



Article Willow, Poplar, and Black Locust Debarked Wood as Feedstock for Energy and Other Purposes

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Abstract: Solid biomass can be used for energy generation and the production of various renewable bioproducts. The aim of this study was to determine the yield and characteristics of wood obtained as debarking residue from 14 genotypes of short-rotation woody crops (SRWCs). These included five Populus genotypes, one Robinia genotype, and eight Salix genotypes, harvested in both annual and quadrennial cycles. The results showed that the highest dry wood yield (12.42 Mg ha⁻¹ y⁻¹ DM) and yield energy value (244.34 GJ ha⁻¹ y⁻¹) were obtained from willow (cultivar Zubr) harvested in a quadrennial cycle. The best effect among the poplar genotypes was achieved for the Hybryda275, and it was particularly marked in the quadrennial harvest cycle. The poorest results were determined for black locust. The Robinia characteristics included the significantly lowest moisture content (31.6%), which was a positive attribute from the energy point of view, but, on the other hand, it had some adverse characteristics-the highest levels of sulfur (0.033% DM), nitrogen (0.38% DM), and ash (0.69% DM). More beneficial properties in this respect were determined for willow and poplar wood. Moreover, willow and poplar wood contained more cellulose—51.8 and 50.0% DM, respectively compared with black locust. Extending the SRWC shoot harvest cycle from annual to quadrennial resulted in an increase in cellulose, lignin, and carbon, higher heating value, and a decrease in nitrogen, sulfur, ash, and moisture content. Therefore, extending the harvest cycle improved the parameters of SRWC wood as an energy feedstock.

Keywords: *Populus; Robinia; Salix;* short-rotation coppice; wood; solid biofuel; higher heating value; ash; sulfur; nitrogen; chlorine; cellulose; lignin

1. Introduction

Solid biomass can be used for energy generation and the production of various renewable bioproducts. Plant biomass is primarily derived from agriculture, forestry, and the processing of materials derived from these industries [1]. Agricultural land can be used to grow perennial industrial crops (PICs), which are a type of lignocellulosic biomass source [2–6]. PICs include a group of short-rotation woody crops (SRWCs) of genera *Eucalyptus, Salix, Populus,* and *Robinia,* which are sources of woody biomass [7,8]. The production and use of SRWCs are seen as bringing environmental, social, and economic benefits [9–12]. According to the biobased economy concept, biomass use should be diverse, and it should include generating various renewable food bioproducts and others, as well as energy generation [13–17].



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Therefore, it is proposed that bark should be separated from wood so that these two materials of different quality can be used in different ways. This approach can result in the valuation and cascaded use of these two materials (wood and bark) obtained from SRWC shoots. Valuable secondary metabolites containing phenolic compounds are obtained from the bark of willow, poplar, and black locust [18–22]. Phenolic glucosides, also known as salicylic glucosides, and salicylic acid (found in willow bark) possess anti-inflammatory, antipyretic, antirheumatic, analgesic, and anticoagulant properties [20,23,24]. Therefore, bark produced as a raw material for manufacturing high-added-value bioproducts will be accompanied by generating wood as SRWC shoot debarking residue. Owing to its properties, such wood can be a valuable material for use in biorefineries and as energy feedstock [16]. However, due to the high diversity among SRWC species, only those that are the most suitable for cultivation from the yield and biomass quality perspective, including the quality of wood as debarking residue, should be selected. It should also be added that, most often, studies in this field cover one type of SRWC, i.e., willow, poplar, or black locust. Therefore, the novelty of this study was the comparison of these three types of SRWC, grown simultaneously under the same environmental (climatic and soil) conditions, at the same time and in two different harvesting cycles. The present study hypothesized that the wood, as a residue from the debarking of SRWC shoots, has interesting and practically important properties. This study seeks to evaluate the quality of wood residue from 14 different genotypes of short-rotation woody crops (SRWCs), both those harvested annually and those harvested every four years, after debarking. The specific objectives included the determination of (i) wood yield and energy value, (ii) thermophysical properties, (iii) elemental composition, and (iv) lignocellulosic composition.

2. Materials and Methods

2.1. Field Experiment

In April 2009, the University of Warmia and Mazury in Olsztyn (UWM) founded a plantation for the purpose of growing short-rotation woody crops (SRWC) in northeast Poland. The field experiment was conducted by the Department of Genetics, Plant Breeding and Bioresource Engineering of the UWM. A two-factorial experiment was conducted, with the first factor being fourteen SRWC genotypes of three different genera, including [25]: one genotype of genus *Robinia: Robinia pseudoacacia* L.; five genotypes of genus *Populus: P. balsamifera* L., UWM2; *P. balsamifera* L., UWM3; *P. nigra* × *P. maximowiczii* (L.) Henry, Max-5; *P. maximowiczii* × *P. trichocarpa* (Henry) Torr. and A. Gray, Androscoggin; *P. maximowiczii* × *P. trichocarpa* (Henry) Torr. and A. Gray, Hybryda275; eight genotypes of genus *Salix: S. alba* L., UWM200; *S. alba* L., UWM035; *S. triandra* L., UWM198; *S. viminalis* × *S. purpurea* L., UWM033; and *S. viminalis* L., cultivar Żubr. Two shoot harvest cycles were studied: annual and quadrennial.

The planting process consisted of 20 cm long willow and poplar cuttings, along with black locust seedlings, which were planted at a density of 18,000 plants per hectare. After every harvest, mineral fertilizers were applied to the crops, which included 90 kg ha⁻¹ of ammonium nitrate for N, 30 kg ha⁻¹ of triple superphosphate for P₂O₅, and 60 kg ha⁻¹ of potassium salt for K₂O. The yield potential and biomass samples for further analysis were obtained by selecting three plots of 45 m² each for each SRWC genotype. The aboveground shoots of all SRWC genotypes were harvested three times in an annual cycle from 2018 to 2020, in late March of each year, and once in the quadrennial cycle. During the harvest period, the shoots of all SRWC genotypes were cut into fragments from all sections along the shoots, packed in PE bags, and transported to the laboratory. Quadrennial leafless shoots (approx. 7 m high, approx. 5 cm in diameter) from each plot were harvested manually with a chainsaw, while annual shoots (approx. 3 m high, approx. 2 cm in diameter) were harvested with a petrol brush cutter. All shoots were electronically weighed immediately

after the harvest. The result was used to calculate the yield of shoots, which was then referred to as a unit area (Mg ha^{-1}).

2.2. Separation of SRWC Shoots into Wood and Bark Fractions and Determination of Wood Yield

The SRWC shoot sections sent to the laboratory were separated into bark and wood using a sharp knife. The bark was used to analyze the concentration of bioactive substances in it and, subsequently, the potential yield of bioactive substances [26]. The wood was analyzed for its potential use in industry and in energy generation. Therefore, the biomass was dried at 105 °C until the sample weight was constant according to PN-EN ISO 18134-2 [27]. The result was used to calculate the wood content of the shoots and their percentage share in the whole dendromass (bark + wood). To ensure accuracy, annual shoots of each genotype were harvested three times, and each year, the analyses were carried out in triplicate. This means that nine analyses were conducted for annual shoots of each genotype. However, for this manuscript, only the mean data from three years for annual shoots are presented. On the other hand, quadrennial shoots of each genotype were harvested only once, and the analyses were carried out in triplicate. The shoot yield (Mg ha⁻¹), percentage share (%) of wood in it, and the dry matter content (%) were used to assess the dry wood yield (Mg ha⁻¹ y⁻¹).

2.3. Laboratory Analyses

After performing the measurements and calculations, the wood samples were ground using a Retsch SM 200 laboratory mill (Retsch, Haan, Germany) and a 1 mm mesh sieve. The ground wood samples were then stored in plastic bags at room temperature. All the analyses were performed at the Energy Feedstock Assessment Laboratory at the Department of Genetics, Plant Breeding and Bioresource Engineering of the UWM. The dynamic method was used to determine the higher heating value (HHV) in an IKA calorimeter C2000 (IKA, Taufen, Germany) according to PN-EN ISO 18125:2017-07 [28]. The weighed portion of approx. 0.5 g was used to determine the HHV. An Eltra Tga Thermostep automatic thermogravimetric analyzer (ELTRA, Neuss, Germany) was used to determine the total ash content, fixed carbon (FC), and volatile matter (VM) content at 550 °C, following the standards PN-EN ISO 18122:2016-01 [29] and PN-EN ISO 18123:2016-01 [30]. The weighed portion of ground wood for an individual analysis was approx. 1.5 g. An Eltra CHS 500 automatic analyzer (ELTRA, Neuss, Germany) was used to determine the total carbon (C), hydrogen (H), and sulfur (S) contents of wood via high-temperature combustion (1350 $^{\circ}$ C), in accordance with PN-EN ISO 16948:2015-07 [31] and PN-EN ISO 16994:2016-10 [32] standards. The weighed portion for the analyses was approx. 0.15 g. The Kjeldahl method with a K-435 analyzer and a BUCHI B-324 (BUCHI, Flawil, Switzerland) distiller were used to determine the total nitrogen (N) content. The total chlorine (Cl) was determined using the Eschka mixture in accordance with the PN-ISO 587:2000 standard [33]. The weighed portion of wood for N and Cl analyses was approx. 1 g each.

The cold water extract (CWE) content was determined by comparing the sample weight before and after extraction. Distilled water (20–25 °C) was used for the sample extraction for 48 h using F57 filtration bags from ANKOM Technology. The samples were washed twice in an Ankom A200 (NY, USA) apparatus and then dried at 105 °C before being weighed. These weighed samples were then used for further analyses. To determine the hot water extract (HWE) content, the samples were extracted for 3 h at 100 °C, dried, and then weighed again at 105 °C. After extracting the hot water, further analyses were carried out to determine the NDF (neutral detergent fiber) according to PN-EN ISO 16472:2007 [34], ADF (acid detergent fiber), and ADL (acid detergent lignin) according to PN-EN ISO 13906:2009 [35] fractions in the wood using an Ankom A200 extraction system. These differences in the results were used to calculate the lignin (Lig), cellulose (Cel), and hemicellulose (Hem) content of the SRWC wood samples. The HWE and NDF contents were used to calculate the content of other soluble substances (OSSs). Each sample was subjected to laboratory analysis three times.

2.4. Energy Value of the Wood

The energy value of wood from a unit area (GJ $ha^{-1} y^{-1}$) was obtained by multiplying the dry wood's HHV (GJ Mg^{-1} DM) by its yield (Mg $ha^{-1} y^{-1}$ DM). Subsequently, the wood yield energy value was expressed as the carbon equivalent (Mg ha^{-1}), assuming that medium-quality hard coal has a calorific value of 25 GJ Mg^{-1} .

2.5. Statistical Analysis

A set of averaged three-year data for annual and one-year harvest data for quadrennial shoots was used in this study. Before any statistical analysis was performed, the normality of the attributes under study was verified using the Shapiro–Wilk test. For statistical analyses, a two-factorial ANOVA was used, with genotype (14 different SRWCs) as the first factor and harvest cycle (annual and quadrennial) as the second factor. Homogeneous groups were determined by applying Tukey's honest significant difference (HSD) test at the significance level of p < 0.05. The study results were presented by calculating the arithmetic mean, the standard error of the mean, and the coefficient of variance for each analyzed attribute of each SRWC genotype. Additionally, the Pearson correlation coefficient between the attributes was also determined. Descriptive statistics were calculated for all SRWC genotypes, including mean, median, minimum and maximum values, lower and upper quartiles, standard deviation, and coefficient of variation. Moreover, two similarity analyses were performed for the 14 SRWC genotypes under study and for the determined wood attributes. In order to analyze the similarities between different genotypes and their wood attributes, a multivariate cluster analysis was conducted. The Ward method was used for agglomeration, while Euclidean distances were utilized to measure the distance between the clusters. Sneath's criterion was used to identify the clusters. Two cut-off lines were applied, the first at $2/3 D_{max}$, and the second at $1/3 D_{max}$, where Dmax is the maximum measure of distance. The results of the cluster analysis were displayed in a dendrogram. All statistical analyses were carried out using STATISTICA 13 software (TIBCO Software Inc., Palo Alto, CA, USA).

3. Results

3.1. SRWC Wood Yield and Its Energy Value

The wood yield and energy value varied significantly depending on the SRWC genotype, as well as the plant harvest cycle and the interaction between these two factors (Table 1). Among the 14 SRWC genotypes studied, the highest wood yield was obtained from the S. viminalis Zubr cultivar, with a yield of 10.65 Mg ha⁻¹ y⁻¹ DM. This was classified as homogeneous group "A" (Figure 1). A second homogeneous group "B" included poplar P. maximowiczii × P. trichocarpa, Hybryda275, and willow S. alba UWM095 as well as, indirectly, another two willow genotypes, S. fragilis UWM195 and S. triandra UWM198. The wood yield for the four genotypes exceeded 8 Mg ha⁻¹ y⁻¹ DM and was lower by 20–29% compared with the wood yield of the willow Żubr cultivar. R. pseudoacacia gave the lowest dry wood yield (3.86 Mg ha⁻¹ y⁻¹ DM), which was lower by 64%. The study found that the plant harvest cycle had a significant impact on the dry wood yield. The quadrennial cycle resulted in a higher yield of dry wood (8.25 Mg ha⁻¹ y⁻¹ DM) compared with the annual cycle, with an increase of almost 37%. The highest dry wood yield in the entire experiment (12.42 Mg ha⁻¹ y⁻¹ DM) was obtained from *S. viminalis* of the Żubr cultivar harvested in the quadrennial cycle (Figure 1, Table A1). The intermediate homogeneous group "ab" included the yield of *P. maximowiczii* × *P. trichocarpa*, Hybryda275 wood, whose yield in the quadrennial cycle was lower by 10%. The lowest dry wood yield was obtained from the *P. balsamifera* UWM3 harvested in the annual cycle, which was lower by as much as 78%. In general, R. pseudoacacia and P. balsamifera UWM3 gave the lowest wood yields, both in the annual and the quadrennial harvest cycles.

Source of Variation	Degrees of Freedom	Dry Wood Yield	МС	VM	Н	S	Cl	For the Rest Features ¹
Genotype	13	< 0.001 *	< 0.001 *	< 0.001 *	0.058	< 0.001 *	< 0.001 *	<0.001 *
Harvest cycle	1	< 0.001 *	< 0.001 *	< 0.001 *	0.007 *	< 0.001 *	0.008 *	< 0.001 *
Genotype \times Harvest cycle	13	< 0.001 *	< 0.001 *	0.001 *	0.056	0.078	0.010 *	< 0.001 *
Error	56							

Table 1. Analysis of variance (*p* values) for the analyzed features.

¹ energy value of wood yield; coal equivalent, higher heating value; ash; FC; C; N; cold water extracts; hot water extracts; other soluble substances; hemicellulose; cellulose; lignin; * significant values (p < 0.05).



Figure 1. Dry wood yield for SRWC genotypes and harvest cycles (Mg ha⁻¹ y⁻¹ DM). A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; genotype × harvest cycle interaction homogeneous groups (see Table A1); error bars—standard error of mean.

The energy value of wood from 1 ha was calculated by multiplying the dry wood yield by its higher heating value (HHV). Obviously, in general, genotypes that give higher wood yields had a higher energy value per unit area. This is why the relationship between the genotypes under study was nearly the same as that of the dry wood yield. The significantly highest wood energy value among the 14 SRWC genotypes under study was calculated for willow of the Żubr cultivar—208.88 GJ ha⁻¹ y⁻¹ DM (Figure 2). A second homogeneous group "B" included the Hybryda275 poplar and S. alba UWM095 willow, and their wood energy value was lower by approx. 21%. The mean energy value of the R. pseudoacacia was lower by as much as 64%. During the quadrennial cycle, the dry wood yield had a higher energy value of nearly 162 GJ ha⁻¹ y⁻¹, which was 37% higher than the annual cycle. The significantly highest wood energy value in the entire experiment (244.34 GJ ha⁻¹ y⁻¹) was calculated for S. viminalis (Żubr cultivar) in the quadrennial harvest cycle (Figure 2, Table A2). It was equivalent to nearly 10 Mg $ha^{-1}y^{-1}$ of hard coal (Table A3). The intermediate homogeneous group "ab" included the Hybryda275 poplar wood in the quadrennial harvest cycle, and the wood energy value was lower by 12% compared with the highest value in this experiment (Figure 2, Table A2). Over 200 GJ ha⁻¹ y⁻¹ was also found for the S. alba UWM095, but the value was lower by 17%. The lowest wood energy value (mere 53 GJ ha⁻¹ y⁻¹) was achieved for *P. balsamifera* UWM3, harvested in the annual cycle. In the annual harvest cycle, the wood energy value did not exceed

100 GJ ha⁻¹ y⁻¹ for seven SRWC genotypes. *R. pseudoacacia* and *P. balsamifera* UWM3 had the lowest energy values in both the annual and quadrennial harvest cycles. The wood energy value, expressed as the coal equivalent, ranged from 2 to 10 Mg ha⁻¹ throughout the experiment (Table A3).



