



Article Greenhouse Gas Emissions from a Main Tributary of the Yangtze River, Eastern China

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Abstract: Rivers and streams are recognized as potential greenhouse gas (GHGs: CO₂, CH₄, and N₂O) sources, contributing to global warming. However, GHG emissions from rivers and streams have received insufficient attention compared to other ecosystems (forests, grasslands, wetlands, etc.). In this study, dissolved GHG concentrations were measured in the Qingyijiang River, the longest tributary in the lower reaches of the Yangtze River, during two campaigns in September 2020 and April 2021. Our results showed that the Qingyijiang River was oversaturated with dissolved GHGs. The dissolved GHG concentration in the surface river water ranged from 8.70 to 67.38 μ M CO₂, 0.03 to 2.06 μ M CH₄, and 12.30 to 32.22 nM N₂O. The average diffusive GHG emission rates were $31.89 \pm 22.23 \text{ mmol CO}_2 \text{ m}^{-2} \text{ d}^{-1}$, 697.22 \pm 939.82 μ mol CH₄ m⁻² d⁻¹, and 18.12 \pm 7.73 μ mol $N_2O \text{ m}^{-2} \text{ d}^{-1}$. The total emissions (CO₂-e) were CO₂ (58%) dominated, while CH₄ (38%) played a moderate role in total emissions. Temporally, average GHG concentrations and fluxes from the studied river in April were higher than those in September. The concentration and flux of CH₄ exhibited high spatial variability, similar to those in most rivers. In contrast, we found that there was no obvious spatial variability in CO₂ and N₂O concentrations but a significant difference among reaches in N₂O fluxes. We found that water temperature and flow velocity were the potential drivers for the regulating spatial variability in GHGs. However, no other observed limnological parameters were found in governing the spatial patterns of GHGs, suggesting a complex combination of factors governing GHG fluxes; thus, these inconspicuous mechanisms underscore the need for further research. Overall, our study suggests that this river acts as a minor source of GHGs relative to other rivers, and CH₄ cannot be ignored when considering aquatic carbon emissions.

Keywords: methane; carbon dioxide; nitrous oxide; Qingyijiang River; flow velocity; global warming potential

1. Introduction

Rivers and streams have been recognized as active transformation sites in transforming organic matter (OM) and the surrounding territorial environments, sediments, and atmosphere when transporting to the oceans [1]. As globally significant emitters of GHGs to the atmosphere, rivers and streams play a disproportionate role in the carbon and nitrogen cycle due to their small coverage area on the planet [2–4]. In recent decades, estimated GHG fluxes from global rivers and streams have varied from 0.23 to 2.35 Pg C yr⁻¹ for



Citation: Miao, Y.; Sun, F.; Hong, W.; Fang, F.; Yu, J.; Luo, H.; Wu, C.; Xu, G.; Sun, Y.; Meng, H. Greenhouse Gas Emissions from a Main Tributary of the Yangtze River, Eastern China. *Sustainability* **2022**, *14*, 13729. https://doi.org/10.3390/su 142113729

Academic Editor: Jiaoyue Wang

Received: 30 August 2022 Accepted: 21 October 2022 Published: 23 October 2022

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Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). CO_2 [1,3,5–8], 1.13 to 22.88 Tg C yr⁻¹ for CH₄ [2,9–12], and 0.19 to 0.68 Tg N yr⁻¹ for N₂O [12–17]. The large, prevailing uncertainty in estimating a GHG budget for global rivers, as reflected in the reported values, is due to the scarcity of available data related to low-order rivers [3,18], and a limited mechanistic understanding of the processes leading to GHG emissions [19].

Although rivers and streams represent key components of freshwater ecosystems, their role in regional or global C and N budgets and underlying biogeochemical processes is still unclear. Theoretically, riverine GHG production processes include the biodegradation of terrestrial and aqueous OM, respiration of phytoplankton, methanogenesis, and nitrification and denitrification (nitrifier and heterotrophic denitrification), while processes such as photosynthesis by phytoplankton, CH₄ oxidation, and excessive denitrification (reduce N_2O to N_2) result in GHG consumption [10,19,20]. Not only are the CO₂, CH₄, and N_2O escaping from river networks dependent on the balance of the aforementioned microbial production and consumption processes inside the rivers, but they are also relative to the gases imported from the catchment [21–23]. Thus, GHG emissions are tightly linked to the factors such as temperature, OM, redox conditions, nitrate, and ammonium that influence these production and consumption processes [10,19,20,24]. However, the relationships between GHGs and those factors are complicated on some occasions. For instance, anoxic streambed sediments are favorable for methanogenesis and denitrification [10,25], leading to CH_4 and N_2O emissions. When more nitrates were transported into anoxic sediments, denitrifying bacteria could out-compete methanogens for substrates and then reduce CH₄ emissions [26]. In a specific river, aquatic plant habitats will promote CO₂ absorption, which can offset CO_2 emission from the open water, ultimately determining the river as a carbon sink or source [27].

In addition, rivers and streams flow through varied landscapes, which may cause differences in abiotic and biotic factors in riverine environments that affect GHG production and consumption processes, and thereby impact riverine GHG fluxes [28–30]. A large number of studies have demonstrated that catchment land-use, including forestland, farmland, and urban areas, can influence the spatial variation in GHG concentrations and fluxes [31–33]. For example, Borges et al. [34] found that rivers draining agriculture (croplands and pastures) had a higher GHG flux than forested systems, which was contributing to excess nutrient and organic matter inputs. The rivers draining urbanized areas enhanced the GHG flux due to nutrient and organic carbon loads [35–37], or receiving external GHGs from sewage [38]. To improve estimates of GHG emissions from running water, it is important to understand the spatiotemporal patterns of GHG fluxes and their drivers in rivers and streams with different catchment environments. Moreover, assessments of CO₂, CH₄, and N₂O concentrations and fluxes simultaneously in a river system across seasons remain scarce.

