

Supplementary material for First Total Synthesis of 5'-O- α -D-Glucopyranosyl Tubercidin

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1. Chemical shifts of naturally occurring nucleoside **2**, Synthetic nucleoside **2** and **17**

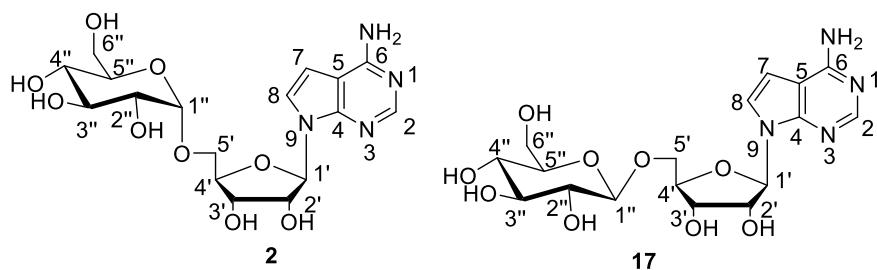


Table S1. ^{13}C NMR chemical shifts of naturally occurring nucleoside **2**, synthetic nucleoside **2** and **17**

Chemical shifts (δ)	Naturally occurring nucleoside 2 / ppm	Synthetic nucleoside 2 / ppm	Synthetic nucleoside 17 / ppm
Position of carbon			
C-6	157.3	157.3	157.5
CH-2	151.5	151.5	151.7
C-4	150.6	150.6	150.7
CH-8	122.2	122.4	122.2
C-5	103.0	102.8	103.1
CH-7	99.8	100.0	100.5
CH-1''	98.7	98.5	103.5
CH-1'	85.8	85.9	87.1
CH-4'	83.0	83.2	83.1
CH-2'	74.0	74.2	77.2
CH-4''	73.4	73.3	74.2
CH-5''	72.9	72.7	74.1
CH-2''	71.8	71.5	77.4
CH-3'	71.1	70.9	71.1
CH-3''	70.2	70.0	70.5
CH ₂ -5'	67.2	67.0	69.5
CH ₂ -6''	60.9	60.9	61.5

Table S2. ^1H NMR chemical shifts and coupling constants of naturally occurring nucleoside 2, synthetic 2 and 17

Chemical shifts (δ) Hydrogen	Naturally occurring nucleoside 2 (coupling constants) / ppm	Synthetic nucleoside 2 (coupling constants) / ppm	Synthetic nucleoside 17 (coupling constants) / ppm
H-2	8.03 (s)	8.09 (s)	8.07 (s)
H-8	7.69(d, $J = 3.7$ Hz)	7.76 (d, $J = 3.2$ Hz)	7.38 (d, $J = 3.7$ Hz)
NH ₂	6.95 (s)	7.20 (s)	7.09 (s)
H-7	6.53 (d, $J = 3.7$ Hz)	6.61 (d, $J = 3.4$ Hz)	6.61 (d, $J = 3.6$ Hz)
H-1'	6.12(d, $J = 7.0$ Hz)	6.14 (d, $J = 7.0$ Hz)	6.08 (d, $J = 5.6$ Hz)
3'-OH	5.19 (d, $J = 4.8$ Hz)	5.21 (d, $J = 4.6$ Hz)	5.17 (d, $J = 4.6$ Hz)
2''-OH	5.19 (d, $J = 4.8$ Hz)	5.21 (d, $J = 4.6$ Hz)	5.03 (d, $J = 3.7$ Hz)
2'-OH	5.10 (d, $J = 4.8$ Hz)	5.10 (d, $J = 4.3$ Hz)	5.26 (d, $J = 5.5$ Hz)
3''-OH	4.93 (d, $J = 3.8$ Hz)	4.92 (d, $J = 4.9$ Hz)	4.95 (s)
4''-OH	4.85 (d, $J = 4.6$ Hz)	4.84 (d, $J = 3.6$ Hz)	4.91 (s)
H-1''	4.70 (d, $J = 3.5$ Hz)	4.72 (d, $J = 3.1$ Hz)	4.21 (d, $J = 7.8$ Hz)
6''-OH	4.50 (d, $J = 4.7$ Hz)	4.49 (m)	4.51 (s)
H-2'	4.41 (td, $J = 6.3$ Hz, 4.8,4.8 Hz)	4.43 (m)	4.36 (d, $J = 5.0$ Hz)
H-4'	4.09 (td, $J = 4.9$ Hz, 4.9,2.1 Hz)	4.11 (m)	4.02–3.95 (m)
H-3'	4.05 (m)	4.08 (d, $J = 1.7$ Hz)	4.14 (d, $J = 4.4$ Hz)
H-5'	3.76 (dd, $J = 11.1,3.1$ Hz)	3.78 (dd, $J = 10.9, 2.8$ Hz)	4.02–3.95 (m)
H-6''	3.64 (ddd, $J = 10.9,4.8,1.2$ Hz)	3.66–3.63 (m)	3.68 (d, $J = 9.2$ Hz)
H-4''	3.45 (m)	3.47–3.42 (m)	3.12–3.04 (m)
H-6' ''	3.44 (m)	3.47–3.42 (m)	3.44 (d, $J = 6.7$ Hz)
H-5'	3.41 (dd, $J = 11.1, 4.9$ Hz)	3.47–3.42 (m)	3.59 (dd, $J = 11.9,5.7$ Hz)
H-5''	3.34 (m)	3.40–3.34 (m)	3.12–3.04 (m)
H-2''	3.26 (m)	3.40–3.34 (m)	3.00 (d, $J = 7.8$ Hz)
H-3''	3.10 (td, $J = 9.3, 3.8$ Hz)	3.13–3.07 (m)	3.15 (d, $J = 9.9$ Hz)

2. Optimization of glycosylation of **11** and **6**

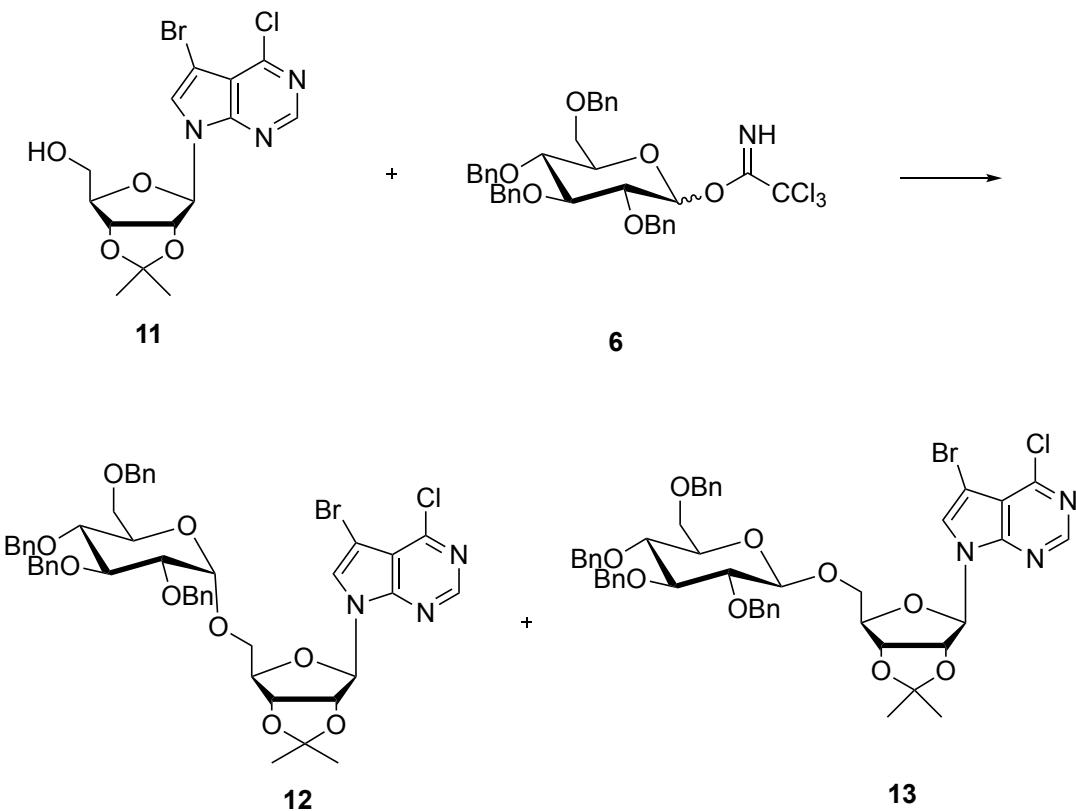
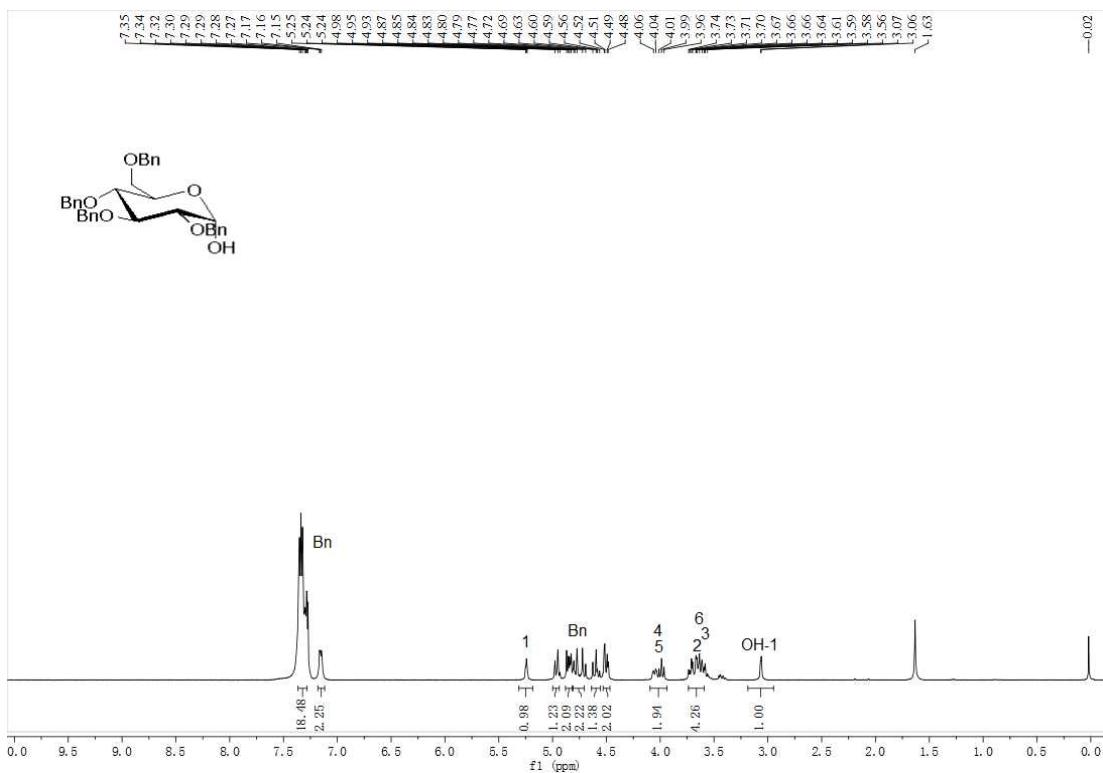


