

## Supplementary Material

# Identification of Effective Anticancer G-Quadruplex-Targeting Chemotypes through the Exploration of a High Diversity Library of Natural Compounds

Chiara Platella <sup>1,†</sup>, Francesca Ghirga <sup>2,†</sup>, Pasquale Zizza <sup>3</sup>, Luca Pompili <sup>3</sup>, Simona Marzano <sup>4</sup>, Bruno Pagano <sup>4</sup>, Deborah Quaglio <sup>2</sup>, Valeria Vergine <sup>2</sup>, Silvia Cammarone <sup>2</sup>, Bruno Botta <sup>2</sup>, Annamaria Biroccio <sup>3</sup>, Mattia Mori <sup>5,\*</sup> and Daniela Montesarchio <sup>1,\*</sup>

**Citation:** Platella, C.; Ghirga, F.; Zizza, P.; Pompili, L.; Marzano, S.; Pagano, B.; Vergine, V.; Cammarone, S.; Botta, B.; Biroccio, A.; et al. Identification of Effective Anticancer G-Quadruplex-Targeting Chemotypes through the Exploration of a High Diversity Library of Natural Compounds. *Pharmaceutics* **2021**, *13*, 1611. <https://doi.org/10.3390/pharmaceutics13101611>

Academic Editors: Jung Min Shin, Eun Sook Lee

Received: 13 August 2021

Accepted: 26 September 2021

Published: 3 October 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).

<sup>1</sup> Department of Chemical Sciences, University of Naples Federico II (Complesso Universitario di Monte S. Angelo), Via Cintia, 21, 80126 Napoli, Italy; chiara.platella@unina.it

<sup>2</sup> Department of Chemistry and Technology of Drugs, “Department of Excellence 2018–2022”, Sapienza University of Rome, P.le Aldo Moro 5, 00185 Rome, Italy; francesca.ghirga@uniroma1.it (F.G.); deborah.quaglio@uniroma1.it (D.Q.); valeria.vergine@uniroma1.it (V.V.); silvia.cammarone@uniroma1.it (S.C.); bruno.botta@uniroma1.it (B.B.)

<sup>3</sup> Oncogenomic and Epigenetic Unit, IRCCS—Regina Elena National Cancer Institute, Via Elio Chianesi 53, 00144, Rome, Italy; pasquale.zizza@ifo.gov.it (P.Z.); luca.pompili@ifo.gov.it (L.P.); annamaria.biroccio@ifo.gov.it (A.B.)

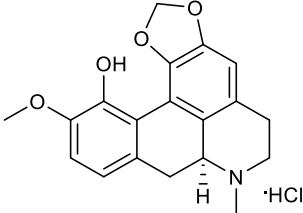
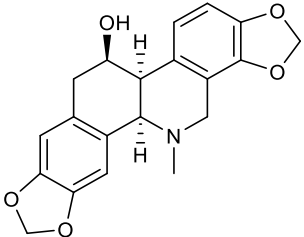
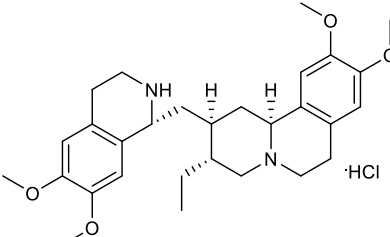
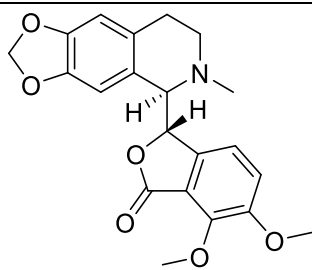
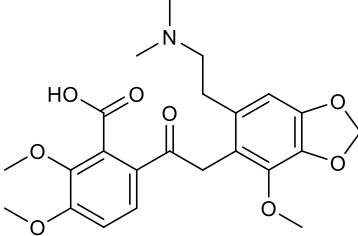
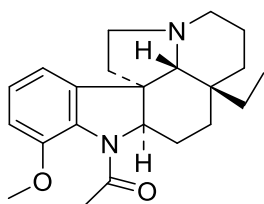
<sup>4</sup> Department of Pharmacy, “Department of Excellence 2018–2022”, University of Naples Federico II, via D.Montesano, 49, 80131 Napoli, Italy; simona.marzano@unina.it (S.M.); bruno.pagano@unina.it (B.P.)

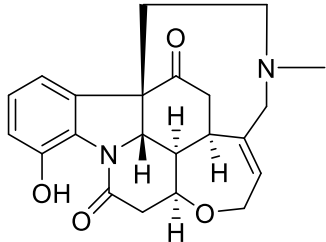
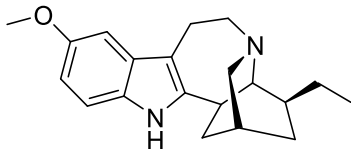
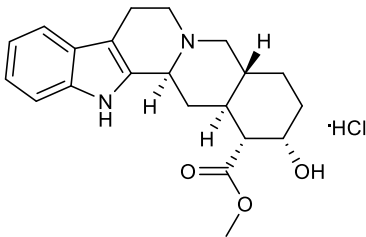
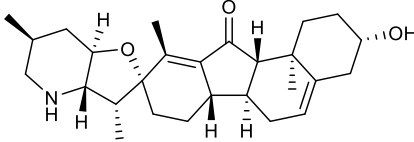
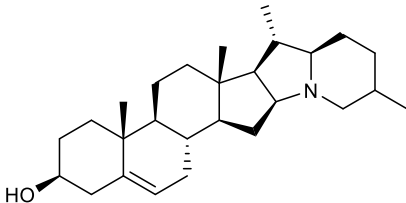
<sup>5</sup> Department of Biotechnology, Chemistry and Pharmacy, “Department of Excellence 2018–2022”, University of Siena, Via Aldo Moro 2, 53100 Siena, Italy

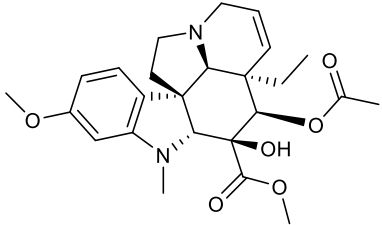
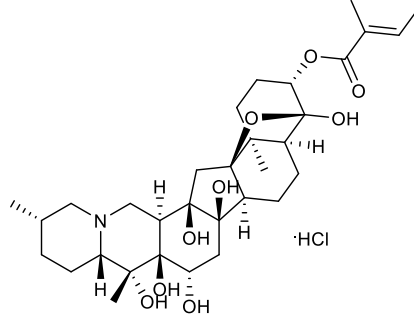
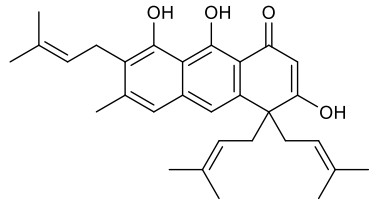
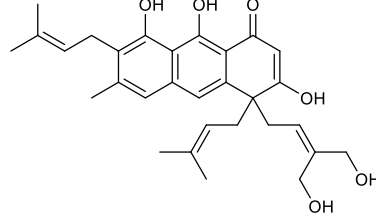
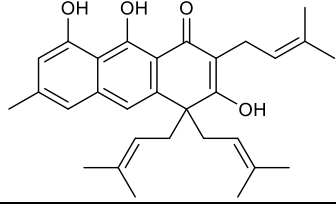
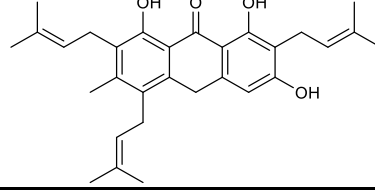
\* Correspondence: mattia.mori@unisi.it (M.M.); daniela.montesarchio@unina.it (D.M.)

† These authors equally contributed to this work.

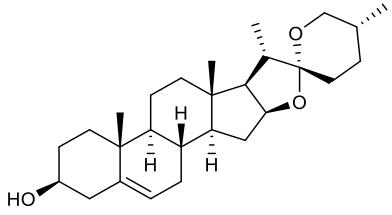
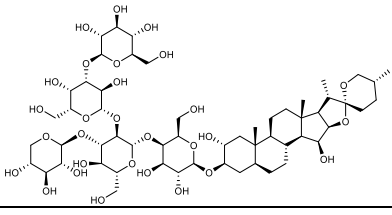
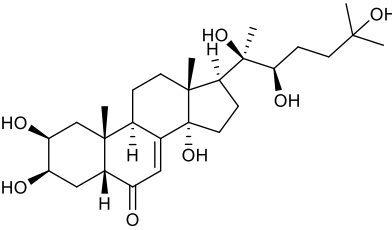
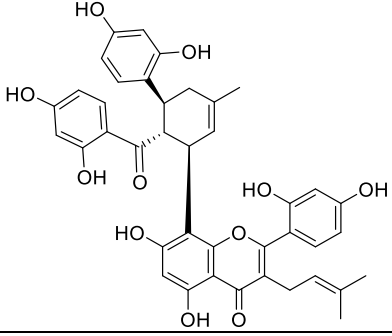
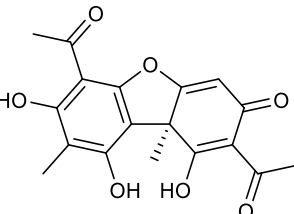
Table S1. Chemical structures, features and natural sources of the 28 natural compounds here investigated.