Figure 2. Dry wood energy value for SRWC genotypes and harvest cycles (GJ ha⁻¹ y⁻¹). A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; genotype × harvest cycle interaction homogeneous groups (see Table A2); error bars—standard error of mean.

3.2. Thermophysical Characteristics of SRWC Wood

The thermophysical characteristics of SRWC wood varied significantly based on genotype, plant harvest cycle, and their interaction (Table 1). The highest mean moisture content (56.01%) was determined in the *P. nigra* \times *P. maximowiczii* Max-5 wood (Table 2). The mean moisture content of the other poplar genotype wood was also high and ranged from 53.5% to 55.8%. The willow wood moisture content was lower, and it lay within a range between 43.3% and 51.41% for the *S. pentandra* UWM035 and *S. dasyclados* UWM155 genotypes, respectively. The significantly lowest moisture content of the wood (31.61%) was determined in *R. pseudoacacia*, the homogeneous group "H". The moisture content of wood at the level of the second experiment factor was significantly differentiated because the annual shoot moisture content was higher by 2.13 pp than that of wood from the quadrennial shoots. The largest disproportion of the attribute (nearly 27 pp.) was observed for the moisture content between the wood obtained from the quadrennial shoots of the Max-5 poplar (57.31%) and *R. pseudoacacia* (Table 2).

The significantly highest HHV (19.85 GJ Mg⁻¹ DM) was determined for the wood of the Max-5 poplar (Table 3). The mean HHV for other poplar and willow genotypes ranged from 18.81 to 19.43 GJ Mg⁻¹ DM for Hybryda275 poplar and *S. pentandra* UWM035. On the other hand, *R. pseudoacacia* was characterized by the significantly lowest value of this attribute—19.36 GJ Mg⁻¹ DM. The HHV for the wood in the quadrennial harvest cycle was slightly (but statistically significantly) higher than that calculated for the annual harvest cycle. The highest HHV (19.87 GJ Mg⁻¹ DM) in the entire study was determined for the wood obtained from quadrennial shoots of the *S. alba* UWM095, while the lowest was for *R. pseudoacacia* (19.33 GJ Mg⁻¹ DM) in the quadrennial harvest cycle (Table 3). The

results showed a significant positive correlation between HHV and the carbon, lignin, and cellulose content, as presented in Table A4.

Table 2. Moisture content of wood for SRWC genotypes and harvest cycles (%).

	Harves		
Genotype —	Annual	Quadrennial	— Mean
R. pseudoacacia	$32.61 \pm 1.13 \text{ m}$	$30.6\pm0.16~\mathrm{m}$	$31.61\pm0.68~\mathrm{H}$
P. nigra $ imes$ P. maximowiczii Max-5	$54.71\pm0.39~\mathrm{abc}$	57.31 ± 0.11 a	$56.01\pm0.61~\mathrm{A}$
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	$53.67\pm1.56~{ m bc}$	$53.37\pm0.18~{ m cd}$	$53.52\pm0.71~\mathrm{BC}$
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	55.13 ± 1.25 abc	$53.22\pm0.30~\mathrm{cde}$	$54.17\pm0.72~\mathrm{AB}$
P. balsamifera UWM2	$56.97\pm0.99~\mathrm{ab}$	$54.72\pm0.08~\mathrm{abc}$	$55.84\pm0.67~\mathrm{A}$
P. balsamifera UWM3	$54.58\pm0.53~\mathrm{abc}$	$55.43\pm0.94~\mathrm{abc}$	$55.00\pm0.52~\mathrm{AB}$
S. alba UWM200	$49.08\pm0.29~\mathrm{ghi}$	$49.41\pm0.42~\mathrm{fghi}$	$49.24\pm0.24~\mathrm{DEF}$
S. alba UWM095	48.57 ± 0.60 hi	46.37 ± 0.13 ijk	$47.47\pm0.57~\mathrm{F}$
S. dasyclados UWM155	$53.07\pm0.36~\mathrm{cde}$	$49.75\pm0.37~\mathrm{efghi}$	$51.41\pm0.78~\mathrm{CD}$
S. fragilis UWM195	$52.48\pm0.39~\mathrm{cdefg}$	$49.58\pm0.48~\mathrm{fghi}$	$51.03\pm0.71~\mathrm{D}$
S. pentandra UWM035	$44.04\pm0.51~\mathrm{jkl}$	$42.60\pm0.16\mathrm{l}$	$43.32\pm0.40~\mathrm{G}$
S. triandra UWM198	52.66 ± 0.79 cdef	$43.87\pm0.52~\mathrm{kl}$	$48.27\pm2.01~\mathrm{F}$
S. viminalis Żubr	$49.99\pm0.78~\mathrm{defgh}$	47.46 ± 0.03 hij	$48.73\pm0.66~\mathrm{EF}$
S. viminalis \times S. purpurea UWM033	$53.86\pm0.54~\mathrm{abc}$	$47.93\pm0.19~\text{hi}$	$50.89\pm1.35~\text{DE}$
Mean	$50.82\pm0.95~\mathrm{X}$	$48.69\pm1.02~\mathrm{Y}$	49.75 ± 0.70

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

Table 3. Higher heating value of wood for SRWC genotypes and harvest cycles (GJ Mg^{-1} DM).

Caracteria	Harves	N/	
Genotype —	Annual	Quadrennial	Mean
R. pseudoacacia	$19.38\pm0.08~\mathrm{fg}$	$19.33 \pm 0.01~{ m g}$	$19.36\pm0.04~\mathrm{F}$
P. nigra \times P. maximowiczii Max-5	$19.84\pm0.06~\mathrm{abc}$	19.86 ± 0.05 a	$19.85\pm0.04~\mathrm{A}$
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	$19.48\pm0.05~\mathrm{defg}$	$19.37\pm0.02~\mathrm{fg}$	$19.43\pm0.03~\mathrm{EF}$
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	$19.55\pm0.04~\mathrm{cdefg}$	19.46 ± 0.02 defg	$19.50\pm0.03~\mathrm{CDEF}$
P. balsamifera UWM2	$19.54\pm0.10~\mathrm{cdefg}$	$19.60\pm0.01~\mathrm{abcdef}$	$19.57\pm0.05~\mathrm{CDE}$
P. balsamifera UWM3	$19.50\pm0.08~{ m defg}$	$19.78\pm0.02~\mathrm{abcd}$	$19.64\pm0.07~\mathrm{BCD}$
S. alba UWM200	19.55 ± 0.09 bcdef	$19.50\pm0.03~\mathrm{defg}$	$19.53\pm0.04~\mathrm{CDE}$
S. alba UWM095	$19.51\pm0.06~\mathrm{defg}$	19.87 ± 0.01 a	$19.69\pm0.09~\text{ABC}$
S. dasyclados UWM155	$19.38\pm0.08~\mathrm{fg}$	$19.58\pm0.01~\mathrm{abcdef}$	$19.48\pm0.06~\mathrm{DEF}$
S. fragilis UWM195	$19.47\pm0.04~\mathrm{defg}$	$19.61\pm0.01~\mathrm{abcdef}$	$19.54\pm0.04~\mathrm{CDE}$
S. pentandra UWM035	$19.87\pm0.06~\mathrm{a}$	$19.75\pm0.02~\mathrm{abcde}$	$19.81\pm0.04~\mathrm{AB}$
S. triandra UWM198	$19.35\pm0.04~ m{g}$	$19.53\pm0.01~{ m defg}$	$19.44\pm0.04~\mathrm{EF}$
S. viminalis Żubr	$19.52\pm0.07~\mathrm{defg}$	$19.67\pm0.02~\mathrm{abcdef}$	$19.59\pm0.05~\text{CDE}$
S. viminalis \times S. purpurea UWM033	$19.34\pm0.16~\mathrm{g}$	$19.67\pm0.01~\mathrm{abcdef}$	$19.50\pm0.10~\text{CDEF}$
Mean	$19.52\pm0.03~\mathrm{Y}$	$19.61\pm0.03~\mathrm{X}$	19.57 ± 0.02

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

The significantly highest mean ash content (0.69% DM) was found for the *R. pseudoa-cacia* wood, homogeneous group "A" (Table 4). The intermediate group "AB" included the wood of two poplar genotypes and two willow ones, and the value of this attribute was lower by 3–4%. The wood of *S. alba* UWM095 had a significantly lower ash content of 0.52% DM, which was 24% lower than *R. pseudoacacia*. The ash content of the wood was significantly affected by the plant harvest cycle. As the harvest cycle became longer, the ash content decreased. Throughout the experiment, the highest ash content (0.81% DM) was found in the wood of annual shoots of *R. pseudoacacia*, homogeneous group "a". On the other hand, the lowest values of the attribute (0.48% DM) were determined in the wood of quadrennial shoots of *S. pentandra* UWM035.

Harves		
Annual	Quadrennial	Mean
0.81 ± 0.04 a	$0.57\pm0.01~ m efghijk$	$0.69\pm0.06~\mathrm{A}$
$0.73\pm0.02~\mathrm{ab}$	0.61 ± 0.02 cdefghi	$0.67\pm0.03~\mathrm{AB}$
$0.62\pm0.03~ m bcdefg$	0.49 ± 0.01 ijk	$0.56\pm0.03~\mathrm{CDE}$
$0.64\pm0.02~ m bcdefg$	0.51 ± 0.02 hijk	$0.57\pm0.03~\mathrm{CDE}$
$0.65\pm0.02~ m bcdef$	$0.59\pm0.01~{ m defghij}$	$0.62\pm0.02~\mathrm{ABC}$
$0.65\pm0.02~\mathrm{bcdef}$	0.69 ± 0.01 bcde	$0.67\pm0.01~\mathrm{AB}$
$0.64\pm0.02~ m bcdefg$	0.58 ± 0.01 efghijk	$0.61\pm0.02~\mathrm{BCD}$
$0.55\pm0.01~{ m fghijk}$	$0.49\pm0.01~{ m jk}$	$0.52\pm0.01~\mathrm{E}$
0.56 ± 0.02 fghijk	0.52 ± 0.01 ghijk	$0.54\pm0.01~\mathrm{DE}$
0.63 ± 0.03 bcdefg	0.57 ± 0.01 fghijk	$0.60\pm0.02~\mathrm{BCD}$
$0.64\pm0.02~ m bcdefg$	$0.48\pm0.01~{ m k}$	$0.56\pm0.04~\mathrm{CDE}$
$0.70\pm0.03~\mathrm{abcd}$	$0.63\pm0.02\mathrm{bcdefg}$	$0.67\pm0.02~\mathrm{AB}$
$0.64\pm0.04~ m bcdefg$	$0.58\pm0.01\mathrm{efghijk}$	$0.61\pm0.02~\mathrm{BCD}$
$0.71\pm0.05~{ m abc}$	0.60 ± 0.02 cdefghi	$0.66\pm0.03~\mathrm{AB}$
$0.65\pm0.01~\mathrm{X}$	$0.56\pm0.01~\mathrm{Y}$	0.61 ± 0.01
	Harves Annual 0.81 ± 0.04 a 0.73 ± 0.02 ab 0.62 ± 0.03 bcdefg 0.64 ± 0.02 bcdefg 0.65 ± 0.02 bcdef 0.65 ± 0.02 bcdef 0.65 ± 0.02 bcdef 0.65 ± 0.02 bcdef 0.64 ± 0.02 bcdefg 0.55 ± 0.01 fghijk 0.56 ± 0.02 fghijk 0.63 ± 0.03 bcdefg 0.64 ± 0.02 bcdefg 0.70 ± 0.03 abcd 0.64 ± 0.04 bcdefg 0.71 ± 0.05 abc 0.65 ± 0.01 X	Harvest CycleAnnualQuadrennial 0.81 ± 0.04 a 0.57 ± 0.01 efghijk 0.73 ± 0.02 ab 0.61 ± 0.02 cdefghi 0.62 ± 0.03 bcdefg 0.49 ± 0.01 ijk 0.64 ± 0.02 bcdefg 0.51 ± 0.02 hijk 0.65 ± 0.02 bcdef 0.59 ± 0.01 defghij 0.65 ± 0.02 bcdef 0.69 ± 0.01 bcde 0.64 ± 0.02 bcdef 0.69 ± 0.01 bcde 0.65 ± 0.02 bcdef 0.69 ± 0.01 bcde 0.64 ± 0.02 bcdefg 0.58 ± 0.01 efghijk 0.55 ± 0.01 fghijk 0.49 ± 0.01 jk 0.56 ± 0.02 fghijk 0.52 ± 0.01 ghijk 0.56 ± 0.02 fghijk 0.57 ± 0.01 fghijk 0.64 ± 0.02 bcdefg 0.63 ± 0.02 bcdefg 0.70 ± 0.03 abcd 0.63 ± 0.02 bcdefg 0.70 ± 0.03 abcd 0.63 ± 0.02 bcdefg 0.64 ± 0.04 bcdefg 0.58 ± 0.01 efghijk 0.71 ± 0.05 abc 0.60 ± 0.02 cdefghi 0.65 ± 0.01 X 0.56 ± 0.01 Y

Table 4. Ash content in wood for SRWC genotypes and harvest cycles (% DM).

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

The significantly highest mean fixed carbon content (19.96% DM) and the lowest volatile matter content (79.37% DM) were determined for the wood of *R. pseudoacacia* from homogeneous groups "A" and "D", respectively (Tables A5 and A6). Further, it was the opposite for the *S. triandra* UWM198 genotype, as the mean fixed carbon content was the lowest (17.67% DM), and the volatile matter was the highest (81.70% DM). The fixed carbon content ranged from 17.67 to 20.01% DM throughout the experiment (Table A5). The volatile matter content ranged from 79.22 to 81.70% DM (Table A6).

3.3. Elemental Composition of SRWC Wood

The content of C, N, Cl, and S elements in SRWC wood was significantly affected by the SRWC genotype, plant harvest cycle, and their interaction. Specifically, the C, N, and Cl contents were differentiated by both factors, while the S content was only affected by the main factors. The harvest cycle, on the other hand, only significantly differentiated the H content (Table 1). The mean carbon content of the genotypes under study lay within a narrow range between 55.03% DM for the Androscoggin poplar and 55.96% DM for *P. balsamifera* UWM3, and homogeneous groups "A" and "B", respectively (Table 5). The wood harvested in the quadrennial cycle contained significantly more carbon (55.86% DM) compared with its annual harvest, although the difference was only 0.7 pp. During the experiment, the wood of quadrennial shoots of *P. balsamifera* UWM3, homogeneous group "a", had the highest carbon content of 56.65% DM. On the other hand, the wood of annual shoots of *R. pseudoacacia*, the homogeneous group "g", had the lowest carbon content of 54.36% DM.

The hydrogen content of the wood of all the genotypes exceeded 6% DM and ranged from 6.34 to 6.54% DM (Table A7). Wood harvested in the annual cycle contained slightly yet significantly more hydrogen (6.47% DM) compared with its quadrennial harvest. The wood of *R. pseudoacacia* contained the highest N level (0.38% DM), group "A" (Table 6). The second homogeneous group, "B", included wood of the poplar Max5, and its content of this element was lower by approx. 23% compared with *R. pseudoacacia*. The lowest mean N content (0.17% DM) was found in the wood *S. pentandra* UWM035, which was lower by approx. 57% compared with *R. pseudoacacia*. The mean N content of the annual cycle (0.30% DM) was higher by 46% compared with its quadrennial cycle. Throughout the experiment, the highest nitrogen content (0.45% DM) was found in the wood of annual shoots of *R. pseudoacacia*, homogeneous group "a". On the other hand, the lowest values of nitrogen content (0.12% DM) were determined in the wood of quadrennial

shoots of three willow genotypes: *S. pentandra, S. triandra,* and *S. viminalis,* homogeneous group "k". The nitrogen content was significantly and positively correlated with the fixed carbon, ash, sulfur, soluble substance, and hemicellulose content (Table A4).

Table 5. Carbon content in wood for SRWC genotypes and harvest cycles (% DM).