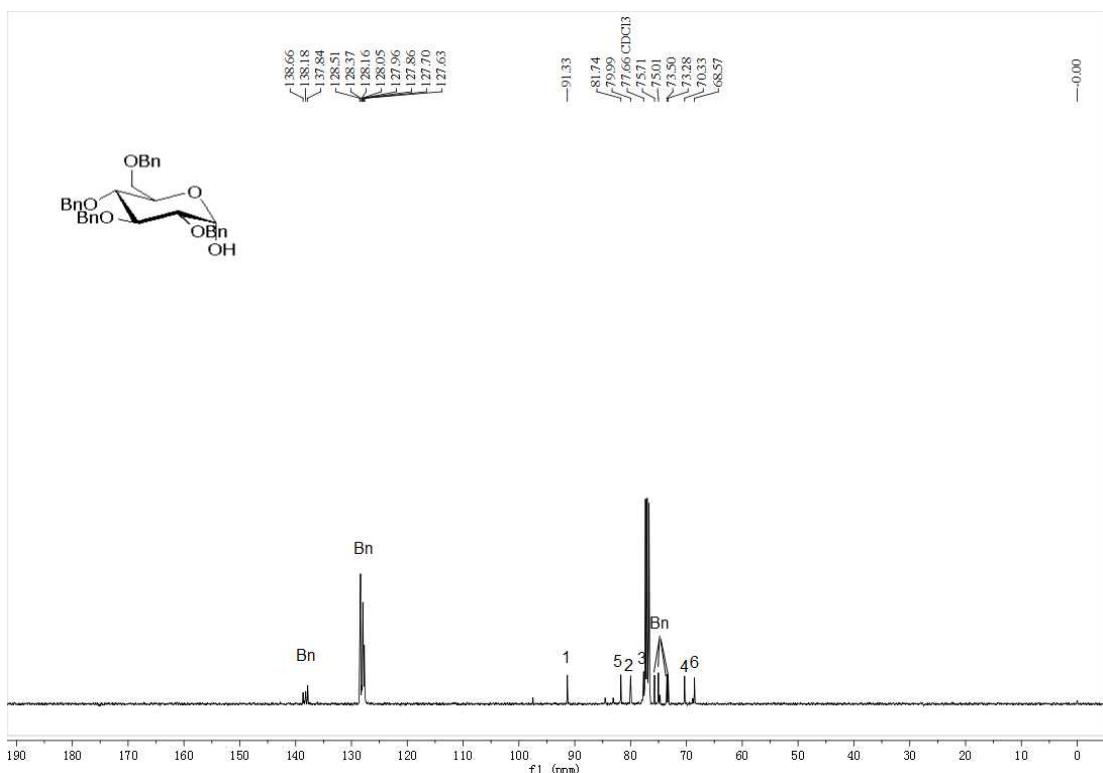
Table S3 Optimization of glycosylation of **11** and **6**

Entry	Solvent	Lewis Acid	Temperature	Ratio of 12:13	Yield
1	DCM	TMSOTf	-78 °C	2.6:1	71%
2	DCM	TMSOTf	-30 °C	4:1	79%
3	DCM	TMSOTf	0 °C	1.6:1	67%
4	DCM	BF ₃ .Et ₂ O	0 °C	1.2:1	43%
5	DCM	BF ₃ .Et ₂ O	-30 °C	3:1	59%
6	DCM	BF ₃ .Et ₂ O	-78 °C	1.5:1	57%

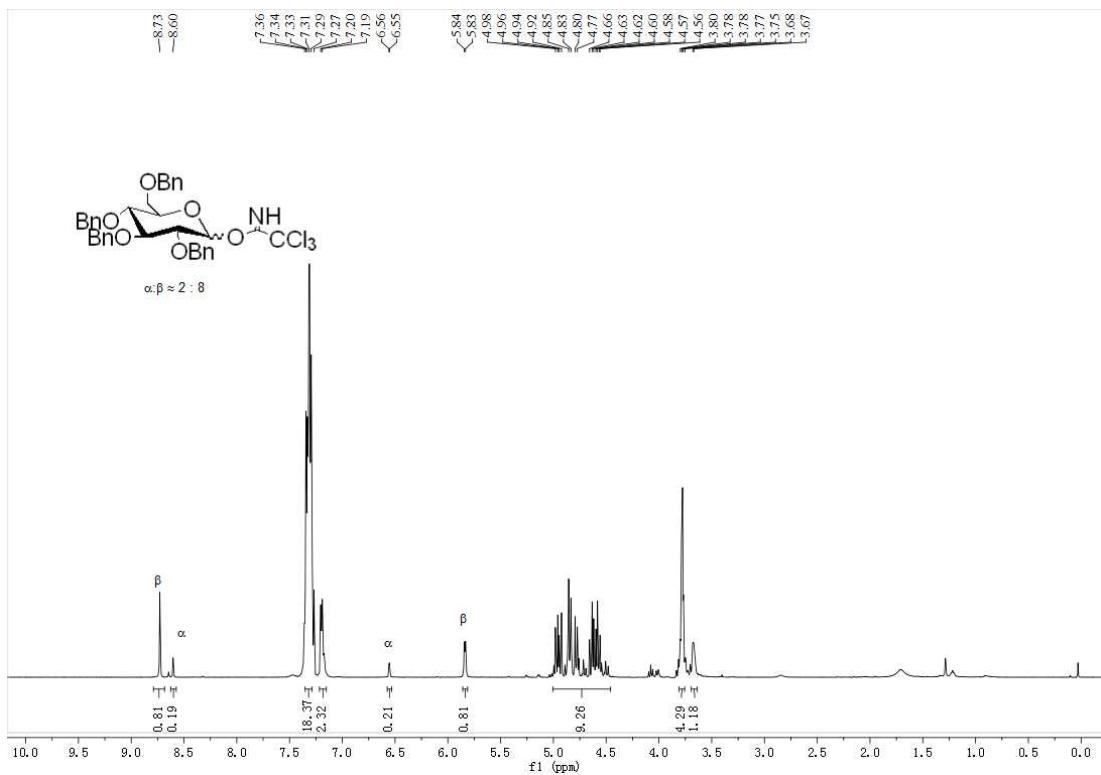
3. ^1H and ^{13}C NMR spectra of compounds 2 – 17.



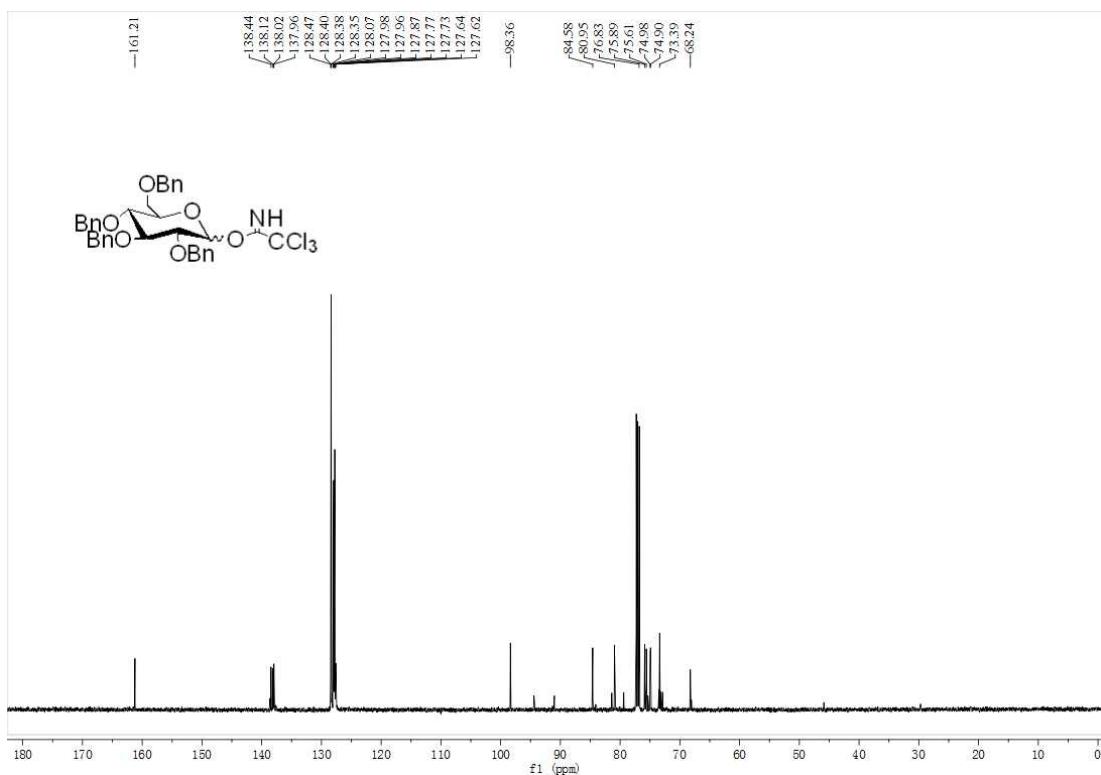
¹H NMR (CDCl_3) of 2,3,4,6-O-tetrabenzyl-D-glucopyranose.



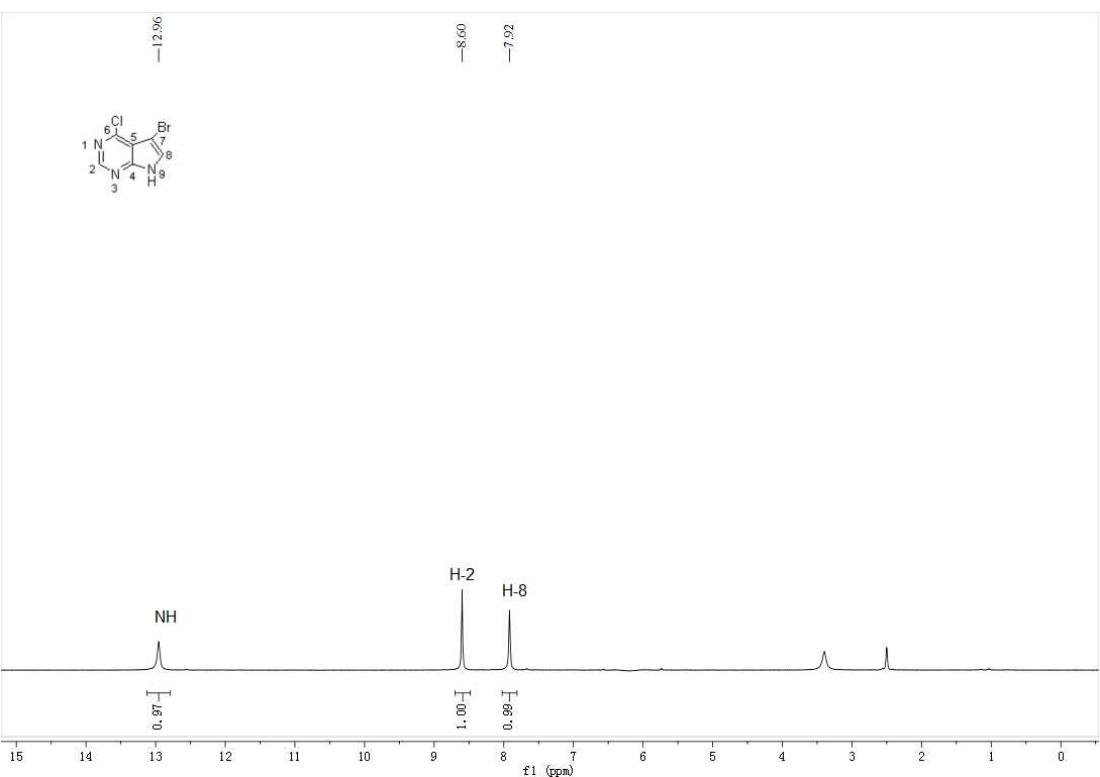
¹³C NMR (CDCl_3) of 2,3,4,6-O-tetrabenzyl-d-glucopyranose.



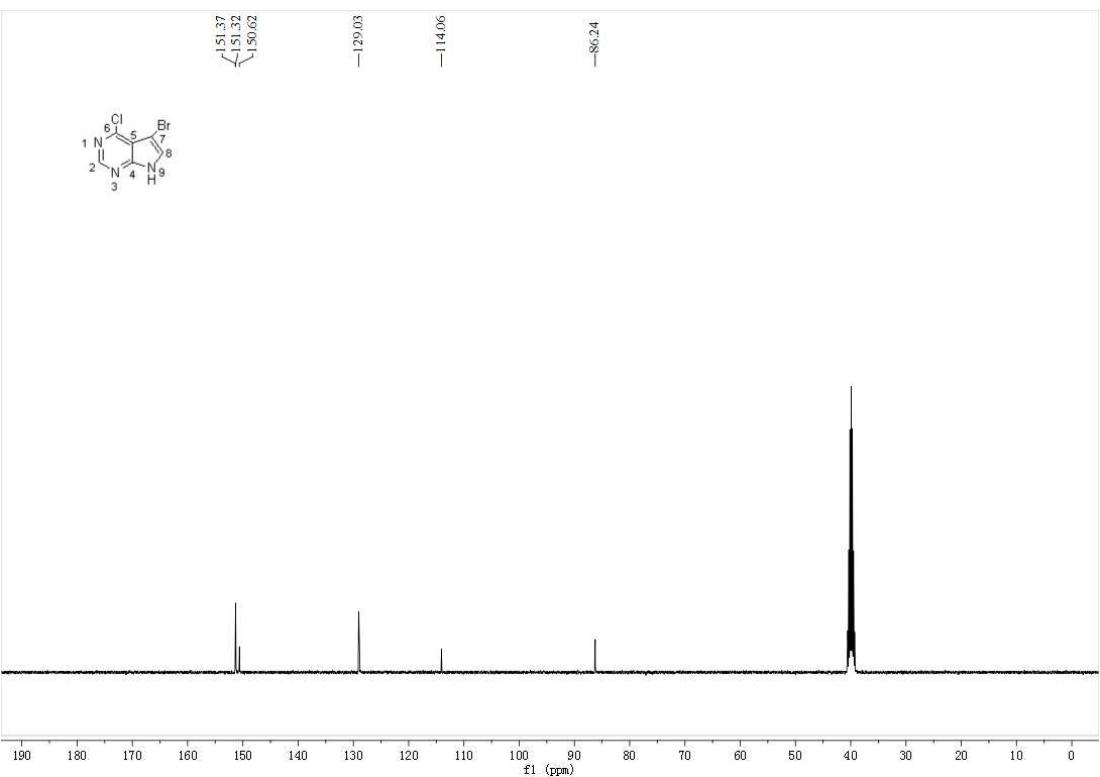
¹H NMR (CDCl₃) of **6**.