To address the dearth of information on the dynamics and driving mechanisms of three major GHGs in a freshwater stream with varying catchment land uses, we examined the concentrations, fluxes, and potential drivers of CO_2 , CH_4 , and N_2O along a 309 km stretch of a subtropical river, the Qingyijiang River, subject to land use changes. The following hypotheses were tested: (1) there would be a notable spatial pattern of GHG emissions in varied reaches, due to the surrounding landscapes, and (2) there would be higher GHG emissions in spring than in autumn. In addition, CH_4 and N_2O emissions were converted into CO_2 -equivalent emissions based on their 100 yr global warming potentials (GWPs), and the relative contribution of three gases to total CO_2 -equivalents was quantified. Our results are intended to fill the knowledge gaps of GHG emissions in subtropical rivers and support the assessment of regional GHG budgets in inland freshwater systems.

2. Materials and Methods

2.1. Site Description

This study was performed in the Qingyijiang catchment (29°54′ to 34°26′ N, 117°38′ to 118°51′E; Figure 1), located in the south of Anhui Province, Eastern China. The Qingyijiang

River originates from the northern fringe of Huangshan Mountain and is the longest tributary in the lower reaches of the Yangtze River. The river has a length of 309 km, with a catchment area of 8487 km² and an average slope of 0.00133 m m⁻¹ [39,40]. According to a previous study, the Qingyijiang River can be divided into three parts due to its terrain differences [39]. In brief, the upper reach of the Qingyijiang River extends north from the source to the Chencun Reservoir, the middle reaches extend between the Chencun Reservoir and Wanzhi, and the lower reaches extend from Wanzhi to the outlet of the Qingyijiang River. This region is characterized by a typically subtropical monsoon climate, with a 30 yr (1981–2010) average annual temperature of 16.1 °C and average annual precipitation of 1431 mm, 71.1% of which falls from March through to September (data from http://data.cma.cn (accessed on 1 October 2022)).



Figure 1. Geographical location of Qingvijiang River catchment and 20 sampling sites.

The Qingyijiang River catchment is mainly overlain by Palaeozoic and Mesozoic rocks (granite, sandstone, conglomerate, limestone, shale, etc.) and Quaternary sediments (fluvial deposit, proluvial deposit, red clay, etc.) [40]. Soils are mostly red soil and yellow soil [41]. Land use types were based on an analysis of the land coverage in the catchment drainage area, which was delineated using a DEM (digital elevation model) with 4.11 m × 4.11 m resolution images obtained from Google Earth. The results showed that the Qingyijiang River catchment was occupied by forestland (63.8%), grassland (11.6%), farmland (11.6%), and unused land (8.2%). In contrast, urban land and the surface water area covered 2.7% and 2.1% of the land area in this catchment, respectively (Figure 2a). In situ measurements and water sample collections were conducted in 20 sampling sites along the Qingyijiang River during the autumn of 2020 (12–15 September 2020) and the spring of 2021 (19–20 April 2021). To explore the potential effects of land use on GHG concentrations and fluxes, the land use percentage coverage in 20 drainage segments was calculated in the buffer zone with a radius of 100 m from each sampling site.



Figure 2. (a) Land use types in the Qingyijiang River catchment and (b) their percentage coverage in 20 drainage segments (circle with radius of 100 m of each sampling site).

2.2. Field Sampling and Analyses

The dissolved GHG concentration was measured by a headspace equilibration method [42]. Briefly, surface waters not exceeding 10 cm in depth were sampled by a 60 mL plastic syringe fitted with a stop-cock in triplicate, from each site. The plastic syringe was to equilibrate 40 mL of water with 20 mL of pure N₂. After vigorously shaking for 1–2 min for gas equilibrium [14,43,44], 20 mL of headspace gas was transferred to airtight TedlarR air sample bags previously flushed with pure N₂ and vacuumed. The ambient air samples were also collected in duplicate or triplicate for analysis. These gas samples were delivered to the laboratory and analyzed within 48 h.

The air temperature (Ta) was measured using a portable digital thermometer (JM 624, Jinming, China). The water temperature (Tw), dissolved oxygen (DO) concentrations, and pH were determined by a portable Multi-Parameter Water Quality Sonde (HQ40d, HACH, U.S.A.) equipped with a pH and a DO probe. The water depth (WD) was measured by a self-made sounding lead and a tape measure. Pre-acid-washed polyethylene bottles were used for storing water samples, which were collected at 10 cm below the surface. Before analysis, the bottles were maintained in cool and dark conditions. Water samples were analyzed for total phosphorous (TP) and ammonium (NH₄⁺-N) using a portable Multi-Parameter Water Quality Monitor (5B-2H, Nanbei Instrument, Zhengzhou, China) in the laboratory within 24 h. The GHG concentrations were analyzed by a modified gas chromatograph (GC-2014, Shimadzu, Kyoto, Japan) in the laboratory, which has been described in detail in Miao et al. [45].

2.3. Dissolved GHG Concentrations and Diffusive Fluxes

The dissolved GHG concentrations (c_g : CO₂ and CH₄ in μ M; N₂O in nM) were calculated using Henry's Law [42]:

$$c_g = p_g K_H \tag{1}$$

where the dissolved GHG concentrations are a function of the partial pressure (p_g : CO₂ and CH₄ in µatm; N₂O in natm) of the given gas in surface river water and Henry's constant (K_H : mol L⁻¹ atm⁻¹) adjusted for ambient water temperature for the given gas. Henry's constant K_H is usually determined by a function of the temperature and salinity [46–48]. Thus:

 $Ln K_{H} (CO_{2}) = -58.0931 + 90.5069 (100/T) + 22.2940 Ln (T/100) + S [(0.027766 - 0.025888(T/100) + 0.0050578 (T/100)^{2}]$ (2) Ln K_H (CH₄) = -68.8862 + 101.4956 (100/T) + 28.7314 Ln (T/100) + S [(-0.076146 + 0.043970 (T/100) - 0.006872 (T/100)^{2}] (3)

 $Ln K_H (N_2 O) = -62.7062 + 97.3066 (100/T) + 24.1406 Ln (T/100) + S [(-0.058420 + 0.033193 (T/100) - 0.0051313 (T/100)^2]$ (4)

where *T* is the surface water temperature in Kelvin and *S* is the salinity in ∞ . In this study, we assumed *S* = 0 in the river.

