

# Low Indirect Land Use Change (ILUC) Energy Crops to Bioenergy and Biofuels—A Review

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The transformation systems vary with several factors like the desired biofuel product, composition of biomass, economics, time and operation conditions from an Microalgal-based biofuels can be obtained through chemical (Table S1), biochemical (Table S2, Table S3 and Table S4) and thermochemical (Table S5, Table S6 and Table S7) conversion pathways [1,2].

**Table S1.** Microalgae raw material studied for chemical conversion (transesterification) pathways towards biofuels.

Transesterification - Biodiesel				
Microalgae	Reaction	Conditions	Yield (%)	Reference
<i>Chlorella protothecoides</i>	Acidic transesterification	30°C; 7h; 160 rpm M/O = 45:1	68%	[3]
<i>Schizochytrium limacinum</i>	Bligh & Dyer extraction-transesterification	90°C; 40 min	55.5%	[4]
	<i>In situ</i> transesterification		FAME content = 66.4%	
			66.3%	
<i>Nannochloropsis oculata</i>	Soxhlet extraction-transesterification	50°C; 4 h; 1100 rpm M/O = 30:1	97.5%	[5]
<i>Chlorella pyrenoidosa</i>	<i>In situ</i> transesterification	90°C; 2h M/A <sup>1</sup> = 4:1 (v w <sup>-1</sup> )	95%	[6]
<i>Nannochloropsis gaditana</i>	<i>In situ</i> transesterification	255°C; 50 min; 70 rpm M/A = 10:1 (v w <sup>-1</sup> )	48%	[7]
<i>Nannochloropsis</i> sp.	<i>In situ</i> transesterification	50°C; 6 h; 1100 rpm M/A = 3.16:100 (w w <sup>-1</sup> )	95%	[8]
<i>Chlorella vulgaris</i>	1st: acidic esterification (radio frequency heating) + hexane extraction	Cell disruption: 90°C; 30 min	FAME yield:	[9]
	2nd: alkali transesterification (radio frequency heating)	55°C; 20 min M/A = 2:5 (v w <sup>-1</sup> )	1st: 58.8%	
			2nd: 79.5%	
<i>Chlorella vulgaris</i>	<i>In situ</i> transesterification using IL <sup>2</sup>	IL: [P4444][For] <sup>3</sup> 102.4°C, 4.6 h	FAME yield:	[10]
		M/A=9:1 (molar)	Theoretical: 98.0%	
		IL/A=8:1 (molar) Water content: 40.6%	Experimental: 98.6%	

<sup>1</sup> M/A - Methanol to biomass (algae) ratio

<sup>2</sup> IL - Ionic liquids

<sup>3</sup> [P4444][For] - Tetrabutylphosphonium formate.

**Table S2.** Microalgae feedstock applied for biochemical conversion (anaerobic digestion) pathways towards biofuels.

Anaerobic digestion - Biogas					
Microalgae	Reaction	Organism	Conditions	Yield (%)	Reference
<i>Chlamydomonas reinhardtii</i>	AD	Inoculum: Sludge from a local sewage plant	38°C; 32 d	587 ml biogas g <sup>-1</sup> VS (66% methane)	[11]
<i>Dunaliella salina</i>	AD	Inoculum: Sludge from a local sewage plant	38°C; 32 d	505 ml biogas g <sup>-1</sup> VS (64% methane)	[11]
<i>Chlorella vulgaris</i>	AD	Inoculum: Sludge from a tertiary WW treatment pond	35°C; 16-28 d	240 ml methane g <sup>-1</sup> VSS <sup>1</sup>	[12]
<i>Phaeodactylum tricornutum</i>	AD	Inoculum: Biomass from a full-scale anaerobic digester treating potato-processing WW	33°C; 30 d; I/M <sup>2</sup> = 3:1	360 ml methane g <sup>-1</sup> VS	[13]
<i>Scenedesmus obliquus</i>	AD	Inoculum: Biomass from a full-scale anaerobic digester treating potato-processing WW	33°C; 30 d; I/M= 3:1	240 ml methane g <sup>-1</sup> VS	[13]
<i>Scenedesmus sp.-AMDD</i>	AD	Inoculum: Biomass from a UASB <sup>3</sup> digester with apple processing WW	35°C; 34-50 d; 150 rpm; I/M= 3:1	410 ml methane g <sup>-1</sup> VS	[14]
<i>Isochrysis sp.</i>	AD	Inoculum: Biomass from a UASB digester with apple processing WW	35°C; 34–50 d; 150 rpm; I/M= 3:1	408 ml methane g <sup>-1</sup> VS	[14]
<i>Arthrospira platensis</i>	Co-digestion with sludge	Inoculum: Digested sludge collected from a lab-scale anaerobic reactor	35°C; 23 d; I/M=2:1; M/S <sup>4</sup> = 2:1	343 ml methane g <sup>-1</sup> VS	[15]

