

Supplementary Materials: Quasi-irreversible Inhibition of CYP2D6 by Berberine

Ha Gyeong Kim, Han Sol Lee, Jang Su Jeon, Young Jae Choi, Yeon Jung Choi, So-Yeol Yoo, Eun-yeong Kim, Kiho Lee, InWha Park, MinKyun Na, Han-Jin Park, Seung-Woo Cho, Jong-Hoon Kim, Jae-Young Lee and Sang Kyum Kim

1. Preparation of Tetrahydroberberrubine

1.1. Extraction and Isolation of Tetrahydroberberrubine

The fruits of *Nandina domestica* (3.7 kg) were extracted three times with MeOH (3L) at room temperature for 7 days to obtain the MeOH extract (360.0 g). After acid-base extraction, an alkaloid containing fraction (NDA, 15.8 g) was separated into 7 fractions (NDA-1 to NDA-7) by MPLC (SNAP Cartridges KP-Sil, 340 g) using a mixed solvent of *n*-Hexane-EtOAc-formic acid (600:10:3, 600:20:5, 600:30:15, 400:30:15, 300:30:15, 200:30:15, and 100% MeOH). Fraction NDA-6 (1.1 g) was purified by C₁₈ HPLC eluting with MeCN-H₂O (linear gradient from 10:90 to 70:30) to yield tetrahydroberberrubine (200 mg).

Tetrahydroberberrubine-acetate: green gum; $[\alpha]_D^{20}$ -188 (c 0.5, MeOH); ¹H NMR (methanol-*d*₄, 300 MHz) δ_H 6.88 (1H, d, *J* = 8.1 Hz, H-11), 6.87 (1H, s, H-1), 6.73 (1H, d, *J* = 8.1 Hz, H-12), 6.68 (1H, s, H-4), 5.94 (2H, s, -OCH₂O-), 4.53 (1H, d, *J* = 15.8 Hz, H₂-8), 4.18 (1H, dd, *J* = 4.2, 11.7 Hz, H-14), 3.98 (1H, d, *J* = 15.8 Hz, H₂-8), 3.83 (3H, s, -OCH₃), 3.59 (1H, m, H₂-13), 3.52 (1H, m, H₂-6), 3.16 (1H, m, H₂-5), 2.94 (2H, m, H₂-6 and H₂-13), 2.87 (1H, m, H₂-5), 1.96 (acetate); ¹³C NMR (methanol-*d*₄, 75 MHz) δ_C 148.7 (C-10), 148.6 (C-3), 147.0 (C-2), 143.9 (C-9), 128.0 (C-14a), 126.8 (C-8a), 126.0 (C-14a), 120.2 (C-12a), 118.0 (C-12), 112.2 (C-11), 109.3 (C-4), 106.5 (C-1), 102.6 (-OCH₂O-), 61.1 (C-14), 56.6 (-OCH₃), 53.6 (C-8), 51.8 (C-6), 34.9 (C-13), 27.9 (C-5), 21.2 (acetate).

1.2. Structure Determination of Tetrahydroberberrubine

The ¹H NMR spectroscopic data exhibited resonances assignable to two tetrasubstituted benzene rings (δ_H 6.88, 1H, d, *J* = 8.1 Hz, H-11; 6.87, 1H, s, H-1; 6.73, 1H, d, *J* = 8.1 Hz, H-12; 6.68, 1H, s, H-4), one dioxymethylene (δ_H 5.94, 2H, s), one methoxy group (δ_H 3.83, 3H, s), one methine (δ_H 4.18, 1H, dd, *J* = 4.2, 11.7 Hz, H-14), and four methylenes (δ_H 4.53, 1H, d, *J* = 15.8 Hz, H₂-8; 3.98, 1H, d, *J* = 15.8 Hz, H₂-8; 3.59, 1H, m, H₂-13; 3.52, 1H, m, H₂-6; 3.16, 1H, m, H₂-5; 2.94, 2H, m, H₂-6 and H₂-13; 2.87, 1H, m, H₂-5). Nineteen carbons signals indicating two benzene rings (δ_C 148.7, 148.6, 147.0, 143.9, 128.0, 126.8, 126.0, 120.2, 118.0, 112.2, 109.3, 106.5), one dioxymethylene (δ_C 102.6), one methoxy group (δ_C 56.6), one methine (δ_C 61.1), and four methylenes (δ_C 53.6, 51.8, 34.9, 27.9) were observed in the ¹³C NMR spectroscopic data. The structure was deduced as tetrahydroberberrubine on the basis of NMR data interpretation. However, the chemical shifts were slightly different from those in the literature [1] due to the presence of acetate (δ_H 1.96 and δ_C 21.2). Thus, the structure was identified to be tetrahydroberberrubine-acetate (Figure S9). The negative value of specific rotation ($[\alpha]_D^{20}$ -188) revealed the absolute configuration of 14S.

