

Article

The Emissions of Carbon Dioxide, Methane, and Nitrous Oxide during Winter without Cultivation in Local Saline-Alkali Rice and Maize Fields in Northeast China

Hao Zhang ^{1,2}, Jie Tang ^{1,2,*}, Shuang Liang ^{3,*}, Zhaoyang Li ², Ping Yang ², Jingjing Wang ² and Sining Wang ²

- Key Laboratory of Groundwater Resources and Environment, Ministry of Education, Jilin University, Changchun 130012, China; zhao14@mails.jlu.edu.cn
- ² College of Environment and Resources, Jilin University, Changchun 130012, China; zhaoyang@jlu.edu.cn (Z.L.); yangping83@jlu.edu.cn (P.Y.); jingjing16@mails.jlu.edu.cn (J.W.); ronggz15@mails.jlu.edu.cn (S.W.)
- ³ Key Laboratory of Songliao Aquatic Environment, Ministry of Education, Jilin Jianzhu University, Changchun 130118, China
- * Correspondence: tangjie@jlu.edu.cn (J.T.); liangshuang13@mails.jlu.edu.cn (S.L.); Tel.: +86-130-6920-9064 (J.T.); +86-137-5604-1683 (S.L.); Fax: +86-431-8515-9440 (J.T. & S.L.)

Received: 28 August 2017; Accepted: 19 October 2017; Published: 23 October 2017

Abstract: Agricultural ecosystems are important contributors to atmospheric greenhouse gasses (GHGs); however, in situ winter emission data in saline-alkali fields are scarce. Gas samples were collected during different periods, from three rice (R1–R3) and three maize (M1–M3) fields with different soil pH levels and salinity conditions. Carbon dioxide (CO₂) emissions in the rice and maize fields decreased with decreasing temperature during the freezing period and increased with the rising temperature during the thawing period, with the majority of winter CO₂ emissions occurring during these two periods. Peaks in methane (CH₄) emissions were observed during the freezing period in the rice fields and during the snow-melting period in the rice and maize fields. CH₄ emissions in the rice fields and CH₄ uptake rates in the maize fields were significantly (P < 0.05) related to surface soil temperature. Nitrous oxide (N₂O) emissions remained relatively low, except for during the peaks observed during the snow-melting period in both the rice and maize fields, leading to the high GHG contribution of the snow-melting period throughout the winter. Higher pH and salinity conditions consistently resulted in lower CO₂, CH₄, and N₂O emissions, CH₄ uptake, and lower global warming potential (GWP). These results can contribute to the assessment of the GWP during winter in saline-alkali regions.

Keywords: greenhouse gas; saline-alkali field; rice field; maize field; seasonal freeze-thaw

1. Introduction

Global warming could alter the earth's ecosystems by rising sea levels caused by glacier melting, changes in biome distribution, and food production, and an elevated minimum temperature in winter, and it influences the survival and development of humans. Global temperatures have increased by 0.85 °C in the past 130 years, and are predicted to continue to increase by 1.2 °C to 4.8 °C by the end of the 21st century [1]. According to the 2013 Intergovernmental Panel on Climate Change (IPCC), the increased atmospheric concentration of greenhouse gases (GHGs), including carbon dioxide (CO₂), methane (CH₄), and nitrous oxide (N₂O), are responsible for past, current, and predicted future global



warming by substantially increasing the greenhouse effect [1]. As a result, GHG emissions are receiving increased attention by the international community.

 CO_2 , CH_4 , and N_2O are the main gases affecting global warming [2], contributing to approximately 60%, 20%, and 6% of the greenhouse effect, respectively [3]. Although increasing atmospheric CO_2 concentrations account for most the GHGs (IPCC, 2013) [1], N_2O and CH_4 are equally as important because of their unique radiative properties and long residence time in the atmosphere result in their global warming potential (GWP) being 298 and 25 times that of CO_2 over a period of 100 years, respectively (IPCC, 2007) [3]. Consequently, considering the emissions of these three gasses when assessing the greenhouse effect is important.

Terrestrial ecosystems are important sources of atmospheric CO₂, CH₄, and N₂O [1,4], and agricultural lands, which occupy 37% of the earth's land surface [5], release large amounts of these GHGs into the atmosphere [5,6]. The most recent studies focusing on CO₂, CH₄, and N₂O emissions from agricultural ecosystems examined the effects of management practices, such as different tillage [6–9] or fertilization practices [7,10–13], environmental changes including changes in temperature [14] and moisture [7,12,15,16], soil depth [17], and soil physicochemical properties [18] on GHG emissions. Studies regarding GHG fluxes on seasonally frozen soil in alkali-saline soil regions are lacking.

Seasonal freeze-thaw is an important meteorological event in boreal areas. GHG emissions during the freeze-thaw period likely lead to variations in the atmospheric GHG concentrations [19–21] and influence the greenhouse effect [22]. Some studies have shown that CO₂ emissions are inhibited, to a large degree, by freezing, and are activated by thawing [23,24]. However, other studies have shown that freezing enhances CO₂ emissions [25]. Similarly, it has been reported that freezing significantly inhibits CH₄ emissions, whereas thawing activates CH₄ emissions [24,26]. Other studies have found that the freeze-thaw cycles has inconsistent effects on CH₄ emissions [27]. Although some studies have shown that freezing [25] or freeze-thaw cycles [28,29] enhance N₂O emissions, other studies suggest that N₂O emissions decrease due to freezing and increase due to thawing [24,30,31].

In addition, most previous freeze-thaw experiments investigated areas with soils with pH values below 7.0, with a limited focus on saline-alkali soil. Saline-alkali stress inhibits microbial biomass and affects the microbial community and structure [32,33]. High salinity inhibits microbial activities and the availability of substrate [34], and prevents the production of enzymes [35]. Therefore, the pH and salinity conditions may impact CO_2 , CH_4 , and N_2O emissions, but more research is required in this area.

The Western Jilin Province in Northeast China is one of the largest soda saline-alkaline areas on earth. The area is an important agricultural production base in China, predominantly consisting of rice paddies and maize lands. Although winter impacts the soil for over six months of the year, from the end of October until the end of April of the following year, continuous field monitoring data of CO_2 , CH_4 , and N_2O emissions throughout the winter in the area remain scarce. Three rice fields and three maize fields of different pH and salinity values were examined for this study. The overall objective was to (1) determine the CO_2 , CH_4 , and N_2O emissions in rice and maize fields during winter and to assess their contribution to the GWP; and (2) to analyze their emissions under different pH and salinity values. The results of the current study supplement existing basal data on CO_2 , CH_4 , and N_2O emissions of saline-alkali rice and maize fields during winter and provide a scientific basis for the accurate assessment of the GWP.

2. Materials and Methods

2.1. Study Site

The sampling sites were in Songyuan City of the Western Jilin Province $(123^{\circ}091'E-124^{\circ}221'E, 44^{\circ}571'N-45^{\circ}461'N)$, Northeast China. The local climate is characterized by long, cold winters, with large amounts of snow, with the lowest air temperature reaching $-21^{\circ}C$ (Figure 1). The average

minimum air temperature during winter of the recent five years (2011–2015) was -12.0 °C with a lowest air temperature -35.6 °C in 2011. The local rice and maize fields experience four periods throughout the winter: freezing, hard frost, snow-melting, and thawing (Figure 1). The freezing period begins when the minimum air temperature drops to approximately 0 °C and ends when the surface soil freezes completely. Subsequently, the soil experiences hard frost under colder temperatures. The snow-melting period begins when the snow starts to melt due to increasing temperatures. The thawing period begins when snow disappears and the soil has thawed completely.