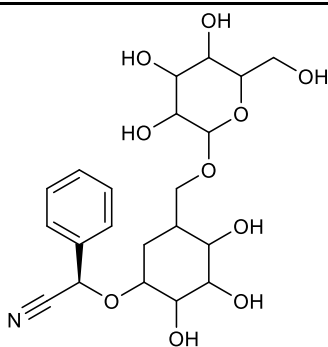
Cpd	Common Name (Library Code)	Chemical Structure	M. W.	Molecular Formula	Source	Reference
Alkaloids						
1	<b>Bulbocapnine·HCl</b> (BBN196)		325.36 361.82 (+HCl)	C <sub>19</sub> H <sub>19</sub> NO <sub>4</sub> ·HCl	Species: <i>Corydalis cava</i> (Papaveraceae family)	[1]
2	<b>Chelidonine</b> (BBN192)		353.37	C <sub>20</sub> H <sub>19</sub> NO <sub>5</sub>	Species: <i>Chelidonium majus</i> L. (Papaveraceae family)	[2]
3	<b>Emetine·HCl</b> (BBN39)		480.65 517.11 (+HCl)	C <sub>29</sub> H <sub>40</sub> N <sub>2</sub> O <sub>4</sub> ·HCl	Species: <i>Psychotria ipecacuanha</i> Stokes (Rubiaceae family)	[3]
4	<b>Hydrastine</b> (BBN40)		383.40	C <sub>21</sub> H <sub>21</sub> NO <sub>6</sub>	Species: <i>Hydrastis canadensis</i> L. (Ranunculaceae family)	[4]
5	<b>Narceine</b> (BBN254)		445.46	C <sub>23</sub> H <sub>27</sub> NO <sub>8</sub>	Species: <i>P. somniferum</i> L. (Papaveraceae family).	[5]
6	<b>Aspidospermine</b> (BBN44)		354.49	C <sub>22</sub> H <sub>30</sub> N <sub>2</sub> O <sub>2</sub>	Aspidosperma species: <i>Aspidosperma album</i> , <i>Aspidosperma australe</i> , <i>Aspidosperma exaltatum</i> , <i>Aspidosperma peroba</i> , <i>Aspidosperma polynuron</i> ,	[6]

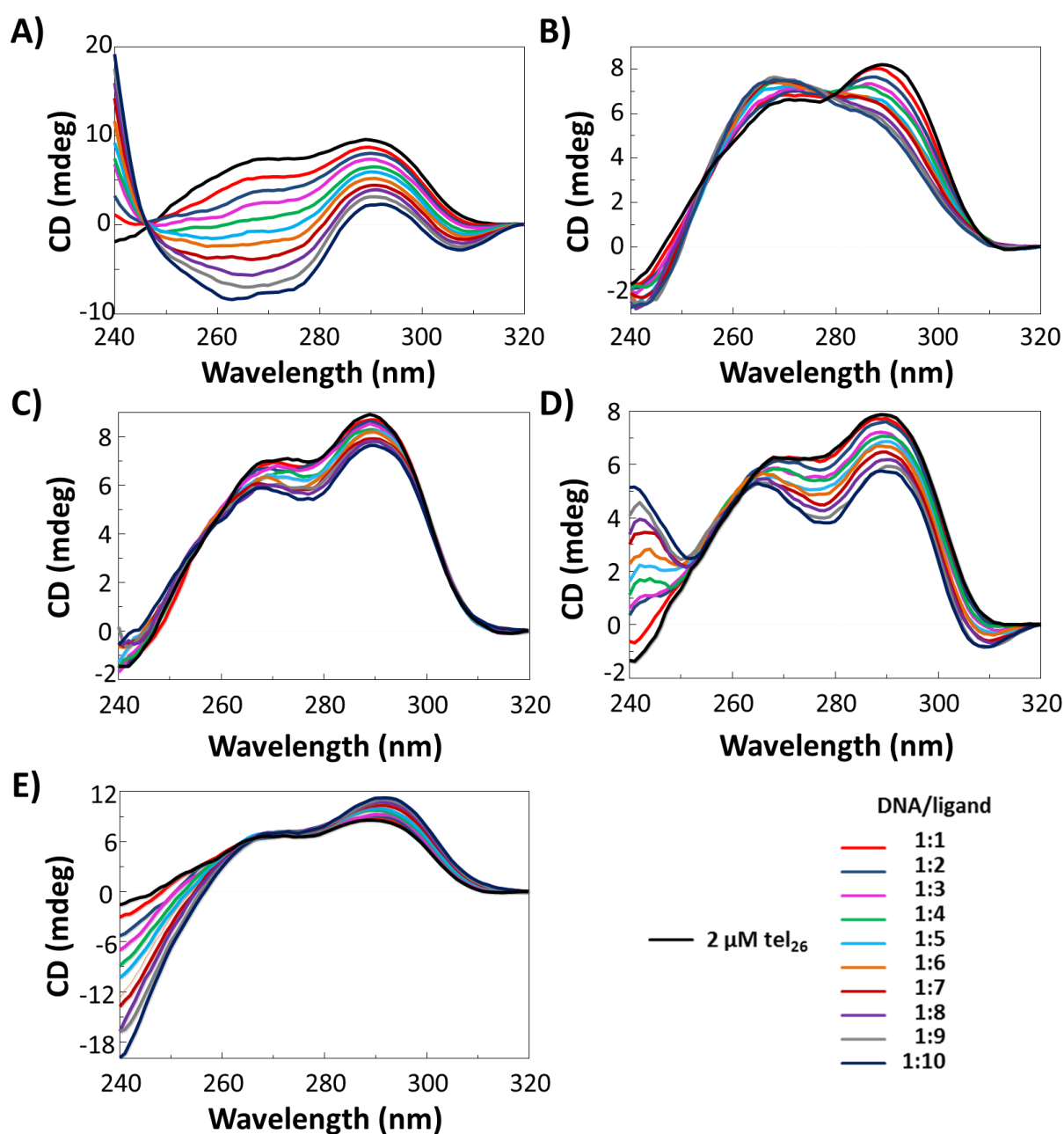
						<i>Aspidosperma pyricollum</i> , <i>Aspidosperma pyrifolium</i> , <i>Aspidosperma quebracho-blanco</i> , <i>Aspidosperma quirandy</i> , <i>Aspidosperma sessiflorum</i> , <i>Aspidosperma rhombeosignatum</i> (Apocynaceae family)
7	Vomicine (BBN186)		380.44	C <sub>22</sub> H <sub>24</sub> N <sub>2</sub> O <sub>4</sub>	Species: <i>Strychnos icaja</i> (Loganiaceae family)	[7, 8]
8	Ibogaine (BBN236)		310.43	C <sub>20</sub> H <sub>26</sub> N <sub>2</sub> O	Species: <i>Tabernanthe iboga</i> (Apocynaceae family)	[9]
9	Yohimbine·HCl (BBN174)		354.44 390.90 (+HCl)	C <sub>21</sub> H <sub>26</sub> N <sub>2</sub> O <sub>3</sub> ·HCl	Species: <i>Aspidosperma discolor</i> A. DC., <i>Aspidosperma excelsum</i> Benth, <i>Aspidosperma eburneum</i> F. Allem, <i>Aspidosperma marcgravianum</i> Woodson, <i>Aspidosperma oblongum</i> A. DC (Apocynaceae family)	[10]
10	Jervine (BBN47)		425.60	C <sub>27</sub> H <sub>39</sub> NO <sub>3</sub>	Veratrum species: <i>V. album</i> ; <i>V. dahuricum</i> ; <i>V. lobelianum</i> ; <i>V. nigrum</i> ; <i>V. nigrum</i> var. <i>ussuriense</i> ; <i>V. patulum</i> ; <i>V. stenophyllum</i> ; <i>V. taliense</i> (Buxaceae family)	[11]
11	Solanidine (BBN209)		397.65	C <sub>27</sub> H <sub>43</sub> NO	Veratrum species: <i>V. mentzeanum</i> ; <i>V. taliense</i> (Buxaceae family) Fritillaria species: <i>F.</i>	[11]

