



Immune therapy for breast cancer in 2010—hype or hope?

A. Florescu MSc MD, E. Amir MD,*
N. Bouganim MD,† and M. Clemons MD†*

ABSTRACT

The identification of numerous breast cancer antigens has generated increasing enthusiasm for the application of immune-based therapies in breast malignancies. Although the use of monoclonal antibodies has revolutionized the “targeted therapy” of breast cancer, and the immunomodulatory effects of bisphosphonates continue to be evaluated, few studies to date have demonstrated widespread utility for other forms of immunotherapy. The present review assesses modern research and explores whether the hopes for immunotherapy can overcome the hype.

KEY WORDS

Breast cancer, immunotherapy, cancer vaccine, cytokine, monoclonal antibody

1. INTRODUCTION

Despite significant advances in the systemic management of breast cancer, metastatic disease (mBC) remains incurable, and in the adjuvant setting, the overall 5-year survival rate for women with node-positive breast cancer has reached a relative plateau. A possible explanation for this situation is that, by the time of breast cancer diagnosis, a substantial number of patients have already developed micrometastases that are never fully eradicated, even with “optimal” adjuvant treatment.

Despite arising from normal host cells, tumour cells can exhibit some degree of immunogenicity, and the immune system has a number of mechanisms by which it acts against them. A possible mechanism for targeting micrometastatic foci or limited disease is to enhance the immune response of the patient to the tumour cells. By enhancing immunogenicity, a robust and lasting immune response might be attained. Renal cell carcinoma and melanoma are the most eloquent examples of the efficacy of immunologic manipulation in eradicating a disseminated cancer¹.

In breast cancer, components of the immune system are thought to play important roles in several parts of the metastatic cascade². Various immune strategies have been tested as therapy for breast cancer: vaccine therapy, administration of exogenous cytokines, monoclonal antibodies, and gene therapy. As a guide to the practicing clinician, the present article reviews current progress and future research into breast cancer immunotherapy.

2. METHODS

In April 2010, we searched MEDLINE (1950 to April 2010) for English-language studies on immunotherapy in breast cancer. The American Society of Clinical Oncology (ASCO) database of abstracts was similarly searched for the years 2005–2010. Subject headings used included “immunotherapy,” “immunology,” “cancer vaccines,” “cytokines,” and “antibodies, monoclonal,” in combination with “breast neoplasm” and “therapeutics.” We refined the search by identifying specific categories of immunotherapies: cancer vaccines, cytokine-based therapy, therapeutic monoclonal antibodies, CpG (cytosine and guanine separated by a phosphate) deoxynucleotides, and bisphosphonates.

This search strategy identified 2984 articles, which were screened by title for relevance. In addition, reference lists of other review articles on the topic were screened, and any relevant articles were included.

3. RESULTS

3.1 Cancer Vaccines

Malignant cells can generate a tumour-specific immune response. This response occurs when an antigen specific to the tumour cells is recognized by the immune system. The antigen is then cleared, neutralized, or destroyed. This innate process may be re-created by the administration of a vaccine containing tumour-specific antigens.

Vaccination strategies are defined as “active specific immunotherapy” because the host immune system is activated *de novo* or is re-stimulated to mount an effective tumour-specific reaction against malignant cells. Theoretically, cancer vaccines have the potential to induce a more robust immune response with minimal toxicity. Multiple antigens have been studied for their ability to stimulate an effective immune response. The ones with the most promise in breast cancer are mucin 1 (MUC1), carcinoembryonic antigen (CEA), the human epidermal growth factor receptor 2 (HER2/*neu*), and alpha-lactalbumin. Table 1 includes a summary of stage of development and proven utility for various cancer vaccines.

3.1.1 MUC1

The membrane-associated glycoprotein MUC1 is a found on the apical surface of many epithelial cells. It is overexpressed in more than 70% of cancers, making it an attractive antigen for research. Preclinical studies demonstrated that MUC1 could induce a humoral

immune response, but not a cellular response^{3–6}. Overall, clinical trials using MUC1 have been variable and inconclusive in stimulating an effective immune response. Initial phase I data showed a weak anti-MUC1 response, leading to development of several modifications and adjuvants aimed at enhancing immunogenicity^{7–11}. Despite modifications, clinical outcome studies failed to show tumour regression or other important clinical endpoints. Phase II trials also failed to provide convincing evidence of a role for this vaccine in tumour regression^{12,13}.

One variation of the MUC1 vaccine that met some success in early trials is Theratope (Biomira, Edmonton, AB), designed by linking a synthetic sialyl-Tn (STn) that emulates a unique carbohydrate in human cancer with the protein carrier keyhole limpet hemocyanin (KLH). In a phase II trial in MBC, overall survival (OS) was significantly improved in women treated with Theratope ($n = 50$; OS: 19.1 months) compared with historical controls ($n = 104$; OS: 9.2 months)¹⁴. Following up on this positive trend, Biomira completed

TABLE 1 Developmental stage and utility of immunotherapies in breast cancer

Immunotherapy	References				Useful? ^a
	Preclinical	Phase I	Phase II	Phase III	
Cancer vaccines					
Mucin 1	3–6	7–11	12–14	Theratope ^{15,b}	–
				Stimuvax ^c	–
				(NCT00925548 ^d)	
Carcinoembryonic antigen		16	17	NCT00217373 ^d	–
				NCT00003125 ^d	*
				NCT00085241 ^d	*
HER2/ <i>neu</i>		18–24	25–27	NCT00854789 ^d	+
Alpha-lactalbumin	28	Trials planned			*
Cytokine therapy					
Interferon α	29	30	NCT00227656 ^b		–
Tumour necrosis factor α		31			–
Interferon γ	32, 33				–
Interleukin 10	34–36				–
Interleukin 12	37–39	40			*
Interleukin 2	41,42		43		–
Granulocyte–macrophage colony-stimulating factor	44–47				*
Flt-3 ligand	48	49			–

^a A minus sign (–) means that the therapy is not currently useful, based on available data; a plus sign (+) means that the therapy is useful, with potential to be incorporated into practice, based on available data; and an asterisk (*) means that the data are insufficient to comment on utility (trials still recruiting participants or trials not completed).

^b Biomira, Edmonton, AB.

^c Oncothyreon, Seattle, WA, U.S.A.

^d For details, search using the trial number at www.clinicaltrials.gov/ct2/search.

a multicentre, double-blind phase III trial, randomizing 1038 mbc patients to receive either Theratope or KLH and the adjuvant detox-B stable emulsion¹⁵. The primary endpoints of that trial, time to disease progression and OS, were not different between the two groups. The only survival benefit was found in *post hoc* analyses for women on hormonal therapy who received Theratope and who developed high antibody titres against naturally clustered STn antigens (OS: 41 months vs. 25 months).