<sup>1</sup> VSS - Volatile Suspended Solids.<sup>2</sup> I/M - Inoculum to microalgae ratio.<sup>3</sup> UASB - Upflow Anaerobic Sludge Blanket.<sup>4</sup> M/S - Microalgae to sludge ratio.**Table S3.** Microalgae feedstock studied for biochemical conversion (alcoholic fermentation) pathways towards biofuels.

Alcoholic fermentation - Bioethanol					
Microalgae	Reaction	Organism	Conditions	Yield (%)	Reference
<i>Chlamydomonas reinhardtii</i> UTEX 90	SHF	<i>Saccharomyces cerevisiae</i> S288C	30°C; pH: 5; 24 h; 160 rpm; Yeast: 10 % (v/v)	29.2%	[16]
<i>Chlorococum sp.</i>	SHF	<i>Saccharomyces bayanus</i>	30°C; 60 h; 200 rpm; Substrate: 10 g L <sup>-1</sup> ; Yeast: 3% (v/v)	38%	[17]
<i>Chlorella vulgaris</i>	SHF	<i>Escherichia coli</i> SJL2526	37°C; pH: 7; 24h; 150 rpm; Substrate: 5 g L <sup>-1</sup>	40%	[18]

<i>Schizocytrium</i> <i>sp.</i>	SSF	<i>Escherichia coli</i> KO11	37°C; 72 h; 150 rpm	5.5%	[19]
<i>Scenedesmus</i> <i>obliquus</i>	SHF	<i>Kluyveromyces</i> <i>marxianus</i> IGC 2671 <i>Saccharomyces</i> <i>carlsbergensis</i> ATCC 6269 <i>Saccharomyces</i> <i>bayanus</i>	30°C; pH: 5.2; 78 h; 150 rpm; Substrate: 500 ml microalga hydrolysate; Yeast: 300 mg dw <sup>-1</sup> L <sup>-1</sup>	<i>K. marxianus</i> : 11.7 g L <sup>-1</sup> <i>S. carlsbergensis</i> : 11.2 g L <sup>-1</sup> <i>S. bayanus</i> : 9 g L <sup>-1</sup>	[20]
<i>Chlorella</i> <i>sp.</i>	SHF SSF	<i>Zymomonas</i> <i>mobilis</i> ATCC 29191	30°C; 60 h; 200 rpm; Substrate: 20 g L <sup>-1</sup> ; Bacterium: 10% (v v <sup>-1</sup> )	SHF: 17.8% SSF: 21.4%	[21]
<i>Arthrospira</i> <i>platensis</i>	SHF	<i>Sacharomyces</i> <i>cerevisiae</i> CAT-1	30°C; 20 h; Substrate: 20 g L <sup>-1</sup> ; Bacterium: 10% (v v <sup>-1</sup> )	83.36% 1.57 g L <sup>-1</sup> h <sup>-1</sup> (16h)	[22]
<i>Arthrospira</i> <i>sp.</i>	SHF	<i>Sacharomyces</i> <i>cerevisiae</i> ATCC 26603	30°C; 24 h; B/M <sup>1</sup> : 25% (v v <sup>-1</sup> ); Bacterium: 10% (v v <sup>-1</sup> )	78.9% 0.72 g L <sup>-1</sup> h <sup>-1</sup>	[23]
<i>Arthrospira</i> <i>sp.</i>	SSF	<i>Sacharomyces</i> <i>cerevisiae</i> CAT-1	30°C; 72 h; 130 rpm; Substrate: 20 % (15% corn starch + 5% alga) (w v <sup>-1</sup> ); Bacterium: 10% (v v <sup>-1</sup> )	55 g L <sup>-1</sup>	[24]

<sup>1</sup> B/M - Biomass to molasses ratio.

**Table S4.** Microalgae feedstock applied for biochemical conversion (biological H<sub>2</sub> production) pathways towards biofuels.