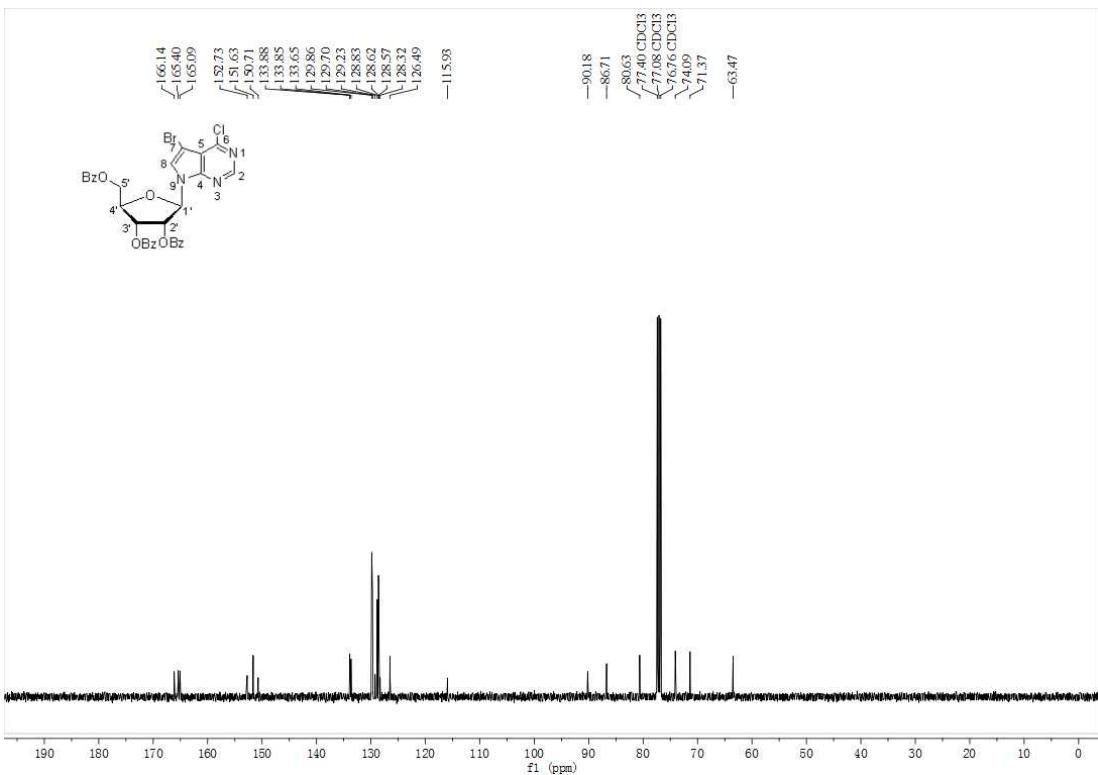
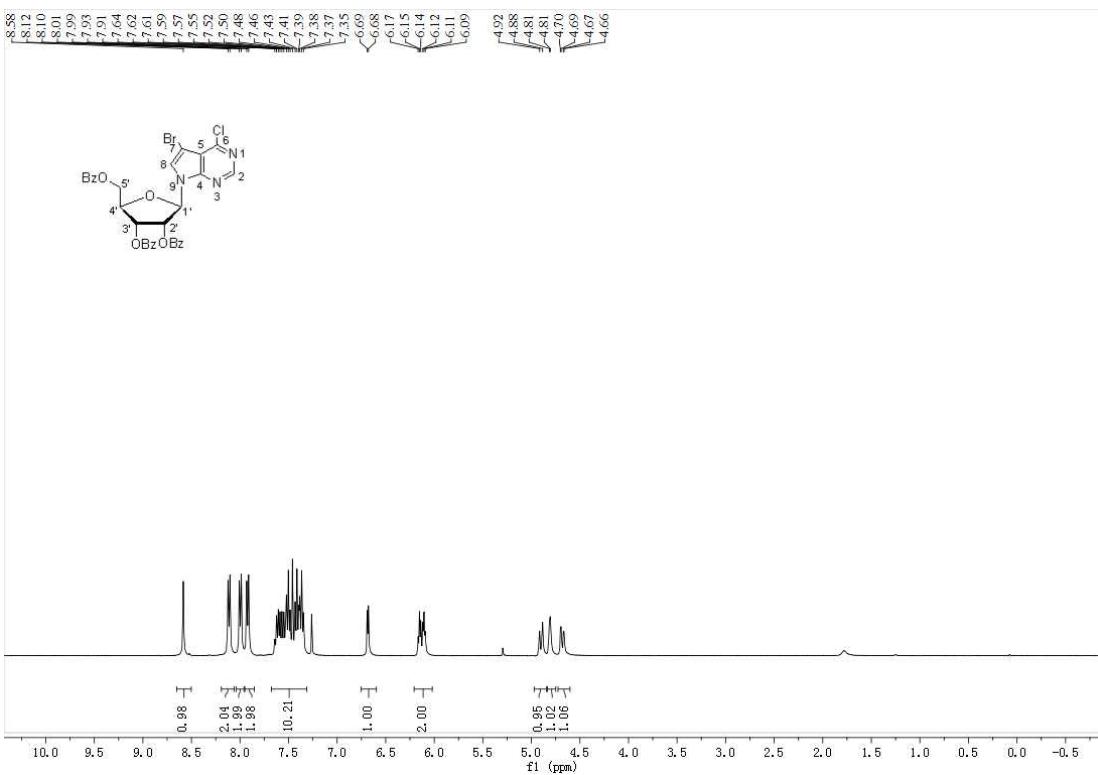
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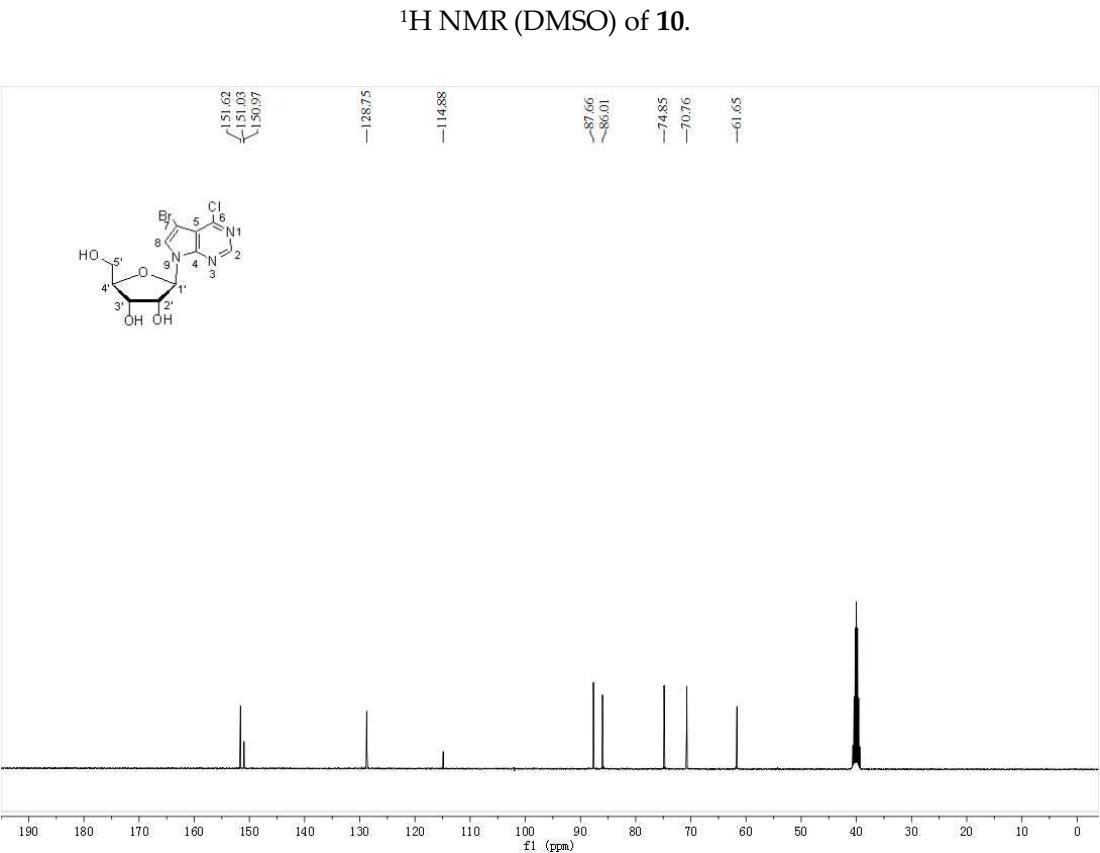
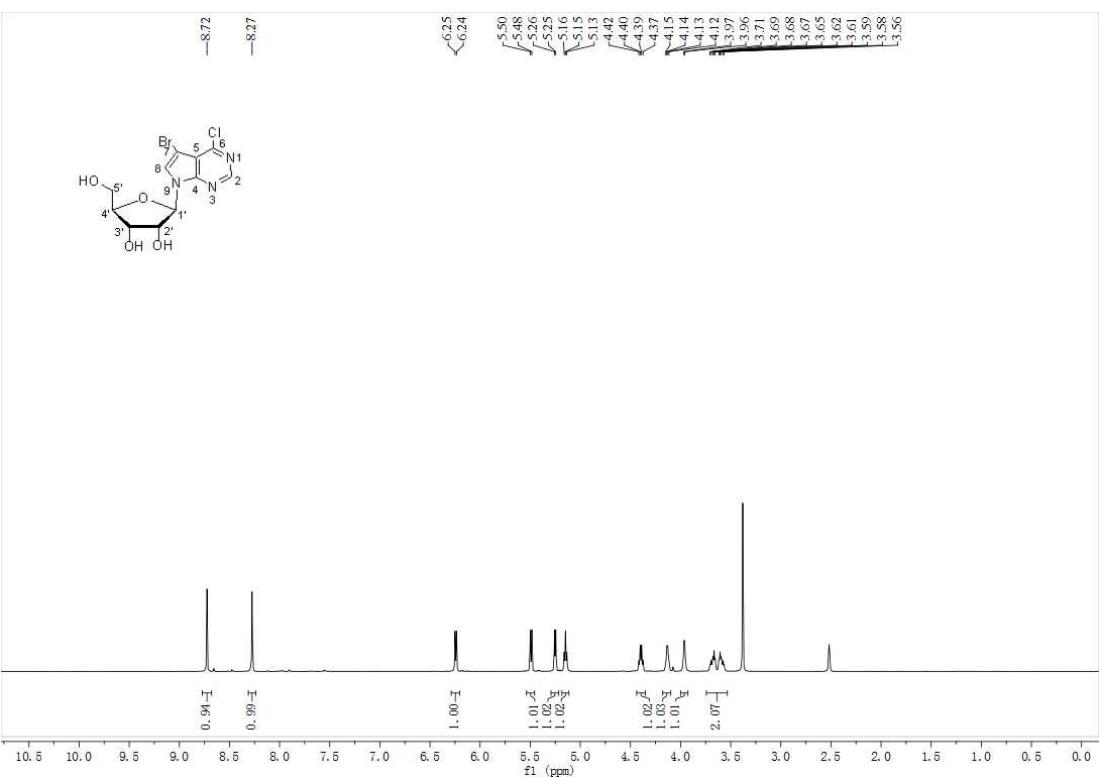
¹H NMR (DMSO) of 8.