The saturation ratio (s_g) for surface-dissolved GHGs was calculated as follows:

$$s_g = c_g / C_{eq}$$

where C_{eq} is the equilibrium concentration of gas g (CO₂ and CH₄ in μ M; N₂O in nM), which was calculated using the ambient atmospheric GHG partial pressure and the temperature-dependent solubility for GHGs [46–48]. In our study, when $s_g > 1$, it denotes that the water is an atmospheric GHG source; when $s_g < 1$, it denotes that the water is an atmospheric GHG sink.

The GHG diffusive fluxes (f_g : CO₂ in mmol m⁻² d⁻¹; N₂O and CH₄ in µmol m⁻² d⁻¹) between surface water and the atmosphere were determined by the thin boundary layer model as follows:

$$f_g = k_g \left(c_g - C_{eq} \right) \tag{5}$$

where k_g is the gas transfer velocity (cm h⁻¹). In this study, the k_g is determined by the gas transfer velocity normalized to a Schmidt number of 600 (k_{600}) [49]:

$$k_g = k_{600} \left(SC/600 \right)^{-0.5} \tag{6}$$

where *SC* is the dimensionless Schmidt number of a given gas. The Schmidt number *SC* for CO_2 , CH_4 , and N_2O is calculated for the surface water temperature *t* (°C) from the following equations [50]:

$$SC_{CO_2} = 1911.1 - 118.11 t + 3.4527 t^2 - 0.04132 t^3$$
 (7)

$$SC_{CH_4} = 1897.8 - 114.28 t + 3.2902 t^2 - 0.047608 t^3$$
 (8)

$$SC_{N_2O} = 2055.6 - 137.11 t + 4.3173 t^2 - 0.05435 t^3$$
 (9)

 k_{600} is the gas transfer velocity (cm h⁻¹) normalized to a temperature of 20 °C in water with a Schmidt (*SC*) number of 600. Various empirical models have been developed to estimate k_{600} in rivers and streams [49]. As k_g is not measured directly, the average k_{600} derived from the following models was used to calculate the diffusive GHG flux for error reduction.

Using the model by Raymond et al. [49]:

$$k_{600} = 8.42 + 11838 \ vS \tag{10}$$

$$k_{600} = 3965 \ (vS)^{0.76} \tag{11}$$

$$k_{600} = 4842 \ v^{0.85} \ S^{0.77} \tag{12}$$

Using the model by Alin et al. [51]:

$$k_{600} = 13.82 + 35 v \tag{13}$$

where *v* is the flow velocity (m s⁻¹), *S* is the average river slope (m m⁻¹) reported in Li [39].

2.4. Global Warming Potential (GWP)

The global warming potential (GWP) of the total GHG fluxes from the Qingyijiang River was calculated by summation of the GHG using the GWP of each gas. Over the 100-year time horizon, fluxes of CH_4 and N_2O could be converted to their CO_2 equivalents by multiplying radiative forcing factors of 30 and 273, respectively [52].

2.5. Statistical Analyses

All the data are expressed as arithmetical means \pm 1 standard deviation in this study. The normality of the measured variables was examined by the Shapiro–Wilk test, and

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the nonnormal distributed variables were log-transformed. Differences in physical and chemical parameters, GHG concentrations, and fluxes among seasons and varied reaches were determined with one-way analysis of variance (ANOVA) with the LSD test. The coefficient of variation (CV) for GHG concentrations and fluxes was calculated by dividing the standard deviation of the variables measured on each sampling campaign by the mean value. Pearson correlation analysis was performed to explore the relationships between the dissolved GHG concentrations, diffusive GHG fluxes, and physical and chemical parameters. The statistical analyses were performed by the SPSS 22 statistical package (IBM SPSS Inc., Chicago, IL, USA).

3. Results

3.1. Variation in Stream Physical and Chemical Parameters

The characteristics of the stream's physical and chemical parameters are shown in Table 1. The lowest surface water temperature was found in the upper reach and the highest in the lower reach in September, while the lowest surface water temperature was found in the middle reach and the highest in the upper reach. The ANOVA analysis results showed no significant difference among the reaches for water temperature (see Table S1 in the supplementary materials). The middle part of the river presented the lowest depth in September and highest depth in April. The lowest river flow velocity was found in the lower reach with an average value of 0.028 ± 0.013 in September and 0.033 ± 0.006 in April. The pH recorded in the middle reach was lower than those from the upper and lower reaches in September but higher than those from the upper and lower reaches in April. The highest average level of ammonium (NH₄⁺-N) was recorded in the middle reach, with a value of 0.30 ± 0.33 mg L⁻¹ in September and 0.15 ± 0.12 in April.

Table 1. Summary of the physical and chemical variables measured in the surface water of the three river reaches during the observation period.

