

Supplementary Materials:

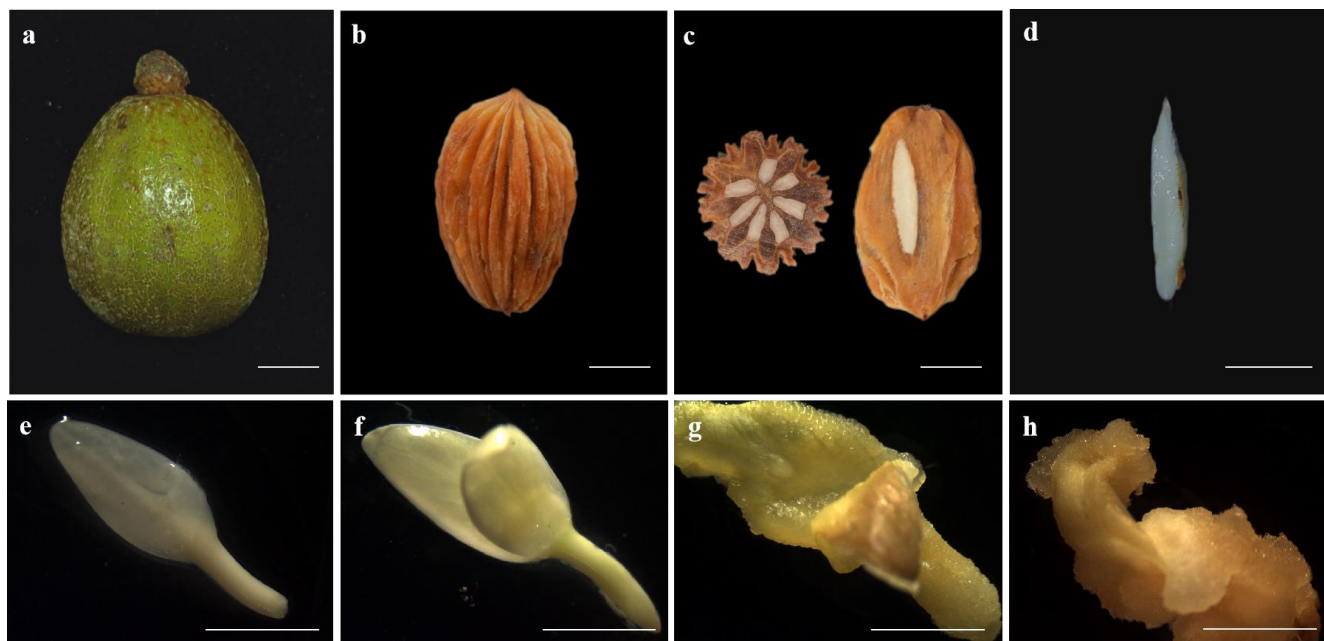


Figure S1: Seed structure and callus induction of *D. involucrata*: (a) Immature *D. involucrata* seeds. (b) Lignified seeds without exocarp and mesocarp. (c) Cross and vertical cutting of seeds. (d) Immature zygotic embryo surrounded by endosperm. (e~h) immature zygotic embryo 0 d, 5 d, 10 d, 15 d. Bar: 1 cm;