	Harves	Harvest Cycle		
Genotype –	Annual	Quadrennial	Mean	
R. pseudoacacia	$54.36\pm0.21~{ m g}$	$56.10\pm0.09~\mathrm{abcd}$	$55.23\pm0.40~\mathrm{AB}$	
P. nigra \times P. maximowiczii Max-5	55.56 ± 0.09 abcdef	$56.26\pm0.09~\mathrm{ab}$	$55.91\pm0.17~\mathrm{A}$	
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	$55.11\pm0.12~\mathrm{cdefg}$	55.37 ± 0.05 bcdefg	$55.24\pm0.08~\mathrm{AB}$	
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	55.42 ± 0.23 bcdefg	$54.65\pm0.01~{ m fg}$	$55.03\pm0.20~\mathrm{B}$	
P. balsamifera UWM2	$54.84\pm0.30~\mathrm{efg}$	$56.14\pm0.05 m ab$	$55.49\pm0.32~\mathrm{AB}$	
P. balsamifera UWM3	55.28 ± 0.38 cdefg	56.65 ± 0.13 a	$55.96\pm0.35~\mathrm{A}$	
S. alba UWM200	55.22 ± 0.13 cdefg	$56.55\pm0.09~\mathrm{ab}$	$55.88\pm0.31~\mathrm{A}$	
S. alba UWM095	55.59 ± 0.22 abcdef	$56.25\pm0.12~\mathrm{ab}$	$55.92\pm0.18~\mathrm{A}$	
S. dasyclados UWM155	55.37 ± 0.29 bcdefg	$56.16\pm0.15~\mathrm{ab}$	$55.76\pm0.23~\mathrm{AB}$	
S. fragilis UWM195	$54.87\pm0.45~\mathrm{efg}$	55.54 ± 0.23 abcdefg	$55.20\pm0.27~\mathrm{AB}$	
S. pentandra UWM035	55.91 ± 0.19 abcde	$55.61\pm0.06~\mathrm{abcdef}$	$55.76\pm0.11~\mathrm{AB}$	
S. triandra UWM198	$54.76\pm0.40~\mathrm{efg}$	55.46 ± 0.12 abcdefg	$55.11\pm0.24~\mathrm{B}$	
S. viminalis Żubr	$54.89\pm0.18~ m defg$	$55.59\pm0.14~ m abcdef$	$55.24\pm0.19~\text{AB}$	
5. viminalis $ imes$ S. purpurea UWM033	$55.05\pm0.45~\mathrm{cdefg}$	55.69 ± 0.22 abcdef	$55.37\pm0.27~\text{AB}$	
Mean	$55.16\pm0.09~\mathrm{Y}$	$55.86 \pm 0.09 \text{ X}$	55.51 ± 0.07	

A, B—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

Table 6. Nitrogen content in wood for SRWC genotypes and harvest cycles (% DM).

Constants	Harves	Maria	
Genotype	Annual	Quadrennial	- Mean
R. pseudoacacia	0.45 ± 0.01 a	$0.31\pm0.01~{ m cd}$	$0.38\pm0.03~\mathrm{A}$
P. nigra $ imes$ P. maximowiczii Max-5	$0.43\pm0.01~\mathrm{ab}$	0.16 ± 0.02 ijk	$0.29\pm0.06~\mathrm{B}$
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	$0.29\pm0.02~{ m de}$	0.17 ± 0.01 hijk	$0.23\pm0.03~\mathrm{CDE}$
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	0.29 ± 0.03 de	0.18 ± 0.01 hijk	$0.24\pm0.03~\mathrm{CDE}$
P. balsamifera UWM2	$0.32\pm0.01~{ m cd}$	$0.13\pm0.01\mathrm{jk}$	$0.23\pm0.04~\mathrm{CDE}$
P. balsamifera UWM3	$0.38\pm0.02~{ m bc}$	0.16 ± 0.02 ijk	$0.27\pm0.05~\mathrm{BC}$
S. alba UWM200	$0.24\pm0.01~\mathrm{efgh}$	0.19 ± 0.02 ghij	$0.21\pm0.01~\mathrm{DEF}$
S. alba UWM095	$0.31\pm0.01~{ m cd}$	$0.20\pm0.02~{ m fghi}$	$0.26\pm0.03~\mathrm{BCD}$
S. dasyclados UWM155	$0.29\pm0.02~{ m de}$	0.16 ± 0.01 ijk	$0.23\pm0.03~\mathrm{CDE}$
S. fragilis UWM195	$0.26\pm0.03~{ m def}$	0.14 ± 0.02 ijk	$0.20\pm0.03~\mathrm{EFG}$
S. pentandra UWM035	$0.21\pm0.01~{ m fghi}$	$0.12\pm0.02~{ m k}$	$0.17\pm0.02~{ m G}$
S. triandra UWM198	$0.26\pm0.03~{ m def}$	$0.12\pm0.01~{ m k}$	$0.19\pm0.03~\mathrm{EFG}$
S. viminalis Żubr	$0.23\pm0.01~\mathrm{efgh}$	$0.12\pm0.01~{ m k}$	$0.18\pm0.03~\mathrm{FG}$
S. viminalis \times S. purpurea UWM033	$0.26\pm0.01~{ m def}$	$0.13\pm0.01~\text{jk}$	$0.19\pm0.03~\text{EFG}$
Mean	$0.30\pm0.01~{ m X}$	$0.16\pm0.01~\mathrm{Y}$	0.23 ± 0.01

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

The wood of *R. pseudoacacia* also contained the significantly largest amount of sulfur (0.033% DM), homogeneous group "A" (Table 7). The second group "B" included the wood of *P. balsamifera* UWM3, and its sulfur level was lower by approx. 20% compared with *R. pseudoacacia*. Further, the lowest S content (0.020% DM) was found in the wood of *S. pentandra* UWM035 and three hybrid poplar genotypes in the homogeneous group "D". The other genotypes were grouped together in the intermediate homogeneous groups. The mean sulfur content of wood obtained during the annual harvest cycle was 0.027% dry matter, which was significantly higher (by 23%) compared to the quadrennial cycle. Throughout the experiment, the wood of annual shoots of *R. pseudoacacia* had the highest

sulfur content (0.038% dry matter), while the lowest values of this attribute (0.013% dry matter) were found in the wood of quadrennial shoots of the Max5 poplar. The sulfur content was significantly and positively correlated with the ash, nitrogen, soluble substance, and hemicellulose content (Table A4).

Table 7. Sulfur content in wood for SRWC genotypes and harvest cycles (% DM).

Canadama	Harve	Harvest Cycle		
Genotype —	Annual	Quadrennial	— Mean	
R. pseudoacacia	0.038 ± 0.002	0.029 ± 0.001	$0.033 \pm 0.002 \text{ A}$	
P. nigra \times P. maximowiczii Max-5	0.026 ± 0.001	0.013 ± 0.001	$0.020 \pm 0.003 \text{ D}$	
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	0.024 ± 0.002	0.016 ± 0.002	$0.020\pm0.002~\mathrm{D}$	
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	0.023 ± 0.001	0.017 ± 0.001	$0.020 \pm 0.001 \ { m D}$	
P. balsamifera UWM2	0.025 ± 0.001	0.017 ± 0.001	$0.021\pm0.002\mathrm{CD}$	
P. balsamifera UWM3	0.029 ± 0.002	0.025 ± 0.001	$0.027\pm0.001~\mathrm{B}$	
S. alba UWM200	0.029 ± 0.001	0.023 ± 0.001	$0.026\pm0.002~\mathrm{BC}$	
S. alba UWM095	0.026 ± 0.001	0.024 ± 0.002	$0.025\pm0.001~\mathrm{BCD}$	
S. dasyclados UWM155	0.026 ± 0.001	0.023 ± 0.001	$0.024\pm0.001~\mathrm{BCD}$	
S. fragilis UWM195	0.025 ± 0.003	0.021 ± 0.002	$0.023\pm0.002~\text{BCD}$	
S. pentandra UWM035	0.025 ± 0.004	0.015 ± 0.001	$0.020 \pm 0.003 \text{ D}$	
S. triandra UWM198	0.028 ± 0.006	0.024 ± 0.001	$0.026\pm0.003~\mathrm{BC}$	
S. viminalis Żubr	0.026 ± 0.002	0.024 ± 0.001	$0.025\pm0.001~\mathrm{BCD}$	
S. viminalis \times S. purpurea UWM033	0.027 ± 0.003	0.020 ± 0.001	$0.024\pm0.002~\text{BCD}$	
Mean	$0.027\pm0.001~\mathrm{X}$	$0.021\pm0.001~\mathrm{Y}$	0.024 ± 0.001	

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; no letter indicates no significant differences, \pm —standard error of mean.

The highest mean chlorine content (0.019% DM) was found in the wood of two willow genotypes: *S. fragilis* UWM195 and *S. viminalis* × *S. purpurea* UWM033, group "A" (Table A8). The wood of *P. balsamifera* UWM2, which belonged to the homogeneous group "C", had the lowest Cl content (0.013% DM) among all the species tested. The other genotypes were grouped in intermediate homogeneous groups. The mean Cl content of the wood obtained in the annual harvest cycle (0.017% DM) was significantly higher (by 8%) than the content obtained in the quadrennial cycle. The highest Cl content throughout the experiment (0.020% DM) was found in the wood of annual shoots of *S. triandra* and *S. viminalis* × *S. purpurea*. On the other hand, the lowest values of the attribute (0.012% DM) were found in the wood of annual shoots of *P. balsamifera* UWM2.

3.4. Lignocellulosic Composition of SRWC Wood

The SRWC genotype, plant harvest cycle, and their interaction significantly differentiated cold and hot water extracts, other soluble substances, and the hemicellulose, cellulose, and lignin content (Table 1). *P. balsamifera* UWM3 had the highest cold water extract content (4.62% DM) among all the woods (Table A9). The wood of *R. pseudoacacia* belonged to the second homogeneous group, "B". On the other hand, the wood of *S. pentandra* UWM035 had the lowest cold water extract content (2.54% DM). The amount of cold water extract content in wood is affected by the plant harvest cycle. As the harvest cycle becomes longer, the content decreases by approximately 41%. The wood from annual shoots of *P. balsamifera* UWM3 had the highest cold water extract content (5.73% DM), while the wood from quadrennial shoots of willow, cultivar Żubr, had the lowest (1.44% DM).

The significantly highest hot water extract content (7.80% DM) was found in the wood of *R. pseudoacacia* (Table A10). The second homogeneous group, "B", included the wood of *P. balsamifera* UWM2. The lowest hot water extract content (4.73% DM) was found in the wood of *P. max.* \times *P. trich.* Androscoggin. The plant harvest cycle also significantly affected the hot water extract content of the wood. As the harvest cycle became longer, there was an approximate 30% decrease in the cold water extract content. The wood from annual shoots of *P. balsamifera* UWM3 had the highest cold water extract content, which was 9.42% DM.

On the other hand, the wood of quadrennial shoots of *P. max.* \times *P. trich.* Androscoggin had the lowest cold water extract content, which was 3.69% DM.

The wood of *S. fragilis* UWM195 had the highest content of other soluble substances (6.39% DM) compared to other woods (Table A11). However, the wood of willow, cultivar Żubr, had the lowest level of other soluble substances (3.53% DM). The content of other soluble substances in wood was also affected by the plant harvest cycle. As the harvest cycle became longer, the content decreased by approximately 15%. The wood from annual shoots of *S. fragilis* UWM195 had the highest content of other soluble substances (7.48% DM), while the wood of annual shoots of willow, cultivar Żubr, had the lowest content (3.35% DM).

The significantly highest mean hemicellulose content (27.30% DM) was found in the *R. pseudoacacia* wood (Table 8). The second group, "B", included the wood of *P. balsamifera* UWM2, and the biopolymer content of it was lower by approx. 6% compared with *R. pseudoacacia*. In this study, it was found that the wood of two willow genotypes, *S. pentandra* UWM035 and *S. alba* UWM095, belonging to the homogeneous group "I", had the lowest hemicellulose content, which was 25% less than the highest content. The mean hemicellulose content of wood obtained in the annual harvest cycle was 24.77% DM, which was significantly higher (by 14%) than that of the quadrennial harvest cycle. The significantly highest hemicellulose content throughout the experiment (28.32% DM) was found in the *R. pseudoacacia* wood from annual shoots, and the lowest (18.45% DM) was in the *S. pentandra* UWM 035 wood from quadrennial shoots. The ash, nitrogen, hot water extract, and cold water extract contents showed a significant positive correlation with the hemicellulose content (Table A4).

Complement	Harves		
Genotype	Annual	Quadrennial	Mean
R. pseudoacacia	$28.32\pm0.20~\mathrm{a}$	$26.27\pm0.09~\mathrm{cd}$	$27.30\pm0.47~\mathrm{A}$
P. nigra \times P. maximowiczii Max-5	$24.62\pm0.29~\mathrm{ef}$	$19.80\pm0.05\mathrm{l}$	$22.21\pm1.09~G$
P. max. \times P. trich. Hybryda275	$27.10\pm0.05\mathrm{bc}$	22.48 ± 0.11 hi	$24.79\pm1.03~\mathrm{C}$
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	$26.90\pm0.24\mathrm{bc}$	$21.47\pm0.21~\mathrm{jk}$	$24.18\pm1.22~\mathrm{D}$
P. balsamifera UWM2	$27.36\pm0.09~\mathrm{b}$	$23.94\pm0.01~{ m fg}$	$25.65\pm0.76~\mathrm{B}$
P. balsamifera UWM3	$25.43\pm0.08~\mathrm{de}$	$20.94\pm0.14~{\rm k}$	$23.18\pm1.01~\mathrm{E}$
S. alba UWM200	$23.71\pm0.26~\mathrm{fg}$	$19.29\pm0.13~\mathrm{lm}$	$21.50\pm1.01~\mathrm{H}$
S. alba UWM095	21.88 ± 0.13 ij	$19.07\pm0.14~\mathrm{lm}$	$20.47\pm0.63~\mathrm{I}$
S. dasyclados UWM155	$24.51\pm0.07~\mathrm{ef}$	$22.07\pm0.40~\mathrm{ij}$	$23.29\pm0.57~\mathrm{E}$
S. fragilis UWM195	$23.42\pm0.02~\mathrm{g}$	$19.01\pm0.14~\mathrm{lm}$	$21.21\pm0.99~\mathrm{H}$
S. pentandra UWM035	22.41 ± 0.13 hi	18.45 ± 0.26 m	$20.43\pm0.90~\mathrm{I}$
S. triandra UWM198	$23.20\pm0.06~\mathrm{gh}$	$21.23\pm0.12~\mathrm{jk}$	$22.22\pm0.44~\mathrm{G}$
S. viminalis Żubr	$24.11\pm0.04~\mathrm{fg}$	21.69 ± 0.14 ijk	$22.90\pm0.54~\mathrm{EF}$
S. viminalis \times S. purpurea UWM033	$23.88\pm0.19~\mathrm{fg}$	$20.85\pm0.20~\mathrm{k}$	$22.36\pm0.69~\text{FG}$
Mean	$24.77\pm0.30~\mathrm{X}$	$21.18\pm0.32~\mathrm{Y}$	22.98 ± 0.29

Table 8. Hemicellulose content in wood for SRWC genotypes and harvest cycles (% DM).

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

The wood of *S. pentandra* UWM035 had the highest mean cellulose content (52.81% DM) (Table 9). This biopolymer content also exceeded 52% DM in three other willow genotypes. However, the cellulose content of poplar wood ranged from 49.24 to 50.85% DM for *P. nigra* \times *P. maximowiczii* Max5 and Hybryda275 poplar. Further, the lowest cellulose content (45.26% DM) was found in the *R. pseudoacacia* wood, homogeneous group "I", which meant that it was lower by 14% than the highest mean value for *S. pentandra* UWM035. The mean cellulose content of wood obtained in the quadrennial harvest cycle (53.71% DM) was higher (by 11%) compared with its annual harvest cycle. The significantly highest cellulose content throughout the experiment (56.97% DM) was found in the wood

of the quadrennial shoots of willow *S. triandra* UWM198, and the lowest (43.69% DM) was in the wood of the annual shoots of *R. pseudoacacia*.