The most developed MUC1 vaccine is Stimuvax (Oncothyreon, Seattle, WA, U.S.A.), which was, until recently, recruiting patients for a phase III trial in hormone-sensitive advanced or metastatic breast cancer (NCT00925548). That trial was suspended in March 2010 when a patient in a phase II trial of Stimuvax acquired encephalitis.

Although MUC1 is an attractive tumour-specific antigen, the evidence available to date for MUC1-based vaccines has not shown altered clinical outcomes in breast cancer.

3.1.2 CEA

The glycoprotein CEA is expressed in carcinomas of breast, colon, lung, pancreas, and gastrointestinal tract. It has a role in cancer spread, promoting cancer cell adhesion.

As has been the case with MUC1 vaccines, development of an adequate CEA-based vaccine has involved various modifications. Priming with a recombinant CEA–*Vaccinia* virus and boosting with recombinant avian pox virus CEA was shown to be more effective at inducing CEA-specific immune responses⁵⁰. In another strategy, tricom (triad of co-stimulatory molecules B7.1, intercellular adhesion molecule-1, and lymphocyte function-associated antigen-3) was added.

A clinical trial of CEA–tricom enrolled 58 cancer patients with CEA-overexpressing tumours who were treated with 8 dose levels of CEA–tricom vaccine. After a 4-vaccination schedule, 6 of 33 patients had moderate levels of immunoglobulin G, 1 lung cancer patient had a complete clinical response, and 40% of patients had stable disease¹⁶.

A small phase II trial in 15 mbc patients looked at the benefit of Panvac (Therion Biologics, Cambridge, MA, U.S.A.), a recombinant pox virus vaccine targeting both MUC1 and CEA¹⁷. Panvac was administered with granulocyte–macrophage colony-stimulating factor (GM-CSF) in 11 patients and with GM-CSF and docetaxel in 3. In the non-chemotherapy arm, 3 of 11 patients had disease stabilization for at least 6 months, 2 of 11 had tumour size reduction not meeting partial response criteria, and 1 had a confirmed partial response. Of patients receiving vaccine and docetaxel, one had a 50% reduction in the size of a chest wall lesion, and another had improvement of disease on bone scan.

Evaluation of anti-CEA vaccines is ongoing in more advanced clinical trials. Scenarios include efficacy in combination with other anticancer therapies

such as interferon alfa [IFN α (NCT00217373)], interleukin-2 [IL-2 (NCT00003125)], and radiation therapy (NCT00085241).

3.1.3 HER2/neu

The tyrosine receptor protein HER2/*neu* is expressed in epithelial tumours and overexpressed in approximately 20% of primary breast cancers. Several phase I trials of vaccination against HER2/*neu* have demonstrated safety and immunogenicity, with only rare grade 3 toxicities^{18,19}. Two small phase I studies have provided some promising data on a HER2/*neu* dendritic cell (DC) vaccine, lapuleucel-T^{20,21}.

Vaccine optimization remains an issue, with several studies demonstrating short-lived low-magnitude immune responses, especially in the absence of immune adjuvants^{22–25}. Clinical outcome studies are more scarce, but promising. A recent study that demonstrated immunogenicity for a HER2/*neu* vaccine in combination with GM-CSF in the adjuvant setting was presented in abstract form at the ASCO annual meeting in 2008. Patients with advanced breast cancer who had been vaccinated with HER2/*neu* vaccines as part of previous phase I/II trials ($n = 75$) were followed for a median of 2.7 years. In those with HER2/*neu*-specific T-cell immunity, a significant, dose-dependent increase in OS was seen²⁶.

Peoples *et al.* conducted a trial of HER2/*neu* vaccine in 53 node-positive breast cancer patients treated with standard therapies and free of active disease, but at high risk of recurrence²⁷. Women positive for the human leukocyte antigen A2 [HLA-A2+ ($n = 24$)] were vaccinated against HER2/*neu*; the patients negative for HLA-A2 constituted the control group. The treatment arms were not balanced: tumours in the HER2/*neu*-vaccination arm were larger, more poorly differentiated, and less hormone-sensitive. Despite those differences, outcomes favoured the treatment group: at 22 months' follow-up, disease-free survival in the treatment arm was 86% as compared with 60% in the control arm. Recurrence rates were 8% and 21% respectively; time to recurrence was 11 months and 8 months respectively. This trial has been expanded and is still aiming to enrol a total of 95 patients (NCT00854789).

In another trial, the same group of investigators enrolled 186 women in remission from a previously treated breast cancer. Of those patients, 101 (HLA-A2+ and -A3+) were given a HER2/*neu* vaccine and GM-CSF as an adjunct. The remaining 85 patients constituted a control group ($n = 85$). Recurrence rates at a median follow-up of 20 months were 5.6% in the vaccinated group and 14.2% in the control group⁵¹.

Based on these limited data, HER2/*neu* vaccines hold some promise for the future.

3.1.4 Alpha-Lactalbumin

Alpha-lactalbumin is a breast-specific differentiation protein expressed in high amounts in most human

breast carcinomas and in mammary epithelial cells only during lactation. Recent data show that immunoreactivity against alpha-lactalbumin provides substantial protection against growth of autochthonous tumours in transgenic mouse models of breast cancer²⁸. It was concluded that alpha-lactalbumin vaccination has the potential to provide safe and effective primary protection against the development of breast cancer for women in their post-childbearing, premenopausal years, when lactation is avoidable and the risk for developing breast cancer is high. Clinical studies are currently being planned.

3.2 Cytokine Therapy

Cytokines are regulatory molecules secreted by lymphocytes and macrophages. They alter the function of target cells by binding to cell-surface receptors. Cytokines provide important intercellular signals in inflammation, immunity, and the biology of tumour and endothelial cells.

In vivo data have shown that, despite the presence of adequate infiltration by lymphocytes into breast carcinomas, the immune response appears to be downregulated—a circumstance thought to result

from the release of “inhibitory” cytokines into the tumour microenvironment². Furthermore, breast cancer is weakly immunogenic and poorly recognized by the immune system⁵². Some available data demonstrate that certain cytokines are able to promote the generation or activation of antitumour effector cells, including DCs or lymphokine-activated killer (LAK) cells. Once activated, these cells can result in inhibition and, in certain instances, regression of tumour growth⁵³.

The foregoing mechanisms of action have led to numerous hypotheses about clinical activity for cytokines in breast cancer when delivered alone or in combination with chemotherapy or other biologic agents. Those hypotheses have included a reduction in the immunosuppressive effects on the microenvironment, enhanced innate or adaptive immunity, and potentially direct cytotoxicity. In Table I, the stage of development is outlined for various cytokines; Table II lists the roles of cytokines that have been studied in breast cancer.

