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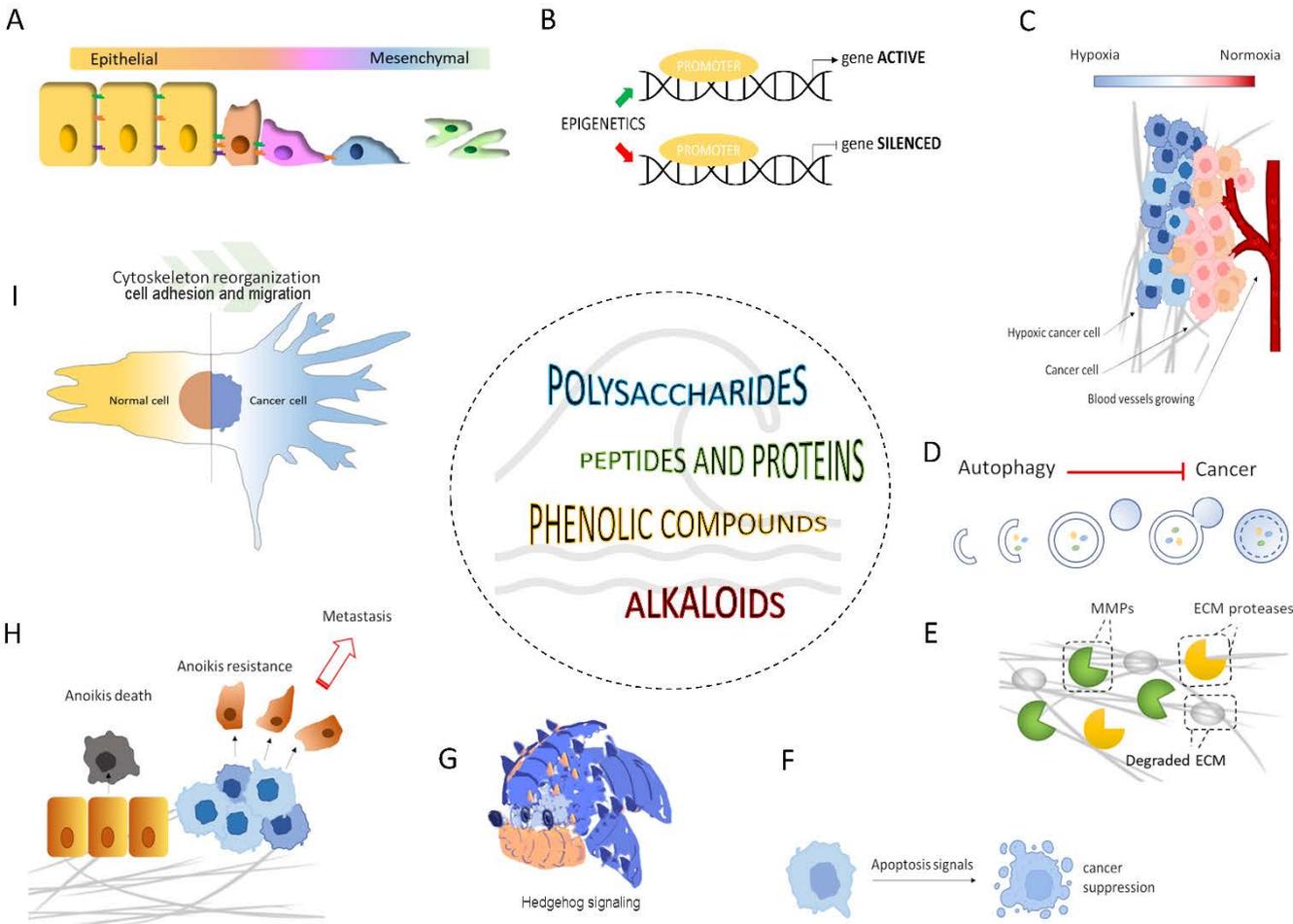


Figure S1. Schematic representation of potential molecular targets of marine migrastatics (2017-2022): (A) epithelial-mesenchymal transition; (B) epigenetic control; (C) neo-angiogenesis and hypoxia; (D) autophagy; (E) ECM degradation by MMPs and other proteases; (F) apoptosis; (G) Hedgehog signalling pathway; (H) resistance to anoikis; (I) cytoskeletal reorganization, cell adhesion and migration. Click on each picture to view the involved migrastatic marine compound.