Complexed	Harves		
Genotype —	Annual	Quadrennial	– Mean
R. pseudoacacia	$43.69\pm0.03\mathrm{l}$	46.83 ± 0.06 j	$45.26\pm0.7~{ m G}$
P. nigra $ imes$ P. maximowiczii Max-5	$45.16\pm0.22~k$	53.32 ± 0.02 ef	$49.24\pm1.83~\mathrm{F}$
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	$48.08\pm0.20\mathrm{i}$	$53.62\pm0.18~\mathrm{de}$	$50.85\pm1.24~\mathrm{E}$
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	$47.74\pm0.19~\mathrm{i}$	$53.53\pm0.10~\mathrm{de}$	$50.63 \pm 1.30~\mathrm{E}$
P. balsamifera UWM2	$45.23\pm0.16~\mathrm{k}$	$54.25\pm0.09~cd$	$49.74\pm2.02~\mathrm{F}$
P. balsamifera UWM3	$46.38\pm0.01~{ m j}$	$52.96\pm0.15~\mathrm{ef}$	$49.67\pm1.47~\mathrm{F}$
S. alba UWM200	$50.58\pm0.13~{ m g}$	$53.27\pm0.21~\mathrm{ef}$	$51.93\pm0.61\mathrm{CD}$
S. alba UWM095	$48.51\pm0.12\mathrm{\ddot{i}}$	$52.64\pm0.21~\mathrm{f}$	$50.58\pm0.93~\mathrm{E}$
S. dasyclados UWM155	$49.76\pm0.14~\mathrm{gh}$	$54.80\pm0.15\mathrm{bc}$	$52.28 \pm 1.13~\mathrm{ABC}$
S. fragilis UWM195	46.00 ± 0.21 jk	$54.64\pm0.27\mathrm{bc}$	$50.32\pm1.94~\mathrm{E}$
S. pentandra UWM035	$50.60\pm0.03~\mathrm{g}$	$55.03\pm0.17\mathrm{bc}$	$52.81\pm0.99~\mathrm{A}$
S. triandra UWM198	$46.01\pm0.14~\mathrm{jk}$	56.97 ± 0.09 a	$51.49\pm2.45~\mathrm{D}$
S. viminalis Żubr	$50.18\pm0.15~{ m gh}$	$55.26\pm0.01~\mathrm{b}$	$52.72 \pm 1.14~\mathrm{AB}$
S. viminalis $ imes$ S. purpurea UWM033	$49.57\pm0.05h$	$54.80\pm0.3~{ m bc}$	$52.18\pm1.18~\text{BC}$
Mean	$47.68\pm0.34~\mathrm{Y}$	$53.71\pm0.35~\mathrm{X}$	50.69 ± 0.41

Table 9. Cellulose content in wood for SRWC genotypes and harvest cycles (% DM).

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

The significantly highest mean lignin content (17.86% DM) was found in the wood of poplar *P. nigra* \times *P. maximowiczii* Max5 (Table 10). The lignin content was significantly lower in the other poplar genotypes, and it was the significantly lowest in the wood of *P. balsamifera* UWM2—13.44% DM, homogeneous group "I". The lignin content of the wood of willow genotypes lay within the range of 14.07 to 17.12% DM for *S. triandra* UWM198 and *S. alba* UWM200, respectively. The wood of *R. pseudoacacia* had a lignin content of nearly 15% DM. The quadrennial harvest cycle produced wood with a mean lignin content of 15.88% DM, which was 3% higher than that of the annual cycle. The significantly highest lignin content throughout the experiment (18.33% DM) was found in the wood from quadrennial shoots of *P. balsamifera* UWM2. The lignin content was correlated significantly positively with HHV as well as fixed and elemental carbon content (Table A4).

Table 10. Lignin content in wood for SRWC genotypes and harvest cycles (% DM).

Canatyna	Harves	Maar	
Genotype	Annual	Quadrennial	Mean
R. pseudoacacia P. nigra × P. maximowiczii Max-5 P. max. × P. trich. Hybryda275 P. max. × P. trich. Androscoggin P. balsamifera UWM2 P. balsamifera UWM3 S. alba UWM200 S. alba UWM200 S. alba UWM095 S. dasyclados UWM155 S. fragilis UWM195 S. pentandra UWM035 S. triandra UWM198	$\begin{array}{c} 14.23 \pm 0.13 \ \text{klmn} \\ 17.39 \pm 0.25 \ \text{b} \\ 15.22 \pm 0.02 \ \text{ghij} \\ 15.20 \pm 0.10 \ \text{ghij} \\ 13.84 \pm 0.14 \ \text{lmno} \\ 13.77 \pm 0.01 \ \text{mno} \\ 17.02 \pm 0.16 \ \text{bc} \\ 16.50 \pm 0.29 \ \text{cde} \\ 14.60 \pm 0.15 \ \text{jklm} \\ 14.95 \pm 0.12 \ \text{hijk} \\ 17.17 \pm 0.11 \ \text{bc} \\ 14.69 \pm 0.10 \ \text{ikl} \end{array}$	$\begin{array}{c} 15.73 \pm 0.07 \ \mathrm{efg} \\ 18.33 \pm 0.13 \ \mathrm{a} \\ 16.05 \pm 0.13 \ \mathrm{def} \\ 17.42 \pm 0.09 \ \mathrm{b} \\ 13.04 \pm 0.12 \ \mathrm{o} \\ 15.50 \pm 0.10 \ \mathrm{ghi} \\ 17.21 \pm 0.24 \ \mathrm{bc} \\ 16.79 \pm 0.07 \ \mathrm{bcd} \\ 14.60 \pm 0.15 \ \mathrm{jklm} \\ 16.02 \pm 0.19 \ \mathrm{def} \\ 17.04 \pm 0.17 \ \mathrm{bc} \\ 13.46 \pm 0.13 \ \mathrm{no} \end{array}$	$\begin{array}{c} 14.98 \pm 0.34 \ \mathrm{FG} \\ 17.86 \pm 0.24 \ \mathrm{A} \\ 15.64 \pm 0.20 \ \mathrm{E} \\ 16.31 \pm 0.50 \ \mathrm{CD} \\ 13.44 \pm 0.20 \ \mathrm{I} \\ 14.64 \pm 0.39 \ \mathrm{G} \\ 17.12 \pm 0.14 \ \mathrm{B} \\ 16.64 \pm 0.15 \ \mathrm{BC} \\ 14.60 \pm 0.09 \ \mathrm{GH} \\ 15.49 \pm 0.26 \ \mathrm{EF} \\ 17.10 \pm 0.10 \ \mathrm{B} \\ 14.07 \pm 0.28 \ \mathrm{H} \end{array}$
<i>S. viminalis</i> Żubr <i>S. viminalis</i> × <i>S. purpurea</i> UWM033	15.71 ± 0.29 efg 15.90 ± 0.15 ef	15.21 ± 0.07 ghij 15.95 ± 0.25 def	$15.46 \pm 0.17 \text{ EF}$ $15.93 \pm 0.13 \text{ DE}$
Mean	$15.44\pm0.19~\mathrm{Y}$	$15.88\pm0.23~\mathrm{X}$	15.66 ± 0.15

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

3.5. General Characteristics of SRWC Wood

Table 11 presents statistics for the data on wood characteristics for all of the 14 SRWC genotypes harvested in the two harvest cycles. These results show the lowest variation (coefficient of variation < 5%) for the following attributes: VM (0.8%), HHV (0.9%), C (1.2%), H (1.4%), and FC (3.4%). The mean values for these parameters were the following: 80.46% DM, 19.57 GJ Mg⁻¹ DM, 50.51% DM, 6.45% DM, and 18.96% DM. The variation of cellulose and lignin content was also low (coefficient of variation < 10%), given the fact that the experiment dealt with the wood of several different SRWC genotypes in two different harvest cycles. The range (minimum-maximum) within which those practically important attributes lay was large: 43.6–57.1% DM for cellulose and 12.8–18.5% DM for lignin. The mean values for these attributes were 50.7 and 15.7% DM, respectively. The highest variability (40.3%) was observed for the N content, which ranged between 0.11 and 0.47% DM. High variability was also observed in cold water extracts (coefficient of variation 35.3%), dry wood yield (36.9%), and the energy value of wood (37.1%). The high variability of dry wood yield shows how important it is to select the right SRWC genotype and the right plant harvest cycle, as it is essential for achieving a satisfying dendromass yield, especially since the minimum dry wood yield was barely 2.7 Mg ha⁻¹ y⁻¹, and the maximum was 13.1 Mg ha⁻¹ y⁻¹. In consequence, the energy value of wood was highly diverse and ranged from 52.5 to 256.8 GJ ha⁻¹ y⁻¹.

Table 11. Selected statistical ana	ysis indicators for all features of	of the SRWC wood (N Valid = 84).
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Feature	Mean	Minimum Value	Maximum Value	Lower Quartile	Upper Quartile	Standard Deviation	Coefficient of Variation (%)
Dry wood yield (Mg ha^{-1} y^{-1} DM)	6.73	2.69	13.06	4.93	8.54	2.48	36.89
Dry wood energy value (GJ ha ^{-1} y ^{-1})	131.83	52.52	256.85	97.57	166.92	48.87	37.07
Coal equivalent (Mg ha ^{-1} y ^{-1})	5.27	2.10	10.27	3.90	6.68	1.95	37.07
Moisture content (%)	49.75	30.43	58.77	47.53	54.05	6.45	12.96
Higher heating value (MJ kg $^{-1}$ DM)	19.57	19.08	19.97	19.44	19.69	0.18	0.94
Ash content (% DM)	0.61	0.47	0.86	0.57	0.65	0.08	13.52
Fixed carbon (% DM)	18.96	17.63	20.39	18.50	19.33	0.65	3.43
Volatile matter (% DM)	80.46	79.04	81.81	80.06	80.94	0.67	0.83
C (% DM)	55.51	53.98	56.87	55.16	56.00	0.66	1.19
H (% DM)	6.45	6.30	6.70	6.37	6.50	0.09	1.41
N (% DM)	0.23	0.11	0.47	0.16	0.31	0.09	40.29
S (% DM)	0.024	0.012	0.041	0.020	0.027	0.006	23.68
Cl (% DM)	0.017	0.010	0.023	0.014	0.019	0.003	17.35
Cold water extracts (% DM)	3.31	1.25	5.82	2.33	4.13	1.17	35.33
Hot water extracts (% DM)	6.16	3.62	9.76	4.87	7.30	1.67	27.10
Other soluble substances (% DM)	4.51	3.27	7.50	3.86	5.03	0.98	21.73
Hemicellulose (% DM)	22.98	17.98	28.67	21.14	24.55	2.69	11.72
Cellulose (% DM)	50.69	43.64	57.12	47.17	54.01	3.76	7.42
Lignin (% DM)	15.66	12.82	18.54	14.71	16.78	1.35	8.62

A cluster analysis based on the values of all the attributes of the wood from the 14 SRWC genotypes harvested in two different harvest cycles at the cut-off point of $2/3 D_{max}$ allowed grouping them into four main clusters (Figure 3a). R. pseudoacacia made its own cluster. Two genotypes of poplar P. balsamifera, UWM2 and UWM3, made a second cluster. Two genotypes of willow, S. fragilis UWM195 and S. triandra UWM198, made a third cluster. The remaining three genotypes of poplar and six genotypes of willow made a fourth joint cluster. When the analysis accuracy increased, nine clusters were identified at $1/3 D_{max}$. The first three clusters remained unchanged, as with the cut-off at $2/3 D_{max}$. The fourth cluster was broken down into six smaller ones. The four genotypes *P. nigra* \times *P.* maximowiczii Max5, S. alba UWM095, S. pentandra UWM035, and S. viminalis cultivar Zubr made four independent clusters. The other two poplar hybrids and S. dasyclados UWM155 made another cluster. The last cluster included two further willow genotypes: S. alba UWM200 and *S. viminalis* × *S. purpurea* UWM033. In addition, the analysis of SRWC wood characteristics revealed the presence of two distinct clusters when the cut-off was set at $2/3 D_{max}$ (as shown in Figure 3b). One of these clusters contained eight attributes, namely ash, sulfur, nitrogen, fixed carbon content, cold water extracts, hot water extracts, other soluble substances, and hemicellulose. A second cluster included the other 11 analyzed



parameters: dry wood yield, energy value of wood yield, coal equivalent, moisture, HHV, volatile matter content, C, H, Cl, cellulose, and lignin. When the analysis accuracy increased, five clusters were identified at $1/3 D_{max}$.

Figure 3. Dendrogram of a hierarchical cluster analysis showing the similarities of SRWC genotypes (a) and their wood characteristics (b). The red vertical line marks the Sneath criterion $(2/3 D_{max})$ and $(1/3 D_{max})$. D—linage distance; D_{max} —maximum linage distance. *Robinia pseudoacacia* (RpUWM1); *P. nigra* × *P. maximowiczii* Max-5 (Pn × mMax5); *P. maximowiczii* × *P. trichocarpa* Hybryda275 (Pm × tHybryda275); *P. maximowiczii* × *P. trichocarpa*, Androscoggin (Pm × tAndroscoggin); *P. balsamifera* UWM2 (PbUWM2); *P. balsamifera* UWM3 (PbUWM3); *S. alba* UWM200 (SaUWM200); *S. alba* UWM095 (SaUWM095); *S. dasyclados* UWM155 (SdUWM155); *S. fragilis* UWM195 (SfUWM195); *S. pentandra* UWM035 (SpUWM035); *S. triandra* UWM198 (StUWM198); *S. viminalis* × *S. purpurea* UWM033 (Sv × pUWM033); *S. viminalis* cultivar Żubr (SvŻubr).

4. Discussion

4.1. Wood Yield and Its Energy Value

Wood is the major component of SRWC shoot lignocellulosic biomass, whose share can vary depending on many factors, including species, genotype, and harvest cycle. The dry wood content of the dry biomass yield, obtained from an annual cycle in the current experiment, was 62.8% DM, and ranged from 52.0 to 72.8% DM for P. nigra \times P. maximowiczii Henry Max-5 and S. triandra UWM198, respectively. These values were higher in the quadrennial harvest cycle, with the mean wood content being 79.3% DM and ranging from 73.5 to 84.8% DM for P. balsamifera UWM3 and S. triandra UWM198, respectively. The wood content of annual willow shoots, in another study, was 71.9% DM, and ranged from 62.3 to 74.9% DM [36]. This means that these levels were close to those found in the current study. Therefore, one should note that when bark is obtained from SRWC shoots, large amounts of pure wood are left as residues after shoot debarking, which can be used in a variety of ways in biorefineries, the paper industry, or for energy generation. Obviously, the wood yield in the current experiment was strongly determined by the genotype and harvest cycle. Nevertheless, it was high in the case of most genotypes, especially in the quadrennial harvest cycle, with a mean level of 8.3 Mg ha⁻¹ y⁻¹ DM, ranging from nearly 5 to over 12 Mg ha⁻¹ y⁻¹ DM, for *R. pseudoacacia* and *S. viminalis* cultivar Zubr, respectively. For the annual harvest cycle, the values ranged from 3 to nearly 9 Mg ha⁻¹ y⁻¹ DM for these genotypes. Moreover, the willow genotypes gave a higher yield in the annual harvest cycle than the poplar or *R. pseudoacacia* genotypes. However, the difference in the wood yield between willow and poplar was not so great in the longer harvest cycle. In the quadrennial cycle, the highest yield may have been given by *S. viminalis*, cultivar Zubr, but the yield from P. max. × P. trich. Hybryda275 was practically the same. A high yield from

annual willow shoots was also found in a different study [36]. The highest wood yield (over 9 Mg ha⁻¹ y⁻¹ DM) in that experiment was for S. purpurea \times S. daphnoides hybrids. The dry biomass yield of *S. alba* and *S. viminalis* was high (over 13 Mg ha⁻¹ y⁻¹ DM) [37], whereas for S. dasyclados it was 11 Mg ha⁻¹ y⁻¹ DM [38]. If the wood content of those annual shoots was taken to be approx. 63%, its yield would be 7–8 Mg ha⁻¹ y⁻¹ DM. The biomass yield from poplar, willow, and black locust from three quadrennial consecutive harvest cycles, depending on the various soil enriching options, was 9.1, 8.5, and 4.8 Mg ha⁻¹ y⁻¹ DM [8]. If the wood content of those quadrennial SRWC shoots was taken to be approx. 79%, its yield would be 7.2, 6.7, and 3.8 Mg ha⁻¹ y⁻¹ DM, respectively. With a similar assumption of the wood content of the SRWC biomass, the wood yield from four clones of *P. balsamifera* from two quadrennial harvest cycles would be 5.5 Mg ha⁻¹ y⁻¹ DM, with it being significantly higher in the second harvest cycle—8.0 Mg ha⁻¹ y⁻¹ DM [39]. Therefore, the cited potential levels of the wood yield were similar to those in the current study for the poplar genotypes of the same species. Further, other data show that the potential wood yield from various poplar species harvested in various cycles would be approx. $12-15 \text{ Mg ha}^{-1} \text{ y}^{-1} \text{ DM } [40]; 9-14 \text{ Mg ha}^{-1} \text{ y}^{-1} \text{ DM } [41,42].$ Lower levels of a potential poplar wood yield (4–8 Mg ha⁻¹ y⁻¹ DM) were achieved in different studies [7,43]. In North America, the potential wood yield of different willow species harvested in different cycles varied within a range of 8 to 11 Mg ha⁻¹ y⁻¹ DM [44–46], and within a potentially wide range of 2 to 18 Mg ha⁻¹ y⁻¹ DM in Europe [47–49]. The potential wood yield from black locust would range from 2 to 8 Mg ha⁻¹ y⁻¹ DM [50–53]. Therefore, although the SRWC wood yield obtained in this study was not the highest, it was satisfactory and comparable to the literature reports.