2. Supplementary Tables

Table S1. MRM parameters for mass spectrometric detection of analytes using API4000 Q-TRAP system.

Metabolite	Transition (<i>m/z</i>)	Retention time (min)	Declustering potential (mV)	Collision energy (mV)	Mode
Acetaminophen	152→110	2.96	46	23	ESI+
7-Hydroxycoumarin	163→107	3.26	56	31	ESI+
Hydroxybupropion	256→238	3.07	66	17	ESI+
N-Desethylamodiaquine	328→283	2.95	51	30	ESI+
4-Hydroxytolbutamide	287→171	3.36	86	59	ESI+
4-Hydroxymephenytoin	235→150	3.21	60	17	ESI+
Dextrorphan	258→157	3.05	71	51	ESI+
6-Hydroxychlorzoxazone	184→120	3.25	81	-26	ESI-
1-Hydroxymidazolam	342→324	3.23	96	29	ESI+
6β-Hydroxytestosterone	305→269	3.47	81	21	ESI+
Berberrubine	322→307	16.88	36	39	ESI+
Thalifendine	322→307	16.08	46	31	ESI+
Demethyleneberberine	324→309	15.37	41	32	ESI+
Jatrorrhizine	338→323	16.29	46	31	ESI+
Demethylenethalifendine (M1)	310→295	14.24	36	26	ESI+

ESI+: positive electrospray ionization; ESI-: negative electrospray ionization.

Table S2. Putative metabolites of berberine identified in pooled HLM using 6530 Q-TOF LC-MS/MS.

	Compound	Formula	Expected <i>m/z</i> ([M+H] ⁺)	Observed <i>m/z</i> ([M+H] ⁺)	Mass error (ppm)	<i>t_R</i> (min)	Fragment ions from Q-TOF
Reference	Berberine	C ₂₀ H ₁₈ NO ₄ ⁺	336.1230	336.1227	-0.9	20.09	321.0993, 320.0923, 306.0767, 304.0974, 292.0969, 278.0808, 275.0930
	Berberrubine	C ₁₉ H ₁₆ NO ₄ ⁺	322.1074	322.1074	0.0	18.91	307.0845, 279.0888
	Thalifendine	C ₁₉ H ₁₆ NO ₄ ⁺	322.1074	322.1075	0.3	18.05	307.0841, 279.0894
	Demthyleneberberine	C ₁₉ H ₁₈ NO ₄ ⁺	324.1230	324.1217	-4.0	16.06	309.0974, 308.0911, 294.0749, 292.0950, 280.0958, 266.0812, 263.0925
	Jatrorrhizine	C ₂₀ H ₂₀ NO ₄ ⁺	338.1387	338.1383	-1.2	18.35	323.1151, 322.1079, 308.0910, 306.1123, 294.1119, 280.0966, 279.0883, 277.1104
	M1	C ₁₈ H ₁₆ NO ₄ ⁺	310.1074	No reference			No reference
Incubation for 120 min	Berberine	C ₂₀ H ₁₈ NO ₄ ⁺	336.1230	336.1235	1.5	20.09	321.0997, 320.0928, 306.0766, 304.0973, 292.0977, 278.0811, 275.0942
	Berberrubine	C ₁₉ H ₁₆ NO ₄ ⁺	322.1074	N.D.	N.A.	N.A.	N.D.
	Thalifendine	C ₁₉ H ₁₆ NO ₄ ⁺	322.1074	322.1074	0.0	18.08	307.0847, 279.0886
	Demthyleneberberine	C ₁₉ H ₁₈ NO ₄ ⁺	324.1230	324.1223	-2.2	16.11	309.0989, 308.0908, 294.0757, 292.0963, 280.0961, 266.0813, 263.0932
	Jatrorrhizine	C ₂₀ H ₂₀ NO ₄ ⁺	338.1387	338.1382	-1.5	18.37	323.1145, 322.1076, 308.0914, 306.1124, 294.1121, 280.0961, 279.0885, 277.1091
	M1	C ₁₈ H ₁₆ NO ₄ ⁺	310.1074	310.1080	1.9	13.38	295.0824, 267.0890