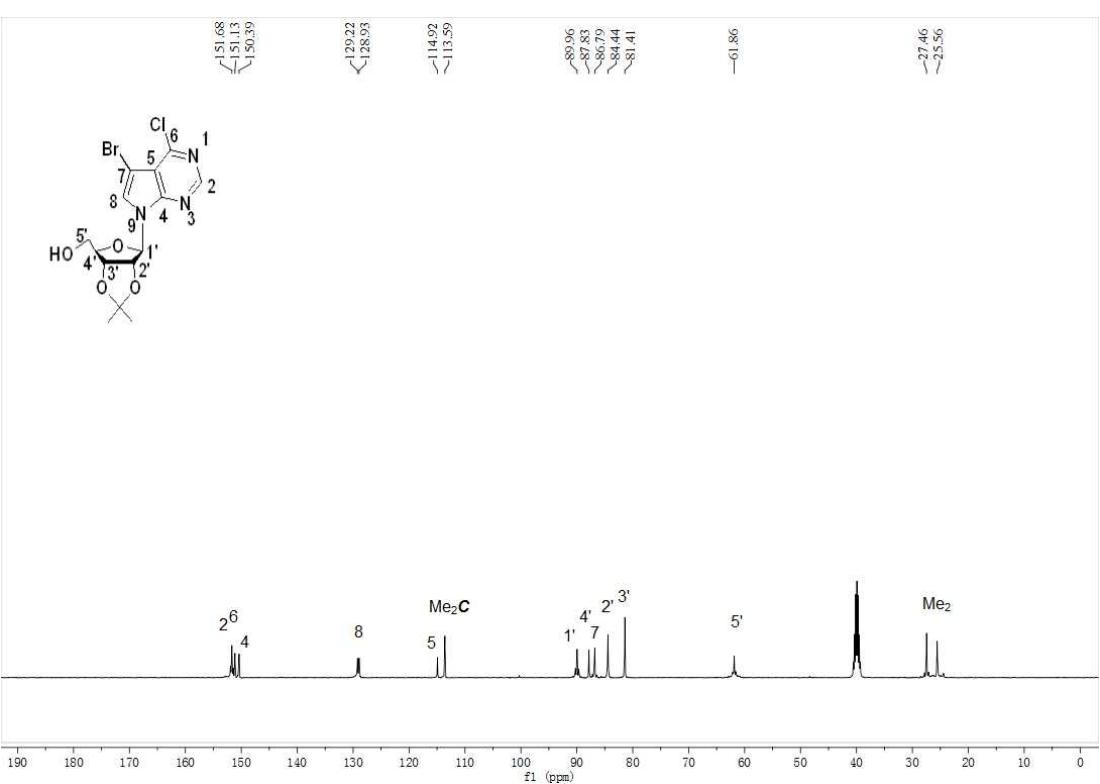
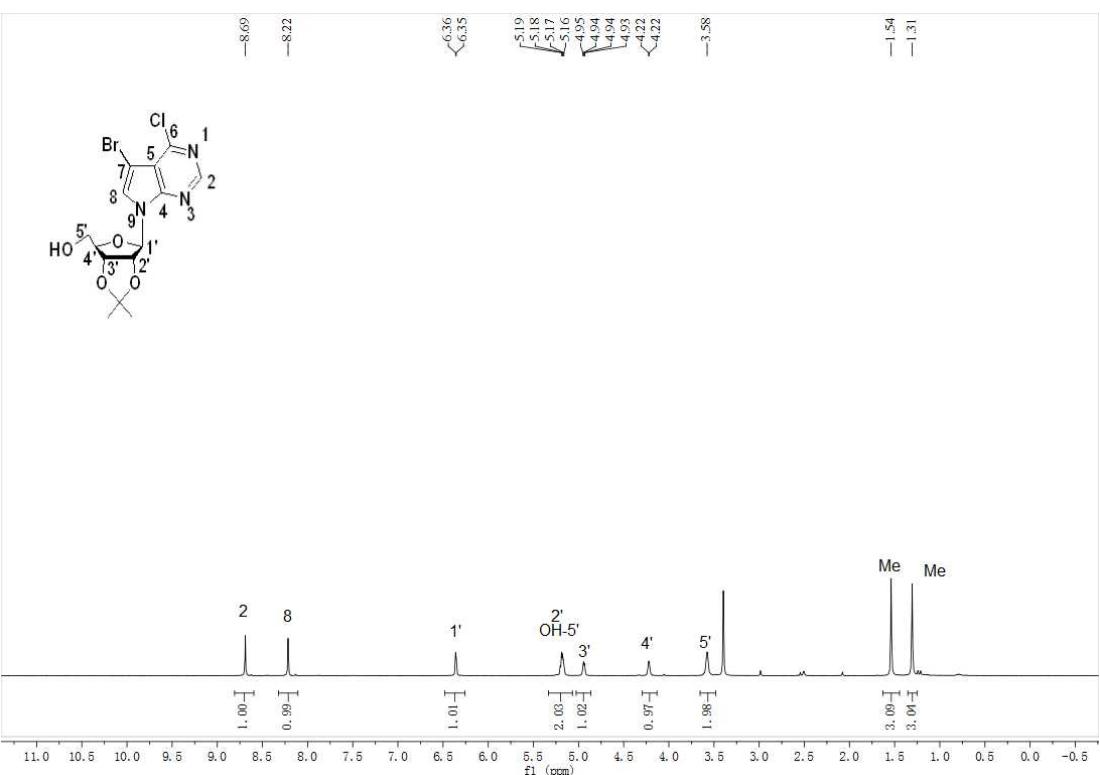


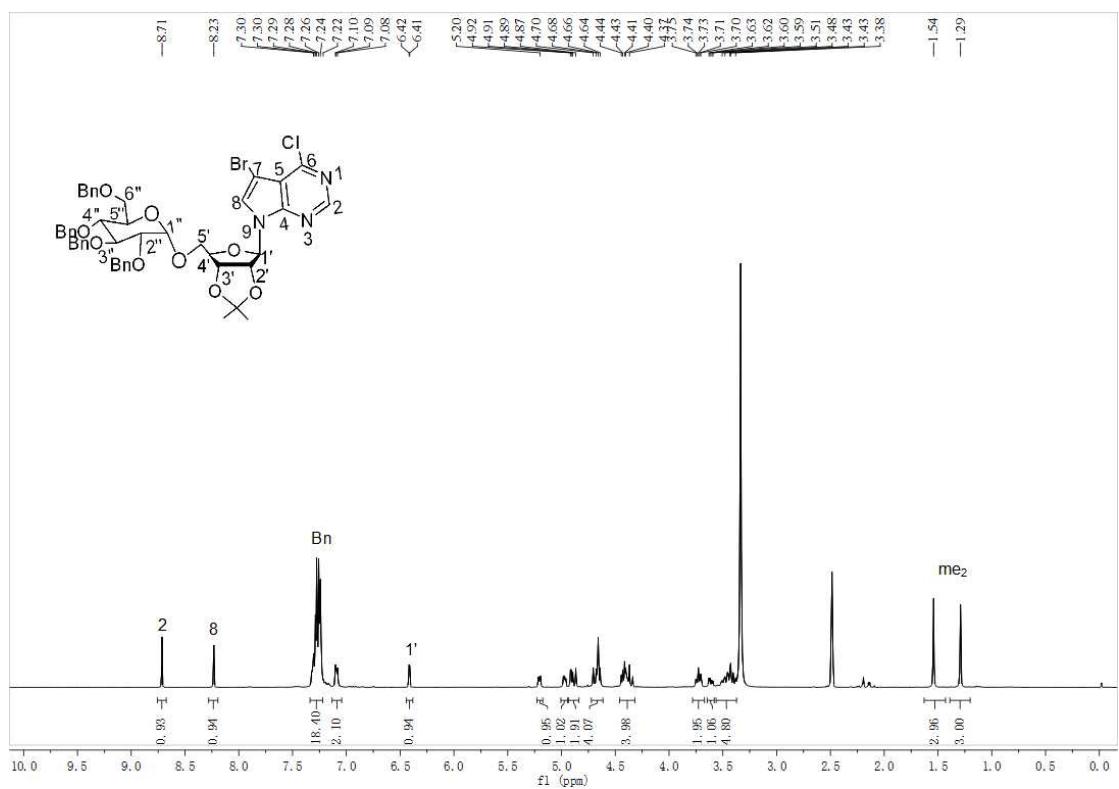
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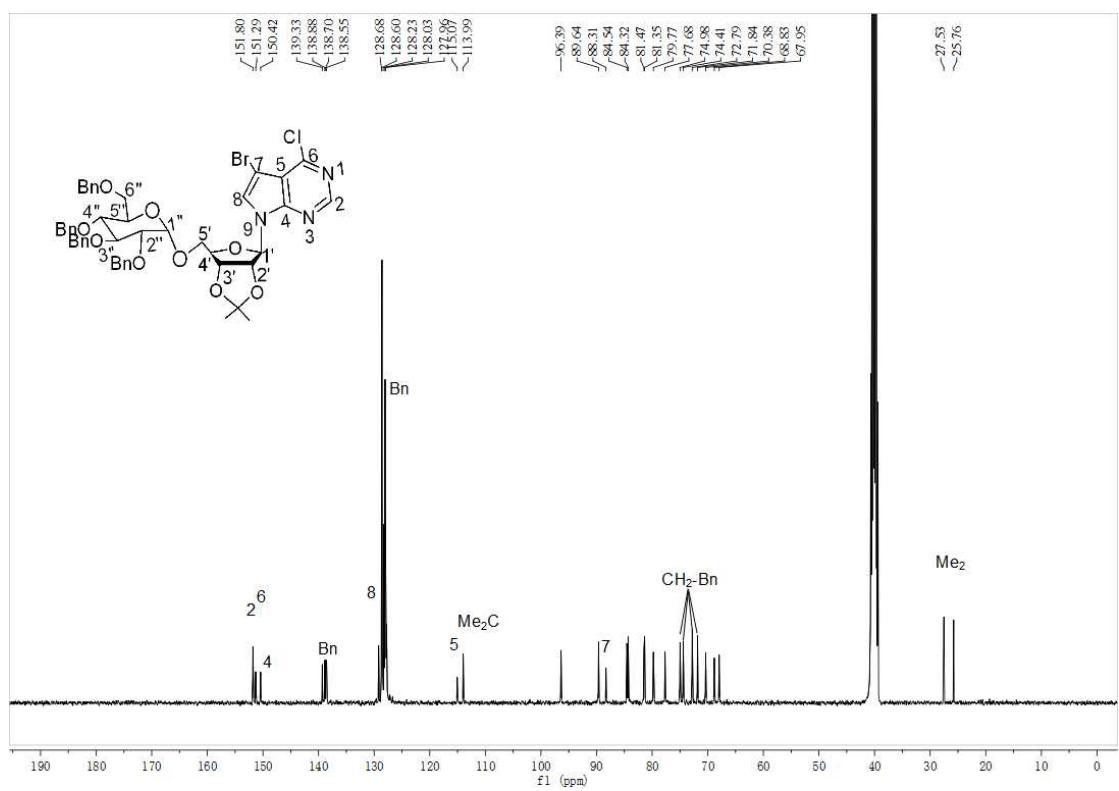
¹³C NMR (CDCl₃) of **9**.



