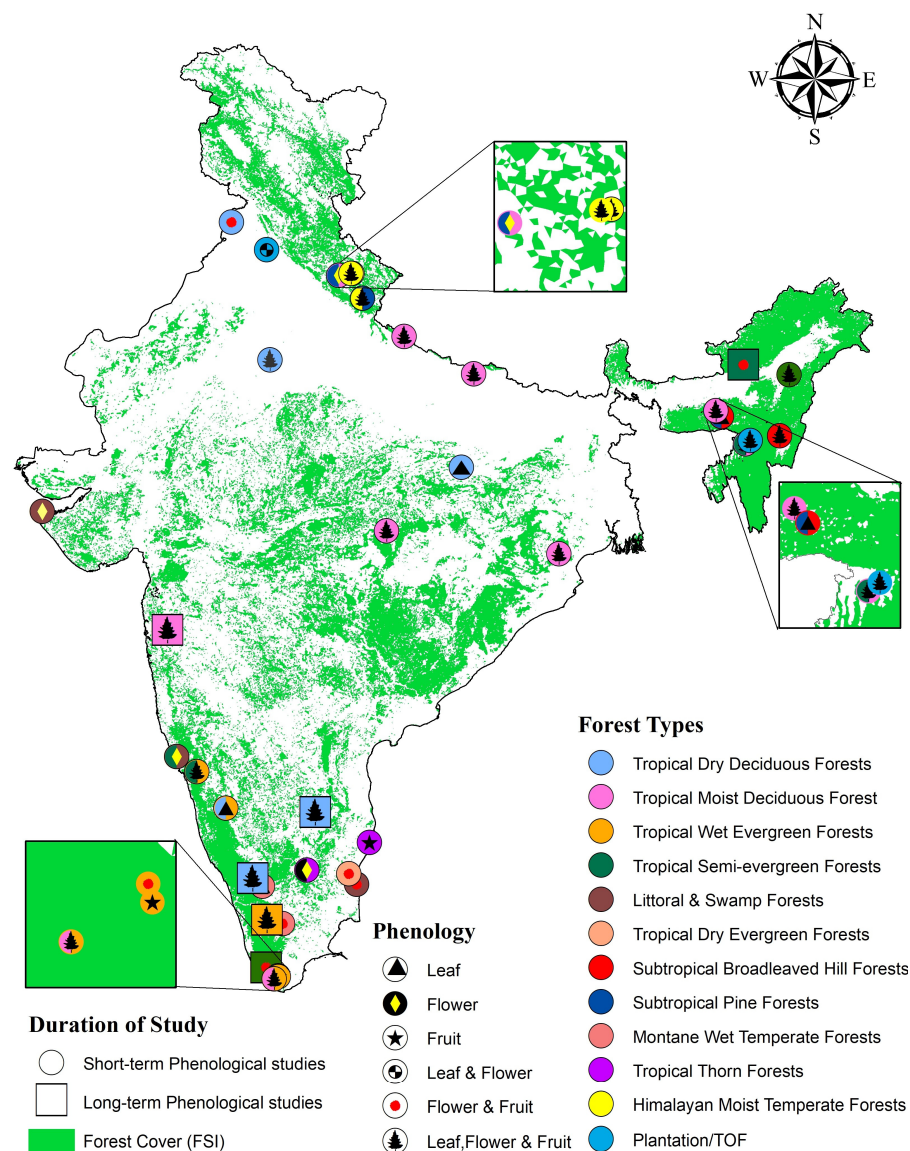


Figure 6. Ground-based forest phenology monitoring sites in India. (Based on the past literature since 1975. We identified 37 ground observation sites as follows: 31 short-term and six long-term).

It is crucial to establish a long-term phenological monitoring network to better understand the drivers and responses to climate change. Short-term studies of 3–5 years (the average period of most financed research projects or a university doctorate thesis) are profoundly inadequate to capture the intrinsic unpredictability of natural systems caused by environmental stochasticity, as evidenced by numerous recent studies conducted worldwide. Indeed, the results of such short-term research can be quite deceiving. Given the absence of a comprehensive long-term ground-based phenological monitoring system across India's diverse forest types and the limitations of satellite-based monitoring, there is a pressing need for a new monitoring network. Daily phenological observations using time-lapse digital cameras, such as PhenoCam, offer a solution to address these challenges. This approach provides high temporal and spatial resolutions, allowing for cost-effective, labor-efficient data collection, long-term monitoring, and reliable canopy phenological monitoring. Adopting digital repeat photography techniques can facilitate the establishment of an extensive and reliable phenological monitoring network in India [54,81,82].

6. Role of the PhenoCam Network in Supporting Forest Management in India

Long-term datasets on ecosystem dynamics are crucial for effective forest management and biodiversity conservation. Unfortunately, in the case of India, there is currently a limited number of long-term observation sites, and access to such datasets is restricted [83]. Introducing a PhenoCam network across various forest types in India could be a valuable addition to forest management efforts. PhenoCams are widely used globally to understand and predict changes in vegetation patterns and phenology, which have significant implications for ecosystem health and climate research.

Gross primary productivity is an important indicator of forest health as well as an indicator of climate change [84]. Wang et al. (2020) showed that GPP derived from PhenoCam shows a higher correlation (R^2 varied from 0.80 to 0.87) with GPP derived from an eddy covariance tower with high statistical significance ($p < 0.01$) [25]. Similarly, PhenoCam-based analysis (Excess green index and GCC) on plant health shows a significant relation to the ground-based metrics of stand health, which includes vigor, mortality, foliar disease occurrence, and root disease symptoms at both the plot and tree level [85]. Several studies have also indicated that camera-derived LAI and chlorophyll concentration correlate with ground-based observations [40,86,87]. Similarly, PhenoCam-derived GCC demonstrates a significant correlation with aboveground green biomass (AGB) in grassland ecosystems [53].

PhenoCam data have proven valuable in tracking global climate events. For instance, Cremonese et al. (2017) effectively utilized time series PhenoCam data from grasslands in the Western European Alps to monitor the impact of heat waves [7]. Similarly, Gonçalves et al. (2020) examined the response of forests to an unexplained drought triggered by El Niño in 2015–2016 and observed post-drought fluctuations in leaf demography using multi-year PhenoCam data [88]. In light of the changing climatic conditions, the introduction of PhenoCam in India's forests holds great promise for enhancing our understanding of ecosystem dynamics, facilitating informed decision-making, and supporting proactive conservation strategies.

In addition to direct monitoring, PhenoCam data can be also used to validate satellite-based products, which are highly used for forest management and decision making in the current scenario. Validating satellite-based phenological products is important for understanding variations in seasonal and inter-annual biome responses due to climatic variability [89]. The following are some of the most widely used satellite-based vegetation products to understand forest dynamics: (a) the Normalized Difference Vegetation Index (NDVI), (b) the Enhanced Vegetation Index (EVI), (c) MODIS LAI, and (d) MODIS and AVHRR Fraction of Absorbed Photosynthetic Active Radiation (fPAR). Currently, data from the eddy covariance tower (EC) [87,90], ground-based measurements of LAI, time-lapse digital photographs, and tower radiation sensors (PAR) are being used to calibrate these products. However, in India, the number of active eddy covariance sites is very limited. Therefore, the establishment of the Indian PhenoCam Network could significantly increase the number of calibration points for satellite-based products, addressing the current gap in phenological monitoring in the South Asian region. Additionally, this could help in validating satellite-derived phenological matrices (e.g., MCD12Q2) and proceed one step closer to qualifying phenological matrices with products from satellites to become one of the essential climate variables (ECVs).