N.A.: not applicable; N.D.: not detected; *t_R*: retention time.

Table 3. Putative M1 metabolite produced in pooled HLM incubated with thalifendine or demethyleneberberine.

Precursor molecules	Incubation time (min) ^a	M1 signal at <i>m/z</i> 310 ($10^3 \times \text{cps}$) ^b
Thalifendine	0	6 ± 1
	30	2200 ± 125
Demethyleneberberine	0	107 ± 3
	30	432 ± 27

^a Thalifendine or demethyleneberberine (1 μM) was incubated with pooled HLM in the presence of NADPH-generating system; ^b Each value represents the mean $\pm$ SD for three separate samples.

3. Supplementary Figures

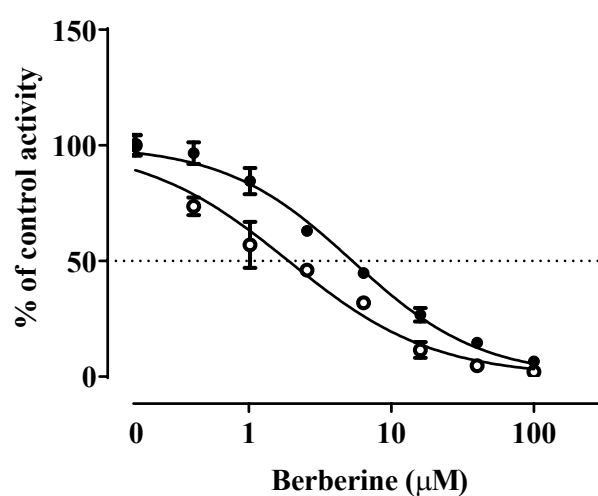


Figure S1. Changes in the inhibition curves of rhCYP2D6 by berberine after pre-incubation with (empty circle, ○) or without (solid circle, ●) NADPH for 30 min. The activity is expressed as the percentage of control samples containing no inhibitor (100%). Data show the mean $\pm$ SD of three separate samples.

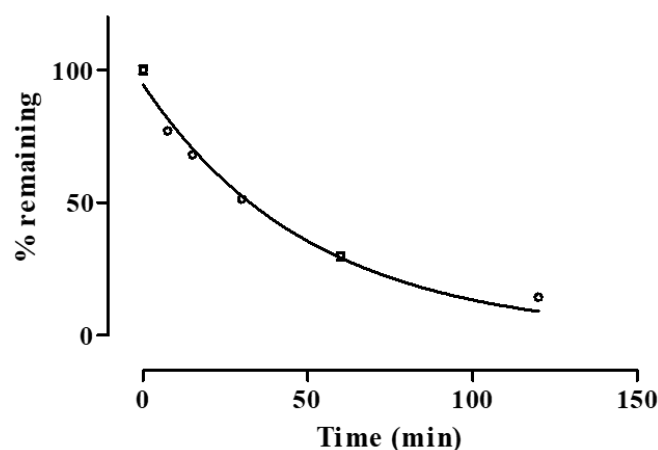


Figure S2. Metabolic stability of berberine in pooled HLM incubated with NADPH-generating system. HLM were incubated with 1 μ M berberine for 120 min. Each value represents the mean $\pm$ SD for three separate samples.