3.2.1 IFN α

Laboratory data show that IFN α has antiproliferative effects on the growth of MCF-7 cell lines. In addition,

TABLE II Roles of cytokines in breast cancer

<i>Cytokine</i>	<i>Role</i>	<i>Reference</i>
Interferon α	Antiproliferative effects	Kamamura <i>et al.</i> , 1998 ³³
Tumour necrosis factor α	Induces apoptosis and necrosis Activates lymphokine activated killer (LAK) cells	Purohit <i>et al.</i> , 2002 ⁵³ Wang <i>et al.</i> , 2003 ⁵⁴
Interferon γ	Activates macrophages/monocytes Differentiation of naïve T-helper cells Increases activity of LAK cells	Pulaski and Ostrand-Rosenberg, 2002 ³² Schreiber and Schreiber, 2003 ⁵⁵
Interleukin 10	Modulates monocyte/macrophage function	Kundu and Fulton, 1997 ³⁶ Venetsanakos <i>et al.</i> , 1997 ³⁴ Dorsey <i>et al.</i> , 2002 ³⁵
Interleukin 12	Stimulates T cells, natural killer (NK) cells	Cavallo <i>et al.</i> , 1999 ³⁷
Interleukin 2	Proliferation of LAK, NK, T cells	Rosenberg <i>et al.</i> , 1985 ⁵⁶ Mule <i>et al.</i> , 1987 ⁵⁷ Addison <i>et al.</i> , 1995 ⁴¹ Stewart <i>et al.</i> , 1999 ⁴²
Granulocyte-macrophage colony-stimulating factor	Growth and differentiation of dendritic cells (DCs)	Caux <i>et al.</i> , 1992 ⁵⁸ Dranoff <i>et al.</i> , 1993 ⁴⁶ Disis <i>et al.</i> , 1996 ⁴⁵
Flt-3 ligand	Stimulates early progenitor DCs	Brasel <i>et al.</i> , 1996 ⁴⁸ Maraskovsky <i>et al.</i> , 1996 ⁵⁹

induction of a number of IFN-inducible genes was shown to be a primary effect of IFN α ²⁹. A clinical study to determine the effectiveness of IFN α and IL-2 recruited women who had previously received 1–2 prior chemotherapy regimens for measurable inoperable, recurrent, or metastatic breast cancer. Of 40 patients accrued to the study, 32 were evaluable for response. Toxicities were frequent but manageable. The most common grades 3 and 4 toxicities were lymphopenia (17%) and malaise or fatigue (24%). No complete responses were observed, but 1 patient had a partial response (3%), and 6 patients had stable disease (19%) for an undetermined duration. It was therefore concluded that IFN α and IL-2 were ineffective ³⁰. A further clinical study exploring the combination of IFN α and capecitabine has been completed, but results are not yet available (NCT00227656).

3.2.2 TNF α

The multifunctional cytokine tumour necrosis factor α (TNF α) plays a key role in apoptosis, cell survival, and oncogenesis ⁶⁰. It is able to induce apoptosis and necrosis, activate cytolytic effector cells, and upregulate the expression of intercellular adhesion molecule 1 on tumour cells, an important event in the interaction with LAK cells ⁵⁴. The interaction of TNF α with the TNF receptors 1 and 2 activates several signal transduction pathways, leading to the diverse functions of TNF α . The TNF receptor 1 signalling molecules have been elucidated reasonably well, but regulation of their signalling remains unclear.

The anti-neoplastic effect of TNF α has been shown to be substantially augmented by co-administration of IL-12 ³¹. In one study, investigators activated human peripheral blood DCS to produce TNF α , which had a direct antitumour effect against breast cancer cell lines *in vitro*, leading to growth inhibition and apoptosis ⁵⁷. Those observations led to the hypothesis that TNF α should be included as part of immunotherapeutic treatment regimens. However, caution should be used in adding TNF α , because potential side effects include the induction of an autoimmune response from generalized T-cell activation ⁵⁵.

3.2.3 Interferon γ

Interferon γ (IFN γ) is a pro-inflammatory mediator that has an important function in the activation of the monocyte–macrophage lineage. It has been demonstrated that IFN γ is a critical component in regulating an innate phagocytic response against mBC ³². In addition, IFN γ has been shown to increase the activity of LAK cells in patients with breast cancer ³³. The use of IFN γ is currently limited because of the need for systemic delivery, which is associated with significant side effects and toxicity, including fever, fatigue, nausea, vomiting, and neurotoxicity. For this agent to be feasible in breast cancer, an alternative delivery system such as intratumoral instillation or conjugation to a cancer-specific antibody needs to be

developed, allowing for cytokine release to be more targeted. Development of the required drug delivery system is ongoing.

3.2.4 IL-10

The pleiotropic cytokine IL-10 can be immunosuppressive or immunostimulatory, depending on its relative concentration in the local microenvironment. Its principal action is to modulate monocyte and macrophage function. Laboratory studies using human breast cancer cells have shown that tumour tissue contains high levels of IL-10 messenger RNA. Interestingly, normal breast tissue showed little-to-no messenger RNA expression ³⁴. Conversely, however, animal studies showed that systemic administration of recombinant human IL-10 to animals bearing malignant mammary tumours led to significant inhibition of tumour growth and increased inflammatory cellular infiltration ³⁵. Kundu and Fulton ³⁶ showed that IL-10 expression downregulates class I major histocompatibility complex expression in tumour cells, leading to enhanced natural killer (NK) cell-mediated tumour cell lysis. In the tumour microenvironment, IL-10 is therefore considered to have concentration-dependent activity, with low concentrations facilitating tumour growth, but higher concentrations having considerable antitumour effects. As in the case of IFN γ , a method of targeted delivery of these agents is needed before they can be further investigated in breast cancer.

3.2.5 IL-12

As a pivotal factor for the initiation of cellular immunity, IL-12 has multiple stimulatory effects on T cells and NK cells. In mice harbouring spontaneously metastasizing mammary adenocarcinomas, antitumour effects have been demonstrated for local and systemic recombinant murine IL-12 ³⁷. Recently, in a murine model of breast cancer, intratumoral administration of polylactic acid–encapsulated microspheres containing IL-12, TNF α , and GM-CSF, was shown to enhance T-cell infiltration, resulting in tumour regression ³⁶. Tumour-specific T cells in the lymph nodes and spleen were also activated by this process, allowing for an additional long-term immune response to be activated.