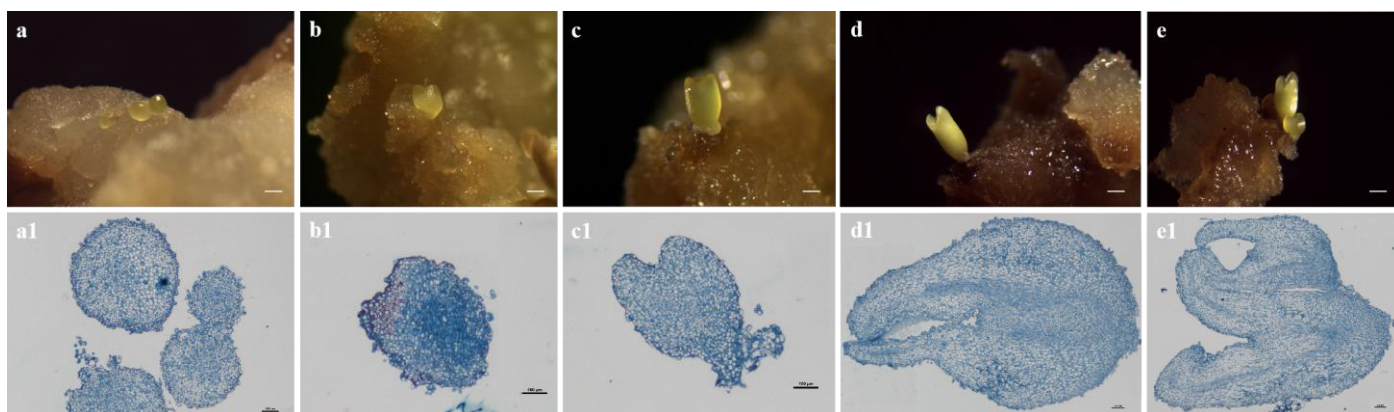


Figure S2: Different developmental stages and histology of somatic embryos from *D. involucrata*. (a,a1) Globular stage somatic embryo. (b,b1) Heart-shaped stage somatic embryo. (c,c1) Torpedo-shaped stage somatic embryo. (d,d1) Cotyledonary stage somatic embryo. (e,e1) Mature somatic embryo. (a~e) Bar: 1 mm. (a1~e1) Bar: 100 μ m;

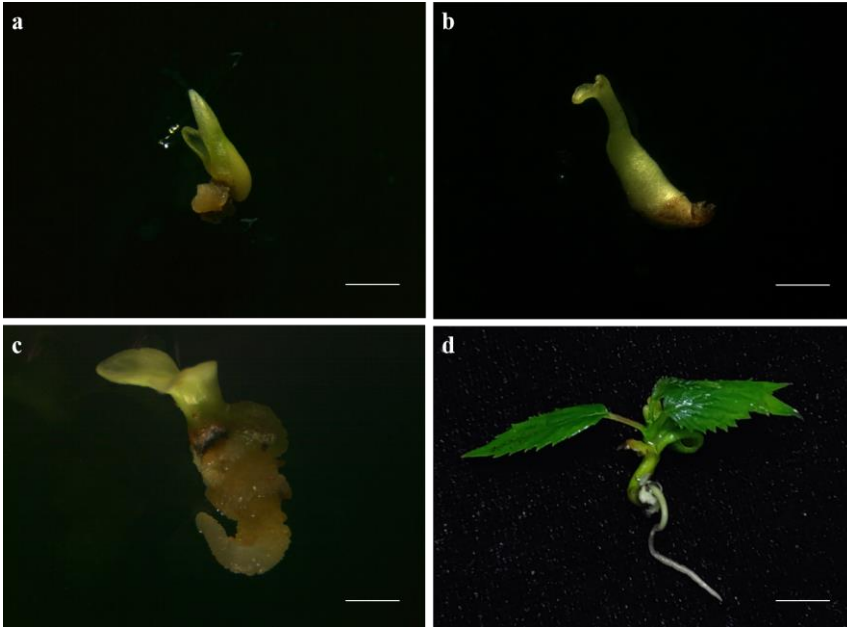


Figure S3: Somatic embryo germination and conversion of *D. involucrata*. (a) Anomalous germinated somatic embryo with cotyledons only. (b) Anomalous germinated somatic embryo with root only. (c) Normally germinated somatic embryo with both shoot and root. (d) Conversion plantlets. (a~c) Bar: 1 mm. (d) Bar: 1cm;

