

Supplementary material

Individual pattern response to CO₂-induced acidification stress in *Haliotis rufescens* suggests stage-specific acclimatization during its early life history

Ricardo Gómez-Reyes ¹, Clara E. Galindo-Sánchez ², Fabiola Lafarga-De la Cruz ², José M. Hernández-Ayón ², Enrique Valenzuela-Wood ³ and Laura López-Galindo ^{3*}

To achieve the control of pH in seawater, a set of three water flow systems was assembled (Figure S1). Excluding the control bioassay (pH 8.0), independent pH controllers (AquaMedic, Germany) were calibrated using the National Bureau of Standards (NBS) and configured to maintain the water at the desired pH during the experiment to moderate (7.8) and low (7.6) pH levels, respectively. Pure CO₂ was injected using a ceramic CO₂ diffuser and mixed between the reservoirs (100 L) to drop and maintain the experimental water pH. The experimental seawater used during the bioassays was filtered (1 μm nominal-sized filter cartridges) and ultra-violet (UV) sterilized; the flux rate was maintained at 100 mL min⁻¹ in the experimental tanks. Additionally, a complete drain-off and refill with new experimental filtered seawater (FSW) were performed twice from the second reservoir during the bioassay. The FSW temperature was maintained (16 ± 0.6, mean ± S.E.) by setting the laboratory air conditioner to 16 °C.

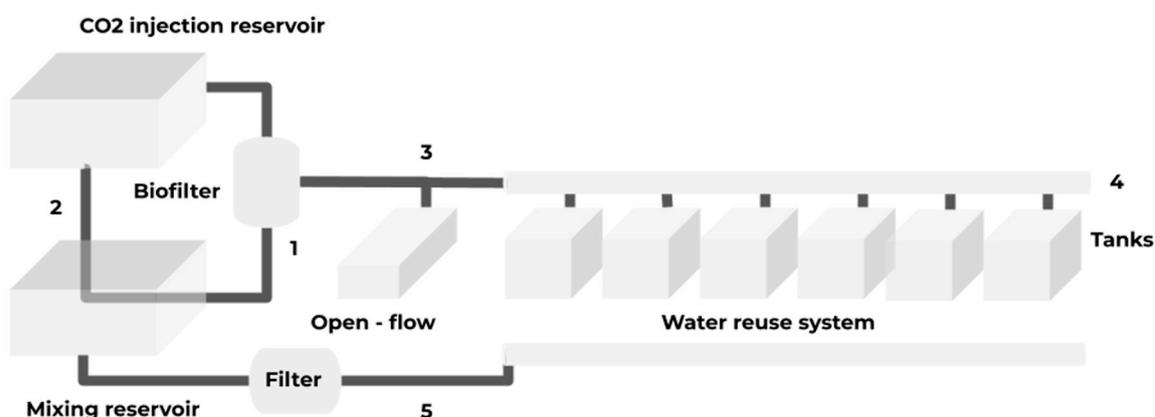


Figure S1. CO₂ injection system designed for the acidification with moderate (7.8) and low pH (7.6) challenges. These systems may contribute to water quality stabilization by managing the removal of solids and metabolism by-products (i.e., biofiltration) during larval development, including ammonia, and enabling control over gas transfer (i.e., pCO₂) and temperature. Numbers indicate water circulation. (1) Water is pumped from the mixing reservoir to the biofilter and CO₂ injection reservoir; then, (2) CO₂-mixed water goes back from the CO₂ injection reservoir to the mixing reservoir (20 L min⁻¹). (3) Before hatching, the eggs are challenged to acidified water in a slow current open-flow system (5 mL min⁻¹) maintained to the desired pH. (4) The CO₂-mixed water stream flows through six tanks (30 mL min⁻¹, 28 × 11 × 10 cm), where

the hatched larvae are placed until the treatment ends. (5) Additional baffles and sieves with mesh sizes from 100 to 150 μm are included in the tanks to prevent the larvae from escaping when the reused water returns to the mixing reservoir system during the experiment.

