

Supporting Information

Diisoprenyl Cyclohexene-Type Meroterpenoids with Cytotoxic Activity from a Mangrove Endophytic Fungus Aspergillus sp. GXNU-Y85

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Figure S1 ^1H NMR spectrum of compound **1** in CD_3OD

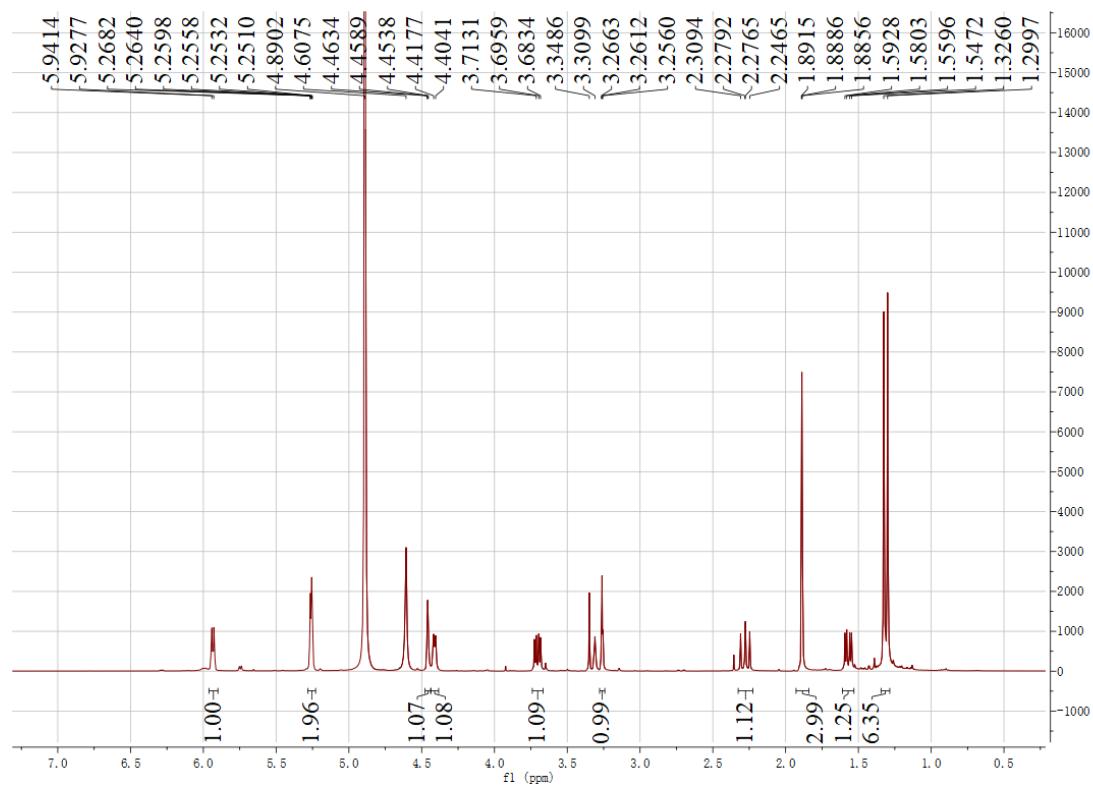


Figure S2 ^{13}C NMR spectrum of compound **1** in CD_3OD

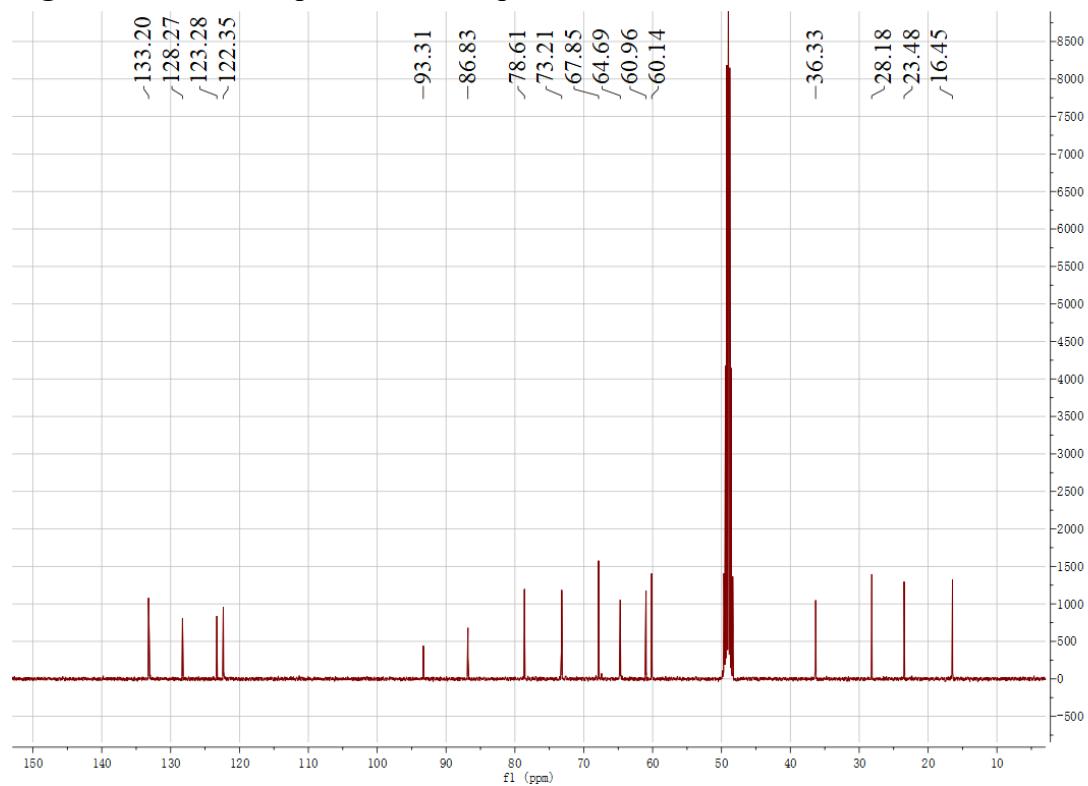


Figure S3 ^1H - ^1H COSY spectrum of compound **1** in CD_3OD

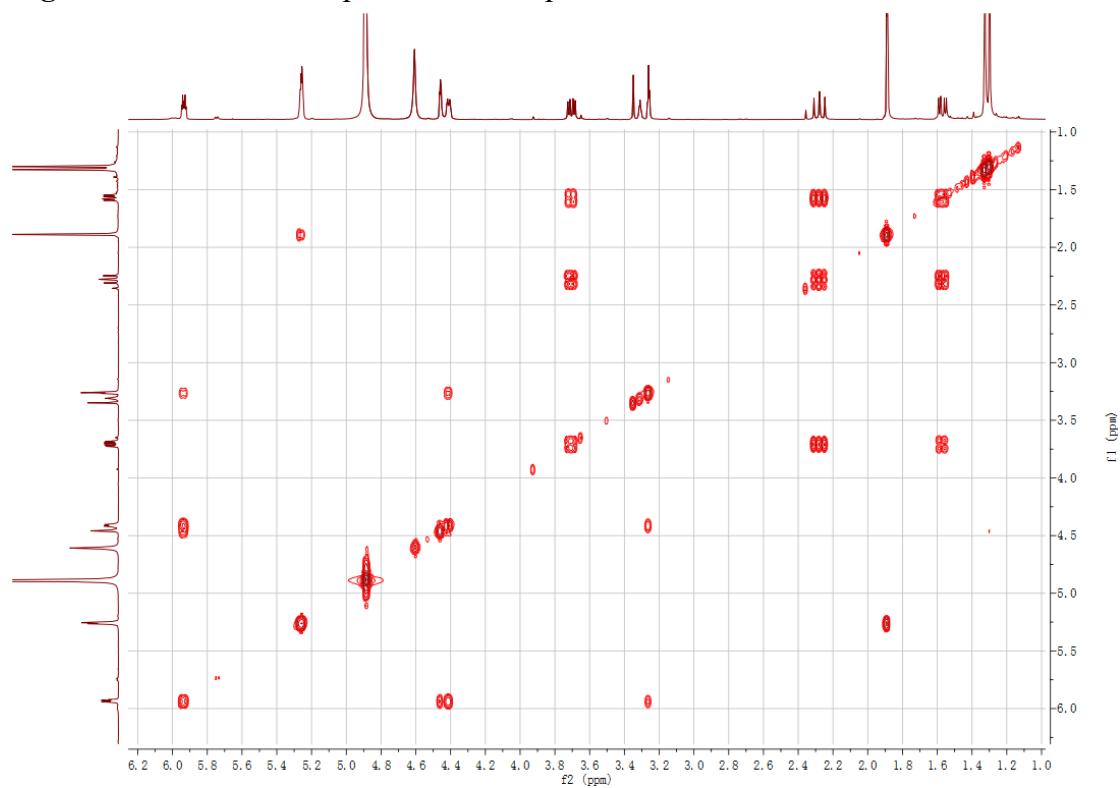


Figure S4 HSQC spectrum of compound **1** in CD_3OD

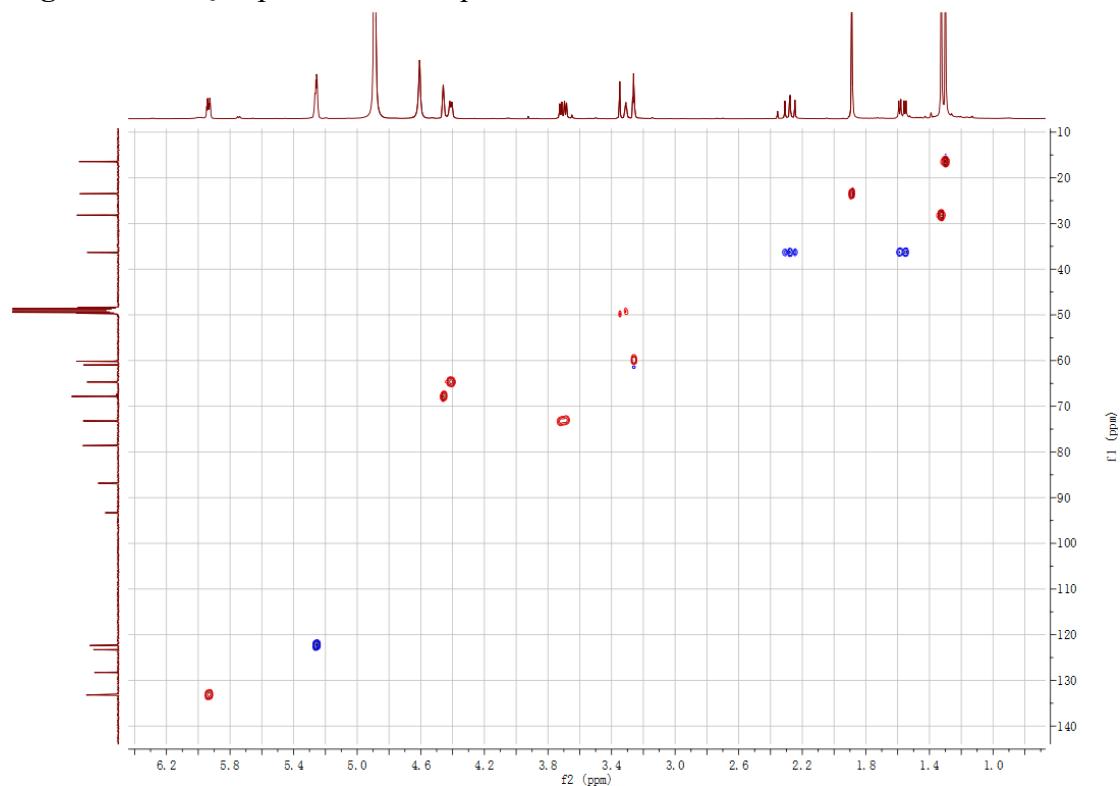


Figure S5 HMBC spectrum of compound **1** in CD₃OD

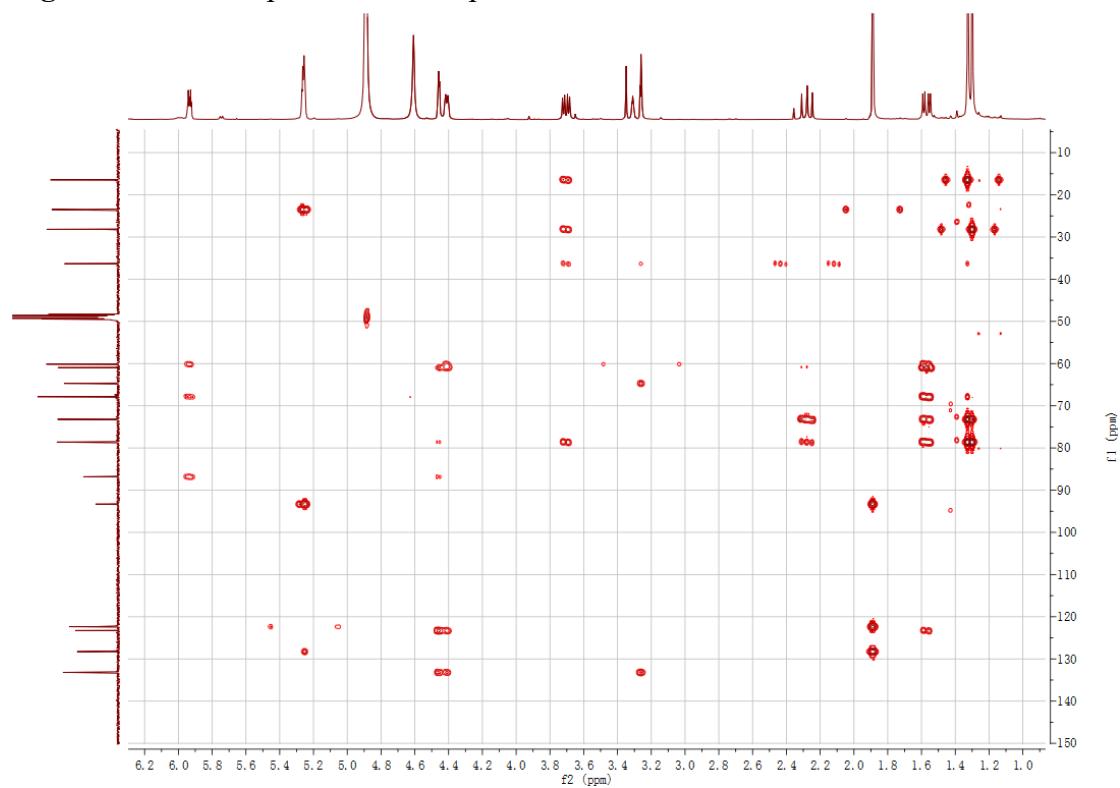


Figure S6 NOESY spectrum of compound **1** in CD₃OD

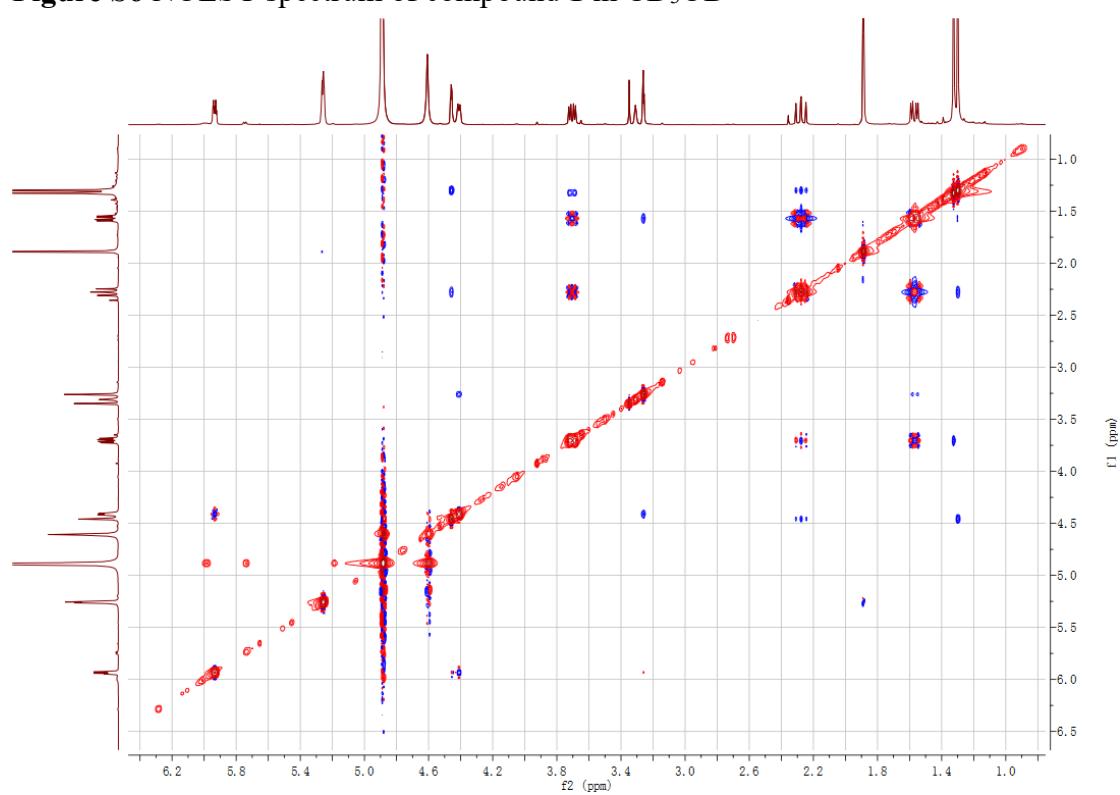