Indian forest calendar for management. Presently, available information about the Indian forests is limited to broad and sporadic measurements (monthly observations in field plots). Such data are inadequate for generating spatial maps on phenology, health, biomass, and other essential datasets. The Forest Survey of India (FSI) conducts satellite-based mapping and monitoring, producing a national forest area assessment report every two years. However, it is insufficient to meet the needs of managing Indian forests effectively. To address this limitation and enhance forest management strategies, the establishment of a long-term PhenoCam network across various Indian forests is crucial. By implementing PhenoCams, which provide continuous and accurate data on shifting phenology, health

indicators, biomass estimation, and gross primary productivity, researchers can gain a deeper understanding of the ecological impacts of climate change on different species. Moreover, phenologically informed forest management is crucial for arresting increasing forest fire events in Indian forests, which see surface fires starting with the accumulation of litter during EOS (senescence). The predictive modeling of meteorological variables crucial for defining EOS is the need of the hour.

Improvements in the Dynamic Global Vegetation Models (DGVMs) and understanding the impact of climate change on forest ecosystems in India. Vegetative phenology holds significant importance in DGVMs. These models utilize information on plant biogeography and biogeochemical processes to predict ecosystem fluxes and climate fluctuations in the context of global change scenarios [91]. One crucial input for DGVMs is GPP, which is currently estimated using satellite data and still requires validation in the South Asian region. Recent studies have demonstrated the potential to estimate GPP [25] and LAI [86] using PhenoCam data. This indicates that PhenoCam technology can contribute to improving and strengthening satellite-derived GPP data, enhancing the accuracy of DGVM predictions, and making them more applicable to the unique conditions of the South Asian region. By integrating PhenoCam data into DGVMs, researchers can gain a better understanding of the impacts of climate change on forest ecosystems in India and refine management strategies accordingly.

7. Conclusions

Ground-based phenological observations, as well as PhenoCams, are mainly concentrated in developed and temperate regions, leading to a paucity of observations in the global south, especially across tropical forests. The expansion of long-term forest monitoring, especially the monitoring of phenology, is necessary for improving our understanding of the impact of global change on forest ecosystems globally. Ground-based phenological observations are constrained by a number of limitations, including high recurring costs. PhenoCam represents a cost-effective, efficient, and reliable alternative for ground-based phenological observations. We propose setting up a PhenoCam network covering different forest types in India. This network not only helps in filling a crucial data gap in the South Asian region, but it also provides a template for the expansion of the PhenoCam network in similar geographies across the tropics.

The establishment of an Indian PhenoCam network has the potential to support forest management in India. By providing continuous and accurate data on phenology, health indicators, biomass estimation, and gross primary productivity, this network can enhance our understanding of ecosystem dynamics and conservation strategies and enable informed decision making. Through the integration of PhenoCam data with Dynamic Global Vegetation Models, we can improve future predictions at a local level and refine management strategies in response to the impacts of climate change on forest ecosystems. The implementation of PhenoCam technology could be a valuable addition to India's forest management efforts, addressing the limitations of current monitoring approaches and contributing to the sustainable management of forest resources.

The PhenoCam network has already been successfully implemented in other countries, and it has shown promise in improving our understanding of forest ecosystems and their response to climate change. In India, the network could be particularly valuable for monitoring the impact of heatwaves and droughts on ecosystems, which are becoming increasingly common in the country. By providing detailed insights into how forests adapt to these challenges, this network could help us prioritize conservation efforts and ensure the long-term sustainability of India's forests.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/rs15245642/s1>, Table S1: PhenoCam studies world-wide [92–133]. Table S2: PhenoCam networks worldwide. Table S3: Indian Ground based forest phenology monitoring sites [134–164].

Author Contributions: R.K.C., C.P.S., M.D.B. and C.J. conceptualized the idea of the review. K.J. collected the data and carried out the systematic review with the help from R.K.C. and C.P.S. K.J. and R.K.C. wrote the draft manuscript. M.D.B., C.J. and C.P.S. reviewed multiple drafts and helped shape the final version. All authors have read and agreed to the published version of the manuscript.

Funding: We acknowledge the funding support from ISRO's Geosphere Biosphere Programme (IGBP) for collaborative research.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Lieth, H.H. Contributions to phenology seasonality research. *Int. J. Biometeorol.* **1976**, *20*, 197–199. [[CrossRef](#)]
- Richardson, A.D.; Keenan, T.F.; Migliavacca, M.; Ryu, Y.; Sonnentag, O.; Toomey, M. Climate change, phenology, and phenological control of vegetation feedbacks to the climate system. *Agric. For. Meteorol.* **2013**, *169*, 156–173. [[CrossRef](#)]
- Rihan, H.Z.; Kareem, F.; El-Mahrouk, M.E.; Fuller, M.P. Artificial seeds (principle, aspects and applications). *Agronomy* **2017**, *7*, 71. [[CrossRef](#)]
- Chaurasia, A.N.; Parmar, R.M.; Dave, M.G.; Mehta, N.; Kallaje, R.; Sahu, A.; Murthy, I.K.; Singh, C.P.; Krishnayya, N.S.R. Syncing phenology phase and canopy spectral reflectance of common tree species of four forest covers in India. *Curr. Sci.* **2021**, *120*, 567. [[CrossRef](#)]
- Nord, E.A.; Lynch, J.P. Plant phenology: A critical controller of soil resource acquisition. *J. Exp. Bot.* **2009**, *60*, 1927–1937. [[CrossRef](#)] [[PubMed](#)]
- Zhao, M.; Peng, C.; Xiang, W.; Deng, X.; Tian, D.; Zhou, X.; Yu, G.; He, H.; Zhao, Z. Plant phenological modeling and its application in global climate change research: Overview and future challenges. *Environ. Rev.* **2013**, *21*, 1–14. [[CrossRef](#)]
- Cremonese, E.; Filippa, G.; Galvagno, M.; Siniscalco, C.; Oddi, L.; di Cella, U.M.; Migliavacca, M. Heat wave hinders green wave: The impact of climate extreme on the phenology of a mountain grassland. *Agric. For. Meteorol.* **2017**, *247*, 320–330. [[CrossRef](#)]
- Beil, I.; Kreyling, J.; Meyer, C.; Lemcke, N.; Malyshev, A.V. Late to bed, late to rise—Warmer autumn temperatures delay spring phenology by delaying dormancy. *Glob. Chang. Biol.* **2021**, *27*, 5806–5817. [[CrossRef](#)]
- Meng, L.; Zhou, Y.; Gu, L.; Richardson, A.D.; Peñuelas, J.; Fu, Y.; Wang, Y.; Asrar, G.R.; De Boeck, H.J.; Mao, J.; et al. Photoperiod decelerates the advance of spring phenology of six deciduous tree species under climate warming. *Glob. Chang. Biol.* **2021**, *27*, 2914–2927. [[CrossRef](#)]
- Jeganathan, C.; Nishant, N. Scrutinising MODIS and GIMMS vegetation indices for extracting growth rhythm of natural vegetation in India. *J. Indian Soc. Remote Sens.* **2014**, *42*, 397–408. [[CrossRef](#)]
- Zohner, C.M.; Renner, S.S.; Sebald, V.; Crowther, T.W. How changes in spring and autumn phenology translate into growth-experimental evidence of asymmetric effects. *J. Ecol.* **2021**, *109*, 2717–2728. [[CrossRef](#)]
- Parker, V.T.; Stickrod, M.A. Reproductive phenological shifts and other phylogenetic trait changes in the Arbutioideae (Ericaceae) in the context of drought, seed predation, and fire. *Botany* **2022**, *100*, 387–399. [[CrossRef](#)]
- Caparros-Santiago, J.A.; Rodriguez-Galiano, V.; Dash, J. Land surface phenology as indicator of global terrestrial ecosystem dynamics: A systematic review. *ISPRS J. Photogramm. Remote Sens.* **2021**, *171*, 330–347. [[CrossRef](#)]
- Keenan, T.F.; Gray, J.; Friedl, M.A.; Toomey, M.; Bohrer, G.; Hollinger, D.Y.; Munger, J.W.; O'keefe, J.; Schmid, H.P.; Wing, I.S.; et al. Net carbon uptake has increased through warming-induced changes in temperate forest phenology. *Nat. Clim. Chang.* **2014**, *4*, 598–604. [[CrossRef](#)]
- Piao, S.; Ciais, P.; Friedlingstein, P.; Peylin, P.; Reichstein, M.; Luyssaert, S.; Margolis, H.; Fang, J.; Barr, A.; Chen, A.; et al. Net carbon dioxide losses of northern ecosystems in response to autumn warming. *Nature* **2008**, *451*, 49–52. [[CrossRef](#)] [[PubMed](#)]
- Workie, T.G.; Debella, H.J. Climate change and its effects on vegetation phenology across ecoregions of Ethiopia. *Glob. Ecol. Conserv.* **2018**, *13*, e00366. [[CrossRef](#)]
- Brown, M.E.; De Beurs, K.M.; Marshall, M. Global phenological response to climate change in crop areas using satellite remote sensing of vegetation, humidity and temperature over 26 years. *Remote Sens. Environ.* **2012**, *126*, 174–183. [[CrossRef](#)]
- Saderi, S.; Rathgeber, C.B.; Rozenberg, P.; Fournier, M. Phenology of Wood Formation in Larch (*Larix decidua* Mill.) Trees Growing Along a 1000-m elevation gradient in the French Southern Alps. *Ann. For. Sci.* **2019**, *76*, 89. [[CrossRef](#)]
- Ovaskainen, O.; Meyke, E.; Lo, C.; Tikhonov, G.; Delgado, M.d.M.; Roslin, T.; Gurarie, E.; Abadonova, M.; Abduraimov, O.; Adrianova, O.; et al. Chronicles of nature calendar, a long-term and large-scale multitaxon database on phenology. *Sci. Data* **2020**, *7*, 47. [[CrossRef](#)]
- Suresh, H.S.; Sukumar, R. Influence of climatic variability on tree phenology in the tropical dry forests of Mudumalai, southern India. *Trop. For. A Chang. World* **2009**, *5*, 39–61.
- Singh, S.L.; Sahoo, U.K. Shift in phenology of some dominant tree species due to climate change in Mizoram, North-East India. *Indian J. Ecol.* **2019**, *46*, 132–136.