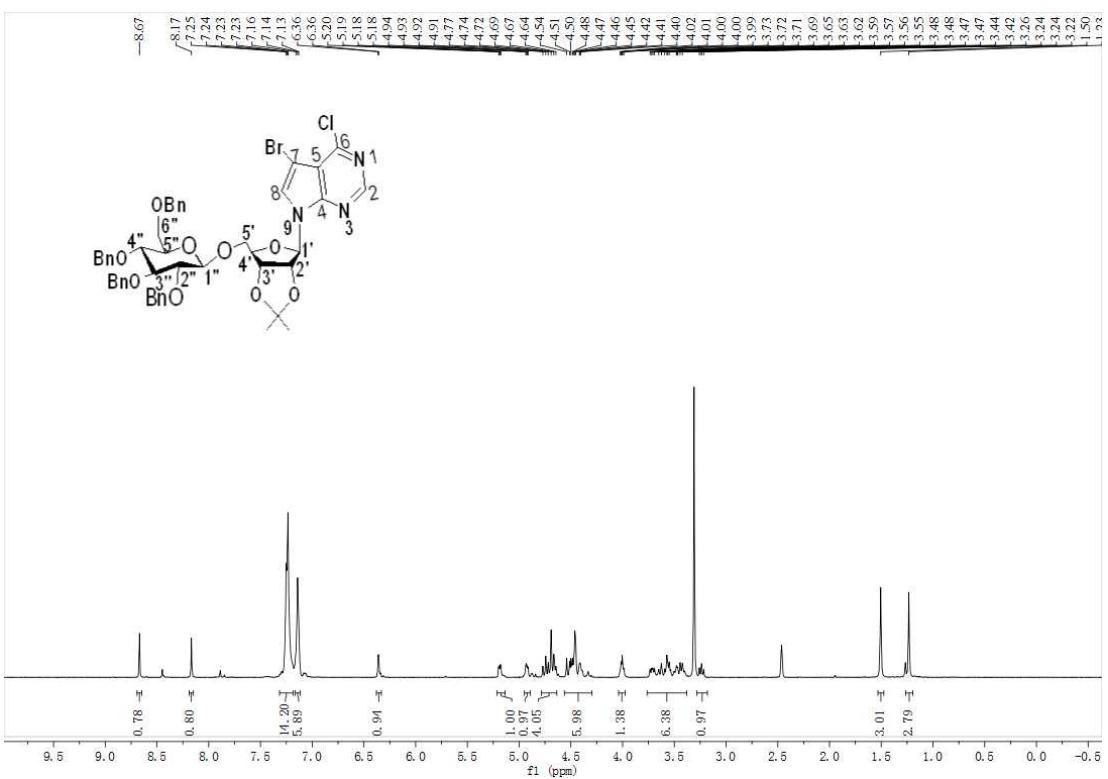




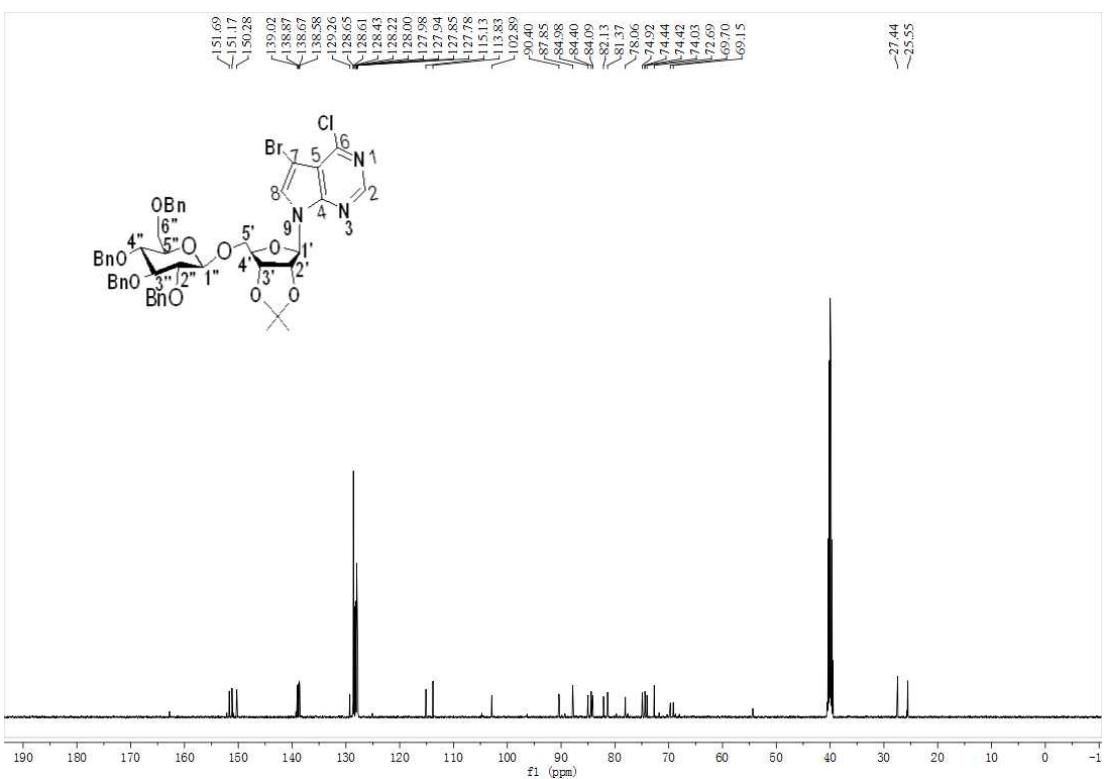
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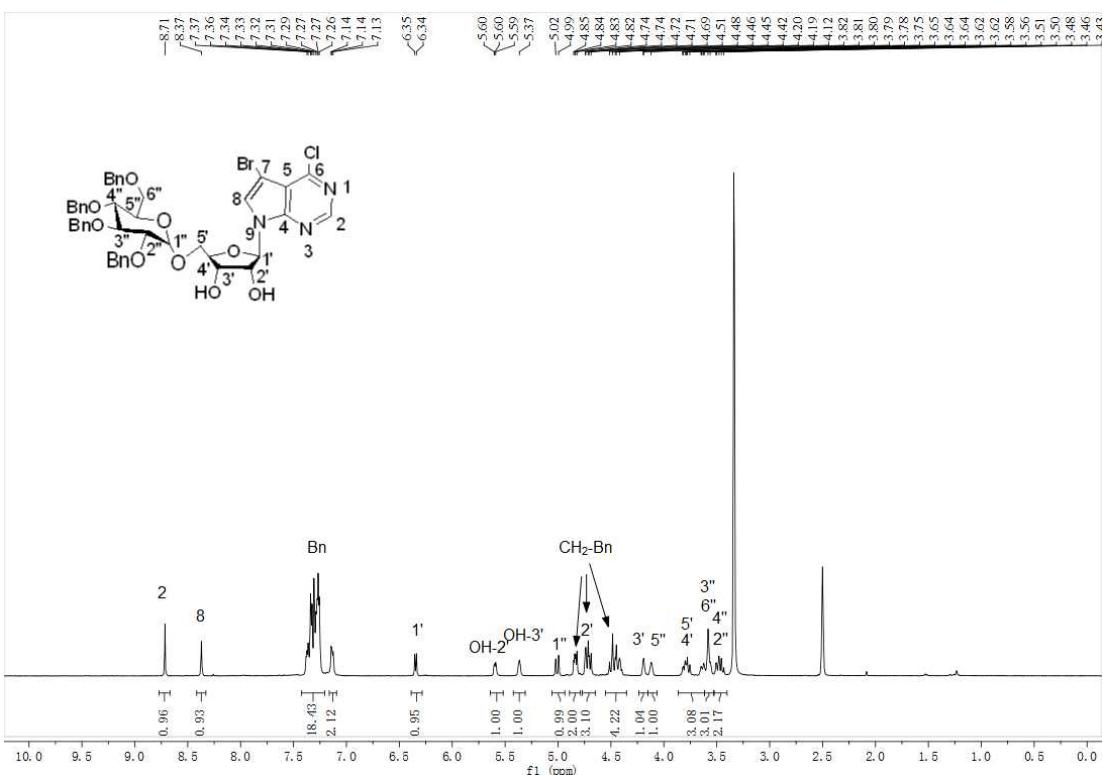
¹³C NMR (DMSO) of **12**.



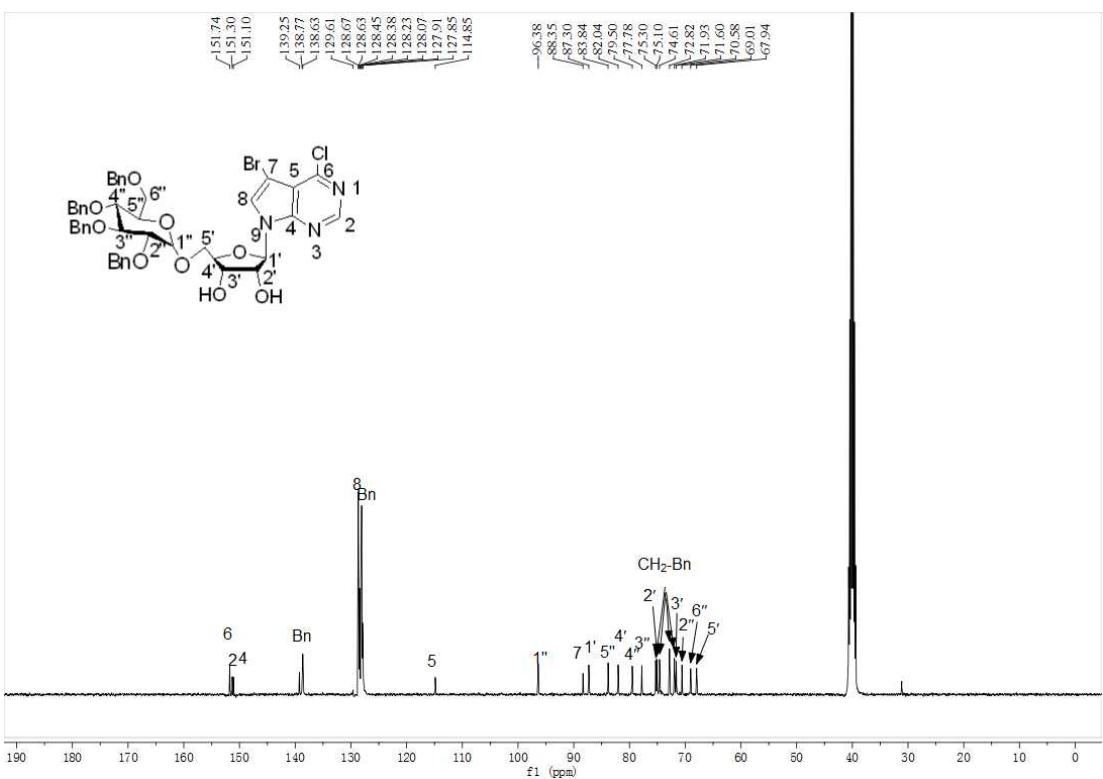
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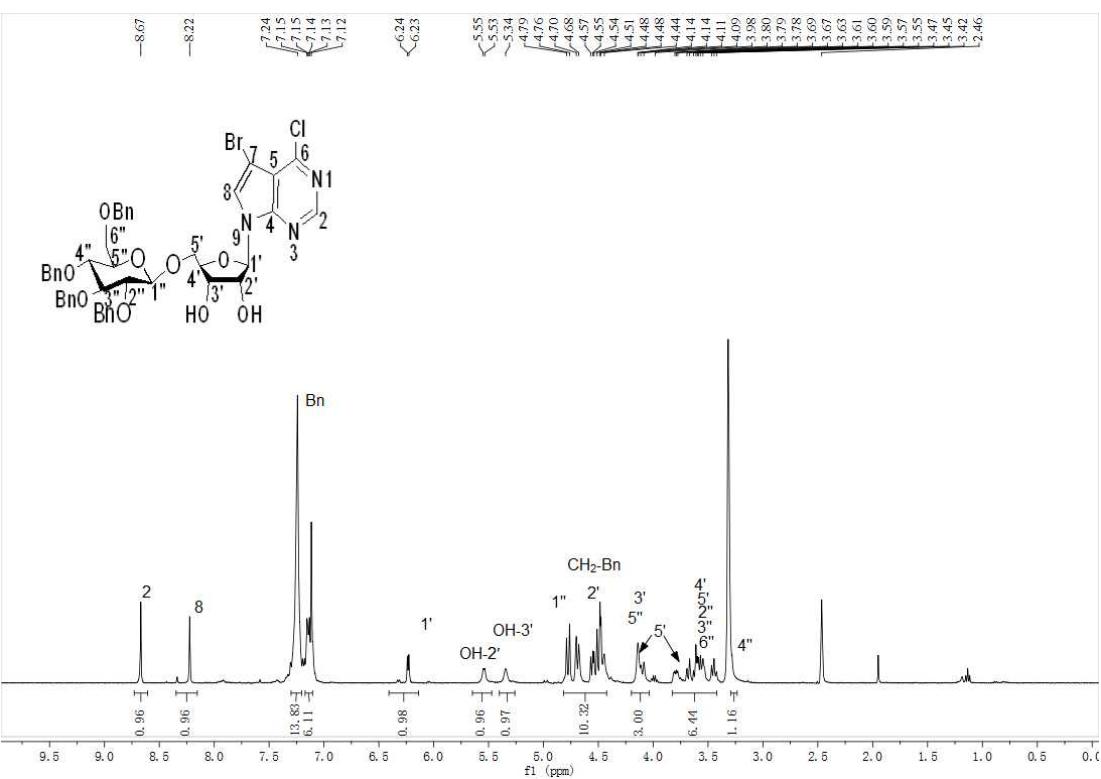
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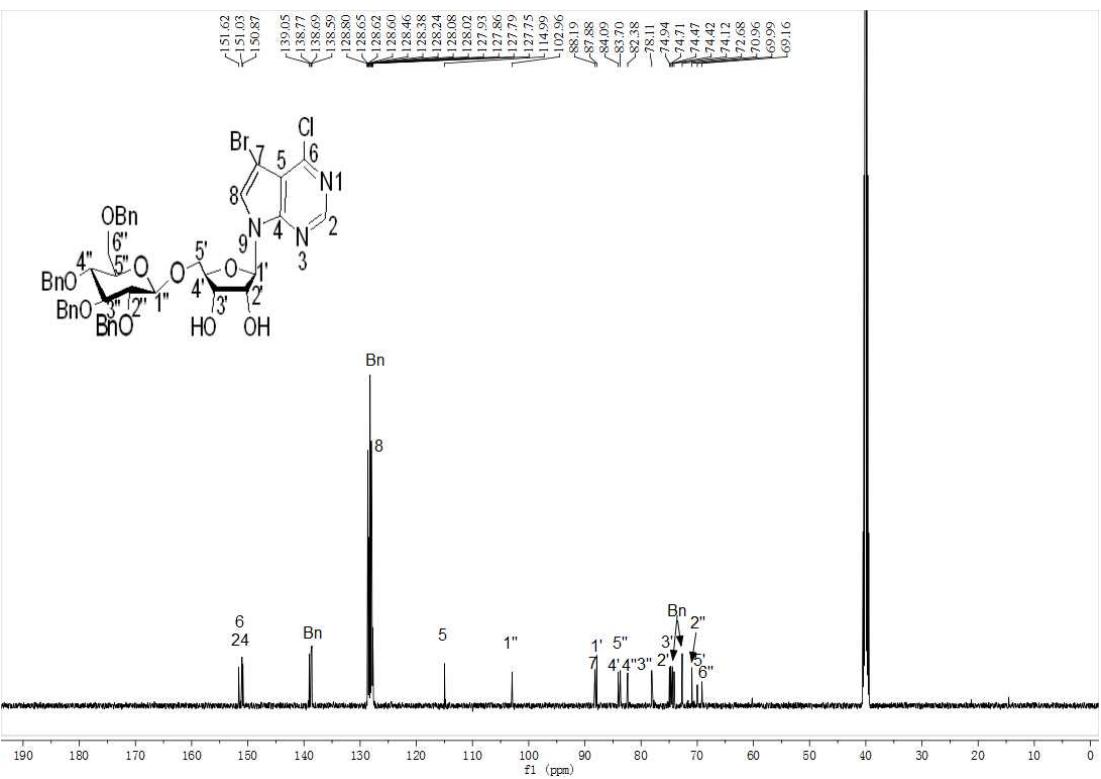
¹H NMR (DMSO) of **14**.