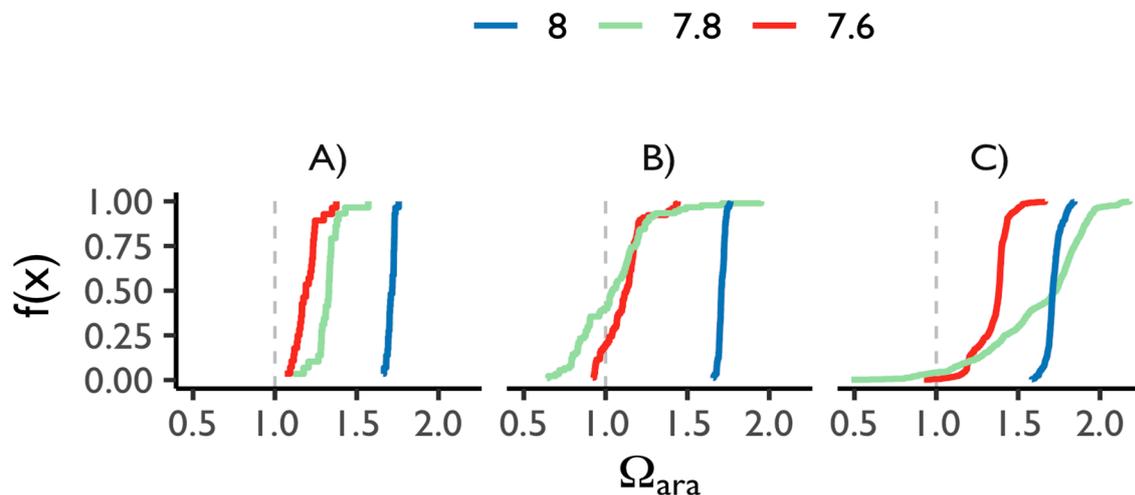


Figure S2. Empirical cumulative distribution (y axis) of predicted saturation levels of CaCO_3 with respect to aragonite (x axis) values obtained during the bioassay with the following pH conditions: control (pH 8.0, blue line), moderate (pH 7.8, green line) and low (pH 7.6, red line). (A) Before hatching, (B) during veliger maturation and (C) upon settlement (see top color legend for the acidification treatment). Values of Ω_{ara} below 1.0 (vertical dotted line in gray) represent an under-saturation level, meaning the water is corrosive. During veliger maturation and settlement, the half and less than a quarter of the Ω_{ara} values were below this threshold for the pH challenges that mimic predicted near-future ocean acidification scenarios (green and red cumulative Ω_{ara} trends). The indices of the model performances were AIC: 26.073, BIC: 31.029, R^2 (adj.): 0.232, RMSE: 0.362 and Sigma: 0.467. The model was validated using the `check_model` function in the R package ‘performance’ version 0.7.2 (Lüdecke et al., 2021)[79].

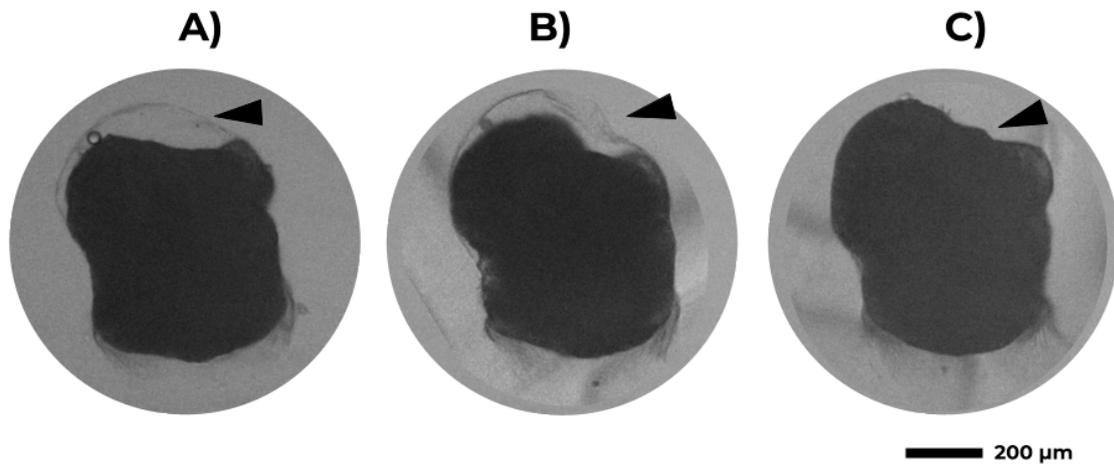


Figure S3. After 24 HPF, the shell first appears as a dome or cap on the posterior end of the larval body (black arrowhead). Individuals were scored as one of the possible morphological groups: **(A)** normal shelled, **(B)** abnormal shelled and **(C)** unshelled.