Figure S7 HR-ESI-MS spectrum of compound 1

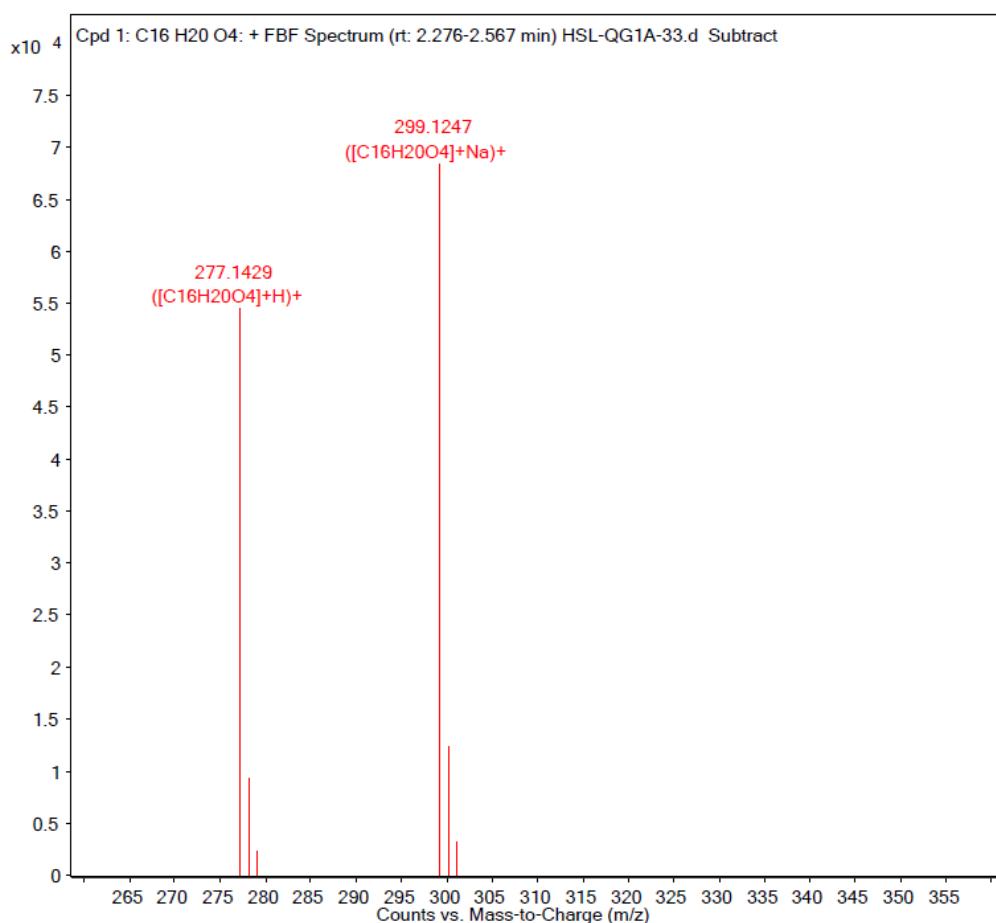


Figure S8 UV spectrum of compound 1

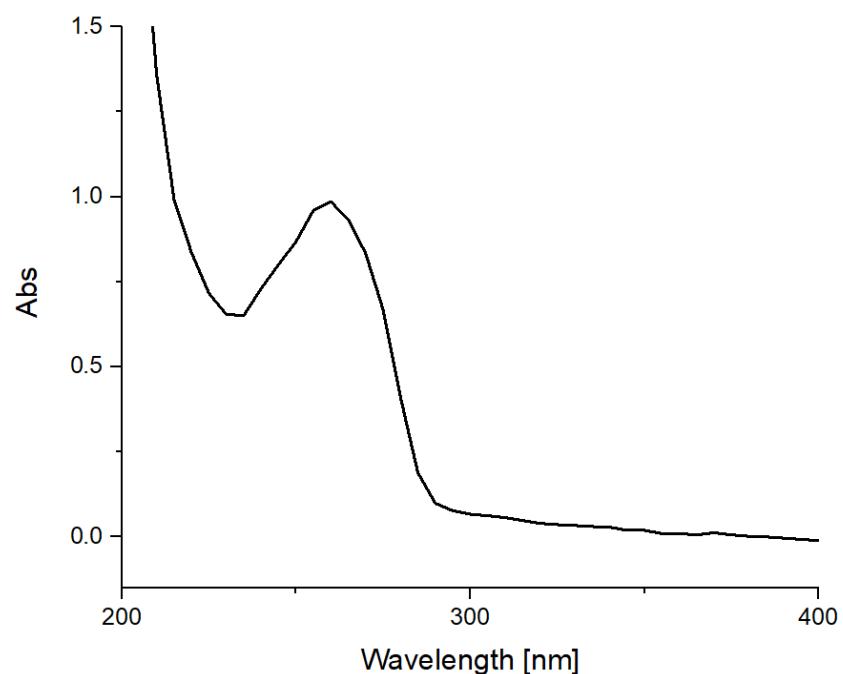


Figure S9 IR spectrum of compound 1

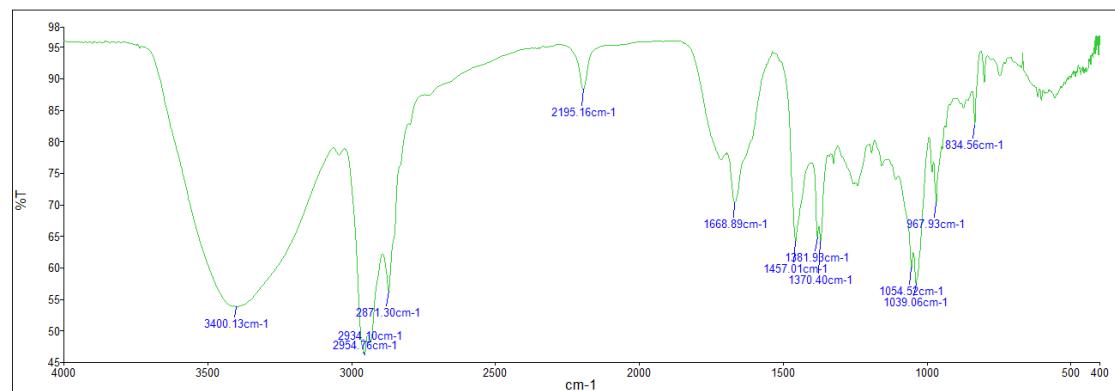


Figure S10 ^1H NMR spectrum of compound 2 in CD_3OD

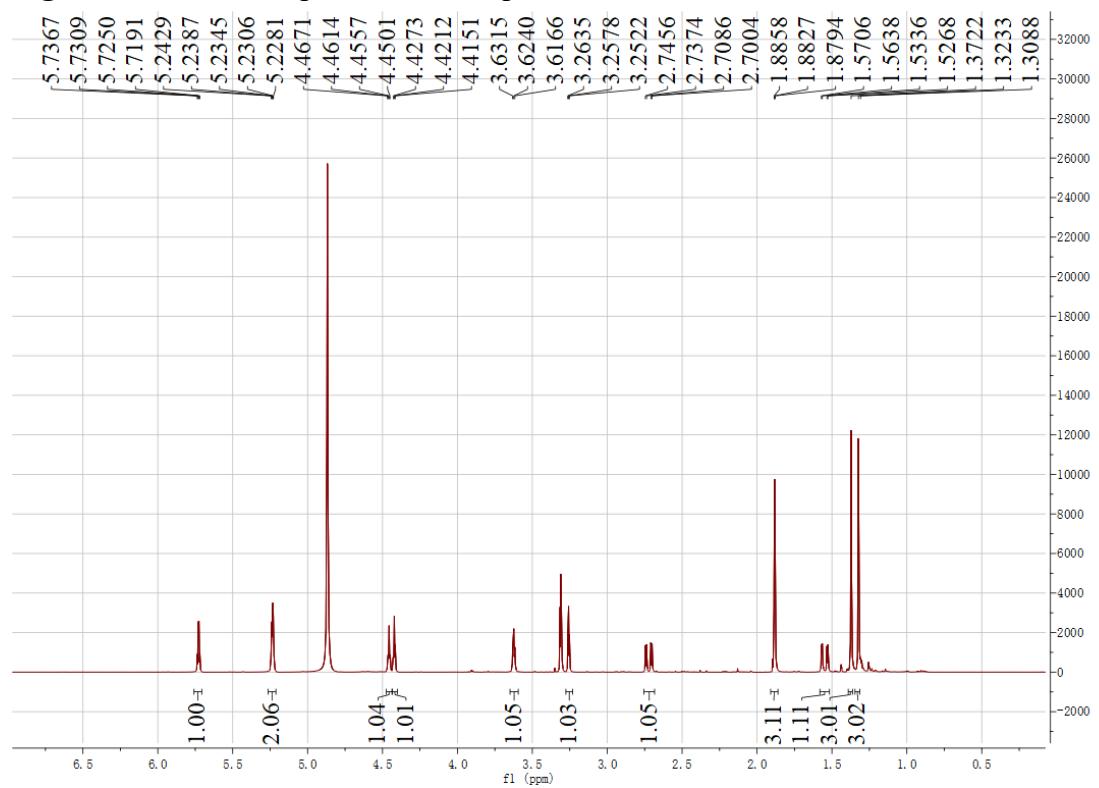


Figure S11 ^{13}C NMR spectrum of compound **2** in CD_3OD

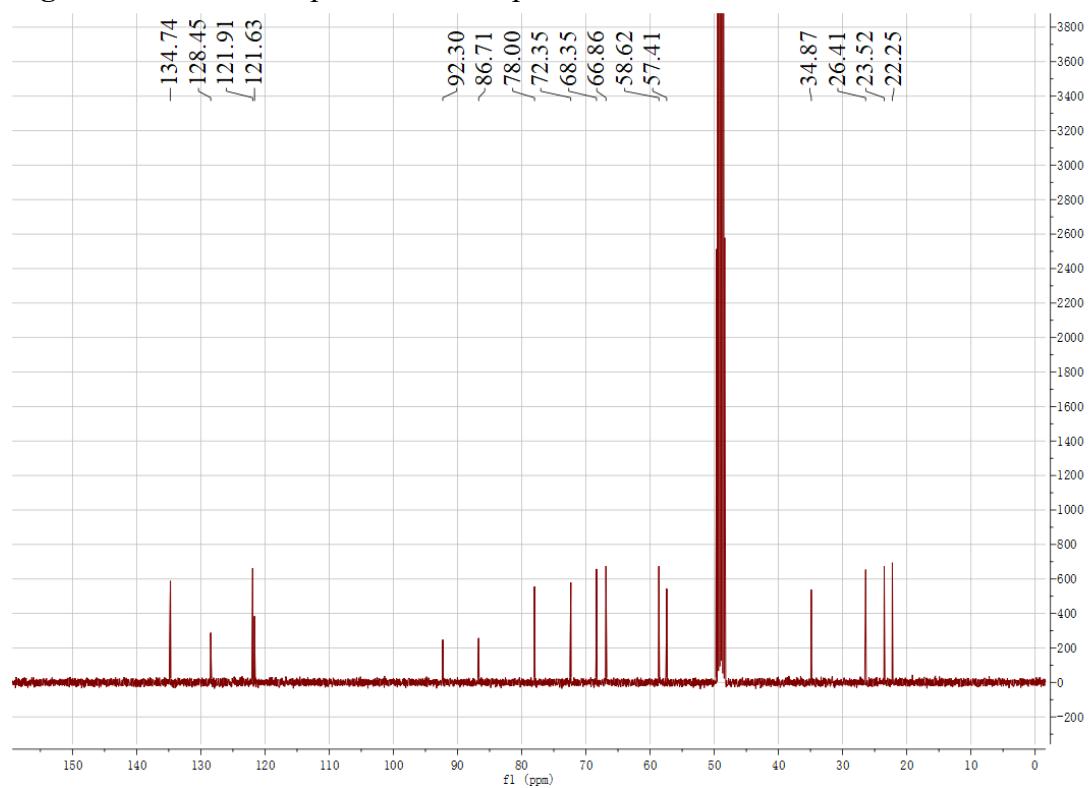


Figure S12 ^1H - ^1H COSY spectrum of compound **2** in CD_3OD

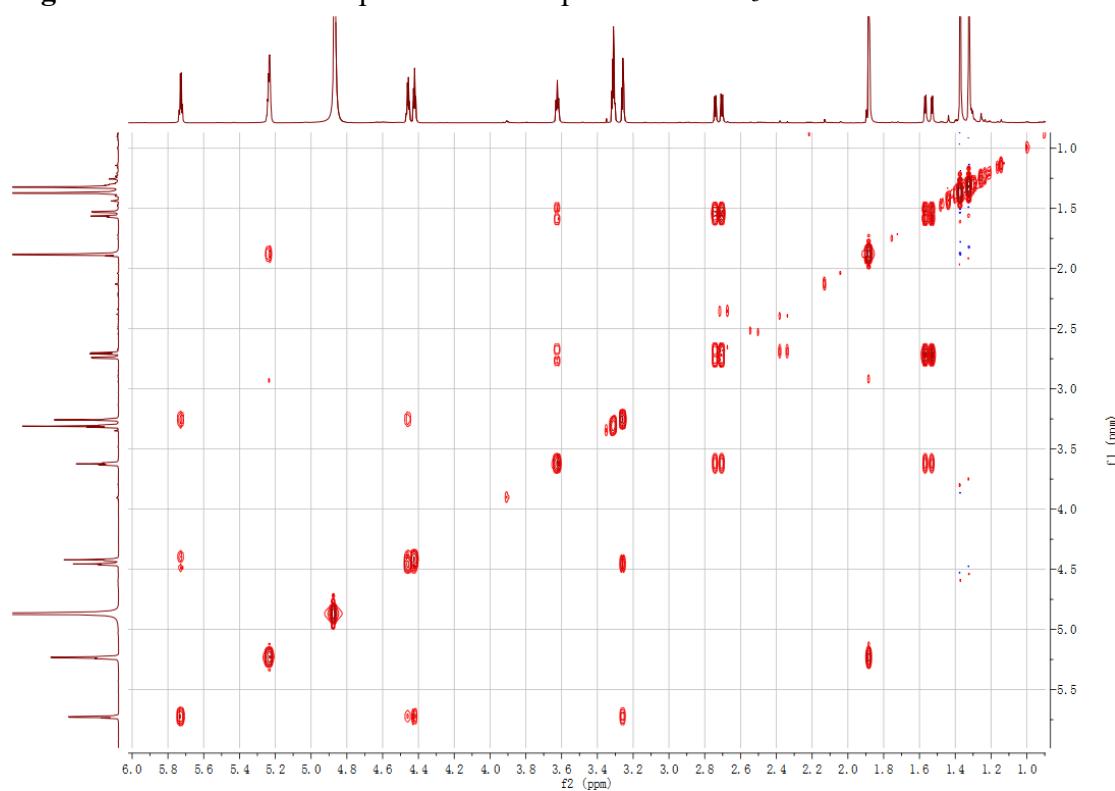


Figure S13 HSQC spectrum of compound **2** in CD₃OD

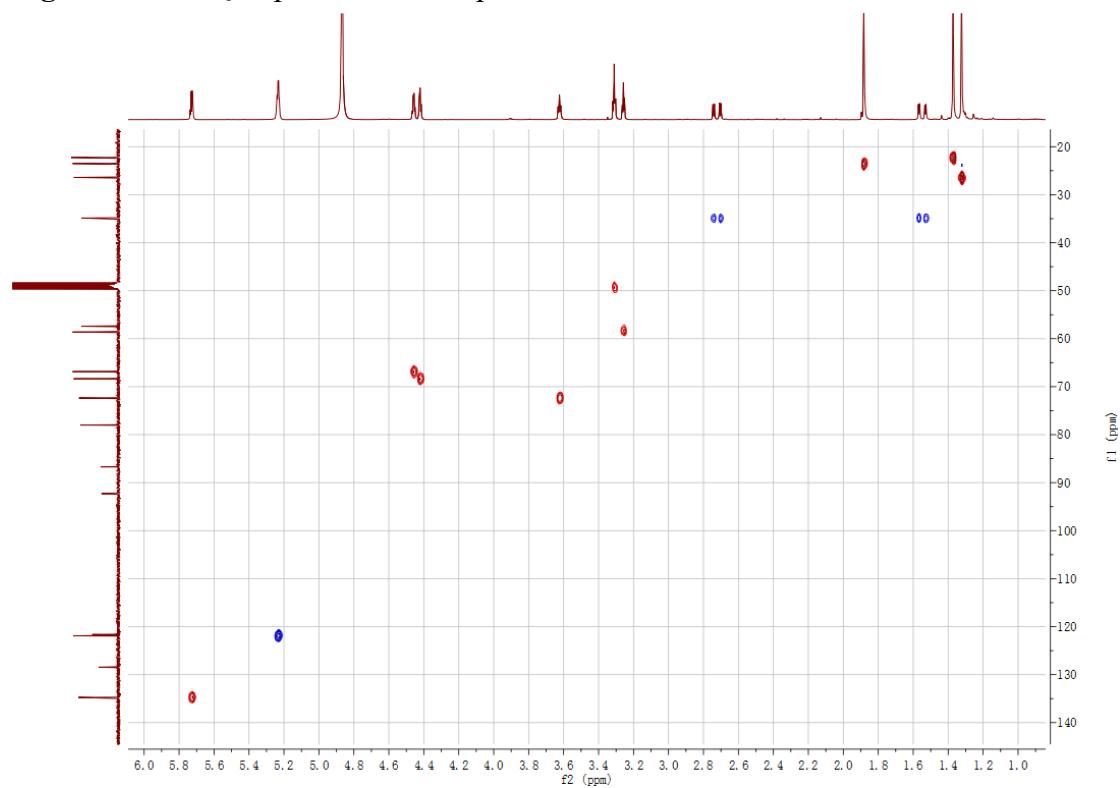


Figure S14 HMBC spectrum of compound **2** in CD₃OD

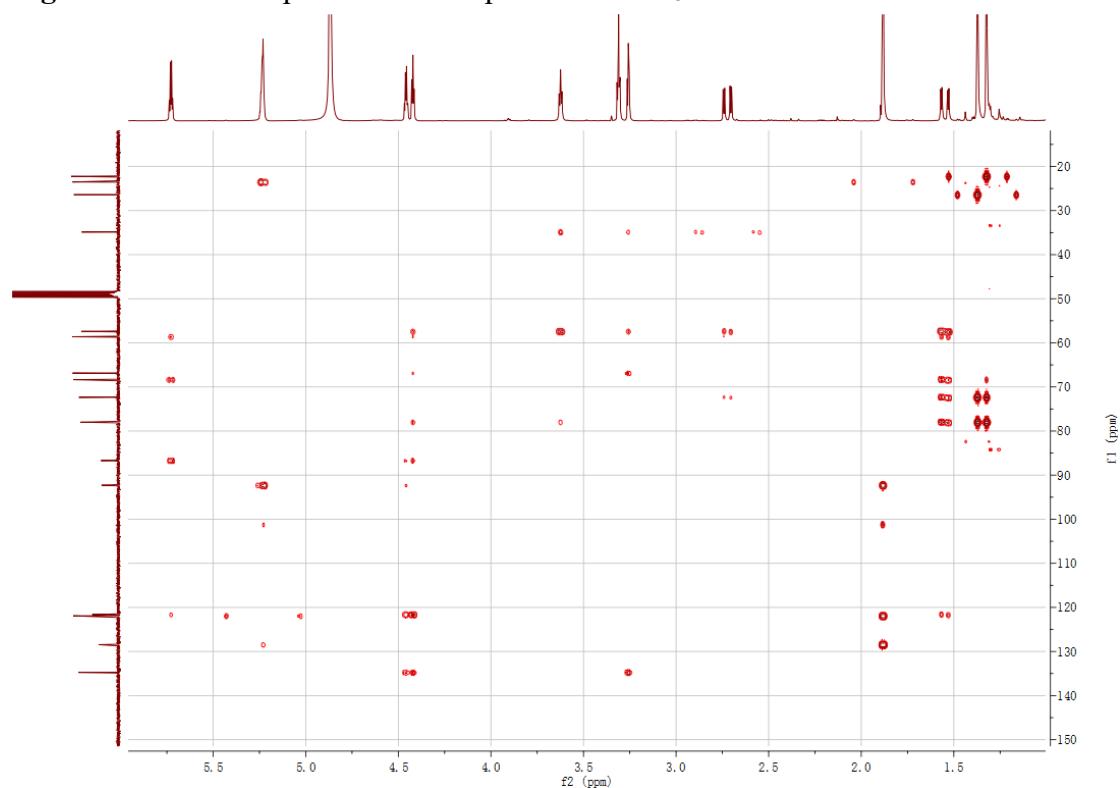


Figure S15 NOESY spectrum of compound **2** in CD₃OD