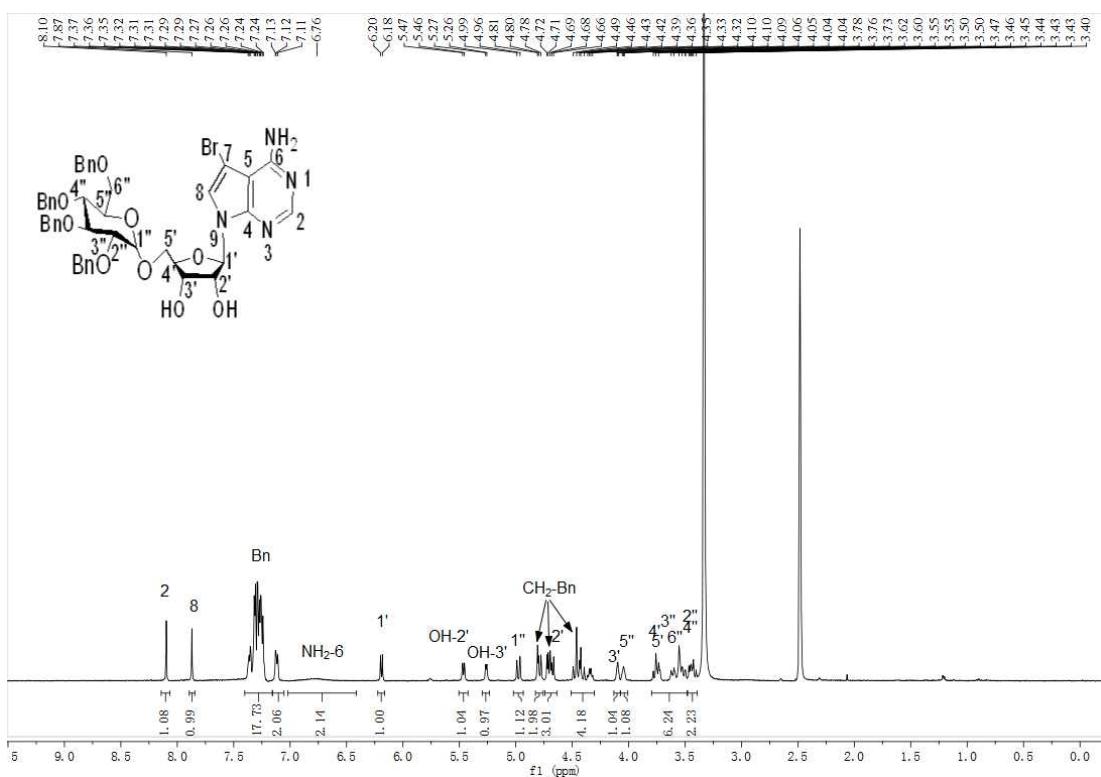
¹³C NMR (DMSO) of **14**.



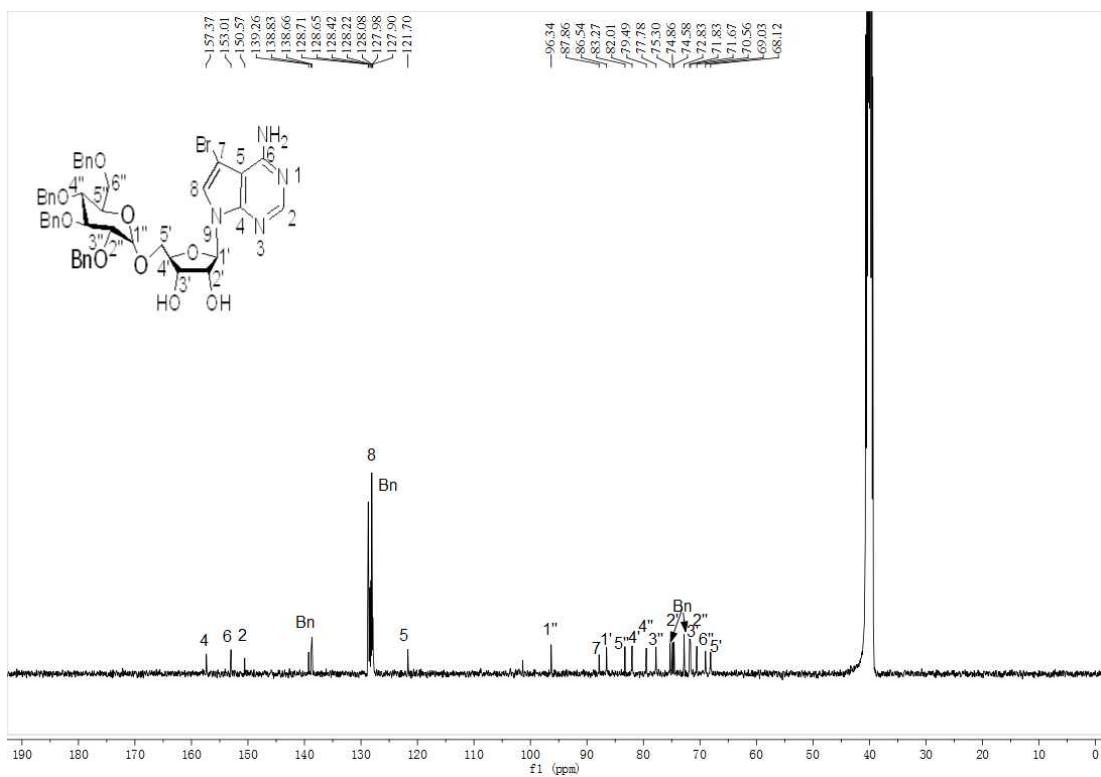
¹H NMR (DMSO) of **14'**.



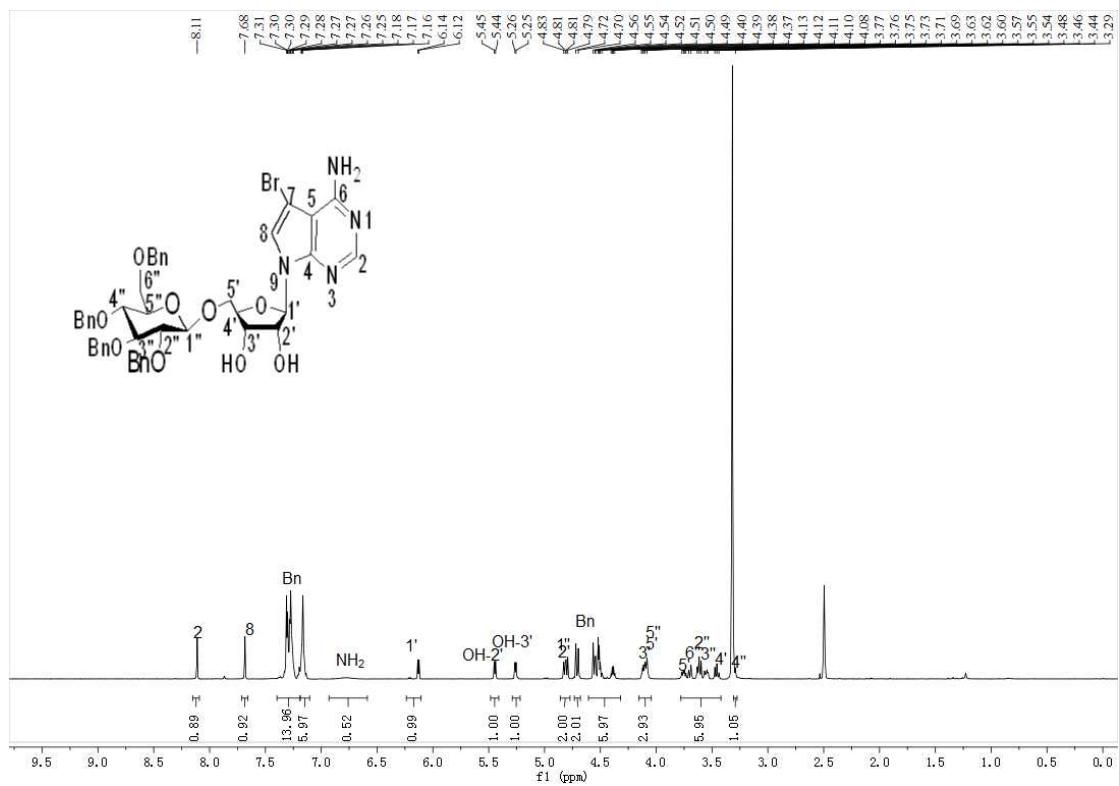
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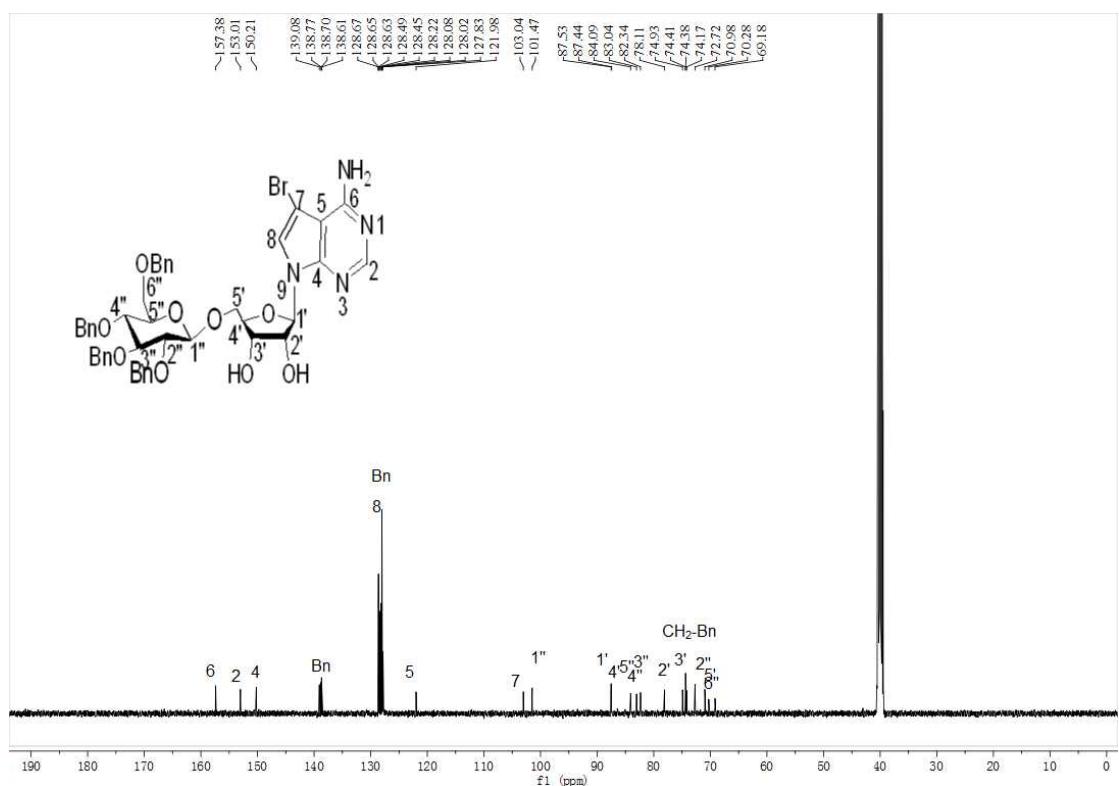
¹H NMR (DMSO) of 15.