Month	Reaches	Ta (°C)	Tw (°C)	WD (m)	Flow Velocity (m/s)	Air Pressure (hPa)	DO (mg/L)	рН	NH4 ⁺ -N (mg/L)
	Upper	24.4 ± 1.8	24.3 ± 1.3	0.30 ±	$0.065 \pm$	99.86 ±	$6.32 \pm$	7.88 \pm	0.13 ±
		d	d	0.21 ^a	0.045 ^a	0.39 b	0.78	0.48 ^a	0.17 ^a
September	Middle	24.8 ± 3.0	24.8 ± 0.9	$0.26 \pm$	$0.039 \pm$	100.14 \pm	_	$7.51 \pm$	$0.30 \pm$
		а	а	0.15 ^a	0.030 ^a	0.79 ^a		0.19 ^b	0.33 ^a
	Tanan	23.0 ± 0.6	25.2 ± 2.2	$0.45~\pm$	0.028 \pm	100.73 \pm		7.56 \pm	$0.25 \pm$
	Lower	а	а	0.10 ^a	0.013 ^a	0.06 ^{ab}	-	0.24 ^{ab}	0.09 ^a
	Unnor	25.4 ± 4.3	20.1 ± 2.1	$0.25 \pm$	$0.048~\pm$	99.78 \pm	$8.68~\pm$	7.78 \pm	$0.12 \pm$
April	Opper	а	а	0.21 ^a	0.042 ^a	0.63 ^c	1.17 ^a	0.64 ^a	0.11 ^a
	Middle	28.0 ± 3.6	19.8 ± 2.0	$0.29 \pm$	0.048 \pm	100.40 \pm	$8.32 \pm$	7.96 \pm	$0.15 \pm$
		а	а	0.19 ^a	0.023 ^a	0.60 ^b	0.81 ^a	0.58 ^a	0.12 ^a
	Lower	27.7 ± 2.4	20.3 ± 1.0	$0.27 \pm$	$0.033 \pm$	101.25 \pm	$8.65 \pm$	7.78 \pm	$0.09 \pm$
		а	а	0.09 ^a	0.006 ^a	0.06 ^a	0.99 ^a	0.23 ^a	0.06 ^a
	TT	24.9 ± 3.2	22.2 ± 2.7	$27.9~\pm$	$0.057 \pm$	99.82 \pm	7.64 \pm	7.83 ± 0.5	$0.13 \pm$
All data	Upper	а	а	20.6 ^a	0.043 ^a	0.51 ^c	1.56 ^a	5 ^a	0.14 ^a
	Middle	26.4 ± 3.6	22.3 ± 3.0	$27.4 \pm$	$0.044~\pm$	100.27 \pm	7.73 \pm	7.73 \pm	$0.23 \pm$
		а	а	16.6 ^a	0.026 ^a	0.69 ^b	1.82 ^a	0.48 ^a	0.25 ^a
	-	25.3 ± 3.0	22.7 ± 3.1	$36.2 \pm$	$0.031 \pm$	$100.99 \pm$	$8.65 \pm$	$7.67 \pm$	$0.17 \pm$
	Lower	a	a	13.1 ^a	0.009 ^a	0.29 ^a	0.99 ^a	0.24 ^a	0.11 ^a

Note: The different letters in a column indicate the significant difference among reaches (p < 0.05). Ta, air temperature; Tw, water temperature; WD, water depth; DO, dissolved oxygen; NH₄⁺-N, ammonium.

3.2. Dissolved GHG Concentrations in Surface River Water

The dissolved GHG concentrations in the surface water of the Qingyijiang River varied from 8.70 to 67.38 μ M (mean: 28.17 \pm 10.31 μ M) for CO₂, 0.03 to 2.06 μ M (mean: 0.27 \pm 0.36 μ M) for CH₄, and 12.30 to 32.22 nM (mean: 16.16 \pm 3.16 nM) for N₂O during

the observation period. The saturation ratios ranged from 0.68 to 3.33 (mean: 1.86 \pm 0.57) for CO₂, from 10.49 to 752.31 (mean: 93.02 \pm 130.51) for CH₄, and from 1.42 to 4.01 (mean: 1.89 \pm 0.45) for N₂O.

Temporally, dissolved GHG concentrations observed in September were lower than those in April (Table 2). For all sampling sites, the average dissolved CO_2 , CH_4 , and N_2O concentrations in April were 16.8%, 137.5%, and 5.5%, respectively, higher than those in September.

The spatial variability in dissolved GHG concentrations was assessed by the coefficient of variation (CV) in this study. The CV values were 7.28% for dissolved CO₂ concentrations and 3.59% for dissolved N₂O concentrations in autumn, indicating that both gases were homogenously distributed, while both of them were more spatially heterogeneous than in spring (CV: 47.11% for CO₂ and 26.80% for N₂O). Compared with CO₂ and N₂O, dissolved CH₄ concentrations exhibited greater spatial variability within the river during the sampling period (CV: 85.33% in autumn and 125.14% in spring). However, the ANOVA test showed that no significant difference existed among reaches for dissolved CO₂ concentrations in either season, and for dissolved CH₄ and N₂O concentrations in spring (p > 0.05) (Table S1). The significant difference among reaches for CH₄ and N₂O only occurred in autumn. Higher dissolved N₂O concentrations were observed in the upper reaches (mean: 16.08 ± 0.48 nM) than those in the middle reaches (mean: 15.43 ± 0.49 nM), while the opposite pattern was shown for dissolved CH₄ concentrations.

3.3. Diffusive GHG Fluxes

Based on the water–air gas exchange model, the calculated diffusive GHG fluxes in the river ranged from -8.53 to 121.57 mmol CO₂ m⁻² d⁻¹, 66.94 to 5395.84 µmol CH₄ m⁻² d⁻¹, and 8.77 to 56.73 µmol N₂O m⁻² d⁻¹, as a net GHG source for the atmosphere (Table 2). The spatiotemporal patterns of diffusive GHG fluxes mimicked the spatial and seasonal variability observed in the GHG concentrations. All three gas fluxes observed in spring were higher than those in autumn, with the average fluxes of 31.89 ± 22.23 mmol CO₂ m⁻² d⁻¹, 697.22 ± 939.82 µmol CH₄ m⁻² d⁻¹, and 18.12 ± 7.73 µmol N₂O m⁻² d⁻¹. The results of the one-way ANOVA test revealed that no significant difference in GHG fluxes was observed between seasons.