Studies combining IL-12, chemotherapy, and trastuzumab have also been undertaken. Sequential (but not concurrent) use of paclitaxel and IL-12 reduced tumour burden in mice ^{38,39}. A phase I study in 21 patients with metastatic HER2/*neu*-positive tumours administered paclitaxel and trastuzumab followed by IL-12 ⁴⁰. Clinical response was observed in 10 of 21 patients: 1 complete response, 4 partial responses, and 5 stable disease at 1 year of follow-up. These data are promising, but results from more advanced trials are needed.

3.2.6 IL-2

Because of its proliferative effect on LAK, NK, and T cells, IL-2 also plays an important role in immunomodulation.

When administered alone or in combination with LAK cells, IL-2 has led to regression of a variety of established tumours^{34,56}. In a murine model, intratumoral injection of an adenovirus expressing the IL-2 gene resulted in regression of established murine breast cancer⁴¹. Studies exploring the efficacy of the antitumour effects of IL-2 have not produced similar results. Attempts use gene therapy to produce IL-2 in human breast tumours (by depositing an adenoviral vector encoding IL-2 into subcutaneous tumour) did not result in clinical responses despite measureable levels of IL-2 protein being detected in tumour biopsies⁴². A recent phase II study of trastuzumab with pulsed intermediate-dose IL-2 did not result in any objective responses⁴³.

3.2.7 GM-CSF

Granulocyte-macrophage colony-stimulating factor is a growth and differentiation factor for human DCs⁵⁸. These specialized antigen-presenting cells are believed to be responsible for stimulating naïve T cells. In experimental models, DCs have also been shown to augment secondary immune responses better than other antigen-presenting cells do⁴⁴. In rodent models, GM-CSF can, when mixed with a soluble antigen, be used as a vaccine adjuvant to induce both an antibody and a T-cell antigen-specific immune response⁴⁵. Transfection of tumour cells with DNA encoding GM-CSF, or direct injection of GM-CSF into established tumours, both resulted in antitumour immune responses in several murine models^{46,47}. Administration of GM-CSF as a vaccine adjuvant has also been shown to result in the mounting of an effective immune response, but the extent to which GM-CSF augmented the activity of the vaccine was unclear. Clinical data are pending.

3.2.8 Flt-3 Ligand

Flt-3 ligand is a potent stimulator of early progenitor DCs⁵⁹. Animal studies have demonstrated that, when administered systemically to mice, Flt-3 ligand increased circulating progenitor DCs that retain an antigen-presenting function and the capacity to stimulate proliferation of antigen-specific T cells⁴⁸. Flt-3 ligand is therefore currently under investigation as an anticancer agent and a possible vaccine adjuvant. In a phase I trial of 10 patients with HER2/*neu*-overexpressing cancer, participants received HER2/*neu* peptide-based vaccine that targeted the intracellular domain of the HER2/*neu* protein and Flt-3 ligand. In 5 patients, the HER2/*neu* peptide-based vaccine was given alone, and in 5 patients, the vaccine was admixed with GM-CSF on day 7 of the Flt-3 ligand cycle. Proliferative responses by T cells to the HER2/*neu* peptides and intracellular domain protein suggested that vaccine regimens including Flt-3 ligand as an adjuvant were not effective in eliciting a significant HER2/*neu* protein-specific proliferative response from the T-cells⁴⁹.

3.3 Trastuzumab

The HER2/*neu* proto-oncogene is a member of the human epidermal growth factor receptor family^{61,62}. Its overexpression is associated with more aggressive disease and worse prognosis. Trastuzumab (Herceptin: Hoffman-La Roche, Mississauga, ON) is a monoclonal antibody targeting the HER2/*neu* extracellular receptor, thereby inhibiting growth signals mediated by that pathway.

After clinical trials showed improved disease-free survival with trastuzumab treatment, trastuzumab was approved by the U.S. Food and Drug Administration in 1998 for the treatment of HER2/*neu*-positive metastatic breast cancer. An overview of the trastuzumab clinical program is beyond the scope of this article, but a detailed review has been published by Shak⁶³.

3.3.1 Single-Agent Trastuzumab Therapy

In women with MBC who progress on standard chemotherapy, single-agent trastuzumab was shown to produce a 15% response rate, with a median duration of response of 9.1 months⁶⁴. Other trials have also demonstrated improved efficacy with trastuzumab monotherapy. Vogel *et al.* randomized 114 women to two-weekly dose regimens of trastuzumab⁶⁵. The overall objective response rate was 26%, ranging from 35% in tumours with heavy immunohistochemical staining for HER2/*neu* to no response in weakly-to-moderately staining tumours. Based on that study, trastuzumab demonstrated safety and efficacy as first-line monotherapy in MBC overexpressing the HER2/*neu* oncogene.

3.3.2 Trastuzumab in Combination with Chemotherapy: the Metastatic Setting

Preclinical data suggested a synergistic effect between trastuzumab and several chemotherapeutic agents, including platinum agents, taxanes, cyclophosphamide, and anthracyclines^{66–70}. Positive phase II data have been complemented by strong phase III data showing that trastuzumab has a synergistic effect in combination with platinum-based chemotherapy⁷¹.

3.3.3 Trastuzumab in Combination with Chemotherapy in Early or Locally Advanced Breast Cancer

Evidence for the use of trastuzumab in combination with chemotherapy in early-stage breast cancer comes from four large trials—HERA (the Herceptin Adjuvant trial)^{72,73}, the National Surgical Adjuvant Breast and Bowel Project B31 trial, the North Central Cancer Treatment Group N9831 trial, and the Breast Cancer International Research Group BCIRG006 study⁷⁴—which all concluded that trastuzumab taken for 1 year after primary treatment reduces recurrence and improves survival in early-stage HER2/*neu*-overexpressing breast cancer.

3.3.4 Trastuzumab and IL-12

Preclinical studies have shown that NK cells secrete potent immunostimulatory cytokines in response to dual stimulation with trastuzumab-coated tumour cells and IL-12⁷⁵. Co-administration of these agents in mice resulted in enhanced NK cell activity. In a phase I trial of IL-12 and trastuzumab in 15 patients with HER2/*neu*-overexpressing malignancies, 1 complete response was reported in a woman with mBC, and 2 patients experienced stabilization of bone disease lasting 10 and 12 months respectively⁷⁶. Antibody-dependent cellular cytotoxicity against tumour targets did not correlate with clinical response or IL-12 dose. Unfortunately, the addition of IL-12 did not enhance the activity of trastuzumab.

