

# Accelerated Endothelialization of Nanofibrous Scaffolds for Biomimetic Cardiovascular Implants

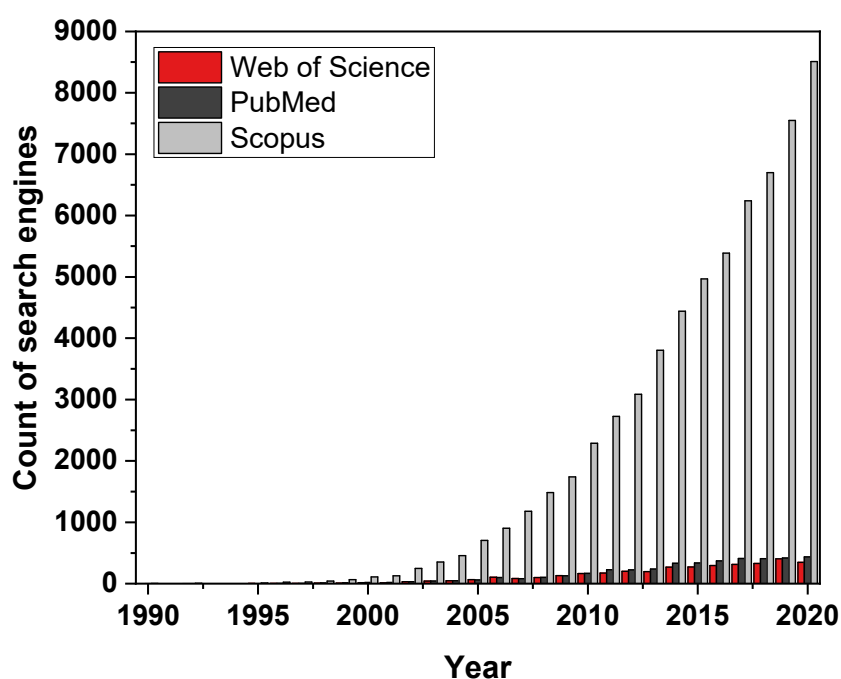
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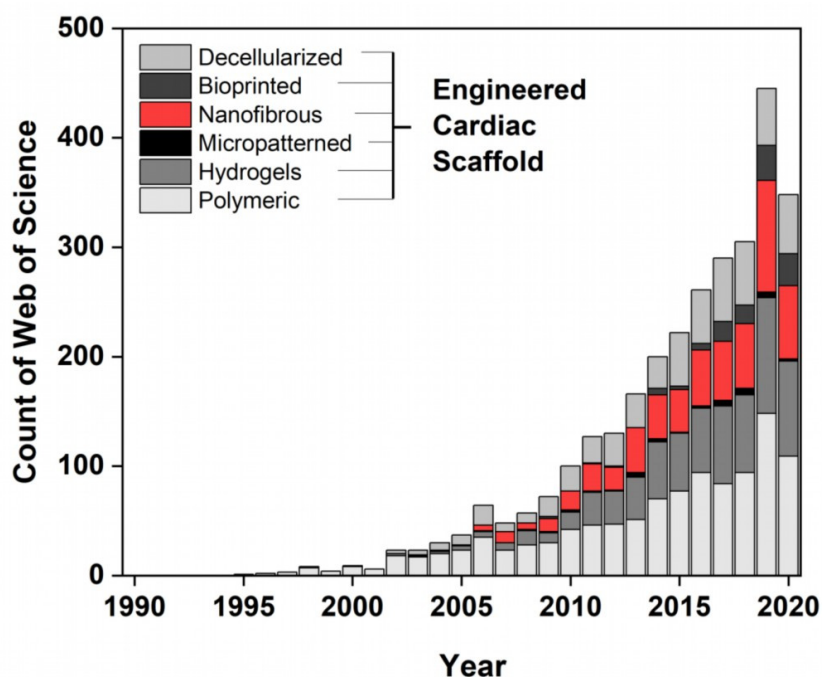
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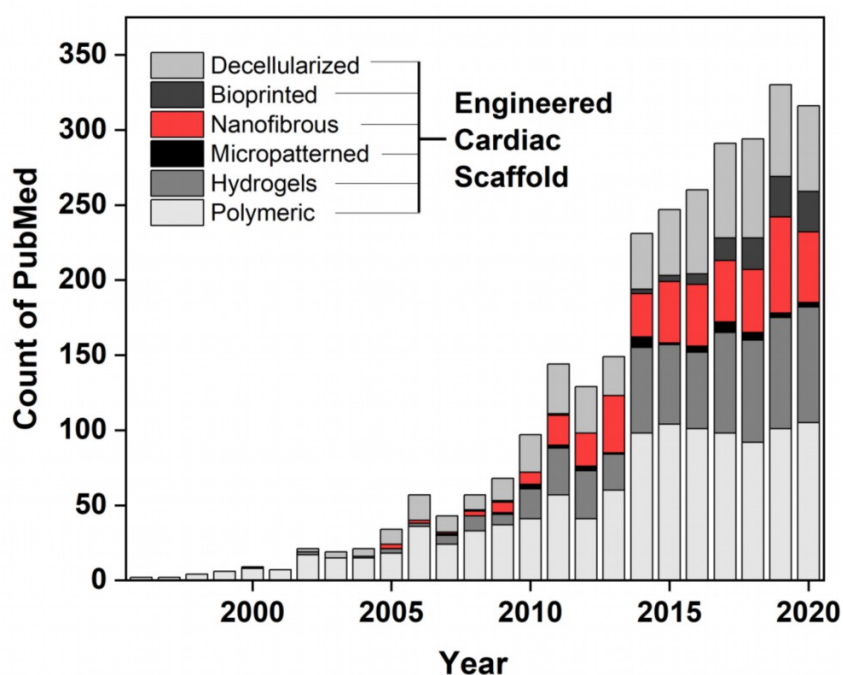
## Section 1. Introduction