22. Bolton, D.K.; Gray, J.M.; Melaas, E.K.; Moon, M.; Eklundh, L.; Friedl, M.A. Continental-scale land surface phenology from harmonized Landsat 8 and Sentinel-2 imagery. *Remote Sens. Environ.* **2020**, *240*, 111685. [\[CrossRef\]](#)
23. Bórnez, K.; Descals, A.; Verger, A.; Peñuelas, J. Land surface phenology from VEGETATION and PROBA-V data. *Assess. Over Deciduous For. Int. J. Appl. Earth Obs. Geoinf.* **2020**, *84*, 101974.
24. Julien, Y.; Sobrino, J.A. Global land surface phenology trends from GIMMS database. *Int. J. Remote Sens.* **2009**, *30*, 3495–3513. [\[CrossRef\]](#)
25. Wang, H.; Jia, G.; Epstein, H.E.; Zhao, H.; Zhang, A. Integrating a PhenoCam-derived vegetation index into a light use efficiency model to estimate daily gross primary production in a semi-arid grassland. *Agric. For. Meteorol.* **2020**, *288*, 107983. [\[CrossRef\]](#)
26. Mohapatra, J.; Singh, C.P.; Tripathi, O.P.; Pandya, H.A. Remote sensing of alpine treeline ecotone dynamics and phenology in Arunachal Pradesh Himalaya. *Int. J. Remote Sens.* **2019**, *40*, 7986–8009. [\[CrossRef\]](#)
27. Prakash Singh, C.; Mohapatra, J.; Pandya, H.A.; Gajmer, B.; Sharma, N.; Shrestha, D.G. Evaluating changes in treeline position and land surface phenology in Sikkim Himalaya. *Geocarto Int.* **2020**, *35*, 453–469. [\[CrossRef\]](#)
28. Seyednasrollah, B.; Young, A.M.; Hufkens, K.; Milliman, T.; Friedl, M.A.; Froking, S.; Richardson, A.D. Tracking vegetation phenology across diverse biomes using Version 2.0 of the PhenoCam Dataset. *Sci. Data* **2019**, *6*, 222. [\[CrossRef\]](#)
29. Liu, F.; Wang, X.; Wang, C. Measuring vegetation phenology with near-surface remote sensing in a temperate deciduous forest: Effects of sensor type and deployment. *Remote Sens.* **2019**, *11*, 1063. [\[CrossRef\]](#)
30. Gonsamo, A.; Chen, J.M.; Price, D.T.; Kurz, W.A.; Wu, C. Land surface phenology from optical satellite measurement and CO₂ eddy covariance technique. *J. Geophys. Res. Biogeosci.* **2012**, *117*. [\[CrossRef\]](#)
31. Berra, E.F.; Gaulton, R.; Barr, S. Assessing spring phenology of a temperate woodland: A multiscale comparison of ground, unmanned aerial vehicle and Landsat satellite observations. *Remote Sens. Environ.* **2019**, *223*, 229–242. [\[CrossRef\]](#)
32. Richardson, A.D.; Jenkins, J.P.; Braswell, B.H.; Hollinger, D.Y.; Ollinger, S.V.; Smith, M.-L. Use of digital webcam images to track spring green-up in a deciduous broadleaf forest. *Oecologia* **2007**, *152*, 323–334. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Watson, C.J.; Restrepo-Coupe, N.; Huete, A.R. Multi-scale phenology of temperate grasslands: Improving monitoring and management with near-surface phenocams. *Front. Environ. Sci.* **2019**, *7*, 14. [\[CrossRef\]](#)
34. Shajahan, S.; Cannayen, I.; Hendrickson, J. Monitoring plant phenology using phenocam: A review. In Proceedings of the 2016 ASABE Annual International Meeting, Orlando, FL, USA, 17–20 July 2016; American Society of Agricultural and Biological Engineers: St. Joseph, MI, USA, 2016; p. 1. [\[CrossRef\]](#)
35. Hufkens, K.; Friedl, M.; Sonnentag, O.; Braswell, B.H.; Milliman, T.; Richardson, A.D. Linking near-surface and satellite remote sensing measurements of deciduous broadleaf forest phenology. *Remote Sens. Environ.* **2012**, *117*, 307–321. [\[CrossRef\]](#)
36. Westergaard-Nielsen, A.; Lund, M.; Hansen, B.U.; Tamstorf, M.P. Camera derived vegetation greenness index as proxy for gross primary production in a low Arctic wetland area. *ISPRS J. Photogramm. Remote Sens.* **2013**, *86*, 89–99. [\[CrossRef\]](#)
37. Klosterman, S.T.; Hufkens, K.; Gray, J.M.; Melaas, E.; Sonnentag, O.; Lavine, I.; Mitchell, L.; Norman, R.; Friedl, M.A.; Richardson, A.D. Evaluating remote sensing of deciduous forest phenology at multiple spatial scales using PhenoCam imagery. *Biogeosciences* **2014**, *11*, 4305–4320. [\[CrossRef\]](#)
38. Hufkens, K.; Keenan, T.F.; Flanagan, L.B.; Scott, R.L.; Bernacchi, C.J.; Joo, E.; Brunsell, N.A.; Verfaillie, J.; Richardson, A.D. Productivity of North American grasslands is increased under future climate scenarios despite rising aridity. *Nat. Clim. Chang.* **2016**, *6*, 710–714. [\[CrossRef\]](#)
39. Toomey, M.; Friedl, M.A.; Froking, S.; Hufkens, K.; Klosterman, S.; Sonnentag, O.; Baldocchi, D.D.; Bernacchi, C.J.; Biraud, S.C.; Bohrer, G.; et al. Greenness indices from digital cameras predict the timing and seasonal dynamics of canopy-scale photosynthesis. *Ecol. Appl.* **2015**, *25*, 99–115. [\[CrossRef\]](#)
40. Keenan, T.; Darby, B.A.; Felts, E.; Sonnentag, O.; Friedl, M.A.; Hufkens, K.; O’Keefe, J.; Klosterman, S.; Munger, J.W.; Toomey, M.; et al. Tracking forest phenology and seasonal physiology using digital repeat photography: A critical assessment. *Ecol. Appl.* **2014**, *24*, 1478–1489. [\[CrossRef\]](#)
41. Richardson, A.D.; Hufkens, K.; Milliman, T.; Aubrecht, D.M.; Chen, M.; Gray, J.M.; Johnston, M.R.; Keenan, T.F.; Klosterman, S.T.; Kosmala, M.; et al. Tracking vegetation phenology across diverse North American biomes using PhenoCam imagery. *Sci. Data* **2018**, *5*, 180028. [\[CrossRef\]](#)
42. Chandra, S.; Singh, A.; Mathew, J.R.; Singh, C.P.; Pandya, M.R.; Bhattacharya, B.K.; Solanki, H.; Nautiyal, M.C.; Joshi, R. Phenocam observed flowering anomaly of *Rhododendron arboreum* Sm. in Himalaya: A climate change impact perspective. *Environ. Monit. Assess.* **2022**, *194*, 877. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Mann, H.M.; Iosifidis, A.; Jepsen, J.U.; Welker, J.M.; Loonen, M.J.; Høye, T.T. Automatic flower detection and phenology monitoring using time-lapse cameras and deep learning. *Remote Sens. Ecol. Conserv.* **2022**, *8*, 765–777. [\[CrossRef\]](#)
44. Delpierre, N.; Soudani, K.; Berveiller, D.; Dufrêne, E.; Hmimina, G.; Vincent, G. “Green pointillism”: Detecting the within-population variability of budburst in temperate deciduous trees with phenological cameras. *Int. J. Biometeorol.* **2020**, *64*, 663–670. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Kurc, S.A.; Benton, L.M. Digital image-derived greenness links deep soil moisture to carbon uptake in a creosotebush-dominated shrubland. *J. Arid Environ.* **2010**, *74*, 585–594. [\[CrossRef\]](#)
46. Liu, Y.; Wu, C.; Sonnentag, O.; Desai, A.R.; Wang, J. Using the red chromatic coordinate to characterize the phenology of forest canopy photosynthesis. *Agric. For. Meteorol.* **2020**, *285*, 107910. [\[CrossRef\]](#)

