



# Multiplexed *tat*-targeting CRISPR-Cas9 protects T cell from acute HIV-1 infection with inhibition of viral escape

# **Supplementary Information**

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**Figure S1.** GFP-expression measurement as a preliminary assay to determine transduction titer of lentivirus. GFP-expressing pseudovirus was constructed via co-transfection of pWPT-GFP (Addgene #12255), psPAX2 (Addgene #12260) and VSV-G expressing plasmid pHIT/G. Lentiviral p24 titer was measured using p24 ELISA, p24-titrated viruses were transduced into 293T cells, and GFP expression was visualized 5 days after transduction using fluorescent microscope. Fluorescent micrograph of cells after transduced with 200 pg, 2 ng and 20 ng of lentiviruses are shown. Based on GFP expression, we determined viral titer equivalent to 20 ng of p24 as MOI of 1.



**Figure S2.** Cellular growth kinetics of Cas9-gRNA-tranduced MT4 T cells, compared to mock-infected and Cas9-only-transduced cells. Cells were seeded at the same density (5x10<sup>4</sup> cells mL<sup>-1</sup>) in 24-well and cultured in 1 mL of RPMI 1640. Viable cells were counted with trypan blue exclusion. Average values (±SE) of three replicates are shown.

tatB sample			revA sample			
AAAAGCCTTAGGCATCTCCTATG	GCAGGAAGA	A ref	GGGGACCCGACAGGCCCGAA	.GGAATAGAAGAAG	ref	
CCTTAGGCATCTCCTATG	GC	gRNA	GACAGGCCCGAA	GGAATAGA	gRNA	
AAAAGCCTTAGGCATCTCCTAAG	GCAGGAAGA	A O	GGGGACCCGAGGCCCGAA	GGAATAGAAGAAG	-2	
AAAAGCCTTAGGCATCTCCTAT	GAAGA	A -5	GGGGACCCGACGAA	GGAATAGAAGAAG	-6	
AAAAGCCTTAGGCATCTCCTAAAAGGCATCTCTTAG	GCAGGAAGA	A +13	GGGGACCCGAGTGGCCCGAA	GGAATAGAAGAAG	0	
AAAAGCCTTAGGCATCTCCTCTG	GCAGGAAGA	A O	GGGGACCCGACGGGAGGCCCGAA	GGAATAGAAGAAG	+3	
AAAAGCCTTAGGCATCTCCTAAG	GCAGGAAGA	A 0	GGGGACCCGACGGGAATAAGGCCCGAA	GGAATAGAAGAAG	+/	
AAAAGCCTTAGGCATCTCCTATA	GCAGGAAGA	A O	GGGGACCCGAA	.GGAATAGAAGAAG	-9	
AAAAGCCTTAGGCATCTCCTAT	GAAGA	A -5	GGGGACCCGACACCCCCGAA	GGAATAGAAGAAG	-0	
AAAAGCCTTAGGCATCTCCTCT	-CAGGAAGA	A -2	GGGGACCCGACTTTTGGCCCGAA	.GGAATAGAAGAAG	+3	
AAAAGCCTTAGGCATCTCCTATAAGG	GCAGGAAGA	A +3	GGGGACCCGACTTTAGGCCCGAA	.GGAATAGAAGAAG	+3	
AAAAGCCTTAGGCATCTCC	GCAGGAAGA	A -4	GAGGCCCGAA	.GGAATAGAAGAAG	-9	
AAAAGCCTTAGGCATCTCCG	GCAGGAAGA	A -3	GGGGACCCGACAACCCCGAA	.GGAATAGAAGAAG	0	
AAAAGCCTTAGGCATCTCTTATG	GCAGGAAGA	A U	GGGGACCCTGGCCCGAA	.GGAATAGAAGAAG	-3	
AAAAGCCTTAGGCA	GGAAGA	A -12	GGGGACCCC	-CAATAGAAGAAG	-12	
AAAAGG	GCAGGAAGA	A -17	GGGGACCCGACGGCCCGAA	.GGAATAGAAGAAG	-1	
AAAAGCCTTAGGCATCTCCTG	GCAGGAAGA	A -2	GGGGACCCGAGGCCCGAA	.GGAATAGAAGAAG	-2	
AAAAGCCTTAGGCATCTCCTATTG	GCAGGAAGA	A +1	GGGGACCCGAGACCCCCGAA	.GGAATAGAAGAAG	0	
AAAAGCCTTAGGCATCTCCTATG	GCAGGAAGA	A wt	GGGGACCCGTCCGAA	.GGAATAGAAGAAG	-5	
AAAAGCCTTAGGCATCTCCTATG	GCAGGAAGA	A wt	GGGGACCCGACAGGCCCGAA	.GGAATAGAAGAAG	wt	
AAAAGCCTTAGGCATCTCCTATG	GCAGGAAGA	A wt	GGGGACCCGACAGGCCCGAA	GGAATAGAAGAAG	wt	
AAAAGCCTTAGGCATCTCCTATG	GCAGGAAGA	A wt	GGGGACCCGACAGGCCCGAA	GGAATAGAAGAAG	wt	
tatA target region in tatrevABC sample		ta	tC target region in <i>tatrevABC</i> sample			
GCCAG <mark>TAGATCCTAGACTAGAGCCCTGG</mark> AAGCA	ref	AGCATCCA	GGAAGTCAGCCTAAAACTGCTTGTA	ref		
TAGATCCTAGACTAGAGCCC	gRNA		GGAAGTCAGCCTAAAACTGC	gRNA		
GCCAG <mark>TAGATCCTAGACTAGAGCCCCCTGG</mark> AAGCA	+2	AGCATCCA	GTCAGCCTAAAACTGCTTGTA	-4		
GCC <mark>TGG</mark> AAGCA	-22	AGCATCCA	.GGTACACAGCCTAAAACTGCTTGTA	0		
GCCAG <mark>TAGATCCTAGACTAGAGG</mark> AAGCA	-5	AGCATCCA	.GGTTGTCAGCCTAAAACTGCTTGTA	0		
GCCAGTAGATCCTAGACTCCA	-16	AGCATCCA	.GGAGCCTAAAACTGCTTGTA	-5		
GCCAG <mark>TAGATCCTAGACTAGAGC</mark> AAGCA	-5	AGCATCCA	GGAA-AGTCAGCCTAAAACTGCTTGTA	+1		
GCCAG <mark>TAGATCCTAGACTAGCCCCCTGG</mark> AAGCA	0	AGCATCCA	GGAAAAACTGCTTGTA	-9		
GCCAGTAGATCCTAGACTAGAGGGAAGCA	-4	AGCATCCA	<b>GGAAGAGTCAGCCTAAAACTGCTTGTA</b>	+2		
GCCAGTAGATAAGCA	-18	AGCATCCA	.GGACAGCCTAAAACTGCTTGTA	-3		
GCCAGTAGGTCCTGGAAGCA	-13	AGCA	GTCAGCCTAAAACTGCTTGTA	-8		
GCCAGTAGATCCTAGACTAGAGCTCCCTGGAAGCA	+2	AGCATCCA	.GGACTCAGCCTAAAACTGCTTGTA	-1		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	.GGTCCTAAAACTGCTTGTA	-6		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	AGTCAGCCTAAAACTGCTTGTA	-3		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	GGTCAGCCTAAAACTGCTTGTA	-3		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	GGATCTCAGCCTAAAACTGCTTGTA	0		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	GGCTGCTTGTA	-14		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	GGAAGTCAGCCTAAAACTGCTTGTA	wt		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	GGAAGTCAGCCTAAAACTGCTTGTA	wt		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	GGAAGTCAGCCTAAAACTGCTTGTA	wt		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	GGAAGTCAGCCTAAAACTGCTTGTA	wt		
GCCAGTAGATCCTAGACTAGAGCCCTGGAAGCA	wt	AGCATCCA	GGAAGTCAGCCTAAAACTGCTTGTA	wt		