3.4 CpG Oligodeoxynucleotides

Unmethylated CpG oligodeoxynucleotides (CpG ODNs) are characteristic of bacterial DNA and have established immunostimulatory properties⁷⁷. CpG ODNs have been studied in combination with monoclonal antibodies and with cytotoxic chemotherapy in epithelial malignancies. Wang *et al.*⁷⁸ evaluated CpG ODNs in several human and murine breast cancer models, showing tumour inhibition of at least 40% in all models. Addition of trastuzumab led to enhanced antitumour activity, resulting in more than 96% inhibition of tumour growth. It was suggested that the potentiating effects of CpG ODNs on monoclonal antibodies may be mediated by activation of NK cells, which mediate antibody-dependent cellular cytotoxicity.

Clinical data supporting CpG use are sparse in the breast cancer literature. A phase I trial was initiated by Pfizer (NCT00031278), aiming to characterize the safety and efficacy of antisense oligodeoxynucleotide CpG 7909 in combination with trastuzumab in mBC. The trial was terminated because of a modification to the route of administration. A continuation study was initiated for those previously treated with CpG 7909, and results of the latter study are awaited.

3.5 Bisphosphonates

The $\gamma\delta$ T cells are a small subset of peripheral T cells that recognize unique phosphoantigens lacking the requirements of classical antigen-presenting molecules. They have been shown to have potent antitumour activity *in vitro*^{79–82}.

Bisphosphonates are potent activators of $\gamma\delta$ T cells, inducing cytokine secretion and cell-mediated cytotoxicity by inhibition of farnesyl pyrophosphate synthase of the mevalonate pathway, leading to accumulation of isoprenoids⁸³.

Kunzmann *et al.*⁸⁴ showed that bisphosphonates can induce expansion of $\gamma\delta$ T cells and thereby have antiproliferative and antitumour functions. Compared with untreated cultures, pamidronate-treated

bone marrow cultures from 24 patients with multiple myeloma showed reduced survival. In metastatic renal carcinoma, a synthetic phosphoantigen (bromohydrin pyrophosphate) amplified $\gamma\delta$ T cells in 11 of 15 patients⁸⁵.

Wilhelm *et al.*⁸⁶ treated 19 patients with non-Hodgkin lymphoma or multiple myeloma with a combination of pamidronate and IL-2. Significant activation and proliferation of $\gamma\delta$ T cells was seen in 5 of 9 patients, and objective responses were observed in 3 of 9 patients. Only patients with significant proliferation of $\gamma\delta$ T cells responded to treatment.

Mattarollo *et al.*⁸⁷ demonstrated synergy with a combination of chemotherapy, bisphosphonates, and $\gamma\delta$ T cells. Chemotherapy and bisphosphonates both sensitize tumour cell lines to $\gamma\delta$ T cell-mediated cytotoxicity, but the combination was shown to enhance the effect. Similar results were seen in the ABCSG-12 study (Austrian Breast and Colorectal Cancer Study Group), which showed improved disease-free survival by 36% (hazard ratio: 0.64; $p = 0.01$) in hormone-positive premenopausal patients receiving zoledronic acid with endocrine treatment. The mechanism could again partly reflect induction of $\gamma\delta$ T cells.

4. CONCLUSIONS

The application of immunotherapeutic principles to the treatment and prevention of breast cancer has been ongoing for many decades. Although cytokines, cancer vaccines, and other host factors have been extensively studied in breast cancer, the therapeutic efficacy of these approaches remains unproven. The recent identification of tumour-specific immunity and of several breast cancer antigens has generated enthusiasm for the application of immune-based therapies. Although monoclonal antibodies, cytokines, and vaccines have all individually shown some promise, and although the immunomodulatory effects of bisphosphonates have taken a front seat in the treatment of breast cancer, it is likely that the best strategy to combat breast cancer will be a multimodality strategy. Clearly, different strategies demonstrate benefit in different patient populations. It may be that the best results will be obtained from vaccines in combination with a variety of antigens, or from vaccine and antibody combinations. Nonspecific and specific immunotherapy combinations may be another potent strategy. The effect of any of the aforementioned strategies in combination with more traditional cancer therapies is another avenue, given that some duration benefits have been seen with cytokines and chemotherapy. Given the mechanisms of immunotherapy, these treatments are most likely to work in the adjuvant setting and not in the setting in which they are usually tested: the heavily treated patient with mBC.

It is hoped that ongoing studies will yield the breakthrough needed to establish immunotherapy as a viable option in the treatment of breast cancer.

5. CONFLICT OF INTEREST DISCLOSURES

The authors declare that no financial conflict of interest exists.