						<i>camtschatcensis</i> ; <i>F. tortifolia</i> ; <i>F. ussuriensis</i> (Liliaceae family)
12	<b>Vindoline (BBN218)</b>		456.53	$C_{25}H_{32}N_2O_6$	Species: <i>Catharanthus roseus</i> (Apocynaceae family)	[12]
13	<b>Veratrine.HCl (BBN173)</b>		591.73 628.19 (+HCl)	$C_{32}H_{49}NO_9 \cdot HCl$	Species: <i>Veratrum lobelianum</i> ( <u>Melanthiaceae</u> family)	[11]
<b>Phenolic Compounds</b>						
<b>Anthranoids</b>						
14	<b>Ferruginin A (BBN240)</b>		460.60	$C_{30}H_{36}O_4$	Species: <i>Vismia baccifera</i> var. <i>ferruginea</i> and <i>Vismia decipiens</i> (Hypericaceae family)	[13, 14]
15	<b><math>\gamma, \gamma'</math>-OH-Ferruginin A (BBN35)</b>		492.61	$C_{30}H_{36}O_6$	Species: <i>Vismia guaranirangae</i> (Hypericaceae family)	[13, 15]
16	<b>Ferruginin B (BBN161)</b>		460.61	$C_{30}H_{36}O_4$	Species: <i>Vismia baccifera</i> var. <i>ferruginea</i> and <i>Vismia decipiens</i> (Hypericaceae family)	[13, 14]
17	<b>Ferruanthrone (BBN257)</b>		460.61	$C_{30}H_{36}O_4$	Species: <i>Vismia baccifera</i> var. <i>ferruginea</i> and <i>Vismia decipiens</i> (Hypericaceae family)	[13, 14]

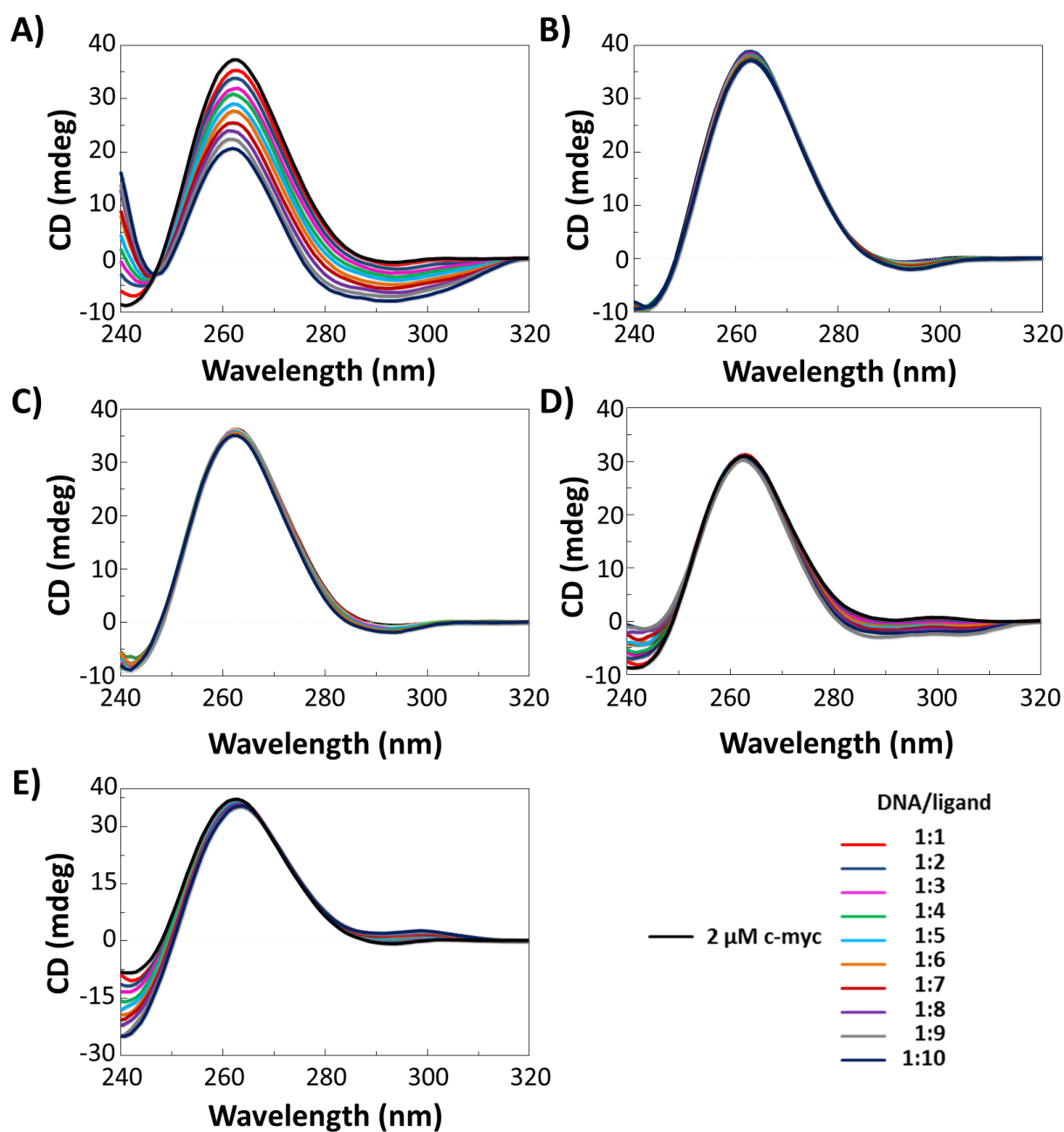
## Steroids

23	Diosgenin (BBN256)		414.63	C <sub>27</sub> H <sub>42</sub> O <sub>3</sub>	Species: <i>Dracaena cambodiana</i> and <i>Chlorophytum laxum</i> (Asparagaceae family) Species: <i>Balanites aegyptiaca</i> (Zygophyllaceae or Balanitaceae family)	[21]
24	Digitonin (BBN253)		1229.32	C <sub>56</sub> H <sub>92</sub> O <sub>29</sub>	Species: <i>Digitalis lanata</i> and <i>Digitalis purpurea</i> (Scrophulariaceae family)	[22]
25	20-OH-Ecdysone (BBN258)		480.64	C <sub>27</sub> H <sub>44</sub> O <sub>7</sub>	Species: <i>Chenopodium quinoides</i> Willd. (Chenopodiaceae family)	[23]
Diels Alder type adduct						
26	Kuwanon G (BBN99)		692.72	C <sub>40</sub> H <sub>36</sub> O <sub>11</sub>	Species: <i>Morus nigra</i> (Moraceae family)	[24]
Dibenzofuran						
27	Usnic acid (BBN66)		344.32	C <sub>18</sub> H <sub>16</sub> O <sub>7</sub>	Species: <i>Ramalina hierensis</i> (Ramalinaceae family)	[25]
Cyanogenic glycoside						

28	Amygdaline (BBN96)		455.46	$C_{21}H_{29}NO_{10}$	Seeds of numerous members of Rosaceae family	[26]
----	-----------------------	---	--------	-----------------------	--	------

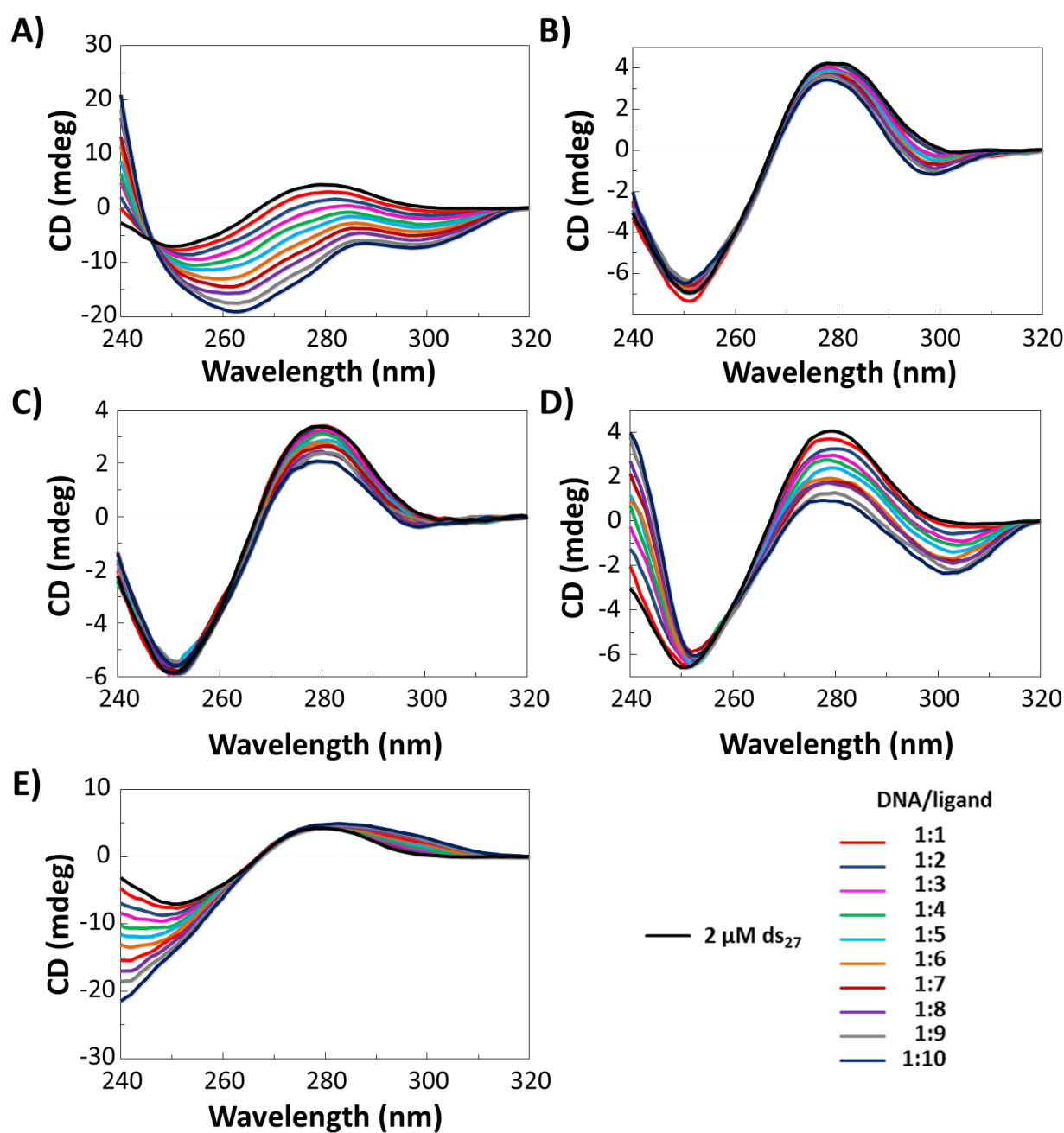


**Figure S1.** CD spectra of 2  $\mu$ M solutions of tel<sub>26</sub> G4 in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7) in the presence of increasing amounts (up to 10 equivalents) of (A) Bulbocapnine, (B) Chelidonine, (C) Ibogaine, (D) Rotenone and (E) Vomicine.