To go back to the pictures click on the table

Epithelial-to-Mesenchymal Transition

| Compound | Marine source |
|-----------------------------|--|
| Actinomycin V | <i>Streptomyces sp.</i> |
| Jorunnamycin A | <i>Xestospongia sp.</i> |
| Manzamine A | <i>Haliclona sp., Xestospongia sp. Pellina sp.</i> |
| α O-conotoxin GeXIVA | <i>C. generalis</i> |

To go back to the pictures click on the table

Epigenetic control

| Compound | Marine source |
|-----------------|--------------------------------------|
| Chromopeptide A | <i>Chromobacterium sp., HS-13-94</i> |

To go back to the pictures click on the table

Neo-angiogenesis and hypoxia

| Compound | Marine source |
|---------------------------------------|---------------------------|
| AATP | <i>H. discus hannai</i> |
| BABP | <i>H. discus hannai</i> |
| Crude sulfate polysaccharides extract | <i>P. tetrastromatica</i> |
| 7-Phloroeckol | <i>E. Cava</i> |

To go back to the pictures click on the table

Autophagy

| Compound | Marine source |
|--------------------|-----------------------------------|
| BFP-3 | <i>B. fuscopurpurea</i> |
| Catechin | <i>P. Oceanica</i> |
| Chlorogenic acid | <i>P. Oceanica</i> |
| Dihydroauroglaucin | <i>E. chevalieri</i> |
| Epicatechin | <i>P. Oceanica</i> |
| Ferulic acid | <i>P. boergeseni, P. oceanica</i> |
| Gallic acid | <i>P. boergeseni, P. oceanica</i> |

To go back to the pictures click on the table

ECM degradation by MMPs and other proteases

| Compound | Marine source |
|---------------------------------------|--|
| AATP | <i>H. discus hannai</i> |
| BABP | <i>H. discus hannai</i> |
| Catechin | <i>P. oceanica</i> |
| Chlorogenic acid | <i>P. Oceanica</i> |
| Crude sulfate polysaccharides extract | <i>P. tetrastromatica</i> |
| Dieckol | <i>E. Cava</i> |
| Epicatechin | <i>P. Oceanica</i> |
| Ferulic acid | <i>P. boergeseni, P. oceanica,</i> |
| Fucoidan-like STPC2 | <i>S. thunbergii</i> |
| Gallic acid | <i>P. boergeseni, P. oceanica</i> |
| Grassystatin F | <i>Cyanobacterium VPG 14-61</i> |
| Laminaran sulfate | <i>F. evanescens</i> |
| Nobilamide I | <i>Saccharomonospora sp., strain CNQ-490</i> |
| PBN11-8 | <i>B. pumilus</i> |
| Phlorofucofuroeckol | <i>E. cava</i> |
| SIP-SII | <i>S. maindroni</i> |

To go back to the pictures click on the table

Apoptosis

| Compound | Marine source |
|-----------------------|-----------------------------------|
| BFP-3 | <i>B. fuscopurpurea</i> |
| Caffeic acid | <i>P. boergeseni</i> |
| Ferulic acid | <i>P. boergeseni, P. oceanica</i> |
| Gallic acid | <i>P. boergeseni, P. oceanica</i> |
| N-V protease (or NAP) | <i>N. virens</i> |
| Quercetin | <i>P. boergeseni</i> |
| Rutin | <i>P. boergeseni</i> |

To go back to the pictures click on the table

Hedgehog signaling

| Compound | Marine source |
|-------------|-----------------------|
| SPUP | <i>U. Pinnatifida</i> |

To go back to the pictures click on the table

Resistance to anoikis

| Compound | Marine source |
|-----------------------|-------------------------|
| Jorunnamycin A | <i>Xestospongia sp.</i> |

To go back to the pictures click on the table

Cytoskeletal reorganization, cell adhesion and migration

| Compound | Marine source |
|-----------------------------|--------------------------------------|
| Aeroplysinin-1 | <i>A. aerophoba</i> |
| Chromopeptide A | <i>Chromobacterium sp., HS-13-94</i> |
| CLP | <i>Marine microorganisms</i> |
| Dieckol | <i>E. Cava</i> |
| Exopolysaccharide 11 | <i>Bacillus sp. 11</i> |
| Fucoidan | <i>C. frondosa</i> |
| Molassamide | <i>Cyanobacterium DRTO-73</i> |
| PBN11-8 | <i>B. pumilus</i> |
| Phlorofucofuroeckol | <i>E. Cava</i> |
| SIP-SII | <i>S. maindroni</i> |
| Sulfated galactans | <i>G. fisheri</i> |
| Viriditoxin | <i>P. variotii</i> |