The CV values of the diffusive GHG fluxes were 69.70% for CO₂, 134.80% for CH₄, and 42.64% for N₂O, and showed considerable spatial variations on the site scale throughout the sampling period. However, no significant difference in diffusive CO₂ and CH₄ fluxes was observed among the reaches in either season (p > 0.05; Table 2), while diffusive N₂O fluxes significantly differed among the reaches in the following order: upper (19.40 ± 2.98 µmol m⁻² d⁻¹) > middle (16.30 ± 2.63 µmol m⁻² d⁻¹) > lower (14.60 ± 0.32 µmol m⁻² d⁻¹) (p < 0.05; Table 2). Overall, lower CO₂ emissions and higher CH₄ and N₂O emissions were observed in the middle reach as compared to the other reaches.

Month	Reaches	с _{СО2} (µМ)	s _{CO2}	f_{CO_2} (mmol m ⁻² d ⁻¹) #	с _{СН4} (µМ)	s _{CH4}	$f_{\rm CH_4} \ (\mu { m mol} \ { m m}^{-2} \ { m d}^{-1})^{\#}$	$c_{\mathrm{N_2O}}$ (nM)	$s_{ m N_2O}$	f _{N2O} (μmol m ⁻² d ⁻¹) #
	Upper	$25.50\pm2.05~^{\rm a}$	$1.68\pm0.09~^{\rm a}$	$\begin{array}{c} 29.71 \pm 6.76 \\ (15.49 \pm \\ 9.93{\sim}43.58 \pm \\ 7.89) \end{array}$	$0.09\pm0.06~^{\rm b}$	29.33 ± 16.70 ^b	$283.84 \pm 153.65 \\ {}^{b} (134.43 \pm \\ 78.22 \sim 422.07 \pm \\ 234.32)$	$16.08\pm0.48~^{\rm a}$	$1.74\pm0.11~^{\rm ab}$	$\begin{array}{c} 19.40 \pm 2.98 \ ^{a} \\ (9.95 \pm \\ 5.35 {\sim} 28.53 \pm \\ 3.32) \end{array}$
September	Middle	$25.82\pm1.60~^{\rm a}$	$1.66\pm0.09~^{a}$	$\begin{array}{c} 25.12 \pm 2.71 \text{ a} \\ (4.76 \pm \\ 6.16{\sim}41.89 \pm \\ 3.57) \end{array}$	0.23 ± 0.19 a	69.26 ± 55.29 ^a	644.76 ± 536.99 ^a (161.00 \pm 281.30~1056.12 \pm 872.91)	$15.43\pm0.49~^{\text{b}}$	$1.76\pm0.07~^{\rm a}$	$\begin{array}{c} 16.30 \pm 2.63 \\ (3.36 \pm \\ 4.99{\sim}27.06 \pm \\ 2.35) \end{array}$
	Lower	$27.91\pm1.26~^{\rm a}$	$1.79\pm0.24~^{\rm a}$	$\begin{array}{c} 28.31 \pm 6.57 \text{ a} \\ (1.13 \pm \\ 0.32{\sim}49.11 \pm \\ 10.51) \end{array}$	$0.18\pm0.0~7~^{ab}$	$54.60\pm20.18~^{ab}$	$\begin{array}{c} 494.17 \pm 211.03 \\ {}^{ab} \ (18.81 \pm \\ 5.76{\sim}866.28 \pm \\ 381.03) \end{array}$	$15.48\pm0.51~^{ab}$	$1.70\pm0.02~^{\rm b}$	$\begin{array}{c} 14.60 \pm 0.32 \ {}^{\mathrm{b}} \\ (0.59 \pm \\ 0.12 {\sim} 25.44 \pm \\ 1.26) \end{array}$
	Overall	25.99 ± 1.89	1.69 ± 0.12	$\begin{array}{c} 27.66 \pm 5.60 \\ (9.04 \pm \\ 9.63{\sim}43.73 \pm \\ 6.97) \end{array}$	0.16 ± 0.14	49.09 ± 40.59	$\begin{array}{c} 459.76 \pm 387.46 \\ (127.72 \pm \\ 184.64 {\sim} 742.32 \pm \\ 641.55) \end{array}$	15.73 ± 0.56	1.74 ± 0.08	$\begin{array}{c} 17.44 \pm 3.15 \\ (5.91 \pm \\ 6.01 27.48 \pm \\ 2.85) \end{array}$
	Upper	$29.78\pm18.44~^{\rm a}$	1.86 ± 0.90 a	$\begin{array}{r} 36.00 \pm 41.26 \ ^{\rm a} \\ (16.34 \pm \\ 25.55{\sim}51.65 \pm \\ 62.33) \end{array}$	0.37 ± 0.64 ^a	132.59 ± 235.67	$973.67 \pm 1694.78 \\ ^{a} (314.38 \pm \\ 384.78 \sim 1499.93 \\ \pm 2702.71)$	$16.27\pm6.29~^{\rm a}$	1.89 ± 0.81 a	$\begin{array}{c} 18.27 \pm 14.84 \text{ a} \\ (6.54 \pm \\ 4.06{\sim}27.94 \pm \\ 23.89) \end{array}$
April	Middle	30.64 ± 11.73 $^{\rm a}$	2.13 ± 0.73 $^{\rm a}$	$35.99 \pm 23.26^{\text{ a}}$ (8.56 \pm 8.66~61.09 \pm 40.17)	0.40 ± 0.33 ^a	145.15 ± 115.09	967.45 ± 897.37 ^a (267.40 \pm 467.51~1578.76 \pm 1351.09)	17.59 ± 2.38 ^a	$2.24\pm0.37~^{\rm a}$	$\begin{array}{c} 21.04 \pm 6.02 \\ (4.29 \pm \\ 3.86 \\ 35.71 \pm \\ 9.91) \end{array}$
	Lower	$31.33\pm9.89~^{\rm a}$	$2.32\pm0.51~^{\rm a}$	$\begin{array}{c} 36.84 \pm 16.62 \\ ^{a} \\ (2.13 \pm \\ 1.60 \\ -64.78 \pm \\ 29.60) \\ \end{array}$	0.33 ± 0.24 a	128.12 ± 94.23 $^{\rm a}$	730.36 ± 537.08 a (41.15 ± 34.89~1278.70 ± 936.61)	14.95 ± 1.19 a	1.89 ± 0.13 a	$\begin{array}{c} 14.46 \pm 1.87 \\ (0.78 \pm \\ 0.46 \\ \sim 25.38 \pm \\ 3.32) \end{array}$
	Overall	30.36 ± 14.30	2.04 ± 0.77	$\begin{array}{r} 36.12 \pm 30.75 \\ (11.10 \pm \\ 18.20 \times 57.40 \pm \\ 48.50) \end{array}$	0.38 ± 0.47	136.94 ± 171.03	934.69 ± 1242.65 (254.60 ± 389.81~1498.28 \pm 1962.36)	16.60 ± 4.45	2.03 ± 0.60	$ \begin{array}{r} 18.81 \pm 10.57 \\ (4.78 \pm \\ 4.06 \sim 30.67 \pm \\ 17.21) \\ \end{array} $

