



# Article Identification of Eight *Pterocarpus* Species and Two *Dalbergia* Species Using Visible/Near-Infrared (Vis/NIR) Hyperspectral Imaging (HSI)

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Abstract: Pterocarpus santalinus is considered among the finest luxury woods in the world and has potential commercial and medicinal value. Due to its rich hue and high price, Pterocarpus santalinus has often been substituted and mislabeled with other woods of lower economic value. To maintain the order of the timber market and the interests of consumers, it is necessary to establish a fast and reliable method for Pterocarpus species identification. In this study, wood samples of Pterocarpus santalinus and nine other wood samples commonly used for counterfeiting were analyzed by visible light/near-infrared (Vis/NIR) hyperspectral imaging (HSI). The spectral data were preprocessed with different algorithms. Principal component analysis (PCA) was applied in different spectral ranges: 400~2500 nm, 400~800 nm, and 800~2500 nm. Partial least squares discriminant analysis (PLS-DA) and square support vector machine (SVM) modeling methods were performed for effective discrimination. The best classification model was SVM combined with a normalization preprocessing method in whole spectral range (400~2500 nm), with prediction accuracy higher than 99.8%. The results suggest that the use of Vis/NIR-HSI in combination with chemometric approaches can be used as an effective tool for the discrimination of Pterocarpus santalinus.

Keywords: PLA-DA; SVM; chemometric; wood identification

## 1. Introduction

*Pterocarpus santalinus*, popularly known as "red sanders", is a slow-growing forest legume tree that can attain a harvestable size of 70 cm DBH at 80~100 years. *Pterocarpus santalinus* has potential commercial and medicinal value, but due to poor natural regeneration, illegal logging, over-exploitation, and microclimate changes causing the degradation of natural populations, it has become an endangered species and was listed as a vulnerable species by the International Union for Conservation of Nature in 1998 [1]. The value of wood per ton was US \$12,000 [2] and the global demand was estimated at 3000 tons per year [3]. *Pterocarpus santalinus* is considered among the finest luxury woods in the world [4] and is in high demand for musical instruments, toys, furniture, and handicrafts [5]. The wood is also considered as antipyretic, astringent, anthelmintic, and diaphoretic in indigenous medicine [6].

Illegal trading and harvesting of *Pterocarpus santalinus* appears to be widespread due to its rich hue and high price. It is often substituted and mislabeled with other woods of lower economic value, like *Pterocarpus soyauxii*, *Pterocarpus tinctorius*, *Dalbergia louvelii*, and *Pterocarpus erinaceus*, in the timber market [7]. Numerous seizures of *Pterocarpus santalinus* have been reported in China and other countries [7]. The identification of



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**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). *Pterocarpus santalinus* wood is difficult because it shares morphological and anatomical similarities with other *Pterocarpus* species and some *Dalbergia* species [8]. Moreover, the Convention on International Trade in Endangered Species (CITES) has limited resources to assist in the identification of *Pterocarpus* wood [9].

To maintain the order of the timber market and the interests of consumers, it is necessary to establish a fast and reliable method for *Pterocarpus* species identification. Traditional wood identification methods were mainly carried out from the perspective of physics and anatomy. Despite the effectiveness of these methods, they have intrinsic disadvantages of high cost, lack of speed, and generally are only accurate to the genus level [10,11]. By contrast, optical and chemotaxonomical methods had proven useful in wood identification [12] and extremely similar wood species can be distinguished by analyzing their spectral characteristics.

Visible light/near-infrared (Vis/NIR) hyperspectral imaging (his) is a simple, green, effective, eco-friendly, and qualitative spectroscopic analytical technique [13]. NIR responds to the energy changing when the non-resonant molecular vibration changes from the ground state to the excited state, mainly reflecting the overtone and combination of hydrogen groups X–H [14]. Vis, as a very narrow part of the electromagnetic spectrum near the NIR, has a similar spectral response to NIR light [15,16]. Spectroscopic analysis combing Vis and NIR is an efficient and fast modern technology for quantitative and qualitative detections. Coupled with suitable modeling methods, Vis/NIR has been successfully applied in many fields, such as the petrochemical [17,18], agricultural [19,20], food [21,22], and pharmaceutical [23] industries. In recent years, Vis/NIR has also shown great potential in forestry applications [24–26]. Compared with Vis/NIR, Vis/NIR-HSI provides simultaneous determination of the physical and chemical properties of the sample as well as their spatial distribution, which overcomes some limitations of Vis/NIR spectroscopy. Therefore, Vis/NIR-HSI is more suitable for the analysis of heterogeneous samples and allows more reliable qualitative identification using both spatial and spectral information. Although this technique requires strong professional judgment, it has greater potential when combined with machine learning algorithms [27]