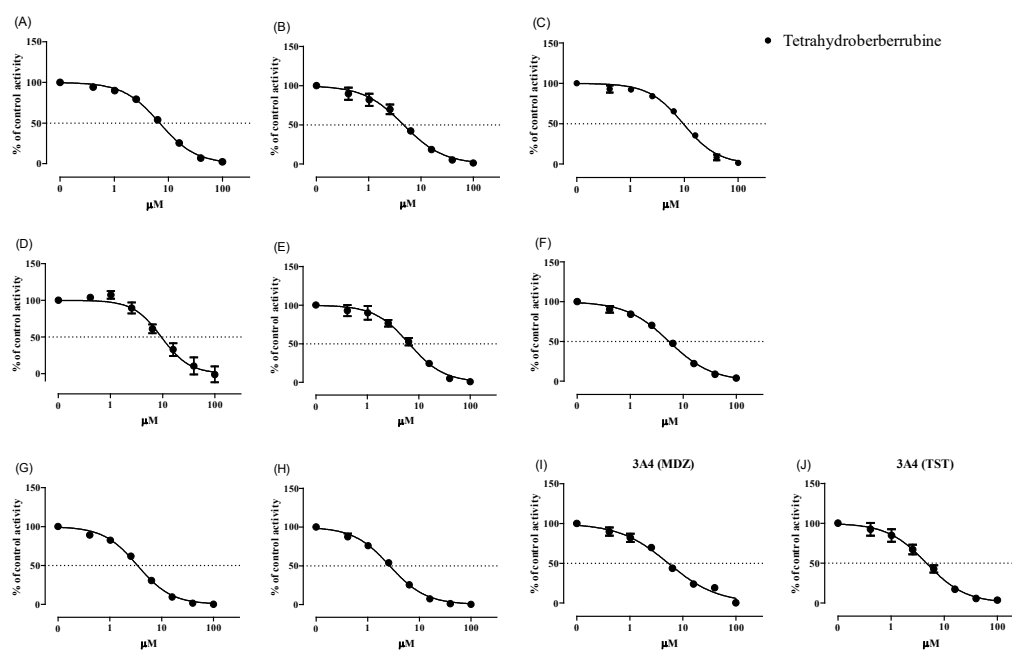


Figure S3. Effects of tetrahydroberberrubine on CYP1A2 (A), CYP2A6 (B), CYP2B6 (C), CYP2C8 (D), CYP2C9 (E), CYP2C19 (F), CYP2D6 (G), CYP2E1 (H), CYP3A4 (I, midazolam), and CYP3A4 (J, testosterone) in pooled HLM. The activity is expressed as the percentage of control samples containing no inhibitor (100%) Data show the mean $\pm$ SD of three separate samples.