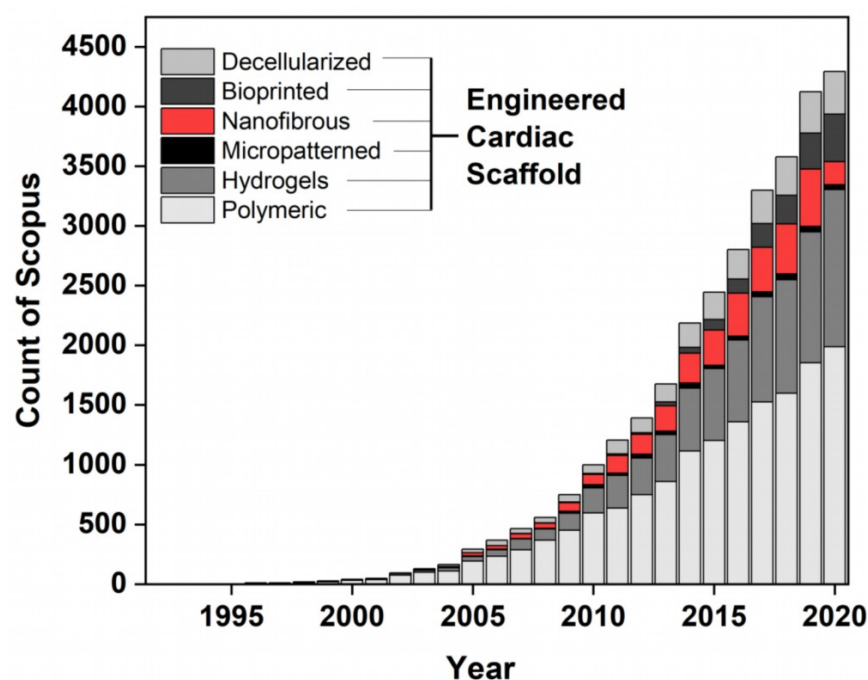
**Figure S1:** The annual number of publications for the last 20 years with topic “engineer\* cardi\* scaffold\*” provided by the search engine of Scopus, PubMed and Web of Science before the 15<sup>th</sup> February, 2022.



**Figure S2:** The annual number of publications for the last 20 years with topic “engineer\* cardi\* scaffold\*” associated with supplementary search terms: “polymer\*”, “hydrogel\*”, “micropattern\*”, “nanofib\*”, “bioprint\*” or “decellular\*”, provided by the search engine of Web of Science before 15<sup>th</sup> February 2022.

