

Supplementary material

APMV-1 (avirulent)	111	G-G- R -Q-G- R -L	117
APMV-3 (wisconsin)	96	P- R -P-S-G- R -L	102
APMV-3 (Netherlands)	101	A- R -P- R -G- R -L	107
APMV-1 (virulent)	111	G- R - R -Q- K - R -F	117

Figure S1: F-Prot. cleavage site motif in APMVs: Basic amino acids (R=arginine, K=lysine) are in bold, mentioned numbers indicate amino acids position in the F protein [17].

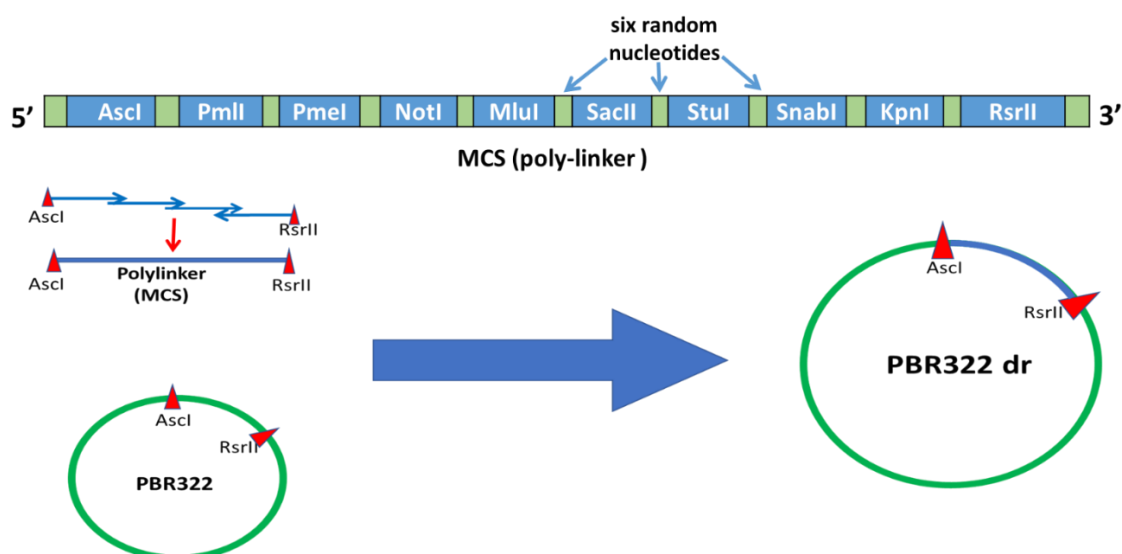


Figure S2: Preparation of the cloning plasmid vector PBR322/dr with the polylinker: A 116 nucleotides poly-linker was designed to have the same restriction enzyme sites order used to fragment the whole genome of APM3 strain Wisconsin (restriction enzyme sites were separated by random six-nucleotides). The poly-linker was created by conventional PCR using three forward and one reverse overlapping primers. The resulting gene fragment was cloned into the modified plasmid vector PBR322/dr.

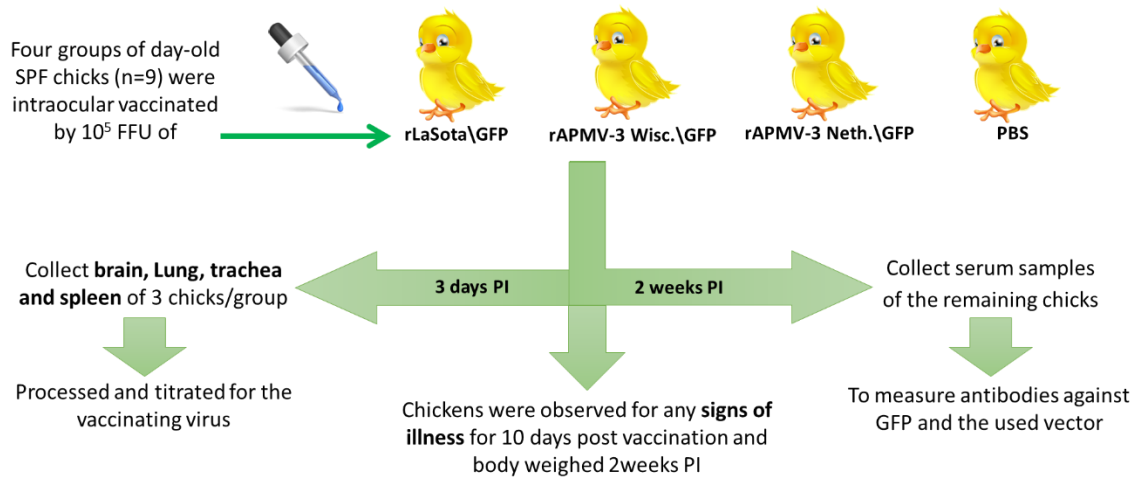


Figure S3: Schematic diagram of the in-vivo experiment for evaluation of rAPMV-3 Wisc.\GFP, rAPMV-3 Neth.\GFP and rLasota\GFP as vaccine vectors in chickens.