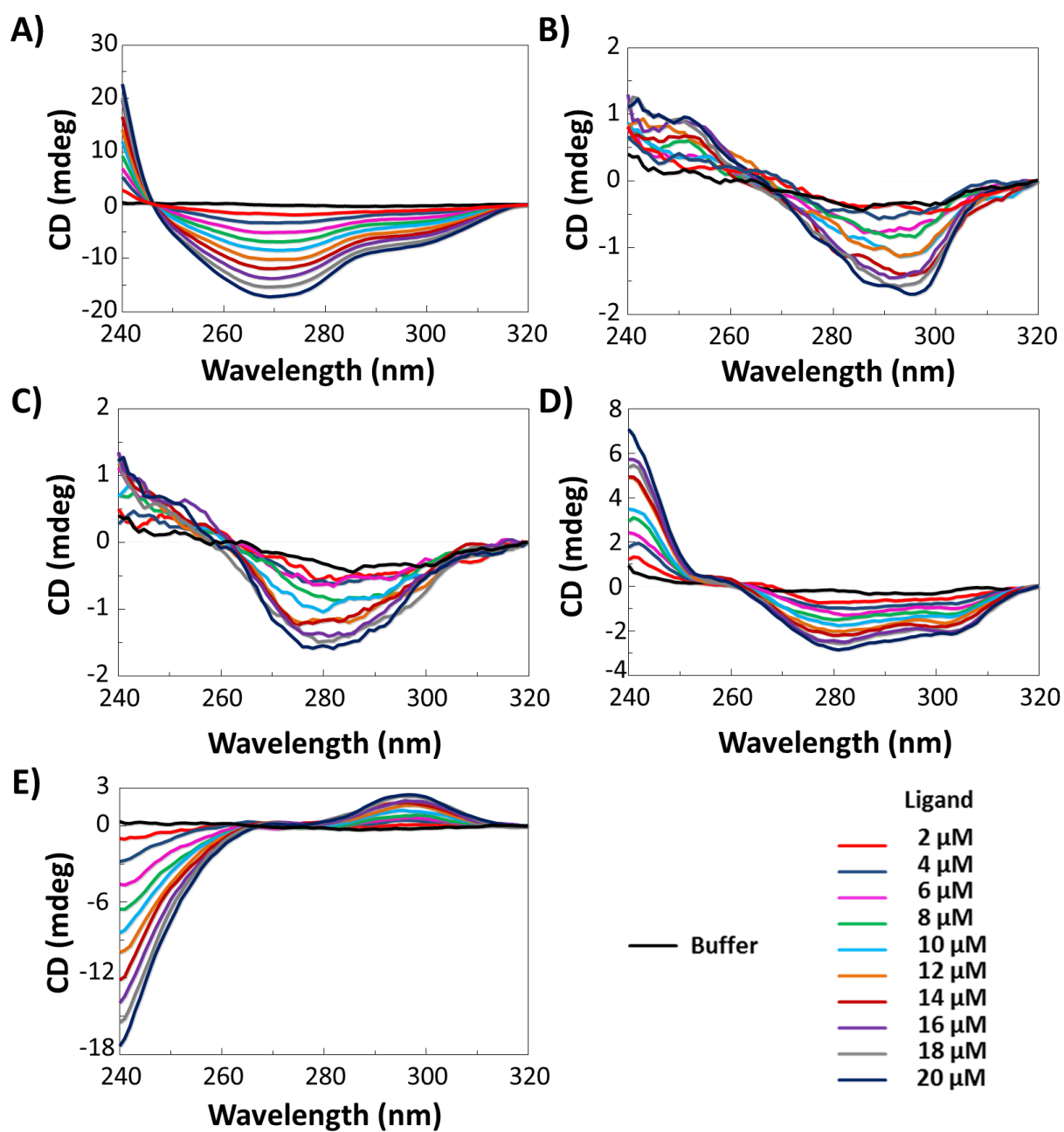


**Figure S2.** CD spectra of 2  $\mu$ M solutions of c-myc G4 in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7) in the presence of increasing amounts (up to 10 equivalents) of (A) Bulbocapnine, (B) Chelidonium, (C) Ibogaine, (D) Rotenone and (E) Vomicine.