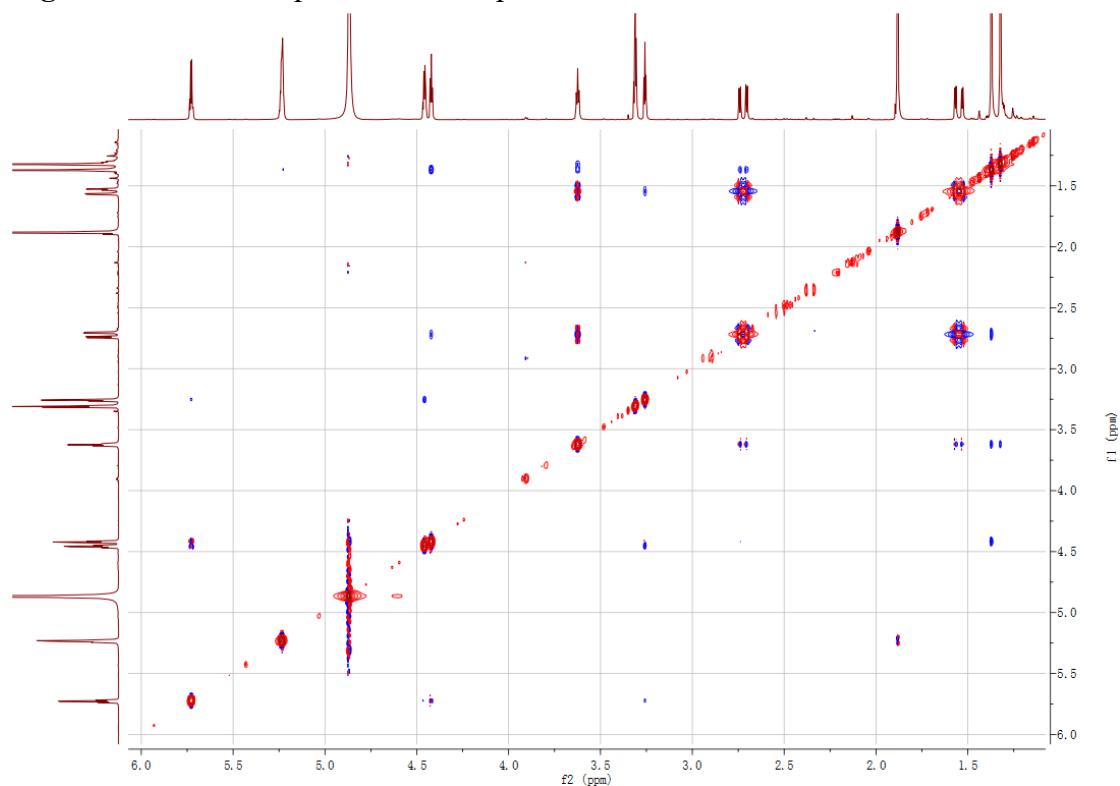


Figure S16 HR-ESI-MS spectrum of compound **2**

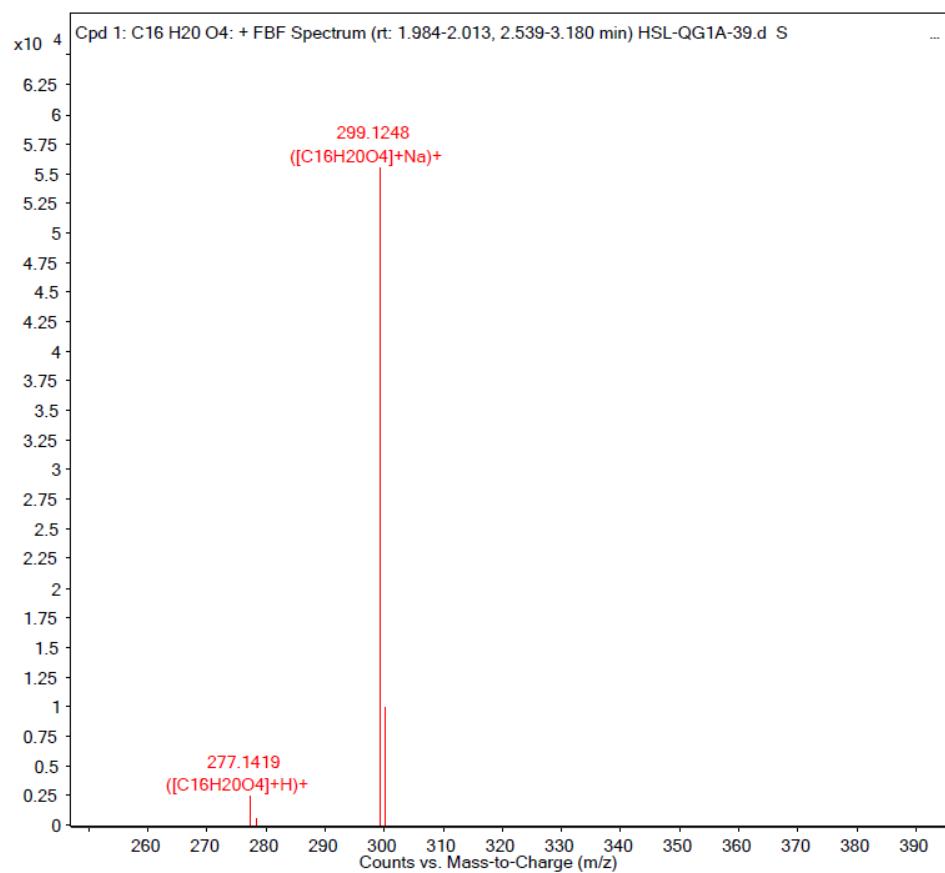


Figure S17 UV spectrum of compound 2

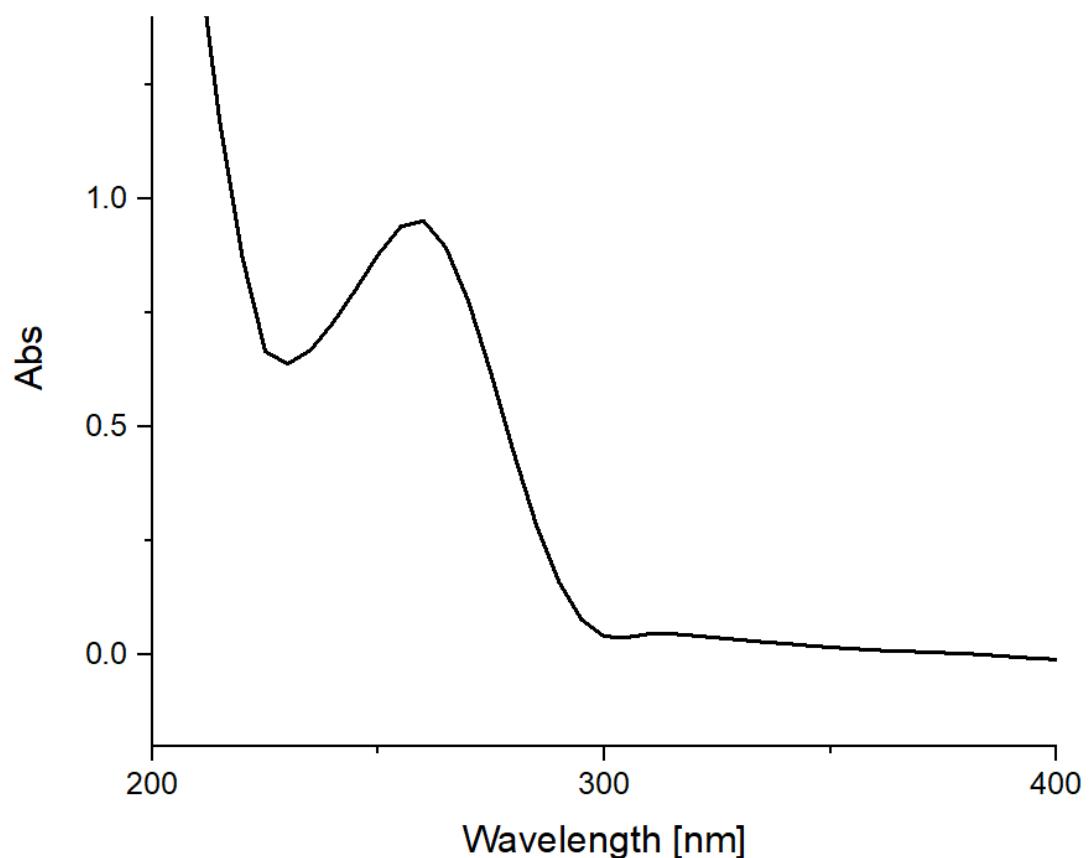


Figure S18 IR spectrum of compound 2

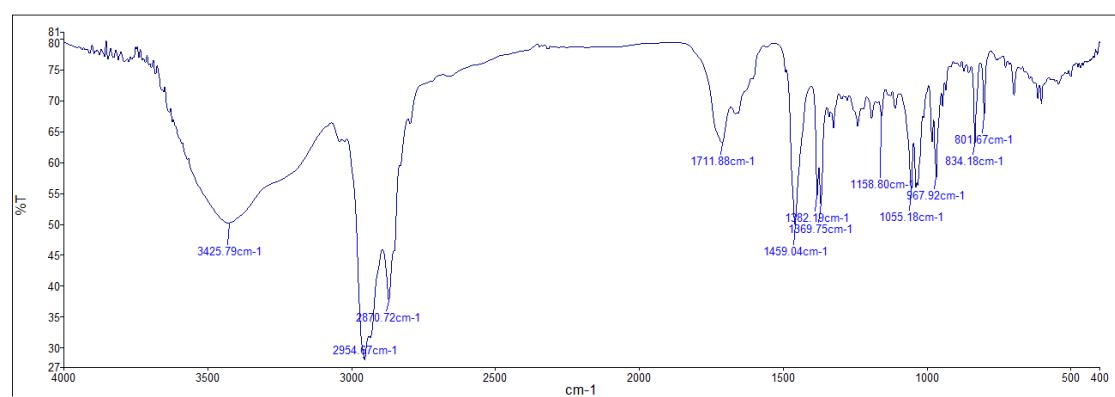


Figure S19 ^1H NMR spectrum of compound **3** in CD_3OD

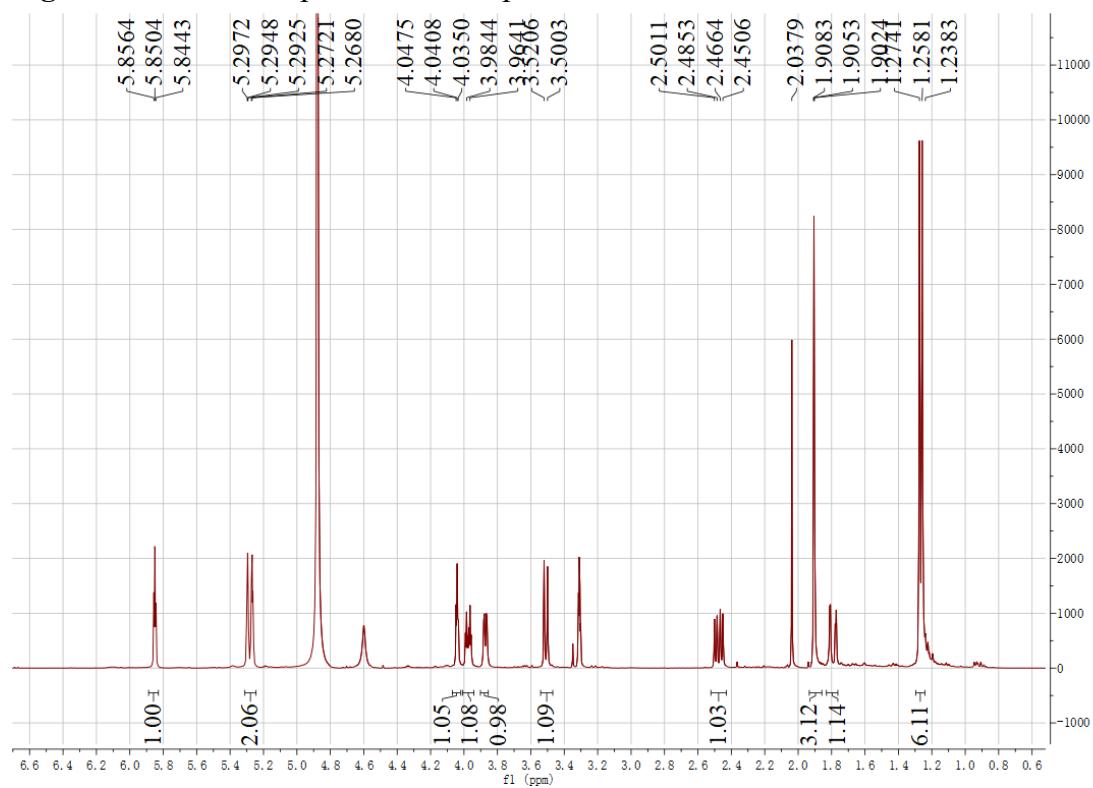


Figure S20 ^{13}C NMR spectrum of compound **3** in CD_3OD

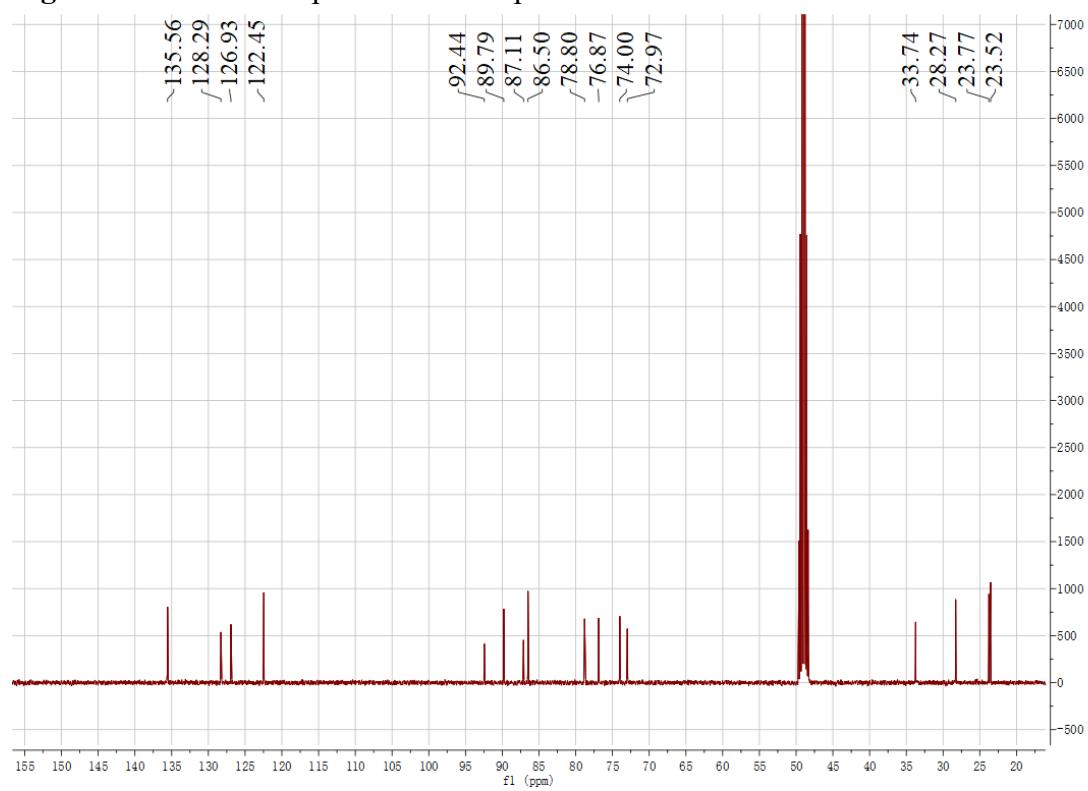


Figure S21 ^1H - ^1H COSY spectrum of compound **3** in CD_3OD

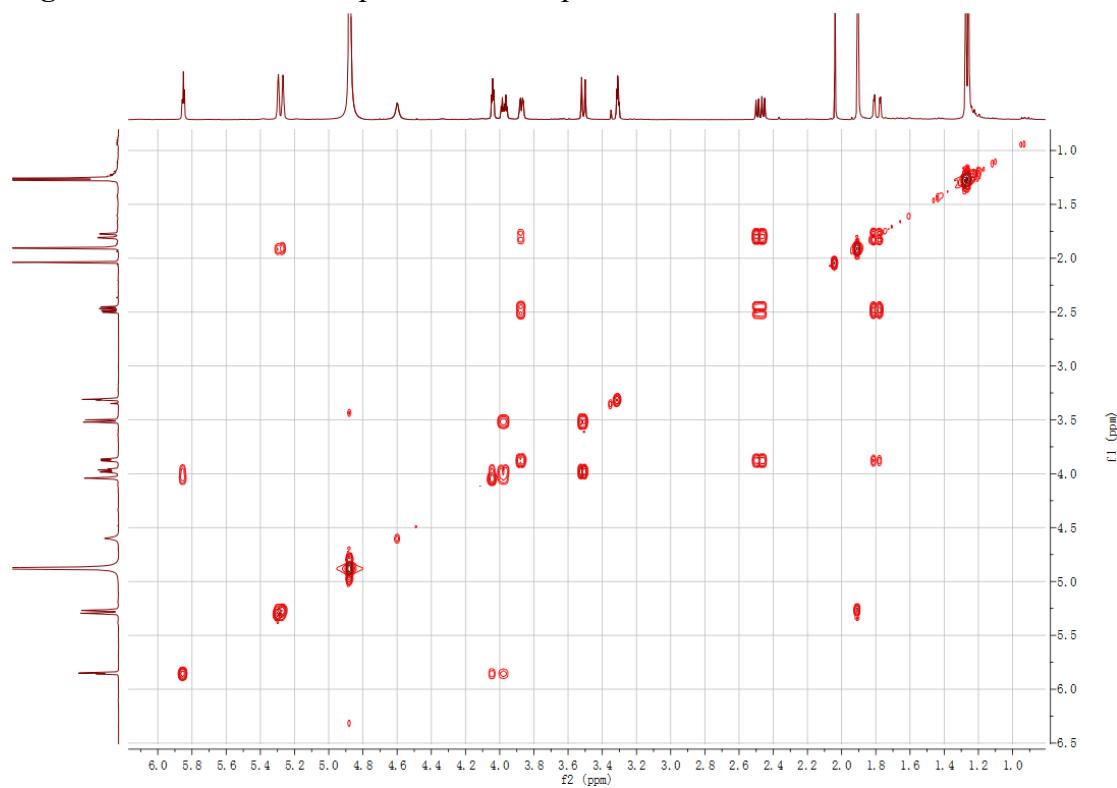


Figure S22 HSQC spectrum of compound **3** in CD_3OD

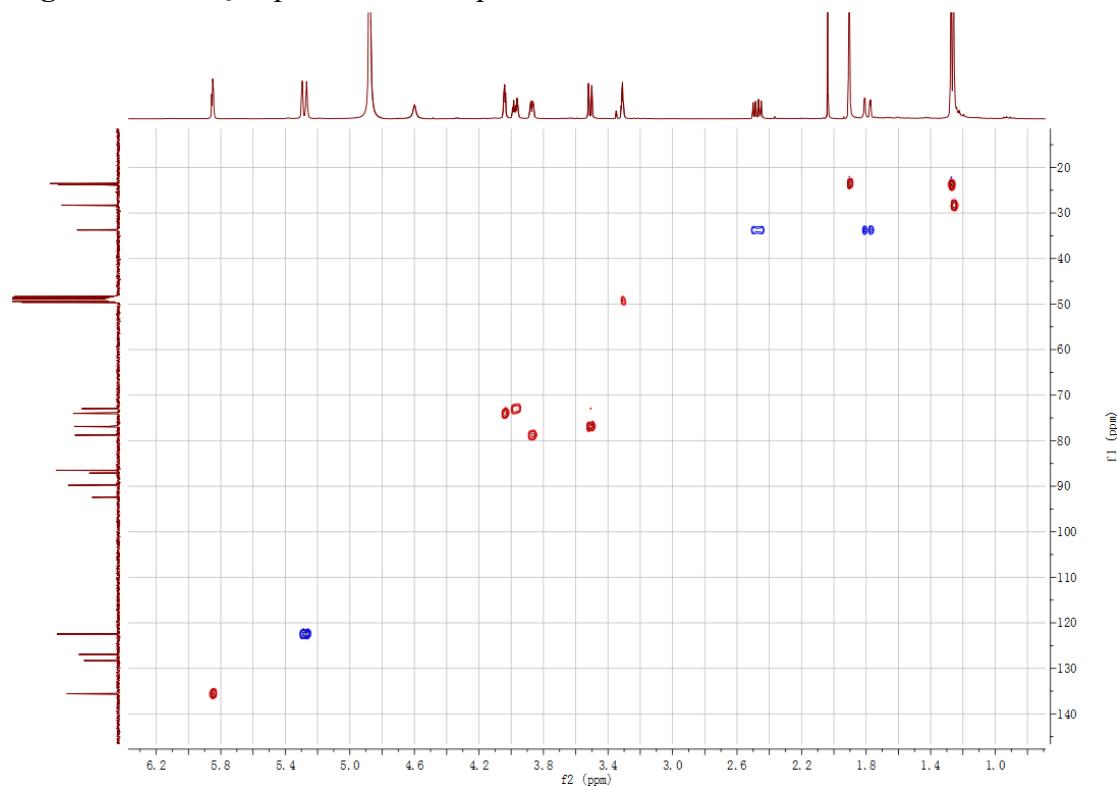


Figure S23 HMBC spectrum of compound **3** in CD₃OD

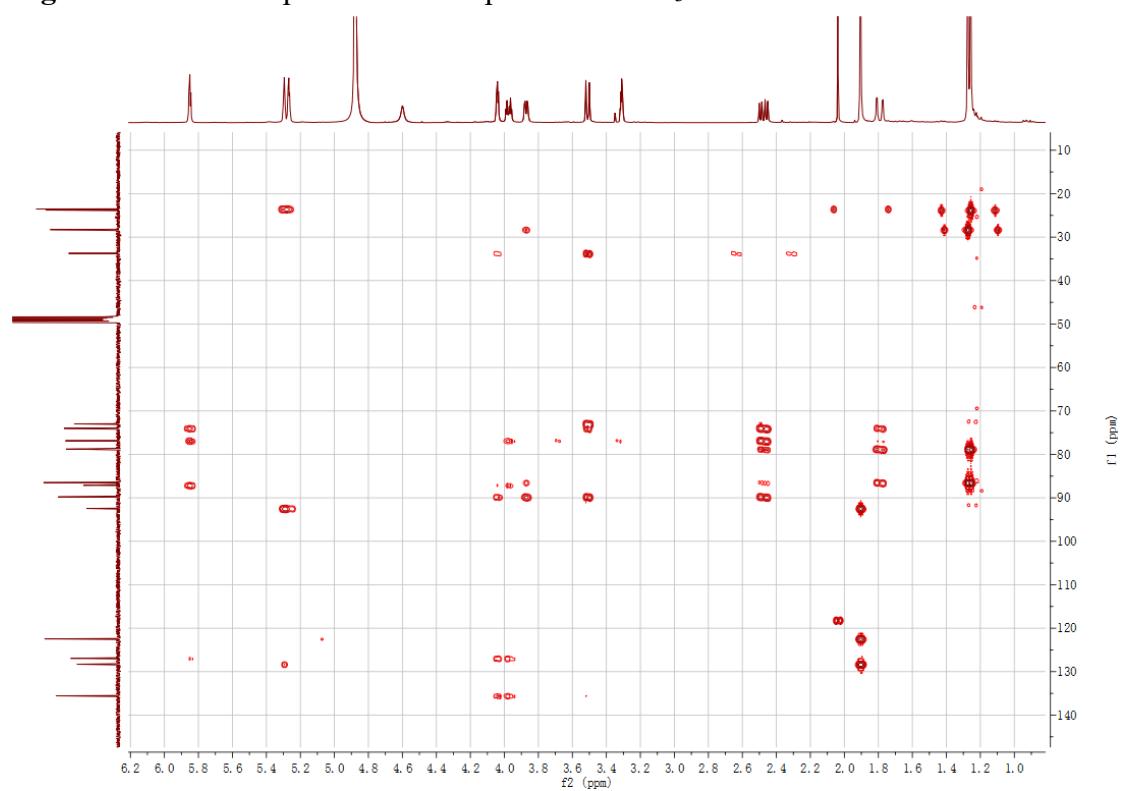


Figure S24 NOESY spectrum of compound **3** in CD₃OD