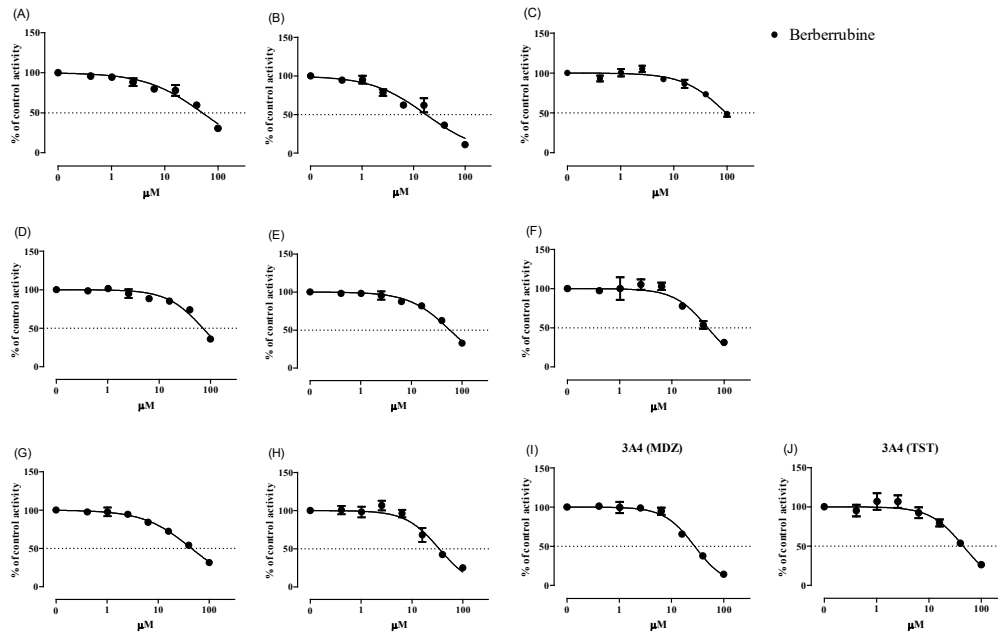


Figure S4. Effects of berberubine on CYP1A2 (A), CYP2A6 (B), CYP2B6 (C), CYP2C8 (D), CYP2C9 (E), CYP2C19 (F), CYP2D6 (G), CYP2E1 (H), CYP3A4 (I, midazolam), and CYP3A4 (J, testosterone) in pooled HLM. The activity is expressed as the percentage of control samples containing no inhibitor (100%) Data show the mean $\pm$ SD of three separate samples.

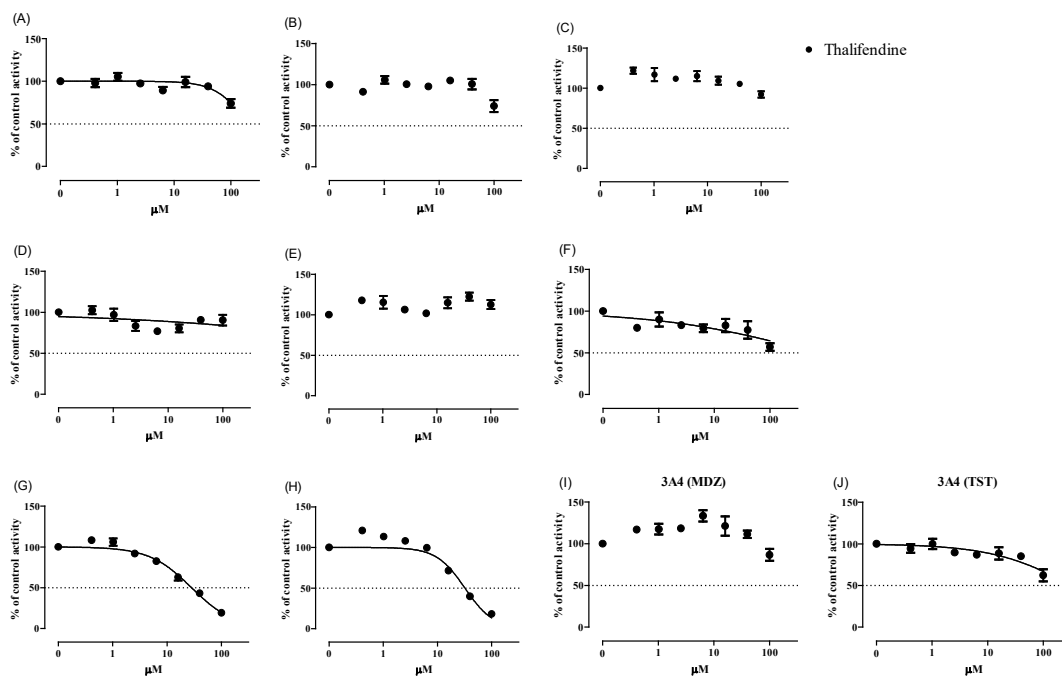


Figure S5. Effects of thalifendine on CYP1A2 (A), CYP2A6 (B), CYP2B6 (C), CYP2C8 (D), CYP2C9 (E), CYP2C19 (F), CYP2D6 (G), CYP2E1 (H), CYP3A4 (I, midazolam), and CYP3A4 (J, testosterone) in pooled HLM. The activity is expressed as the percentage of control samples containing no inhibitor (100%) Data show the mean $\pm$ SD of three separate samples.