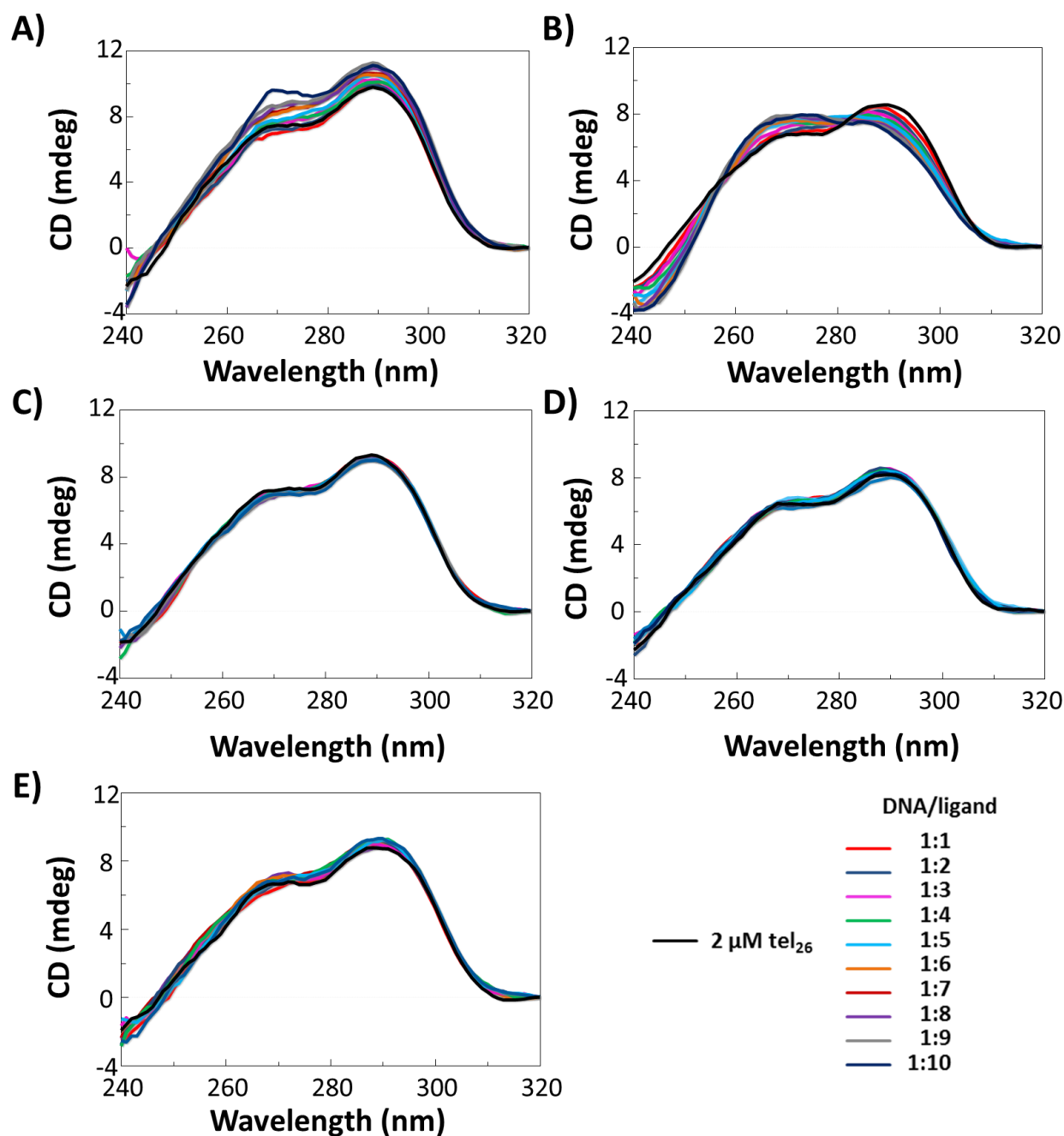




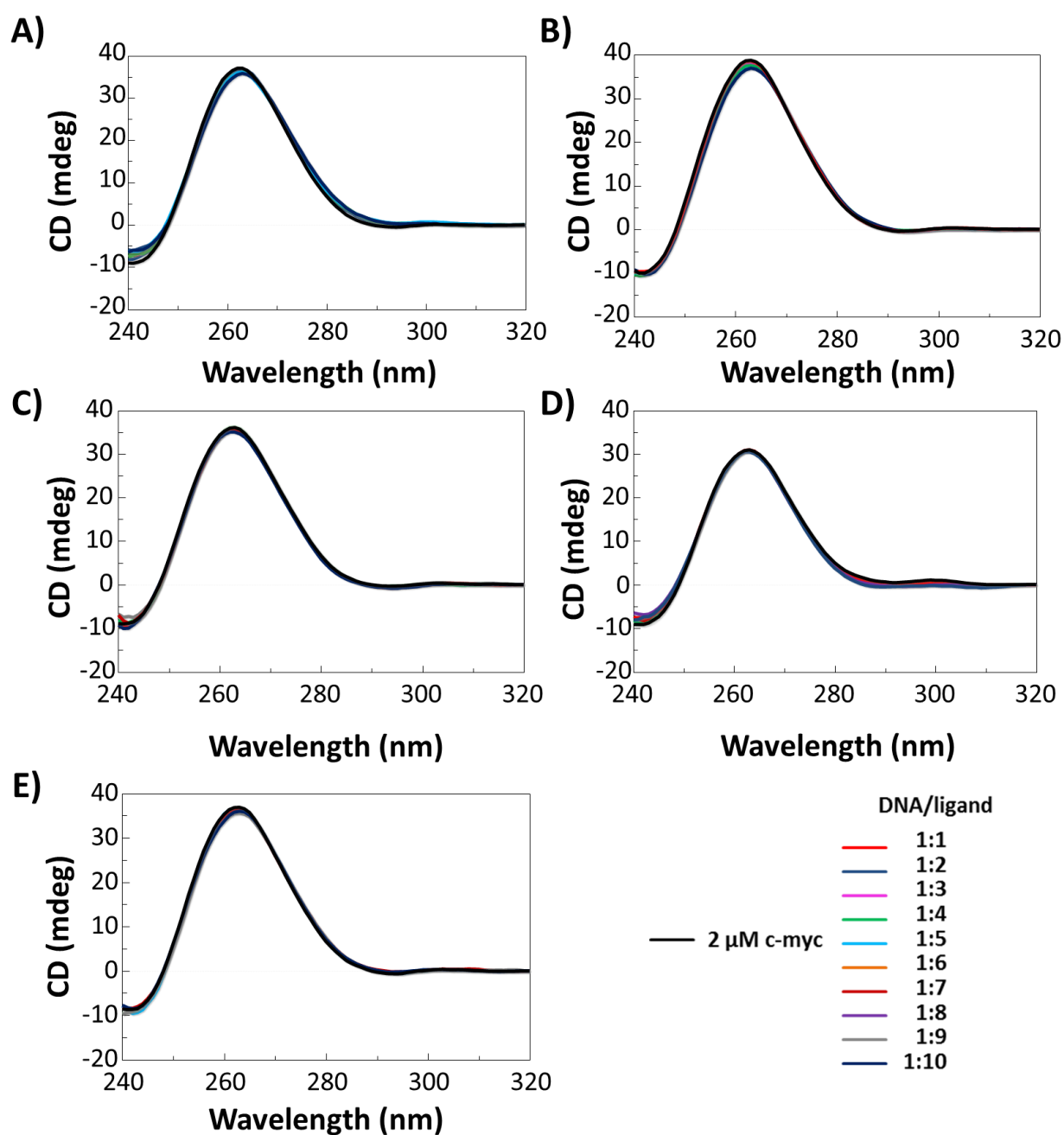
**Figure S3.** CD spectra of 2  $\mu$ M solutions of  $ds_{27}$  duplex in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7) in the presence of increasing amounts (up to 10 equivalents) of (A) Bulbocapnine, (B) Chelidonine, (C) Ibogaine, (D) Rotenone and (E) Vomicine.



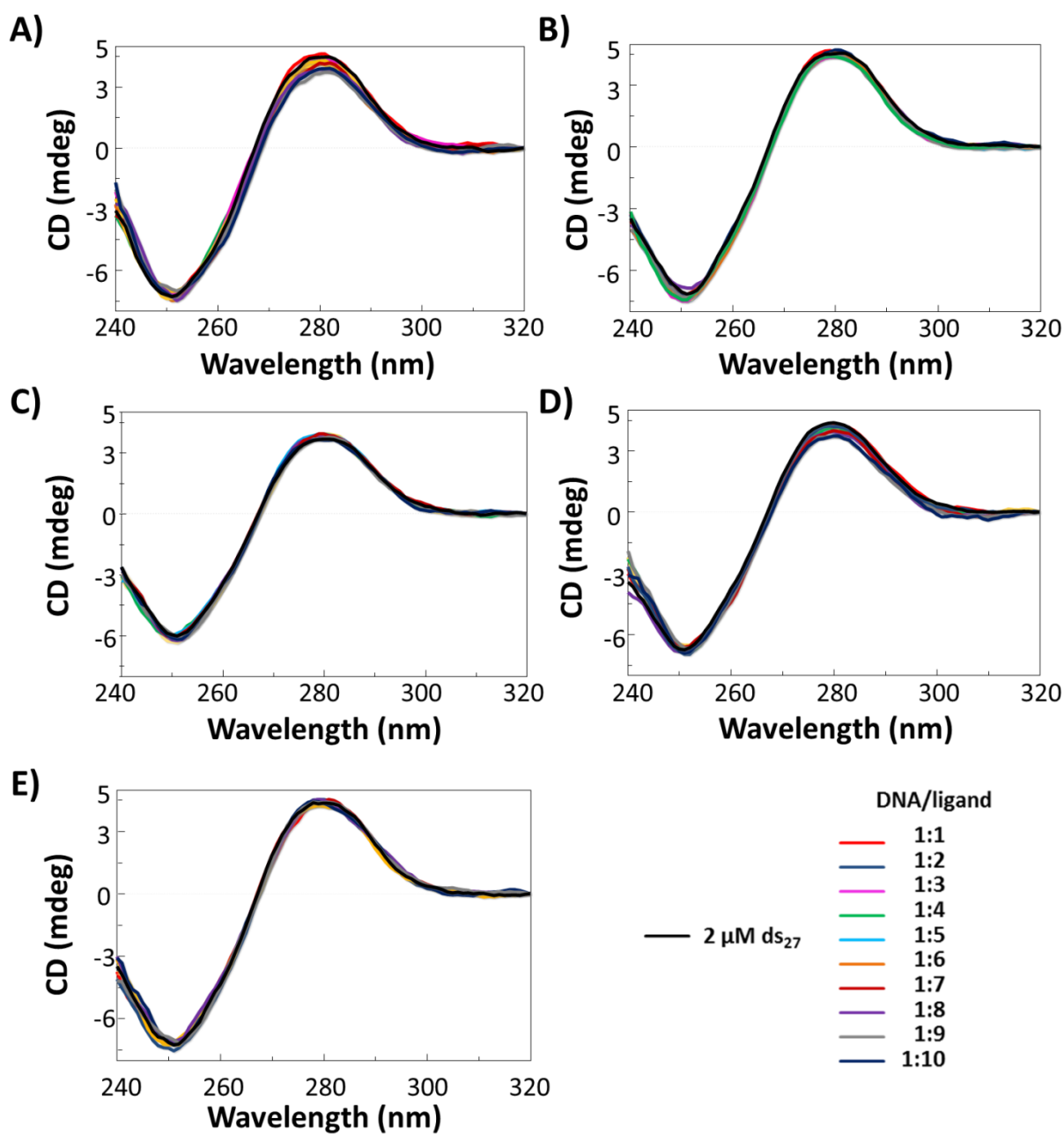
**Figure S4.** CD spectra of solutions (from 2 to 20  $\mu\text{M}$ ) of (A) Bulbocapnine, (B) Chelidonine, (C) Ibogaine, (D) Rotenone and (E) Vomicine in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7).