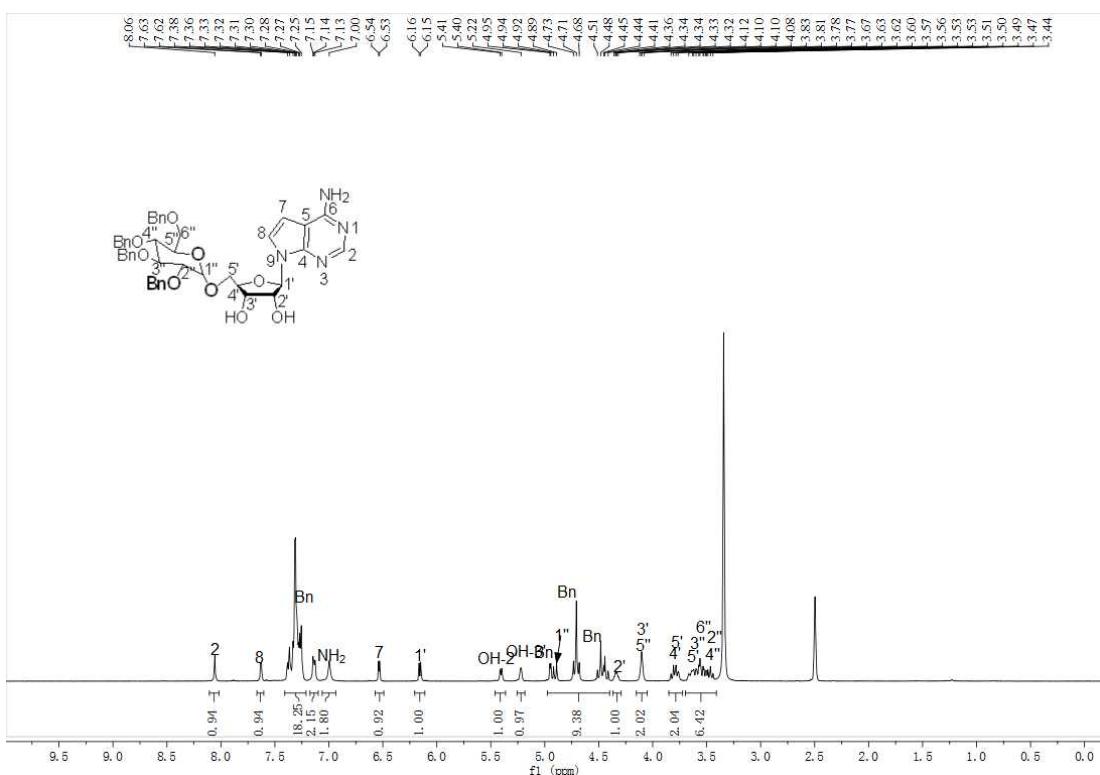
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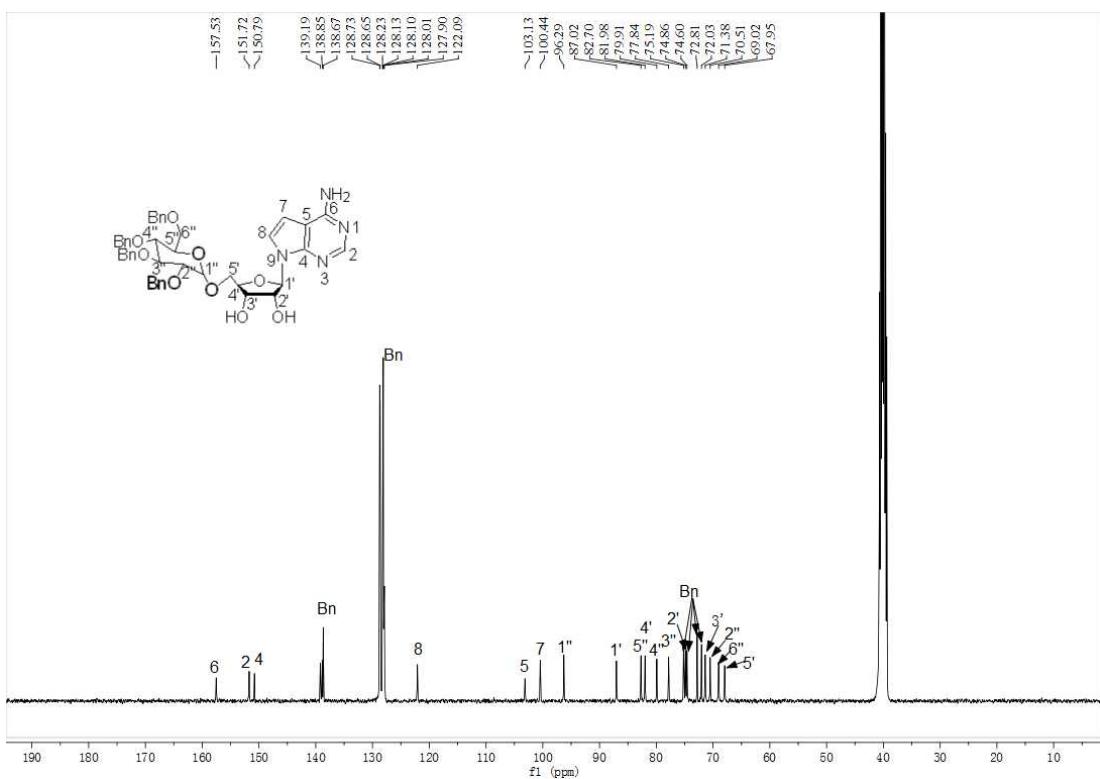
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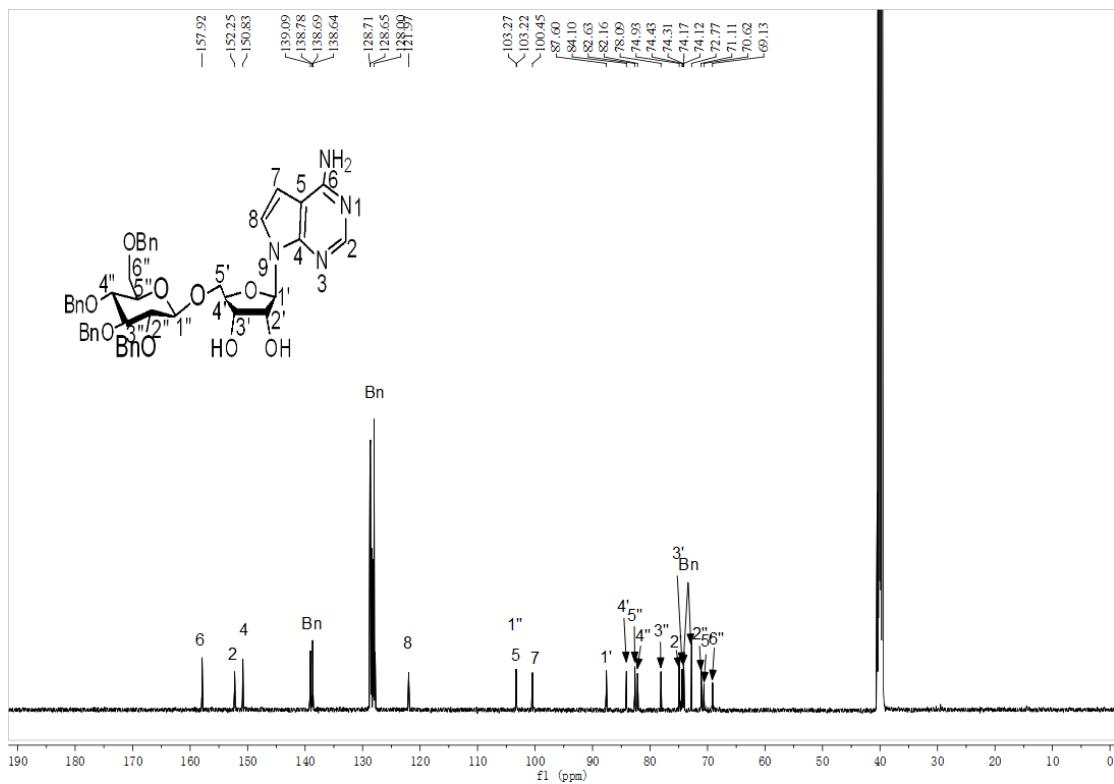
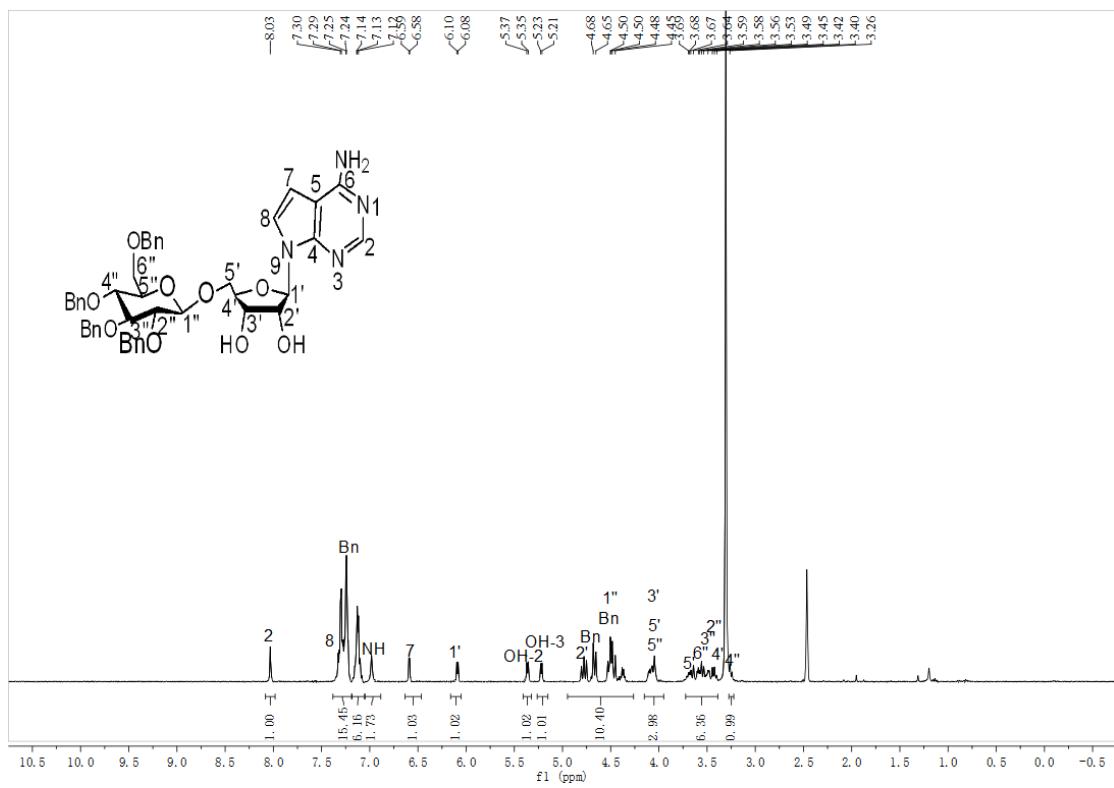
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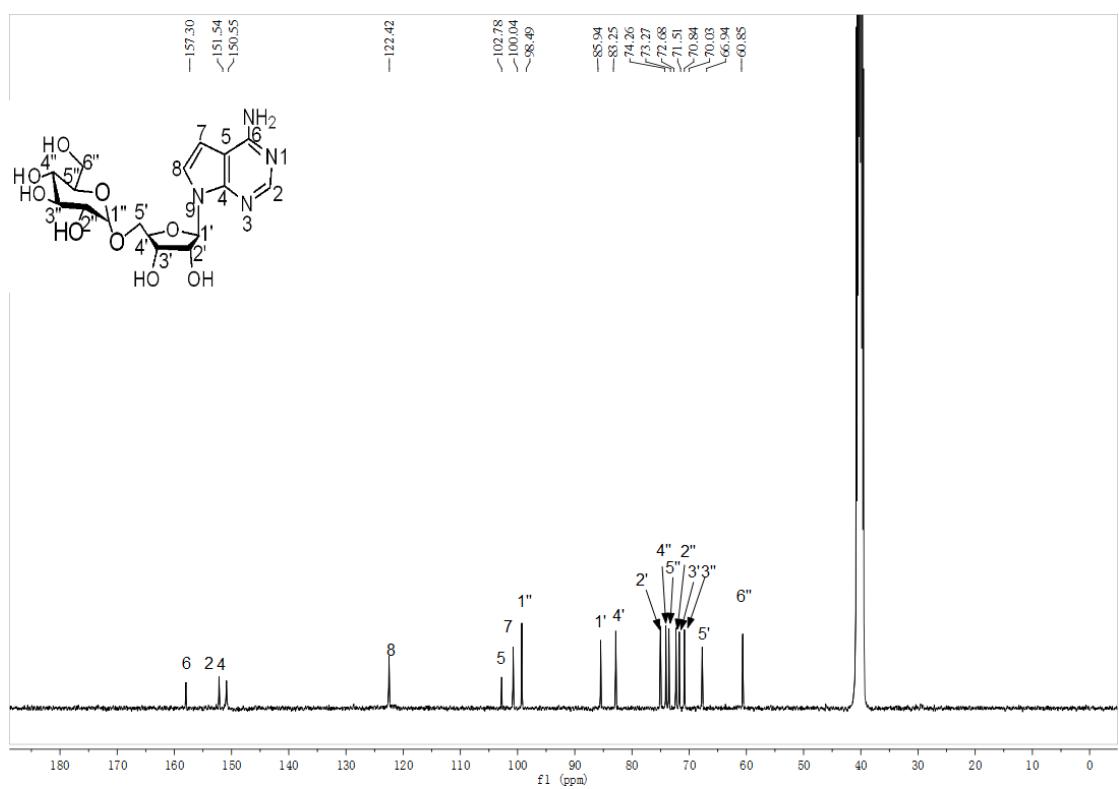
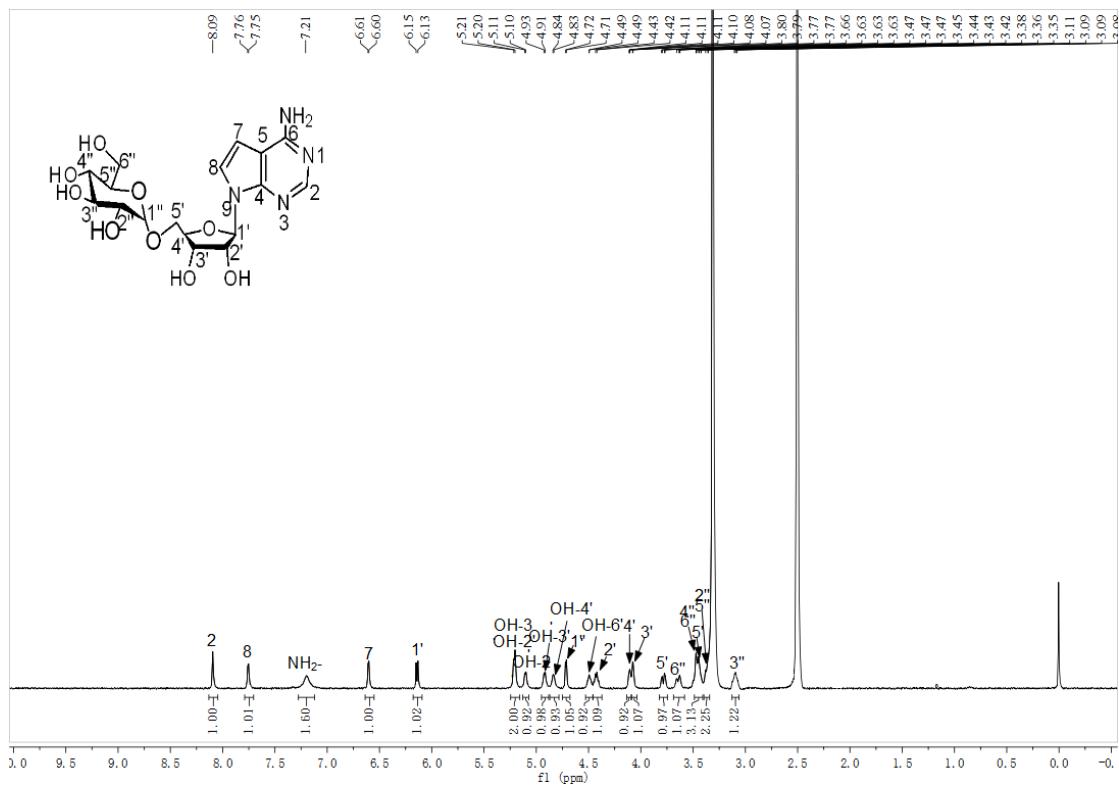