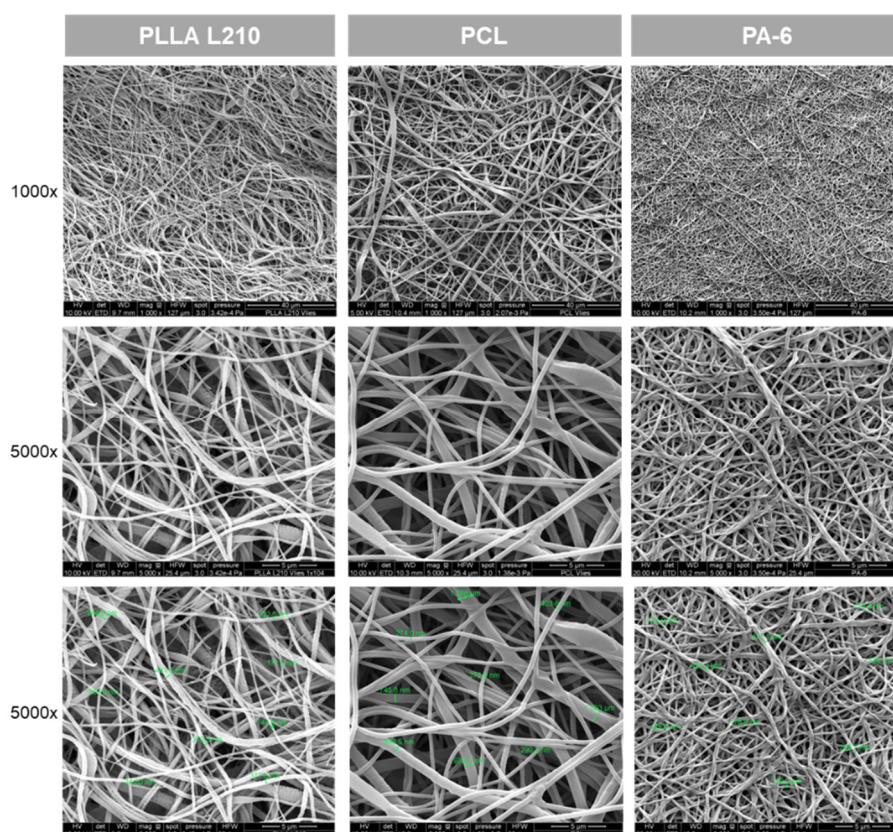


**Figure S3:** The annual number of publications for the last 20 years with topic “engineer\* cardi\* scaffold\*” associated with supplementary search terms: “polymer\*”, “hydrogel\*”, “micropattern\*”, “nanofib\*”, “bioprint\*” or “decellular\*”, provided by the search engine of PubMed before before 15th February 2022.

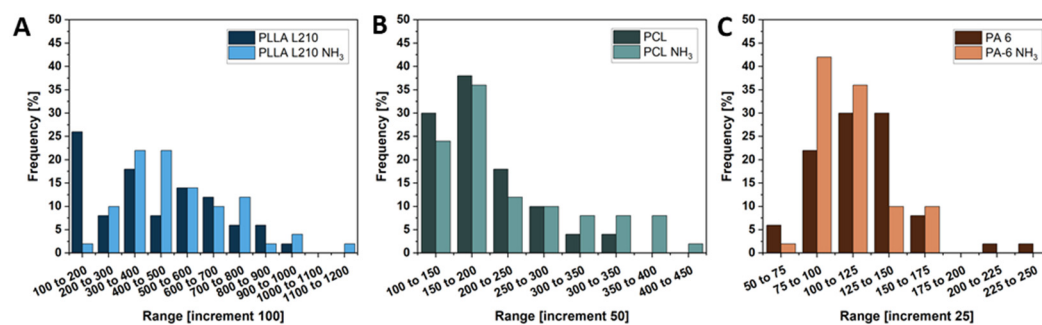


**Figure S4:** The annual number of publications for the last 20 years with topic “engineer\* cardi\* scaffold\*” associated with supplementary search terms: “polymer\*”, “hydrogel\*”, “micropattern\*”, “nanofib\*”, “bioprint\*” or “decellular\*”, provided by the search engine of Scopus before 15th February 2022.