In this study, wood samples of *Pterocarpus santalinus* (expensive) and nine other highly similar species (less expensive) were analyzed by the Vis/NIR-HSI technique with standard normal variate (SNV), Savitzky–Golay (SG) smoothing, normalization, and multiple scattering correction (MSC) preprocessing methods. Different classification models have been compared and ranked, with the aim of establishing a reliable approach for the identification of these woods. Furthermore, there are few studies comparing and discerning which wavelength ranges are suitable for wood identification [28,29]. Using inappropriate wavelength ranges not only reduces the accuracy of the identification but also increases the computing costs. This study compared the reliability of spectral data in different ranges (Vis range, NIR range, entire range) in order to select the optimal range for the identification of *Pterocarpus santalinus*.

#### 2. Materials and Methods

## 2.1. Samples

The scientific classification and number of wood samples are presented in Table 1. Wood samples were obtained from China National Forestry and Grassland Administration Wildlife Criminal Evidence Identification Center (Nanjing Forest Police College) including eight *Pterocarpus* species and two *Dalbergia* species. We included only species that had at least 20 individuals, with 4~8 samples taken from each individual. A total of 800 wood samples were obtained. In order to develop models, 600 samples were randomly selected as the calibration set for model calibration and 200 samples were selected as the validation set for validation.

Air-dried samples (moisture content of 11.5%–12%) were polished using sandpaper to reduce the roughness of surface and cut to 20 cubic mm (sap wood). According to our

previous research, a transverse section of wood is more suitable for wood identification [27]. In this study, spectral data were scanned in a transverse section of the samples.

Table 1. The plant materials used in the study.

Scientific Classification	Calibration Set	Validation Set		
Pterocarpus soyauxii	60	20		
Pterocarpus tinctorius var. chrysothris	60	20		
Pterocarpus santalinus	60	20		
Pterocarpus erinaceus	60	20		
Pterocarpus indicus	60	20		
Pterocarpus macrocarpus	60	20		
Dalbergia louvelii	60	20		
Dalbergia melanoxylon	60	20		
Pterocarpus tinctorius	60	20		
Pterocarpus angolensis	60	20		

## 2.2. Equipment and Spectra Acquisition

Vis/NIR spectra of wood samples were collected in a darkroom using a Vis/NIR spectrophotometer (ImSpectorV 10E, Specim, Oulu, Finland). The light source was provided by a 350 W halogen lamp (Illumination Technologies, Liverpool, NY, USA) at a 45° angle. The distance between the light source and the surface of the wood sample was 350 mm. The spectroradiometer was located approximately 170 mm from the surface of the wood to be analyzed. The detection wavelength range of the spectrometer was 400~2500 nm and the sampling range of the Vis/NIR spectrophotometer was 6.2 nm. The light source was turned on 15 min earlier to allow the halogen lamps to warm up prior to analysis. Before spectrum acquisition, the instrument was calibrated with a black (covered camera lens) and white image (99.9% reflectance Teflon white plate). Wood samples were placed on a black cloth. For each sample, 80 spectra were scanned and averaged (Figures 1 and 2).



Figure 1. Vis-HSI of wood samples. (a) Pterocarpus soyauxii, (b) Pterocarpus tinctorius var. chrysothris,
(c) Pterocarpus santalinus, (d) Pterocarpus erinaceus, (e) Pterocarpus indicus, (f) Pterocarpus macrocarpus,
(g) Dalbergia louvelii, (h) Dalbergia melanoxylon, (i) Pterocarpus tinctorius, (j) Pterocarpus angolensis.



Figure 2. NIR-HSI of wood samples. (a) Pterocarpus soyauxii, (b) Pterocarpus tinctorius var. chrysothris,
(c) Pterocarpus santalinus, (d) Pterocarpus erinaceus, (e) Pterocarpus indicus, (f) Pterocarpus macrocarpus,
(g) Dalbergia louvelii, (h) Dalbergia melanoxylon, (i) Pterocarpus tinctorius, (j) Pterocarpus angolensis.

## 2.3. Model Development

Wood identification is needed in qualitative analysis. Principal component analysis (PCA), partial least squares discriminant analysis (PLS-DA), and square support vector machine (SVM) modeling methods were performed separately for the Vis range (400 nm~800 nm), the NIR range (800 nm~2500 nm), and the entire spectral range with PLS-toolbox 802 (Eigenvector Research, Inc., Manson, WA, USA). In addition, SNV, SG smoothing, normalization, and MSC preprocessing methods were employed before model development.