The energy value of SRWC wood, as determined in this study, is primarily dependent on the wood yield. This is why the energy value ranges from 53 to 244 GJ ha⁻¹ y⁻¹ for P. balsamifera UWM3 in the annual harvest cycle and S. viminalis, cultivar Zubr, in the quadrennial harvest cycle. In general, higher values were found for all the SRWC genotypes in the quadrennial (mean: 162 GJ ha⁻¹ y⁻¹) than in the annual cycle (102 GJ ha⁻¹ y⁻¹). The energy value of willow wood in annual cycles was 136 GJ ha⁻¹ y⁻¹ and ranged from 93 to 200 GJ ha⁻¹ y⁻¹ depending on the genotype [36]. The cited range was close to that found in this study for willow genotypes harvested in the annual cycles (96 to 173 GJ ha⁻¹ y⁻¹). These values apply to the energy value of wood alone (without bark), which is why they should be regarded as satisfactory and high. The research in this regard usually provides the energy value of the whole SRWC biomass (wood + bark). This shows that when biomass is harvested as an energy feedstock, it is obtained as a whole, without being separated into fractions, i.e., bark and wood. For example, the whole poplar, willow, and black locust biomass energy values from three consecutive quadrennial harvest cycles were 142, 137, and 81 GJ ha⁻¹ y⁻¹, respectively [8]. If the wood content of biomass of the four-year SRWC shoots was taken to be 79%, then its potential energy value would be 112 GJ ha⁻¹ y⁻¹ for poplar, 108 GJ ha⁻¹ y⁻¹ for willow, and 64 GJ ha⁻¹ y⁻¹ for black locust. Considering this percentage share of wood in poplar biomass, the energy value of the four clones of P. *balsamifera* in two quadrennial harvest cycles would be 91 GJ ha⁻¹ y⁻¹, and it would be 135 GJ ha⁻¹ y⁻¹ in the second harvest cycle [39]. Assuming similar wood content, poplar biomass in a quadrennial cycle has an energy value nearly half as low [54]. Further, a potentially higher wood energy value (202–213 GJ ha⁻¹ y⁻¹) was achieved by growing poplar in various harvest cycles, with fertilization and watering [55,56]. A high energy value of black locust biomass (190 GJ ha⁻¹ y⁻¹), which would be equivalent to approx. 150 GJ ha⁻¹ y⁻¹ of the energy value of wood alone, was achieved in the six-year harvest cycle [57]. This would be potentially twice as high compared with the mean value for R. pseudoacacia in our experiment. The significant variations in energy values of SRWC wood and the entire biomass (wood + bark) are due to several factors, such as the selection of species and genotypes, the harvest cycle, soil and climate conditions, and the agrotechnical procedures used in the production process. These factors affect the biomass yield and the energy stored in it.

4.2. SRWC Wood Characteristics

Considering SRWC wood as a potential energy feedstock, one should note its thermophysical characteristics and elemental composition. The moisture content is a basic parameter that immediately impacts the calorific value. In general, *R. pseudoacacia* contained the significantly lowest amounts of moisture (a mean of 31.6%) compared with willow (a mean of 48.8%) and poplar, which contained the highest amounts of moisture (a mean of 54.9%). The moisture content of the whole biomass (wood + bark), as measured in a different study in three consecutive quadrennial harvest cycles (depending on various soil enrichment options), was also the significantly lowest (a mean of 38.9%) [58]. The willow and poplar biomass moisture content was also higher (49.8% and 56.5%, respectively). A lower moisture content (approx. 40%) of *R. pseudoacacia* was also found in a different study [59], compared with willow biomass (approx. 50%) [60,61] and poplar (50–60%) [39,40,62].

Therefore, the low moisture content of black locust wood was a positive characteristic. However, this wood's characteristics were worse in terms of the content of sulfur (0.033% DM), nitrogen (0.38% DM), and ash (0.69% DM) compared with the values for poplar and willow wood, for which these values were lower. It should be emphasized that nitrogen and sulfur in solid biofuel are not desirable, as they cause higher NOx and SO_2 emissions during combustion, and the ash can cause technological problems. Low levels of sulfur (0.031% DM), nitrogen (0.32% DM), and ash (0.59% DM) were also found in the wood of ten genotypes of willow harvested in annual cycles [36]. However, the nitrogen and sulfur content of willow was even lower in this study. Moreover, willow wood contained the lowest levels of nitrogen and ash in this study. The biomass of willow exhibited the lowest ash content with 1.25% dry matter, and the values were higher by 12% and 34%, respectively, for black locust and poplar in a different study in which the whole biomass (wood + bark) was analyzed [58]. These ash content levels were much higher compared with those in pure wood, as determined in whole SRWC biomass (wood + bark). The bark is known to contain more ash than wood, not only in SRWC but also in forest dendromass [63]. The ash content of biomass can vary considerably (1-3% DM), depending on the genotype and soil [62,64,65]. It is similar to poplar (0.98–3.12% DM) [40,61,62] and black locust (0.17–3.3% DM) [66]. Despite the large fluctuations in SRWC biomass ash content, it is typically lower than that of straw, semi-woody, or palm kernel shell biomass [67,68]. It is important to note that less ash in solid biofuel is more energy-efficient. However, different installations (e.g., depending on power output and technology used) have different expectations regarding ash content. For example, for small automatic pellet-fueled boilers (up to 10–30 kW) that generate heat for single-family homes, the ash content is expected to be below 1% DM. On the other hand, in the ISO standard for wood pellets of the highest class A1, the ash content should be $\leq 0.7\%$ DM, and in class A2 it should be $\leq 1.2\%$ DM. However, larger local woodchip-fueled installations have a considerably higher tolerance for ash content. For the use of woody biomass for energy purposes under Polish conditions, when ordering woodchip supplies, the ash content is expected to be below 3% DM and sometimes below 5% DM. Thus, the results of the present study showed that the debarked wood of all SRWC genotypes met the highest expectations in terms of ash content.

The lignocellulosic composition of SRWC wood should be noted when analyzing its potential usability as a raw material for integrated biorefineries or for the paper industry. Cellulose is widely used, and its highest level in this study was found in willow wood (51.8% DM), followed by poplar (50.0% DM), and the lowest levels of this biopolymer were found in wood of *R. pseudoacacia* (45.3% DM). Moreover, it was found in other studies [16] that willow wood contained higher levels of cellulose (56.0% DM) compared with poplar (51.0% DM) and black locust (52.0% DM). This biopolymer content of the SRWC species, as reported in that study, was higher by several pp compared with the mean values determined in this study, and this was particularly visible in *R. pseudoacacia*. Moreover, the cellulose content of pure SRWC wood was higher by several pp compared with its content of whole biomass (wood + bark). This is also reflected in the cellulose content of triennial

willow shoots, whose mean level was 44.4% DM [69]. A study by Przybysz et al. [70] found that 'Hybryda275' poplar had a higher cellulose content (52.4% DM) in older wood, which is consistent with the levels found in this study for this genotype during the quadrennial harvest cycle. Quite diverse, but also often very high cellulose content (54–59%) and low lignin content (17–22%) in willow biomass were reported by Baker et al. [71]. A similar lignin content (16–22%) in S. *viminalis* biomass was also determined by Gao et al. [72]. In contrast, a significantly higher lignin content (27-32% DM) in SRWC biomass was found in studies conducted in Northern Ireland and Canada [73]. However, in the present study, SRWC wood was characterized by significantly lower lignin contents. Of these, willow wood had the highest average lignin content, but it averaged only 15.8% DM, and its lowest amounts (15.0% DM) were found in wood of R. pseudoacacia. Lignin content was also determined, where a higher level was found in the wood of black locust (17.0% DM) than in willow (16.3% DM) and the highest in poplar wood (18.3% DM) [16]. Moreover, those values were higher by several pp than those determined in this study. A similar lignin content (18.0% DM) was found in the wood of the Hybryda275 poplar [70]. It was much higher in triennial willow shoots (25% DM) [69]. The R. pseudoacacia wood analyzed in the current study contained the highest mean level of hemicellulose (27.3% DM), and its lowest amounts (21.8% DM) were found in the wood of the willow genotypes. In a separate study conducted by Stolarski et al. [16], hemicellulose content was analyzed in various types of wood. The cited study found that black locust had the highest hemicellulose content (23.5% DM) compared to poplar (20.1% DM) and willow (19.2% DM). However, these values were still lower than the hemicellulose content found in the wood analyzed in the current study. The hemicellulose content of pure SRWC wood, as determined in that study, was higher by several pp compared with its content of whole biomass (wood + bark). A higher hemicellulose content (over 26% DM) was found in triennial willow biomass [69], as well as in poplar [70]. This value was similar to the annual harvest cycle of black locust, certain poplar genotypes, and some willow genotypes in the current study.

Both this study and the literature reports on SRWC biomass yield and quality show that most of the species and genotypes under study can find diverse applications because of varied amounts and quality of the biofeedstock (wood, bark, or wood and bark together) and because of the requirements and expectations of a specific industry branch. Willow biomass, including mainly *S. viminalis* genotypes harvested in longer (e.g., quadrennial) harvest cycles, seems to be an interesting feedstock for the energy industry because of the biomass quality, yield, and energy value. Meanwhile, the bark of *P. balsamifera* UWM2 proved to be an interesting source of bioactive substances, containing their highest total concentration [26]. The current study has shown that willow and poplar wood could be interesting for the energy and paper industry and integrated biorefineries because of their high cellulose content. The black locust contained the highest hemicellulose level, but the wood yield from this genotype was much lower than from most willow and poplar genotypes. However, it should also be noted that the choice of debarked wood as feedstock may also depend on the processes such as combustion, pyrolysis, or gasification.

5. Conclusions

The current study has found that the wood yield and its energy value were significantly influenced by the genotype, even within the same SRWC genus. The harvest cycle also played an important role, with longer cycles resulting in higher values for these parameters. The *Salix* genus had the highest dry wood yield and energy value, with *S. viminalis*, cultivar Żubr, producing the most in both the annual and quadrennial harvest cycles. The best effect for genus *Populus* was achieved for *P. maximowiczii* \times *P. trichocarpa* Hybryda275, and it was particularly marked in the quadrennial harvest cycle. The poorest results were determined for *R. pseudoacacia*. When it comes to the energy-related characteristics, those of *R. pseudoacacia* included the significantly lowest moisture content, which was a positive attribute, but, on the other hand, it had some adverse characteristics—the highest levels of

nitrogen, sulfur, and ash. More beneficial properties in this respect were determined for willow and poplar wood. Moreover, willow and poplar wood contained more cellulose compared with black locust. However, it is important to conduct more research in order to assess the economic and environmental viability of producing bioenergy and different bioproducts using SRWC wood, bark, or a combination of both as feedstock. This will provide a complete evaluation of the practical usability of these materials for various purposes. It is also worth noting that the production of SRWC biomass in a quadrennial harvest cycle can be challenging for farmers, as the waiting time for the first revenues from this type of production is much longer compared with the production viable, long-term contracts must be signed between farmers and bioenergy facilities that use this solid biofuel for energy purposes. Such contracts serve as a guarantee for the farmer that their investment in SRWC biomass production will bring the intended benefits. Additionally, bioenergy facilities would have a stable supply of good-quality energy feedstock.

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Conflicts of Interest: The authors declare no conflicts of interest.

Appendix A

Table A1. Dry wood yield for SRWC genotypes and harvest cycles (Mg ha⁻¹ y⁻¹ DM).

Canatura	Harves	Maara	
Genotype	Annual	Quadrennial	Mean
R. pseudoacacia	2.90 ± 0.06 mn	4.82 ± 0.38 hijklm	3.86 ± 0.46 J
P. nigra $ imes$ P. maximowiczii Max-5	4.61 ± 0.33 ijklmn	8.46 ± 0.60 cdefg	6.53 ± 0.91 DEF
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	$5.80\pm0.14~ m ghijk$	$11.15\pm0.78~ m{ab}$	$8.48 \pm 1.25~\mathrm{B}$
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	4.46 ± 0.04 jklmn	$9.13\pm0.30~\mathrm{cd}$	$6.79\pm1.05~\mathrm{DE}$
P. balsamifera UWM2	3.15 ± 0.07 lmn	$7.01\pm0.38~\mathrm{efghi}$	$5.08\pm0.88~\mathrm{HIJ}$
P. balsamifera UWM3	2.74 ± 0.04 n	5.68 ± 0.39 ghijk	$4.21\pm0.68~\mathrm{IJ}$
S. alba UWM200	5.03 ± 0.32 hijkl	6.29 ± 0.02 ghijk	5.66 ± 0.32 EFG
S. alba UWM095	6.53 ± 0.28 fghij	10.21 ± 0.55 bc	$8.37\pm0.87~\mathrm{B}$
S. dasyclados UWM155	5.40 ± 0.35 hijk	$7.47\pm0.29~\mathrm{defgh}$	$6.44\pm0.51~\mathrm{DEF}$
S. fragilis UWM195	6.74 ± 0.35 fghij	$9.62\pm0.47\mathrm{bc}$	$8.18\pm0.69~\mathrm{BC}$
S. pentandra UWM035	5.35 ± 0.42 hijk	$8.76\pm0.17~\mathrm{cde}$	$7.06\pm0.79~\mathrm{CD}$
S. triandra UWM198	6.53 ± 0.14 fghij	8.63 ± 0.33 cdef	$7.58\pm0.50~\mathrm{BCD}$
S. viminalis Żubr	8.89 ± 0.45 cde	12.42 ± 0.34 a	$10.65\pm0.83~\mathrm{A}$
S. viminalis $ imes$ S. purpurea UWM033	4.98 ± 0.12 hijkl	5.79 ± 0.43 ghijk	$5.38\pm0.27\mathrm{GHI}$
Mean	$5.22\pm0.26~\mathrm{Y}$	$8.25\pm0.34~\mathrm{X}$	6.73 ± 0.27

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

	Harves		
Genotype –	Annual	Quadrennial	- Mean
R. pseudoacacia	$56.19\pm1.25\mathrm{no}$	93.19 ± 7.31 jklmn	$74.69\pm8.91~\mathrm{I}$
P. nigra \times P. maximowiczii Max-5	$91.41\pm 6.50~\mathrm{klmno}$	167.97 ± 11.87 cdef	$129.69\pm18.16~\text{DE}$
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	113.06 ± 2.71 hijkl	$216.05\pm15.20~\mathrm{ab}$	$164.56\pm24.04~\mathrm{B}$
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	87.11 ± 0.79 lmno	$177.59\pm5.88~\mathrm{bcd}$	$132.35\pm20.41~\text{DE}$
P. balsamifera UWM2	61.54 ± 1.37 mno	$137.37\pm7.47~\mathrm{efgh}$	$99.46\pm17.29\mathrm{GH}$
P. balsamifera UWM3	$53.38\pm0.80~\mathrm{o}$	112.33 ± 7.79 hijkl	$82.85\pm13.64~\mathrm{HI}$
S. alba UWM200	98.28 ± 6.29 jklm	122.69 ± 0.34 hijkl	$110.49\pm6.14~\text{EFG}$
S. alba UWM095	127.28 ± 5.52 hijk	$202.88 \pm 11.03 \text{ bc}$	$165.08 \pm 17.78 \; \mathrm{B}$
S. dasyclados UWM155	104.75 ± 6.83 ijkl	$146.26\pm5.74~\mathrm{defg}$	$125.51\pm10.10~\mathrm{DEF}$
S. fragilis UWM195	131.28 ± 6.84 ghji	$188.57\pm9.26~\mathrm{bc}$	$159.92 \pm 13.81 \text{ BC}$
S. pentandra UWM035	106.32 ± 8.29 ijkl	172.99 ± 3.39 cde	$139.65 \pm 15.44 \text{ CD}$
S. triandra UWM198	126.37 ± 2.71 hijk	$168.60\pm 6.45~\mathrm{cdef}$	$147.48\pm9.95~\mathrm{BCD}$
S. viminalis Żubr	173.43 ± 8.82 cde	$244.34\pm6.78~\mathrm{a}$	$208.88\pm16.62~\mathrm{A}$
S. viminalis $ imes$ S. purpurea UWM033	$96.22\pm2.39~\mathrm{jklm}$	113.89 \pm 8.55 hijkl	$105.06\pm5.6~\text{FGH}$
Mean	$101.9\pm5.04~\mathrm{Y}$	$161.77\pm6.78~\mathrm{X}$	131.83 ± 5.33

Table A2. Dry wood energy value for SRWC genotypes and harvest cycles (GJ $ha^{-1} y^{-1}$).