Biological H <sub>2</sub> production - Biohydrogen					
Microalgae	Reaction	Organism	Conditions	Yield (%)	Reference
<i>Chlamydomonas</i> <i>reinhardtii</i> 137c	Biophotolysis (S deprivation)	-	25°C; pH: 7.2; 120 h; I <sup>1</sup> : 110 µmol m <sup>-2</sup> s <sup>-1</sup>	2.5 ml H <sub>2</sub> L <sup>-1</sup> culture h <sup>-1</sup>	[25]
<i>Chlorella</i> <i>sorokiniana</i> Ce	Biophotolysis (S deprivation)	-	30°C; pH: 7.2; 220 h I: 120 µmol m <sup>-2</sup> s <sup>-1</sup>	1.35 ml H <sub>2</sub> L <sup>-1</sup> culture h <sup>-1</sup>	[26]
<i>Chlamydomonas</i> <i>reinhardtii</i>	Biophotolysis (S deprivation)	-	120 h I: 100 µmol m <sup>-2</sup> s <sup>-1</sup>	5.2 ml H <sub>2</sub> L <sup>-1</sup> culture	[27]
<i>Chlorella</i> <i>vulgaris</i>	Biophotolysis (S deprivation)	-	27°C; 52 h I: 120 µmol m <sup>-2</sup> s <sup>-1</sup> 24 h light/48 h dark	530 ml H <sub>2</sub> L <sup>-1</sup> culture 34.8 ml H <sub>2</sub> L <sup>-1</sup> culture h <sup>-1</sup>	[28]
<i>Chlorella</i> <i>vulgaris</i>	DF	<i>Clostridium</i> <i>butyricum</i>	37°C; 30 h; 220 rpm; Substrate: 5 g L <sup>-1</sup> ; Bacterium: 20%	81 ml H <sub>2</sub> g <sup>-1</sup> (dw)	[29]

(v v <sup>-1</sup> )					
<i>Nannochloropsis</i> sp.	DF	<i>Enterobacter aerogenes</i>	30°C; 6 h; 220 rpm; Substrate: 2.5 g L <sup>-1</sup> ; Bacterium: 10% (v v <sup>-1</sup> )	60.6 ml H <sub>2</sub> g <sup>-1</sup> (dw)	[30]
<i>Chlorella pyrenoidosa</i>	DF	Hydrogen producing and photosynthetic bacteria	DF: 35°C; pH: 6; 48 h; Substrate: 20 g L <sup>-1</sup> ; PF <sup>2</sup> : 30°C; pH: 8; 96 h; Substrate: 15 mM SMPs <sup>3</sup> ; I: 120 µmol m <sup>-2</sup> s <sup>-1</sup>	198.3 ml g <sup>-1</sup> TVS	[31]
<i>Scenedesmus obliquus</i>	DF	<i>Enterobacter aerogenes</i>	30°C; 6 h; 220 rpm; Substrate: 2.5 g L <sup>-1</sup> ; Bacterium: 10% (v v <sup>-1</sup> )	57.6 ml H <sub>2</sub> g <sup>-1</sup> VS (wet biomass)	[32]
<i>Scenedesmus obliquus</i>	DF	<i>Clostridium butyricum</i>	37°C; 48 h; 150 rpm; Substrate: 50 g L <sup>-1</sup> ; Bacterium: 1% (v v <sup>-1</sup> )	113.1 ml H <sub>2</sub> g <sup>-1</sup> VS (dried biomass)	[32]
<i>Scenedesmus obliquus</i>	DF	<i>Clostridium butyricum</i>	37°C; 96 h; 150 rpm; Substrate: 50 g L <sup>-1</sup> (dw); Bacterium: 1% (v v <sup>-1</sup> )	116.3 ml H <sub>2</sub> g <sup>-1</sup> alga	[33]
<i>Spirogyra</i> sp.	DF	<i>Clostridium butyricum</i> DSM 10702	37°C; 144 h; 150 rpm; Substrate: 10 g L <sup>-1</sup> (dw); Bacterium: 1% (v v <sup>-1</sup> )	47 ml H <sub>2</sub> g <sup>-1</sup> alga <sup>dw</sup> 156 ml H <sub>2</sub> g <sup>-1</sup> total sugars	[34]
<i>Spirogyra</i> sp.	DF (SBR <sup>4</sup> )	<i>Clostridium butyricum</i> DSM 10702	37°C; 10-14 h; 150 rpm; Substrate: 10 g <sub>sugar</sub> L <sup>-1</sup> ; Bacterium: 10% (v v <sup>-1</sup> )	324 ml H <sub>2</sub> L <sup>-1</sup> h <sup>-1</sup> 4.4 L H <sub>2</sub> L <sup>-1</sup> FM	[35]
<i>Chlorella</i> sp.	DF	Mixed acidogenic bacteria	35°C; 150 h; pH 7; 150 rpm; Substrate: 80-100 g <sub>VS</sub> L <sup>-1</sup> ; Bacteria: 25% (v v <sup>-1</sup> )	22 ml H <sub>2</sub> g <sup>-1</sup> VS	[36]
<i>Chlorella vulgaris</i>	Biophotolysis (S deprivation)	-	30°C; 120 h; pH 7.5; I (purple light): 140 µmol m <sup>-2</sup> s <sup>-1</sup>	60.4 ml H <sub>2</sub> L <sup>-1</sup> 39.18 ml H <sub>2</sub> L <sup>-1</sup> d <sup>-1</sup>	[37]
<i>Scenedesmus obliquus</i>	Biophotolysis (S deprivation)	-	30°C; 120 h; pH 7.5; I (purple light): 140 µmol m <sup>-2</sup> s <sup>-1</sup>	128 ml H <sub>2</sub> L <sup>-1</sup> 204 ml H <sub>2</sub> L <sup>-1</sup> d <sup>-1</sup>	[37]