47. Gutman, G.; Skakun, S.; Gitelson, A. Revisiting the use of red and near-infrared reflectances in vegetation studies and numerical climate models. *Sci. Remote Sens.* **2021**, *4*, 100025. [\[CrossRef\]](#)
48. Moore, C.E.; Beringer, J.; Evans, B.; Hutley, L.B.; Tapper, N.J. Tree–grass phenology information improves light use efficiency modelling of gross primary productivity for an Australian tropical savanna. *Biogeosciences* **2017**, *14*, 111–129. [\[CrossRef\]](#)
49. Menzel, A.; Helm, R.; Zang, C. Patterns of late spring frost leaf damage and recovery in a European beech (*Fagus sylvatica* L.) stand in south-eastern Germany based on repeated digital photographs. *Front. Plant Sci.* **2015**, *6*, 110. [\[CrossRef\]](#)
50. Anderson, H.B.; Nilsen, L.; Tømmervik, H.; Karlsen, S.R.; Nagai, S.; Cooper, E.J. Using ordinary digital cameras in place of near-infrared sensors to derive vegetation indices for phenology studies of high arctic vegetation. *Remote Sens.* **2016**, *8*, 847. [\[CrossRef\]](#)
51. Cui, T.; Martz, L.; Lamb, E.G.; Zhao, L.; Guo, X. Comparison of grassland phenology derived from MODIS satellite and PhenoCam near-surface remote sensing in North America. *Can. J. Remote Sens.* **2019**, *45*, 707–722. [\[CrossRef\]](#)
52. Alberton, B.; Almeida, J.; Helm, R.; Torres, R.D.S.; Menzel, A.; Morellato, L.P.C. Using phenological cameras to track the green up in a cerrado savanna and its on-the-ground validation. *Ecol. Inform.* **2014**, *19*, 62–70. [\[CrossRef\]](#)
53. Inoue, T.; Nagai, S.; Kobayashi, H.; Koizumi, H. Utilization of ground-based digital photography for the evaluation of seasonal changes in the aboveground green biomass and foliage phenology in a grassland ecosystem. *Ecol. Inform.* **2015**, *25*, 1–9. [\[CrossRef\]](#)
54. Filippa, G.; Cremonese, E.; Migliavacca, M.; Galvagno, M.; Sonnentag, O.; Humphreys, E.; Hufkens, K.; Ryu, Y.; Verfaillie, J.; di Cella, U.M.; et al. NDVI derived from near-infrared-enabled digital cameras: Applicability across different plant functional types. *Agric. For. Meteorol.* **2018**, *249*, 275–285. [\[CrossRef\]](#)
55. Nagai, S.; Ichie, T.; Yoneyama, A.; Kobayashi, H.; Inoue, T.; Ishii, R.; Suzuki, R.; Itioka, T. Usability of time-lapse digital camera images to detect characteristics of tree phenology in a tropical rainforest. *Ecol. Inform.* **2016**, *32*, 91–106. [\[CrossRef\]](#)
56. Sakamoto, T.; Gitelson, A.A.; Nguy-Robertson, A.L.; Arkebauer, T.J.; Wardlow, B.D.; Suyker, A.E.; Verma, S.B.; Shibayama, M. An alternative method using digital cameras for continuous monitoring of crop status. *Agric. For. Meteorol.* **2012**, *154*, 113–126. [\[CrossRef\]](#)
57. Tsuchida, S.; Nishida, K.; Iwao, K.; Kawato, W.; Oguma, H.; Iwasaki, A. Phenological eyes network for validation of remote sensing data. *J. Remote Sens. Soc. Jpn.* **2005**, *25*, 282–288. [\[CrossRef\]](#)
58. Kampe, T.U.; Johnson, B.R.; Kuester, M.A.; Keller, M. NEON: The first continental-scale ecological observatory with airborne remote sensing of vegetation canopy biochemistry and structure. *J. Appl. Remote Sens.* **2010**, *4*, 043510. [\[CrossRef\]](#)
59. Wingate, L.; Ogée, J.; Cremonese, E.; Filippa, G.; Mizunuma, T.; Migliavacca, M.; Moisy, C.; Wilkinson, M.; Moureaux, C.; Wohlfahrt, G.; et al. Interpreting canopy development and physiology using a European phenology camera network at flux sites. *Biogeosciences* **2015**, *12*, 5995–6015. [\[CrossRef\]](#)
60. Moore, C.E.; Brown, T.; Keenan, T.F.; Duursma, R.A.; van Dijk, A.I.J.M.; Beringer, J.; Culvenor, D.; Evans, B.; Huete, A.; Hutley, L.B.; et al. Reviews and syntheses: Australian vegetation phenology: New insights from satellite remote sensing and digital repeat photography. *Biogeosciences* **2016**, *13*, 5085–5102. [\[CrossRef\]](#)
61. Brown, T.B.; Hultine, K.R.; Steltzer, H.; Denny, E.G.; Denslow, M.W.; Granados, J.; Henderson, S.; Moore, D.; Nagai, S.; SanClements, M.; et al. Using phenocams to monitor our changing Earth: Toward a global phenocam network. *Front. Ecol. Environ.* **2016**, *14*, 84–93. [\[CrossRef\]](#)
62. FSI. Chapter 2: Forest Cover. 2021. Available online: <https://fsi.nic.in/isfr-2021/chapter-2.pdf> (accessed on 28 June 2022).
63. Champion, H.G.; Seth, S.K. *A Revised Survey of the Forest Types of India*; Government of India: New Delhi, India, 1968.
64. Das, T.; Das, A.K. Vegetative and reproductive phenology of some multipurpose tree species in the homegardens of Barak Valley, northeast India. *Int. J. Biometeorol.* **2013**, *57*, 185–196. [\[CrossRef\]](#) [\[PubMed\]](#)
65. Tripathi, A.K.; Pandey, P.C.; Sharma, J.K.; Triantakoustantis, D.; Srivastava, P.K. Climate Change and Its Impact on Forest of Indian Himalayan Region: A Review. *Clim. Chang.* **2022**, 207–222. [\[CrossRef\]](#)
66. Prasad, S.N.; Hegde, M. Phenology and seasonality in the tropical deciduous forest of Bandipur, South India. *Proc. Plant Sci.* **1986**, *96*, 121–133. [\[CrossRef\]](#)
67. Ralhan, P.K.; Khanna, R.; Singh, S.P.; Singh, J.S. Certain phenological characters of the shrub layer of Kumaun Himalayan forests. *Vegetatio* **1985**, *63*, 113–119. [\[CrossRef\]](#)
68. Murali, K.S.; Sukumar, R. Leaf flushing phenology and herbivory in a tropical dry deciduous forest, southern India. *Oecologia* **1993**, *94*, 114–119. [\[CrossRef\]](#) [\[PubMed\]](#)
69. Ganesh, T.; Davidar, P. Flowering phenology and flower predation of *Cullenia exarillata* (Bombacaceae) by arboreal vertebrates in Western Ghats, India. *J. Trop. Ecol.* **1997**, *13*, 459–468. [\[CrossRef\]](#)
70. Boojh, R.; Ramakrishnan, P.S. Growth strategy of trees related to successional status II. Leaf dynamics. *For. Ecol. Manag.* **1982**, *4*, 375–386. [\[CrossRef\]](#)
71. Shukla, R.P.; Ramakrishnan, P.S. Phenology of trees in a sub-tropical humid forest in north-eastern India. *Vegetatio* **1982**, *49*, 103–109. [\[CrossRef\]](#)
72. Omesh, B.; Anoop, K.; Mishra, A.K.; Nayan, S.; Behera, S.K.; Chaudhary, L.B. Phenological study of two dominant tree species in tropical moist deciduous forest from the northern India. *Int. J. Bot.* **2012**, *8*, 66–72.
73. Datta, A.; Rane, A. Phenology, seed dispersal and regeneration patterns of *Horsfieldia kingii*, a rare wild nutmeg. *Trop. Conserv. Sci.* **2013**, *6*, 674–689. [\[CrossRef\]](#)