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

Table A3. Coal equivalent of wood energy value for SRWC genotypes and harvest cycles $(Mg ha^{-1} y^{-1})$.

Caratan	Harve	st Cycle		
Genotype	Annual	Quadrennial	Mean	
R. pseudoacacia	2.25 ± 0.05	3.73 ± 0.29	2.99 ± 0.36	
P. nigra \times P. maximowiczii Max-5	3.66 ± 0.26	6.72 ± 0.47	5.19 ± 0.73	
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	4.52 ± 0.11	8.64 ± 0.61	6.58 ± 0.96	
<i>P. max.</i> \times <i>P. trich.</i> Androscoggin	3.48 ± 0.03	7.10 ± 0.24	5.29 ± 0.82	
P. balsamifera UWM2	2.46 ± 0.05	5.49 ± 0.30	3.98 ± 0.69	
P. balsamifera UWM3	2.14 ± 0.03	4.49 ± 0.31	3.31 ± 0.55	
S. alba UWM200	3.93 ± 0.25	4.91 ± 0.01	4.42 ± 0.25	
S. alba UWM095	5.09 ± 0.22	8.12 ± 0.44	6.60 ± 0.71	
S. dasyclados UWM155	4.19 ± 0.27	5.85 ± 0.23	5.02 ± 0.40	
S. fragilis UWM195	5.25 ± 0.27	7.54 ± 0.37	6.40 ± 0.55	
S. pentandra UWM035	4.25 ± 0.33	6.92 ± 0.14	5.59 ± 0.62	
S. triandra UWM198	5.05 ± 0.11	6.74 ± 0.26	5.90 ± 0.40	
S. viminalis Żubr	6.94 ± 0.35	9.77 ± 0.27	8.36 ± 0.66	
S. viminalis \times S. purpurea UWM033	3.85 ± 0.10	4.56 ± 0.34	4.20 ± 0.22	
Mean	4.08 ± 0.2	6.47 ± 0.27	5.27 ± 0.21	

Item	Dry Wood Yield	Dry Wood Energy Value	Coal Equivalent	Moisture	нну	Ash	FC	VM	С	н	N	S	Cl	Cold Water Extracts	Hot Water Extracts	Other Soluble Substances	Hemicellulose	Cellulose	Lignin
Dry wood yield	1.00	1.00 *	1.00 *	0.03	0.20	-0.57 *	-0.38 *	0.41 *	0.18	-0.09	-0.70 *	-0.55 *	0.06	-0.72 *	-0.61 *	-0.21	-0.65 *	0.70 *	0.24 *
Dry wood energy value		1.00	1.00 *	0.03	0.22 *	-0.57*	-0.37*	0.41 *	0.19	-0.10	-0.70 *	-0.55 *	0.06	-0.73 *	-0.61 *	-0.21	-0.66 *	0.71 *	0.25 *
Coal equivalent Moisture HHV Ash FC VM C H N S Cl Cold water extracts Hot water extracts Other soluble substances Hemicellulose Cellulose Lignin			1.00	0.03 1.00	0.22 * 0.11 1.00	-0.57 * 0.01 -0.09 1.00	$-0.37 * -0.19 \\ 0.03 \\ 0.33 * \\ 1.00$	$0.41 * 0.20 \\ 0.04 \\ -0.39 * \\ -0.96 * \\ 1.00$	$\begin{array}{c} 0.19 \\ -0.01 \\ 0.49 * \\ -0.24 * \\ 0.02 \\ 0.00 \\ 1.00 \end{array}$	$\begin{array}{c} -0.10\\ 0.03\\ -0.13\\ -0.14\\ -0.02\\ 0.05\\ -0.05\\ 1.00\end{array}$	$\begin{array}{c} -0.70\ ^*\\ -0.10\\ -0.26\ ^*\\ 0.54\ ^*\\ 0.57\ ^*\\ -0.55\ ^*\\ -0.43\ ^*\\ 0.12\\ 1.00\end{array}$	$\begin{array}{c} -0.55 \\ -0.36 \\ -0.28 \\ 0.51 \\ \end{array} \\ \begin{array}{c} 0.22 \\ -0.25 \\ -0.40 \\ 0.18 \\ 0.69 \\ \end{array} \\ \begin{array}{c} 1.00 \end{array}$	$\begin{array}{c} 0.06 \\ -0.03 \\ -0.24 * \\ 0.17 \\ 0.17 \\ -0.24 * \\ -0.20 \\ 0.01 \\ 0.05 \\ 1.00 \end{array}$	$\begin{array}{c} -0.73 \\ 0.13 \\ -0.34 \\ 0.52 \\ * \\ 0.36 \\ * \\ -0.28 \\ 0.09 \\ 0.78 \\ 0.09 \\ 0.78 \\ * \\ 0.55 \\ * \\ -0.02 \\ 1.00 \\ \end{array}$	$\begin{array}{c} -0.61 * \\ -0.05 \\ -0.26 * \\ 0.48 * \\ -0.39 * \\ -0.33 * \\ 0.08 \\ 0.76 * \\ 0.58 * \\ 0.05 \\ 0.89 * \\ 1.00 \end{array}$	$\begin{array}{c} -0.21 \\ 0.07 \\ -0.08 \\ 0.28 * \\ 0.14 \\ -0.17 \\ -0.20 \\ -0.08 \\ 0.35 * \\ 0.29 * \\ 0.20 \\ 0.48 * \\ 0.59 * \\ 1.00 \end{array}$	$\begin{array}{c} -0.66 \\ -0.05 \\ -0.45 \\ 0.52 \\ * \\ 0.32 \\ * \\ -0.35 \\ * \\ -0.48 \\ * \\ 0.07 \\ 0.75 \\ * \\ 0.54 \\ * \\ -0.02 \\ 0.66 \\ * \\ 0.53 \\ * \\ 0.02 \\ 1.00 \end{array}$	$\begin{array}{c} 0.71 * \\ 0.04 \\ 0.32 * \\ -0.57 * \\ -0.53 * \\ 0.54 * \\ 0.54 * \\ 0.46 * \\ -0.13 \\ -0.91 * \\ -0.62 * \\ -0.13 \\ -0.85 * \\ -0.85 * \\ -0.85 * \\ -0.51 * \\ -0.78 * \\ 1.00 \end{array}$	$\begin{array}{c} 0.25 \\ 0.00 \\ 0.41 \\ -0.26 \\ * \\ 0.27 \\ 0.23 \\ 0.18 \\ -0.14 \\ -0.27 \\ 0.19 \\ -0.39 \\ -0.35 \\ * \\ -0.08 \\ -0.50 \\ * \\ 0.18 \\ 1.00 \end{array}$

Table A4. The Pearson correlation coefficients for the analyzed features in the SRWC wood.

* Significant values (p < 0.05).

Annual 20.01 ± 0.07 a 19.92 ± 0.27 ab 19.22 ± 0.17 abcde 19.12 ± 0.23 abcde	Quadrennial 19.91 ± 0.28 ab 19.89 ± 0.03 ab 17.96 ± 0.15 fg	19.96 ± 0.13 A 19.90 ± 0.12 A
20.01 ± 0.07 a 19.92 ± 0.27 ab 19.22 ± 0.17 abcde 10.12 ± 0.23 abcde	19.91 ± 0.28 ab 19.89 ± 0.03 ab 17.96 ± 0.15 for	$19.96 \pm 0.13 \text{ A} \\ 19.90 \pm 0.12 \text{ A} \\ 19.50 \pm 0.20 \text{ CDF}$
18.13 ± 0.23 abcde 18.98 ± 0.29 abcdef 18.89 ± 0.15 bcdef 18.74 ± 0.08 cdefg 19.54 ± 0.38 abc 19.38 ± 0.44 abcd 18.69 ± 0.23 cdefg	$18.54 \pm 0.01 \text{ cdefg} \\ 18.54 \pm 0.01 \text{ cdefg} \\ 18.39 \pm 0.05 \text{ defg} \\ 18.48 \pm 0.05 \text{ cdefg} \\ 19.14 \pm 0.03 \text{ abcde} \\ 19.02 \pm 0.21 \text{ abcdef} \\ 18.34 \pm 0.04 \text{ defg} \\ 18.22 \pm 0.15 \text{ efg} \\ 18.24 \pm$	$18.39 \pm 0.30 \text{ CDE} \\18.84 \pm 0.17 \text{ BCDE} \\18.68 \pm 0.19 \text{ BCDE} \\18.68 \pm 0.12 \text{ BCDE} \\18.94 \pm 0.10 \text{ BCD} \\19.28 \pm 0.23 \text{ AB} \\18.86 \pm 0.30 \text{ BCDE} \\18.46 \pm 0.16 \text{ DE} \\18.46 \pm 0.16 DE$
$\begin{array}{c} 18.50 \pm 0.23 \text{cdefg} \\ 18.50 \pm 0.37 \text{cdefg} \\ 18.80 \pm 0.21 \text{cdef} \\ 19.19 \pm 0.11 \text{abcde} \\ 19.49 \pm 0.16 \text{abc} \\ \end{array}$	$\begin{array}{c} 19.17 \pm 0.07 {\rm obs} \\ 19.17 \pm 0.07 {\rm abcde} \\ 17.67 \pm 0.02 {\rm g} \\ 18.68 \pm 0.03 {\rm cdefg} \\ 18.85 \pm 0.08 {\rm bcdef} \end{array}$	$18.84 \pm 0.23 \text{ BCDE} \\ 18.24 \pm 0.27 \text{ D} \\ 18.94 \pm 0.13 \text{ BCD} \\ 19.17 \pm 0.16 \text{ BC} \\ 10.06 \pm 0.077 \\ 10.075 \\ $
	$\begin{array}{l} 19.13 \pm 0.23 \text{ abcde} \\ 19.13 \pm 0.23 \text{ abcde} \\ 18.98 \pm 0.29 \text{ abcdef} \\ 18.98 \pm 0.29 \text{ abcdef} \\ 18.74 \pm 0.08 \text{ cdefg} \\ 19.54 \pm 0.38 \text{ abc} \\ 19.38 \pm 0.44 \text{ abcd} \\ 18.69 \pm 0.23 \text{ cdefg} \\ 18.50 \pm 0.37 \text{ cdefg} \\ 18.50 \pm 0.37 \text{ cdefg} \\ 18.80 \pm 0.21 \text{ cdef} \\ 19.19 \pm 0.11 \text{ abcde} \\ 19.49 \pm 0.16 \text{ abc} \\ \hline 19.18 \pm 0.09 \text{ X} \end{array}$	19.13 \pm 0.23 abcde17.96 \pm 0.01 cdefg19.13 \pm 0.23 abcde18.54 \pm 0.01 cdefg18.98 \pm 0.29 abcdef18.54 \pm 0.05 cdefg18.94 \pm 0.08 cdefg19.14 \pm 0.03 abcde19.54 \pm 0.38 abc19.02 \pm 0.21 abcdef19.38 \pm 0.44 abcd18.34 \pm 0.04 defg18.69 \pm 0.23 cdefg19.17 \pm 0.07 abcde18.80 \pm 0.21 cdef17.67 \pm 0.02 g19.19 \pm 0.11 abcde18.68 \pm 0.03 cdefg19.19 \pm 0.16 abc18.87 \pm 0.08 bcdef

Table A5. Fixed carbon content in wood for SRWC genotypes and harvest cycles (% DM).

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

Table A6. Volatile matter content in wood for SRWC genotypes and harvest cycles (% DM).

Construe	Harves	Maan	
Genotype	Annual	Quadrennial	Mean
R. pseudoacacia P. nigra × P. maximowiczii Max-5 P. max. × P. trich. Hybryda275 P. max. × P. trich. Androscoggin P. balsamifera UWM2 P. balsamifera UWM3 S. alba UWM200 S. alba UWM200 S. alba UWM095 S. dasyclados UWM155 S. fragilis UWM195 S. pentandra UWM035 S. triandra UWM198	$79.22 \pm 0.06 h$ $79.92 \pm 0.52 defgh$ $80.16 \pm 0.20 cdefgh$ $80.23 \pm 0.25 cdefgh$ $80.37 \pm 0.30 bcdefgh$ $80.46 \pm 0.14 bcdefgh$ $80.65 \pm 0.10 abcdefg$ $79.91 \pm 0.38 defgh$ $80.66 \pm 0.46 cdefgh$ $80.67 \pm 0.24 abcdefg$ $80.86 \pm 0.39 abcdef$ $80.50 \pm 0.24 abcdefg$	$\begin{array}{l} 79.52 \pm 0.28 \ \text{efgh} \\ 79.50 \pm 0.01 \ \text{gh} \\ 81.55 \pm 0.15 \ \text{ab} \\ 80.95 \pm 0.01 \ \text{abcdef} \\ 81.02 \pm 0.05 \ \text{abcdef} \\ 80.28 \pm 0.03 \ \text{cdefgh} \\ 80.49 \pm 0.21 \ \text{abcdefg} \\ 81.14 \pm 0.04 \ \text{abcd} \\ 81.21 \pm 0.15 \ \text{abc} \\ 80.35 \pm 0.07 \ \text{bcdefgh} \\ 81.70 \pm 0.02 \ \text{abcdefgh} \\ 81.70 \pm 0.02 \ \text{abcdefgh} \\ \end{array}$	$\begin{array}{c} 79.37 \pm 0.14 \ \mathrm{D} \\ 79.71 \pm 0.25 \ \mathrm{CD} \\ 80.85 \pm 0.33 \ \mathrm{AB} \\ 80.59 \pm 0.20 \ \mathrm{AB} \\ 80.70 \pm 0.20 \ \mathrm{AB} \\ 80.65 \pm 0.11 \ \mathrm{AB} \\ 80.47 \pm 0.10 \ \mathrm{ABC} \\ 80.20 \pm 0.23 \ \mathrm{BC} \\ 80.60 \pm 0.32 \ \mathrm{AB} \\ 80.94 \pm 0.18 \ \mathrm{AB} \\ 80.61 \pm 0.21 \ \mathrm{AB} \\ 81.10 \pm 0.29 \ \mathrm{AB} \\ 81.10 \pm 0.29 \ \mathrm{AB} \end{array}$
S. viminalis Zubr S. viminalis \times S. purpurea UWM033	80.17 ± 0.15 cdergn 79.80 \pm 0.19 efgh	80.74 ± 0.03 abcdefg 80.55 ± 0.08 abcdefg	80.45 ± 0.14 ABC 80.17 ± 0.19 BC
Mean	$80.21\pm0.09~\mathrm{Y}$	$80.70\pm0.10~\mathrm{X}$	80.46 ± 0.07

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

Table A7. Hydrogen content in wood for SRWC genotypes and harvest cycles (% DM).

Conotyna	Harves	Maar		
Genotype	Annual	Quadrennial	Wiedii	
R. pseudoacacia	6.42 ± 0.04	6.44 ± 0.01	6.43 ± 0.02	
P. nigra $ imes$ P. maximowiczii Max-5	6.37 ± 0.06	6.40 ± 0.02	6.39 ± 0.03	
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	6.41 ± 0.03	6.54 ± 0.02	6.48 ± 0.03	
<i>P. max.</i> \times <i>P. trich.</i> And roscoggin	6.44 ± 0.04	6.51 ± 0.01	6.47 ± 0.02	
P. balsamifera UWM2	6.47 ± 0.01	6.36 ± 0.01	6.41 ± 0.03	
P. balsamifera UWM3	6.51 ± 0.10	6.34 ± 0.01	6.42 ± 0.06	
S. alba UWM200	6.52 ± 0.10	6.49 ± 0.02	6.50 ± 0.04	
S. alba UWM095	6.53 ± 0.03	6.52 ± 0.01	6.52 ± 0.01	
S. dasyclados UWM155	6.52 ± 0.04	6.50 ± 0.02	6.51 ± 0.02	
S. fragilis UWM195	6.41 ± 0.05	6.36 ± 0.02	6.38 ± 0.03	
S. pentandra UWM035	6.57 ± 0.09	6.35 ± 0.03	6.46 ± 0.06	
S. triandra UWM198	6.46 ± 0.04	6.34 ± 0.01	6.40 ± 0.03	
S. viminalis Żubr	6.47 ± 0.04	6.42 ± 0.02	6.45 ± 0.02	
S. viminalis \times S. purpurea UWM033	6.54 ± 0.09	6.43 ± 0.01	6.48 ± 0.05	
Mean	$6.47\pm0.02~\mathrm{X}$	$6.43\pm0.01~\mathrm{Y}$	6.45 ± 0.01	

X, Y—harvest cycle homogeneous groups; no letter indicates no significant differences, \pm —standard error of mean.