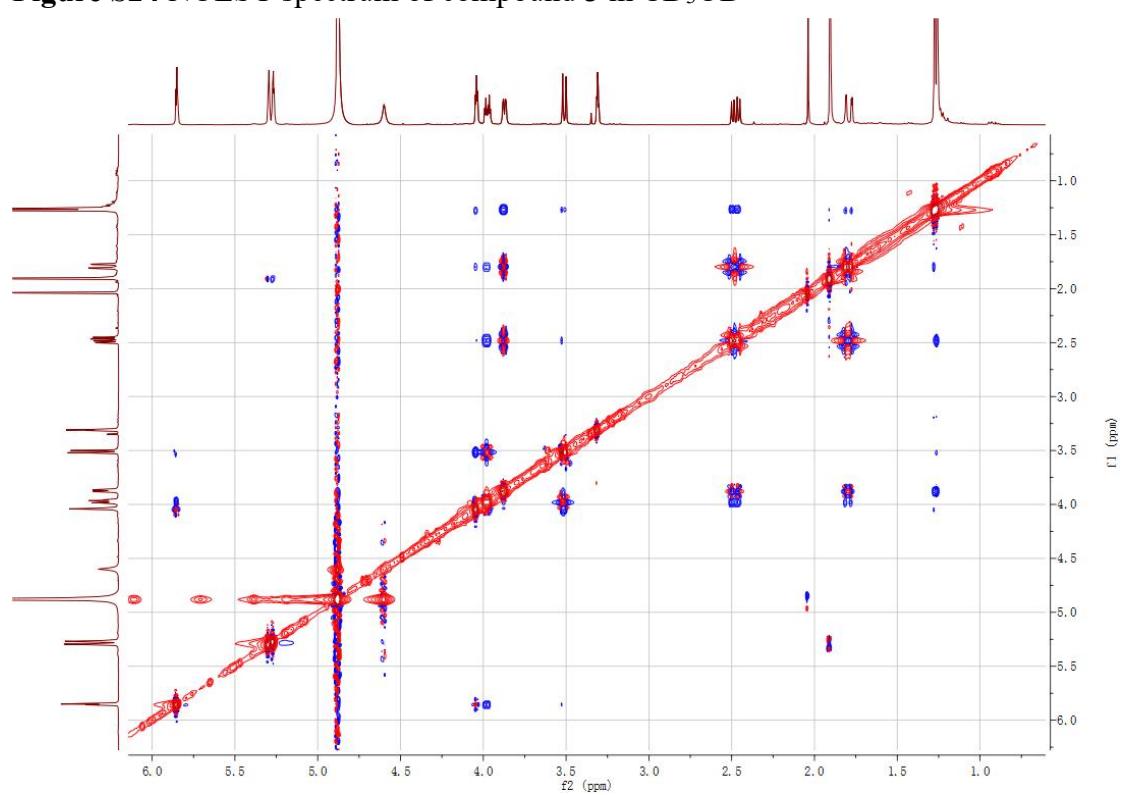


Figure S25 HR-ESI-MS spectrum of compound 3

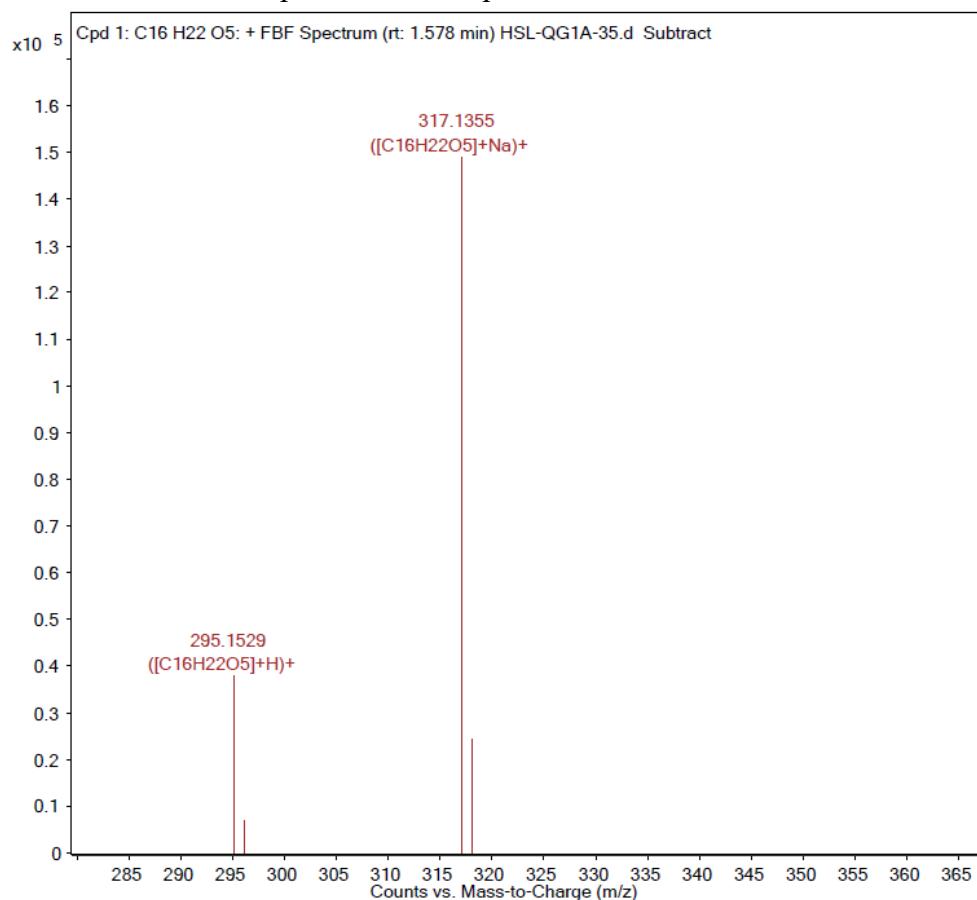


Figure S26 UV spectrum of compound 3

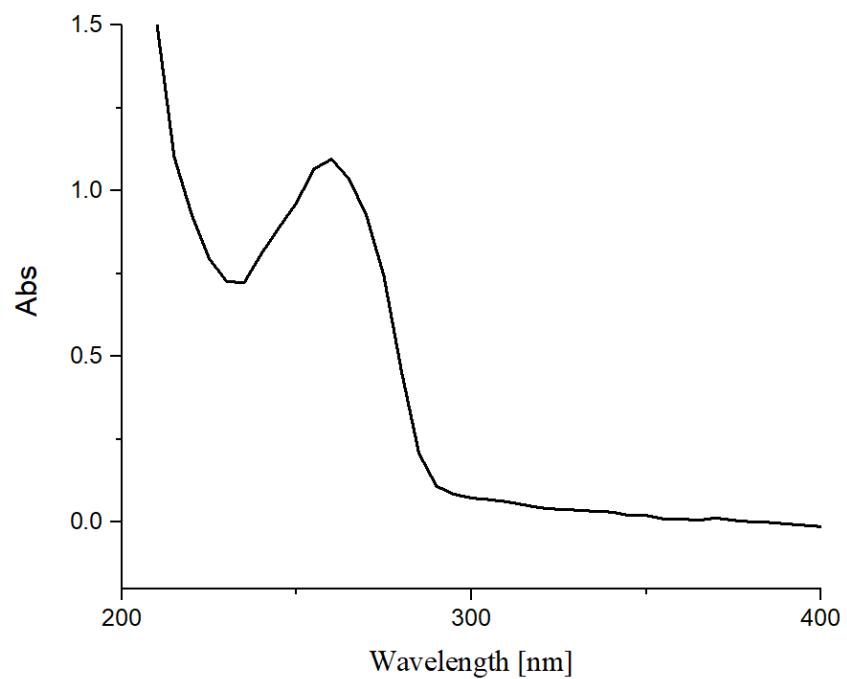


Figure S27 IR spectrum of compound 3

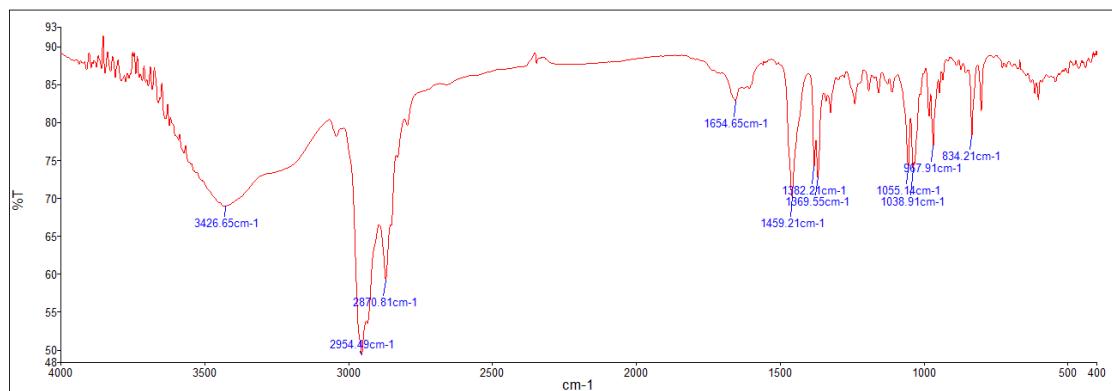


Figure S28 ^1H NMR spectrum of compound 4 in CD_3OD

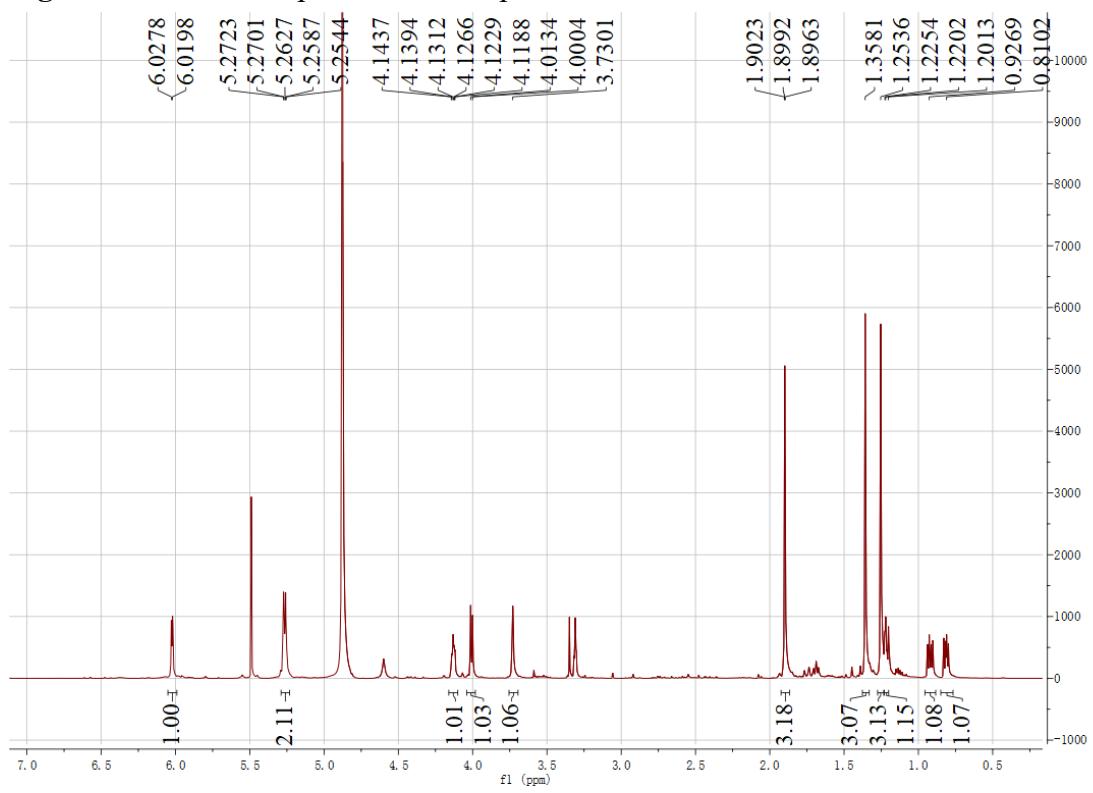


Figure S29 ^{13}C NMR spectrum of compound **4** in CD_3OD

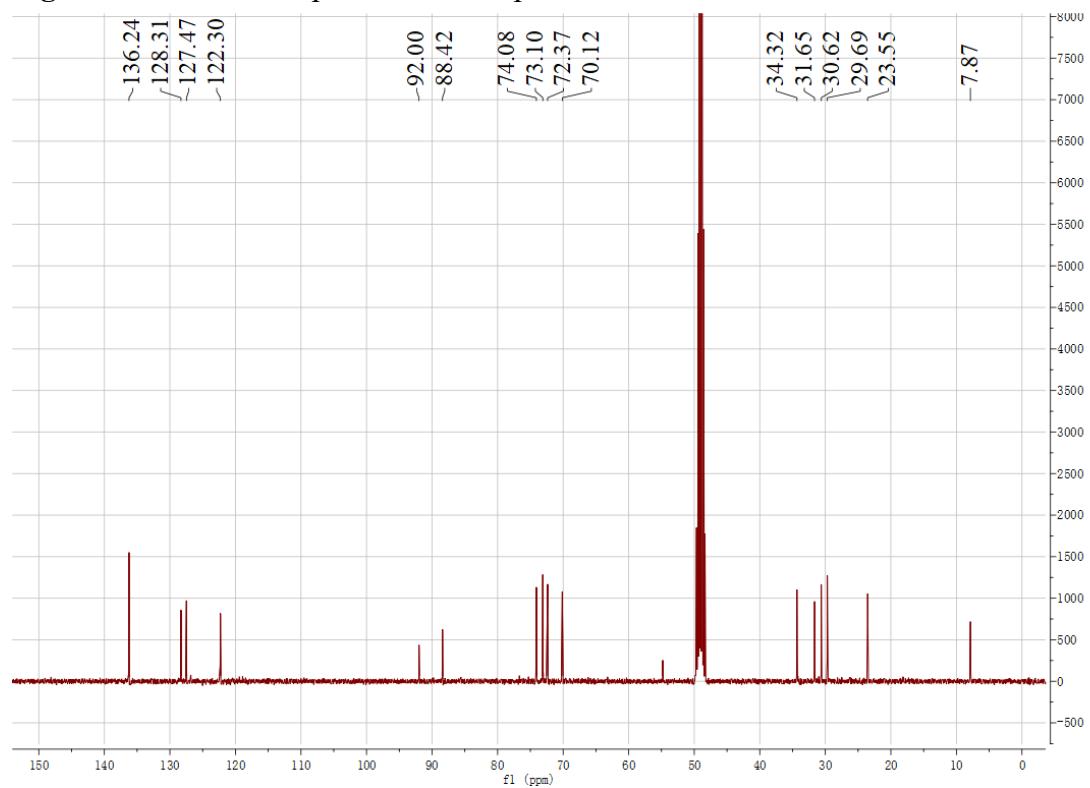


Figure S30 ^1H - ^1H COSY spectrum of compound **4** in CD_3OD

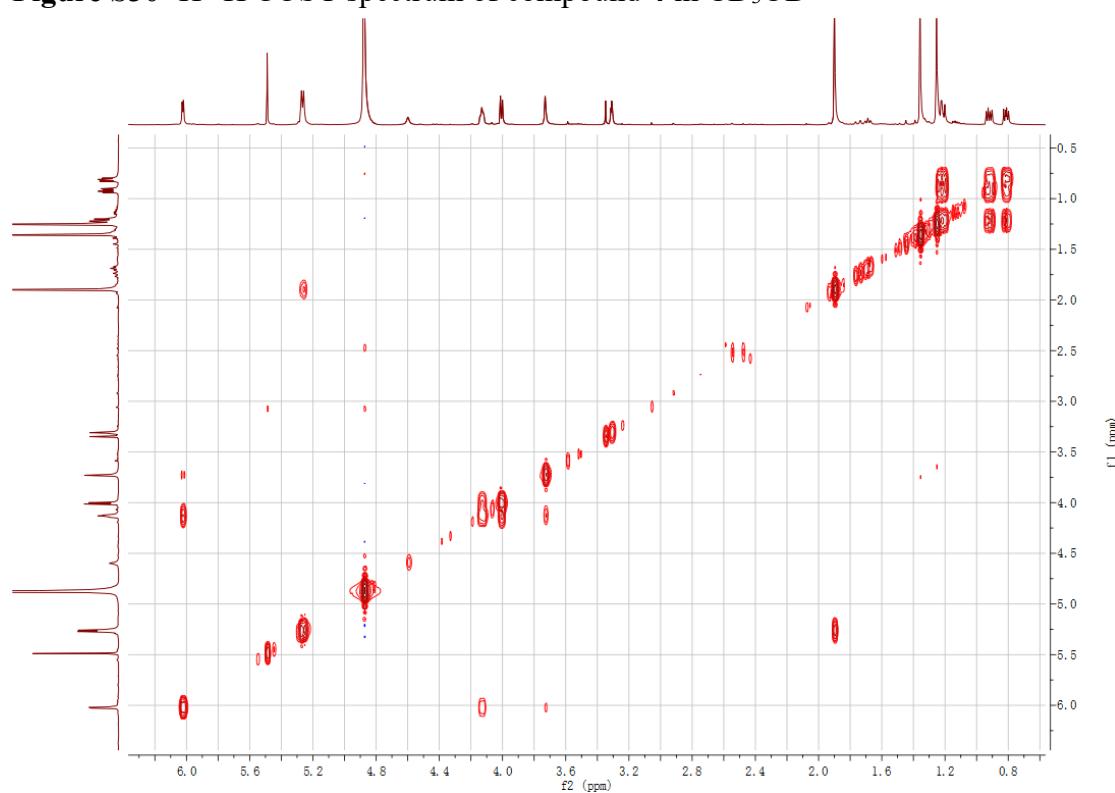


Figure S31 HSQC spectrum of compound **4** in CD₃OD

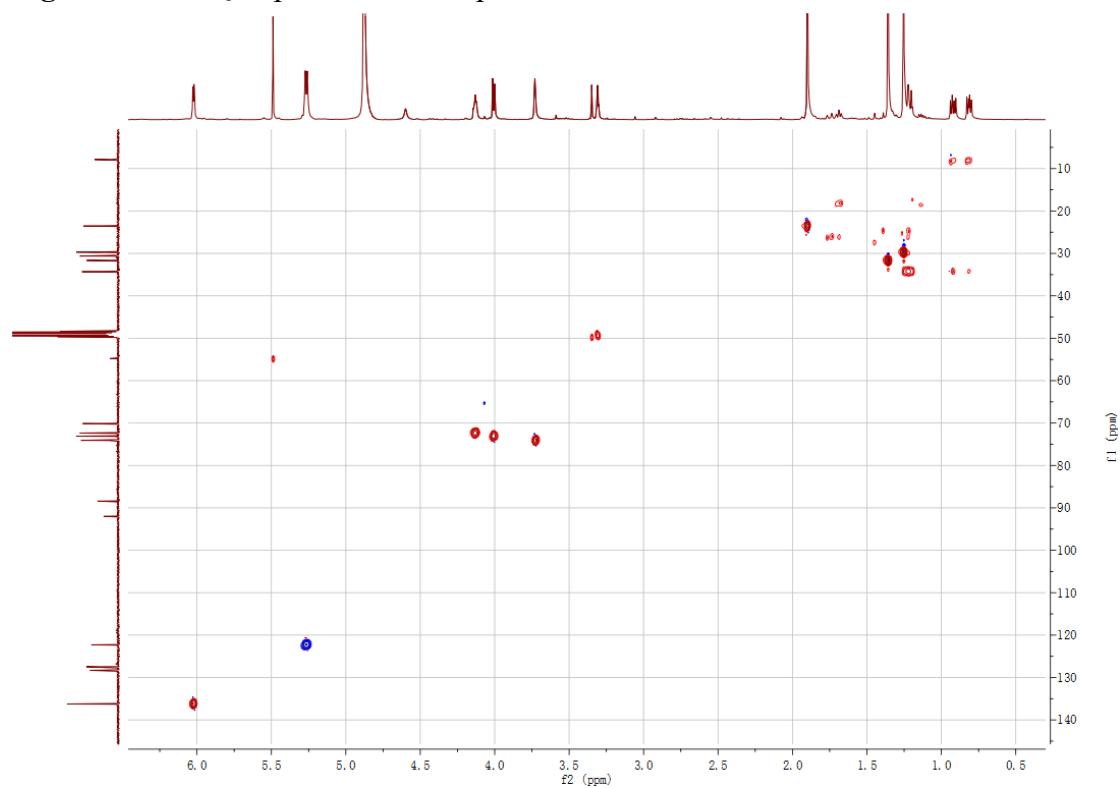


Figure S32 HMBC spectrum of compound **4** in CD₃OD

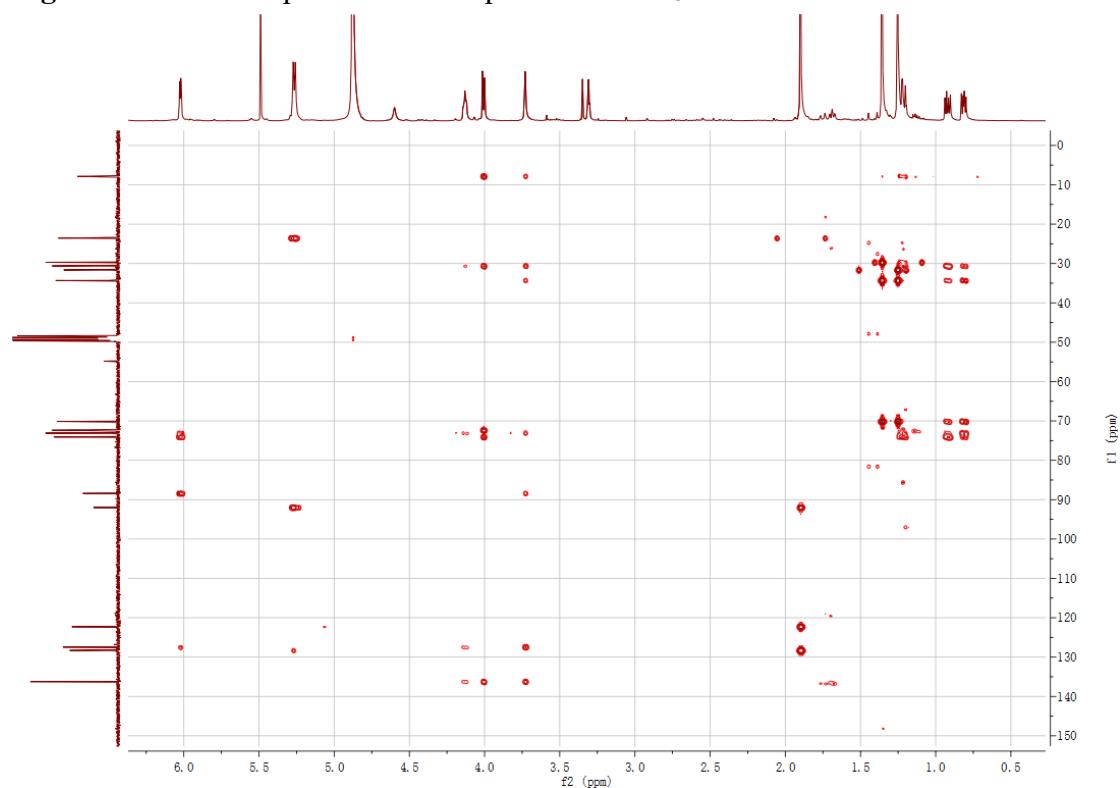