<sup>1</sup> I - Light Intensity.<sup>2</sup> PF - Photo-Fermentation.<sup>3</sup> SMPs - Soluble Metabolite Products.<sup>4</sup> SBR - Sequential Batch Reactor.

**Table S5.** Microalgae raw material studied for thermochemical conversion (gasification) pathways towards biofuels.

Gasification - Syngas				
Microalgae	Reaction	Conditions	Yield	Reference
<i>Chlorella vulgaris</i>	Supercritical water gasification	600°C; 240 bar; 2 min; 7.3 wt% loading	53%	[38]
<i>Nannochloropsis</i> sp.	Supercritical water gasification	450°C; 40 min; 4.8 wt% loading	16.3 mmol H <sub>2</sub> g <sup>-1</sup> 5.1 mmol methane g <sup>-1</sup>	[39]
<i>Chlorella vulgaris</i>	Supercritical water gasification	500°C; 36 MPa; 30 min; Catalyst: 1.67M NaOH	12 mmol H <sub>2</sub> g <sup>-1</sup>	[40]
<i>Nannochloropsis oculata</i>	Steam gasification	700°C; 30 min; Catalyst: Fe <sub>2</sub> O <sub>3</sub> -CeO <sub>2</sub>	413 cm <sup>3</sup> H <sub>2</sub> g <sup>-1</sup> 278 cm <sup>3</sup> CO <sub>2</sub> g <sup>-1</sup>	[41]
<i>Nannochloropsis gaditana</i>	Supercritical water gasification	663°C; 24 MPa; 128 s; 3-5 wt% loading	73-97%	[42]
<i>Chlorella vulgaris</i>	Steam gasification	800°C; 10 min	61.7%	[43]
<i>Chlorella vulgaris</i>	Chemical looping gasification (CLG)	800°C; 10 min; Oxygen carrier: Fe <sub>2</sub> O <sub>3</sub>	81.6% 1.05 Nm <sup>3</sup> syngas kg <sup>-1</sup>	[43]
<i>Chlorella pyrenoidosa</i>	Supercritical water gasification	430°C; 2-13 MPa; 60 min	5.6 mmol H <sub>2</sub> g <sup>-1</sup> 8.2 mmol methane g <sup>-1</sup>	[44]
<i>Chlorella vulgaris</i>	Supercritical water gasification	385°C; 26 MPa; 30 min; Catalyst: Ru/charcoal	87% C gasification	[45]
<i>Chlorella vulgaris</i> (lipid-extracted) <i>Arthrospira platensis</i> (lipid-extracted)	Steam gasification	800°C; 10°C min <sup>-1</sup> ; Catalyst: Ni-CaO	Yield: 497.29 ml H <sub>2</sub> g <sup>-1</sup> Purity: 63.15% Yield: 435 ml H <sub>2</sub> g <sup>-1</sup> Purity: 45.78%	[46]