Figure 1. Temperatures of the air and soil during the field experiment.

2.2. Sampling and Measurement of CO₂, CH₄, and N₂O Fluxes

Three rice fields (R1: 20.4 m \times 30.7 m; R2: 22.6 m \times 36.8 m; R3: 15.5 m \times 30.3 m; rice type: Tonghe 837) and three maize fields (M1: 10.0 m \times 44.8 m; M2: 12.3 m \times 46.2 m; M3: 13.7 m \times 46.8 m; maize type: Tongyu 9585) with different pH and salinity values (Table 1) were selected.

		рН	Salinity (%)	Moisture (%)	Bulk Density (g·cm ^{−3})	Nitrate−N (mg·kg ⁻¹)	Total N (g∙kg ⁻¹)	Soil Organic C (g·kg ⁻¹)
	R1	7.6	0.16	28.68	1.28	3.87	2.11	22.45
Rice Field (Silty Loam)	R2	8.6	0.27	28.35	1.33	2.45	1.79	17.58
	R3	9.1	0.41	27.98	1.34	1.86	0.75	13.46
	M1	7.34	0.10	15.93	1.36	8.65	1.87	17.23
Maize Field (Silty Loam)	M2	7.76	0.19	15.44	1.29	7.86	1.24	12.24
	M3	8.43	0.25	15.16	1.37	3.36	0.89	8.65

Table 1. The physicochemical properties of the soil at a depth of 0–20 cm before gas sampling.

All of the surface litter and stubble were removed after harvest to avoid their uneven distribution and participation in microbial respiration, which might affect GHG emission rates. Gas sampling was conducted from 16 October 2014 to 30 April 2015 using static chambers. Soil samples from a depth of 0–20 cm were collected before the first gas sampling from the sampling plots, to determine the soil properties, such as pH, salinity, soil organic carbon, et al. The pH, salinity, moisture (gravimetric water content), and soil organic carbon (C) were determined by the methods described by Tang et al. [36]. Nitrate-N was measured by the methods described by Bissett et al. [37].

Gas samples were collected every 10 days during the hard frost period and every two days during other periods. Each static chamber (bottomless, 50 cm \times 50 cm \times 50 cm) was composed of transparent plexiglass wrapped in silver paper to reflect sunshine, thereby avoiding rising temperatures. Each

static chamber was equipped with an internal circulating fan to ensure complete gas mixing. A base (without a top or bottom, 50 cm \times 50 cm \times 30 cm) with a seal groove was buried 30 cm into the soil in each plot. There were three plots in each field. Each plot was marked by a stick and equipped with a static chamber combined with a base (Figure 2). The static chamber was embedded into the groove, and a saturated NaCl solution was added to the groove to seal the chamber while gas sampling. The samples of all of the fields were collected between 9:00 a.m. and 11:00 a.m., representing 1-day average flux [38]. The samples of three plots in one field were collected at the same time. Gas samples were collected into vacuum bags using a 50-mL airtight syringe. In 30 min, four samples from a single field were collected every 10 min. The soil temperatures at the surface (0 cm depth) and at a depth of 15 cm were recorded by an electronic geothermometer (Type: TP101, Shenzhen Riters Instrumentation Co., Ltd, Shenzhen, China), which has a temperature range of -50 °C to 300 °C and a testing length of 15 cm. Soil temperatures and snow cover thickness were from one of the six fields because the six fields were close in one plain.



Figure 2. The gas sampling equipment in each plot. The base was fixed in the soil. The snow was kept intact as much as possible when the static chamber was placed on the base each time prior to sampling.

Concentrations of CO_2 , CH_4 , and N_2O in the gas samples were analysed by gas chromatograph (GC, Agilent 7820A, Santa Clara, CA, USA) within 24 h following gas sampling. The gas chromatograph configurations for analysing concentrations of CO_2 , CH_4 , and N_2O were as described by Zhang et al. [39]. The GHG flux was calculated based on the rate of change in gas concentration within the chamber, which was estimated as the slope of the linear regression between concentration and time. The flux was calculated per the equation described by Zhang et al. [39].

2.3. Statistical Methods

The mean flux was calculated as:

$$\overline{F} = \frac{1}{n} \sum_{i=1}^{n} F_i \tag{1}$$

where \overline{F} (CO₂ g·m⁻²·h⁻¹; CH₄, N₂O µg·m⁻²·h⁻¹) is the mean flux of one period; F_i is the flux of CO₂, CH₄, or N₂O during the *i*th sampling time during one period; and *n* is the total number of samples during one period.

The cumulative GHG flux was calculated as follows:

$$F_i = m\overline{F_i} \times 24 \times 10000 \tag{2}$$

where F_j (CO₂ kg·ha⁻¹; CH₄, N₂O g·ha⁻¹) is the cumulative flux of CO₂, CH₄, or N₂O during the *j*th period; $\overline{F_j}$ is the mean flux of CO₂, CH₄, or N₂O during the *j*th period; and *m* is the total number of days during the *j*th period.

$$F_T = \sum_{j=1}^4 F_j \tag{3}$$

where F_T (CO₂ kg·ha⁻¹; CH₄, N₂O g·ha⁻¹) is the cumulative flux of CO₂, CH₄, or N₂O across the entire winter.

GWP (kg CO₂ eq·ha⁻¹) was calculated as follows:

$$GWP = F_{T-CO_2} + 25F_{T-CH_4} + 298F_{T-N_2O}$$
(4)

The statistical analyses were performed using SPSS 22.0 software (International Business Machines Corporation, New York, NY, USA). All figures were drawn using Sigmaplot 12.5 (Systat Software, Inc., San Jose, CA, USA).

3. Results

3.1. Variations in Snow and Temperatures

Temperatures fluctuated widely during winter (Figure 2). Surface soil temperature directly reflected the variations in air temperature. The soil temperature at a depth of 15 cm dropped with decreasing air temperature, and the lowest recorded temperature was -8.5 °C during the hard frost period. From 16 October 2014 to 1 December 2014 was the freezing period. The surface soil temperature was above zero from 15 March 2015 during the snow-melting period. The soil temperature at 15 cm depth was above zero from 27 March 2015 during the thawing period. The minimum air temperature sometimes was below zero during the snow-melting, and thawing periods. These differences in temperature resulted in the freeze-thaw cycles occurring during these periods.

Snow began on 5 November 2014 (Figure 3). The maximum snow cover thickness was 40.0 cm on 1 March 2015, after which the snow melted rapidly and disappeared completely by 27 March 2015. Snowpack insulated the soil from the air and acted to keep the soil warm.



Figure 3. Snow cover thickness during the field experiment in the four periods. The values were obtained from one of the six experimental fields.

3.2. Variations in the Flux of CO_2 , CH_4 , and N_2O

For the flux in CO_2 , the rice fields showed similar trends as the maize fields (Figures 4A and 5A). The flux in CO_2 to the atmosphere decreased with decreasing soil temperature during the freezing period, and remained at the lowest level during the hard frost period in the six fields. The maximum CO_2 emission rate during the freezing period was higher than during other periods in the three rice fields. The CO_2 emission rates began to increase after the snow melted and subsequently reached

a peak during the early thawing period in the six fields. Thereafter, continuously rising CO_2 emission rates were observed in the six fields. R1 had higher fluxes of CO_2 than R2 and R3, and M1 had higher fluxes of CO_2 than M2 and M3 during both the freezing and thawing periods.