 Table 2. Descriptive statistics of surface water GHG concentration and diffusive GHG fluxes across the water-atmosphere interface during sampling period.

Month	Reaches	с _{СО2} (µМ)	s _{CO2}	f_{CO_2} (mmol m ⁻² d ⁻¹) [#]	с _{СН4} (µМ)	$s_{ m CH_4}$	$f_{\rm CH_4} \ (\mu { m mol} \ { m m}^{-2} \ { m d}^{-1})^{\#}$	c _{N2O} (nM)	s_{N_2O}	$f_{ m N_2O}$ (µmol m $^{-2}$ d $^{-1}$) #
	Upper	27.64 ± 12.92 ^a	1.77 ± 0.63 $^{\rm a}$	$\begin{array}{c} 32.86 \pm 28.87 \text{ a} \\ (15.92 \pm \\ 18.81 47.61 \pm \\ 43.30) \end{array}$	$0.23\pm0.47~^{a}$	$80.96 \pm 170.56~^{\rm a}$	$\begin{array}{c} 628.76 \pm 1220.14 \\ {}^{a} \ (224.40 \pm \\ 284.82 {\sim} 961.00 \pm \\ 1941.86) \end{array}$	$16.17\pm4.33~^{\rm a}$	1.81 ± 0.56 $^{\rm a}$	$\begin{array}{c} 18.84 \pm 10.40 \ ^{\rm a} \\ (8.24 \pm \\ 4.93{\sim}28.24 \pm \\ 16.55) \end{array}$
All data	Middle	$28.23\pm8.47~^{a}$	$1.90\pm0.56~^{\rm a}$	$\begin{array}{c} 30.56 \pm 16.95 \text{ a} \\ (6.66 \pm \\ 7.52{\sim}51.49 \pm \\ 29.28) \end{array}$	$0.32\pm0.28~^a$	107.20 ± 95.62 ^a	$\begin{array}{c} 806.10 \pm 733.57 \\ {}^{a} \ (214.20 \pm \\ 376.76 \\ -1317.44 \\ \pm \ 1131.50) \end{array}$	16.51 ± 2.00	$2.00\pm0.36~^a$	$\begin{array}{c} 18.67 \pm 5.11 \\ (3.83 \pm \\ 4.33 \text{~~} 31.38 \pm \\ 8.27) \end{array}$
	Lower	$29.62\pm6.58~^{\rm a}$	$2.06\pm0.46~^{a}$	$\begin{array}{c} 32.58 \pm 12.23 \text{ a} \\ (1.63 \pm \\ 1.17 \text{~~} 56.95 \pm \\ 21.62) \end{array}$	$0.26\pm0.18~^{a}$	91.36 ± 73.05 ^a	$\begin{array}{c} 612.27 \pm 387.21 \\ {}^{a} (29.98 \pm \\ 649.74 {\sim} 1072.49 \\ \pm 678.23) \end{array}$	15.21 ± 0.87 ^a	$1.80\pm0.14~^{a}$	$\begin{array}{c} 14.53 \pm 1.20 \ ^{\rm a} \\ (0.69 \pm \\ 0.32 {\sim} 25.41 \pm \\ 2.25) \end{array}$
	Overall	28.17 ± 10.31	1.86 ± 0.57	$\begin{array}{c} 31.89 \pm 22.23 \\ (10.07 \pm \\ 14.41 {\sim} 50.57 \pm \\ 34.89) \end{array}$	0.27 ± 0.36	93.02 ± 130.51	$\begin{array}{c} 697.22 \pm 939.82 \\ (191.16 \pm \\ 307.84{\sim}1120.30 \\ \pm 1491.00) \end{array}$	16.16 ± 3.16	1.89 ± 0.45	$\begin{array}{c} 18.12 \pm 7.73 \\ (5.34 \pm \\ 5.10 {\sim} 29.07 \pm \\ 12.28) \end{array}$

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Note: The different letters in a column indicate significant differences among reaches (p < 0.05). c_{CO_2} , c_{CH_4} , and c_{N_2O} represent dissolved CO₂, CH₄, and N₂O concentrations, respectively. s_{CO_2} , s_{CH_4} , and s_{N_2O} represent saturations of dissolved CO₂, CH₄, and N₂O in river water, respectively. f_{CO_2} , f_{CH_4} , and f_{N_2O} represent diffusive CO₂, CH₄, and N₂O fluxes, respectively. # The ranges of fluxes estimated by the different models are presented in the brackets.

The global warming potentials of CH₄ and N₂O were over the 100-year horizon, which were 30 and 273 times larger than that of CO₂, respectively [52]. For the Qingyijiang River, CH₄ and N₂O emissions would translate into 20.92 and 2.47 mmol CO₂ eq m⁻² d⁻¹, which accounted for 37.84% and 4.47% of the three GHG emissions, respectively.