Figure S33 NOESY spectrum of compound 4 in CD₃OD

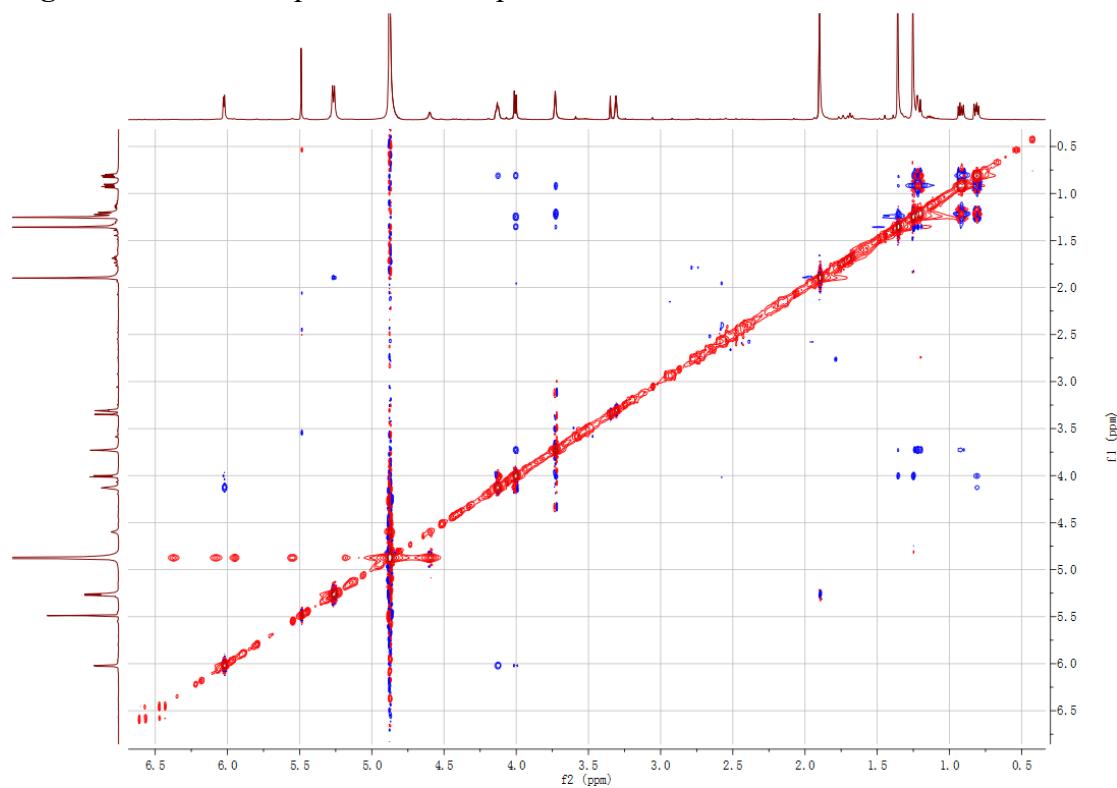


Figure S34 HR-ESI-MS spectrum of compound 4

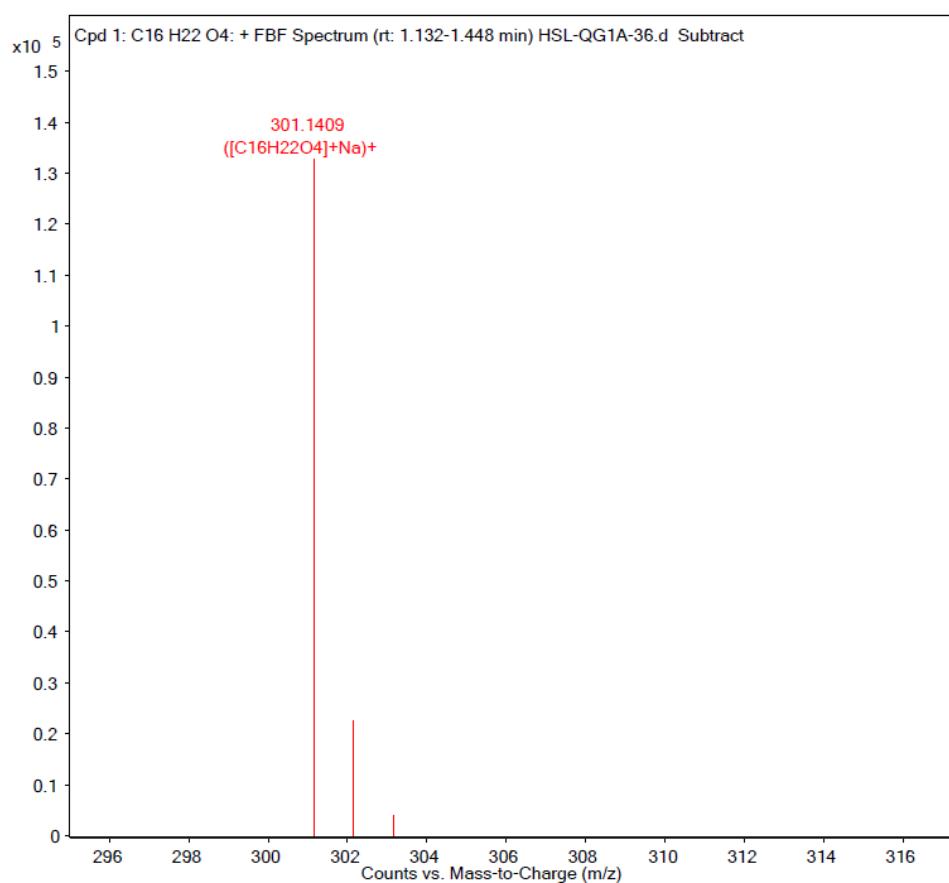


Figure S35 UV spectrum of compound 4

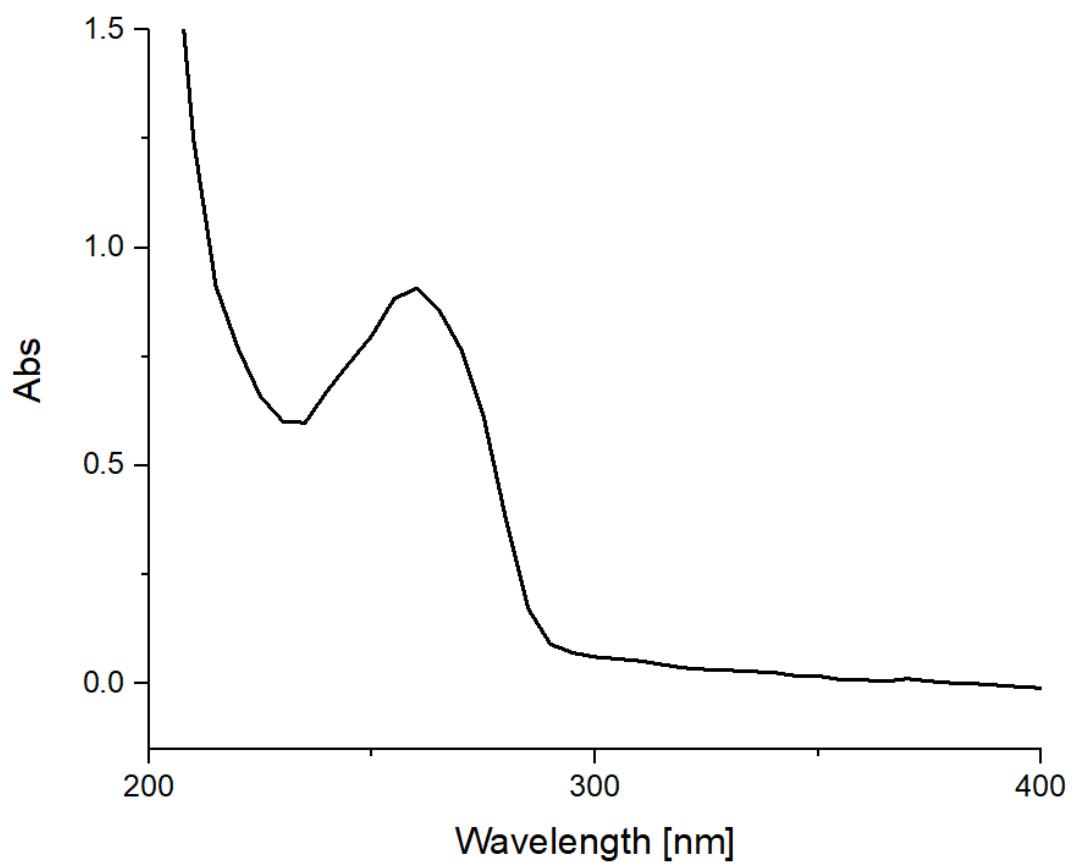


Figure S36 IR spectrum of compound 4

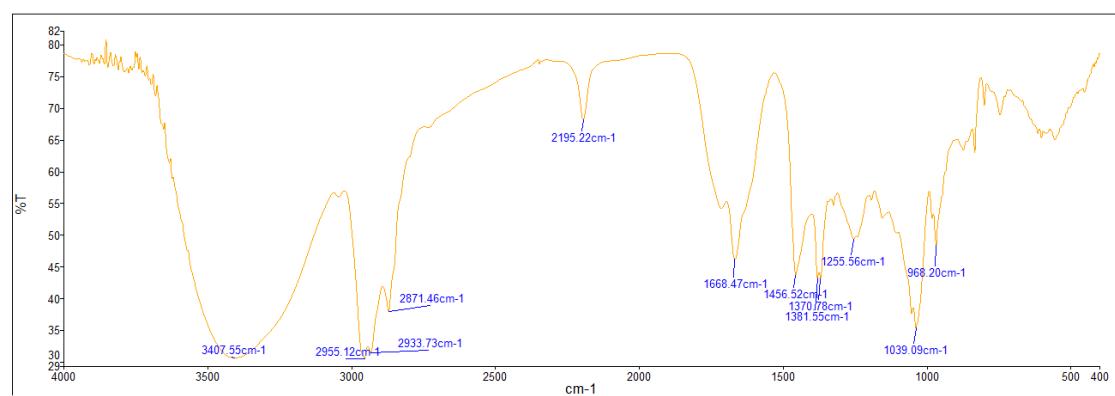


Figure S37 ^1H NMR spectrum of compound **5** in CD_3OD

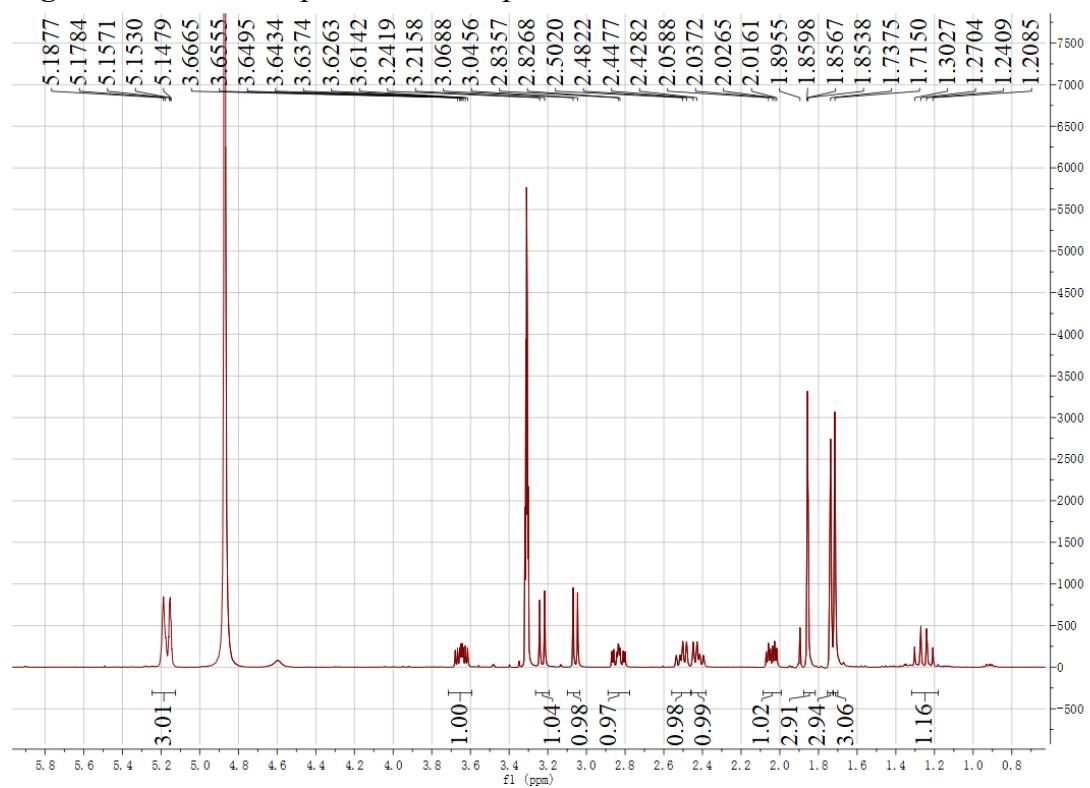


Figure S38 ^{13}C NMR spectrum of compound **5** in CD_3OD

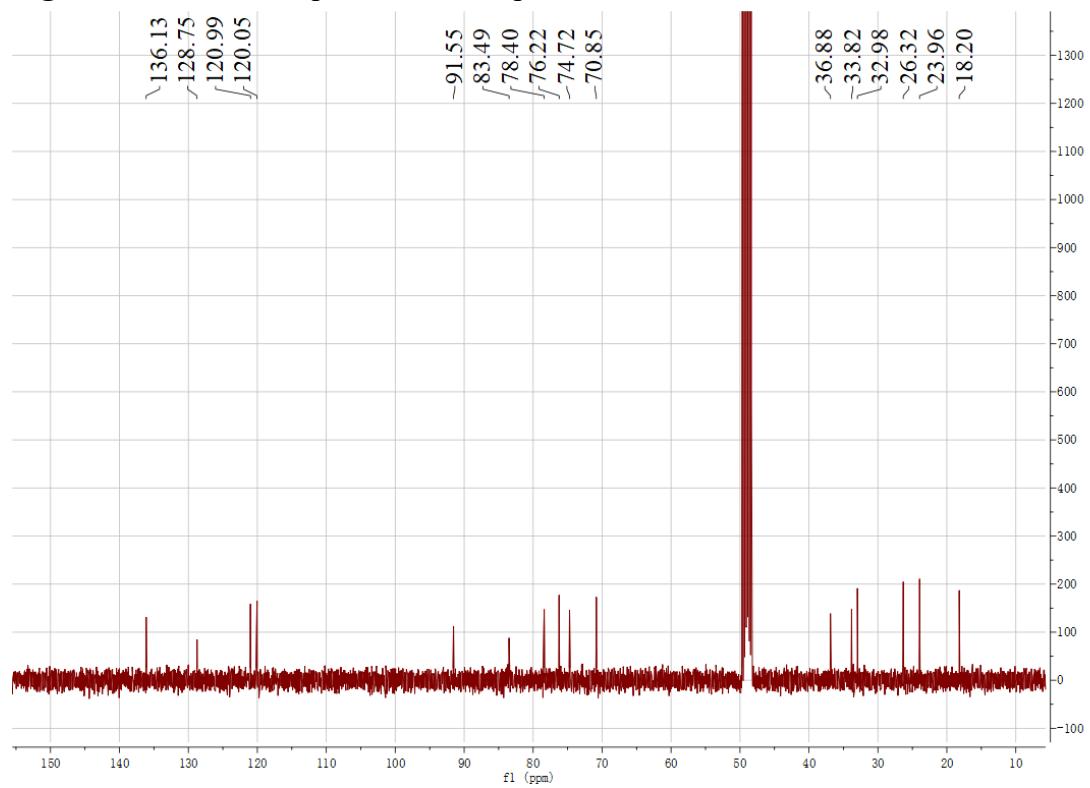


Figure S39 ^1H - ^1H COSY spectrum of compound **5** in CD_3OD

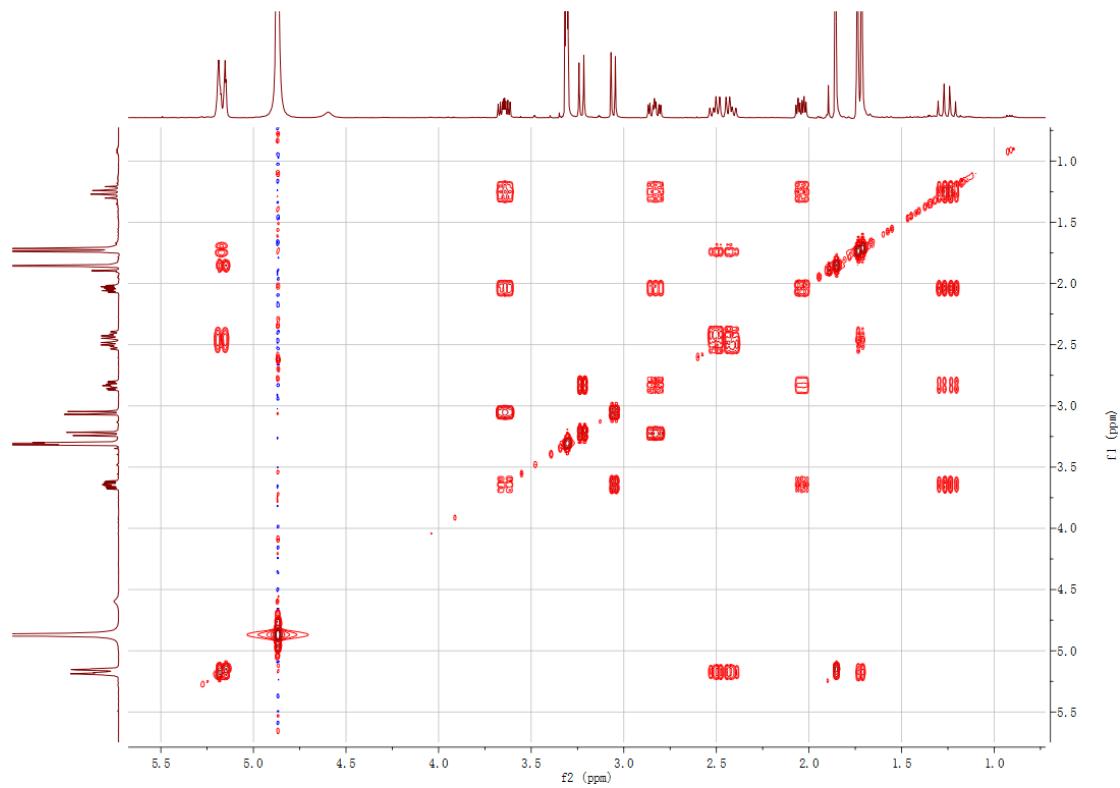


Figure S40 HSQC spectrum of compound **5** in CD_3OD

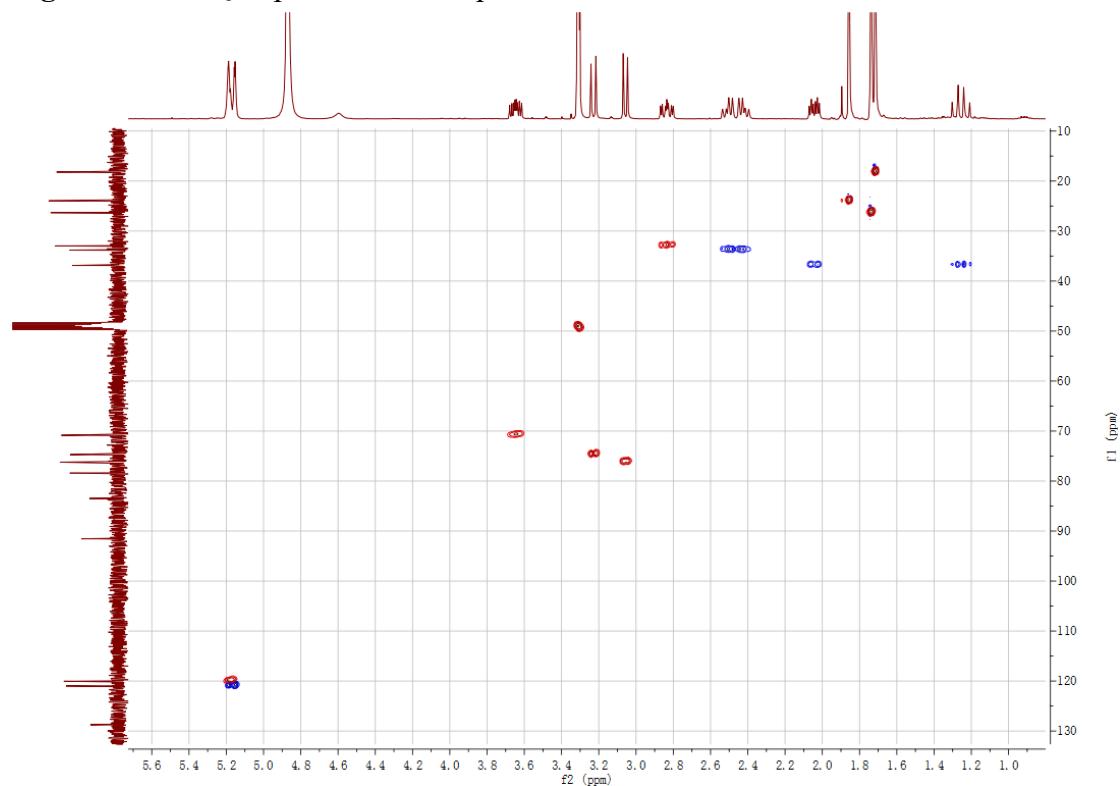


Figure S41 HMBC spectrum of compound **5** in CD₃OD