Figure 4. Measured mean flux of (**A**) Carbon Dioxide (CO₂); (**B**) Methane (CH₄); and (**C**) Nitrous Oxide (N₂O) in the three rice fields.



Figure 5. Measured mean flux of (A) CO₂; (B) CH₄; and (C) N₂O in the three maize fields.

CH₄ was assimilated by the maize fields during the freezing, hard frost, and thawing periods. During the freezing period, CH₄ fluctuated widely in the six fields, and two CH₄ emission peaks in the three rice fields and a CH₄ uptake peak in the three maize fields were observed. Significant CH₄ emission peaks occurred during the snow-melting period in each of the six fields. The peak values in the six fields followed the order: R1 (126.5 μ g·m⁻²·h⁻¹) > R2 (112.6 μ g·m⁻²·h⁻¹) > R3 (96.8 μ g·m⁻²·h⁻¹) > M1 (22.6 μ g·m⁻²·h⁻¹) > M2 (18.9 μ g·m⁻²·h⁻¹) > M3 (9.2 μ g·m⁻²·h⁻¹). The CH₄ emission peaks in the maize fields during the snow-melting period occurred before those in the rice fields. The CH₄ emissions rates followed the order: R1 > R2 > R3 during the freezing, snowmelting, and thawing periods in the three rice fields. The CH₄ uptake rates followed the order: M1 > M2 > M3 during the freezing and thawing periods in the three maize fields. Gradually increasing CH₄ uptake rates in the three maize fields were observed during the thawing period; however, the CH₄ emission rates experienced a decrease after 24 April 2015.

A single significant peak in N₂O emissions was observed in each of the six fields. The N₂O emission rates of the three rice fields stayed at the lowest level during the hard frost period. Explosive N₂O emissions occurred during the snow-melting period, extending into the initial several days of the thawing period in the six fields. The peak N₂O emission values followed the order: R1 (290.2 μ g·m⁻²·h⁻¹) > M1 (270.5 μ g·m⁻²·h⁻¹) > R2 (227.8 μ g·m⁻²·h⁻¹) > M2 (208.9 μ g·m⁻²·h⁻¹) > M3 (150.4 μ g·m⁻²·h⁻¹) > R3 (146.5 μ g·m⁻²·h⁻¹). However, the duration of high-level N₂O emissions in the rice fields was longer than that in maize fields.

3.3. Mean Flux of CO_2 , CH_4 , and N_2O during the Different Periods

The mean flux of CO₂, CH₄, and N₂O during the different periods fluctuated widely in the six fields (Figures 6 and 7). The mean fluxes of CO₂, CH₄, and N₂O during the same period followed the order R1 > R2 > R3 in the rice fields, and generally followed the order M1 > M2 > M3 in the maize fields.



Figure 6. Mean flux of (**A**) CO₂; (**B**) CH₄; and (**C**) N₂O during the different periods in the rice fields. Bars indicate standard errors of the means (n = 3).



Freezing period Hard frost periodSnow melting period Thawing period

Figure 7. Mean flux of CO₂ (**A**); CH₄ (**B**); and N₂O (**C**) during different periods in maize fields. Bars indicate standard errors of the means (n = 3).

All of the rice and maize fields acted as CO_2 sources during winter. The mean emission fluxes of CO_2 during the different periods followed the order of freezing period > thawing period > snow-melting period > hard frost period in the three rice fields (Figure 6A) and followed the order of thawing period > freezing period > snow-melting period > hard frost period in the maize fields (Figure 7A).

Rice fields acted as important CH_4 sources during the freezing, snow-melting, and thawing periods, and as CH_4 sinks during the hard frost period (Figure 6B). The snow-melting period exhibited a higher flux of CH_4 as compared to the other periods in the rice fields. Maize fields were important CH_4 sinks in winter, except for a period of CH_4 release during the snow-melting period, and the freezing period showed the highest mean flux of CH_4 (Figure 7B).

Rice and maize fields acted as N_2O sources in winter, except for a relatively minor uptake of N_2O by the maize fields during the hard frost period (Figures 6C and 7C). The mean N_2O emission fluxes during the snow-melting period in all six fields were notably higher than those during other periods.

3.4. Cumulative Emissions, Contribution Rates, and GWPs of CO₂, CH₄, and N₂O

In general, the cumulative flux of CO₂, CH₄, and N₂O in winter followed the order: R1 > R2 > R3 and M1 > M2 > M3 (Table 2).

		Freezing Period	Hard Frost Period	Snow-Melting Period	Thawing Period	Total
CO₂ (kg·ha ⁻¹)	R1	1103.2	45.7	13.0	482.6	1644.5
	R2	893.7	46.2	7.9	319.7	1267.5
	R3	621.7	42.2	7.4	220.9	892.3
	M1	425.7	61.0	12.6	471.6	970.8
	M2	327.7	58.6	11.9	329.3	727.6
	M3	226.2	59.9	9.9	232.7	528.8
	R1	638.9	-174.8	356.0	183.5	1003.7
	R2	550.7	-170.8	320.0	144.9	844.9
CII (-h - 1)	R3	412.1	-182.2	270.9	109.5	610.4
CH ₄ (g·na ⁻¹)	M1	-437.1	-223.3	47.0	-158.1	-771.6
	M2	-344.4	-221.2	35.2	-116.5	-646.9
	M3	-304.4	-215.2	20.1	-84.6	-584.1
	R1	119.6	61.8	670.8	244.0	1096.1
	R2	96.4	51.0	498.3	166.9	812.5
$N \cap (-h - 1)$	R3	74.9	49.1	327.1	94.0	545.1
$N_2 O (g \cdot na^{-1})$	M1	78.6	-62.3	537.2	111.8	665.2
	M2	51.7	-62.9	397.8	75.8	462.5
	M3	32.8	-78.4	294.0	41.5	289.8

Table 2. Cumulative emissions of carbon dioxide (CO_2), methane (CH_4), and nitrous oxide (N_2O) in the rice and maize fields.

The total cumulative CO₂ emissions during winter in rice fields reached 1644.5 kg·ha⁻¹, 1267.5 kg·ha⁻¹, and 892.3 kg·ha⁻¹ in R1, R2, and R3, respectively (Table 2). The freezing period contributed 67.1%, 70.5%, and 69.7%, and the thawing period contributed 29.3%, 25.2%, and 24.8% of the total cumulative CO₂ emissions in R1, R2, and R3, respectively. The total cumulative CO₂ emission during winter in M1 was 970.8 kg·ha⁻¹, which was 243.3 kg·ha⁻¹ and 442.1 kg·ha⁻¹ higher than in M2 and M3, respectively. The freezing period associated with the thawing period contributed 92.4%, 90.3%, and 86.8% of the total cumulative CO₂ emissions in M1, M2, and M3, respectively.