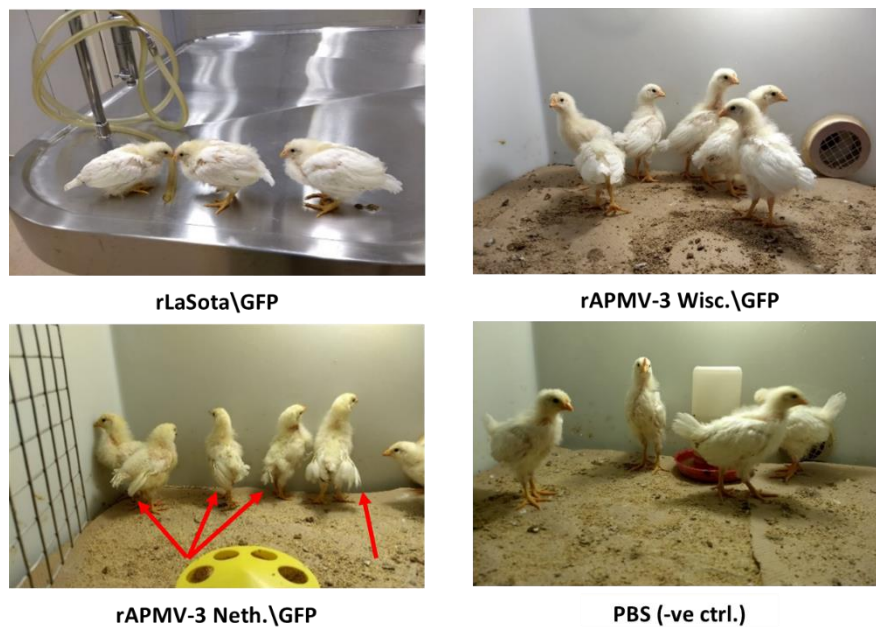


Figure S4: Pathological changes observed in vaccinated chicks (two weeks post immunization): Chicks vaccinated by rAPMV-3 Neth.\GFP are showing ruffled-abnormal feathering and stunted growth. Chicks vaccinated by rLaSota\GFP and rAPMV-3 Wisc.\GFP are showing normal body size with no signs of illness. Negative control group was mock vaccinated by PBS .

Table S1: Primers used for creation of cDNA sub-genomic fragments

Subgenomic Fragment	Primer Direction	Primers (5'-3')	Fragment size (Kbp)
I	Forward Reverse	ATT <u>CGGCGCGCCT</u> AATACGACTCACTATAGGGactaaacagaaagttaataagtgttg gtctgc <u>AcGtgggattgggtgctgatgg</u>	1.6 Kbp
II	Forward Reverse	caatcca <u>CgTg</u> cagacaggccaccgcctcc gcaatag <u>TtTaaa</u> Caagagagctagatgggttg	1.5 Kbp
III	Forward Reverse	tctctt <u>GtttAaAc</u> tattgctttataaaaaacc ggtaag <u>GCggcCgc</u> agtgagtgatttattcctg	1.4 Kbp
IV	Forward Reverse	ctcactgc <u>GgccGC</u> cttaccactagtaacaaattac gggtgtac <u>GCgtgtgt</u> ctcaacatgggactgc	2.1 Kbp
V	Forward Reverse	agacacac <u>GCgt</u> acaccctagtttctagtaaaacc ttatct <u>CcgCgg</u> caaaggtgcaataatcaattc	2.1 Kbp
VI	Forward Reverse	cctttgcc <u>GcgGag</u> ataactaagcatttaag actctt <u>aggcctg</u> ctacagctgcag	2.3 Kbp
VII	Forward Reverse	tgtagcagg <u>ccta</u> agagtcgcatc gcattata <u>cgtact</u> atttgctc	1 Kbp
VIII	Forward Reverse	aatagt <u>tacgtata</u> atgcaacc gattacgg <u>taccaa</u> actgagtgtc	2 Kbp
IX	Forward Reverse	cagtttgg <u>taccg</u> taatcgtaatac ggtc <u>CGGACCGC</u> GAGGAGGTGGAGATGCCATGCCGACCCactaaacaaaagtata taaatggtttaattaac ⁺	2.3 Kbp

Table S2: Nucleotide changes involved in pAPMV-3 Wisc.FLC

A

Subgenomic Fragment		Nucleotid number	Original nucleotide	Replacement nucleotide	Original sequence	New sequence	Introduced RE sites
I		Before (1)	<u>GGCGCGCCTAATACGACTCACTATAGGG</u>				AscI + T7 promotor
		1590	A	C	caagcg 1588-1593	<u>caCgTg</u>	PmlI site
II		1592	C	T			
		3074	A	G	attttatc 3074-3081	<u>GtttAaAc</u>	PmeI site
		3078	T	A			
III		3080	T	A	gccgccag 4389-4396	<u>gcGgccGC</u>	NotI site
		4391	C	G			
		4395	A	G			
IV		4396	G	C	accagt 6515-6520	<u>acGCgt</u>	MluI site
		6517	C	G			
V		6518	A	C	ccacgt 8575-8580	<u>ccGcgG</u>	SacII site
		8577	A	G			
VI	VI d	8580	T	G	aggcct 10896 -10901	<u>aggcct</u>	StuI site
			---	---			
	VI c				tacgta 11916 -11921	<u>tacgta</u>	SnaBI site
			---	---			
	VI b				ggtacc 13925 -13930	<u>ggtacc</u>	KpnI site
			---	---			
	VI a	After (16182)	<u>GGGTCGGCATGGCATCTCCACCTCCTCGCGGTCCG</u>				

B

Deleted RE site	Original sequence	Corrected sequence	Changed nucleotide
SacII	ccgcgg 6386-6391	TcgAgg	6358
			6361
RsrII	cggaccg 1291-1297	cCgATcg	1264
			1267
RsrII	cggaccg 2682-2688	TggCccg	2682
			2685