The value of accuracy rate, sensitivity, and specificity were used to determine the performance of models. The accuracy rate is the ratio of true positives to the total number of samples. Sensitivity allows for the assessment of how well the model can identify samples that belong to a particular class, and specificity measures the capacity of the model to reject nonbelonging samples. In this study, three statistical parameters were considered to evaluate the models. The flowchart for the identification of wood samples using Vis/NIR-HSI technique is shown in Figure 3.



**Figure 3.** Flowchart of eight *Pterocarpus* species and two *Dalbergia* species' identification using Vis/NIR-HSI method.

## 3. Results

#### 3.1. Spectroscopic Characterization

Vis/NIR are highly suitable for assessment of heterogeneous organic matter, including wood and wood products. The Vis/NIR spectrum contains information regarding both the physical state and chemical composition of measured wood samples. The spectral peak position and its shape corresponds to the presence of specific functional groups possessing dipole momentum [30].

Figure 4 shows that the eight *Pterocarpus* species and two *Dalbergia* species had different absorbance patterns, with prominent absorption peaks at 780, 980, 1240, 1660, 1930, 2080, 2230, and 2350 nm. The variation in the Vis range would be related to ASTA values (extractable color) [31]. The peak around 980 nm may associated with the third stretching overtone of the C-H bonds [32]. The peaks at 1240 nm may correspond to cellulose [33], while the peak at 1660 nm corresponds to a C-H stretching vibration in the first and second overtone. The peak at approximately 1930 nm corresponds to an O-H stretching vibration or O-H-O combination of deformation, and might be due to water peaks [34]. The peaks at 2080 and 2230 nm correspond to an N-H stretching vibration and are mainly related

to lipids, carbohydrates, or protein macromolecular organic matter [35]. The peak at around 2350 nm most likely originates from fat [36,37]. The above characteristic peaks are associated with the chemical compound content in the wood samples. Overall, the NIR range provided more useful information than Vis range, which may be more suitable for wood identification.



Figure 4. (a) Average Vis spectra of wood samples. (b) Average NIR spectra of wood samples.

## 3.2. Principal Component Analysis

A PCA model was applied to preprocessed data to investigate the grouping possibility in the data set. The best results of the PCA model were achieved by a normalization algorithm, and therefore the 3D scatterplots of ten tree species were created based on normalization preprocessed spectra. PCA were performed on three spectral ranges (400~800 nm, 800~2500 nm, 400~2500 nm) including all wood samples. For three spectral ranges, the first three principal components (PCs) can describe the most variance. In Figure 5a, PC1 explains 79.2% of total variance, and PC2 explains 14.7% of variance. Figure 5b represents 95.5% of total variance (PC1 = 79.1%, PC2 = 9.2%, PC = 7.2%). When including the entire spectral range, the first three main components explain 90.3% of data variance (PC1 = 66.7%, PC2 = 14.4%, PC = 9.2%).



**Figure 5.** Principal Component Analysis 3D scatterplots of spectral data from distinct wood samples. (a) Spectral data range from 400 nm to 800 nm, (b) spectral data range from 800 nm to 2500 nm, (c) spectral data range from 400 nm to 2500 nm.

Figure 5 illustrates the ten tree species that were clustered into ten groups. However, there was a moderate overlap between wood samples of *Pterocarpus soyauxii* and *Pterocarpus tinctorius var. chrysothris* and a small overlap between *Pterocarpus soyauxii* and *Pterocarpus tinctorius*. This indicates that *Pterocarpus soyauxii* and *Pterocarpus tinctorius var. chrysothris* were similar to each other in wood spectra. Moreover, the overlap of the groups is stronger when using only the Vis range, compared to using the NIR and entire spectral range. PCA scatterplots of spectral data from 800 nm to 2500 nm show the best clustering compared to the other spectral data ranges. The Vis range was previously reported to be related to pigment [38]. Some of the wood samples in this study had a similar pigment composition [39], which may lead to close Vis spectral characteristics. From the outcomes of PCA analysis, it can be concluded that the results of PCA model are dependent on the range of spectral data.

#### 3.3. Results Using PLS-DA

In this study, PLS-DA and SVM were used for spectral data classification. Both chemometric methods belong to the group of supervised techniques and need complete information regarding the membership of each wood sample to a certain category. These algorithms are capable of classifying an unknown sample into one of the pre-defined classes on the basis of its spectral pattern [40].