6. REFERENCES

1. Yang JC, Childs R. Immunotherapy for renal cell carcinoma. *J Clin Oncol* 2006;24:5576–83.
2. Stewart TH. Evidence for immune facilitation of breast cancer growth and for the immune promotion of oncogenesis in breast cancer. *Medicina (B Aires)* 1996;56(suppl 1):13–24.
3. Ding L, Lalani EN, Reddish M, *et al.* Immunogenicity of synthetic peptides related to the core peptide sequence encoded by the human MUC1 mucin gene: effect of immunization on the growth of murine mammary adenocarcinoma cells transfected with the human MUC1 gene. *Cancer Immunol Immunother* 1993;36:9–17.
4. Acres RB, Hareuveni M, Balloul JM, Kieny MP. *Vaccinia* virus MUC1 immunization of mice: immune response and protection against the growth of murine tumors bearing the MUC1 antigen. *J Immunother Emphasis Tumor Immunol* 1993;14:136–43.
5. Apostolopoulos V, Xing PX, McKenzie IF. Murine immune response to cells transfected with human MUC1: immunization with cellular and synthetic antigens. *Cancer Res* 1994;54:5186–93.
6. Zhang S, Graeber LA, Helling F, *et al.* Augmenting the immunogenicity of synthetic MUC1 peptide vaccines in mice. *Cancer Res* 1996;56:3315–19.
7. Goydos JS, Elder E, Whiteside TL, Finn OJ, Lotze MT. A phase I trial of a synthetic mucin peptide vaccine. Induction of specific immune reactivity in patients with adenocarcinoma. *J Surg Res* 1996;63:298–304.
8. Reddish M, MacLean GD, Koganty RR, *et al.* Anti-MUC1 class I restricted CTLs in metastatic breast cancer patients immunized with a synthetic MUC1 peptide. *Int J Cancer* 1998;76:817–23.
9. Karanikas V, Hwang LA, Pearson J, *et al.* Antibody and T cell responses of patients with adenocarcinoma immunized with mannan–MUC1 fusion protein. *J Clin Invest* 1997;100:2783–92.
10. Musselli C, Ragupathi G, Gilewski T, Panageas KS, Spinat Y, Livingston PO. Reevaluation of the cellular immune response in breast cancer patients vaccinated with MUC1. *Int J Cancer* 2002;97:660–7.
11. Scholl SM, Balloul JM, Le Goc G, *et al.* Recombinant *Vaccinia* virus encoding human MUC1 and IL2 as immunotherapy in patients with breast cancer. *J Immunother* 2000;23:570–80.
12. Scholl S, Squiban P, Bizouarne N, *et al.* Metastatic breast tumour regression following treatment by a gene-modified *Vaccinia* virus expressing MUC1 and IL-2. *J Biomed Biotechnol* 2003;2003:194–201.
13. Rochlitz C, Figlin R, Squiban P, *et al.* Phase I immunotherapy with a modified *Vaccinia* virus (MVA) expressing human MUC1 as antigen-specific immunotherapy in patients with MUC1-positive advanced cancer. *J Gene Med* 2003;5:690–9.
14. Miles D, Papazisis K. Rationale for the clinical development of STn-KLH (Theratope) and anti-MUC-1 vaccines in breast cancer. *Clin Breast Cancer* 2003;3(suppl 4):S134–8.
15. Ibrahim NK, Murray J, Parker J, Finke L, Miles D on behalf of the Theratope Study Group. Humoral immune responses to naturally occurring STn in metastatic breast cancer patients treated with STn-KLH vaccine [abstract 2547]. *J Clin Oncol* 2004;22:174. [Available online at: www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst_detail_view&confID=26&abstractID=883; cited December 13, 2010]
16. Marshall JL, Gulley JL, Arlen PM, *et al.* Phase I study of sequential vaccinations with fowlpox-CEA (6D) tricom alone and sequentially with *Vaccinia*-CEA (6D) tricom, with and without granulocyte–macrophage colony-stimulating factor, in patients with carcinoembryonic antigen-expressing carcinomas. *J Clin Oncol* 2005;23:720–31.
17. Mohebtash M, Madan RA, Gulley JL, *et al.* Panvac vaccine alone or with docetaxel for patients with metastatic breast cancer [abstract 3035]. *J Clin Oncol* 2008;26:. [Available online at: www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst_detail_view&confID=55&abstractID=30165; cited December 13, 2010]
18. Disis ML, Schiffman K, Guthrie K, *et al.* Effect of dose on immune response in patients vaccinated with an HER-2/*neu* intracellular domain protein-based vaccine. *J Clin Oncol* 2004;22:1916–25.
19. Wiltshcke C, Wiedermann U, Zurbriggen R, *et al.* A phase I study to evaluate safety, immunogenicity and antitumour activity of a HER2 multi-peptide virosome vaccine in patients with metastatic breast cancer [abstract 3055]. *J Clin Oncol* 2008;26:. [Available online at: www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst_detail_view&confID=55&abstractID=35404; cited December 13, 2010]
20. Peethambaram PP, Melisko ME, Rinn KJ, *et al.* Phase I trial of immunotherapy with lapuleucel-T (APC8024) in patients with refractory metastatic tumors that express HER-2/*neu*. *Clin Cancer Res* 2009;15:5937–44.
21. Park JW, Melisko ME, Esserman LJ, Jones LA, Wollan JB, Sims R. Treatment with autologous antigen-presenting cells activated with the HER-2 based antigen lapuleucel-T: results of a phase I study in immunologic and clinical activity in HER-2 overexpressing breast cancer. *J Clin Oncol* 2007;25:3680–7.
22. Knutson KL, Schiffman K, Cheever MA, Disis ML. Immunization of cancer patients with a HER-2/*neu*, HLA-A2 peptide, p369–377, results in short-lived peptide-specific immunity. *Clin Cancer Res* 2002;8:1014–18.
23. Morse MA, Clay TM, Colling K, *et al.* HER2 dendritic cell vaccines. *Clin Breast Cancer* 2003;3(suppl 4):S164–72.
24. Czerniecki BJ, Roses RE, Koski GK. Development of vaccines for high-risk ductal carcinoma in situ of the breast. *Cancer Res* 2007;67:6531–4.
25. Moviglia GA, Gaeta C, Gabriela V, *et al.* Clinical response of patients with advanced breast cancer treated with dendritic cell vaccine with and without thymalfasin [abstract 2593]. *J Clin Oncol* 2005;23:. [Available online at: www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst_detail_view&confID=34&abstractID=33625; cited December 14, 2010]
26. Disis ML, Strickler JH, Wallace D, *et al.* Cellular immune parameters associated with improved long-term survival in advanced stage breast cancer patients after active immunization with a HER2-specific vaccine [abstract 3015]. *J Clin Oncol* 2008;26:. [Available online at: www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst_detail_view&confID=55&abstractID=35885; cited December 14, 2010]
27. Peoples GE, Gurney JM, Hueman MT, *et al.* Clinical trial results of a HER2/*neu* (E75) vaccine to prevent recurrence in high-risk breast cancer patients. *J Clin Oncol* 2005;23:7536–45.