The net cumulative CH₄ emissions during winter were 1003.7 g·ha⁻¹, 844.9 g·ha⁻¹, and 610.4 g·ha⁻¹ and the cumulative CH₄ uptake during the hard frost period were 174.8 g·ha⁻¹, 170.8 g·ha⁻¹, and 182.2 g·ha⁻¹ in R1, R2, and R3, respectively. The CH₄ emissions were concentrated during the freezing and snow-melting periods in the three rice fields. Cumulative CH₄ rates of 818.6 g·ha⁻¹, 682.1 g·ha⁻¹, and 604.2 g·ha⁻¹ were assimilated, and 47.0 g·ha⁻¹, 35.2 g·ha⁻¹, and 20.1 g·ha⁻¹ were released by M1, M2, and M3, respectively. Approximately 80.7%, 82.9%, and 86.0% of the CH₄ uptake rates were observed during the freezing and hard frost periods in M1, M2, and M3, respectively.

The net cumulative N₂O emissions during winter were 1096.1 g·ha⁻¹, 812.5 g·ha⁻¹, and 545.1 g·ha⁻¹ in R1, R2, and R3, respectively. The snow-melting period contributed 61.2%, 61.3%, and 60.0%, and the thawing period contributed 22.3%, 20.5%, and 17.2% of the total cumulative N₂O emissions in R1, R2, and R3, respectively. M1, M2, and M3 released a total of 727.5 g·ha⁻¹, 525.3 g·ha⁻¹, and 368.2 g·ha⁻¹ N₂O during winter, and assimilated 62.3 g·ha⁻¹, 62.9 g·ha⁻¹, and 78.4 g·ha⁻¹, N₂O during the hard frost period, respectively. The snow-melting period dominated the N₂O emissions in winter in the maize fields.

The GWP values were 1996.2 kg CO₂ eq· ha⁻¹, 1530.8 kg CO₂ eq· ha⁻¹, 1070.0 kg CO₂ eq· ha⁻¹, 1149.8 kg CO₂ eq· ha⁻¹, 849.2 kg CO₂ eq· ha⁻¹, and 600.5 kg CO₂ eq· ha⁻¹ in R1, R2, R3, M1, M2, and M3, respectively (Table 3).

Table 3. Global warming potentials of all fields during winter.

	R1	R2	R3	M1	M2	M3
GWP (kg CO_2 eq ha ⁻¹)	1996.2	1530.8	1070.0	1149.8	849.2	600.5

3.5. Pearson Correlation Analysis between CO_2 , CH_4 , and N_2O Flux and Temperatures during Winter

The CH₄ emission flux in rice fields was significantly related to the temperatures of the soil at the surface and at a depth of 15 cm (P < 0.05; Table 4). The CH₄ emission flux in rice fields was significantly related to the temperatures of the soil at the surface and at a depth of 15 cm. CH₄ uptake rate was notably (P < 0.01) related to the soil surface temperature, whereas it was not related to the soil temperature at a 15-cm depth. The N₂O flux showed no significant correlation with the temperature of soil either at the surface or at a 15-cm depth (Table 4).

	Temperature Location	R1	R2	R3	M1	M2	M3
CO ₂	Surface Soil	0.840 **	0.784 **	0.789 **	0.927 **	0.923 **	0.929 **
	15 cm Depth	0.401 **	0.336 **	0.341 **	0.708 **	0.682 **	0.693 **
CH ₄	Surface Soil	0.361 **	0.339 **	0.308 *	-0.602 **	-0.552 **	-0.463 **
	15 cm Depth	0.309 *	0.295 *	0.313 *	-0.151	-0.101	0.003
N ₂ O	Surface Soil 15 cm Depth	$-0.182 \\ 0.164$	$-0.187 \\ 0.144$	$-0.182 \\ 0.135$	-0.217 0.118	-0.22 0.113	-0.224 0.111

Table 4. Pearson correlation analysis between CO₂, CH₄, and N₂O flux and temperatures during winter.

Note: * significant at the 0.05 level; ** significant at the 0.01 level.

4. Discussion

4.1. CO₂ Flux

Temperature is one of the key factors affecting the production and emission of CO_2 [40]. The soil temperature, both at the surface and at a depth of 15 cm, was significantly (P < 0.01) related to CO_2 emission in the six fields. Microbial biomass decreased with decreasing temperatures [41,42], and freezing soil insulated O_2 from the air, restraining aerobic respiration and the release of CO_2 . Furthermore, increasing solubility and adsorption of CO_2 with the decrease in temperature led to its retention [43]. Therefore, CO₂ flux decreased gradually during the freezing period and remained at the lowest level during the hard frost period. Although soil temperatures at the surface and a depth of 15 cm (<0 °C) during the snow-melting period increased somewhat, they both remained around 0 °C. As the subsurface soil was frozen, little CO_2 was released during the snow-melting period. Previous studies showed that dissolved organic matter was released from the broken aggregates [44-46] during freeze-thaw cycles and freezing [46], as well as from the lyzed microbial cells at low temperature [42]. The readily available sources of dissolved organic matter supplied a massive carbon source for recovering microbes and stimulated microbial respiration [42], until the sources were exhausted. Therefore, CO₂ emission peaks occurred during the early thawing period in M1, M2, R1, R2, whereas R3, and M3 did not exhibit obvious CO_2 emission peaks during the early thawing period, possibly because of the high salinity conditions in R3 and M3 restrained microbial activities [33], reduced microbial biomass [47], and decelerated the consumption of readily available dissolved organic matter. Thereafter, microbial activities increased and soil thawed with the rising temperatures in the six fields, leading to an increased CO₂ emission flux.

Soil organic carbon is the substrate used for microbial respiration and is therefore vital to the process. Low soil organic carbon content is an indicator of low microbial activity and of low CO₂ emissions [48]. Furthermore, a higher salinity resulted in a smaller microbial biomass [48] and inhibited the decomposition [49] and transformation [50] of organic matter. The high pH in alkaline soil facilitated CO₂ absorption intensity and reduced the CO₂ emission flux [51]. Results showed that CO₂ fluxes with pH 7.6–7.8 were significantly lower than with soil with a pH of below 7.0 [52]. Annual CO₂ fluxes decreased with increasing pH in the range of 7.37–8.72 and were negative with a pH greater than 9 [53]. Consequently, the emission flux and cumulative emissions of CO₂ followed the decreasing order of R1, R2, then R3 and M1, M2, then M3.

4.2. CH_4 Flux

CH₄ is produced by methanogen under strict anaerobic conditions [54]. The anoxic conditions caused by high moisture resulted in the rice fields becoming CH₄ sources.

The CH_4 emission flux in rice fields was significantly related to the soil temperature, both at the surface and at 15-cm depth, which is consistent with the conclusion that the release of CH_4 is temperature-sensitive and that lower temperatures result in lower CH_4 emissions [55,56].

The two CH₄ emission peaks occurred during the early freezing period (22 October 2014 to 5 November 2014) in every rice field, perhaps because the surface soil temperature dropped below 0 °C before every peak. The freezing of the soil surface caused by the low air temperature strengthened the anoxic condition and promoted the production of CH₄, which was emitted when the temperature increased to above zero. Furthermore, freeze-thaw cycles occurring around the second peak had a positive effect on CH₄ emissions. Thereafter, CH₄ production and emissions were inhibited by the lower temperatures and frozen soil.