74. Suresh, H.S.; Sukumar, R. Phenology of *Ficus* spp. in a tropical dry forest, Mudumalai, south India. *J. For. Res.* **2018**, *29*, 1129–1138. [[CrossRef](#)]
75. Ramaswami, G.; Datta, A.; Reddy, A.; Quader, S. Tracking phenology in the tropics and in India: The impacts of climate change. *Implic. Clim. Chang. Indian Biodivers. Overv.* **2019**, *16*, 45–59.
76. Prabakaran, C.; Singh, C.P.; Panigrahy, S.; Parihar, J.S. Retrieval of forest phenological parameters from remote sensing-based NDVI time-series data. *Curr. Sci.* **2013**, *105*, 795–802.
77. Jeganathan, C.; Dash, J.; Atkinson, P.M. Remotely sensed trends in the phenology of northern high latitude terrestrial vegetation, controlling for land cover change and vegetation type. *Remote Sens. Environ.* **2014**, *143*, 154–170. [[CrossRef](#)]
78. Rajan, H.; Jeganathan, C. Understanding Spatio-temporal Pattern of Grassland Phenology in the western Indian Himalayan State. *J. Indian Soc. Remote Sens.* **2019**, *47*, 1137–1151. [[CrossRef](#)]
79. Sharma, R.; Kaur, S.; Uniyal, S.K. Tracking the seasonal dynamics of Himalayan birch using a time-lapse camera. *Folia Geobot.* **2021**, *56*, 125–138. [[CrossRef](#)]
80. Parihar, J.S.; Goroshi, S.; Singh, R.P.; Krishnayya, N.S.R.; Sirsayya, M.B.; Kumar, A.; Rawat, L.S.; Sonakia, A. Observation of forest phenology using field-based digital photography and satellite data. *Curr. Sci.* **2013**, *105*, 1740–1747.
81. Liu, Z.; An, S.; Lu, X.; Hu, H.; Tang, J. Using canopy greenness index to identify leaf ecophysiological traits during the foliar senescence in an oak forest. *Ecosphere* **2018**, *9*, e02337. [[CrossRef](#)]
82. Graham, E.A.; Riordan, E.C.; Yuen, E.M.; Estrin, D.; Rundel, P.W. Public Internet-connected cameras used as a cross-continental ground-based plant phenology monitoring system. *Glob. Chang. Biol.* **2010**, *16*, 3014–3023. [[CrossRef](#)]
83. Negi, V.S.; Pathak, R.; Rawal, R.S.; Bhatt, I.D.; Sharma, S. Long-term ecological monitoring on forest ecosystems in Indian Himalayan Region: Criteria and indicator approach. *Ecol. Indic.* **2019**, *102*, 374–381. [[CrossRef](#)]
84. Sarkar, D.P.; Shankar, B.U.; Parida, B.R. Machine learning approach to predict terrestrial gross primary productivity using topographical and remote sensing data. *Ecol. Inform.* **2022**, *70*, 101697. [[CrossRef](#)]
85. Reid, A.M.; Chapman, W.K.; Prescott, C.E.; Nijland, W. Using excess greenness and green chromatic coordinate colour indices from aerial images to assess lodgepole pine vigour, mortality and disease occurrence. *For. Ecol. Manag.* **2016**, *374*, 146–153. [[CrossRef](#)]
86. Toda, M.; Richardson, A.D. Estimation of plant area index and phenological transition dates from digital repeat photography and radiometric approaches in a hardwood forest in the Northeastern United States. *Agric. For. Meteorol.* **2018**, *249*, 457–466. [[CrossRef](#)]
87. Migliavacca, M.; Galvagno, M.; Cremonese, E.; Rossini, M.; Meroni, M.; Sonnentag, O.; Cogliati, S.; Manca, G.; Diotri, F.; Busetto, L.; et al. Using digital repeat photography and eddy covariance data to model grassland phenology and photosynthetic CO₂ uptake. *Agric. For. Meteorol.* **2011**, *151*, 1325–1337. [[CrossRef](#)]
88. Gonçalves, N.B.; Lopes, A.P.; Dalagnol, R.; Wu, J.; Pinho, D.M.; Nelson, B.W. Both near-surface and satellite remote sensing confirm drought legacy effect on tropical forest leaf phenology after 2015/2016 ENSO drought. *Remote Sens. Environ.* **2020**, *237*, 111489. [[CrossRef](#)]
89. Restrepo-Coupe, N.; Huete, A.; Davies, K.; Cleverly, J.; Beringer, J.; Eamus, D.; van Gorsel, E.; Hutley, L.B.; Meyer, W.S. MODIS vegetation products as proxies of photosynthetic potential: A look across meteorological and biologic driven ecosystem productivity. *Biogeosci. Discuss.* **2015**, *12*, 19213–19267.
90. Tian, F.; Cai, Z.; Jin, H.; Hufkens, K.; Scheffinger, H.; Tagesson, T.; Smets, B.; Van Hoolst, R.; Bonte, K.; Ivits, E.; et al. Calibrating vegetation phenology from Sentinel-2 using eddy covariance, PhenoCam, and PEP725 networks across Europe. *Remote Sens. Environ.* **2021**, *260*, 112456. [[CrossRef](#)]
91. Foley, J.A.; Levis, S.; Prentice, I.C.; Pollard, D.; Thompson, S.L. Coupling dynamic models of climate and vegetation. *Glob. Chang. Biol.* **1998**, *4*, 561–579. [[CrossRef](#)]
92. Nagai, S.; Maeda, T.; Gamo, M.; Muraoka, H.; Suzuki, R.; Nasahara, K.N. Using digital camera images to detect canopy condition of deciduous broad-leaved trees. *Plant Ecol. Divers.* **2011**, *4*, 79–89. [[CrossRef](#)]
93. Zhang, X.; Jayavelu, S.; Liu, L.; Friedl, M.A.; Henebry, G.M.; Liu, Y.; Schaaf, C.B.; Richardson, A.D.; Gray, J. Evaluation of land surface phenology from VIIRS data using time series of PhenoCam imagery. *Agric. For. Meteorol.* **2018**, *256–257*, 137–149. [[CrossRef](#)]
94. Nagai, S.; Saitoh, T.M.; Noh, N.J.; Yoon, T.K.; Kobayashi, H.; Suzuki, R.; Nasahara, K.N.; Son, Y.; Muraoka, H. Utility of information in photographs taken upwards from the floor of closed-canopy deciduous broadleaved and closed-canopy evergreen coniferous forests for continuous observation of canopy phenology. *Ecol. Inform.* **2013**, *18*, 10–19. [[CrossRef](#)]
95. Descals, A.; Verger, A.; Yin, G.; Peñuelas, J. Improved estimates of arctic land surface phenology using sentinel-2 time series. *Remote Sens.* **2020**, *12*, 3738. [[CrossRef](#)]
96. Snyder, K.A.; Huntington, J.L.; Wehan, B.L.; Morton, C.G.; Stringham, T.K. Comparison of landsat and land-based phenology camera normalized difference vegetation index (NDVI) for dominant plant communities in the great basin. *Sensors* **2019**, *19*, 1139. [[CrossRef](#)] [[PubMed](#)]
97. Andresen, C.; Tweedie, C.; Loughheed, V. Climate and nutrient effects on Arctic wetland plant phenology observed from phenocams. *Remote Sens. Environ.* **2018**, *205*, 46–55. [[CrossRef](#)]