### Section 3.1. Morphology of polymeric nonwovens

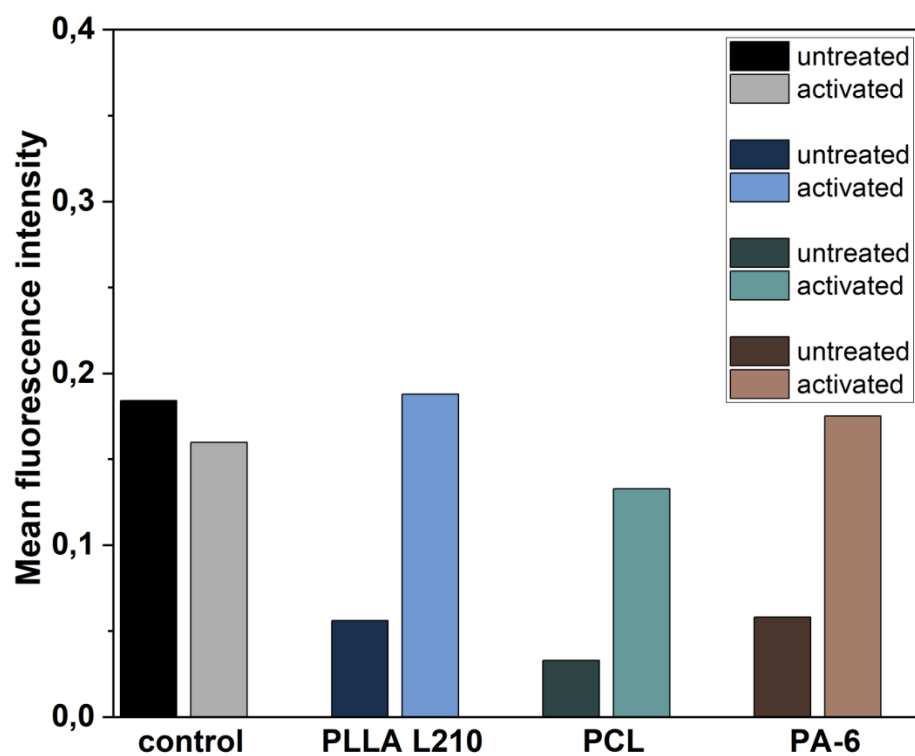


**Figure S5:** Representative SEM images of PLLA L210, PCL and PA-6 nonwovens at magnification 1000x and 5000x.



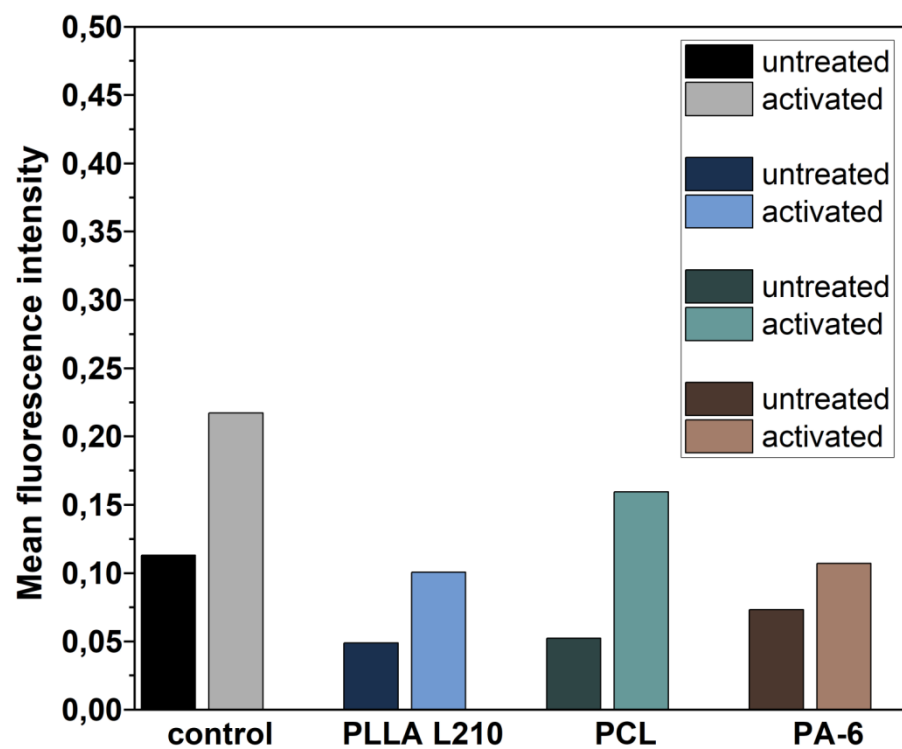
**Figure S6:** Frequency of the respective fiber diameters within the individually defined increments for untreated and activated (A) PLLA L210, (B) PCL and (C) PA-6 nonwovens based on 50 individual measurement points (where for each polymer 10 fibers were measured manually in 5 SEM images).

### Section 3.6. Endothelial actin cytoskeleton formation on polymeric nonwovens



**Figure S7.** Fluorescence intensity of F-actin formation (phalloidin-TRITC for F-actin) in human endothelial EA.hy926 cells grown for 48 h on untreated and NH<sub>3</sub>-plasma functionalized polymeric nonwovens and polystyrene control surface (n = 1).

### Section 3.7. Expression of endothelial cell-specific PECAM-1 marker



**Figure S8.** Mean fluorescence intensity of CD 31 expression by human endothelial EA.hy926 cells grown for 48 h on untreated and NH<sub>3</sub>-plasma functionalized polymeric nonwovens (n = 1).