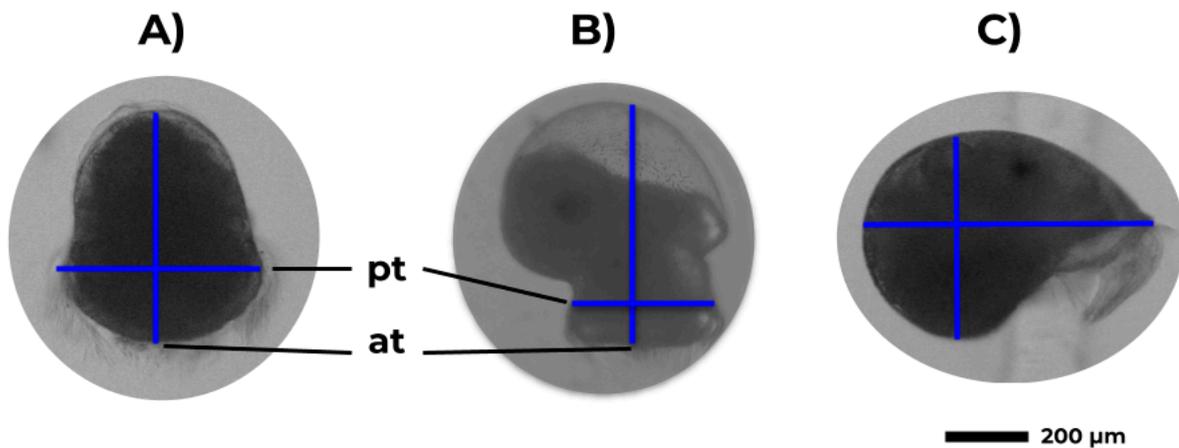


Figure S4. Morphological measurement of the width and length of larvae at 30, 48, 60 and 108 hours post fertilization (HPF). The presence of larvae observed at **30 HPF** include larvae with morphological features related to the trochophore stage **(A)** and early veliger stage **(B)**. Here, the measurement was made horizontally along the prototrochal ciliary band (pt) and vertically from the anterior apical tuft (at) towards the upper zone of the larvae. **(C)** Subsequent measurements at 48, 60 and **108 HPF** considered the maximum length and width of the shell. Images are not to scale.

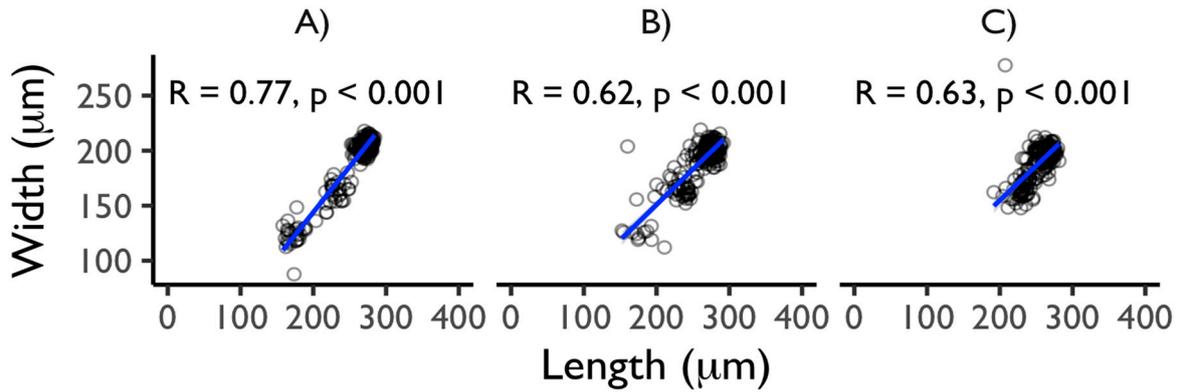


Figure S5. Morphometric assessment consisting of the width (x-axis) and the length (y-axis) during the abalone pelagic life history for **(A)** control (8.0), **(B)** moderate (7.8) and **(C)** low (7.6) pH. The larvae's body widths and lengths are combined as a parameter to estimate growth using the square root product from the length x width. The Spearman correlation test was used for each treatment to calculate a monotonic relationship value (**R**).