Construis	Harves	Maar	
Genotype	Annual	Quadrennial	- Iviean
R. pseudoacacia	0.017 ± 0.003 abc	0.016 ± 0.002 abc	$0.017 \pm 0.001 \text{ ABC}$
P. nigra $ imes$ P. maximowiczii Max-5	$0.015\pm0.002~\mathrm{abc}$	$0.019\pm0.001~\mathrm{ab}$	$0.017\pm0.001~\mathrm{ABC}$
<i>P. max.</i> \times <i>P. trich.</i> Hybryda275	$0.018 \pm 0.001 \text{ abc}$	$0.016\pm0.002~\mathrm{abc}$	$0.017\pm0.001~\mathrm{ABC}$
<i>P. max.</i> \times <i>P. trich.</i> And roscoggin	$0.018\pm0.001~\mathrm{abc}$	$0.014\pm0.002~\mathrm{abc}$	$0.016\pm0.001~\mathrm{ABC}$
P. balsamifera UWM2	$0.012 \pm 0.001 \ { m c}$	$0.014\pm0.001~\mathrm{abc}$	$0.013 \pm 0.001 \ { m C}$
P. balsamifera UWM3	$0.015\pm0.002~\mathrm{abc}$	$0.013\pm0.001~{ m bc}$	$0.014\pm0.001~\mathrm{BC}$
S. alba UWM200	$0.018 \pm 0.001 \text{ abc}$	$0.018\pm0.001~\mathrm{abc}$	$0.018\pm0.001~\mathrm{A}$
S. alba UWM095	$0.017\pm0.002~\mathrm{abc}$	$0.015\pm0.002~\mathrm{abc}$	$0.016\pm0.002~\mathrm{ABC}$
S. dasyclados UWM155	$0.017 \pm 0.001 \text{ abc}$	$0.014\pm0.002~\mathrm{abc}$	$0.015\pm0.001~\mathrm{ABC}$
S. fragilis UWM195	$0.019\pm0.001~\mathrm{ab}$	$0.019\pm0.001~\mathrm{abc}$	$0.019 \pm 0.001~{ m A}$
S. pentandra UWM035	$0.016 \pm 0.001 \text{ abc}$	$0.017\pm0.001~\mathrm{abc}$	$0.017\pm0.001~\mathrm{ABC}$
S. triandra UWM198	0.020 ± 0.001 a	$0.013\pm0.002~{ m bc}$	$0.017\pm0.002~\mathrm{ABC}$
S. viminalis Żubr	$0.018\pm0.002~\mathrm{abc}$	$0.018\pm0.001~\mathrm{abc}$	$0.018\pm0.001~\mathrm{AB}$
S. viminalis \times S. purpurea UWM033	0.020 ± 0.001 a	$0.018\pm0.001~\mathrm{abc}$	$0.019\pm0.001~\mathrm{A}$
Mean	$0.017 \pm 0.001 \text{ X}$	$0.016 \pm 0.001 ~{ m Y}$	0.017 ± 0.001

Table A8. Chlorine content in wood for SRWC genotypes and harvest cycles (% DM).

A, B, C—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

Table A9. Cold water extract content in wood for SRWC genotypes and harvest cycles (% DM).

Canakuna	Harves	Maar	
Genotype	Annual	Quadrennial	Iviean
R. pseudoacacia P. nigra × P. maximowiczii Max-5 P. max. × P. trich. Hybryda275 P. max. × P. trich. Androscoggin P. balsamifera UWM2 P. balsamifera UWM3 S. alba UWM200 S. alba UWM095 S. dasyclados UWM155 S. fragilis UWM195 S. pentandra UWM035	$\begin{array}{c} 4.03 \pm 0.24 \mathrm{cdef} \\ 4.78 \pm 0.14 \mathrm{bc} \\ 3.79 \pm 0.01 \mathrm{def} \\ 3.98 \pm 0.01 \mathrm{cdef} \\ 5.28 \pm 0.17 \mathrm{ab} \\ 5.73 \pm 0.05 \mathrm{a} \\ 2.50 \pm 0.04 \mathrm{ijkl} \\ 4.14 \pm 0.13 \mathrm{cde} \\ 4.11 \pm 0.09 \mathrm{cde} \\ 4.38 \pm 0.04 \mathrm{cd} \\ 3.08 \pm 0.03 \mathrm{ghij} \end{array}$	$\begin{array}{l} 4.04\pm 0.11\ {\rm cdef}\\ 1.64\pm 0.07\ {\rm mn}\\ 1.94\pm 0.23\ {\rm lmn}\\ 1.92\pm 0.02\ {\rm lmn}\\ 2.69\pm 0.17\ {\rm ijkl}\\ 3.51\pm 0.02\ {\rm efgh}\\ 3.27\pm 0.01\ {\rm fghi}\\ 2.50\pm 0.12\ {\rm ijkl}\\ 2.87\pm 0.07\ {\rm hijk}\\ 2.31\pm 0.06\ {\rm jklm}\\ 2.00\pm 0.04\ {\rm lmn}\\ \end{array}$	$\begin{array}{c} 4.03 \pm 0.12 \text{ B} \\ 3.21 \pm 0.71 \text{ DE} \\ 2.86 \pm 0.43 \text{ EF} \\ 2.95 \pm 0.46 \text{ EF} \\ 3.99 \pm 0.59 \text{ BC} \\ 4.62 \pm 0.50 \text{ A} \\ 2.89 \pm 0.17 \text{ EF} \\ 3.32 \pm 0.38 \text{ DE} \\ 3.49 \pm 0.28 \text{ CD} \\ 3.34 \pm 0.46 \text{ DE} \\ 2.54 \pm 0.24 \text{ F} \end{array}$
S. triandra UWM198 S. viminalis Żubr S. viminalis × S. purpurea UWM033 Mean	$ \begin{array}{r} 4.77 \pm 0.10 \text{bc} \\ 3.81 \pm 0.51 \text{def} \\ 3.91 \pm 0.08 \text{def} \\ \end{array} $	$2.24 \pm 0.09 \text{ klmn} \\ 1.44 \pm 0.12 \text{ n} \\ 1.98 \pm 0.22 \text{ lmn} \\ 2.45 \pm 0.12 \text{ Y}$	$3.51 \pm 0.57 \text{ CD} \\ 2.62 \pm 0.58 \text{ F} \\ 2.94 \pm 0.45 \text{ EF} \\ \hline 3.31 \pm 0.13 \\ \hline$

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

Table A10. Hot water extract content in wood for SRWC genotypes and harvest cycles (% DM).

Canatura	Harve	Maan	
Genotype	Annual	Quadrennial	- Iviean
R. pseudoacacia P. nigra × P. maximowiczii Max-5 P. max. × P. trich. Hybryda275 P. max. × P. trich. Androscoggin P. balsamifera UWM2 P. balsamifera UWM3 S. alba UWM200 S. alba UWM095 S. dasyclados UWM155 S. fragilis UWM195 S. prestandra UWM035	$\begin{array}{c} 8.32 \pm 0.22 \ {\rm bc} \\ 8.04 \pm 0.16 \ {\rm cd} \\ 5.46 \pm 0.03 \ {\rm ijk} \\ 5.78 \pm 0.13 \ {\rm hij} \\ 9.04 \pm 0.16 \ {\rm ab} \\ 9.42 \pm 0.21 \ {\rm a} \\ 4.93 \pm 0.04 \ {\rm lmn} \\ 7.27 \pm 0.12 \ {\rm def} \\ 6.72 \pm 0.34 \ {\rm efg} \\ 8.16 \pm 0.09 \ {\rm c} \\ 5.80 \pm 0.08 \ {\rm hij} \end{array}$	$\begin{array}{c} 7.29 \pm 0.06 \ \mathrm{de} \\ 4.11 \pm 0.02 \ \mathrm{no} \\ 4.38 \pm 0.05 \ \mathrm{mno} \\ 3.69 \pm 0.04 \ \mathrm{o} \\ 5.38 \pm 0.02 \ \mathrm{ijk} \\ 5.21 \pm 0.11 \ \mathrm{klm} \\ 6.15 \pm 0.21 \ \mathrm{ghi} \\ 6.43 \pm 0.06 \ \mathrm{fgh} \\ 4.69 \pm 0.44 \ \mathrm{lmn} \\ 5.02 \pm 0.02 \ \mathrm{klm} \\ 5.02 \pm 0.01 \ \mathrm{klm} \end{array}$	$\begin{array}{c} 7.80 \pm 0.25 \ \mathrm{A} \\ 6.08 \pm 0.88 \ \mathrm{DE} \\ 4.92 \pm 0.24 \ \mathrm{GH} \\ 4.73 \pm 0.47 \ \mathrm{H} \\ 7.21 \pm 0.82 \ \mathrm{B} \\ 7.31 \pm 0.95 \ \mathrm{AB} \\ 5.54 \pm 0.29 \ \mathrm{EF} \\ 6.85 \pm 0.20 \ \mathrm{BC} \\ 5.70 \pm 0.52 \ \mathrm{EF} \\ 6.59 \pm 0.70 \ \mathrm{CD} \\ 5.45 \pm 0.18 \ \mathrm{FG} \end{array}$
S. triandra UWM198 S. vininalis Żubr S. viminalis × S. purpurea UWM033	9.49 \pm 0.11 a 6.65 \pm 0.16 efg 6.32 \pm 0.01 gh	$\begin{array}{c} 4.62 \pm 0.15 \mathrm{lmn} \\ 4.11 \pm 0.03 \mathrm{no} \\ 4.78 \pm 0.05 \mathrm{lmn} \end{array}$	$7.06 \pm 1.09 \text{ BC} \\ 5.38 \pm 0.57 \text{ FG} \\ 5.55 \pm 0.35 \text{ EF} $
Mean	$7.24\pm0.23~\mathrm{X}$	$5.07\pm0.15~\mathrm{Y}$	6.16 ± 0.18

A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times harvest cycle interaction homogeneous groups; \pm —standard error of mean.

Genotype

R. pseudoacacia

P. nigra × P. maximowiczii Max-5

P. max. \times *P. trich.* Hybryda275

P. max. \times *P. trich.* Androscoggin

P. balsamifera UWM2

P. balsamifera UWM3

S. alba UWM200

S. alba UWM095

S. dasyclados UWM155

S. fragilis UWM195

S. pentandra UWM035

S. triandra UWM198

S. viminalis Żubr

 3.90 ± 0.02 ijklm

 $3.39\pm0.02\ mn$

 $5.39\pm0.08~cd$

 4.06 ± 0.11 hijkl

 $5.07\pm0.06~de$

 3.84 ± 0.04 jklmn

 $5.30\pm0.05\ d$

 $4.38\pm0.10~\text{ghi}$

 3.71 ± 0.19 jklmn

 3.72 ± 0.09 jklmn

Table A11. Other soluble substance content in wood for SRWC genotypes and harvest cycles (% DM).

S. viminalis \times S. purpurea UWM033	4.33 ± 0.02 hgij	3.62 ± 0.11 klmn	$3.98\pm0.17\text{DE}$		
Mean	$4.86\pm0.17~\mathrm{X}$	$4.15\pm0.10~\mathrm{Y}$	4.51 ± 0.11		
A, B, C, etc.—genotype homogeneous groups; X, Y—harvest cycle homogeneous groups; a, b, c, etc.—genotype \times					

harvest cycle interaction homogeneous groups; ±-standard error of mean.

References

1. Stolarski, M.J.; Warmiński, K.; Krzyżaniak, M.; Olba–Zięty, E.; Akincza, M. Bioenergy Technologies and Biomass Potential Vary in Northern European Countries. Renew. Sustain. Energy Rev. 2020, 133, 110238. [CrossRef]

 $4.38\pm0.08~\text{ghi}$

 $4.52\pm0.05~\text{fgh}$

 $5.01\pm0.11~def$

 3.76 ± 0.06 jklmn

 $5.84\pm0.16\ c$

 $4.41\pm0.13~\mathrm{gh}$

 $7.48\pm0.01~\text{a}$

 4.03 ± 0.03 hijkl

 $6.61\pm0.19\,b$

 $3.35\pm0.02\ n$

- 2. Ceotto, E.; Castelli, F.; Moschella, A.; Diozzi, M.; Di Candilo, M. Cattle Slurry Fertilization to Giant Reed (Arundo donax L.): Biomass Yield and Nitrogen Use Efficiency. Bioenergy Res. 2015, 8, 1252–1262. [CrossRef]
- 3. Ceotto, E.; Castelli, F.; Moschella, A.; Diozzi, M.; Di Candilo, M. Poplar Short Rotation Coppice Is Not a First Choice Crop for Cattle Slurry Fertilization: Biomass Yield and Nitrogen-Use Efficiency. Ind. Crops Prod. 2016, 85, 167–173. [CrossRef]
- Jankowski, K.J.; Dubis, B.; Sokólski, M.M.; Załuski, D.; Bórawski, P.; Szempliński, W. Biomass Yield and Energy Balance of 4. Virginia Fanpetals in Different Production Technologies in North-Eastern Poland. Energy 2019, 185, 612–623. [CrossRef]
- Jankowski, K.J.; Dubis, B.; Kozak, M. Sewage Sludge and the Energy Balance of Jerusalem Artichoke Production—A Case Study 5. in North-Eastern Poland. Energy 2021, 236, 121545. [CrossRef]
- Scordia, D.; Papazoglou, E.G.; Kotoula, D.; Sanz, M.; Ciria, C.S.; Pérez, J.; Maliarenko, O.; Prysiazhniuk, O.; von Cossel, M.; 6. Greiner, B.E.; et al. Towards Identifying Industrial Crop Types and Associated Agronomies to Improve Biomass Production from Marginal Lands in Europe. GCB Bioenergy 2022, 14, 710–734. [CrossRef]
- 7. Dillen, S.Y.; Djomo, S.N.; Al Afas, N.; Vanbeveren, S.; Ceulemans, R. Biomass Yield and Energy Balance of a Short-Rotation Poplar Coppice with Multiple Clones on Degraded Land during 16 Years. Biomass Bioenergy 2013, 56, 157–165. [CrossRef]
- Stolarski, M.J.; Stachowicz, P. Black Locust, Poplar or Willow? Yield and Energy Value in Three Consecutive Four-Year Harvest 8. Rotations. Ind. Crops Prod. 2023, 193, 116197. [CrossRef]
- Monti, A.; Zanetti, F.; Scordia, D.; Testa, G.; Cosentino, S.L. What to Harvest When? Autumn, Winter, Annual and Biennial 9. Harvesting of Giant Reed, Miscanthus and Switchgrass in Northern and Southern Mediterranean Area. Ind. Crops Prod. 2015, 75, 129–134. [CrossRef]
- 10. Amaducci, S.; Facciotto, G.; Bergante, S.; Perego, A.; Serra, P.; Ferrarini, A.; Chimento, C. Biomass Production and Energy Balance of Herbaceous and Woody Crops on Marginal Soils in the Po Valley. GCB Bioenergy 2017, 9, 31–45. [CrossRef]
- Von Cossel, M.; Lewandowski, I.; Elbersen, B.; Staritsky, I.; Van Eupen, M.; Iqbal, Y.; Mantel, S.; Scordia, D.; Testa, G.; Cosentino, 11. S.L.; et al. Marginal Agricultural Land Low-Input Systems for Biomass Production. Energies 2019, 12, 3123. [CrossRef]
- 12. Radzikowski, P.; Matyka, M.; Berbeć, A.K. Biodiversity of Weeds and Arthropods in Five Different Perennial Industrial Crops in Eastern Poland. Agriculture 2020, 10, 636. [CrossRef]
- Parajuli, R.; Knudsen, M.T.; Dalgaard, T. Multi-criteria Assessment of Yellow, Green, and Woody Biomasses: Pre-screening of 13. Potential Biomasses as Feedstocks for Biorefineries. Biofuels Bioprod. Biorefining 2015, 9, 545–566. [CrossRef]
- Parajuli, R.; Dalgaard, T.; Jørgensen, U.; Adamsen, A.P.S.; Knudsen, M.T.; Birkved, M.; Gylling, M.; Schjørring, J.K. Biorefining in 14. the Prevailing Energy and Materials Crisis: A Review of Sustainable Pathways for Biorefinery Value Chains and Sustainability Assessment Methodologies. Renew. Sustain. Energy Rev. 2015, 43, 244–263. [CrossRef]
- Oleszek, M.; Kowalska, I.; Oleszek, W. Phytochemicals in Bioenergy Crops. Phytochem. Rev. 2019, 18, 893–927. [CrossRef] 15.
- 16. Stolarski, M.J.; Warmiński, K.; Krzyżaniak, M.; Tyśkiewicz, K.; Olba-Zięty, E.; Graban, Ł.; Lajszner, W.; Załuski, D.; Wiejak, R.; Kamiński, P.; et al. How Does Extraction of Biologically Active Substances with Supercritical Carbon Dioxide Affect Lignocellulosic Biomass Properties? Wood Sci. Technol. 2020, 54, 519-546. [CrossRef]