Tables S1 and S2, and Figure 6 summarize the accuracy, sensitivity, and specificity for the PLS-DA model and compare different preprocessing algorithms. The PLS-DA model, as a commonly used classification model, has the advantage of less computational cost; it can find the best functional match for a set of data by minimizing the sum of squares of the errors [41]. PLS-DA has been widely used in the analysis of multivariate data. However, the calculation results show that the PLS-DA model performed poorly in the identification of eight *Pterocarpus* species and two *Dalbergia* species. The best results were achieved for the NIR range in both calibration and validation sets. The entire spectral range gave slightly lower accuracy than the NIR range.



**Figure 6.** Mixing matrix between true class and predicted class with PLS-DA model (without preprocessing). (**a**) Spectral data range from 400 nm to 800 nm, (**b**) spectral data range from 800nm to 2500 nm, (**c**) spectral data range from 400 nm to 2500 nm.

In this study, for the sake of suppressing the bad influence of noise, SNV, SG smoothing, normalization, and MSC preprocessing were used to analyze the Vis/NIR spectra data. The results for the preprocessed spectra data were improved compared to the raw spectra data. It is apparent from Table 2 that PLS-DA combined with SG smoothing preprocessing methods can achieve the highest prediction accuracy, while SNV and MSC treatment performed relatively poorly. Additionally, there were some differences in the performance of the preprocessing methods within each spectral range. In this study, the best PLS-DA model was obtained using NIR spectral data with SG smoothing methods with 96.8% and 96.5% accuracy for calibration and validation sets, respectively. Furthermore, it can be seen from Figure 6 that *Pterocarpus santalinus* can be easily confused with *Pterocarpus erinaceus* when using Vis range, which may be related to the highly similar ASTA values of the two species.

	400~800 nm		800~2500 nm		400~2500 nm	
	Calibration Set (%)	Validation Set (%)	Calibration Set (%)	Validation Set (%)	Calibration Set (%)	Validation Set (%)
Preprocessing Raw	88	84.5	96.7	96.5	90.3	94
SNV	79.2	76	92.8	92	86	88
SG Smoothing	88.8	85.5	96.8	96.5	90.3	94
Normalization	88.8	85	96.5	96	90.3	94
MSC	88.8	85.5	92.7	92	85.8	88.5

 Table 2. The accuracy of PLS-DA model with different preprocessing method.

Bold values indicate the best results.

#### 3.4. Results Using SVM

SVM is a supervised machine learning algorithm useful for solving both regression and classification problems. It is a nonlinear classification method that constructs a set of hyperplanes in a high or infinite dimensional space, and a good separation is achieved by the hyperplane that has the largest distance to the nearest training data point of any class [42]. Compared with the PLS-DA model, SVM is not influenced by the distribution of diverse sample classes.

In contrast to the results of PLS-DA model, the performance of the SVM was better. The classification accuracy was lower for the spectral range from 400 nm to 800 nm as compared with the other ranges, but still provided acceptable results. As shown in Table 2, the best choice with SVM model would be with the entire spectral range, but the difference was not significant compared to the NIR range. In the NIR and entire spectral range, the sensitivity and specificity rates present similar values, which means that the error was balanced.

The classification accuracy of the SVM model for various preprocessing methods is illustrated in Table 3. The best results for NIR and entire spectral range were obtained with the normalization method, with 100% (validation set). In spite of weaker classification results for PLS-DA in the Vis range, the SVM model combined with normalization preprocessing was able to identify eight *Pterocarpus* species and two *Dalbergia* species with an accuracy of 96.5% (calibration set) and 95% (validation set). It needs to be noted that the use of the SVM model with Vis ranges does not improve the accuracy of identifying *Pterocarpus santalinus* (Figure 7).

	400~800 nm		800~2500 nm		400~2500 nm	
	Calibration Set (%)	Validation Set (%)	Calibration Set (%)	Validation Set (%)	Calibration Set (%)	Validation Set (%)
Preprocessing Raw	96.3	94.5	99.7	99.5	99.8	99.5
SNV	93.2	92.5	97.3	99.5	95.8	98
SG Smoothing	96.5	95	99.7	99.5	99.8	99.5
Normalization MSC	<b>96.5</b> 92.2	<b>95</b> 91.5	<b>99.7</b> 97.3	<b>100</b> 99.5	<b>99.8</b> 96.7	<b>100</b> 98.5

Bold values indicate the best results.

Table 3. The accuracy of SVM model with different preprocessing method.