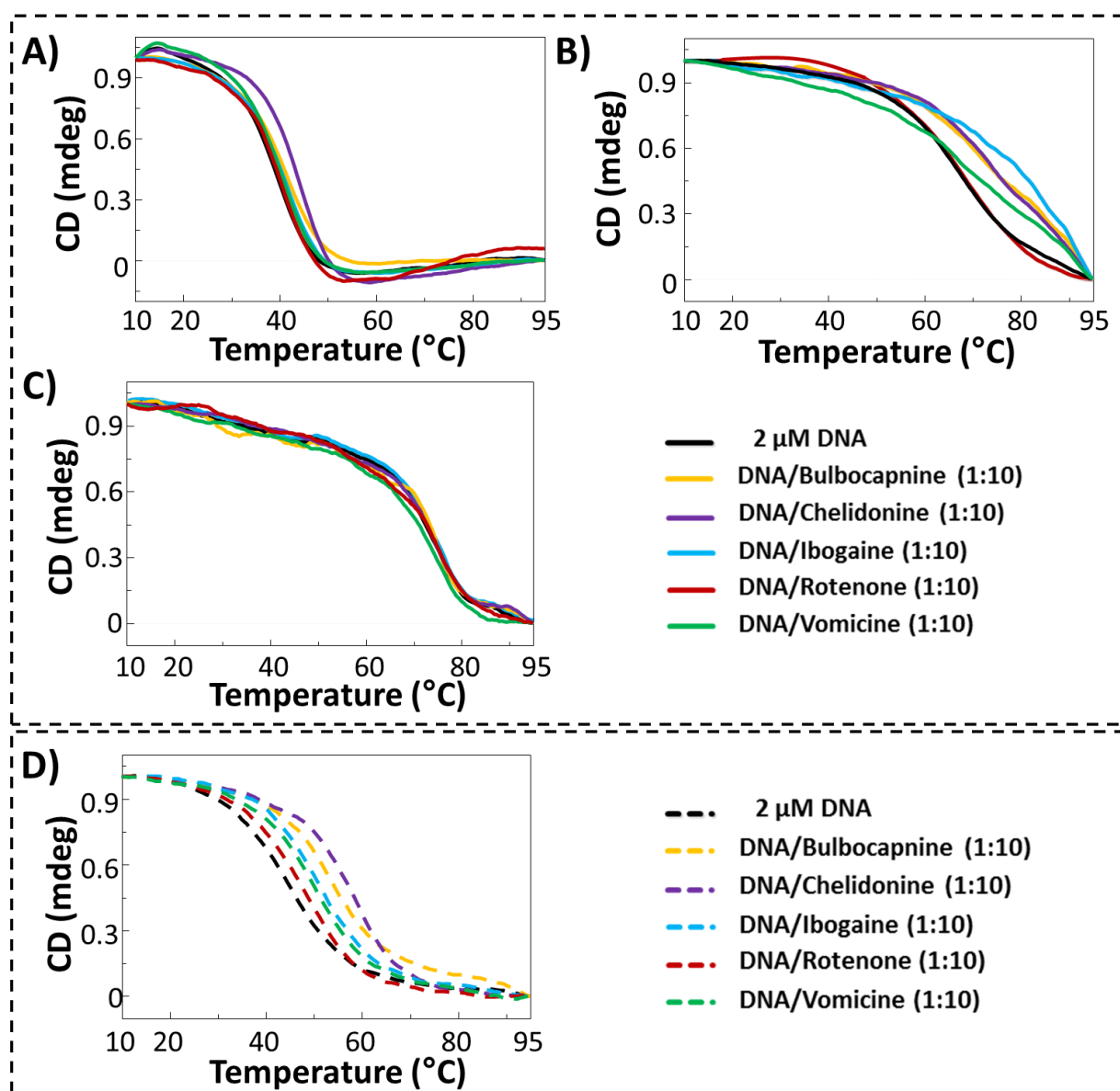
**Figure S5.** CD spectra (after ligand contribution subtraction) of 2  $\mu\text{M}$  solutions of tel<sub>26</sub> G4 in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7) with increasing amounts (up to 10 molar equivalents) of (A) Bulbocapnine, (B) Chelidونية, (C) Ibogaine, (D) Rotenone and (E) Vomoxetine.



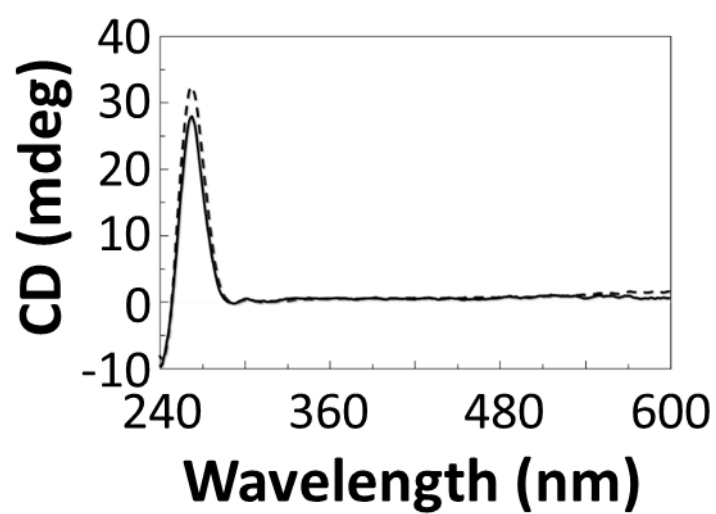
**Figure S6.** CD spectra (after ligand contribution subtraction) of 2  $\mu$ M solutions of c-myc G4 in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7) with increasing amounts (up to 10 molar equivalents) of (A) Bulbocapnine, (B) Chelidonine, (C) Ibogaine, (D) Rotenone and (E) Vomicine.



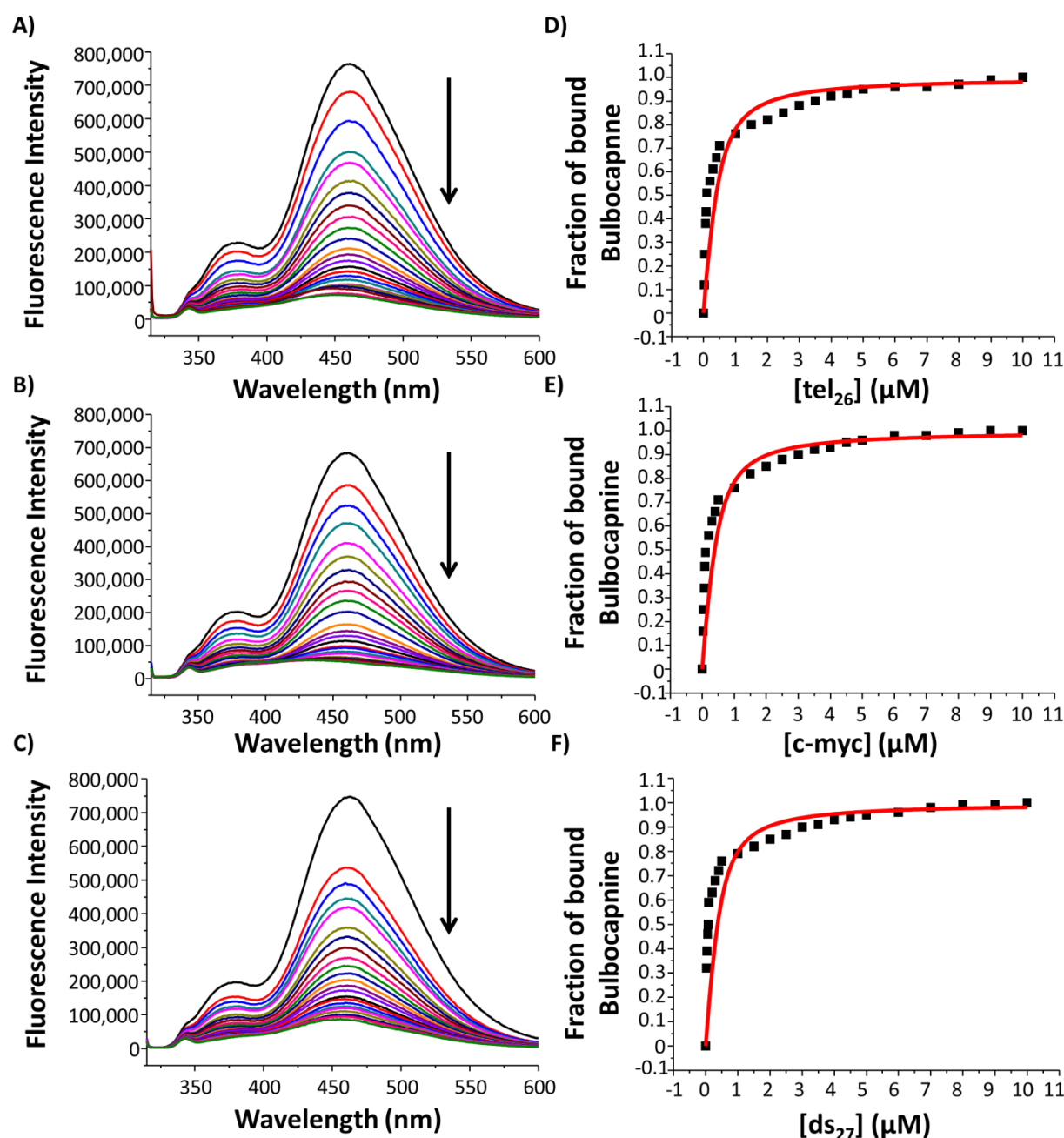
**Figure S7.** CD spectra (after ligand contribution subtraction) of 2  $\mu$ M solutions of ds<sub>27</sub> duplex in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7) with increasing amounts (up to 10 molar equivalents) of (A) Bulbocapnine, (B) Chelidone, (C) Ibogaine, (D) Rotenone and (E) Vomicine.