 $4.14\pm0.11~\text{D}$

 $3.96\pm0.25\,\text{DE}$

 $5.20\pm0.11~\text{B}$

 $3.91\pm0.09~\text{DE}$

 $5.45\pm0.19~B$

 $4.12\pm0.14~\text{DE}$

 $6.39\pm0.49~A$

 $4.21\pm0.09~\text{D}$

 $5.16\pm0.66\ B$

 $3.53\pm0.09\ F$

- 17. Walter, M.; Brzozowski, B.; Adamczak, M. Effect of Supercritical Extract from Black Poplar and Basket Willow on the Quality of Natural and Probiotic Drinkable Yogurt. *Animals* **2021**, *11*, 2997. [CrossRef] [PubMed]
- Sulima, P.; Przyborowski, J.A. Purple Willow (*Salix purpurea* L.) and Its Potential Uses for the Treatment of Arthritis and Rheumatism. In *Bioactive Food as Dietary Interventions for Arthritis and Related Inflammatory Diseases*; Elsevier: Amsterdam, The Netherlands, 2019; pp. 535–551.
- Valette, N.; Perrot, T.; Sormani, R.; Gelhaye, E.; Morel-Rouhier, M. Antifungal Activities of Wood Extractives. *Fungal Biol. Rev.* 2017, 31, 113–123. [CrossRef]
- Montinari, M.R.; Minelli, S.; De Caterina, R. *The First* 3500 years of Aspirin History from Its Roots—A Concise Summary. *Vasc. Pharmacol.* 2019, 113, 1–8. [CrossRef]
- Sulima, P.; Krauze-Baranowska, M.; Przyborowski, J.A. Variations in the Chemical Composition and Content of Salicylic Glycosides in the Bark of Salix Purpurea from Natural Locations and Their Significance for Breeding. *Fitoterapia* 2017, 118, 118–125. [CrossRef]
- Ostolski, M.; Adamczak, M.; Brzozowski, B.; Wiczkowski, W. Antioxidant Activity and Chemical Characteristics of Supercritical CO2 and Water Extracts from Willow and Poplar. *Molecules* 2021, 26, 545. [CrossRef] [PubMed]
- Bonaterra, G.A.; Heinrich, E.U.; Kelber, O.; Weiser, D.; Metz, J.; Kinscherf, R. Anti-Inflammatory Effects of the Willow Bark Extract STW 33-I (Proaktiv[®]) in LPS-Activated Human Monocytes and Differentiated Macrophages. *Phytomedicine* 2010, 17, 1106–1113. [CrossRef]
- Noleto-Dias, C.; Ward, J.L.; Bellisai, A.; Lomax, C.; Beale, M.H. Salicin-7-Sulfate: A New Salicinoid from Willow and Implications for Herbal Medicine. *Fitoterapia* 2018, 127, 166–172. [CrossRef]
- 25. Gil, Ł. Short Rotation Dendromass Bioactive Compound Contents, Thermophysical Properties and Elementary Composition. Ph.D. Thesis, University of Warmia and Mazury in Olsztyn, Olsztyn, Poland, 2021; p. 223.
- 26. Stolarski, M.J.; Gil, Ł.; Warmiński, K.; Krzyżaniak, M.; Olba-Zięty, E. Short Rotation Woody Crops as a Source of Bioactive Compounds Depending on Genotype and Harvest Cycle. *Ind. Crops Prod.* **2022**, *180*, 114770. [CrossRef]
- 27. *PN-EN ISO 18134-2*; Solid Biofuels–Determination of Moisture Content–Dryer Method–Part 2: Total Moisture–Simplified Method. Polish Standardization Committee: Warsaw, Poland, 2014.
- PN-EN ISO 18125:2017-07; Solid Biofuels—Determination of Calorific Value. Polish Standardization Committee: Warsaw, Poland, 2017.
- PN-EN ISO 18122:2016-01; Solid Biofuels—Determination of Ash Content. Polish Standardization Committee: Warsaw, Poland, 2016.
- 30. PN-EN ISO 18123:2016-01; Solid Biofuels—Determination of Volatile Matter Content. Polish Standardization Committee: Warsaw, Poland, 2016.
- PN-EN ISO 16948:2015-07; Solid Biofuels—Determination of Total Content of Carbon, Hydrogen and Nitrogen. Polish Standardization Committee: Warsaw, Poland, 2015.
- 32. *PN-EN ISO 16994:2016-10;* Solid Biofuels—Determination of Total Content of Sulfur and Chlorine. Polish Standardization Committee: Warsaw, Poland, 2016.
- PN-ISO 587:2000; Solid Fuels—Determination of Chlorine Using Eschka Mixture. Polish Standardization Committee: Warsaw, Poland, 2000.
- 34. *PN-EN ISO 16472:2007;* Determination of Amylase-Treated Neutral Detergent Fibre Content (ANDF). Polish Standardization Committee: Warsaw, Poland, 2007.
- 35. *PN-EN ISO 13906:2009;* Determination of Acid Detergent Fibre (ADF) and Acid Detergent Lignin (ADL) Contents. Polish Standardization Committee: Warsaw, Poland, 2009.
- Warmiński, K.; Stolarski, M.J.; Gil, Ł.; Krzyżaniak, M. Willow Bark and Wood as a Source of Bioactive Compounds and Bioenergy Feedstock. *Ind. Crops Prod.* 2021, 171, 113976. [CrossRef]
- Stolarski, M.J.; Szczukowski, S.; Tworkowski, J.; Krzyżaniak, M.; Załuski, D. Willow Biomass and Cuttings' Production Potential over Ten Successive Annual Harvests. *Biomass Bioenergy* 2017, 105, 230–247. [CrossRef]
- Bullard, M.J.; Mustill, S.J.; McMillan, S.D.; Nixon, P.M.I.; Carver, P.; Britt, C.P. Yield Improvements through Modification of Planting Density and Harvest Frequency in Short Rotation Coppice Salix Spp.—1. Yield Response in Two Morphologically Diverse Varieties. *Biomass Bioenergy* 2002, 22, 15–25. [CrossRef]
- 39. Stolarski, M.J.; Warmiński, K.; Krzyżaniak, M. Energy Value of Yield and Biomass Quality of Poplar Grown in Two Consecutive 4-Year Harvest Rotations in the North-East of Poland. *Energies* **2020**, *13*, 1495. [CrossRef]
- Sabatti, M.; Fabbrini, F.; Harfouche, A.; Beritognolo, I.; Mareschi, L.; Carlini, M.; Paris, P.; Scarascia-Mugnozza, G. Evaluation of Biomass Production Potential and Heating Value of Hybrid Poplar Genotypes in a Short-Rotation Culture in Italy. *Ind. Crops Prod.* 2014, 61, 62–73. [CrossRef]
- Guidi, W.; Tozzini, C.; Bonari, E. Estimation of Chemical Traits in Poplar Short-Rotation Coppice at Stand Level. *Biomass Bioenergy* 2009, 33, 1703–1709. [CrossRef]
- 42. Labrecque, M.; Teodorescu, T.I. Field Performance and Biomass Production of 12 Willow and Poplar Clones in Short-Rotation Coppice in Southern Quebec (Canada). *Biomass Bioenergy* **2005**, *29*, 1–9. [CrossRef]
- 43. Bergante, S.; Facciotto, G. Nine Years Measurements in Italian SRC Trial in 14 Poplar and 6 Willow Clones. In Proceedings of the 19th European Biomass Conference and Exhibition, Berlin, Germany, 6–10 June 2011; pp. 6–10.

- 44. Labrecque, M.; Teodorescu, T.I. High Biomass Yield Achieved by Salix Clones in SRIC Following Two 3-Year Coppice Rotations on Abandoned Farmland in Southern Quebec, Canada. *Biomass Bioenergy* **2003**, 25, 135–146. [CrossRef]
- Serapiglia, M.J.; Cameron, K.D.; Stipanovic, A.J.; Abrahamson, L.P.; Volk, T.A.; Smart, L.B. Yield and Woody Biomass Traits of Novel Shrub Willow Hybrids at Two Contrasting Sites. *Bioenergy Res.* 2013, 6, 533–546. [CrossRef]
- Sleight, N.J.; Volk, T.A.; Johnson, G.A.; Eisenbies, M.H.; Shi, S.; Fabio, E.S.; Pooler, P.S. Change in Yield Between First and Second Rotations in Willow (*Salix* Spp.) Biomass Crops Is Strongly Related to the Level of First Rotation Yield. *Bioenergy Res.* 2016, 9, 270–287. [CrossRef]
- 47. Aronsson, P.; Rosenqvist, H.; Dimitriou, I. Impact of Nitrogen Fertilization to Short-Rotation Willow Coppice Plantations Grown in Sweden on Yield and Economy. *Bioenergy Res.* 2014, 7, 993–1001. [CrossRef]
- 48. Nord-Larsen, T.; Sevel, L.; Raulund-Rasmussen, K. Commercially Grown Short Rotation Coppice Willow in Denmark: Biomass Production and Factors Affecting Production. *Bioenergy Res.* 2015, *8*, 325–339. [CrossRef]
- 49. Stolarski, M.J.; Szczukowski, S.; Tworkowski, J.; Krzyżaniak, M.; Załuski, D. Willow Production during 12 Consecutive Years—The Effects of Harvest Rotation, Planting Density and Cultivar on Biomass Yield. *GCB Bioenergy* **2019**, *11*, 635–656. [CrossRef]
- Geyer, W. Biomass Production in the Central Great Plains USA under Various Coppice Regimes. *Biomass Bioenergy* 2006, 30, 778–783. [CrossRef]
- Grünewald, H.; Böhm, C.; Quinkenstein, A.; Grundmann, P.; Eberts, J.; von Wühlisch, G. Robinia pseudoacacia L.: A Lesser Known Tree Species for Biomass Production. *Bioenergy Res.* 2009, 2, 123–133. [CrossRef]
- 52. Gruenewald, H.; Brandt, B.K.V.; Schneider, B.U.; Bens, O.; Kendzia, G.; Hüttl, R.F. Agroforestry Systems for the Production of Woody Biomass for Energy Transformation Purposes. *Ecol. Eng.* **2007**, *29*, 319–328. [CrossRef]
- 53. Rédei, K.; Veperdi, I. The Role of Black Locust (*Robinia pseudoacacia* L.) in Establishment of Short-Rotation Energy Plantations in Hungary. *Int. J. Hortic. Sci.* 2009, 15, 41–44. [CrossRef]
- Vande Walle, I.; Van Camp, N.; Van de Casteele, L.; Verheyen, K.; Lemeur, R. Short-Rotation Forestry of Birch, Maple, Poplar and Willow in Flanders (Belgium) II. Energy Production and CO2 Emission Reduction Potential. *Biomass Bioenergy* 2007, *31*, 276–283. [CrossRef]
- 55. Manzone, M.; Bergante, S.; Facciotto, G. Energy and Economic Evaluation of a Poplar Plantation for Woodchips Production in Italy. *Biomass Bioenergy* 2014, 60, 164–170. [CrossRef]
- Manzone, M.; Calvo, A. Energy and CO2 Analysis of Poplar and Maize Crops for Biomass Production in North Italy. *Renew.* Energy 2016, 86, 675–681. [CrossRef]
- 57. Manzone, M.; Bergante, S.; Facciotto, G. Energy and Economic Sustainability of Woodchip Production by Black Locust (*Robinia pseudoacacia* L.) Plantations in Italy. *Fuel* **2015**, *140*, 555–560. [CrossRef]
- Stachowicz, P.; Stolarski, M.J. Thermophysical Properties and Elemental Composition of Black Locust, Poplar and Willow Biomass. Energies 2022, 16, 305. [CrossRef]
- 59. Gasol, C.M.; Brun, F.; Mosso, A.; Rieradevall, J.; Gabarrell, X. Economic Assessment and Comparison of Acacia Energy Crop with Annual Traditional Crops in Southern Europe. *Energy Policy* **2010**, *38*, 592–597. [CrossRef]
- 60. Mitsui, Y.; Seto, S.; Nishio, M.; Minato, K.; Ishizawa, K.; Satoh, S. Willow Clones with High Biomass Yield in Short Rotation Coppice in the Southern Region of Tohoku District (Japan). *Biomass Bioenergy* **2010**, *34*, 467–473. [CrossRef]
- 61. Tharakan, P.J.; Volk, T.A.; Abrahamson, L.P.; White, E.H. Energy Feedstock Characteristics of Willow and Hybrid Poplar Clones at Harvest Age. *Biomass Bioenergy* 2003, 25, 571–580. [CrossRef]
- 62. Monedero, E.; Hernández, J.; Collado, R. Combustion-Related Properties of Poplar, Willow and Black Locust to Be Used as Fuels in Power Plants. *Energies* 2017, *10*, 997. [CrossRef]
- 63. Gendek, A.; Malatak, J.; Velebil, J. Effect of Harvest Method and Composition of Wood Chips on Their Caloric Value and Ash Content. *Sylwan* **2018**, *162*, 248–257.
- Fabio, E.S.; Volk, T.A.; Miller, R.O.; Serapiglia, M.J.; Kemanian, A.R.; Montes, F.; Kuzovkina, Y.A.; Kling, G.J.; Smart, L.B. Contributions of Environment and Genotype to Variation in Shrub Willow Biomass Composition. *Ind. Crops Prod.* 2017, 108, 149–161. [CrossRef]
- 65. Bajcar, M.; Zaguła, G.; Saletnik, B.; Tarapatskyy, M.; Puchalski, C. Relationship between Torrefaction Parameters and Physicochemical Properties of Torrefied Products Obtained from Selected Plant Biomass. *Energies* **2018**, *11*, 2919. [CrossRef]
- Straker, K.C.; Quinn, L.D.; Voigt, T.B.; Lee, D.K.; Kling, G.J. Black Locust as a Bioenergy Feedstock: A Review. *Bioenergy Res.* 2015, 8, 1117–1135. [CrossRef]
- Jagustyn, B.; Patyna, I.; Skawińska, A. Evaluation of Physicochemical Properties of Palm Kernel Shell as Agro Biomass Used in the Energy Industry. *Chemik* 2013, 67, 552–559.
- Stolarski, M.J.; Śnieg, M.; Krzyżaniak, M.; Tworkowski, J.; Szczukowski, S.; Graban, Ł.; Lajszner, W. Short Rotation Coppices, Grasses and Other Herbaceous Crops: Biomass Properties versus 26 Genotypes and Harvest Time. *Ind. Crops Prod.* 2018, 119, 22–32. [CrossRef]
- 69. Krzyżaniak, M.; Stolarski, M.J.; Waliszewska, B.; Szczukowski, S.; Tworkowski, J.; Załuski, D.; Śnieg, M. Willow Biomass as Feedstock for an Integrated Multi-Product Biorefinery. *Ind. Crops Prod.* **2014**, *58*, 230–237. [CrossRef]
- Przybysz, K.; Małachowska, E.; Martyniak, D.; Boruszewski, P.; Iłowska, J.; Kalinowska, H.; Przybysz, P. Yield of Pulp, Dimensional Properties of Fibers, and Properties of Paper Produced from Fast Growing Trees and Grasses. *Bioresources* 2018, 13, 1372–1387. [CrossRef]

- 71. Baker, P.; Charlton, A.; Johnston, C.; Leahy, J.J.; Lindegaard, K.; Pisano, I.; Prendergast, J.; Preskett, D.; Skinner, C. A Review of Willow (*Salix* spp.) as an Integrated Biorefinery Feedstock. *Ind. Crops Prod.* **2022**, *189*, 115823. [CrossRef]
- 72. Gao, J.; Jebrane, M.; Terziev, N.; Daniel, G. Evaluation of Wood Quality Traits in Salix Viminalis Useful for Biofuels: Characterization and Method Development. *Forests* **2021**, *12*, 1048. [CrossRef]
- 73. Jerbi, A.; Kalwahali-Muissa, M.; Krygier, R.; Johnston, C.; Blank, M.; Sarrazin, M.; Barnabé, S.; Laur, J.; Labrecque, M.; Brereton, N.J.B.; et al. Comparative Wood Anatomy, Composition and Saccharification Yields of Wastewater Irrigated Willow Cultivars at Three Plantations in Canada and Northern Ireland. *Biomass Bioenergy* 2023, 170, 106683. [CrossRef]

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