The CH₄ accumulated under extremely anoxic conditions in the frozen soil during the hard frost period, and was emitted when the soil thawed during the snow-melting period. In addition, the topsoil thawed first, but the deeper soil remained frozen and was not permeable to water. So, fresh melt water fully saturated a small top layer of soil of several centimeters, creating anaerobic conditions favorable to produce of CH₄. Thereby, CH₄ emission peaks were observed in the three rice fields during the snow-melting period. Song et al. similarly noted explosive CH₄ emissions during the freeze-thaw period [39]. After the complete release of accumulated CH₄ in the three rice fields, the CH₄ emission flux decreased and then increased with the rising temperature. The CH₄ fluxes in the three rice fields declined after the 24 April 2015, possibly because the aerobic condition that increased with the evaporation of water restrained the production of CH₄.

The CH₄ uptake rates in the maize fields began to decline from the early freezing period. CH₄ uptake rates were notably (P < 0.01) related to the surface soil temperature, whereas they were not related to the soil temperature at a depth of 15 cm. This corresponds to methane oxidation being sensitive to air temperature [57] as methane oxidation occurs mainly in the surface soil. The observed CH₄ emission peaks in the maize fields during the snow-melting period occurred for the same reason as those in the rice fields. CH₄ uptake rates in the three maize fields increased with the increasing soil surface temperature during the thawing period, which was consistent with the previous observed results in the saline-alkali soil in arid and semi-arid regions [58,59].

The higher moisture in the rice fields might result in deeper frost depths and harder frost than in maize fields, and the rice fields required more time to thaw, taking more time to release accumulated CH_4 as compared to the maize fields at the same soil depth. Therefore, CH_4 emission peaks in the rice fields during the snow-melting period occurred later than those in the maize fields.

The CH₄ uptake peak in maize fields (5 November 2014) during the early freezing period, the increase in CH₄ uptake rates in the maize fields, the period of CH₄ uptake in the rice fields during the late freezing period, and the period of CH₄ uptake during the hard frost period in the six fields, might all be related to the living methane-oxidizing bacteria and methane adsorption by the snow layer [60,61]. However, the physicochemical mechanisms responsible remain unclear. A sudden jump in CH₄ uptake in the maize fields in early December was likely because of the inhibition caused by lower temperatures in the CH₄ uptake by the living methane-oxidizing bacteria and the approximately saturated CH₄ in the snow.

Previous studies have shown that the optimum pH of methanogen is 6.9-7.2 [62]; a higher pH (>7.2) would result in stronger inhibition of CH₄ production. In addition, studies have shown that CH₄ emissions are negatively related to salinity [63,64]. CH₄ is the final product of soil organic carbon under anaerobic decomposition. Higher soil organic carbon content positively affected CH₄ emissions [65,66]. Therefore, the emission flux and the accumulated CH₄ emission in the rice fields followed the decreasing order of R1, R2, and then R3.

The optimum pH for methane oxidation has been reported to be 7.0–7.5 [67]. A rise in pH above the optimal level would result in a decline in methane oxidation. Furthermore, high salinity levels inhibited the activity of methane-oxidizing bacteria [68,69], inhibiting the CH_4 uptake [70]. Hence, M1 exhibited higher CH_4 uptake rates and a higher cumulative CH_4 uptake than those of M2 and M3.

4.3. N₂O Flux

 N_2O emissions showed no significant correlation with the soil temperatures either at the surface or at a 15-cm depth, which was consistent with the findings of Koponen et al. who found that N_2O emissions were not related to soil temperature [71]. The flux of N_2O is driven by its rates of production and consumption. The soil acted as a N_2O sink when the N_2O consumption rate exceeded that of its production [72]. Goldberg et al. similarly found that soil acted as a N_2O sink under arid and aerobic conditions [73]. The maize fields maintained relatively arid and aerobic conditions as compared to the rice fields, which likely explains why the maize fields acted as weak N_2O sinks, whereas the rice fields were sources of N_2O during the hard frost period.

 N_2O is mainly derived from nitrification under aerobic conditions and denitrification under anaerobic conditions. The anaerobic and aerobic conditions occurred consecutively in the soil under the freeze-thaw cycles, thereby promoting concurrent nitrification and denitrification. However, the results of the current study suggest that denitrification mainly contributed to the N_2O emissions during freeze-thaw cycles [74]. The large amount of NO_3^- that accumulated during the lengthy winter [75] stimulated denitrification. In addition, the topsoil thawed first, but the deeper soil remained frozen and was not permeable to water. So, fresh melt water fully saturated a small top layer of soil, creating anaerobic conditions favorable for denitrification. Hence, N_2O emission peaks were observed in the six fields during the snow-melting period and the initial few days of the thawing period. The higher moisture in rice fields led to longer anaerobic condition than in the maize fields that contained lower moisture, thus facilitating denitrification [74] and the longer duration of explosive N_2O emissions in rice fields. Thereafter, aerobic conditions in the six fields were promoted with the evaporation of water, which decreased N_2O emission rates [76].

High soil organic carbon content and low pH favors N_2O production [77]. High NO NO_3^- content also stimulates denitrification and N_2O production [78]. High salinity inhibits nitrification [79]. Hence, accumulated N_2O emissions during winter followed the decreasing order of R1, R2, then R3 and M1, M2, then M3.

5. Conclusions

The rice and maize fields acted as CO_2 sources in winter, and the freezing period combined with the thawing period dominated the cumulative CO_2 emissions. Rice fields were CH_4 sources in winter, and the freezing and snow-melting periods dominated the CH_4 emissions. However, maize fields were CH_4 sinks during winter, with 85.6–89.0% of the CH_4 emissions occurring in the freezing and hard frost periods. Both rice and maize fields acted as N₂O sources, and the snow-melting period dominated the N₂O emissions. Rice fields contributed 1070.0–1996.2 kg CO_2 eq ha⁻¹ to the GWP in the winter. Maize fields contributed 600.5–1149.8 kg CO_2 eq ha⁻¹ to the GWP in the winter. High pH and salinity negatively affected CO_2 , CH_4 , and N₂O emissions, CH_4 uptake, and GWP during winter.

Acknowledgments: We gratefully acknowledge the Foundation of National Natural Science Foundation of China under Grant (51179073, 41471152).