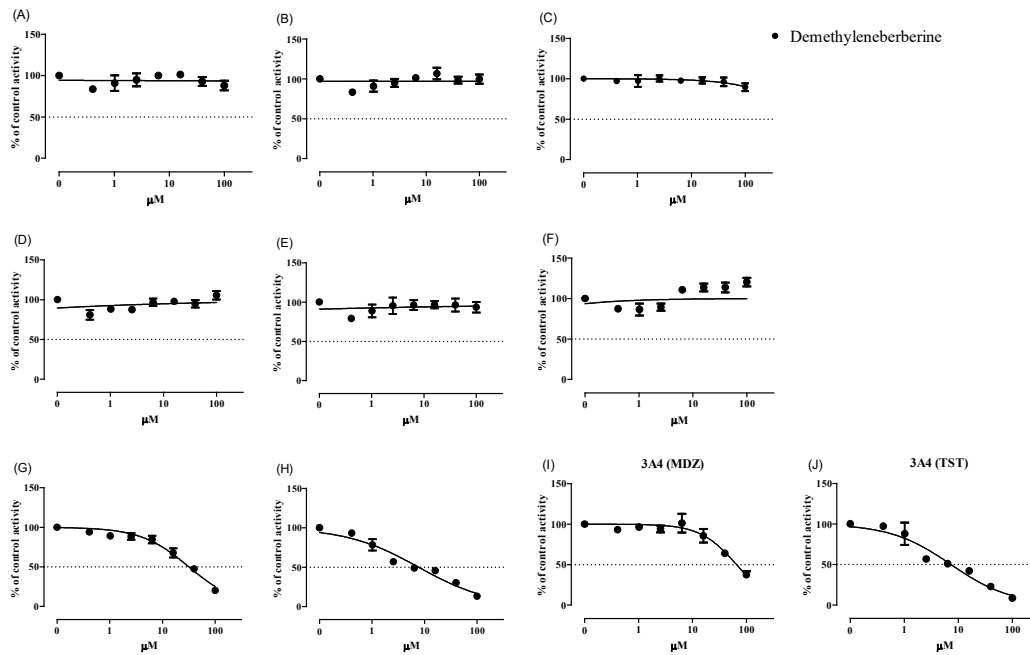


Figure S6. Effects of demethyleberberine on CYP1A2 (A), CYP2A6 (B), CYP2B6 (C), CYP2C8 (D), CYP2C9 (E), CYP2C19 (F), CYP2D6 (G), CYP2E1 (H), CYP3A4 (I, midazolam), and CYP3A4 (J, testosterone) in pooled HLM. The activity is expressed as the percentage of control samples containing no inhibitor (100%) Data show the mean $\pm$ SD of three separate samples.