28. Jaini R, Kesaraju P, Johnson JM, *et al.* An autoimmune strategy for prophylactic breast cancer vaccination. *Nat Med* 2010;16:799–803.
29. Tiwari RK, Wong GY, Mukhopadhyay B, *et al.* Interferon-alpha and gamma mediated gene responses in a human breast carcinoma cell line. *Breast Cancer Res Treat* 1991;18:33–41.
30. Kimmick G, Ratain MJ, Berry D, *et al.* Subcutaneously administered recombinant human interleukin-2 and interferon alfa-2a for advanced breast cancer: a phase II study of the Cancer and Leukemia Group B (CALGB 9041). *Invest New Drugs* 2004;22:83–9.
31. Sabel MS, Skitzki J, Stoolman L, *et al.* Intratumoral IL-12 and TNF- α -loaded microspheres lead to regression of breast cancer and systemic antitumor immunity. *Ann Surg Oncol* 2004;11:147–56.
32. Pulaski BA, Smyth MJ, Ostrand-Rosenberg S. Interferon gamma-dependent phagocytic cells are a critical component of innate immunity against metastatic mammary carcinoma. *Cancer Res* 2002;62:4406–12.
33. Kamamura Y, Takahashi K, Komaki K, Monden Y. Effects of interferon-alpha and gamma on development of LAK activity from mononuclear cells in breast cancer patients. *J Med Invest* 1998;45:71–5.
34. Venetsanakos E, Beckman I, Bradley J, Skinner JM. High incidence of interleukin 10 mRNA but not interleukin 2 mRNA detected in human breast tumours. *Br J Cancer* 1997;75:1826–30.
35. Dorsey R, Kundu N, Yang Q, *et al.* Immunotherapy with interleukin-10 depends on the CXCL12 chemokine inducible protein-10 and monokine induced by IFN-gamma. *Cancer Res* 2002;62:2606–10.
36. Kundu N, Fulton AM. Interleukin-10 inhibits tumor metastasis, downregulates MHC class I, and enhances NK lysis. *Cell Immunol* 1997;180:55–61.
37. Cavallo F, Di Carlo E, Butera M, *et al.* Immune events associated with the cure of established tumors and spontaneous metastases by local and systemic interleukin 12. *Cancer Res* 1999;59:414–21.
38. Mullins DW, Burger CJ, Elgert KD. Paclitaxel enhances macrophage IL-12 production in tumor-bearing hosts through nitric oxide. *J Immunol* 1999;162:6811–18.
39. Pressley JS, Elgert KD. Post-chemotherapeutic administration of interleukin-12 retards tumor growth and enhances immune cell function: combination therapy using paclitaxel and IL-12. *Cancer Invest* 2006;24:351–9.
40. Ramaswamy B, Bekaii-Saab T, Julie R, Young D, Shapiro CL, Carson W. Phase I study on interleukin 12 in combination with trastuzumab/paclitaxel in HER2 positive advanced solid tumors [abstract 174]. *Proc ASCO Breast Cancer Symp* 2007;:. [Available online at: www.asco.org/ASCOv2/Meetings/Abstracts?&vmview=abst_detail_view&confID=52&abstractID=40343; cited December 14, 2010]
41. Addison CL, Braciak T, Ralston R, Muller WJ, Gaudie J, Graham FL. Intratumoral injection of an adenovirus expressing interleukin 2 induces regression and immunity in a murine breast cancer model. *Proc Natl Acad Sci U S A* 1995;92:8522–6.
42. Stewart AK, Lassam NJ, Quirt IC, *et al.* Adenovector-mediated gene delivery of interleukin-2 in metastatic breast cancer and melanoma: results of a phase 1 clinical trial. *Gene Ther* 1999;6:350–63.
43. Mani A, Roda J, Young D, *et al.* A phase II trial of trastuzumab in combination with low-dose IL-2 in patients with metastatic breast cancer who have previously failed trastuzumab. *Breast Cancer Res Treat* 2009;117:83–9.
44. Levin D, Constant S, Pasqualini T, Flavell R, Bottomly K. Role of dendritic cells in the priming of CD4+ T lymphocytes to peptide antigen *in vivo*. *J Immunol* 1993;151:6742–50.
45. Disis ML, Bernhard H, Shiota FM, *et al.* Granulocyte-macrophage colony-stimulating factor: an effective adjuvant for protein and peptide-based vaccines. *Blood* 1996;88:202–10.
46. Dranoff G, Jaffee E, Lazenby A, *et al.* Vaccination with irradiated tumor cells engineered to secrete murine granulocyte-macrophage colony-stimulating factor stimulates potent, specific, and long-lasting antitumor immunity. *Proc Natl Acad Sci U S A* 1993;90:3539–43.
47. Kurane S, Arca MT, Aruga A, Krinock RA, Krauss JC, Chang AE. Cytokines as an adjuvant to tumor vaccines: efficacy of local methods of delivery. *Ann Surg Oncol* 1997;4:579–85.
48. Brasel K, McKenna HJ, Morrissey PJ, *et al.* Hematologic effects of Flt3 ligand *in vivo* in mice. *Blood* 1996;88:2004–12.
49. Disis ML, Rinn K, Knutson KL, *et al.* Flt3 ligand as a vaccine adjuvant in association with HER-2/neu peptide-based vaccines in patients with HER-2/neu-overexpressing cancers. *Blood* 2002;99:2845–50.
50. Hodge JW, McLaughlin JP, Kantor JA, Schlom J. Diversified prime and boost protocols using recombinant Vaccinia virus and recombinant non-replicating avian pox virus to enhance T-cell immunity and antitumor responses. *Vaccine* 1997;15:759–68.
51. Peoples GE, Holmes JP, Hueman MT, *et al.* Combined clinical trial results of a HER2/neu (E75) vaccine for the prevention of recurrence in high-risk breast cancer patients: U.S. Military Cancer Institute Clinical Trials Group Study I-01 and I-02. *Clin Cancer Res* 2008;14:797–803.
52. Allan CP, Turtle CJ, Mainwaring PN, Pyke C, Hart DN. The immune response to breast cancer, and the case for DC immunotherapy. *Cytotherapy* 2004;6:154–63.
53. Purohit A, Newman SP, Reed MJ. The role of cytokines in regulating estrogen synthesis: implications for the etiology of breast cancer. *Breast Cancer Res* 2002;4:65–9.
54. Wang H, Czura CJ, Tracey KJ. Tumor necrosis factor. In: Thomson AW, Lotze MT, eds. *The Cytokine Handbook*. 4th ed. London, U.K.: Academic Press; 2003: 837–60.
55. Schreiber GH, Schreiber RD. Interferon gamma. In: Thomson AW, Lotze MT, eds. *The Cytokine Handbook*. 4th ed. London, U.K.: Academic Press; 2003: 567–601.
56. Rosenberg SA, Mule JJ, Spiess PJ, Reichert CM, Schwarz SL. Regression of established pulmonary metastases and subcutaneous tumor mediated by the systemic administration of high-dose recombinant interleukin 2. *J Exp Med* 1985;161:1169–88.
57. Mule JJ, Yang JC, Afreniere RL, Shu SY, Rosenberg SA. Identification of cellular mechanisms operational *in vivo* during the regression of established pulmonary metastases by the systemic administration of high-dose recombinant interleukin 2. *J Immunol* 1987;139:285–94.
58. Caux C, Dezutter-Dambuyant C, Schmitt D, Banchereau J. GM-CSF and TNF-alpha cooperate in the generation of dendritic Langerhans cells. *Nature* 1992;360:258–61.
59. Maraskovsky E, Brasel K, Teepe M, *et al.* Dramatic increase in the numbers of functionally mature dendritic cells in Flt3