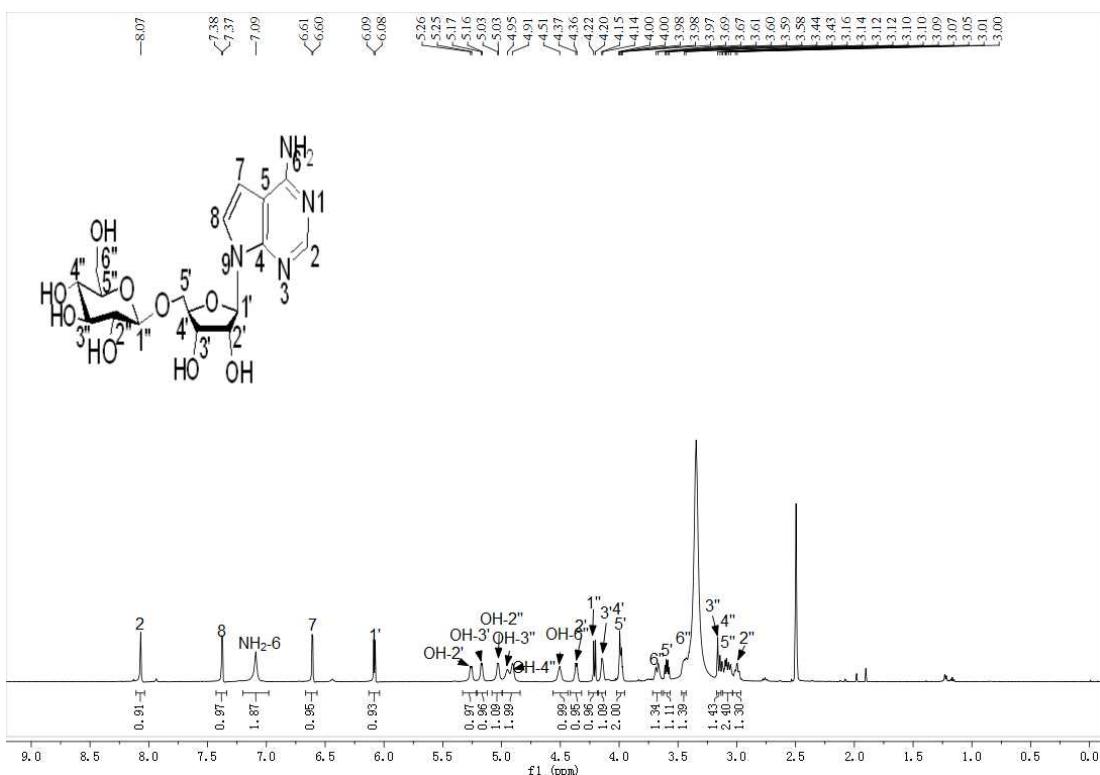
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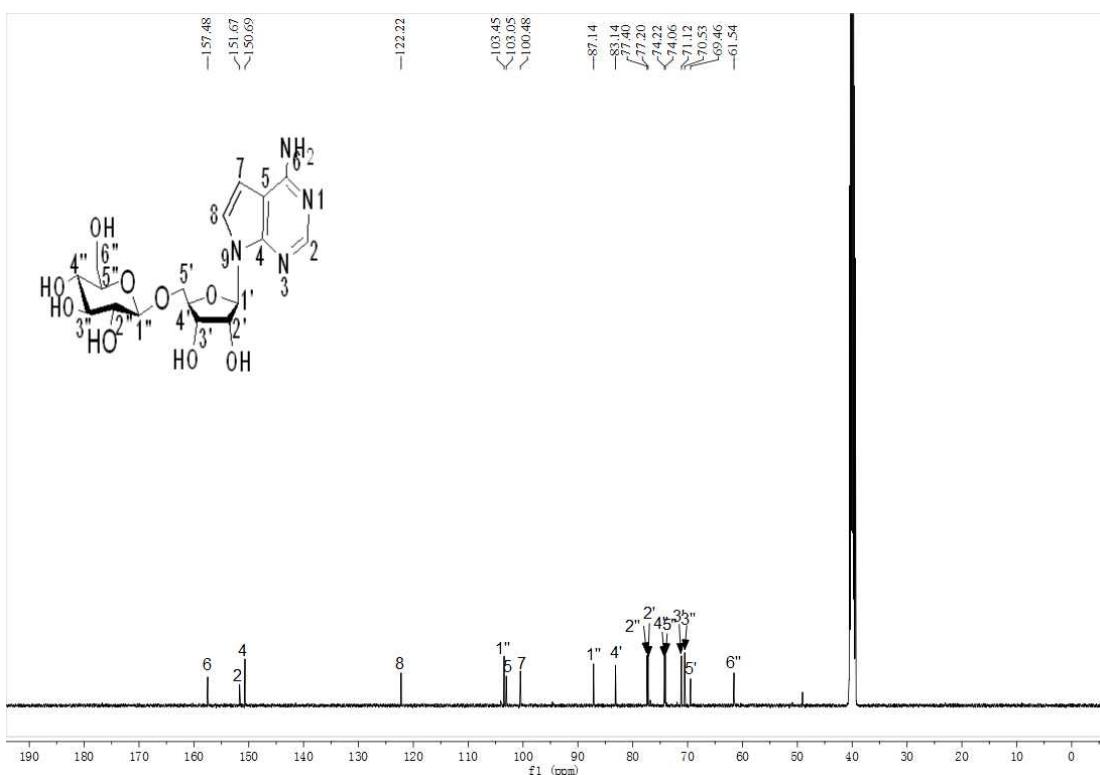
¹³C NMR (DMSO) of 16.







¹H NMR (DMSO) of 17.



¹³C NMR (DMSO) of 17.