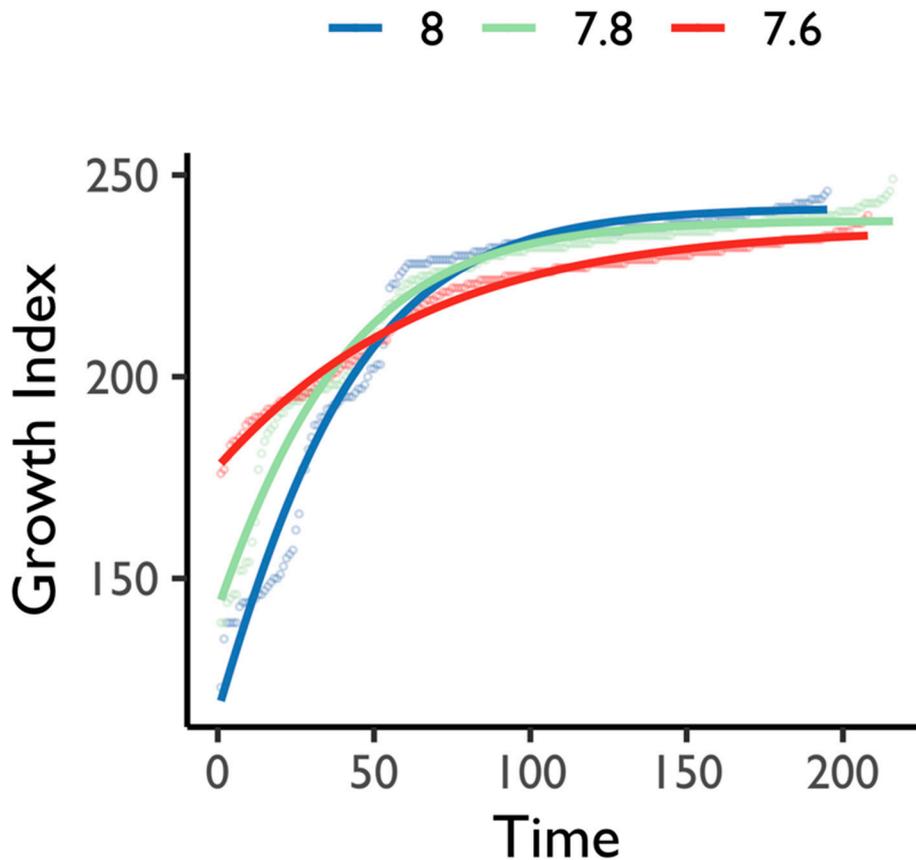


Figure S6. Length-at-stage analysis using Gompertz growth function: $ae^{-be^{-cx}}$.