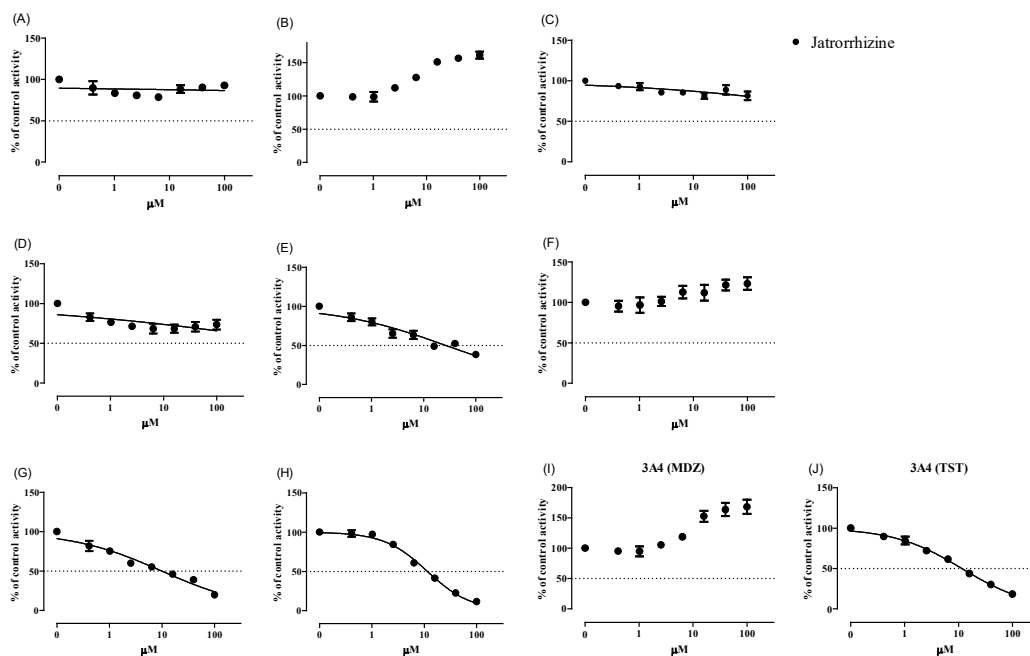


Figure S7. Effects of jatrorrhizine on CYP1A2 (A), CYP2A6 (B), CYP2B6 (C), CYP2C8 (D), CYP2C9 (E), CYP2C19 (F), CYP2D6 (G), CYP2E1 (H), CYP3A4 (I, midazolam), and CYP3A4 (J, testosterone) in pooled HLM. The activity is expressed as the percentage of control samples containing no inhibitor (100%) Data show the mean $\pm$ SD of three separate samples.

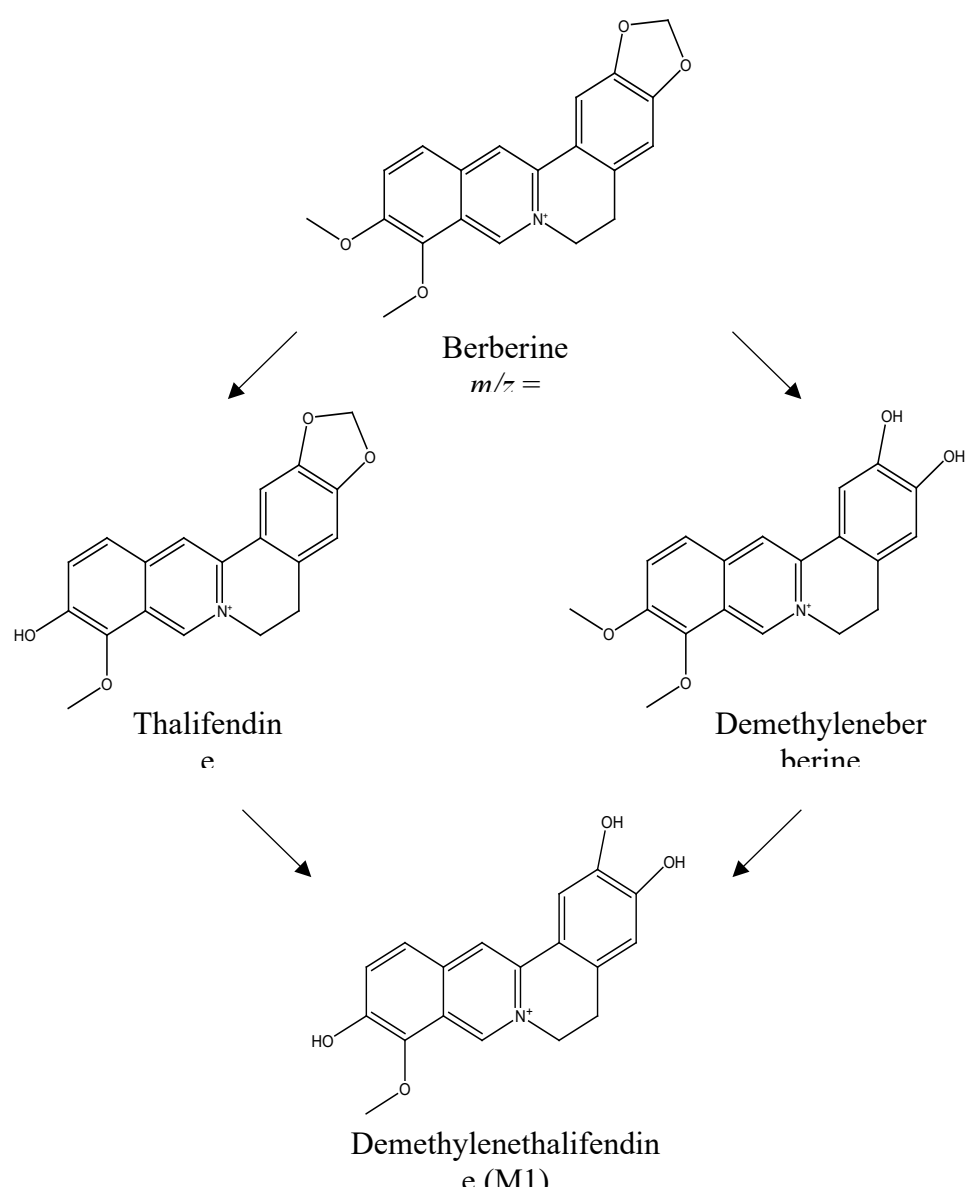


Figure S8. Proposed metabolic pathways of berberine in human liver microsomes.