Author Contributions: Hao Zhang, Jie Tang and Shuang Liang drafted the manuscript and tested the samples, Zhaoyang Li designed the research. The others analyzed the data and collected the samples.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. IPCC. Climate Change 2013: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2013; p. 1535.
- Kong, Y.; Nagano, H.; Kátai, J.; Vágó, I.; Oláh, Á.Z.; Yashima, M.; Inubushi, K. CO₂, N₂O and CH₄ production/consumption potentials of soils under different land-use types in central Japan and eastern Hungary. *Soil Sci. Plant Nutr.* 2013, *59*, 455–462. [CrossRef]
- Solomon, S.; Qin, D.; Manning, M.; Chen, Z.; Marquis, M.; Averyt, K.B.; Tignor, M.; Miller, H.L. Climate Change 2007: The Physical Science Basis, Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change, 1st ed.; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2007; pp. 2–18.
- Friedlingstein, P.; Cox, P.; Betts, R.; Bopp, L.; Von Bloh, W.; Brovkin, V.; Cadule, P.; Doney, S.; Eby, M.; Fung, I.; et al. Climate-carbon cycle feedback analysis, results from the C⁴MIP model intercomparison. *J. Clim.* 2006, 19, 3337–3353. [CrossRef]
- Smith, P.; Martino, D.; Cai, Z.; Gwary, D.; Janzen, H.; Kumar, P.; McCarl, B.; Ogle, S.; O'Mara, F.; Rice, C.; et al. Greenhouse gas mitigation in agriculture. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 2008, 363, 789–813. [CrossRef] [PubMed]
- 6. Ussiri, D.A.N.; Lal, R.; Jarecki, M.K. Nitrous oxide and methane emissions from long-term tillage under a continuous corn cropping system in Ohio. *Soil Tillage Res.* **2009**, *104*, 247–255. [CrossRef]
- Liu, X.J.; Mosier, A.R.; Halvorson, A.D.; Reule, C.A.; Zhang, F.S. Dinitrogen and N₂O emissions in arable soils: Effect of tillage, N source and soil moisture. *Soil Biol. Biochem.* 2007, *39*, 2362–2370. [CrossRef]
- Omonode, R.A.; Vyn, T.J.; Smith, D.R.; Hegymegi, P.; Gál, A. Soil carbon dioxide and methane fluxes from long-term tillage systems in continuous corn and corn-soybean rotations. *Soil Tillage Res.* 2007, 95, 182–195. [CrossRef]
- 9. Hwang, H.Y.; Kim, G.W.; Kim, S.Y.; Mozammel Haque, M.; Khan, M.I.; Kim, P.J. Effect of cover cropping on the net global warming potential of rice paddy soil. *Geoderma* **2017**, *292*, 49–58. [CrossRef]
- Bhattacharyya, P.; Nayak, A.K.; Mohanty, S.; Tripathi, R.; Shahid, M.; Kumar, A.; Raja, R.; Panda, B.B.; Roy, K.S.; Neogi, S.; et al. Greenhouse gas emission in relation to labile soil C, N pools and functional microbial diversity as influenced by 39 years long-term fertilizer management in tropical rice. *Soil Tillage Res.* 2013, 129, 93–105. [CrossRef]
- 11. Zhang, X.X.; Yin, S.; Li, Y.S.; Zhuang, H.L.; Li, C.H.; Liu, C.J. Comparison of greenhouse gas emissions from rice paddy fields under different nitrogen fertilization loads in Chongming Island, Eastern China. *Sci. Total Environ.* **2014**, *472*, 381–388. [CrossRef] [PubMed]
- 12. Maris, S.C.; Teira-Esmatges, M.R.; Arbonés, A.; Rufat, J. Effect of irrigation, nitrogen application, and a nitrification inhibitor on nitrous oxide, carbon dioxide and methane emissions from an olive (*Olea europaea* L.) orchard. *Sci. Total Environ.* **2015**, *538*, 966–978. [CrossRef] [PubMed]
- 13. Shah, A.; Lamers, M.; Streck, T. N₂O and CO₂ emissions from South German arable soil after amendment of manures and composts. *Environ. Earth Sci.* **2016**, *75*, 1–12. [CrossRef]
- Bhattacharyya, P.; Roy, K.S.; Neogi, S.; Dash, P.K.; Nayak, A.K.; Mohanty, S.; Baig, M.J.; Sarkar, R.K.; Rao, K.S. Impact of elevated CO₂ and temperature on soil C and N dynamics in relation to CH₄ and N₂O emissions from tropical flooded rice (*Oryza sativa* L.). *Sci. Total Environ.* **2013**, 461–462, 601–611. [CrossRef] [PubMed]
- Jiang, B.; Yang, S.Y.; Yang, X.B.; Ma, Y.H.; Chen, X.L.; Zuo, H.F.; Fan, D.F.; Gao, L.; Yu, Q.; Yang, W. Effect of controlled drainage in the wheat season on soil CH₄ and N₂O emissions during the rice season. *Int. J. Plant Prod.* 2015, *9*, 273–290.
- 16. Wang, G.S.; Liang, Y.P.; Zhang, Q.; Jha, S.K.; Gao, Y.; Shen, X.J.; Sun, J.S.; Duan, A.W. Mitigated CH₄ and N₂O emissions and improved irrigation water use efficiency in winter wheat field with surface drip irrigation in the North China Plain. *Agric. Water Manag.* **2016**, *163*, 403–407. [CrossRef]
- 17. Wang, Y.Y.; Hu, C.S.; Ming, H.; Oenema, O.; Schaefer, D.A.; Dong, W.X.; Zhang, Y.M.; Li, X.X. Methane, carbon dioxide and nitrous oxide fluxes in soil profile under a winter wheat-summer maize rotation in the North China Plain. *PLoS ONE* **2014**, *9*, e98445. [CrossRef] [PubMed]
- 18. Tang, J.; Liang, S.; Li, Z.Y.; Zhang, H.; Wang, S.N.; Zhang, N. Emission laws and influence factors of greenhouse gases in Saline-Alkali paddy fields. *Sustainability* **2016**, *8*, 163. [CrossRef]