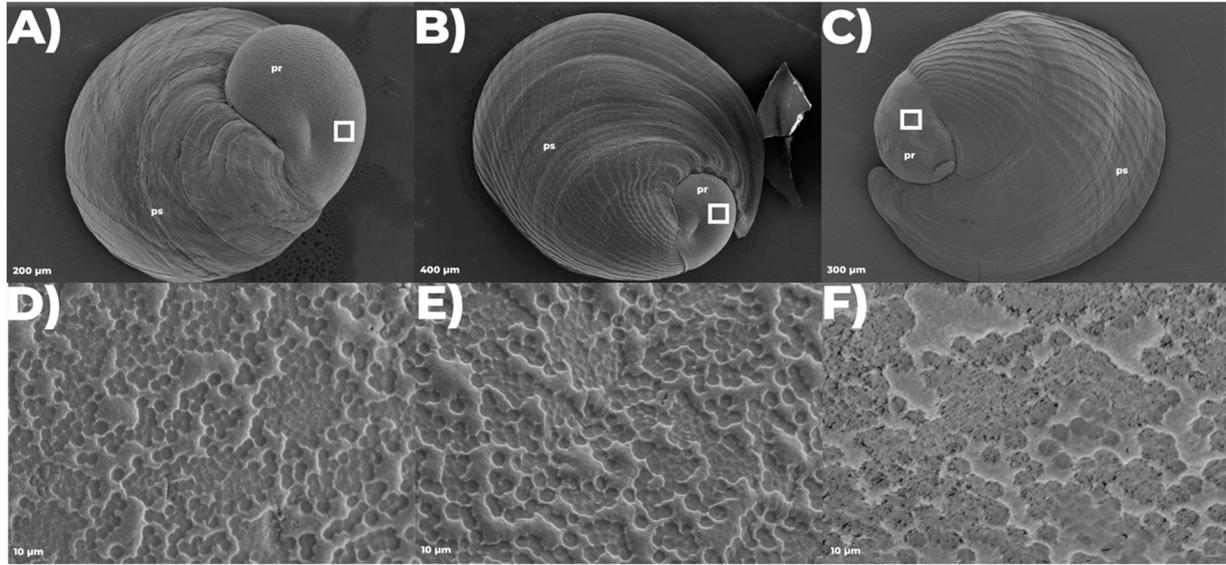


Figure S7. Scanning electron micrographs of *Haliotis rufescens* post-larvae reared under short-term acidification. Dorsal view (200 – 400 μm) of *Haliotis rufescens* 26-day-old post-fertilized larvae showing typical ear-shaped peristomal shell (**ps**) with growth rings and protoconch (**pr**). The control, moderate and low pH are shown, respectively (**A – C**). The laminar section (10 μm) from the protoconch (**pr**, white boxes, **A – C**) is observed in the bottom panel. No evidence of an abnormal granular texture along the protoconch nor microfractures was observed in all the experiments (**D – F**).

Table S1. Specific growth rate from Gompertz growth function: $ae^{-be^{-cx}}$.

	Control	Moderate	Low
Coefficient a	241.844	238.716	236.933
Coefficient b	0.726	0.516	0.288
Coefficient c	0.031	0.030	0.017
Specific growth rate (a*c)	7.532	7.258	4.040
Latency phase (b-1/c)	-8.791	-15.902	-41.763
r-squared	0.977	0.974	0.988
Adjusted r-squared	0.977	0.973	0.988
AIC	1175.593	1205.052	818.517
BIC	1188.685	1218.553	831.867

Table S2. Predicted parameters derived from measured pH_{NBS} records and in situ pH. The indices of model performances were AIC: 26.073, BIC: 31.029, R² (adj.): 0.232, RMSE: 0.362 and Sigma: 0.467. The model was validated using the check_model function in the R package ‘performance’ version 0.7.2 [79].

Assay	Parameter	Stage	Mean ± SD	N records
Low	pH	Embryo	7.5 ± 0.053	28
Moderate	pH	Embryo	7.7 ± 0.022	29
Low	pH	Veliger maturation	7.4 ± 0.104	90
Moderate	pH	Veliger maturation	7.6 ± 0.066	90
Low	pH	Settlement	7.7 ± 0.094	558
Moderate	pH	Settlement	7.8 ± 0.094	550
Control	pH	Settlement	8.1 ± 0.035	308
Moderate	Aragonite	Embryo	1.32 ± 0.108	29
Control	Aragonite	Embryo	1.71 ± 0.022	29
Low	Aragonite	Veliger maturation	1.12 ± 0.088	78
Moderate	Aragonite	Veliger maturation	1.05 ± 9.525	90
Control	Aragonite	Veliger maturation	1.71 ± 0.019	90
Low	Aragonite	Settlement	1.35 ± 46.439	537
Moderate	Aragonite	Settlement	1.63 ± 0.233	543
Control	Aragonite	Settlement	1.72 ± 0.038	368
Low	pCO ₂ _matm	Embryo	930 ± 68.79	28
Moderate	pCO ₂ _matm	Embryo	1020 ± 59.24	29
Control	pCO ₂ _matm	Embryo	581 ± 5.38	29
Low	pCO ₂ _matm	Veliger maturation	1004 ± 115	78
Moderate	pCO ₂ _matm	Veliger maturation	1229 ± 171	90
Control	pCO ₂ _matm	Veliger maturation	582 ± 9	90
Low	pCO ₂ _matm	Settlement	778 ± 92.69	537
Moderate	pCO ₂ _matm	Settlement	787 ± 233.02	543
Control	pCO ₂ _matm	Settlement	580 ± 19.23	370