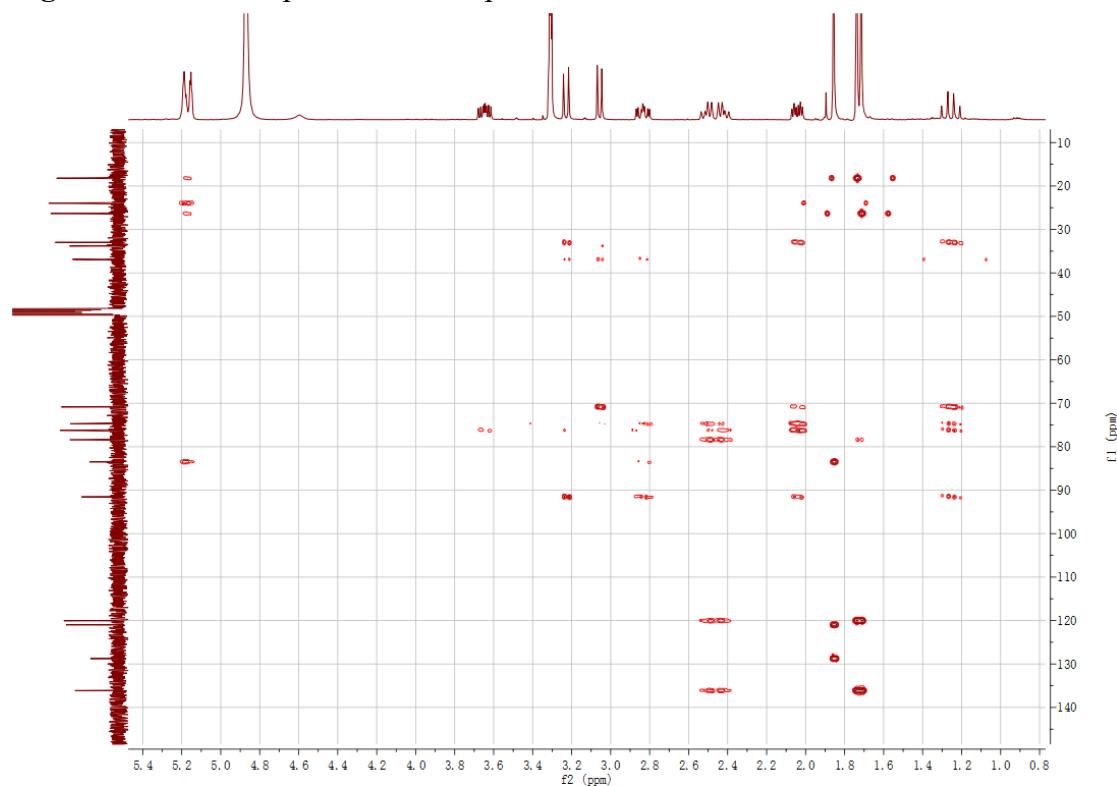


Figure S42 NOESY spectrum of compound **5** in CD₃OD

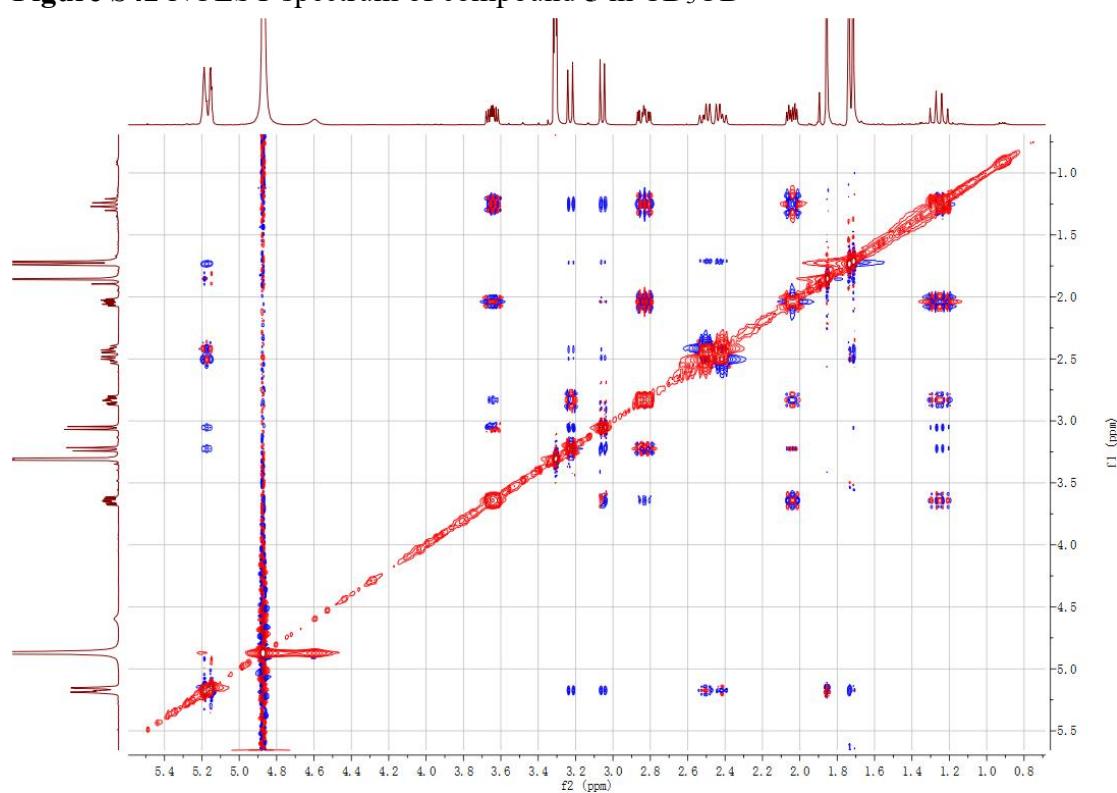


Figure S43 HR-ESI-MS spectrum of compound 5

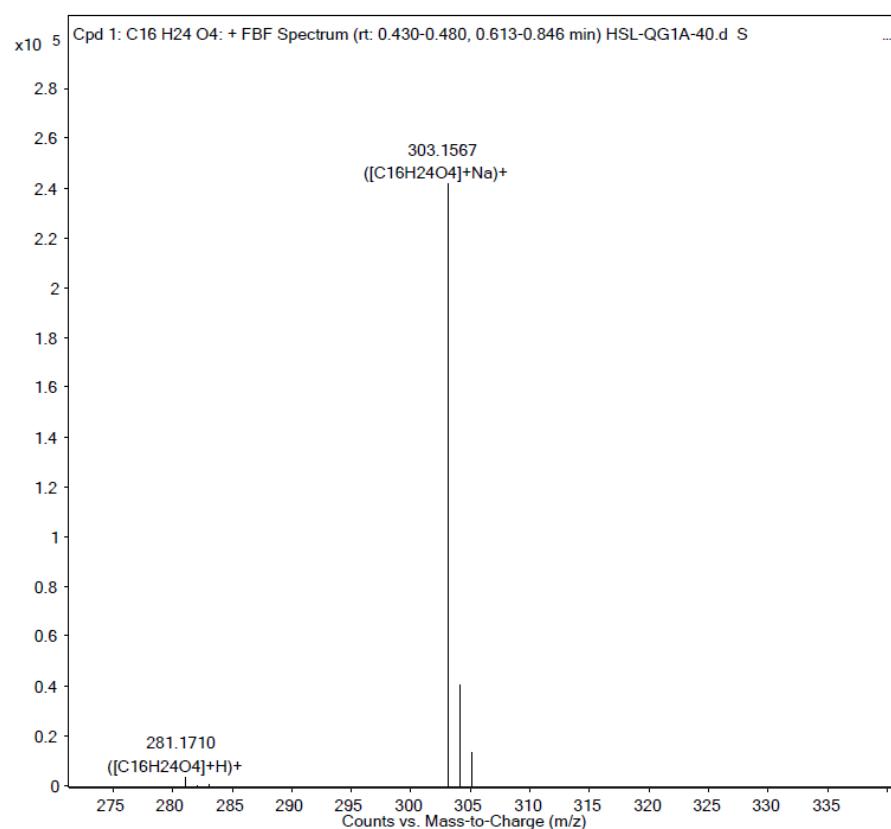


Figure S44 UV spectrum of compound 5

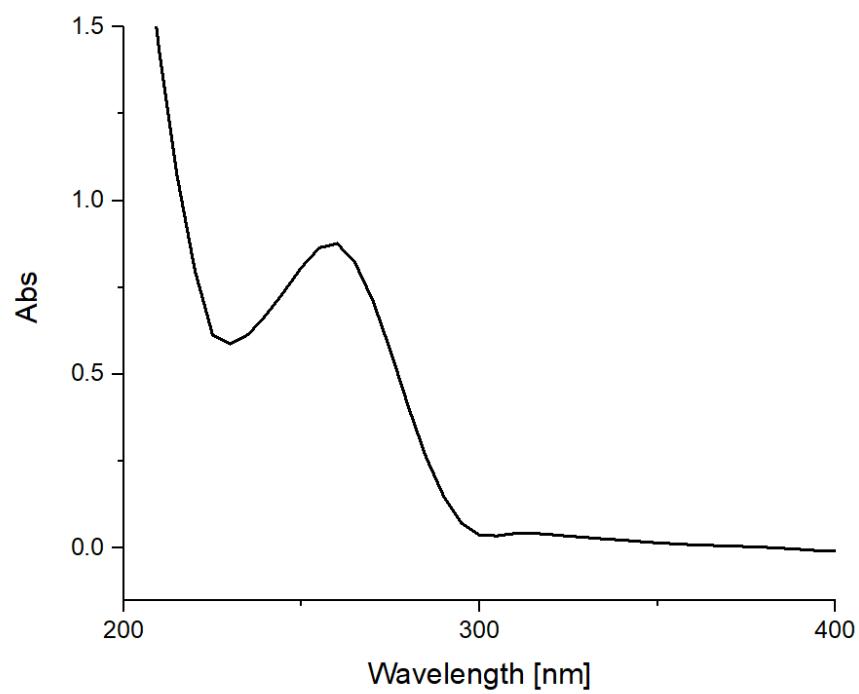
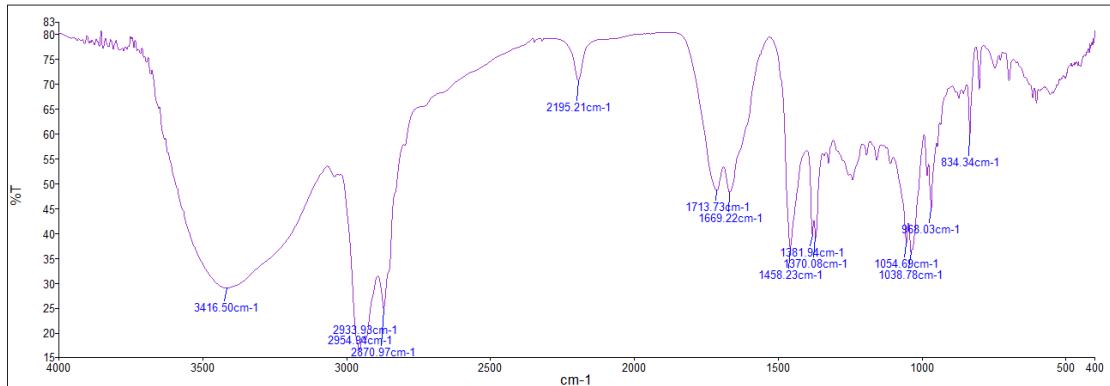


Figure S45 IR spectrum of compound 5



Computational Section

ECD calculation

Conformational analysis was initially performed using Confab at MMFF94 force field for new compound. Room-temperature equilibrium populations were calculated according to Boltzmann distribution law. The conformers with Boltzmann-population from above 1% were chosen for ECD calculations. The chosen conformer was optimized at B3LYP/6-311G** using Density functional theory (DFT). Then, it was further optimized in gas phase using the CPCM polarizable conductor calculation model. The theoretical calculation of ECD was conducted using Time-dependent DFT (TD-DFT) method at B3LYP/6-311G* in methanol. Rotatory strengths for total excited states were calculated. The ECD spectrum is simulated in SpecDis by overlapping Gaussian functions for each transition according to:

$$\Delta\epsilon(E) = \frac{1}{2.297 \times 10^{-39}} \times \frac{1}{\sqrt{2\pi}\sigma} \sum_i^A \Delta E_i R_i e^{-\left(\frac{E-E_i}{2\sigma}\right)^2}$$

where σ represents the width of the band at $1/e$ height, and ΔE_i and R_i are the excitation energies and rotatory strengths for transition i , respectively. $\sigma = 0.2\text{-}0.3$ eV and UV-

Shift = 5 nm and R^{velocity} have been used in this work.