**Figure S3.** Sequencing analysis of mutations accumulated inside HIV-1 proviral DNA after HIV-1 infection, Cas9 cleavage and NHEJ repair. Cellular DNA was extracted from T cells transduced with *tatB, revA*, and *tatrevABC*, 16 days after infection with replicative HIV<sub>NL43</sub>. Target gRNA regions were amplified for respective samples, with *tatA* and *tatC* regions amplified from *tatrevABC*-transduced cells. Amplicons were cloned using TA cloning vector to produce 20 samples for each gRNA target, Sanger sequenced and aligned with reference (topmost). Summary of results were shown in Figure 3. Target gRNA and PAM sequence were shown in yellow and blue box, respectively. Mutations indicated in red and Cas9-cutting site noted under the red triangles. Numbers to the right of sequences signify changes in sequence length after mutation; wt indicated wild-type, non-mutated sequences.



**Figure S4.** Colony PCR screening of constructed plasmids. (a) Schematic for the PCR amplification, shown for px330A-tatrev-6 as example. Amplicon stretched from immediate upstream of the first gRNA to immediate downstream of the last gRNA; amplification was done using primers as indicated. (b) Gel electrophoresis of amplicons. Number of bands indicate the number of gRNAs contained in the assembled clones.



**Figure S5.** Titer of gRNA-containing lentiviral vectors, produced with co-transfection of gRNAcontaining lentiCRISPRv2 with packaging plasmid psPAX2 and VSVG-pseudotyped envelope pHIT/G in Lenti-X 293T cells using FuGENE HD transfection reagent. Lentiviruses were harvested three days after transfection. Titers of the all-in-one multiplexed gRNA vectors are shown, with that of single gRNA *tatA* shown as comparison. Average values (±SE) of three replicates are shown.



**Figure S6.** Kinetics of cell density for selected CRISPR-transduced cells after infection with HIV<sub>NL4-3</sub> in titers equivalent to 2 ng (left) and 20 ng (right) of p24, as counted using trypan blue exclusion. All WT cells died from 15 days onwards. Half of cell culture were passaged every 15 days. Assays were performed three times with average values (±SE) as shown.

#### Uncropped blot and gel images

#### Original Western Blot images of Figure 1b

Original uncropped Western blot images of Figure 1b. Figures were not doctored or joined together from different photographs. Blotted PVDF membranes were incubated with anti-Cas9 (top) and anti-beta-actin (bottom) as loading control.



## Original electrophoresis gel image of Figure 2b

Original uncropped gel images of Figure 2b. Gel image were not doctored or joined together from different photographs.



# **Original Western Blot images of Figure 5b**

Original uncropped Western blot images of Figure 5b. Membrane images were not doctored or joined together from different photographs. Blotted PVDF membranes were incubated with anti-FLAG (top) and anti-beta-actin (bottom).



### Original electrophoresis gel image of Figure S4

Original uncropped gel images of Figure S4. Figures were not doctored or joined together from different images.