- 19. Song, C.C.; Wang, Y.S.; Wang, Y.Y.; Zhao, Z.C. Emission of CO₂, CH₄ and N₂O from freshwater marsh during freeze-thaw period in Northeast of China. *Atmos. Environ.* **2006**, *40*, 6879–6885. [CrossRef]
- Tang, H.; Xiao, X.; Tang, W.; Wang, K.; Sun, J.; Li, W.; Yang, G. Effects of winter covering crop residue incorporation on CH₄ and N₂O emission from double-cropped paddy fields in southern China. *Environ. Sci. Pollut. Res.* 2015, 22, 12689–12698. [CrossRef] [PubMed]
- 21. Merbold, L.; Steinlin, C.; Hagedorn, F. Winter greenhouse gas fluxes (CO₂, CH₄ and N₂O) from a subalpine grassland. *Biogeosciences* **2013**, *10*, 3185–3203. [CrossRef]
- 22. Hao, Q.J.; Wang, Y.S.; Song, C.C.; Huang, Y. Contribution of winter fluxes to the annual CH₄, CO₂ and N₂O emissions from freshwater marshes in the Sanjiang Plain. *J. Environ. Sci.* **2006**, *18*, 270–275.
- 23. Kurganova, I.N.; Lopes de Gerenyu, V.O. Contribution of abiotic factors to CO₂ emission from soils in the freeze–thaw cycles. *Eurasian Soil Sci.* **2015**, *48*, 1009–1015. [CrossRef]
- 24. Liang, W.; Shu, Y.; Zhang, H.; Yue, J.; Huang, G.H. Greenhouse Gas Emissions from Northeast China Rice Fields in Fallow Season. *Pedosphere* **2007**, *17*, 630–638. [CrossRef]
- 25. Sehy, U.; Dyckmans, J.; Ruser, R.; Munch, J.C. Adding dissolved organic carbon to simulate freeze–thaw related N₂O emissions from soil. *J. Plant Nutr. Soil Sci.* **2004**, *167*, 471–478. [CrossRef]
- 26. Mastepanov, M.; Sigsgaard, C.; Dlugokencky, E.J.; Houweling, S.; Ström, L.; Tamstorf, M.P.; Christensen, T.R. Large tundra methane burst during onset of freezing. *Nature* **2008**, *456*, 628–630. [CrossRef] [PubMed]
- 27. Liu, X.; Qi, Z.M.; Wang, Q.; Ma, Z.W.; Li, L.H. Effects of biochar addition on CO₂ and CH₄ emissions from a cultivated sandy loam soil during freeze-thaw cycles. *Plant Soil Environ.* **2017**, *63*, 243–249.
- 28. Kurganova, I.N.; Lopes de Gerenyu, V.O. Effect of the temperature and moisture on the N₂O emission from some arable soils. *Eurasian Soil Sci.* **2010**, *43*, 919–928. [CrossRef]
- Pelster, D.E.; Chantigny, M.H.; Rochette, P.; Angers, D.A.; Laganiere, J.; Zebarth, B.J.; Goyer, C. Crop residue incorporation alters soil nitrous oxide emissions during freeze–thaw cycles. *Can. J. Soil Sci.* 2013, 93, 415–425. [CrossRef]
- Bandibas, J.; Vermoesen, A.; Degroot, C.J.; Vancleemput, O. The effect of different moisture regimes and soil characteristics on nitrous oxide emission and consumption by different soils. *Soil Sci.* 1994, 158, 106–114. [CrossRef]
- 31. Nyborg, M.; Laidlaw, J.W.; Solberg, E.D.; Malhi, S.S. Denitrification and nitrous oxide emissions from a Black Chernozemic soil during spring thaw in Alberta. *Can. J. Soil Sci.* **1997**, *77*, 153–160. [CrossRef]
- 32. Pankhurst, C.E.; Yu, S.; Hawke, B.G.; Harch, B.D. Capacity of fatty acid profiles and substrate utilization patterns to describe differences in soil microbial communities associated with increased salinity or alkalinity at three locations in South Australia. *Biol. Fertil. Soils* **2001**, *33*, 204–217. [CrossRef]
- 33. Sardinha, M.; Müller, T.; Schmeisky, H.; Joergensen, R.G. Microbial performance in soils along a salinity gradient under acidic conditions. *Appl. Soil Ecol.* **2003**, *23*, 237–244. [CrossRef]
- 34. Pathak, H.; Rao, D.L.N. Carbon and nitrogen mineralization from added organic matter in saline and alkali soils. *Soil Biol. Biochem.* **1998**, *30*, 695–702. [CrossRef]
- 35. Batra, L.; Manna, M.C. Dehydrogenase activity and microbial biomass carbon in salt-affected soils of semiarid and arid regions. *Arid Soil Res. Rehabil.* **1997**, *11*, 295–303. [CrossRef]
- 36. Tang, J.; Liang, S.; Li, Z.; Zhang, H.; Lou, Y.; Wang, J. Effect of freeze-thaw cycles on carbon stocks of saline–alkali paddy soil. *Arch. Agron. Soil Sci.* **2016**, *62*, 1640–1653. [CrossRef]
- Bissett, A.; Abell, G.C.J.; Bodrossy, L.; Richardson, A.E.; Thrall, P.H. Methanotrophic communities in Australian woodland soils of varying salinity. *FEMS Microbiol. Ecol.* 2012, *80*, 685–695. [CrossRef] [PubMed]
- Lin, X.W.; Wang, S.P.; Ma, X.Z.; Xu, G.P.; Luo, C.Y.; Li, Y.N.; Jiang, G.M.; Xie, Z.B. Fluxes of CO₂, CH₄, and N₂O in an alpine meadow affected by yak excreta on the Qinghai-Tibetan plateau during summer grazing periods. *Soil Biol. Biochem.* 2009, 41, 718–725. [CrossRef]
- 39. Zhang, J.B.; Song, C.C.; Yang, W.Y. Cold season CH₄, CO₂ and N₂O fluxes from freshwater marshes in northeast China. *Chemosphere* **2005**, *59*, 1703–1705. [CrossRef] [PubMed]
- 40. Li, H.J.; Yan, J.X.; Yue, X.F.; Wang, M.B. Significance of soil temperature and moisture for soil respiration in a Chinese mountain area. *Agric. For. Meteorol.* **2008**, *148*, 490–503. [CrossRef]
- 41. Zhang, H.; Tang, J.; Liang, S. Effects of snow cover plus straw mulching on microorganisms in paddy soil during winter. *Appl. Soil Ecol.* **2017**, *119*, 339–344. [CrossRef]
- 42. Larsen, K.S.; Jonasson, S.; Michelsen, A. Repeated freeze-thaw cycles and their effects on biological processes in two arctic ecosystem types. *Appl. Soil Ecol.* **2002**, *21*, 187–195. [CrossRef]