- ligand-treated mice: multiple dendritic cell subpopulations identified. *J Exp Med* 1996;184:1953–62.
60. Jablonska E. Serum levels of tumor necrosis factor alpha and production of this cytokine by polymorphonuclear cells in breast cancer patients. *Arch Immunol Ther Exp (Warsz)* 1998;46:93–6.
 61. Slamon DJ, Clark GM, Wong SG, Levin WJ, Ullrich A, McGuire WL. Human breast cancer: correlation of relapse and survival with amplification of the HER-2/*neu* oncogene. *Science* 1987;235:177–82.
 62. Slamon DJ, Godolphin W, Jones LA, *et al.*. Studies of the HER-2/*neu* proto-oncogene in human breast and ovarian cancer. *Science* 1989;244:707–12.
 63. Shak S. Overview of the trastuzumab (Herceptin) anti-HER2 monoclonal antibody clinical program in HER2-overexpressing metastatic breast cancer. Herceptin Multinational Investigator Study Group. *Semin Oncol* 1999;26:71–7.
 64. Cobleigh MA, Vogel CL, Tripathy D, *et al.*. Multinational study of the efficacy and safety of humanized anti-HER2 monoclonal antibody in women who have HER2-overexpressing metastatic breast cancer that has progressed after chemotherapy for metastatic disease. *J Clin Oncol* 1999;17:2639–48.
 65. Vogel CL, Cobleigh MA, Tripathy D, *et al.* Efficacy and safety of trastuzumab as a single agent in first-line treatment of HER2-overexpressing metastatic breast cancer. *J Clin Oncol* 2002;20:719–26.
 66. Pegram MD, Konecny GE, O’Callaghan C, Beryt M, Pietras R, Slamon DJ. Rational combinations of trastuzumab with chemotherapeutic drugs used in the treatment of breast cancer. *J Natl Cancer Inst* 2004;96:739–49.
 67. Slamon D, Pegram M. Rationale for trastuzumab (Herceptin) in adjuvant breast cancer trials. *Semin Oncol* 2001;28:13–19.
 68. Pietras RJ, Pegram MD, Finn RS, Maneval DA, Slamon DJ. Remission of human breast cancer xenografts on therapy with humanized monoclonal antibody to HER-2 receptor and DNA-reactive drugs. *Oncogene* 1998;17:2235–49.
 69. Pegram M, Hsu S, Lewis G, *et al.* Inhibitory effects of combinations of HER-2/*neu* antibody and chemotherapeutic agents used for treatment of human breast cancers. *Oncogene* 1999;18:2241–51.
 70. Pegram MD, Lopez A, Konecny G, Slamon DJ. Trastuzumab and chemotherapeutics: drug interactions and synergies. *Semin Oncol* 2000;27:21–5.
 71. Robert N, Leyland-Jones B, Asmar L, *et al.* Randomized phase III study of trastuzumab, paclitaxel, and carboplatin compared with trastuzumab and paclitaxel in women with HER-2-overexpressing metastatic breast cancer. *J Clin Oncol* 2006;24:2786–92.
 72. Romond EH, Perez EA, Bryant J, *et al.* Trastuzumab plus adjuvant chemotherapy for operable HER2-positive breast cancer. *N Engl J Med* 2005;353:1673–84.
 73. Piccard-Gebhart MJ, Procter M, Leyland Jones B, *et al.* Trastuzumab after adjuvant chemotherapy in HER2 *neu* positive breast cancer. *N Engl J Med* 2005;353:1659–72.
 74. Slamon D, Eiermann W, Robert N, *et al.* Phase III trial comparing doxorubicin and cyclophosphamide followed by docetaxel (AC T) with doxorubicin and cyclophosphamide followed by docetaxel and trastuzumab (AC-TH) with docetaxel, carboplatin and trastuzumab (TCH) in HER2 positive early breast cancer patients: BCIRG 006 study [abstract 62]. *Cancer Res* 2009;69(suppl):.
 75. Parihar R, Dierksheide J, Hu Y, Carson WE. IL-12 enhances the natural killer cell cytokine response to Ab-coated tumor cells. *J Clin Invest* 2002;110:983–92.
 76. Parihar R, Nadella P, Lewis A, *et al.* A phase I study of interleukin 12 with trastuzumab in patients with human epidermal growth factor receptor-2-overexpressing malignancies: analysis of sustained interferon gamma production in a subset of patients. *Clin Cancer Res* 2004;10:5027–37.
 77. Roda JM, Parihar R, Carson WE 3rd. CpG-containing oligodeoxynucleotides act through TLR9 to enhance the NK cell cytokine response to antibody-coated tumor cells. *J Immunol* 2005;175:1619–27.
 78. Wang H, Rayburn ER, Wang W, Kandimalla ER, Agrawal S, Zhang R. Immunomodulatory oligonucleotides as novel therapy for breast cancer: pharmacokinetics, *in vitro* and *in vivo* anticancer activity, and potentiation of antibody therapy. *Mol Cancer Ther* 2006;5:2106–14.
 79. Liu Z, Guo BL, Gehrs BC, Nan L, Lopez RD. *Ex vivo* expanded human Vγ9Vδ2+ γδ T cells mediate innate antitumor activity against human prostate cancer cells *in vitro*. *J Urol* 2005;173:1552–6.
 80. Corvaisier M, Moreau-Aubry A, Diez E, *et al.* V gamma 9 V delta 2 T cell response to colon carcinoma cells. *J Immunol* 2005;175:5481–8.
 81. Kabelitz D, Wesch D, Pitters E, Zöller M. Characterization of tumor reactivity of human V gamma 9V delta 2 gamma delta T cells *in vitro* and in SCID mice *in vivo*. *J Immunol* 2004;173:6767–76.
 82. Kato Y, Tanaka Y, Miyagawa F, Yamashita S, Minato N. Targeting of tumor cells for human gamma delta T cells by nonpeptide antigens. *J Immunol* 2001;167:5092–8.
 83. Knight LA, Conroy M, Fernando A, Polak M, Kurbacher CM, Cree IA. Pilot studies of the effect of zoledronic acid on tumor-derived cells *ex-vivo* in the TP-based tumor chemosensitivity assay. *Anticancer Drugs* 2005;16:969–76.
 84. Kunzmann V, Bauer E, Feurle J, Weissinger F, Tony HP, Wilhelm M. Stimulation of gamma delta T cells by aminobisphosphonates and induction of antiplasma cell activity in multiple myeloma. *Blood* 2000;96:384–92.
 85. Viey E, Fromont G, Escudier B, *et al.* Phosphostim-activated gamma delta T cells kill autologous metastatic renal cell carcinoma. *J Immunol* 2005;174:1338–47.
 86. Wilhelm M, Kunzmann V, Eckstein S, *et al.* Gamma delta T cells for immune therapy of patients with lymphoid malignancies. *Blood* 2003;102:200–6.
 87. Mattarollo SR, Kenna T, Nieda M, Nicol AJ. Chemotherapy and zoledronate sensitize solid tumour cells to Vgamma9Vdelta2 T cells cytotoxicity. *Cancer Immunol Immunother* 2007;56:1285–97.

Correspondence to: Mark Clemons, Division of Medical Oncology, The Ottawa Hospital Cancer Centre, Box 912, 501 Smyth Road, Ottawa, Ontario K1H 8L6.
E-mail: mclemons@toh.on.ca

- * Division of Medical Oncology and Hematology, Princess Margaret Hospital, and Department of Medicine, University of Toronto, Toronto, ON.
† Division of Medical Oncology, The Ottawa Hospital Cancer Centre, and Department of Medicine, University of Ottawa, Ottawa, ON.