98. Baumann, M.; Ozdogan, M.; Richardson, A.D.; Radeloff, V.C. Phenology from Landsat when data is scarce: Using MODIS and Dynamic Time-Warping to combine multi-year Landsat imagery to derive annual phenology curves. *Int. J. Appl. Earth Obs. Geoinf.* **2017**, *54*, 72–83. [\[CrossRef\]](#)
99. Zhang, S.; Buttò, V.; Khare, S.; Deslauriers, A.; Morin, H.; Huang, J.-G.; Ren, H.; Rossi, S. Calibrating PhenoCam data with phenological observations of a black spruce stand. *Can. J. Remote Sens.* **2020**, *46*, 154–165. [\[CrossRef\]](#)
100. Marchin, R.M.; McHugh, I.; Simpson, R.R.; Ingram, L.J.; Balas, D.S.; Evans, B.J.; Adams, M.A. Productivity of an Australian mountain grassland is limited by temperature and dryness despite long growing seasons. *Agric. For. Meteorol.* **2018**, *256–257*, 116–124. [\[CrossRef\]](#)
101. Inoue, T.; Nagai, S.; Saitoh, T.M.; Muraoka, H.; Nasahara, K.N.; Koizumi, H. Detection of the different characteristics of year-to-year variation in foliage phenology among deciduous broad-leaved tree species by using daily continuous canopy surface images. *Ecol. Inform.* **2014**, *22*, 58–68. [\[CrossRef\]](#)
102. Ahrends, H.; Etzold, S.; Kutsch, W.; Stoeckli, R.; Bruegger, R.; Jeanneret, F.; Wanner, H.; Buchmann, N.; Eugster, W. Tree phenology and carbon dioxide fluxes: Use of digital photography for process-based interpretation at the ecosystem scale. *Clim. Res.* **2009**, *39*, 261–274. [\[CrossRef\]](#)
103. Crimmins, M.A.; Crimmins, T.M. Monitoring plant phenology using digital repeat photography. *Environ. Manag.* **2008**, *41*, 949–958. [\[CrossRef\]](#)
104. Linkosalmi, M.; Aurela, M.; Tuovinen, J.-P.; Peltoniemi, M.; Tanis, C.M.; Arslan, A.N.; Kolari, P.; Böttcher, K.; Aalto, T.; Rainne, J.; et al. Digital photography for assessing the link between vegetation phenology and CO₂ exchange in two contrasting northern ecosystems. *Geosci. Instrum. Methods Data Syst.* **2016**, *5*, 417–426. [\[CrossRef\]](#)
105. Jorge, C.; Silva, J.M.N.; Boavida-Portugal, J.; Soares, C.; Cerasoli, S. Using digital photography to track understory phenology in mediterranean cork oak woodlands. *Remote Sens.* **2021**, *13*, 776. [\[CrossRef\]](#)
106. Nijland, W.; Coops, N.; Coogan, S.; Bater, C.; Wulder, M.; Nielsen, S.; McDermid, G.; Stenhouse, G. Vegetation phenology can be captured with digital repeat photography and linked to variability of root nutrition in *Hedysarum alpinum*. *Appl. Veg. Sci.* **2013**, *16*, 317–324. [\[CrossRef\]](#)
107. Paloschi, R.A.; Ramos, D.M.; Ventura, D.J.; Souza, R.; Souza, E.; Morellato, L.P.C.; Nóbrega, R.L.B.; Coutinho, Í.A.C.; Verhoef, A.; Körting, T.S.; et al. Environmental drivers of water use for caatinga woody plant species: Combining remote sensing phenology and sap flow measurements. *Remote Sens.* **2020**, *13*, 75. [\[CrossRef\]](#)
108. Koide, D.; Ide, R.; Oguma, H. Detection of autumn leaf phenology and color brightness from repeat photography: Accurate, robust, and sensitive indexes and modeling under unstable field observations. *Ecol. Indic.* **2019**, *106*, 105482. [\[CrossRef\]](#)
109. Wang, J.; Yang, D.; Detto, M.; Nelson, B.W.; Chen, M.; Guan, K.; Wu, S.; Yan, Z.; Wu, J. Multi-scale integration of satellite remote sensing improves characterization of dry-season green-up in an Amazon tropical evergreen forest. *Remote Sens. Environ.* **2020**, *246*, 111865. [\[CrossRef\]](#)
110. Atkins, J.W.; Stovall, A.E.L.; Yang, X. Mapping temperate forest phenology using tower, UAV, and terrestrial-based sensors. *Drones* **2020**, *4*, 56. [\[CrossRef\]](#)
111. Liu, Y.; Hill, M.J.; Zhang, X.; Wang, Z.; Richardson, A.D.; Hufkens, K.; Filippa, G.; Baldocchi, D.D.; Ma, S.; Verfaillie, J.; et al. Using data from Landsat, MODIS, VIIRS and PhenoCams to monitor the phenology of California oak/grass savanna and open grassland across spatial scales. *Agric. For. Meteorol.* **2017**, *237–238*, 311–325. [\[CrossRef\]](#)
112. Julitta, T.; Cremonese, E.; Migliavacca, M.; Colombo, R.; Galvagno, M.; Siniscalco, C.; Rossini, M.; Fava, F.; Cogliati, S.; Di Cella, U.M.; et al. Using digital camera images to analyse snowmelt and phenology of a subalpine grassland. *Agric. For. Meteorol.* **2014**, *198–199*, 116–125. [\[CrossRef\]](#)
113. Xie, Y.; Civco, D.L.; Silander, J.A., Jr. Species-specific spring and autumn leaf phenology captured by time-lapse digital cameras. *Ecosphere* **2018**, *9*, e02089. [\[CrossRef\]](#)
114. Vrieling, A.; Meroni, M.; Darvishzadeh, R.; Skidmore, A.K.; Wang, T.; Zurita-Milla, R.; Oosterbeek, K.; O'Connor, B.; Paganini, M. Vegetation phenology from Sentinel-2 and field cameras for a Dutch barrier island. *Remote Sens. Environ.* **2018**, *215*, 517–529. [\[CrossRef\]](#)
115. Melaas, E.K.; Friedl, M.A.; Richardson, A.D. Multiscale modeling of spring phenology across Deciduous Forests in the Eastern United States. *Glob. Chang. Biol.* **2016**, *22*, 792–805. [\[CrossRef\]](#) [\[PubMed\]](#)
116. Nezval, O.; Krejza, J.; Světlík, J.; Šigut, L.; Horáček, P. Comparison of traditional ground-based observations and digital remote sensing of phenological transitions in a floodplain forest. *Agric. For. Meteorol.* **2020**, *291*, 108079. [\[CrossRef\]](#)
117. Thapa, S.; Millan, V.E.G.; Eklundh, L. Assessing forest phenology: A multi-scale comparison of near-surface (UAV, spectral reflectance sensor, phenocam) and satellite (MODIS, sentinel-2) remote sensing. *Remote Sens.* **2021**, *13*, 1597. [\[CrossRef\]](#)
118. Brown, L.A.; Dash, J.; Ogutu, B.O.; Richardson, A.D. On the relationship between continuous measures of canopy greenness derived using near-surface remote sensing and satellite-derived vegetation products. *Agric. For. Meteorol.* **2017**, *247*, 280–292. [\[CrossRef\]](#)
119. Henneken, R.; Dose, V.; Schleip, C.; Menzel, A. Detecting plant seasonality from webcams using Bayesian multiple change point analysis. *Agric. For. Meteorol.* **2013**, *168*, 177–185. [\[CrossRef\]](#)
120. Morris, D.E.; Boyd, D.S.; Crowe, J.A.; Johnson, C.S.; Smith, K.L. Exploring the potential for automatic extraction of vegetation phenological metrics from traffic webcams. *Remote Sens.* **2013**, *5*, 2200–2218. [\[CrossRef\]](#)