- 43. Certini, G.; Scalenghe, R. *Soils: Basic Concepts and Future Challenges*; Cambridge University Press: Cambridge, UK; New York, NY, USA, 2006; pp. 75–85.
- Groffman, P.M.; Driscoll, C.T.; Fahey, T.J.; Hardy, J.P.; Fitzhugh, R.D.; Tierney, G.L. Effects of mild winter freezing on soil nitrogen and carbon dynamics in a northern hardwood forest. *Biogeochemistry* 2001, 56, 191–213. [CrossRef]
- 45. Bechmann, M.E.; Kleinman, P.J.A.; Sharpley, A.N.; Saporito, L.S. Freeze–Thaw Effects on Phosphorus Loss in Runoff from Manured and Catch-Cropped Soils. *J. Environ. Qual.* **2005**, *34*, 2301–2309. [CrossRef] [PubMed]
- Fuss, C.B.; Driscoll, C.T.; Groffman, P.M.; Campbell, J.L.; Christenson, L.M.; Fahey, T.J.; Fisk, M.C.; Mitchell, M.J.; Templer, P.H.; Durán, J.; et al. Nitrate and dissolved organic carbon mobilization in response to soil freezing variability. *Biogeochemistry* 2016, 131, 35–47. [CrossRef]
- 47. Tripathi, S.; Kumari, S.; Chakraborty, A.; Gupta, A.; Chakrabarti, K.; Bandyapadhyay, B.K. Microbial biomass and its activities in salt-affected coastal soils. *Biol. Fertil. Soils* **2006**, *42*, 273–277. [CrossRef]
- 48. Yuan, B.C.; Li, Z.Z.; Liu, H.; Gao, M.; Zhang, Y.Y. Microbial biomass and activity in salt affected soils under arid conditions. *Appl. Soil Ecol.* 2007, *35*, 319–328. [CrossRef]
- 49. Wichern, J.; Wichern, F.; Joergensen, R.G. Impact of salinity on soil microbial communities and the decomposition of maize in acidic soils. *Geoderma* **2006**, *137*, 100–108. [CrossRef]
- 50. Mamilov, A.; Dilly, O.M.; Mamilov, S.; Inubushi, K. Microbial eco-physiology of degrading Aral Sea wetlands: Consequences for C-cycling. *Soil Sci. Plant Nutr.* **2004**, *50*, 839–842. [CrossRef]
- 51. Xie, J.X.; Li, Y.; Zhai, C.X.; Li, C.H.; Lan, Z.D. CO₂ absorption by alkaline soils and its implication to the global carbon cycle. *Environ. Geol.* **2009**, *56*, 953–961. [CrossRef]
- 52. Brummell, M.E.; Farrell, R.E.; Siciliano, S.D. Greenhouse gas soil production and surface fluxes at a high arctic polar oasis. *Soil Biol. Biochem.* **2012**, *52*, 1–12. [CrossRef]
- 53. Wang, W.F.; Chen, X.; Pu, Z.; Yuan, X.L.; Ma, J.L. Negative soil respiration fluxes in unneglectable arid regions. *Polish J. Environ. Stud.* **2016**, *24*, 905–908. [CrossRef]
- 54. Houghton, J.T.; Ding, Y.; Griggs, D.J.; Noguer, M.; van der Linden, P.J.; Dai, X.; Maskell, M.; Johnson, C.A. *Climate Change 2001: The Scientific Basis*; Cambridge University Press: Cambridge, UK, 2001.
- 55. Keppler, F.; Hamilton, J.T.; Brass, M.; Rockmann, T. Methane emissions from terrestrial plants under aerobic conditions. *Nature* **2006**, *439*, 187–191. [CrossRef] [PubMed]
- 56. Fey, A.; Conrad, R. Effect of temperature on carbon and electron flow and on the archaeal community in methanogenic rice field soil. *Appl. Environ. Microbiol.* **2000**, *66*, 4790–4797. [CrossRef] [PubMed]
- 57. Zhuang, Q.L.; Chen, M.; Xu, K.; Tang, J.Y.; Saikawa, E.; Lu, Y.Y.; Melillo, J.M.; Prinn, R.G.; McGuire, A.D. Response of global soil consumption of atmospheric methane to changes in atmospheric climate and nitrogen deposition. *Glob. Biogeochem. Cycles* **2013**, *27*, 650–663. [CrossRef]
- 58. Sjogersten, S.; Wookey, P.A. Spatio-temporal variability and environmental controls of methane fluxes at the forest-tundra ecotone in the Fennoscandian Mountains. *Glob. Chang. Biol.* **2002**, *8*, 885–894. [CrossRef]
- 59. Hart, S.C. Potential impacts of climate change on nitrogen transformations and greenhouse gas fluxes in forests: A soil transfer study. *Glob. Chang. Biol.* **2006**, *12*, 1032–1046. [CrossRef]
- 60. Kerbrat, M.; Pinzer, B.; Huthwelker, T.; Gäggeler, H.W.; Ammann, M.; Schneebeli, M. Measuring the specific surface area of snow with X-ray tomography and gas adsorption: Comparison and implications for surface smoothness. *Atmos. Chem. Phys. Discuss.* **2007**, *7*, 10287–10322. [CrossRef]
- 61. Smagin, A.V.; Shnyrev, N.A. Methane fluxes during the cold season: Distribution and mass transfer in the snow cover of bogs. *Eurasian Soil Sci.* **2015**, *48*, 823–830. [CrossRef]
- 62. Wang, Z.P.; Lindau, C.W.; Delaune, R.D.; Patrick, W.H.J. Methane emission and entrapment in flooded rice soils as affected by soil properties. *Biol. Fertil. Soils* **1993**, *16*, 163–168. [CrossRef]
- 63. Denier van der Gon, H.A.C.; Neue, H.U. Methane emission from a wetland rice field as affected by salinity. *Plant Soil* **1995**, *170*, 307–313. [CrossRef]
- 64. Magenheimer, J.F.; Moore, T.R.; Chmura, G.L.; Daoust, R.J. Methane and carbon dioxide flux from a macrotidal salt marsh, Bay of Fundy, New Brunswick. *Estuaries* **1996**, *19*, 139–145. [CrossRef]
- 65. Menéndez, S.; Barrena, I.; Setien, I.; González-Murua, C.; Estavillo, J.M. Efficiency of nitrification inhibitor DMPP to reduce nitrous oxide emissions under different temperature and moisture conditions. *Soil Biol. Biochem.* **2012**, *53*, 82–89. [CrossRef]
- 66. Datta, A.; Santra, S.C.; Adhya, T.K. Effect of inorganic fertilizers (N, P, K) on methane emission from tropical rice field of India. *Atmos. Environ.* **2013**, *66*, 123–130. [CrossRef]

- 67. Smith, N.R.; Kishchuk, B.E.; Mohn, W.W. Effects of wildfire and harvest disturbances on forest soil bacterial communities. *Appl. Environ. Microbiol.* **2008**, *74*, 216–224. [CrossRef] [PubMed]
- Osudar, R.; Matoušů, A.; Alawi, M.; Wagner, D.; Bussmann, I. Environmental factors affecting methane distribution and bacterial methane oxidation in the German Bight (North Sea). *Estuar. Coast. Shelf Sci.* 2015, *160*, 10–21. [CrossRef]
- Serrano-Silva, N.; Valenzuela-Encinas, C.; Marsch, R.; Dendooven, L.; Alcántara-Hernández, R.J. Changes in methane oxidation activity and methanotrophic community composition in saline alkaline soils. *Extremophiles* 2014, 18, 561–571. [CrossRef] [PubMed]
- 70. Chambers, L.G.; Reddy, K.R.; Osborne, T.Z. Short-Term Response of Carbon Cycling to Salinity Pulses in a Freshwater Wetland. *Soil Sci. Soc. Am. J.* 2011, *75*, 2000–2007. [CrossRef]
- 71. Koponen, H.T.; Flöjt, L.; Martikainen, P.J. Nitrous oxide emissions from agricultural soils at low temperatures: A laboratory microcosm study. *Soil Biol. Biochem.* **2004**, *36*, 757–766. [CrossRef]
- 72. Chapuis-lardy, L.; Wrage, N.; Metay, A.; Chotte, J.L.; Bernoux, M. Soils, a sink for N₂O? A review. *Glob. Chang. Biol.* 2007, 13, 1–17. [CrossRef]
- 73. Goldberg, S.D.; Gebauer, G. N₂O and NO fluxes between a Norway spruce forest soil and atmosphere as affected by prolonged summer drought. *Soil Biol. Biochem.* **2009**, *41*, 1986–1995. [CrossRef]
- 74. Ludwig, B.; Wolf, I.; Teepe, R. Contribution of nitrification and denitrification to the emission of N₂O in a freeze-thaw event in an agricultural soil. *J. Plant Nutr. Soil Sci.* **2004**, *167*, 678–684. [CrossRef]
- Zhang, X.R.; Bai, W.; Gilliam, F.S.; Wang, Q.D.; Han, X.; Li, L. Effects of in situ freezing on soil net nitrogen mineralization and net nitrification in fertilized grassland of northern China. *Grass Forage Sci.* 2011, 66, 391–401. [CrossRef]
- Khalil, K.; Renault, P.; Mary, B. Effects of transient anaerobic conditions in the presence of acetylene on subsequent aerobic respiration and N₂O emission by soil aggregates. *Soil Biol. Biochem.* 2005, 37, 1333–1342. [CrossRef]
- Cai, Y.J.; Ding, W.X.; Zhang, X.L.; Yu, H.Y.; Wang, L.F. Contribution of Heterotrophic Nitrification to Nitrous Oxide Production in a Long-Term N-Fertilized Arable Black Soil. *Commun. Soil Sci. Plant Anal.* 2010, 41, 2264–2278. [CrossRef]
- Mulvaney, R.L.; Khan, S.A.; Mulvaney, C.S. Nitrogen fertilizers promote denitrification. *Biol. Fertil. Soils* 1997, 24, 211–220. [CrossRef]
- 79. Badía, D. Potential nitrification rates of semiarid cropland soils from the Central Ebro Valley, Spain. *Arid Soil Res. Rehabil.* **2000**, *14*, 281–292. [CrossRef]



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).