3.4. Relationships between GHG Emissions and Environmental Parameters

The correlation analysis between dissolved GHG concentrations and diffusive GHG fluxes and the measured environmental parameters for each season are shown in Figure 3. In spring, there was a significant negative relationship between CO_2 concentration and flux and DO, and a significant positive relationship between CO_2 concentration and flux and TP, while we observed a significant positive relationship between CH_4 and N_2O fluxes and water temperature. In autumn, the spatial patterns of CO_2 and N_2O fluxes were significantly regulated by flow velocity. However, no significant relationship between CH_4 concentration (or flux) and the environmental variables was found in this study.





Figure 3. Correlation matrix for GHG concentrations and fluxes with physiochemical parameters in the surface water of the Qingyijiang River during the measurement periods: (a) September 2020; (b) April 2021. WT, DO, AP, WD, NH₄, TP, and FV represent the water temperature, dissolved oxygen, air pressure, water depth, ammonium, total phosphorous, and flow velocity, respectively.

4. Discussion

4.1. Comparison of GHG Emissions with Other Rivers

A considerable number of observations in the literature have demonstrated that most of streams and rivers were supersaturated with GHGs relative to the atmosphere [3,10,19,29]. The dissolved GHG concentrations and diffusive GHG fluxes obtained from different temporal and spatial sampling strategies undertaken in the Qingyijiang River were lower than those found in most rivers and streams around the world (Table S3). However, the dissolved CO₂ and N₂O concentrations and diffusive CO₂ and N₂O flux were comparable to those reported from tropical and subtropical rivers [53,54]. Dissolved CH₄ concentrations and diffusive CH_4 fluxes were comparable to those reported from some boreal streams [24]. The lower GHG emissions in this study could be explained as follows: first, they might be associated with the low frequency of sampling measurements, which was unlikely to capture events with high GHG fluxes [55,56]. Second, they might be related to the low GHG production capacity in the studied rivers, which received less pollutant loadings compared to rivers flowing through urban regions or agriculture-dominated catchments [35,36,57]. For example, Wang et al. [58] found that the CO_2 emission from rivers in urban areas was 2–4 times higher than that in remote rural rivers. Yu et al. [35,36] and Hu et al. [54] also found higher CH_4 and N_2O fluxes from rivers draining highly urbanized landscapes. In

addition, rivers draining agricultural landscapes emitted more GHGs than from natural rivers [34,57,59].

Calculating CO₂-equivalent Global Warming Potentials (GWP) on a specified year timescale enabled us to assess the relative contribution of each GHG [60]. In this study, overall CO₂-equivalent emissions for the Qingyijiang River were dominated by CO₂, which was similar to other studies [30,32,61]. It is worth noting that CH₄ and N₂O had a certain contribution to CO₂-equivalent emissions (37.84% for CH₄ and 4.47% for N₂O), which highlights the crucial role of CH₄ and N₂O in river GHG emissions.

4.2. Factors Influencing the GHG Spatial Variation

Previous studies have shown that emissions of GHGs across the river-wateratmosphere interface originate in complex microbial processes (production and consumption) in carbon and nitrogen cycles, which depend on abundant environmental parameters such as temperature, oxygen, nutrient status, and flow velocity [34,57,62]. Temperature (sediment and/or water) has been proven to be a crucial factor for determining GHG concentrations and fluxes, as a result of the influence on the microbial processes of production and consumption. Similar to studies conducted in other rivers and streams [54,57,63], a positive correlation had been found between water temperature and N₂O and CH₄ flux through Pearson correlation analysis (Figure 3), but this relationship could not be established for CO_2 . A considerable number of studies have shown that microbial activity can be stimulated by suitable high water and sediment temperature, thereby affecting CH_4 and N_2O production and emission [10,63–65]. In addition, the solubility of CH₄ and N_2O decreases with increasing temperature; therefore, CH₄ and N₂O are immediately released into the atmosphere at high temperature [54,66]. Furthermore, temperature not only affects respiration, but also affects photosynthesis in aquatic systems [67,68]; the combined effects on CO₂ concentration may have resulted in an inconspicuous temperature dependence of CO_2 emission in this study.

Dissolved oxygen, as another important feature of habitat redox status, plays a crucial role in GHG production and consumption, thereby determining GHG concentrations and fluxes [10,19,62]. For example, CH_4 is predominantly produced via methanogenesis under low- O_2 conditions in aquatic systems [10]. However, recent studies have found that CH_4 could also be produced under an oxic environment that was originally suitable for CH_4 oxidation [69]. In this study, no significant relationship between CH_4 and DO suggests that the spatial heterogeneity of CH₄ emission could not explained by DO concentration. N₂O can be produced via N-cycling pathways such as denitrification and dissimilatory nitrate reduction to ammonia (DNRA) under anaerobic conditions, and nitrification under aerobic conditions [19]. The DO concentrations observed in the Qingyijiang River were high enough for nitrification, and the higher N₂O concentration and NH₄⁺-N concentrations were observed in the middle reach of the river, which suggests that the nitrification pathway may be an important source of N₂O. The negative relationships between DO concentration and CO_2 concentration and flux agree with previous studies for rivers [62,70] (Figure 3), and could be attributed to pelagic respiratory production of CO₂, thus decreasing DO levels.

Land use types in the catchment are important in mediating carbon and nitrogen biogeochemical processes on land, thus influencing inputs of allochthonous materials into adjacent waters [71–73], with the consequence of changing the magnitude of GHG fluxes [33,34]. Agricultural and urban land uses have been found to be crucial for riverine GHG production [30,34,36,37], because nutrient loadings can lead to enhanced in-stream metabolism [32]. However, the impact of land use on river GHG concentrations and fluxes can be complex, and could not be clearly observed in this study (Table S2). Although no significant correlations between GHG concentrations and fluxes and the proportions of land use were found in the present study, higher GHG concentrations were clearly observed in the group with >50% agricultural land use in both seasons (Figure 2b). This suggests the importance of agriculture in generating GHGs in the river network.