121. Melaas, E.K.; Sulla-Menasse, D.; Gray, J.M.; Black, T.A.; Morin, T.H.; Richardson, A.D.; Friedl, M.A. Multisite analysis of land surface phenology in North American temperate and boreal deciduous forests from Landsat. *Remote Sens. Environ.* **2016**, *186*, 452–464. [\[CrossRef\]](#)
122. Peichl, M.; Sonnentag, O.; Nilsson, M.B. Bringing color into the picture: Using digital repeat photography to investigate phenology controls of the carbon dioxide exchange in a boreal mire. *Ecosystems* **2015**, *18*, 115–131. [\[CrossRef\]](#)
123. Cheng, Y.; Vrieling, A.; Fava, F.; Meroni, M.; Marshall, M.; Gachoki, S. Phenology of short vegetation cycles in a Kenyan rangeland from PlanetScope and Sentinel-2. *Remote Sens. Environ.* **2020**, *248*, 112004. [\[CrossRef\]](#)
124. Matiu, M.; Bothmann, L.; Steinbrecher, R.; Menzel, A. Monitoring succession after a non-cleared windthrow in a Norway spruce mountain forest using webcam, satellite vegetation indices and turbulent CO₂ exchange. *Agric. For. Meteorol.* **2017**, *244–245*, 72–81. [\[CrossRef\]](#)
125. Zhao, J.; Zhang, Y.; Tan, Z.; Song, Q.; Liang, N.; Yu, L.; Zhao, J. Using digital cameras for comparative phenological monitoring in an evergreen broad-leaved forest and a seasonal rain forest. *Ecol. Inform.* **2012**, *10*, 65–72. [\[CrossRef\]](#)
126. Yan, D.; Scott, R.; Moore, D.; Biederman, J.; Smith, W. Understanding the relationship between vegetation greenness and productivity across dryland ecosystems through the integration of PhenoCam, satellite, and eddy covariance data. *Remote Sens. Environ.* **2019**, *223*, 50–62. [\[CrossRef\]](#)
127. Songsom, V.; Koedsin, W.; Ritchie, R.J.; Huete, A. Mangrove phenology and water influences measured with digital repeat photography. *Remote Sens.* **2021**, *13*, 307. [\[CrossRef\]](#)
128. Seyednasrollah, B.; Bowling, D.R.; Cheng, R.; Logan, B.A.; Magney, T.S.; Frankenberg, C.; Yang, J.C.; Young, A.M.; Hufkens, K.; Arain, M.A.; et al. Seasonal variation in the canopy color of temperate evergreen conifer forests. *New Phytol.* **2021**, *229*, 2586–2600. [\[CrossRef\]](#) [\[PubMed\]](#)
129. Wheeler, K.I.; Dietze, M.C. Improving the monitoring of deciduous broadleaf phenology using the Geostationary Operational Environmental Satellite (GOES) 16 and 17. *Biogeosciences* **2021**, *18*, 1971–1985. [\[CrossRef\]](#)
130. Cao, M.; Sun, Y.; Jiang, X.; Li, Z.; Xin, Q. Identifying leaf phenology of deciduous broadleaf forests from phenocam images using a convolutional neural network regression method. *Remote Sens.* **2021**, *13*, 2331. [\[CrossRef\]](#)
131. Moon, M.; Richardson, A.D.; Friedl, M.A. Multiscale assessment of land surface phenology from harmonized Landsat 8 and Sentinel-2, PlanetScope, and PhenoCam imagery. *Remote Sens. Environ.* **2021**, *266*, 112716. [\[CrossRef\]](#)
132. Khare, S.; Deslauriers, A.; Morin, H.; Latifi, H.; Rossi, S. Comparing time-lapse PhenoCams with satellite observations across the boreal forest of Quebec, Canada. *Remote Sens.* **2021**, *14*, 100. [\[CrossRef\]](#)
133. Yan, D.; Zhang, X.; Nagai, S.; Yu, Y.; Akitsu, T.; Nasahara, K.N.; Ide, R.; Maeda, T. Evaluating land surface phenology from the Advanced Himawari Imager using observations from MODIS and the Phenological Eyes Network. *Int. J. Appl. Earth Obs. Geoinf.* **2019**, *79*, 71–83. [\[CrossRef\]](#)
134. Shukla, R.P.; Ramakrishnan, P.S. Leaf dynamics of tropical trees related to successional status. *New Phytol.* **1984**, *97*, 697–706. [\[CrossRef\]](#)
135. Bhat, D.M. Phenology of tree species of tropical moist forest of Uttara Kannada district, Karnataka, India. *J. Biosci.* **1992**, *17*, 325–352. [\[CrossRef\]](#)
136. Murali, K.S.; Sukumar, R. Reproductive phenology of a tropical dry forest in Mudumalai, southern India. *J. Ecol.* **1994**, *82*, 759. [\[CrossRef\]](#)
137. Ganesh, T.; Davidar, P. Fruiting phenology and pre-dispersal seed predation in a rainforest in southern Western Ghats, India. *Trop. Fruits Frugivores Search Strong Interactors* **2005**, 139–154.
138. Krishnan, R.M. Reproductive phenology of a wet forest understorey in the Western Ghats, South India. *Glob. Ecol. Biogeogr.* **2002**, *11*, 179–182. [\[CrossRef\]](#)
139. Sundarapandian, S.M.; Chandrasekaran, S.; Swamy, P.S. Phenological behaviour of selected tree species in tropical forests at Kodayar in the Western Ghats, Tamil Nadu, India. *Curr. Sci.* **2005**, *88*, 805–810.
140. Selwyn, M.A.; Parthasarathy, N. Reproductive traits and phenology of plants in tropical dry evergreen forest on the Coromandel coast of India. *Biodivers. Conserv.* **2006**, *15*, 3207–3234. [\[CrossRef\]](#)
141. Sellamuthu, S.; Lalitha, V. Plant diversity and phenological pattern in the montane wet temperate forests of the southern Western Ghats, India. *For. Stud. China* **2010**, *12*, 116–125. [\[CrossRef\]](#)
142. Suresh, H.S.; Sukumar, R. Vegetative phenology of tropical montane forests in the Nilgiris, South India. *J. Natl. Sci. Found. Sri Lanka* **2011**, *39*, 333. [\[CrossRef\]](#)
143. Nanda, A.; Krishnamurthy, Y.L.; Suresh, H.S. Canopy trees leaf phenology in tropical dry deciduous and evergreen forests of Bhadra Wildlife Sanctuary Karnataka, India. *Afr. J. Plant Sci.* **2013**, *7*, 170–175. [\[CrossRef\]](#)
144. Nanda, A.; Suresh, H.S.; Krishna Murthy, Y.L. Leafing phenology of tropical forests of Bhadra wildlife sanctuary, Karnataka, India. *Appl. Sci. Rep.* **2015**, *12*, 33–40.
145. Kushwaha, C.P.; Singh, K.P. Diversity of leaf phenology in a tropical deciduous forest in India. *J. Trop. Ecol.* **2005**, *21*, 47–56. [\[CrossRef\]](#)
146. Bajpai, O.; Pandey, J.; Chaudhary, L.B. Periodicity of different phenophases in selected trees from Himalayan Terai of India. *Agrofor. Syst.* **2017**, *91*, 363–374. [\[CrossRef\]](#)
147. Chaurasia, B.; Shukla, R.P. Changes in reproductive phenology and sex ratio of *Trewia nudiflora* Linn. Growing in sal Forest of North-eastern Uttar Pradesh, India. *Trop. Ecol.* **2016**, *57*, 89–99.