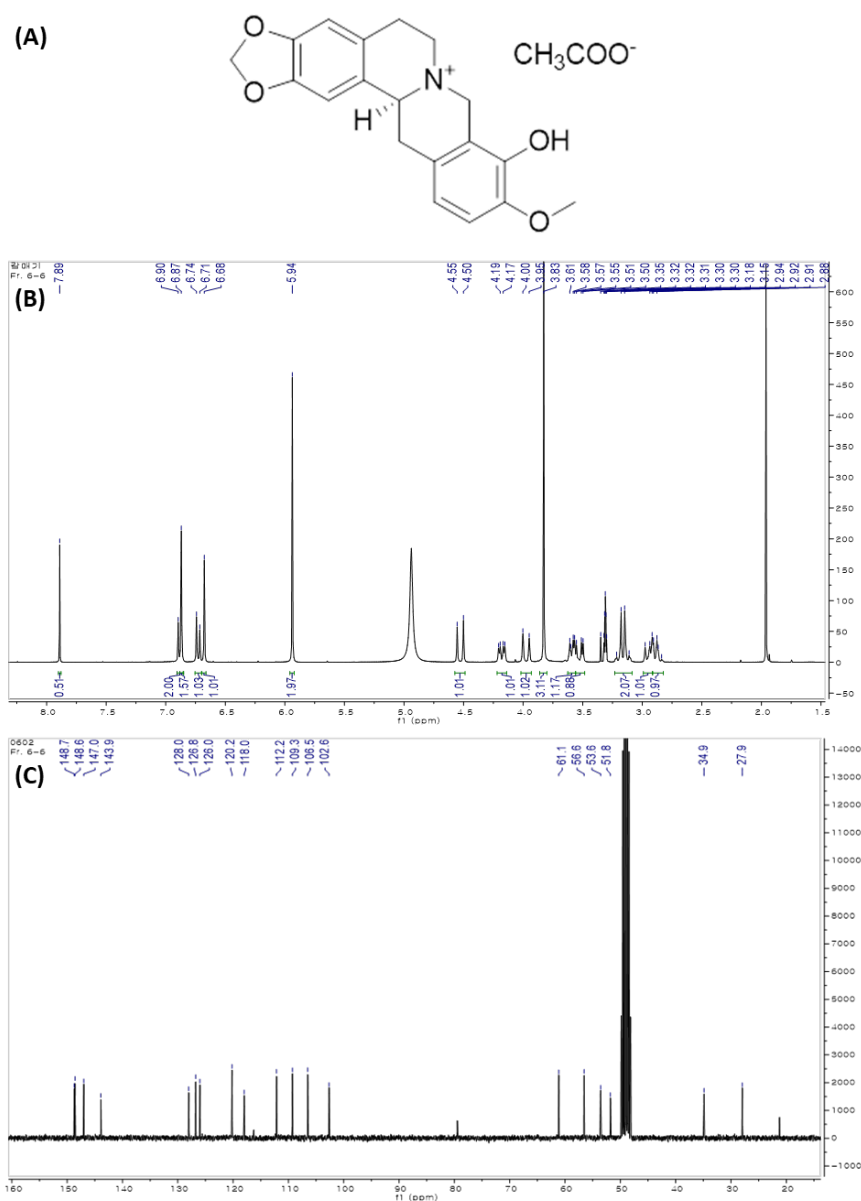


Figure S9. Chemical structure of tetrahydroberberrubine (A) The ^1H NMR (B) and ^{13}C NMR (C) spectroscopic data are presented.

4. Reference

1. Ge, H.X.; Zhang, J.; Dong, Y.; Cui, K.; Yu, B.Y. Unique biocatalytic resolution of racemic tetrahydroberberrubine via kinetic glucosylation and enantio-selective sulphation. *Chem. Commun.* **2012**, *48*, 6127–6129.



© 2020 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).