**Table S6.** Microalgae feedstock studied for thermochemical conversion (pyrolysis) pathways towards biofuels.

Pyrolysis - Bio-oil, biochar, biogas				
Microalgae	Reaction	Conditions	Yield	Reference
<i>Chlorella protothecoides</i>	Slow pyrolysis	500°C; 5 min	Bio-oil: 52%	[47]
<i>Chlorella protothecoides</i>	Fast pyrolysis	450°C; 2-3 s; N <sub>2</sub> Flow = 0.4 m <sup>3</sup> h <sup>-1</sup>	Bio-oil (Autotrophic): 16.6%; Bio-oil (Heterotrophic): 57.9	[48]
<i>Nannochloropsis</i> sp.	Slow pyrolysis	400°C; 120 min; N <sub>2</sub> Flow: 30 ml min <sup>-1</sup>	Bio-oil: 31% Biochar: 28% Biogas: 25%	[49]
<i>Chlorella</i> sp.	Microwave-assisted	750 W; 20 min;	Bio-oil: 28.6%	[50]

	pyrolysis	N <sub>2</sub> Flow: 500 ml min <sup>-1</sup> ; Biomass/Char= 1:5 (w w <sup>-1</sup> )		
<i>Chlorella sp.</i>	Slow pyrolysis	450°C; 30 min; Air Flow: 100 ml min <sup>-1</sup>	Bio-oil: 55% (40% energy recovery) Biochar: 30% Biogas: 15%	[51]
<i>Chlorella vulgaris</i>	Microwave-assisted pyrolysis	1500 W; 20 min; N <sub>2</sub> Flow: 300 ml min <sup>-1</sup>	Bio-oil: 35.83% Biochar: 29.87% Biogas: 33%	[52]
<i>Chlorella vulgaris</i> (lipid-extracted)	Fast pyrolysis	500°C	Bio-oil: 53% Biochar: 31% Biogas: 10%	[53]
<i>Scenedesmus obliquus</i>	Slow pyrolysis	475°C; 30 min; N <sub>2</sub> Flow: 200 ml min <sup>-1</sup>	Bio-oil: 57.6% Bio-char: 25.6% Biogas: 16.8%	[54]
<i>Arthrospira platensis</i>	Slow pyrolysis	550°C; 60 min	Bio-oil: 35% Biochar: 32% Biogas: 12%	[55]
<i>Nannochloropsis sp.</i> <i>Tetraselmis sp.</i> <i>Isochrysis galbana</i>	Fast pyrolysis	500°C; N <sub>2</sub> Flow: 3500 ml min <sup>-1</sup>	Bio-oil: 58-66% Biochar: 21-30% Biogas: 13%	[56]
<i>Chlorella sp.</i>	Slow pyrolysis	550°C; 240 min	Bio-oil: 42.81% Biochar: 55% Biogas: 1.14%	[57]

**Table S7.** Microalgae raw material studied for thermochemical conversion (hydrothermal liquefaction) pathways towards biofuels.

Hydrothermal liquefaction - Bio-oil				
Microalgae	Reaction	Conditions	Yield	Reference
<i>Botryococcus braunii</i>	Liquefaction	300°C; 60 min	57%	[58]
<i>Dunaliella tertiolecta</i>	Liquefaction	300°C; 5 min	43.8% (organic basis)	[59]
<i>Nannochloropsis sp.</i>	Liquefaction	350°C; 60 min	43 wt%	[60]
<i>Nannochloropsis sp.</i>	Liquefaction	350°C; 60 min	57%	[61]
<i>Chlorella vulgaris</i>	Liquefaction	350°C; 60 min	35.8% daf <sup>1</sup>	[62]
<i>Nannochloropsis oculata</i>			34.3% daf	
<i>Porphyridium creuntum</i>			21% daf	
<i>Arthrospira sp.</i>			29% daf	
<i>Desmodesmus sp.</i>	Liquefaction	375°C; 5 min	49%	[63]
<i>Nannochloropsis sp.</i>	Liquefaction	600°C; 1 min	66%	[64]
<i>Chlorella sp.</i>	Liquefaction	350°C; 1.4 min	39.7% daf	[65]
<i>Scenedesmus obliquus</i>	Liquefaction	300°C; 15 min	44% daf	[66]
<i>Tetraselmis sp.</i>	Liquefaction	350°C; 30 min	31%	[67]
<i>Haematococcus pluvialis</i>	Liquefaction	200°C; 150 min	54.2%	[68]

<sup>1</sup> daf - dry ash free basis.

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