148. Mishra, R.K.; Upadhyay, V.P.; Bal, S.; Mohapatra, P.K.; Mohanty, R.C. Phenology of species of moist deciduous forest sites of Similipal biosphere reserve. *Lyonia* **2006**, *11*, 5–17.
149. Yadav, R.K.; Yadav, A.S. Phenology of selected woody species in a tropical dry deciduous forest in Rajasthan, India. *Trop. Ecol.* **2008**, *49*, 25–34.
150. Nautiyal, M.C.; Nautiyal, B.P.; Prakash, V. Phenology and growth form distribution in an alpine pasture at Tungnath, Garhwal, Himalaya. *Mt. Res. Dev.* **2001**, *21*, 168–174. [[CrossRef](#)]
151. Joshi, V.C.; Janarthanam, M.K. The diversity of life-form type, habitat preference and phenology of the endemics in the Goa region of the Western Ghats, India. *J. Biogeogr.* **2004**, *31*, 1227–1237. [[CrossRef](#)]
152. Borah, M.; Devi, A. Phenology, growth and survival of *Vatica lanceaefolia* Bl.: A critically endangered tree species in moist tropical forest of Northeast India. *Trop. Plant Res.* **2014**, *1*, 1–12.
153. Nath, S.; Nath, A.J.; Das, A.K. Vegetative and reproductive phenology of a floodplain tree species *Barringtonia acutangula* from North East India. *J. Environ. Biol.* **2016**, *37*, 215–220.
154. Aruna, R.; Balasubramanian, P. Fruiting phenology of a scrub forest in thiruporur, Eastern Ghats, India. *Int. Lett. Nat. Sci.* **2015**, *44*, 25–30. [[CrossRef](#)]
155. Arunparasath, A.; Gomathinayagam, M. Reproductive phenology of true mangrove species in Pichavaram mangrove forests, Tamilnadu, India—A comparative account. *J. Environ. Treat. Tech.* **2015**, *15*, 17–21.
156. Kaur, G.; Singh, B.P.; Nagpal, A.K. Phenology of Some Phanerogams (Trees and Shrubs) of Northwestern Punjab, India. *J. Bot.* **2013**, *2013*, 712405. [[CrossRef](#)]
157. Kikim, A.; Yadava, P.S. Phenology of tree species in subtropical forests of Manipur in north eastern India. *Trop. Ecol.* **2001**, *42*, 269–276.
158. Pandey, R.; Pandey, C.N. Reproductive Strategy of *Aegiceras corniculatum* L. (Blanco.)—A Mangrove Species, in MNP&S, Gujarat, India. *J. Plant Stud.* **2014**, *3*, 35. [[CrossRef](#)]
159. Singh, K.P.; Kushwaha, C.P. Diversity of flowering and fruiting phenology of trees in a tropical deciduous forest in India. *Ann. Bot.* **2006**, *97*, 265–276. [[CrossRef](#)] [[PubMed](#)]
160. Newton, P.N. The structure and phenology of a moist deciduous forest in the Central Indian Highlands. *Vegetatio* **1988**, *75*, 3–16. [[CrossRef](#)]
161. Thakur, P.S.; Dutt, V.; Thakur, A. Impact of inter-annual climate variability on the phenology of eleven multipurpose tree species. *Curr. Sci.* **2008**, *94*, 1053–1058.
162. Sivaraj, N.; Krishnamurthy, K.V. Flowering phenology in the vegetation of Shervaroys, South India. *Vegetatio* **1988**, *79*, 85–88. [[CrossRef](#)]
163. Sundriyal, R.C. Phenology of some temperate woody species of the Garhwal Himalaya. *Int. J. Ecol. Environ. Sci.* **1990**, *16*, 107–117.
164. Sundriyal, R.C.; Joshi, A.P.; Dhasmana, R. Phenology of high altitude plants at Tungnath in the Garhwal Himalaya. *Trop. Ecol.* **1987**, *28*, 289–299.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.