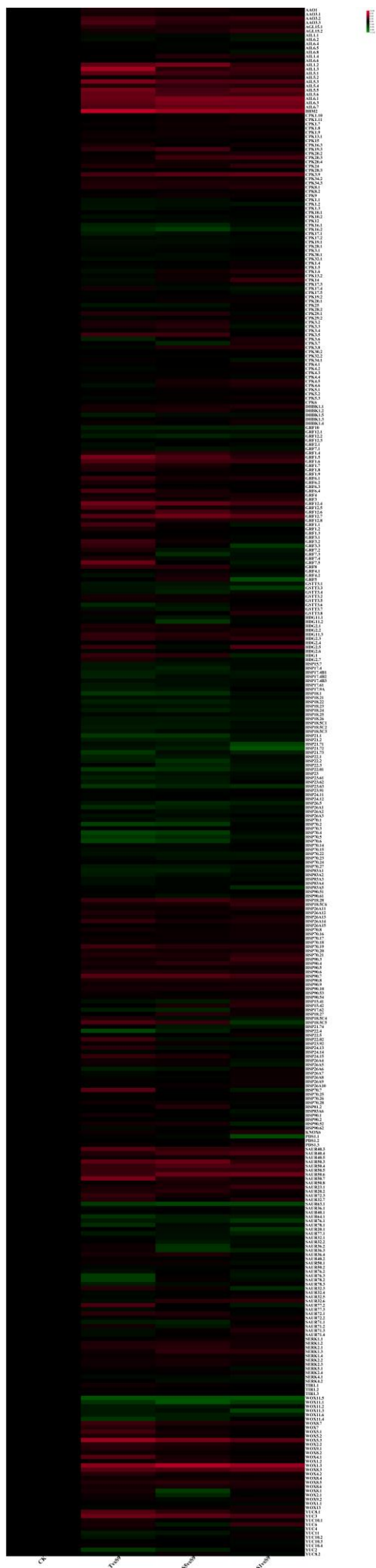


Figure S4: Genes associated with somatic embryogenesis from EC of *D. involucrata*;

Table S1: qRT-PCR primer design;

Gene	Forward primers	Reverse primers
<i>DiPRU1</i>	CAGGATTAGCCAAGAGGT	GGCGAAGGCAGCCATTAC
<i>DiPHI1</i>	TCAACGACATTCTCCACC	CACCCACTCCATTTTACA
<i>DiHIP21</i>	GTGTCTTGCCAAACTCCA	ACCTTCATTATCCCTACCA
<i>DiASPA1</i>	TCAGTGGCATCACCGTTT	ACCCGCAAGGTCCATCAT
<i>DiGOLS1</i>	GAGCCCTAAAGCACTCCACA	GGCGAATCCCACCCTACCT
<i>DiIAA9</i>	TGGTTGTCAGGTGGATAG	TTTGCGTAAAGTGAAGAGT
<i>DiKWL1</i>	CTTGAGGGGCATAAGGATAA	ATGGACTGTGATGGCTGTG
<i>DiASPA2</i>	CAGGCGGACTGCTATTGT	AAACTTGGGAGTGAAACC
<i>DiEP1</i>	TTTGTTGGCAGTGTTGGT	CCGTTTAGGCTTTGTTTC
<i>DiFB60</i>	GGGTTTGCTCACAGGGAT	GCCGTGGAAGTTGAAGGA
<i>DiPGKH</i>	CTTGCCACTAAGGTCTGC	GGATGGTTGGATGGGATT
<i>DiRG1</i>	GGTCACGAAGGGCTCTAT	GAAACTCGGTCTAAATGC
<i>DiCSE</i>	ACCTCCCCAAACACCAAATC	CCGCCAGCAAGGACAAGA
<i>DiKALSE</i>	GAGTCGTCGCTGGTGAGA	CGTGTCGGGTGTTTGTGA
<i>DiACX3</i>	TGAGGGTAGAACTTGTGAGC	ATGCCGTGAAGCCTGTGG
<i>DiCAN2</i>	TAAACAAGGACTCCCAAACA	AGAGGCAGACGCAATACA

Table S2: Reverse transcription reaction system;

Procedure	Reagents name	Volume
Removal of Genomic DNA Reaction	5 × gDNA Emaser Buffer	2 µl
	gDNA Emaser	1 µl
	Total RNA	Calculated according to the concentration, not to exceed 1 µg
	RNase Free ddH ₂ O	Up to 10 µl
Reverse transcription reaction	The product obtained in the previous step	10 µl
	RNase Free ddH ₂ O	4 µl
	5 × PrimeScript Buffer 2	4 µl
	RT Primer Mix	1 µl
	PrimeScript RT Enzyme Mix I	1 µl

Table S3: Reverse transcriptional response program;

Procedure	Temperature	Time
Removal of Genomic DNA Reaction	42 °C	2 min
Reverse transcription reaction	42 °C	15 min
	85 °C	5 s

Table S4: qRT-PCR reaction system;

Reagents Name	Volume
2 × Q3 SYBR qRT-PCR Master Mix	10 µl
RNase Free ddH ₂ O	8 µl
Primer F	0.5 µl
Primer R	0.5 µl
cDNA	1 µl

Table S5: qRT-PCR procedure.

Procedure	Reaction temperature	Reaction time	Number of cycles
pre-denatured	95 °C	60 s	1
	95 °C	10 s	
cyclic reaction	57 °C	10 s	40
	72 °C	30 s	
	95 °C	15 s	
melting curve	65 °C	60 s	1
	95 °C	15 s	
cool down	37 °C	30 s	1