**Figure S8.** CD-melting curves (solid lines) for (A) tel<sub>26</sub>/ligand, (B) c-myc/ligand and (C) ds<sub>27</sub>/ligand mixtures (1:10) in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7), recorded at 290, 262 and 251 nm, respectively, and CD-melting curves (dashed lines) for (D) c-myc/ligand mixtures (1:10) in 0.5 mM KCl, 0.5 mM phosphate buffer, 5% DMSO (pH 7), recorded at 262 nm.

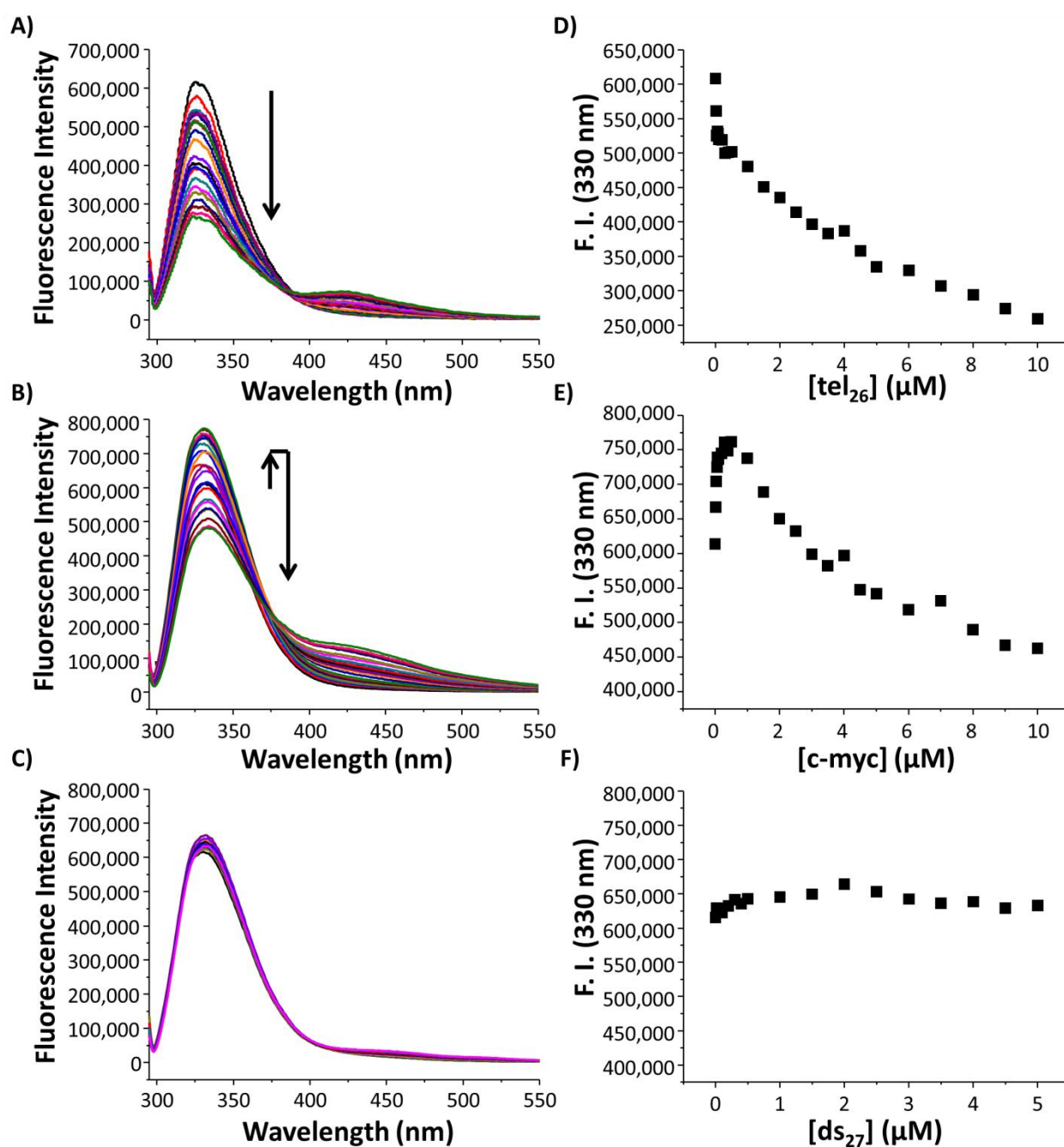


**Figure S9.** CD spectra for 2  $\mu$ M solutions of c-myc G4 in 5 mM KCl, 5 mM phosphate buffer, 5% DMSO (pH 7) (dashed line) or in 0.5 mM KCl, 0.5 mM phosphate buffer, 5% DMSO (pH 7) (solid line).

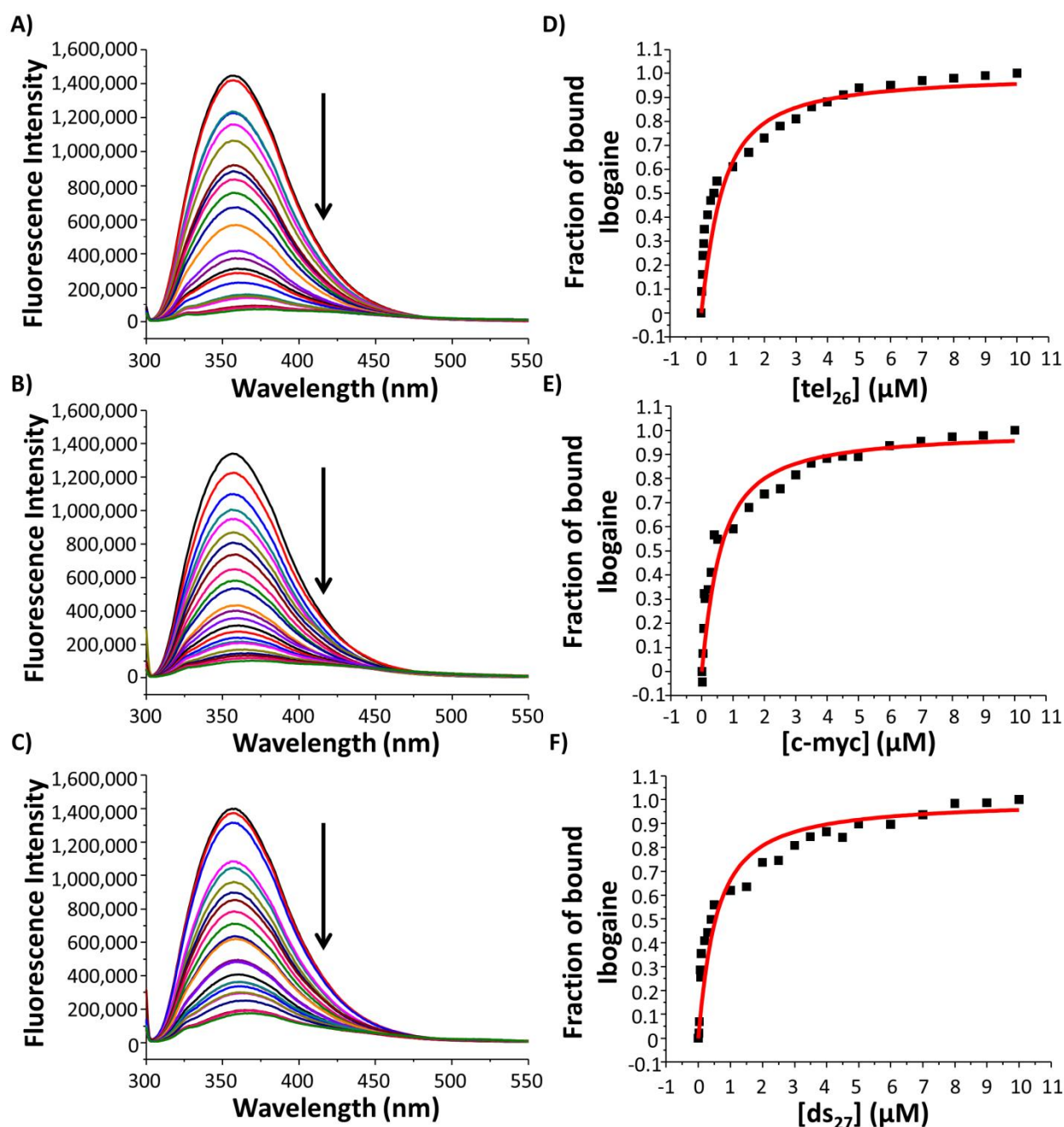


**Figure S10.** Left: Fluorescence emission spectra obtained by adding increasing amounts of (A) tel<sub>26</sub> G4, (B) c-myc G4 and (C) ds<sub>27</sub> duplex to 2 μM solutions of Bulbocapnine. Arrows indicate the variation of fluorescence intensity on increasing DNA concentration. Right: Representative binding curves obtained by plotting the fraction of bound Bulbocapnine to (D) tel<sub>26</sub> G4, (E) c-myc G4 and (F) ds<sub>27</sub> duplex as a function of the DNA concentration. The black squares represent the experimental data; the red lines represent the best fit obtained using an independent and equivalent-sites model.