Table S1. Energies of the dominative conformers of compounds **1-5**.

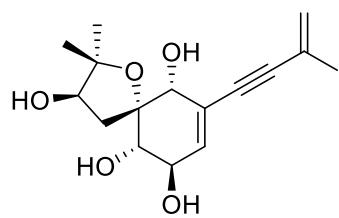
Compounds	No	Structure	E (Hartree)	Population (%)
(1 <i>R</i> , 2 <i>S</i> , 3 <i>R</i> , 4 <i>R</i> , 13 <i>R</i>)- 1	1		-922.53787494	70.79
	2		-922.53685018	29.21
(1 <i>R</i> , 2 <i>S</i> , 3 <i>R</i> , 4 <i>R</i> , 13 <i>R</i>)- 2	1		-922.53356696	56.25
	2		-922.53351014	43.75
(1 <i>R</i> , 2 <i>R</i> , 3 <i>S</i> , 4 <i>R</i> , 13 <i>R</i>)- 3	1		-999.02684494	56.03
	2		-999.02543941	43.97
(1 <i>R</i> , 2 <i>S</i> , 3 <i>R</i> , 4 <i>S</i> , 12 <i>R</i>)- 4	1		-923.74831245	54.12

	2		-923.74788993	39.44
	3		-923.74520715	6.43
(1 <i>S</i> , 2 <i>S</i> , 3 <i>R</i> , 4 <i>S</i> , 6 <i>S</i>)-5	1		-924.97692936	20.84
	2		-924.97677915	20.76
	3		-924.97666752	29.42
	4		-924.97722919	28.98

¹³C NMR calculation

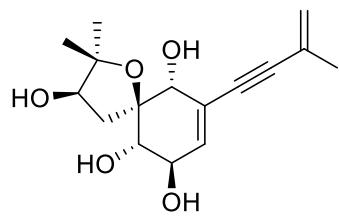
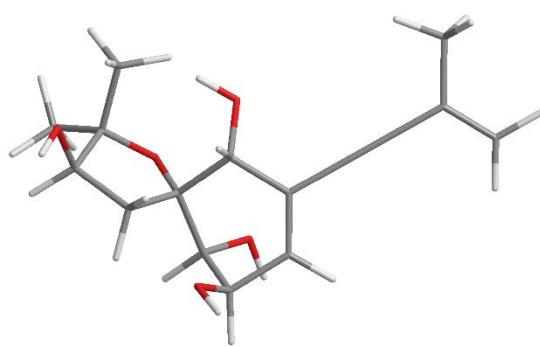
The ¹³C NMR calculation of **3a** and **3b** were calculated with the GIAO method at the MPW1PW91/6-311+G(d,p) level with PCM. The shielding constants (σ) were converted into chemical shifts (δ) by referencing to TMS at 0 ppm ($\delta_{\text{cal}} = \sigma_{\text{TMS}} - \sigma_{\text{cal}}$), where the σ_{TMS} was the shielding constant of TMS calculated at the same level. The calculated chemical shifts were directly performed statistical analyses with experimental chemical shifts by using DP4+ method. The DP4+ probability, and linear correlation coefficients (R^2), were adopted for evaluation of the results.

Figure S46 Conformations of low-energy conformers of structures **3a and **3b**.**



1*R*, 2*R*, 3*S*, 4*R*, 13*R*

3a



1*R*, 2*S*, 3*S*, 4*R*, 13*R*

3b

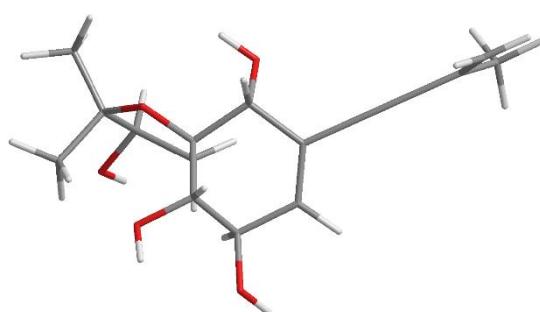


Table S2 DP4+ analysis result table of model compounds **3a** and **3b**.

NO	Experimental	3a	3b
1	135.6	139.9442	139.6799
2	128.3	129.6029	129.5934
3	126.9	126.296	126.5934
4	122.5	122.8981	125.6672
5	92.4	95.1603	95.5666
6	89.8	88.5949	87.1109
7	87.1	86.4094	86.9531
8	86.5	85.6815	86.8295
9	78.8	76.2249	76.3838
10	76.9	75.9158	73.4216
11	74	71.5636	70.2743
12	73	71.7203	68.385
13	33.7	33.2928	29.5577
14	28.3	23.137	20.9278
15	23.8	19.9035	19.7076
16	23.5	16.9438	16.3642

Figure S47 DP4+ probability analysis result.

A	B	C	D	E	F	G	H
Functional	Solvent?		Basis Set		Type of Data		
mPW1PW91	PCM		6-311+G(d, p)		Unscaled Shifts		
	DP4+	100.00%	0.00%	-	-	-	
Nuclei	sp2?	Experimental	Isomer 1	Isomer 2	Isomer 3	Isomer 4	Isomer 5
C	x	135.6	139.9442	139.6799			
C	x	128.3	129.6029	129.5934			
C	x	126.9	126.296	126.5934			
C	x	122.5	122.8981	125.6672			
C		92.4	95.1603	95.5666			
C		89.8	88.5949	87.1109			
C		87.1	86.4094	86.9531			
C		86.5	85.6815	86.8295			
C		78.8	76.2249	76.3838			
C		76.9	75.9158	73.4216			
C		74	71.5636	70.2743			
C		73	71.7203	68.385			
C		33.7	33.2928	29.5577			
C		28.3	23.137	20.9278			
C		23.8	19.9035	19.7076			
C		23.5	16.9438	16.3642			

Figure S48 Regression analyses of experimental versus calculated ¹³C NMR chemical shifts of model compounds **3a** and **3b**

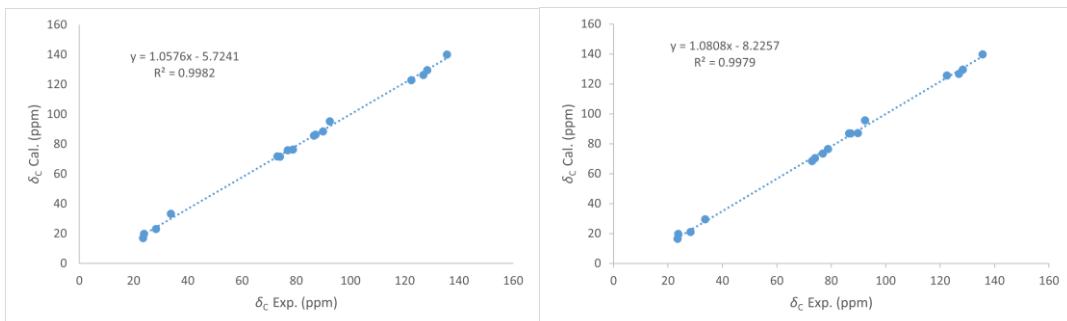


Table S3. ¹H (400 MHz) and ¹³C (100 MHz) NMR, and COSY, HMBC and NOESY assignment of **1** in CD₃OD.

No	1				
	δ_{C} , type	δ_{H} (<i>J</i> in Hz)	COSY	HMBC	NOESY
1	67.8, CH	4.46, t (2.0)		2, 5, 6, 7, 14	12 α , 15
2	61.0, C				
3	60.1, CH	3.26, t (1.8)	4	12	12 β
4	64.7, CH	4.41, dt (5.3, 1.8)	3, 5	2, 6	
5	133.2, CH	5.93, dt (5.3, 2.0)	4	1, 7	
6	123.3, C				
7	86.8, C				
8	93.3, C				
9	128.3, C				

10	122.3, CH ₂	5.26, m		8, 9, 11	
11	23.5, CH ₃	1.89, m		8, 9, 10	
12 α	36.3, CH ₂	2.27, dd (13.3, 12.0)	13	2, 3	1, 15
12 β	36.3, CH ₂	1.57, dd (13.3, 5.0)	13	2, 3	3
13	73.2, CH	3.70, dd (12.0, 5.0)	12	14, 15	16
14	78.6, C				
15	16.5, CH ₃	1.30, s		13	1, 12 α
16	28.2, CH ₃	1.33, s		1, 14	13

Table S4. ^1H (400 MHz) and ^{13}C (100 MHz) NMR, and COSY, HMBC and NOESY assignment of **2** in CD₃OD.

No	2				
	δ_{C} , type	δ_{H} (<i>J</i> in Hz)	COSY	HMBC	NOESY
1	68.4, CH	4.42, t (2.4)		2, 5, 6, 7, 14	12 α , 15
2	57.4, C				
3	58.6, CH	3.26, t (2.3)	4	2, 12	12 β
4	66.9, CH	4.46, dd (4.5, 2.3)	3, 5	6	
5	134.7, CH	5.73, dd (4.5, 2.4)	4	1, 6, 7	
6	121.6, C				
7	86.7, C				
8	92.3, C				
9	128.5, C				
10	121.9, CH ₂	5.24, m		8, 9, 11	
11	23.5, CH ₃	1.88, m		8, 9	
12 α	34.9, CH ₂	2.72, dd (14.8, 3.3)	13	2, 3, 14	1, 15
12 β	34.9, CH ₂	1.55, dd (14.8, 2.7)	13	2, 3, 14	3
13	72.4, CH	3.62, dd (3.3, 2.7)	12	14	15
14	78.0, C				
15	26.4, CH ₃	1.32, s		14	1, 12 α , 13
16	22.3, CH ₃	1.37, s		14	

Table S5. ^1H (400 MHz) and ^{13}C (100 MHz) NMR, and COSY, HMBC and NOESY assignment of **3** in CD₃OD.

No	3				
	δ_{C} , type	δ_{H} (<i>J</i> in Hz)	COSY	HMBC	NOESY
1	74.0, CH	4.04, t (2.5)		2, 5, 6, 12	3, 12 β
2	89.8, C				
3	76.9, CH	3.51, d (8.1)	4	1, 2, 12	1
4	73.0, CH	3.97, dt (8.1, 2.8)	3, 5	2	12 α , 13, 15
5	135.6, CH	5.85, dd (2.8, 2.5)	4	1, 6, 7	
6	126.9, C				
7	87.1, C				
8	92.4, C				
9	128.3, C				

10	122.5, CH ₂	5.28, m		8, 9	
11	23.5, CH ₃	1.90, m		8, 9, 10	
12 α	33.7, CH ₂	1.79, dd (13.9, 1.8)	13	2, 14	4
12 β	33.7, CH ₂	2.48, dd (13.9, 6.3)	13	2, 14	1, 16
13	78.8, CH	3.87, dd (6.3, 1.8)	12	2, 14	4
14	86.5, C				
15	28.3, CH ₃	1.26, s		13, 14, 16	4
16	23.8, CH ₃	1.27, s		14	12 β

Table S6. ^1H (400 MHz) and ^{13}C (100 MHz) NMR, and COSY, HMBC and NOESY assignment of **4** in CD₃OD.

No	4				
	δ_{C} , type	δ_{H} (<i>J</i> in Hz)	COSY	HMBC	NOESY
1	74.1, CH	3.73, s		2, 3, 6, 7	3, 13
2	30.6, C				
3	73.1, CH	4.01, d (5.2)	4	1, 12	1
4	72.4, CH	4.13, m	3, 5		
5	136.2, CH	6.02, d (3.2)	4	1, 6, 7	
6	127.5, C				
7	88.4, C				
8	92.0, C				
9	128.3, C				
10	122.3, CH ₂	5.26, m		8, 9, 11	
11	23.6, CH ₃	1.90, m		9, 10	
12	7.9, CH ₂	0.92, dd (9.7, 4.8) 0.81, dd (7.5, 4.8)	13	2, 3	
13	34.3, CH	1.21, m	12	1	1
14	70.1, C				
15	31.7, CH ₃	1.36, s		13, 14	
16	29.7, CH ₃	1.25, s		13, 14	

Table S7. ^1H (400 MHz) and ^{13}C (100 MHz) NMR, and COSY, HMBC and NOESY assignment of **5** in CD₃OD.

No	5				
	δ_{C} , type	δ_{H} (<i>J</i> in Hz)	COSY	HMBC	NOESY
1	74.7, CH	3.23, d (10.5)	6	8	3, 5 β , 12 β
2	78.4, C				
3	76.2, CH	3.06, d (9.3)	4	12	1, 5 β
4	70.9, CH	3.65, m	3, 5		6
5 α	36.9, CH ₂	2.04, dt (13.2, 4.6)	4, 6		
5 β	36.9, CH ₂	1.26, m	4, 6		1, 3
6	33.0, CH	2.83, m	5, 1	7, 8	4
7	83.5, C				
8	91.6, C				

9	128.8, C				
10	121.0, CH ₂	5.19, m 5.15, m		7, 11	
11	24.0, CH ₃	1.86 m		9, 10	
12 α	33.8, CH ₂	2.51, dd (13.4, 7.9)	13	1, 2, 3	
12 β	33.8, CH ₂	2.42, dd (13.4, 7.8)	13	1, 2, 3	1
13	120.1, CH	5.18, m	12	15, 16	
14	136.1, C				
15	26.3, CH ₃	1.74, s		13, 14, 16	
16	18.2, CH ₃	1.71, s		13, 14, 15	

Table S8. ¹H (400 MHz) and ¹³C (100 MHz) NMR of **6-8** in CD₃OD.

No	6		7		8	
	δ_{C}	δ_{H} (<i>J</i> in Hz)	δ_{C}	δ_{H} (<i>J</i> in Hz)	δ_{C}	δ_{H} (<i>J</i> in Hz)
1	68.3	4.60, t (1.9)	78.0	4.25, s	67.6	4.17, t (2.0)
2	58.1		32.2		63.3	
3	57.5	3.14, t (2.1)	75.3	3.44, m	60.4	3.34, overlapped
4	64.9	4.44, dt (5.5, 1.9)	69.2	4.14, dd (5.2, 1.1)	66.2	4.43, dd (5.0, 2.5)
5	133.0	5.93, dt (5.5, 2.2)	134.4	6.10, d (5.2)	134.3	5.76, dd (4.5, 2.5)
6	123.8		129.4		124.5	
7	86.9		88.2		87.6	
8	93.4		92.5		91.6	
9	128.4		128.4		128.4	
10	122.2	5.24, s	122.3	5.28, dd (3.8, 3.0) 5.26, m	122.2	5.28, m; 5.25, m
11	23.5	1.89, t (1.1)	23.6	1.90, d (1.1)	23.5	1.90, m
12	34.8	2.71, dd (14.8, 3.3) 1.55, dd (14.8, 2.9)	12.3	0.69, m	32.2	2.93, dd (14.7, 8.9) 2.16, dd (14.7, 6.2)
13	72.6	3.64, t (2.9)	33.5	1.69, dd (7.8, 4.6)	118.9	5.13, m
14	78.1		83.9		136.7	
15	22.3	1.39, s	28.5	1.37, s	18.1	1.69, s
16	26.4	1.32, s	26.0	1.20, s	26.0	1.74, s