**Figure 7.** Mixing matrix between true class and predicted class with SVM model (without preprocessing). (a) Spectral data range from 400 nm to 800 nm, (b) spectral data range from 800 nm to 2500 nm, (c) spectral data range from 400 nm to 2500 nm.

## 4. Discussion

*Pterocarpus santalinus* has a higher economic value than *Pterocarpus soyauxii*, *Pterocarpus tinctorius var. chrysothris*, and *Pterocarpus erinaceus*, et al. The identification of *Pterocarpus santalinus* and its counterfeits requires specialist skills. In this study, we have used Vis/NIR-HSI techniques to identify *Pterocarpus santalinus* and its nine common counterfeit woods. It can be seen from the raw Vis-NIR spectra (Figure 4) that the wavelengths with the highest variation for *Pterocarpus santalinus* were mainly at 780, 980, 1240, 1660, 1930, 2080, 2230, and 2350 nm, which were different from its counterfeits. Furthermore, the original average spectra of all samples in the wavelength ranges of 400~955 nm and 800~2500 nm showed different absorbance peaks related to the vibration energy of the molecular bonds of O–H, C–H, C–O, and N–H. This demonstrates the potential of using the Vis/NIR-HSI technique for the identification of wood samples.

In recent years, many studies have focused on the identification of wood using Vis/NIR techniques, [43] but the optimal wavelength range selection has rarely been discussed. Choosing an unsuitable wavelength range not only reduces the accuracy of identification, but it also increases the cost of the equipment. In this study, we developed a classification model with spectral data in different ranges (Vis range, NIR range, entire range). When compared to the results of the NIR spectra, the predictive accuracy of the

classification model developed with full spectra was not enhanced. The reason behind this may be that the Vis range cannot provide enough spectral information, and noise exists, leading to the low quality of spectral data. This also demonstrated that few wavelengths are needed for the identification of *Pterocarpus* and *Dalbergia* species. Wavelengths attributed to wood compounds, such as cellulose and lignin, may have more effects on the prediction accuracy [28,44,45].

The wood samples used in this study were natural and had not been treated with formaldehyde, thermally modified, etc. The treatment of wood can significantly affect its spectral characteristics [46,47]. As treated wood and wood products are very common in the market, further research is needed in this important area. Furthermore, the wood samples selected for this study were all sap wood. Different parts of the rosewood may secrete different substances [48], which may affect the applicability of the models. To build more accurate classification models, wood samples from different parts are needed in future studies.

#### 5. Conclusions

In this study, Vis/NIR-HSI techniques have been proven to classify eight *Pterocarpus* species and two Dalbergia species. The variability of wood samples and the random selection of samples for calibration and validation suggest the robustness of the models. PCA was used to analyze the spectral data of the wood samples. PCA scatterplots of NIR range show the best clustering compared to the other spectral ranges. Moreover, it was found that the differences between Pterocarpus soyauxii and Pterocarpus tinctorius var. chrysothris were not obvious due to similar spectral characteristics. The PLS-DA and SVM models were established based on different spectral ranges of the raw data and preprocessed data. The Vis range offered acceptable classification, whereas the NIR range showed significantly higher identification accuracy based on differences in the absorbance of lipids, carbohydrates, or protein macromolecular organic matter. It was found that the SVM model performed better than the PLS-DA method, as shown by its discrimination accuracy. Satisfactory identification accuracy was obtained, which was 99.5% for both NIR and entire spectral range (validation set). It can be highlighted that the normalization preprocessing method combined with SVM model can achieve the highest accuracy in the whole spectral range (100%, validation set).

This study demonstrated that the Vis/NIR-HSI techniques combined with SVM have the ability to identify wood from the genera *Pterocarpus* and *Dalbergia*. This method is rapid, easy, and non-destructive, which could reduce the enormous workload of wood experts and increase the accuracy of testing.

**Supplementary Materials:** The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/f14061259/s1, Table S1: Results of calibration and validation sets using PLS-DA. Table S2: Results of calibration and validation sets using SVM.

**Author Contributions:** Conceptualization, X.X.; writing—original draft preparation, Z.C.; data curation, H.W.; supervision, H.G.; software, J.N. and X.L. All authors have read and agreed to the published version of the manuscript.

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**Data Availability Statement:** The datasets generated and analyzed during the current study are available in the Figshare repository, [https://doi.org/10.6084/m9.figshare.22284712.v1 (accessed on 16 March 2023)].

Conflicts of Interest: The authors declare no conflict of interest.

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