The water flow velocity is known to be an important driver in determining the spatial variability in GHG fluxes [57,62,64,74]. In the present study, the positive relationships between flow velocity and CO_2 and N_2O fluxes are consistent with a previous study [61]. The strong flow, which affected the near-surface turbulence, led to high gas transfer velocities, thus enhancing the gas evasion [49].

Although the potential drivers influencing the spatial variability in GHGs have been assessed, our findings suggest that it is difficult to predict GHG emissions from the Qingyijiang River based on existing concentration/flux-environmental parameter relationships alone.

4.3. Study Limitations and Future Research

Similar to many other studies, there exist several uncertainties associated with GHG flux estimates in the Qingyijiang River. First, the riverine GHG flux calculation was derived from the gas transfer velocity coefficient (k) and \triangle GHG concentration ($c_g - C_{eq}$), which was associated with high uncertainty. *k* was computed by empirical models, rather than measured directly in the field, which makes the actual gas transfer velocity coefficients unclear. As Raymond et al. [46] compiled, there are many models with which to estimate k; however, there is no consistent principle to determine which one should be chosen. To eliminate the divergence, the average gas transfer velocity of each gas derived from the selected empirical models was applied to calculate GHG flux in this study. Despite this, future studies should determine the gas transfer velocities directly in the field in order to constrain the uncertainty in flux estimates. Secondly, our study focused on gas diffusive fluxes without considering ebullitive fluxes, which were the main emission pathway for CH_4 reported in some streams and rivers [9,55,56]. Although the studied river originates from a mountain area, the lower reaches of the river are located on a plain, where potential ebullitive fluxes may occur. Previous studies have illustrated that ebullition may contribute substantially (> 50%) to total CH_4 flux [9,55,56]; thus, this study's estimation of CH_4 flux was conservative without considering ebullitive fluxes. Whether CH₄ is emitted through bubbles in the lower reaches of our studied river requires further study.

Finally, we also recognized that our flux estimation was biased toward sampling time and frequency. On the one hand, flux measurements conducted in the daytime may lead to bias in flux estimates, as the diel variability of GHG fluxes has been widely reported in rivers [8,75,76]. For instance, Gómez-Gener et al. [8] demonstrated that nocturnal CO₂ emissions from global streams are on average 27% greater than those estimated from diurnal concentrations alone. Chen et al. [56] found that higher CH₄ fluxes occurred during the daytime compared to the nighttime. Some researchers found that N₂O emissions from subtropical rivers during the nighttime were higher [77] or lower [78] than N₂O emissions during the daytime. These results imply that extrapolations based on daytime observations most likely result in overestimated or underestimated fluxes. On the other hand, singletime flux measurements may produce a large uncertainty in flux estimates, which has been reported for other aquatic ecosystems [79,80]. Therefore, consideration of diel variability and sampling frequency in future studies is critical to properly assess total riverine GHG fluxes.

5. Conclusions

In conclusion, here, we have investigated the spatial variability in CO_2 , CH_4 , and N_2O concentration and flux from a tributary of the Yangtze River located in Eastern China. Our results showed that the river was supersaturated in CO_2 , CH_4 , and N_2O at most of the times and at most of sites, relative to the atmosphere, but the estimated GHG fluxes were one and/or two orders of magnitude lower than those found in most rivers and streams around the world. Although the river acts as a minor GHG emitter, the crucial role of CH_4 and N_2O cannot be ignored when considering CO_2 -equivalent emissions. The GHG emissions from the river varied between seasons, with the higher values appearing in spring. However, there was no significant spatial variability in CO_2 concentrations

and fluxes but a significant difference among reaches in N₂O concentrations and fluxes in September. This could be explained by the varied flow velocities in the sampling sites, which determine the gas transfer velocity coefficient. Furthermore, the spatial variation in CH₄ concentration and flux was observed without obvious patterns. Unlike most studies with clear drivers for influencing the spatial variability in GHGs, we only found water temperature, DO, and flow velocity partly regulating GHG emissions, which suggests that it is difficult to predict GHG emissions from this river based on measured parameters alone. Overall, our results highlight an urgent need for more detailed studies on flux measurements to improve the reliability of riverine GHG flux estimates, and process-based measurements to help elucidate the drivers of variability in GHGs from these subtropical rivers.

Supplementary Materials: The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/su142113729/s1, Table S1: Summary of ANOVA examining the effect of reaches on physical and chemical variables, GHG concentrations and fluxes during sampling period. Table S2: Relations between GHG concentrations and fluxes and environmental variables in each campaign. Table S3: Dissolved GHG concentrations in surface waters and diffusive GHG emissions from Qingyijiang River compared with other studies.

Author Contributions: Conceptualization, Y.M. and H.M.; methodology, Y.M.; software, W.H.; investigation, Y.S., H.L. and Y.M.; data curation, Y.M. and H.M.; writing—original draft preparation, Y.M., C.W., F.S. and H.M.; writing—review and editing, Y.M., F.F., J.Y., C.W. and G.X.; visualization, F.S. and W.H.; project administration, Y.M.; funding acquisition, Y.M., H.M. and G.X. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Anhui Provincial Natural Science Foundation (grant no. 2108085MD126), the National Natural Science Foundation of China (grant nos. 41601083 and 41601103), the Opening Foundation of Anhui Province key Laboratory of Environmental Hormone and Reproduction (Fuyang Normal University) (grant no. FSKFKT012), Science and Technology Program of Hebei Academy of Sciences (grant no. 18109), and the University Synergy Innovation Program of Anhui Province (grant no. GXXT-2020–075).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data that support the findings of this study are openly available on request.

Acknowledgments: We would like to thank Qian Cui, Qiangqiang Yang, and Xiancheng Yang of the School of Geography and Tourism, Anhui Normal University for their field assistance.

Conflicts of Interest: The authors declare no conflict of interest.

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