**Figure S11.** Left: Fluorescence emission spectra obtained by adding increasing amounts of (A) tel<sub>26</sub> G4, (B) c-myc G4 and (C) ds<sub>27</sub> duplex to 2 μM solutions of Chelidoniine. Arrows indicate the variation of fluorescence intensity on increasing DNA concentration. Right: Fluorescence intensity at 330 nm vs. concentration of (D) tel<sub>26</sub> G4, (E) c-myc G4 and (F) ds<sub>27</sub> duplex.



**Figure S12.** Left: Fluorescence emission spectra obtained by adding increasing amounts of (A) tel<sub>26</sub> G4, (B) c-myc G4 and (C) ds<sub>27</sub> duplex to 2 μM solutions of Ibogaine. Arrows indicate the variation of fluorescence intensity on increasing DNA concentration. Right: Representative binding curves obtained by plotting the fraction of bound Ibogaine to (D) tel<sub>26</sub> G4, (E) c-myc G4 and (F) ds<sub>27</sub> duplex as a function of the DNA concentration. The black squares represent the experimental data; the red lines represent the best fit obtained using an independent and equivalent-sites model.

## References

1. Meyer, A.; Imming, P. Benzylisoquinoline Alkaloids from the Papaveraceae: The Heritage of Johannes Gadamer (1867–1928). *J. Nat. Prod.* **2011**, *74*, 2482–2487, doi:10.1021/np2005049.
2. Colombo, M.L. Pharmacological activities of chelidonium majusl. (papaveraceae). *Pharmacol. Res.* **1996**, *33*, 127–134, doi:10.1006/phrs.1996.0019.
3. Akinboye, E.S. Biological Activities of Emetine. *Open Nat. Prod. J.* **2011**, *4*, 8–15, doi:10.2174/1874848101104010008.
4. Brown, P.N.; Roman, M.C.; Chang, C.; Jin, C.; Kuriyedath, R.; Lidstone, S.; Ly, L.; Ma, Y.-C.; Sigmund, P.; Smith, R.; et al. Determination of Hydrastine and Berberine in Goldenseal Raw Materials, Extracts, and Dietary Supplements by High-Performance Liquid Chromatography with UV: Collaborative Study. *J. AOAC Int.* **2008**, *91*, 694–701, doi:10.1093/jaoac/91.4.694.
5. Šantavý, F. Chapter 4 Papaveraceae Alkaloids. II. In *The Alkaloids: Chemistry and Physiology*; Elsevier BV, 1979; Vol. 17, pp. 385–544.
6. Guimarães, H.A.; Braz-Filho, R.; Vieira, I.J.C. <sup>1</sup>H and <sup>13</sup>C-NMR Data of the Simplest Plumeran Indole Alkaloids Isolated from *Aspidosperma* Species. *Mol.* **2012**, *17*, 3025–3043, doi:10.3390/molecules17033025.
7. Monache, F.D.; Monache, G.D.; Souza, M.A.D.M.E.; Cavalcanti, M.D.S.; Chiappeta, A. ChemInform Abstract: Isopentenylindole Derivatives and Other Components of *Esenbeckia leiocarpa*. *Chemin-* **1990**, *21*, 435–439, doi:10.1002/chin.199005299.
8. Ohiri, F.; Verpoorte, R.; Svendsen, A. The African strychnos species and their alkaloids: A review. *J. Ethnopharmacol.* **1983**, *9*, 167–223, doi:10.1016/0378-8741(83)90032-6.
9. Alper, K.R. Chapter 1 Ibogaine: A review. *The Alkaloids* **2001**, *56*, 1–38, doi:10.1016/s0099-9598(01)56005-8.
10. Amaral, A.C.F.; Ramos, A.D.S.; Ferreira, J.L.P.; Dos Santos, A.R.; Da Cruz, J.D.; De Luna, A.V.M.; Nery, V.V.C.; De Lima, I.C.; Chaves, M.H.D.C.; Silva, J.R.D.A.; et al. LC-HRMS for the Identification of  $\beta$ -Carboline and Canthinone Alkaloids Isolated from Natural Sources. *Mass Spectrometry* **2017**, *6*, 187–207, doi:10.5772/68075.
11. Li, H.-J.; Jiang, Y.; Li, P. Chemistry, bioactivity and geographical diversity of steroidal alkaloids from the Liliaceae family. *Nat. Prod. Rep.* **2006**, *23*, 735–752, doi:10.1039/b609306j.
12. Song, K.M.; Park, S.W.; Hong, W.H.; Lee, H.; Kwak, S.S.; Liu, J.R. Isolation of vindoline from *Catharanthus roseus* by supercritical fluid extraction. *Biotechnol. Prog.* **1992**, *8*, 583–586, doi:10.1021/bp00018a018.
13. Monache, G.D.; Gonzalez, J.; Bettolo, G. Prenylated benzophenones from *Vismia decipiens*. *Phytochemistry* **1980**, *19*, 2025–2028, doi:10.1016/0031-9422(80)83030-5.
14. Monache, F.; Mc Quhae, M.; Ferrari, F.; Marini-Bettolo, G. Ferruginin A and B and ferruanthrone, new triprenylated anthranoids from *Vismia baccifera* var. *ferruginea*. *Tetrahedron* **1979**, *35*, 2143–2149, doi:10.1016/0040-4020(79)87031-3.
15. Monache, F.D.; Torres, F.F.; Marini-Bettolo, G.B.; De Lima, R.A. Chemistry of *Vismia* Genus. Note V:  $\gamma$ -Hydroxy- and  $\gamma,\gamma'$ -Dihydroxy-Ferruginin A. *J. Nat. Prod.* **1980**, *43*, 487–494, doi:10.1021/np50010a009.
16. Patel, K.; Patel, D.K. Medicinal importance, pharmacological activities, and analytical aspects of aloin: A concise report. *J. Acute Dis.* **2013**, *2*, 262–269, doi:10.1016/s2221-6189(13)60141-9.
17. Monache, F.D.; Ferrari, F.; Marini-Bettolo, G.B.; Maxfield, P.; et al. Vismiones from *Vismia baccifera* var. *dealdata* (H.B.K.): Chemistry and X-ray structure determination. *Gazz. Chim. Ital.* **1979**, *109*, 301–310.
18. Delfel, N.E.; Tallent, W.H.; Carlson, D.G.; Wolff, I.A. Distribution of rotenone and deguelin in *Tephrosia vogelii* and separation of rotenoid-rich fractions. *J. Agric. Food Chem.* **1970**, *18*, 385–390, doi:10.1021/jf60169a053.
19. Irvine, J.E.; Freyre, R.H. Source Materials for Rotenone, Occurrence of Rotenoids in Some Species of the Genus *Tephrosia*. *J. Agric. Food Chem.* **1959**, *7*, 106–107, doi:10.1021/jf60096a002.
20. Gonzalez, J.G.; Olivares, E.M.; Monache, F.D. Citrans and cyclols from *Clusia multiflora*. *Phytochemistry* **1995**, *38*, 485–489, doi:10.1016/0031-9422(94)00642-7.
21. Sobolewska, D.; Galanty, A.; Grabowska, K.; Makowska-Wąs, J.; Wróbel-Biedrawa, D.; Podolak, I. Saponins as cytotoxic agents: an update (2010–2018). Part I—steroidal saponins. *Phytochem. Rev.* **2020**, *19*, 139–189, doi:10.1007/s11101-020-09661-0.
22. Jograna, M.; Patil, D.; Kotwal, S.V., Digitalis species a potent herbal drug: A review on their pharmacognosy and pharmacological activities. *J. Curr. Pharms Res.* **2020**, *10*(4), 3821–3831.
23. Zughdani, M.; Yusufoglu, H.S.; Ekiz, G.; Linden, A.; Çalış, I. Ecdysteroids from the underground parts of *Rhaponticum acule* (L.) DC. *Phytochemistry* **2020**, *180*, 112530, doi:10.1016/j.phytochem.2020.112530.
24. Mascarello, A.; Chiaradia-Delatorre, L.D.; Mori, M.; Terenzi, H.; Botta, B. Mycobacterium tuberculosis-Secreted Tyrosine Phosphatases as Targets Against Tuberculosis: Exploring Natural Sources in Searching for New Drugs. *Curr. Pharm. Des.* **2016**, *22*, 1561–1569, doi:10.2174/1381612822666160112130539.
25. González, A.G.; Pérez, E.M.R.; Barrera, J.B. Biologically Active Compounds from the Lichen *Ramalina hierrensis*. *Planta Medica* **1991**, *57*, A2, doi:10.1055/s-2006-960238.
26. Botelho, A.F.M.; Pierezan, F.; Soto-Blanco, B.; Melo, M.M. A review of cardiac glycosides: Structure, toxicokinetics, clinical signs, diagnosis and antineoplastic potential. *Toxicon* **2019**, *158*, 63–68, doi:10.1016/j.toxicon.2018.11.429.